

Importance of the vegetation-groundwater-stream continuum to understand transformation of biogenic carbon in aquatic systems — A case study based on a pine-maize comparison in a lowland sandy watershed (Landes de Gascogne, SW France)

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- 1 Importance of the vegetation-groundwater-stream continuum to understand transformation of
- 2 biogenic carbon in aquatic systems a case study based on a pine-maize comparison in a lowland
- 3 sandy watershed (Landes de Gascogne, SW France)
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### 24 Abstract

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- During land-aquatic transfer, carbon (C) and inorganic nutrients (IN) are transformed in soils,
- 26 groundwater, and at the groundwater-surface water interface as well as in stream channels and stream
- 27 sediments. However, processes and factors controlling these transfers and transformations are not well
- 28 constrained, particularly with respect to land use effect. We compared C and IN concentrations in shallow
- 29 groundwater and first-order streams of a sandy lowland catchment dominated by two types of land use:
- 30 pine forest and maize cropland. Contrary to forest groundwater, crop groundwater exhibited oxic

conditions all-year round as a result of higher evapotranspiration and better lateral drainage that decreased the water table below the organic-rich soil horizon, prevented the leaching of soil-generated dissolved organic carbon (DOC) in groundwater, and thus limited consumption of dissolved oxygen  $(O_2)$ . In crop groundwater, oxic conditions inhibited denitrification and methanogenesis resulting in high nitrate (NO<sub>3</sub>; on average  $1{,}140 \pm 485 \,\mu\text{mol}\,\,L^{-1}$ ) and low methane (CH<sub>4</sub>;  $40 \pm 25 \,\text{nmol}\,\,L^{-1}$ ) concentrations. Conversely, anoxic conditions in forest groundwater led to lower NO<sub>3</sub> (25  $\pm$  40  $\mu$ mol L<sup>-</sup>1) and higher CH<sub>4</sub> (1,770  $\pm$ 1,830 nmol L<sup>-1</sup>) concentrations. The partial pressure of carbon dioxide (pCO<sub>2</sub>;  $30,650 \pm 11,590$  ppmv) in crop groundwater was significantly lower than in forest groundwater ( $50,630 \pm 26,070$  ppmv), and was apparently caused by the deeper water table delaying downward diffusion of soil CO<sub>2</sub> to the water table. In contrast, pCO<sub>2</sub> was not significantly different in crop  $(4,480 \pm 2,680 \text{ ppmv})$  and forest  $(4,900 \pm 4,500 \text{ ppmv})$ ppmv) streams, suggesting faster degassing in forest streams resulting from greater water turbulence. Although NO<sub>3</sub> concentrations indicated that denitrification occurred in riparian-forest groundwater, crop streams nevertheless exhibited important signs of spring and summer eutrophication such as the development of macrophytes. Stream eutrophication favored development of anaerobic conditions in crop stream sediments, as evidenced by increased ammonia (NH<sub>4</sub><sup>+</sup>) and CH<sub>4</sub> in stream waters and concomitant decreased in NO<sub>3</sub> concentrations as a result of sediment denitrification. In crop streams, dredging and erosion of streambed sediments during winter sustained high concentration of particulate organic C, NH<sub>4</sub><sup>+</sup> and CH<sub>4</sub>. In forest streams, dissolved iron (Fe<sup>2+</sup>), NH<sub>4</sub><sup>+</sup> and CH<sub>4</sub> were negatively correlated with O<sub>2</sub> reflecting the gradual oxygenation of stream water and associated oxidations of Fe<sup>2+</sup>, NH<sub>4</sub><sup>+</sup> and CH<sub>4</sub>. The results overall showed that forest groundwater behaved as source of CO<sub>2</sub> and CH<sub>4</sub> to streams, the intensity depending on the hydrological connectivity among soils, groundwater, and streams. CH<sub>4</sub> production was prevented in cropland in soils and groundwater, however crop groundwater acted as a source of CO2 to streams (but less so than forest groundwater). Conversely, in streams, pCO<sub>2</sub> was not significantly affected by land use while CH<sub>4</sub> production was enhanced by cropland. At the catchment scale, this study found substantial biogeochemical heterogeneity in C and IN concentrations between forest and crop waters, demonstrating the importance of including the full vegetation-groundwater-stream continuum when

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- estimating land-water fluxes of C (and nitrogen) and attempting to understand their spatial and temporal dynamics.
- 59 Keywords: carbon dioxide, methane, groundwater, stream, land use, pine, maize

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### 1. Introduction

62 Despite their small surface area worldwide (Downing et al., 2012), inland waters have been recognized as 63 key component of the global carbon (C) cycle, constituting a preferential pathway of dissolved and particulate C transport from terrestrial ecosystems to the coastal ocean (Cole et al., 2007; Meybeck, 1982; 64 Ludwig et al., 1996b, 1996a; Meybeck, 1987). Inland waters act as significant sources of carbon dioxide 65 (CO<sub>2</sub>) and methane (CH<sub>4</sub>) to the atmosphere because inland waters are generally supersaturated by CO<sub>2</sub> 66 67 and CH<sub>4</sub> compared to the overlying atmosphere (Abril et al., 2014; Bastviken et al., 2011; Borges et al., 2015; Lauerwald et al., 2015; Raymond et al., 2013; Stanley et al., 2016). 68 Inland waters and specifically small streams are tightly connected to their catchment characteristics such 69 70 as hydrology and land use, as they receive large inputs of C from land (mainly from soils and groundwater), which in turn control the stream biogeochemical processes and the water composition 71 72 (Aitkenhead et al., 1999; Deirmendjian and Abril, 2018; Hotchkiss et al., 2015; Johnson et al., 2006; Jones 73 and Mulholland, 1998; McClain et al., 2003; Polsenaere and Abril, 2012; Bodmer et al., 2016; Findlay et 74 al., 2001; Lehrter, 2006). Groundwater discharge has been recognized as an important source of CO<sub>2</sub> in 75 riverine systems, especially in small streams and headwaters (Deirmendjian and Abril, 2018; Hotchkiss et 76 al., 2015; Johnson et al., 2008; Kokic et al., 2015; Marx et al., 2017; Raymond et al., 2013; Wallin et al., 77 2013). On the contrary to riverine CO<sub>2</sub>, riverine CH<sub>4</sub> is likely to originate from wetlands that generally 78 combine a strong hydrological connectivity with riverine waters and a high productivity (Abril et al., 79 2014; Abril and Borges, 2018). Although some studies found low CH<sub>4</sub> concentrations in the groundwater of Belgium (up to 1.1 µmol L<sup>-1</sup>; Borges et al., 2018; Jurado et al., 2017), other studies found high CH<sub>4</sub> 80

concentrations in the groundwater of Great Britain (up to 295 µmol L<sup>-1</sup>; Bell et al., 2017) and in the Appalachian basin of the USA (up to 2,8000 umol L<sup>-1</sup>; Molofsky et al., 2016). Actually, soil moisture, which controls oxic/anoxic conditions in soil, is the main determinant of terrestrial  $CO_2$  or  $CH_4$  production in soil. As a consequence, CH<sub>4</sub> emissions from soils are high under strictly anaerobic conditions in waterlogged soils whereas CO<sub>2</sub> emissions from soils are high under aerobic conditions in drier soils (Christensen et al., 2003; Moore and Knowles, 1989). Croplands affect water mass balance at the plot scale, especially through irrigation and extraction of groundwater, which results in declining water table in many regions worldwide (Foley et al., 2005; Gleick, 2003; Jackson et al., 2001; Postel, 1999; Rosegrant et al., 2002). Investigating spatial dynamics of CO<sub>2</sub> and CH<sub>4</sub> in groundwater in relation with land use is critical better understanding processes governing their terrestrial production and leaching to groundwater. Croplands cover about 40% of the terrestrial ice-free surface and are often associated with degradation of both ground and surface water quality (Asner et al., 2004; Clague et al., 2015; Foley et al., 2005; Hiscock et al., 1991; Ramankutty and Foley, 1999). Intensive agriculture led to an increase of nitrate (NO<sub>3</sub>) entering ground and surface water environments that has fueled aquatic primary production in surface waters and led to low CO<sub>2</sub> and high CH<sub>4</sub> concentrations, the latter being related to enhanced organic matter delivery in sediments (Borges et al., 2018; Carpenter et al., 1998; Clague et al., 2015; Crawford et al., 2016; Jordan and Weller, 1996; Smith, 2003; Zhou et al., 2017). Additionally, aquatic primary production in crop streams is enhanced as a result of low light limitation (clearing of riparian vegetation), and the excessive transport of sediment-bound organic matter and nutrients to surface waters (Bernot et al., 2010; Lamba et al., 2015; Ramos et al., 2015; Young and Huryn, 1999). Soil erosion rates in agricultural landscapes are one to two times larger than those in areas with native vegetation (Montgomery, 2007; Quinton et al., 2010). Indeed, riparian forest is usually considered stream buffer zones that attenuate stream bank erosion and NO<sub>3</sub> inputs from croplands (Balestrini et al., 2016; Cey et al., 1999; Christensen et al., 2013; Stott, 2005; Wynn and Mostaghimi, 2006). Denitrification represents a permanent removal pathway that limits the extent and impact of NO<sub>3</sub> contamination by transforming NO<sub>3</sub>

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to inert dinitrogen  $(N_2)$ . However, incomplete denitrification can produced nitrous oxide  $(N_2O)$ , a major anthropogenic ozone-depleting substance (Ravishankara et al., 2009). On the contrary to croplands, forests are known to export fewer nutrients by limiting runoff and leakage of nutrients (Canton et al., 2012; Onderka et al., 2010).

Land use effects on both water composition and biogeochemical processes have been studied in streams and groundwater (Barnes and Raymond, 2010, 2009; Bernot et al., 2010; Bodmer et al., 2016; Jeong, 2001; Lehrter, 2006; Masese et al., 2017; Raymond and Cole, 2003; Rodrigues et al., 2018; Salvia-Castellví et al., 2005; Vidon et al., 2008; Wilson and Xenopoulos, 2009; Young and Huryn, 1999; Zhang et al., 2018), but land use studies with simultaneous groundwater and stream sampling are more scarce (Bass et al., 2014; Borges et al., 2018; Hu et al., 2016). The objective of this study was to understand how two contrasting types of land use (pine forest and maize cropland) affected C and inorganic nutrient (IN) concentrations in shallow groundwater and in first-order streams of a sandy lowland catchment. We hypothesized that the biogeochemical variability between crop groundwater and forest groundwater was due to agricultural practices that affect N inputs (fertilizer) and water mass balance (irrigation and drainage). We hypothesized that the biogeochemical variability between crop and forest streams originate from differential lateral export of C and IN from two distinct sources (i.e., crop groundwater and forest groundwater) because of a strong hydrological connection between groundwater and streams in the studied catchment.

#### 2. Materials and Methods

### 2.1. Study site

The Leyre catchment (2,100 km²) is located in the southwestern part of France. This is a flat coastal plain with a mean slope lower than 0.125% and a mean altitude lower than 50 m (Jolivet et al., 2007). The lithology is relatively homogeneous and composed of sandy permeable surface layers dating from the

Plio-Quaternary period (Legigan, 1979; Bertran et al., 2009, 2011). The soils are podzols characterized by a low pH ( $\approx$ 4), low nutrient availability, low cationic exchange capacity, and high organic C content that can reach 50 g per kg of soil (Augusto et al., 2010; Lundström et al., 2000). In Leyre sandy podzols, the low clay and silt content causes a low soil water retention (Augusto et al., 2010). The study area was a vast wetland until the 19<sup>th</sup> century, when a wide forest of maritime pine was sown following a landscape drainage campaign resulting from an imperial decree of Napoleon III in 1857 (Jolivet et al., 2007). Currently, the catchment is mainly occupied by C<sub>3</sub> pine forest (approximately 85%), with a modest proportion of C<sub>4</sub> maize cropland (approximately 15%) (Fig. 1; Jolivet et al., 2007). Following catastrophic forest wildfires, the maize croplands were installed during the second half of the 20th century. Consequently, their spatial distribution was not based on soil properties, as confirmed by the similar mean values of soil texture in local croplands and forests (Augusto et al., 2010; Jolivet et al., 2003). During the maize cropping season (usually May to November), farmers irrigate the plots by pumping shallow groundwater (~1-5 m deep) almost daily to maintain adequate soil moisture status, whereas maritime pine stands are never irrigated (Govind et al., 2012). As N is not limiting for tree growth in our study region (Trichet et al., 2009), forests are never fertilized with N. Conversely, croplands generally receive two N fertilizer applications annually, a first at the beginning of May (30–50 kg N ha<sup>-1</sup>), and second at the beginning of June with 200–250 kg N ha<sup>-1</sup> (Canton et al., 2012; Jambert et al., 1997; Ulrich et al., 2002). Additionally, in order to maintain soil pH in the 5.5–6.0 range, local maize croplands are limed with crushed limestone (CaCO<sub>3</sub>) containing a small portion of dolomite (CaMg(CO<sub>3</sub>)<sub>2</sub>) (10 t ha<sup>-1</sup> right after forest conversion and then 0.5 t ha<sup>-1</sup> an<sup>-1</sup>; Jolivet et al., 2003). The climate is oceanic with a mean annual air temperature of 13°C and a mean annual precipitation of 930 mm (Moreaux et al., 2011). Owing to the low slope, the low soil water retention and the high permeability of the soil (i.e., hydraulic conductivity is approximately 40 cm h<sup>-1</sup>, Corbier et al., 2010), the percolation of rain water is fast (55 cm h<sup>-1</sup> on average, Vernier and Castro, 2010). Consequently, surface runoff does not occur as the excess of rainfall percolates into the soil and recharges the shallow groundwater, causing the

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water table to rise. The sandy permeable surface layers contain a free and continuous water table that is strongly interconnected with the superficial river network. This is facilitated by a dense network of drainage ditches, initiated in the 19<sup>th</sup> century and currently maintained by forest managers in order to enhance tree regeneration and growth (Thivolle-Cazat and Najar, 2001). During the sampling period, channels of some crop streams were dredged before they began to flow again. This was done to optimize local cropland drainage and to feed croplands with IN and organic residuals found in the stream sediments. To increase soil permeability and to optimize lateral drainage in local maize croplands, farmers practice subsoiling and agricultural ditches are generally deeper (2.0–2.5m) than forest ditches (1.0m).

# 2.2. Sampling strategy

We defined order 0 as groundwater and order 1 as streams and ditches either having no tributaries or being seasonally dry (from June to December during our sampling period). We selected 17 sampling stations (5 shallow groundwater and 12 first-order streams) within the Leyre catchment (Tab. 1; Fig. 1). The groundwater sampling stations were located in maize cropland (n=2), pine forest (n=2; one is the Bilos station (FR-Bil) of the ICOS Research infrastructure) and in a riparian forest adjacent to a maize cropland (n=1; Tab. 1; Fig. 1). The stream sampling stations were chosen based on the different proportions of croplands in their respective catchments (Tab. 1; Fig. 1). Groundwater was sampled for temperature, electrical conductivity (EC), pH, dissolved oxygen (O<sub>2</sub>), methane (CH<sub>4</sub>), partial pressure of CO<sub>2</sub> (pCO<sub>2</sub>), total alkalinity (TA), dissolved inorganic carbon (DIC), stable isotope composition of the dissolved inorganic carbon ( $\delta^{13}$ C-DIC), dissolved organic carbon (DOC), ammonia (NH<sub>4</sub><sup>+</sup>), nitrate (NO<sub>3</sub><sup>-</sup>) and dissolved iron (Fe<sup>2+</sup>). For groundwater, we took the precaution to renew the water in the piezometers by pumping with a submersible pump before sampling. Groundwater was then sampled once the stabilization (approximately 10 min) of groundwater temperature, pH, EC and O<sub>2</sub> monitored with portable probes was observed. Streams were sampled for the 

same parameters, plus total suspended matter (TSM), particulate organic carbon (POC) and the POC content of the TSM (POC%).

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### 2.3. Field measurements and laboratory analyses

Groundwater and streams were sampled at approximately monthly time intervals between Jan. 2014 and Jul. 2015 (Tab. S1). In total, throughout the sampling period, we sampled 55 groundwaters and 137 stream waters. The pCO<sub>2</sub> in groundwater and streams was measured directly using an equilibrator (Frankignoulle and Borges, 2001; Polsenaere et al., 2013) following the procedure of Deirmendjian and Abril (2018). We stored the total alkalinity (TA) samples in polypropylene bottles after filtration using a syringe equipped with glass fiber filters (GF/F; 0.7 µm). TA was then analyzed on filtered samples by automated electro-titration on 50 mL samples with 0.1N HCl as the titrant. The equivalence point was determined from pH between 4 and 3 with the Gran method (Gran, 1952). Precision based on replicate analyses was better than  $\pm 5 \mu \text{mol L}^{-1}$ . For samples with a very low pH (<4.5), we bubbled the water with atmospheric air in order to degas the CO<sub>2</sub>. Consequently, the initial pH increased above 5, and the TA titration was then performed (Abril et al., 2015). We calculated DIC from pCO<sub>2</sub>, TA, and temperature measurements using the carbonic acid dissociation constants of Millero (1979) and the CO<sub>2</sub> solubility from Weiss (1974), using the CO<sub>2</sub>SYS software (Lewis et al., 1998). The  $\delta^{13}$ C-DIC samples were collected using 120 mL glass serum bottles sealed with a rubber stopper and treated with 0.3 mL of HgCl<sub>2</sub> at 20 g L<sup>-1</sup> to avoid any microbial respiration during storage. Vials were carefully sealed such that no air remained in contact with samples and were stored in the dark to prevent photo-oxidation. The  $\delta^{13}$ C-DIC measurements were performed with the headspace technique using an isotope ratio mass spectrometer coupled to an elemental analyser (EA-IRMS, Micromass

IsoPrime) equipped with a manual gas injection port as described in Gillikin and Bouillon (2007).

203 CH<sub>4</sub> was also measured using a headspace technique in 60 mL glass serum bottles. The headspace was 204 created with 10 mL of N<sub>2</sub> gas. We then injected 0.5 mL of the headspace in a gas chromatograph equipped 205 with a flame ionization detector (GC-FID). 206 DOC samples were obtained after filtration in the field through pre-combusted GF/F (0.7 µm). DOC filtrates were stored in pre-combusted Pyrex vials (25 mL), acidified with 50 µL of 37% HCl to reach pH 207 208 2, and kept at 4 °C in the laboratory before analysis. The DOC concentrations were measured with a 209 SHIMADZU TOC 500 analyzer (in TOC-IC mode), using a technique based on thermal oxidation after a DIC removal step (Sharp, 1993). The repeatability was better than 0.1 mg L<sup>-1</sup>. 210 211 The water for TSM and POC measurements was filtered through pre-weighed and pre-combusted GF/F 212 glass fiber filters (0.7 µm). The filters were dried at 60 °C and stored in the dark, and subsequently, TSM 213 was determined by gravimetry. POC was measured using the same filter. The filters were acidified in 214 crucibles with 2N HCl to remove carbonates and were then dried at 60 °C to remove inorganic carbon and 215 most of the remaining acid and water (Etcheber et al., 2007). POC content was measured by combustion 216 (1500 °C) using a LECO CS 200 analyzer and the CO<sub>2</sub> formed was determined quantitatively by infrared absorption. POC in  $\mu$ mol L<sup>-1</sup> and POC% were then calculated. The uncertainty was  $\pm 0.05\%$  of TSM. 217 218 For IN determination, water was filtered through a 0.20 µm cellulose acetate syringe membrane. Subsamples for Fe<sup>2+</sup> were acidified with 37% HCl to prevent precipitation of iron oxide, whereas 219 subsamples for NH<sub>4</sub> and NO<sub>3</sub> were not acidified but kept frozen until later analyses. Then, NH<sub>4</sub>, NO<sub>3</sub>, 220 and Fe<sup>2+</sup> were analyzed by colorimetry according to standard techniques. NH<sub>4</sub><sup>+</sup> was analyzed following the 221 222 procedure of Harwood and Kühn (1970). NO<sub>3</sub> was analyzed by flow injection analysis following the procedure of Anderson (1979). Fe<sup>2+</sup> was analyzed using the ferrozine method (Stookey, 1970). Precision 223 was  $\pm 10\%$  for NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub>, and was  $\pm 5\%$  for Fe<sup>2+</sup>. 224 EC, temperature, O<sub>2</sub>, and pH were measured using portable probes (WTW®). Before each field trip, the 225 226 pH probe was calibrated using two NBS buffer solutions (4 and 7), the oxygen polarographic probe was

calibrated to 100% in a humid atmosphere and the conductivity probe was calibrated using a salinity standard.

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K-means clustering analysis (MacQueen, 1967) was used to classify waters either as forest-dominated or as cropland-affected (Tab. 1). Indeed, K-means clustering analysis allows partitioning a dataset into k groups (i.e., clusters) pre-specified by the analyst (MacQueen, 1967). Contrary to forest waters at our study site, crop waters exhibit disproportionately higher NO<sub>3</sub> concentration as a result of N fertilizer use on maize cropland (Canton et al., 2012; De Wit et al., 2005; Jambert et al., 1997, 1994). Consequently, in the K-means clustering analysis we used NO<sub>3</sub> concentration data as a proxy to establish a statistical distinction between forest and crop waters (Tab. 1). K-means clustering analysis was performed one time with the groundwater dataset (but excluding the riparian groundwater) and a second time with the firstorder streams dataset. We excluded data from riparian groundwater because we have considered riparian groundwater as a cluster itself (Tab. 1). Principal component analysis (PCA) was used to condense multivariate information on correlated biogeochemical parameters to a set of uncorrelated variables called principal components (further referred to as dimensions). PCA was performed one time with a dataset consisting of each measured parameter in groundwater (but excluding the riparian groundwater) and a second time with the corresponding firstorder streams dataset. PCA was performed separately for groundwater and streams because particulate parameters were not present in groundwater. If PCA were not performed separately for groundwater and streams, all data from groundwater would have been removed from the analysis (indeed, if one parameter is missing for a given sampling station, the sampling station is entirely deleted from the PCA). In addition, performing the PCA separately for groundwater and streams led to information that was more robust with respect to the biogeochemical variability induced by land use, in either groundwater or streams. However,

to observe whether the two groundwater (crop and forest) and two streams (crop and forest) sources could be distinguished mathematically in one PCA, we performed an additional PCA with data from both groundwater and streams that excluded particulate parameters from the analysis. All concentrations data were log-transformed prior to PCA. The PCAs showed the biogeochemical variability across forest, cropland, and hydrological seasons in either groundwater or first-order streams.

Later, non-parametric bivariate analyses (Mann-Whitney statistical tests) were used to estimate if hydrological seasons or increasing stream order significantly influenced the concentration of a biogeochemical parameter. Linear regressions were performed to model the relationships between two variables by fitting a linear equation to observed data.

K-means clustering analysis (package Stats) and PCA analysis (package FactoMineR for analysis and package factoextra for visualization; Kassambra and Mundt, 2017; Lê et al., 2008) were performed with R software version 3.1.4 (R Core Team, 2018). Mann-Whitney tests and linear regressions were performed with Graph Pad Prism version 7 software.

### 3. Results

### 3.1. Hydrology

In previous work based on the same dataset, but excluding cropland sampling stations, we identified two major hydrological seasons (Deirmendjian and Abril, 2018). One defined a high flow period as two relatively short flood events that occurred in Jan. 2014–Mar. 2014 and in Feb. 2015–Mar. 2015, whereas we defined the base flow period as two longer periods of low flow occurring in Apr. 2014–Jan. 2015 and Apr. 2015–Jul. 2015. During high flow, the average and the maximum river flows were 50 m<sup>3</sup> s<sup>-1</sup> and 119 m<sup>3</sup> s<sup>-1</sup>, respectively. During base flow, the average and the minimum river flow were 10 m<sup>3</sup> s<sup>-1</sup> and 5 m<sup>3</sup> s<sup>-1</sup>, respectively. The water tables in the forest, riparian forest, and cropland exhibited similar temporal fluctuations but with a different intensity, and the forest had an overall higher water table depth than the

cropland (Fig. 2). The water table in the riparian area exhibited intermediate depth between the forest and cropland sites (Fig. 2). As surface runoff was negligible in the studied sandy and flat catchment, most of the stream water likely originated from groundwater discharge.

To investigate the temporal variability of the studied biogeochemical parameters, we chose to rely on hydrological regimes (high flow and base flow periods) rather than on temperature periods (seasons). At our study site climate was oceanic (by definition very temperate) and the amplitude of the water temperature was not as high as the amplitude of the river flow. As an example, Leyre River (main stem) flow could be up to 119 m³ s⁻¹ and could be down to 5 m³ s⁻¹, whereas the highest water temperature amplitude occurred in forest streams and was 6.4–25.8 °C (Tab. 2). Additionally, most of the lateral export occurred during the short periods of high flow (up to 90% for DOC, Deirmendjian et al., 2018). Thus, characterizing biogeochemical variability and biogeochemical processes in relation with land use during this hydrological period was important. Furthermore, the seasonality induced by water temperature was to a certain extent included in the defined hydrological regimes since the high flow period was associated with lower water temperatures while the base flow period was associated with higher water temperatures (Tabs. 2-4).

# 3.2. K-means clustering analysis

For both hydrological seasons (i.e., high and base flow), we partitioned each sampling stations (excepting riparian groundwater), into either cropland-affected or forest-dominated waters (Tab. 1). K-means clustering analysis produced satisfactory results. Logically, groundwater located in cropland was classified as crop water whereas groundwater located in forest was classified as forest water (Tab. 1). Stream sampling stations having more than 30% of croplands in their respective catchment were always classified as crop waters (Tab. 1). Stream sampling stations having less than 8% of croplands in their respective catchment were always classified as forest waters excepting two times (Tab. 1). These two stream

sampling stations were located a few kilometers downstream from important maize croplands.

Specifically, one station was a ditch strongly vegetated during the base flow period that showed signs of N fertilizer uptake from upstream cropland and, therefore, this ditch was logically classified as a crop station during base flow (Tab. 1). One station was a stream that exhibited a high water flow during the high flow period, which probably increased the upstream cropland influences during this hydrological period and, therefore, this stream was logically classified as a crop station during high flow (Tab. 1).

Excepting one strictly forested headwater, the other sampled streams were not strictly forested or cropped (Tab. 1). Consequently, as explained further, some biogeochemical variability between forest and crop streams was introduced by simple water mixing from two distinct sources: forest groundwater and crop groundwater. We used the term crop stream to indicate a stream classified as a crop-affected one, although

#### 3.3. Land use influence on water composition of shallow groundwater

such stream was a forest stream affected by cropland rather that a strictly crop stream.

PCA on the groundwater dataset revealed that groundwater biogeochemical variability was strongly dependent on land use (maize cropland vs. pine forest) and hydrological seasons (base flow vs. high flow) (Figs. 3a, b). The first three PCA dimensions covered 44%, 17.5% and 10.5% of the total variance within the dataset, respectively (Figs. 3a, b).

PCA dimension 1 clearly separated forest groundwater from crop groundwater based on two groups of variables negatively correlated with one another (Fig. 3a). One group of variables was characterized crop groundwater and was composed of EC, NO<sub>3</sub>-, δ<sup>13</sup>C-DIC, and O<sub>2</sub>, whereas the second group of variables was characterized forest groundwater and was composed of DIC, pCO<sub>2</sub>, CH<sub>4</sub>, Fe<sup>2+</sup>, and NH<sub>4</sub>+ (Fig. 3a). Indeed, we observed that the yearly average of EC (+270 μS cm<sup>-1</sup>), NO<sub>3</sub>- (+1,115 μmol L<sup>-1</sup>), δ<sup>13</sup>C-DIC (+6.9‰), and O<sub>2</sub> (+200 μmol L<sup>-1</sup>) were higher in crop groundwater than in forest groundwater and were significantly and positively affected by cropland cover (Tab. 2; Figs. 4b, c, f, j). Conversely, we observed

higher DIC (+1,010  $\mu$ mol L<sup>-1</sup>), pCO<sub>2</sub> (+19,985 ppmv), CH<sub>4</sub> (+1,730  $\mu$ mol L<sup>-1</sup>), Fe<sup>2+</sup> (+14.1  $\mu$ mol L<sup>-1</sup>), and 323  $NH_4^+$  (+4.1 µmol L<sup>-1</sup>) in forest groundwater than in crop groundwater; these were significantly and 324 positively affected by forest cover (Tab. 2; Figs. 4d, e, g, h, k). In riparian groundwater, EC, NO<sub>3</sub>, δ<sup>13</sup>C-325 DIC, O<sub>2</sub>, DIC, pCO<sub>2</sub>, and CH<sub>4</sub> exhibited intermediate values between the groundwater of forest and crop 326 sites, whereas Fe<sup>2+</sup> and NH<sub>4</sub><sup>+</sup> were low and close to those found in crop groundwater (Tab. 2; Figs. 4b, c, 327 328 d, e, f, g, h, j, k). 329 In crop groundwater, EC,  $NO_3^-$ ,  $\delta^{13}$ C-DIC, and  $O_2$  were not significantly affected by hydrological seasons (Figs. 3a, S2c, h, j, k, l). However,  $\delta^{13}$ C-DIC (+1.1%), NO<sub>3</sub><sup>-</sup> (+120  $\mu$ mol L<sup>-1</sup>), and DOC (+130  $\mu$ mol L<sup>-1</sup>) 330 were slightly higher (but not significantly) during base flow compared to high flow (Tab. 3; Figs. S2c, h, j, 331 332 k, l). In forest groundwater, pCO<sub>2</sub>, DIC, CH<sub>4</sub>, and DOC were significantly affected by hydrological 333 seasons (Tab. 3; Figs. S2g, h, k, l). DOC (+1,490 µmol L<sup>-1</sup>) was significantly higher during high flow, whereas pCO<sub>2</sub> (+30,980 ppmv), DIC (+1,330  $\mu$ mol L<sup>-1</sup>) and CH<sub>4</sub> (+1,780 nmol L<sup>-1</sup>) were significantly 334 335 higher during base flow (Tab. 3; Figs. S2 and S4g, h, k, l). In crop and riparian groundwater, we also 336 observed higher pCO<sub>2</sub> and DIC values during base flow, but with lower intensities than in forest 337 groundwater (Tab. 3; Figs. S2h, k).

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# 3.4. Land use influence on water composition of first-order streams

Figures 3c-d present PCA based on first-order streams data set. The first three PCA dimensions covered 28.6%, 18.5% and 13.8% of the total variance within the dataset, respectively (Figs. 3c, d).

Interestingly, the PCA based on the first-order streams dataset did not clearly separate crop streams from forest streams as it did for groundwater dataset (Figs. 3a-d). This implied lower spatial variability in streams in relation to land use in than in groundwater (Tabs. 2-4; Fig. 4). Nevertheless, a land use gradient was observed on PCA dimension 2 (Figs. 3c, d). PCA dimension 2 was best defined by a group of variables composed of EC, CH<sub>4</sub>, NO<sub>3</sub>-, NH<sub>4</sub>+, DOC, TSM, and POC, which collectively characterized crop

streams (Figs. 3c, d). On a yearly average basis, significantly higher EC (+105 µS cm<sup>-1</sup>), CH<sub>4</sub> (+220 µmol  $L^{-1}$ ),  $NO_3^{-1}$  (+265 µmol  $L^{-1}$ ),  $NH_4^{+1}$  (+4.3 µmol  $L^{-1}$ ), DOC (+135 µmol  $L^{-1}$ ), TSM (+3.3 mg  $L^{-1}$ ), and POC(+70 μmol L<sup>-1</sup>) were observed in crop streams compared to forest streams (Tab. 2; Figs. 4b, c, d, g, l, m, o). High CH<sub>4</sub>, NH<sub>4</sub><sup>+</sup>, and DOC concentrations were characteristics of forest groundwater, but in streams, these parameters were characteristic of crop streams (Figs. 3a-d). In addition, pCO<sub>2</sub>, DIC,  $\delta^{13}$ C-DIC and O<sub>2</sub> were not able to separate crop streams from forest streams as they did for groundwaters (Figs. 3a-d). On a yearly average basis, no significant differences were observed between crop and forest streams for these four parameters (Figs. 3f, h, j, k). The relatively low temporal variability between high and base flow periods observed in both crop and forest streams for the studied parameters did not allow the PCA based on stream data set to clearly separate base flow samples from high flow samples (Tab. 4; Figs. 3c-d, S3, S4). Nevertheless, in crop streams, pH (+0.2) and  $\delta^{13}$ C-DIC (+3.8%) were significantly higher during base flow while pCO<sub>2</sub> (-1,160 ppmv), NO<sub>3</sub> (-130 µmol L<sup>-1</sup>) and DOC (-230 µmol L<sup>-1</sup>) were significantly lower during the same period (Tab. 4; Figs. S3a, j, c, h, l, m, o). In streams, TSM and POC were significantly higher during high flow but with a higher intensity in crop streams than in forest streams (Tab. 4; Fig. S3m, o). Interestingly, in both forest and crop streams, POC% was not significantly affected by hydrological regime (Tab. 4; Fig. S3n).

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### 3.5. Upstream-downstream distribution of biogeochemical parameters

To explore the influence of land use on water composition at the groundwater-stream continuum, we observed the upstream-downstream (groundwater-stream) distribution of biogeochemical parameters along forest and crop continuums (Tab. 2; Fig. 4). Along both types of continuum, some parameters (i.e., pCO<sub>2</sub>, TA, DIC,  $\delta^{13}$ C-DIC, pH, O<sub>2</sub>) exhibited the same upstream-downstream distribution, whereas other

parameters (i.e., EC, NO<sub>3</sub>, NH<sub>4</sub><sup>+</sup>, Fe<sup>2+</sup>, CH<sub>4</sub>, DOC) exhibited a different upstream-downstream distribution (Tab. 2; Fig. 4).

In crop and forest continuums, we observed strong spatial patterns for pCO<sub>2</sub>, TA, DIC, δ<sup>13</sup>C-DIC, and pH:

pCO<sub>2</sub> and DIC decreased while TA remained more or less constant, and  $\delta^{13}$ C-DIC and pH increased (Tab. 2; Figs. 4a, h, i, j, k). However, a larger decrease in pCO<sub>2</sub> levels in the forest continuum suggested a more intense degassing in forest streams (Tab. 2; Fig. 4h). We also observed an increase in O<sub>2</sub> in both types of continuum, which could result from stream ventilation, although with a higher intensity in forest continuum (Tab. 2; Figs. 4f, h).

EC decreased downstream in the crop continuum, but did not in the forest continuum where EC remained very stable and much lower than in the crop continuum (Tab. 2; Fig. 4b). NO<sub>3</sub><sup>-</sup> decreased downstream between groundwater and streams in croplands, and in contrast, in forests NO<sub>3</sub><sup>-</sup> increased downstream between groundwaters and streams (Tab. 2; Fig. 4c). NH<sub>4</sub><sup>+</sup>, Fe<sup>2+</sup>, and CH<sub>4</sub> decreased in the forest continuum but they increased in the crop continuum (Tab. 2; Figs. 4d, e, g). DOC significantly decreased in the forest continuum but remained stable in the crop continuum (Tab. 2; Fig. 4l). TSM and POC were significantly higher in crop relative to forest streams; however, similarly high POC% (28%) was observed in both types of streams (Tab. 2; Fig. 4m, n, o).

# 3.6. Biogeochemistry dynamics in the groundwater-stream continuum

PCA with groundwater and streams datasets indicated mathematically that streams were fed with two distinct sources: forest groundwater mostly characterized by high pCO<sub>2</sub>, DIC, and CH<sub>4</sub> concentrations and crop groundwater mostly characterized by high NO<sub>3</sub><sup>-</sup> concentrations (Figs. 5). Forest and crop streams were characterized by higher O<sub>2</sub>,  $\delta^{13}$ C-DIC and pH values than in groundwater (Figs. 5). In this PCA, the distinction between forest and crop streams was primarily a function of NO<sub>3</sub><sup>-</sup>, crop streams points were moved upward along dimension 2 (Figs. 5). Throughout the sampling period, we observed a negative

394 linear relationship (R<sup>2</sup>=0.6, p<0.001, n=192) between CO<sub>2</sub> and O<sub>2</sub> for all sampled groundwater and 395 streams (Fig. 6a). On the one side, stream samples were mostly characterized by high O<sub>2</sub> (mean was 290 396 umol L<sup>-1</sup>) and low CO<sub>2</sub> (mean was 4,480 ppmv), excepting some forest streams during summer that were characterized by low O<sub>2</sub> (down to 110 µmol L<sup>-1</sup>) and high CO<sub>2</sub> (up to 27,200 ppmv) (Tab. 2; Figs. 6a, S4f, 397 398 h). On the other side, anoxic conditions associated with high CO<sub>2</sub> (mean was 50,630 ppmv) were 399 characteristic of forest groundwater, whereas crop groundwater exhibited O<sub>2</sub> (mean was 220 µmol L<sup>-1</sup>) and 400 CO<sub>2</sub> (mean was 30,650 ppmy) intermediate between streams and forest groundwater (Tab. 2; Fig. 6a). In 401 forest groundwater, DOC was negatively and linearly correlated with  $CO_2$  (R<sup>2</sup>=0.4, p < 0.001, n=22) 402 suggesting that part of groundwater CO<sub>2</sub> came from degradation of groundwater DOC (Fig. 6b). A 403 comparison of CO<sub>2</sub> and CH<sub>4</sub> for all sampled groundwaters and streams showed that a large portion of the 404 CO<sub>2</sub> and CH<sub>4</sub> in forest streams could come from forest groundwater discharge (Tab. 2; Fig. 6e). In crop streams, CH<sub>4</sub>, NH<sub>4</sub><sup>+</sup>, and Fe<sup>2+</sup> could not originate from crop groundwater discharge since they had much 405 lower CH<sub>4</sub>, NH<sub>4</sub><sup>+</sup>, and Fe<sup>2+</sup> concentrations than crop streams (Tab. 2; Fig. 6e, 7e, 7f). We observed a 406 407 positive linear relationship between CH<sub>4</sub> and NH<sub>4</sub><sup>+</sup> (R<sup>2</sup>=0.4, p<0.001, n=53) in crop streams, 408 demonstrating that these two compounds may come from the same source (Fig. 7d). Conversely, a comparison of O<sub>2</sub> and NH<sub>4</sub><sup>+</sup> or Fe<sup>2+</sup> in forest streams indicated that NH<sub>4</sub><sup>+</sup> and Fe<sup>2+</sup> were mostly discharged 409 through forest groundwater (Fig. 7e, f). In forest streams, the negative linear relationship between O<sub>2</sub> and 410  $NH_4^+$  (R<sup>2</sup>=0.1, p-value<0.05, n=70), Fe<sup>2+</sup> (R<sup>2</sup>=0.5, p-value<0.001, n=77), or  $CH_4$  (R<sup>2</sup>=0.1, p-value<0.001, 411 412 n=77) suggested oxidation of these reduced compounds in the stream water column (Fig. 7e, f). We 413 observed a gradient of NO<sub>3</sub> concentration, from high values to low values, between crop groundwater (mean was 1,140 μmol L<sup>-1</sup>), to riparian groundwater and crop streams (310 and 340 μmol L<sup>-1</sup>, 414 415 respectively), to forest streams (75 µmol L<sup>-1</sup>) and to forest groundwater (25 µmol L<sup>-1</sup>) (Tab. 2; Fig. 7a, b, 416 c). In crop streams, a large share of riverine NO<sub>3</sub> could be discharged through crop groundwater. 417 Conversely, NO<sub>3</sub> concentration in forest streams could not be explained by NO<sub>3</sub> concentration in forest 418 groundwater (Tab. 2; Fig. 7a, b, c, d). In crop groundwater, high NO<sub>3</sub> concentrations were associated with

low CH<sub>4</sub> concentrations. In crop streams, high NO<sub>3</sub> concentrations could be related to high CH<sub>4</sub> concentrations (Fig. 7c).

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#### 4. Discussion

### 4.1. Water table depth in relation to land use

At the studied catchment scale, lithology, topography, soils, and precipitation are more or less uniform (Augusto et al., 2010; Jolivet et al., 2003). At the plot scale, spatial variability of water table depth in relation to land use was thus necessarily dependent on how water outputs (drainage, evapotranspiration or groundwater storage) of the water mass balance were human-affected (Govind et al., 2012; Stella et al., 2009). Local forests are never irrigated, conversely, irrigation with extraction of groundwater (that decreases groundwater storage) in local croplands could strongly bias the water mass balance at the plot scale since about half of the water diverted for irrigation is rapidly consumed through evapotranspiration (e.g., Jackson et al; 2001). Additionally, evapotranspiration in maize croplands is typically higher than in forests owing to the larger stomatal conductance that makes the exchange of C and water between the biosphere and the atmosphere much easier (Govind et al., 2012; Stella et al., 2009). Other studies have shown that the combination of subsoiling practices (increasing soil permeability) with deep agricultural ditches in croplands also affected water mass balance at the plot scale by enhancing lateral drainage of groundwater (Evans et al., 1996; Robinson et al., 1985). From an 8 year survey of local cropland, Juste et al (1982) showed that lateral drainage strongly affected the water mass balance at the plot scale as it represented an annual mean of 637 mm (70% of the amount of precipitation), whereas precipitation was estimated at 922 mm. At the forest plot scale, lateral drainage represented an annual mean of 182 mm (20% of the amount of precipitation), whereas precipitation was estimated at 895mm (Deirmendjian et al, 2018). At the study site, deeper water table in croplands was thus a consequence of a higher evapotranspiration and more lateral drainage than in forest. As explained further, water table depth is an

important determinant for understanding the biogeochemical variability in groundwater in relation to land use.

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# 4.2. Dynamic of $O_2$ , DOC, DIC and $\delta^{13}$ C-DIC in groundwater: a combination of hydrological,

### physical and metabolic processes

In other aquifer systems worldwide, several studies have observed a significant positive correlation between groundwater O<sub>2</sub> concentration and depth to water (Datry et al., 2004; Foulquier et al., 2010; Goldscheider et al., 2006; McMahon and Chapelle, 2008; Pabich et al., 2001). Where the water table is close to the soil surface, groundwater O<sub>2</sub> consumption is likely rapid because of incomplete degradation of soil-generated labile DOC in the unsaturated zone. On the contrary, where the water table is far from the soil surface, strong oxygen depletion in the vicinity of the water table does not occur since the higher residence time of infiltrating water results in almost complete degradation of soil-generated DOC in the unsaturated zone (Malard and Hervant, 1999; Starr and Gillham, 1993). A regional study in forest soils of Switzerland (Hagerdon et al., 2000) and a study compiling a global database of soil carbon (Camino-Serrano et al., 2014) both found that soil-generated DOC was preferentially mobilized under reducing conditions in soils because of dissolution of Fe oxides. Deeper water tables in croplands do not reach topsoil that exhibits high labile organic C content. Thus, reducing conditions in topsoil and the leaching of soil-generated DOC are prevented as is the consumption of the groundwater O<sub>2</sub> stock, as occurs in forests during high flow stages (Tab. 3; Fig. S2l; Deirmendjian et al. 2018). Therefore, groundwater pCO<sub>2</sub> was higher in the forest during high flow than it was in cropland and riparian sites (Tab.3; Figs. 4h, S2l, h). This also explains the negative correlation between DOC and CO<sub>2</sub> observed only in forest groundwater (Fig. 6b). During base flow, we observed a clear land use spatial pattern among cropland, riparian forest, and forest sites (Tab. 3; Fig. S2h). We hypothesize that this difference was a consequence of a less intense soil respiration in croplands during summer. From simultaneous eddy covariance measurements over pine forests and maize croplands of the study area, Stella et al (2009) confirmed that ecosystem respiration was

lower in croplands than in forests over the whole year. However, Stella et al (2009) also showed that ecosystem respiration was larger during the growing season of the maize, because of increased of soil respiration in response to the higher soil water content caused by irrigation. In forest sites, groundwater pCO<sub>2</sub> increases during the summer because soil CO<sub>2</sub> diffuses downward and then is dissolved into the water table (Deirmendjian et al., 2018; Tsypin and Macpherson, 2012). A deeper water table in cropland suggests a less efficient CO<sub>2</sub> transfer from soil air to water table. Higher soil moisture in croplands due to irrigation probably delays soil CO<sub>2</sub> diffusion to groundwater. The  $\delta^{13}$ C-DIC signature of forest groundwater was typical of a signature that originated from respiration of soil organic matter derived from C<sub>3</sub> plants (O'Leary, 1988; Vogel et al., 1993). The studied forest soils have no natural carbonate minerals (Augusto et al., 2010) and DIC originating from silicate weathering has the same isotopic signature as DIC originating from soil respiration (Das et al., 2005; Polsenaere and Abril, 2012; Wachniew, 2006). Crop groundwater had a heavier  $\delta^{13}$ C-DIC signature than forest groundwater and this discrepancy resulted from distinct processes. Liming in cropland brings artificial carbonates into crop soil and DIC originating from carbonate weathering produced DIC with a  $\delta^{13}$ C value of approximately half of that of soil CO<sub>2</sub> as carbonate rocks have a  $\delta^{13}$ C of approximately 0 %, making  $\delta^{13}$ C-DIC less negative (Clark and Fritz, 1997; Salomons and Mook, 1986). Irrigation with extraction of groundwater could also increase the  $\delta^{13}$ C-DIC signature by enhancing the degassing rate of  $^{12}$ CO<sub>2</sub> relative to  $^{13}\text{CO}_2$  (Deirmendjian and Abril, 2018; Polsenaere and Abril, 2012). Changes in the  $\delta^{13}\text{C-DIC}$  signature could also originate from respiration of soil organic matter derived from maize, a C<sub>4</sub> plant with a heavier δ<sup>13</sup>C signature than C<sub>3</sub> forest plants (O'Leary, 1988; Vogel et al., 1993), as observed in the study region (Quénéa et al., 2006). Indeed, after three decades of cultivation, the remaining carbon from the forest pool was mostly recalcitrant and its degradation probably did not affect the  $\delta^{13}$ C-DIC pool (Jolivet et al., 1997).

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### 4.3. Dynamics of IN and CH<sub>4</sub> in groundwater: the influence of groundwater O<sub>2</sub>

Subsurface and groundwater redox zonation is driven by the spatial and temporal distribution of O<sub>2</sub> that serves as the primary terminal electron acceptor during the degradation of organic C. In crop groundwater, high O<sub>2</sub> concentrations inhibited methanogenesis, as this process is strictly anaerobic and thus resulted in very low CH<sub>4</sub> concentrations (Tab. 2; Figs. 4f, g; Borges et al., 2018; Ciais et al., 2010; Jurado et al., 2017; Klüber and Conrad, 1998). Conversely, forest and riparian groundwater exhibited anoxic conditions that allowed methanogenesis to occur and created higher CH<sub>4</sub> concentration in forest sites compared to cropland sites (Tab. 2; Figs. 4f, g). High water table stages in forested areas cause anoxia in soils, forcing plants and microorganisms to switch to anaerobic metabolism (Naumburg et al., 2005; Bakker et al., 2006, 2009). Thus, in riparian and forest areas, we expected a positive relationship between water table and groundwater CH<sub>4</sub> but, to the contrary, we observed a negative relationship between these two parameters  $(R^2 = 0.25, p < 0.05; data not shown)$ . This implies that methanogenesis primarily occurs in deeper layers of forest soils, especially in summer. Fe<sup>2+</sup> and NH<sub>4</sub><sup>+</sup> accumulates in forest groundwater because anoxic conditions inhibit nitrification and iron oxidation (Tab. 2; Figs. 4d, e; Jambert et al., 1994; Widdel et al., 1993). In groundwater, anoxic conditions enable heterotrophic denitrification, whereas an O<sub>2</sub> threshold of 30–60 umol L<sup>-1</sup> completely inhibits heterotrophic denitrification (Balestrini et al., 2016; Cey et al., 1999; Christensen et al., 2013; Jambert et al., 1994; Kolbjørn Jensen et al., 2017; Korom, 1992). In strictly forest sites, denitrification in groundwater is usually limited by the scarcity of NO<sub>3</sub>, whereas in strictly crop sites denitrification is often limited by organic C availability (Tab. 2; Jambert et al., 1994; Starr and Gillham, 1993). Thus, N fertilizer application associated with different groundwater denitrification rates in the different plots creates the observed spatial pattern of groundwater NO<sub>3</sub> concentration in crop, riparian and forest sites (Tab. 2; Fig. 4c). In local maize croplands, Jambert et al (1997) found that 13% of the N fertilizers inputs were converted to N<sub>2</sub> gas, demonstrating that denitrification could occur in these oxic crop soils and groundwater. Although oxic conditions are not favorable for groundwater denitrification, some studies in agricultural catchments do describe this process at relatively high O<sub>2</sub> (150 µmol L<sup>-1</sup>)

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levels (McAleer et al., 2017; Otero et al., 2009). In crop soils, Rubol et al. (2016) investigated the spatiotemporal dynamics in oxidative microbial activity and the development of anoxic micro zones (i.e., anoxic hot-spots) at the microscopic level (µm to cm). They found that labile C addition resulted in maximum rates of local metabolic activity within a few minutes and led to the subsequent formation of anoxic hotspots and thus, both oxic and anoxic conditions coexisted closely within a small volume of crop soils. Consequently, denitrification probably occurs in anoxic microsites in waterlogged soil during irrigation as higher soil moisture results in lower soil oxygen concentration, lower redox potential and higher leaching of soil DOC (Hagedorn et al., 2000; Jambert et al., 1997; Rubol et al., 2012; Silver et al., 1999). N fertilizer load in local croplands is 25 g N m<sup>-2</sup> yr<sup>-1</sup> (Jambert et al., 1997), whereas export (using drainage of 637 mm yr<sup>-1</sup> and the average NO<sub>3</sub> concentration in crop groundwater) of NO<sub>3</sub> through crop groundwater was estimated at 10 g N m<sup>-2</sup> yr<sup>-1</sup> (40% of the annual N fertilizer load), and export (using same drainage and the average NO<sub>3</sub> concentration in riparian groundwater) of NO<sub>3</sub> through riparian groundwater was estimated at 2.8 g N m<sup>-2</sup> yr<sup>-1</sup> (8% of the annual N fertilizer load). This shows the importance of riparian groundwater to attenuate N inputs from adjacent croplands to streams, otherwise a large portion of the annual N fertilizer load would have been leached into adjacent streams rather than being denitrified or used by plants. In riparian groundwater adjacent to a farm in the New York state (USA), Anderson et al. (2014) found that total groundwater denitrification was equivalent to 32% of manure N spread on the adjacent upland field. (Mekala et al., 2017) simulated the transport and dynamics of N in an agricultural soil under flooded conditions and concluded that relatively shallow aguifers with sandy soil are vulnerable to NO<sub>3</sub> contamination at around 10 days if continuous irrigation is practiced. They also stated that NO<sub>3</sub> had higher leaching potential than NH<sub>4</sub> or DOC. At our study site, irrigation and associated desorption of DOC and NO<sub>3</sub> could explain their slight increase in crop groundwater during base flow (Tab. 3, Figs. 1, S2c). In a storm infiltration basin in Florida (USA), O'Reilly et al. (2012) found that concomitant peaks in groundwater O<sub>2</sub> and NO<sub>3</sub> concentrations after storm rainfall were a consequence of organic N leaching, indicating that there were short periods of ammonification and

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nitrification. In crop groundwater of Wallonia (Belgium), when groundwater  $O_2$  levels are higher than 125  $\mu$ mol  $L^{-1}$  (as at the study site), nitrification rather than denitrification promotes the accumulation of  $N_2O$  in groundwater (Jurado et al., 2017).

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### 4.4. Stream biogeochemical functioning: mostly a function of groundwater composition

NO<sub>3</sub> inputs to streams cause stream eutrophication (Carpenter et al., 1998; Jordan and Weller, 1996; Smith, 2003; Zhou et al., 2017). This is consistent with our field observations where we observed that crop streams were highly vegetated with macrophytes during base flow stages. Compared to high flow conditions, crop stream eutrophication was accompanied by higher pH and  $\delta^{13}$ C-DIC, and lower pCO<sub>2</sub>caused by preferential <sup>12</sup>CO<sub>2</sub> uptake during the macrophyte plant photosynthesis (Tab. 4; Figs. S3a, h, j; De Carvalho et al., 2009; Raven et al., 2002). The development of macrophytes in crop streams modifies flow and can cause a significant drop in water velocity, which in turn, gives rise to extensive deposition and retention of sediment beneath the macrophytes (Cotton et al., 2006; Sand-Jensen and Pedersen, 1999). This leads to seasonal accumulation of organic matter, a predominance of anoxic conditions in stream sediments, and thus the occurrence of methanogenesis as evidenced by peaks in dissolved CH<sub>4</sub> during base flow (Tab. 4; Figs. S3g; Borges et al., 2018; Crawford et al., 2016; Sanders et al., 2007). Crop stream CH<sub>4</sub> concentration was 390 nmol L<sup>-1</sup> during base flow (Tab. 2), a concentration significantly lower (1,430 nmol L<sup>-1</sup>) than chalk streams impacted by macrophyte vegetation in England (Sanders et al., 2007). This discrepancy probably resulted from the increased in silt and clay fraction during summer of the underlying sediment in chalk streams (Sanders et al., 2007). This would suggest that the permeability of chalk stream sediment became lower than that of sandy stream sediment and created stronger reducing conditions in chalk stream sediments, which likely increased the potential for methanogenesis (Baker et al., 1999; Findlay, 1995; Kankaala et al., 2005; Morrice et al., 1997). Sanders et al. (2007) also showed that the chalk streams' emissions of CH<sub>4</sub> to the atmosphere were approximately 50 times lower than the CH<sub>4</sub> production in stream sediments, illustrating the high potential for CH<sub>4</sub> oxidation

in the water column of crop stream. During base flow, a second explanation for higher CH<sub>4</sub> (and NH<sub>4</sub><sup>+</sup>) concentrations in crop streams relative to forest streams could be differential hydrology. Drainage plot is a function of the water table height (hydraulic gradient, Darcy's law). During base flow, the water table in cropland was deeper than in forest (e.g., 4 m deeper in Sep. 2014; Fig. 2), and so during this period, potentially more forest groundwater was drained into crop streams. However, in forest streams we usually did not observe CH<sub>4</sub> (or NH<sub>4</sub><sup>+</sup>) concentrations as high as in crop streams (Tab. 2; Fig. 7d) indicating that CH<sub>4</sub> (or NH<sub>4</sub><sup>+</sup>) in crop streams primarily originated from crop stream sediments rather than from higher discharge of forest groundwater. In crop streams, CH<sub>4</sub> was correlated with NH<sub>4</sub><sup>+</sup> but not correlated with NO<sub>3</sub> or DOC (Figs. 6d, 7c, d). Such relationships were also observed in the Meuse river basin (Belgium) (Borges et al., 2018) and in a global meta-analysis of riverine CH<sub>4</sub> (Stanley et al., 2016). In contrast, this does not fit the conceptual model of Schade et al (2016) developed from data in New Hampshire streams, whereby the CH<sub>4</sub> was positively correlated with DOC, while negatively related to NO<sub>3</sub>. Sandy sediments of low order stream beds impacted by eutrophication are significant areas of NO<sub>3</sub> reduction over the spring and summer, lowering DOC and NO<sub>3</sub> concentrations in stream water (Tab. 4; Figs. S3c, 1; Böhlke et al., 2009; Mulholland et al., 2008; Sanders et al., 2007). Additionally, the decreased of stream velocity during base flow increased residence times of NO<sub>3</sub> in the hyporheic zone and the time for denitrification (Bardini et al., 2012). In a small stream dominated by maize cropland in the USA, Böhlke et al. (2009) demonstrated that denitrification mainly occurred in sediments and not in the water column since integrated rates of pore-water denitrification derived from <sup>15</sup>N tracer profiles within the hyporheic zone were similar to the reach-scale rates derived from measurements in the stream. In crop streams, a portion of the NO<sub>3</sub> variability between the two hydrological periods could also result from higher drainage of forest groundwater during base flow, which would dilute the NO<sub>3</sub> signal from crop groundwater. Considering the flat catchment topography, a minor portion of TSM and POC in streams originates from soil erosion and surface runoff. The most frequent effects of dredging on aquatic ecosystems are changes

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in the concentration of suspended solids, turbidity and light penetration (Lewis et al., 2001; Newell et al., 1998). Higher concentrations of POC (and TSM) observed in crop streams were also caused by macrophyte biomass developed in summer became a sediment trap. When stream discharge was sufficiently energetic, it re-suspended all the accumulated sediment and removed this litter. Moreover, we observed peaks of CH<sub>4</sub> and NH<sub>4</sub><sup>+</sup> in crop streams during high flow (Tab. 4; Figs. S3d, g), suggesting that dredging or streambed erosion of crop streams also release CH<sub>4</sub> and NH<sub>4</sub><sup>+</sup> from the sediment. In forest streams, we observed significantly lower concentrations of Fe<sup>2+</sup>, NH<sub>4</sub><sup>+</sup>, and CH<sub>4</sub> than in forest groundwater and significant negative linear relationships between O<sub>2</sub> on the one side and Fe<sup>2+</sup>, NH<sub>4</sub><sup>+</sup>, or CH<sub>4</sub> (Tab. 2; Figs. 4d, e, g, 7e, f). This suggests there were low O<sub>2</sub> concentration groundwater inputs with high concentrations of reduced compounds and that the stream water was gradually oxygenated, which induced Fe<sup>2+</sup> and CH<sub>4</sub> oxidations and nitrification. Mulholland et al. (2000) studied N cycling by adding <sup>15</sup>N-labeled NH<sub>4</sub><sup>+</sup> into a forest stream in eastern Tennessee (USA). They concluded that the residence time of NH<sub>4</sub><sup>+</sup> in the water column was low (5 min) and that nitrification was an important sink for NH<sub>4</sub><sup>+</sup>, accounting for 19% of total ammonium uptake. In forest streams, the NH<sub>4</sub><sup>+</sup> concentration was approximately 3 µmol L<sup>-1</sup> lower than in forest groundwater and thus did not explained the NO<sub>3</sub> increase of 50 µmol L<sup>-1</sup> (Tab. 2; Figs. 3c, d). Up to 76% of N exports from local forest are in organic forms but these N exports are very low (< 0.2 g N m<sup>-2</sup> yr<sup>-1</sup>; De Wit et al., 2005; Rimmelin, 1998; Vernier et al., 2003), so in-stream mineralization of organic N coupled to nitrification could not explain NO<sub>3</sub> concentrations in forest streams. Since the sampled forest streams are not strictly forested, NO<sub>3</sub> concentration are explained by simple hydrological mixing between crop and forest groundwater (Tab. 2). In streams, pCO<sub>2</sub> was lower and O<sub>2</sub> was higher than in groundwater (Tab. 2; Figs. 3f, h, 5a). This shows that gas exchange between stream water and the atmosphere occurs quickly, favored by low stream depth and strong concentration gradients between the two compartments. Some authors (e.g., Bodmer et al. 2016; Borges et al., 2018) found elevated pCO<sub>2</sub> in crop streams rather than in forest streams, due to higher levels of dissolved and particulate organic matter in crop dominated systems compared to the forested

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ones that facilitated the in-stream degradation of organic matter. Moreover, land uses are expected to change the composition of terrestrial soil organic matter leached to streams, shifting from vegetation- to microbe-derived organic matter with greater agricultural land use and potentially higher emissions in crop streams (Fuss et al., 2017; Graeber et al., 2015; Wilson and Xenopoulos, 2009). Those results contrasted with ours because we found no difference in pCO<sub>2</sub> between crop and forest streams. Forest groundwater did have higher pCO<sub>2</sub> than crop groundwater, indicating a more intense degassing in forest streams. The similar  $\delta^{13}$ C-DIC signatures in forest and crop streams despite the strong difference between crop and forest groundwater suggests faster isotopic equilibration of DIC resulting from degassing. The greater gas transfer velocity in forest streams is a consequence of the abundance of coarse woody debris which generates higher levels of water turbulence (e.g., Bodmer et al. 2016), and is consistent with our field observations. A lower gas transfer velocity lower in crop streams results from stream calibration reducing turbulent flow, and macrophyte vegetation that protects the water surface from wind shear.

#### 5. Conclusion

The present study demonstrates that C and IN concentrations in shallow groundwater and in first-order streams are strongly sensitive to land use. In sandy lowland catchments, simultaneous measurements of biogeochemical parameters in groundwater and streams are crucial for identifying and quantifying biogeochemical processes involved at the groundwater-stream interface. We also show that a statistical clustering analysis based on NO<sub>3</sub><sup>-</sup> dataset enables partitioning of groundwater and streams into cropaffected or forest-dominated waters. Such a classification could be useful to river managers and policy makers. The water table had greater depth in croplands and was a crucial parameter necessary for understanding groundwater biogeochemical variability in relation to land use. Higher water table stages in forests created anoxic conditions and increased soil leaching. Conversely, in croplands, the deeper water table prevented anoxic conditions, creating different groundwater compositions from forest groundwater and inhibiting the denitrification of the N fertilizers, which led to groundwater NO<sub>3</sub><sup>-</sup> accumulation. Despite

642 the occurrence of groundwater denitrification in riparian and forest sites, N fertilizers inputs in crop 643 streams were still high enough to generate eutrophic conditions in these streams. Eutrophication resulted 644 in a biogeochemical cascading effect, which sustained high CH<sub>4</sub> concentration and lowered NO<sub>3</sub>. High 645 CO<sub>2</sub> and CH<sub>4</sub> production occurs in forest soils and groundwater, but these two gases exhibit lower 646 concentrations in forest streams, indicating intense degassing or oxidation. 647 The groundwater-stream interface is a biogeochemical hotspot and hot moment for C emissions and N 648 removal processes (McClain et al., 2003). Future studies focusing on the groundwater-stream interface in 649 relation to land use are needed to better understand C and N dynamics in aquatic systems in order to 650 correctly close C and N budgets at regional and global scales. 651 652 Acknowledgments This research is part of the CNP-Leyre project funded by the Cluster of Excellence COTE at the 653 654 Université de Bordeaux (ANR-10-LABX-45). 655 656 **Bibliography** Abril, G., Borges, A.V., 2018. Carbon leaks from flooded land: do we need to re-plumb the inland water 657 658 active pipe? Biogeosciences Discuss. 2018, 1-46. https://doi.org/10.5194/bg-2018-239 659 Abril, G., Bouillon, S., Darchambeau, F., Teodoru, C.R., Marwick, T.R., Tamooh, F., Ochieng Omengo, F., Geeraert, N., Deirmendjian, L., Polsenaere, P., Borges, A.V., 2015. Technical Note: Large 660 661 overestimation of pCO2 calculated from pH and alkalinity in acidic, organic-rich freshwaters. Biogeosciences 12, 67-78. https://doi.org/10.5194/bg-12-67-2015 662 Abril, G., Martinez, J.-M., Artigas, L.F., Moreira-Turcq, P., Benedetti, M.F., Vidal, L., Meziane, T., Kim, J.-H., 663 Bernardes, M.C., Savoye, N., Deborde, J., Souza, E.L., Albéric, P., Landim de Souza, M.F., Roland, 664 665 F., 2014. Amazon River carbon dioxide outgassing fuelled by wetlands. Nature 505, 395–398. 666 https://doi.org/10.1038/nature12797 667 Aitkenhead, J.A., Hope, D., Billett, M.F., 1999. The relationship between dissolved organic carbon in stream water and soil organic carbon pools at different spatial scales. Hydrol. Process. 13, 1289-668

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Stream Order	Description	Catchment area (km²) <sup>a</sup>	Crop (%) <sup>b</sup>	Forest (%) <sup>b</sup>	Urban (%) <sup>b</sup>	During High Flow <sup>c</sup>	During Base Flow <sup>c</sup>
1	Ditch	1.0	86.5	13.5	0.0	C	С
1	Ditch	1.3	53.8	46.2	0.0	C	C
1	Ditch	11.3	44.2	55.8	0.0	C	C
1	Ditch	13.4	42.5	57.5	0.0	C	C
1	Stream	57.0	30.7	69.3	0.0	C	C
1	Stream	16.8	7.8	92.2	0.0	F	F
1	Ditch	7.9	5.8	94.2	0.0	F	F
1	Ditch	2.3	5.2	94.8	0.0	F	C
1	Stream	16.0	4.6	93.8	1.6	C	F
1	Stream	34.0	3.8	96.2	0.0	F	F
1	Stream	31.0	2.3	97.7	0.0	F	F
1	Headwater	0.3	0.0	100.0	0.0	F	F
0	Groundwater in a riparian forest but very near (5m) a maize cropland (P1)					R	R
0	Groundwater in maize cropland (P2)					C	C
0	Groundwater in maize cropland (P3)					C	C
0	Groundwater in pine forest (P4)					F	F
0	Groundwater in pine forest (P5)					F	F

Table 1. Characteristics of groundwater and stream sampling stations, ranked in decreasing order of cropland percentage in their respective subcatchments. <sup>a</sup> delimited with a geographic information system software (ArcGIS 10.5®) using an hydrological database in a polyline form (BD CARTHAGE®) and a digital elevation model (BD ALTI®, resolution of 25m), which both have been made available by the national geographic institute of France (http://www.ign.fr/). <sup>b</sup> retrieved with the CORINE land cover 2006 database (EEA, 2014) using a geographic information system software (ArcGIS 10.5®). <sup>c</sup> C, F, R corresponding to crop, forest and riparian waters, respectively, either during high or base flow. Piezometer 1 (P1) is located in a riparian mixed pine and oak forest near a first-order stream and near a maize cropland, which where P2 is located. P2 and P3 are located in the middle of two different maize croplands of 0.6 km² and 6 km², respectively. P5 is located in an 11-years old pine plot of 0.6 km² and is part of the ICOS (name is FR-Bil) research infrastructure (http://icos-ri.eu), whereas P4 is located in another pine forest (approximately same age as P5 pine forest). The depth of piezometers (from the soil surface to the bottom of the piezometer) is 5.3m for P1, 4.9m for P2, 9.1m for P3, 5m for P4 and P5.

	Crop continuum		Forest cont	Riparian groundwater	
	Groundwaters (22)	Streams (59)	Groundwaters (22)	Streams (78)	Groundwaters (11)
рН	4.5±0.2	6.0±0.3	4.5±0.3	5.8±0.5	4.7±0.1
_	4.3-5.0	5.4-7.0	3.7-4.8	4.2-6.9	4.4-4.8
Temperature (°C)	14.5±1.8	13.6±4.2	12.8±1.8	12.9±3.9	14.9±2.4
•	10.7-17.5	6.4-25.8	8.5-15.1	4.8-22.1	11.8-17.9
EC (µS cm <sup>-1</sup> )	360±70	220±55	90±10	115±30	160±50
•	220-465	75-370	65-115	70-200	95-270
$NO_3^-$ (µmol $L^{-1}$ )	$1,140\pm485$	340±200	25±40	75±70	310±260
	260-1,785	10-950	0-120	0-275	40-860
$NH_4^+$ (µmol $L^{-1}$ )	$0.4\pm0.8$	6.1±7.0	4.5±7.0	1.8±1.7	$0.4\pm0.4$
,	0-3.5	0-40	0.3-30	0-7.8	0-1.5
$Fe^{2+}$ (µmol L <sup>-1</sup> )	$0.9\pm0.4$	$5.9 \pm 4.4$	15±15	$7.9 \pm 12.0$	$0.6\pm0.5$
•	0.1-1.9	0.1-22	$0.9\pm56$	$0.6\pm58$	0.2-1.5
$O_2$ (µmol $L^{-1}$ )	220±65	290+45	20±30	280±50	100±70
- ,	100-315	160-400	0-110	110-370	0-170
$CH_4$ (nmol $L^{-1}$ )	40±25	460±950	$1,770\pm1,830$	240±300	1,470±1,490
. ,	15-130	20-4,900	50-6,700	20-2,370	30-4,150
pCO <sub>2</sub> (ppmv)	30,650±11,590	$4,480\pm2,680$	50,630±26,070	$4,900\pm4,500$	42,950±28,560
	19,000-60,550	1,040-14,080	7,680-116,380	1,000-27,200	17,300-103,300
$TA (\mu mol L^{-1})$	90±25	100±50	70±30	90±50	70±15
•	35-130	30-300	30-135	30-280	45-85
$\delta^{13}$ C-DIC (‰)	-19.8±1.3	-18.2±3.5	-26.7±1.0	-19.8±2.8	-25.2±1.1
, ,	-22 to -17.6	-27.6 to -11.3	-28.8 to -24	-27.6 to -14	-27.9 to -23.4
DIC (µmol L <sup>-1</sup> )	1,450±480	315±135	$2,460\pm1,130$	320±210	1,960±1,150
,	820-2,590	90-650	570-5,370	120-1,280	940-4,480
DOC (µmol L <sup>-1</sup> )	510±150	605±320	930±930	470±250	400±100
, ,	275-880	220-2,290	310-3,670	190-1,725	280-620
$TSM (mg L^{-1})$		5.6±8.6		2.3±1.7	
\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \		0.1-50.5		0.4-8.2	
POC (%)		28±10		28±10	
` '		0-50		10-80	
POC (µmol L <sup>-1</sup> )		120±180		50±35	
4 /		0-1,100		0-170	

Tab. 2. Values of carbon and ancillary parameters throughout sampling period (Jan. 2014-Jul. 2015) in crop and forest continuums and in riparian groundwater. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed average±standard deviations and the range.

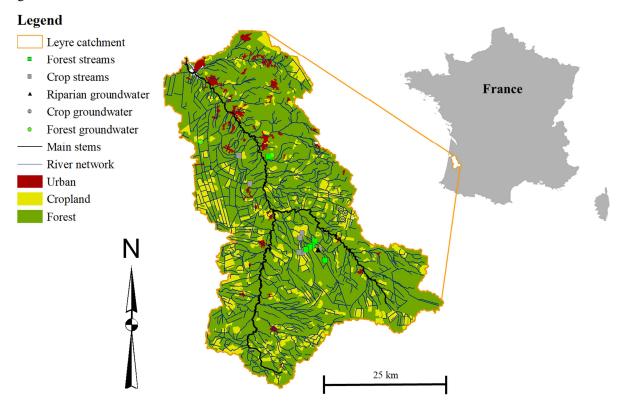
				Groundwater		
	Cropland_HF (4)	Cropland_BF (18)	Forest_HF (6)	Forest_BF (16)	Riparian_HF (2)	Riparian_BF (9)
pН	4.6±0.3	4.5±0.2	4.4±0.3	4.5±0.3	4.7±0.1	4.6±0
	4.3-4.9	4.3-5.0	4.0-4.8	3.7-4.8	4.6-4.8	4.4-4.
Temperature (°C)	12.8±1.7	14.9±1.6	$10.8 \pm 1.4$	13.5±1.4	12.2±0.6	15.6±2
	10.7-14.5	11.6-17.5	8.5-12.2	10.7-15.1	11.8-12.6	12.1-17
EC (μS cm <sup>-1</sup> )	370±60	360±70	90±15	90±10	200±20	150±5
	320-460	220-470	70-115	70-115	185-215	95-27
$NO_3^-$ (µmol L <sup>-1</sup> )	$1,040\pm300$	$1,160\pm420$	30±50	20±40	510±20	260±27
	760-1,320	260-1785	0-120	0-120	500-520	40-86
$NH_4^+$ (µmol L <sup>-1</sup> )	$0.5\pm0.4$	$0.4\pm0.9$	$3.3\pm2.2$	$5.0\pm8.0$	$0.3\pm0.1$	$0.4\pm0$
•	0.1-1	0-3.5	1.1-7	0-3-30	0.2-0.3	0-1
$Fe^{2+}$ (µmol L <sup>-1</sup> )	$0.8\pm0.2$	$0.9\pm0.4$	$10.0\pm8.2$	15±15	$0.4\pm0.2$	$0.7\pm0$
•	0.7-1.1	0.1-1.9	2.7-25.5	0.9-56.6	0.2-0.5	0.2-1
$O_2 (\mu \text{mol } L^{-1})$	250±90	220±70	20±20	20±30	170±0	100±8
	180-310	100-320	0-40	0-110	170-170	0-20
CH <sub>4</sub> (nmol L <sup>-1</sup> )	30±3	50±25	480±630	$2,260\pm1,900$	$1,460\pm2,010$	1,470±1,50
	25-30	16-130	50-1,700	50-6,700	40-2,880	30-4,13
pCO <sub>2</sub> (ppmv)	$22,050\pm2,000$	32,560±12,000	$28,100\pm11,580$	$59,080\pm25,060$	21,530±5,950	47,700±29,59
	19,800-24,270	19,000-60,550	7,680-39,000	29,685-116,400	17,320-25,740	20,600-103,30
TA (µmol L <sup>-1</sup> )	85±2	92±30	95±40	65±30	83±2	60-
	82-86	35-130	60-135	30-100	82-85	45-
$\delta^{13}$ C-DIC (‰)	-20.7±1.1	-19.6±1.3	-26.6±1.3	$-26.8\pm1.0$	-26.9±1.4	-24.9±0
` '	-22 to -19.7	-21.9 to -17.6	-27.6 to -24.0	-28.8 to -25.3	-27.9 to -25.9	-25.7 to -23
DIC (µmol L <sup>-1</sup> )	$1,100\pm180$	$1,520\pm490$	$1,500\pm550$	$2,830\pm1,080$	1,160±315	$2,140\pm1,2$
<b>'</b>	930-1,300	820-2,590	570-2,040	1,650-5,370	940-1,380	1,020-4,4
DOC (µmol L <sup>-1</sup> )	420±120	550±140	$2,230\pm1,440$	740±380	310±50	420±1
٧ /	320-590	340-880	575-3,670	310-1,720	275-350	310-6

Tab. 3. Values of carbon and ancillary parameters throughout sampling period (Jan. 2014-Jul. 2015) in different types of groundwater across hydrological seasons. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed the average±standard deviations and the range. We defined six groups that are Cropland\_HF/Cropland\_BF, Forest\_HF/Forest\_BF and Riparian\_HF/Riparian\_BF corresponding to groundwaters during high flow (HF) or base flow (BF); in either cropland, forest or riparian forest.

	First-order streams					
	Cropland_HF (22)	Cropland_BF (37)	Forest_HF (23)	Forest_BF (55)		
рН	5.9±0.3	6.1±0.4	5.7±0.6	6.1±0.4		
	5.4-6.6	5.5-7.0	4.2-6.8	5.0-6.9		
Temperature (°C)	10.2±1.6	15.7±3.9	9.0±1.9	14.6±3.3		
_	6.4-12.1	9.1-25.8	4.8-12	8.1-22.1		
EC (μS cm <sup>-1</sup> )	230±50	220±60	110±20	120±30		
	145-340	75-370	80-150	70-200		
$NO_3^-$ (µmol $L^{-1}$ )	420±220	290±170	95±70	65±70		
	180-950	8.5-705	0-275	0-275		
$NH_4^+$ (µmol $L^{-1}$ )	$7.0\pm 8.4$	5.5±6.0	1.7±1.7	1.7±1.7		
•	0.5-38.7	0-25.3	0.3-7.8	0-6.9		
$Fe^{2+}$ (µmol $L^{-1}$ )	$6.7 \pm 3.8$	5.4±4.7	5.7±3.0	8.8±14.0		
	1.6-15.7	0.1-22	2.6-13.6	0.6-57.1		
$O_2$ (µmol $L^{-1}$ )	290±50	290±40	300±40	270±60		
	190-400	160-370	210-370	110-360		
CH <sub>4</sub> (nmol L <sup>-1</sup> )	580±1,080	390±880	185±190	270±340		
	30-4,380	20-4,900	40-980	20-2,370		
pCO <sub>2</sub> (ppmv)	5,200±2,370	$4,040\pm2,790$	4,200±2,430	5,200±5,100		
	1,040-10,740	1,220-14,080	1,240±11,690	1,010-27,20		
TA (μmol L <sup>-1</sup> )	105±50	100±50	70±40	95±55		
	40-300	30-255	35-195	30-280		
δ <sup>13</sup> C-DIC (‰)	-20.6±3.9	-16.8±2.4	-22.1±2.5	-18.9±2.3		
	-27.6 to -11.3	-22.3 to -12.4	-27.6 to -16.8	-23.1 to -14.0		
DIC (µmol L <sup>-1</sup> )	380±130	280±120	300±150	330±230		
	1,000-650	90-600	150-750	120-1,280		
DOC (µmol L <sup>-1</sup> )	750±400	520±230	540±305	450±220		
	300-2,290	220-1,520	260-1725	190-154		
TSM (mg L <sup>-1</sup> )	9.3±11.5	3.1±4.9	$2.8 \pm 1.7$	2.1±1.		
	0.9-51	0.1-27	0.5-6.6	0.4-8.		
POC (%)	26±10	30±10	29±8	29±1		
	15-48	16-48	20-50	12-8		
POC (µmol L <sup>-1</sup> )	190±250	65±100	65±40	40±30		
	0-1100	0.3-540	0-170	0.5-140		

Tab. 4. Values of carbon and ancillary parameters throughout sampling period (Jan. 2014-Jul. 2015) in different types of streams across hydrological seasons. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed the average±standard deviations and the range. We defined four groups that are Cropland\_HF/Cropland\_BF, Forest\_HF/Forest\_BF corresponding to streams during high flow (HF) or base flow (BF); in either cropland-affected or forest-dominated land use.

Figure 1: Land use map of the Leyre catchment showing river network and the sampling locations of groundwaters and streams.



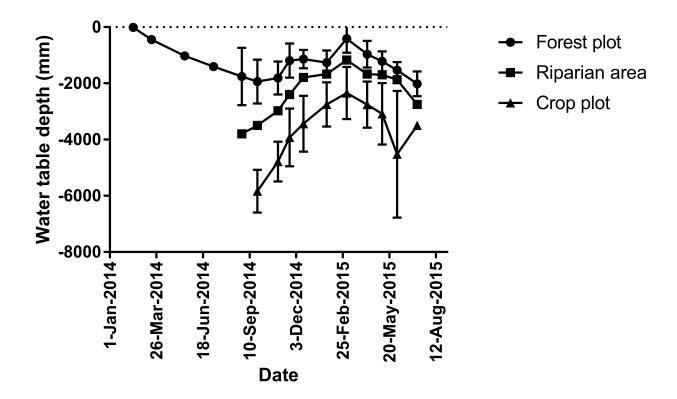


Figure 2: Water table depth during the sampling period (Jan. 2014-Jul. 2015) across land use in the Leyre catchment. The water table in riparian area is the water table at P1 (Tab. 1). The water table in crop plot is the average±standard deviations of water tables at P2 and P3 (Tab. 1). The water table in forest plot is the average±standard deviations of water tables at P4 and P5 (Tab. 1).

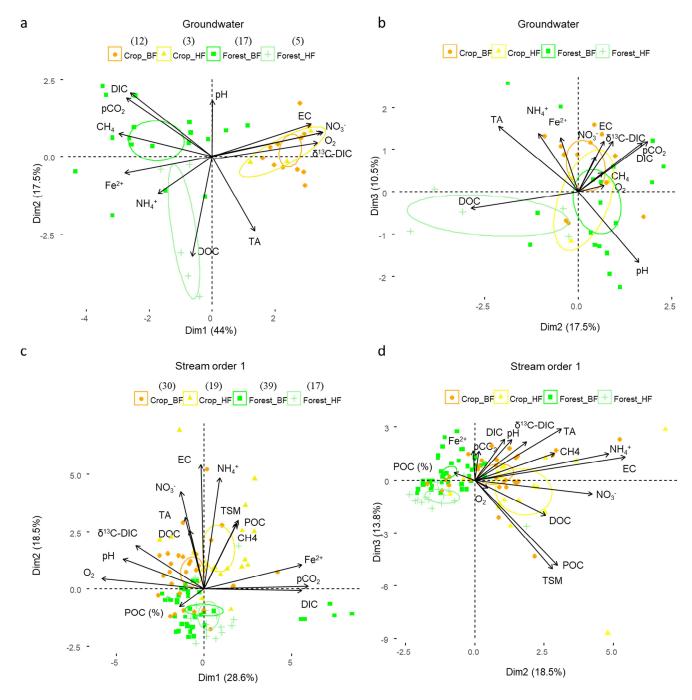


Figure 3: Principal component analysis (PCA) of shallow groundwater dataset (a-b) and first-order streams dataset (c-d). We represented only the first three dimensions. Numbers between brackets are corresponding to the sampling size. The sampling size in the PCAs did not correspond exactly to the sampling size in Tables 3 and 4 because R software deletes stations from the analysis with a missing value for one parameter. In these PCAs, we used all the quantitative variables measured in this study. In each PCA, we plotted as well the individuals separated in four groups. The first group corresponds to cropland-affected samples during high flow (Crop\_HF), the second group corresponds to cropland-affected samples during base flow (Crop\_BF), the third group corresponds to forest-dominated samples during base flow (Forest\_BF).

The mean value of each qualitative group has 95% chance to be within the corresponding confidence ellipse.

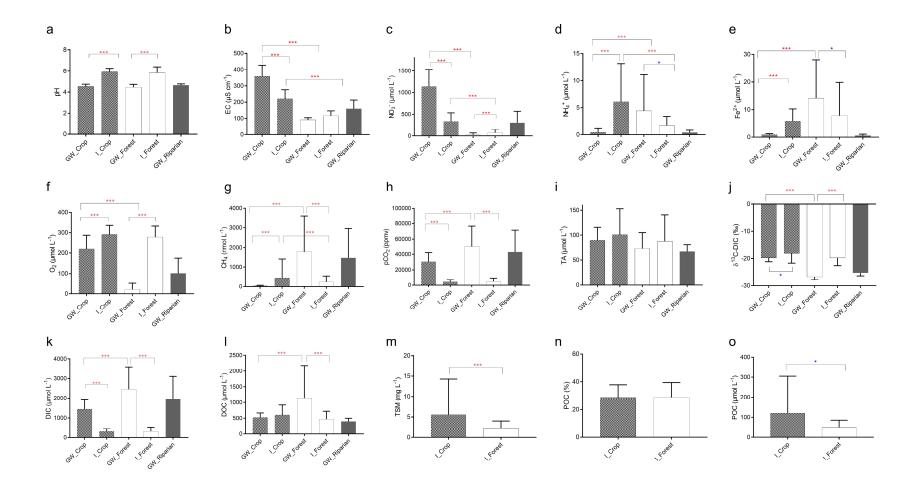


Figure 4: Values of carbon and ancillary parameters throughout the sampling period (Jan. 2014-Jul. 2015) in groundwater and streams across land use. Histograms represent the mean with standard deviations of a given parameter. We defined four groups that are GW\_Forest/GW\_Crop and I\_Forest/I\_crop corresponding to groundwaters and streams order 1 either dominated by forests or croplands. A fifth group is GW\_Riparian and corresponding to riparian groundwater. Then, based on Mann-Whitney statistical analysis, we compared GW\_Crop VS I\_Crop, GW\_Forest VS I\_Forest, GW\_Crop VS GW\_Forest, I\_Crop VS I\_Forest. Three red stars (\*\*\*) indicate that data were significantly different with p < 0.001. One blue star (\*) indicates that data were significantly different with p < 0.05).

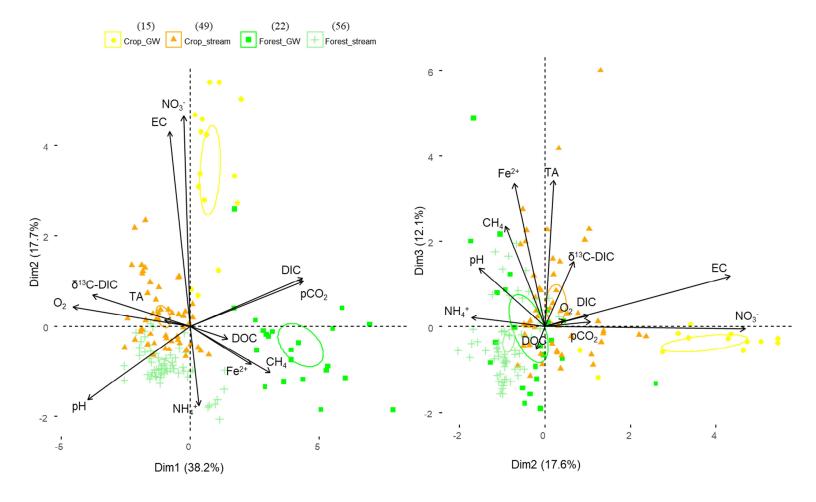


Figure 5: Principal component analysis (PCA) of shallow groundwater and stream datasets. We represented only the first three dimensions. Numbers between brackets are corresponding to the sampling size. The sampling size in the PCA did not correspond exactly to the sampling size in Table 2 because R software deletes stations from the analysis with a missing value for one parameter. In these multivariate statistical analyses, we used all the quantitative variables measured in this study. We defined four groups that are Crop\_GW/Forest\_GW and Crop\_stream/Forest\_stream, which are corresponding to groundwater and streams order 1, either dominated by forests or croplands. The mean value of each qualitative group has 95% chance to be within the corresponding confidence ellipse

Figure 6: Scatter plots of (a)  $CO_2$  (ppmv) vs.  $O_2$  ( $\mu$ mol  $L^{-1}$ ), (b)  $CO_2$  (ppmv) vs. DOC ( $\mu$ mol  $L^{-1}$ ), (c)  $CH_4$ (nmol  $L^{-1}$ ) vs.  $O_2$  ( $\mu$ mol  $L^{-1}$ ), (d)  $CH_4$  vs. DOC ( $\mu$ mol  $L^{-1}$ ), (e)  $CH_4$ (nmol  $L^{-1}$ ) vs.  $CO_2$  (ppmv), and (f)  $O_2$  ( $\mu$ mol  $L^{-1}$ ) vs. DOC ( $\mu$ mol  $L^{-1}$ ), in all sampled groundwater and streams

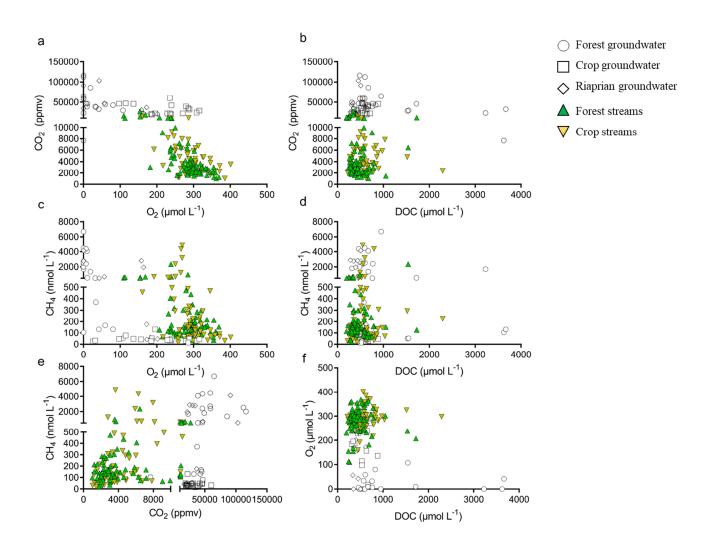
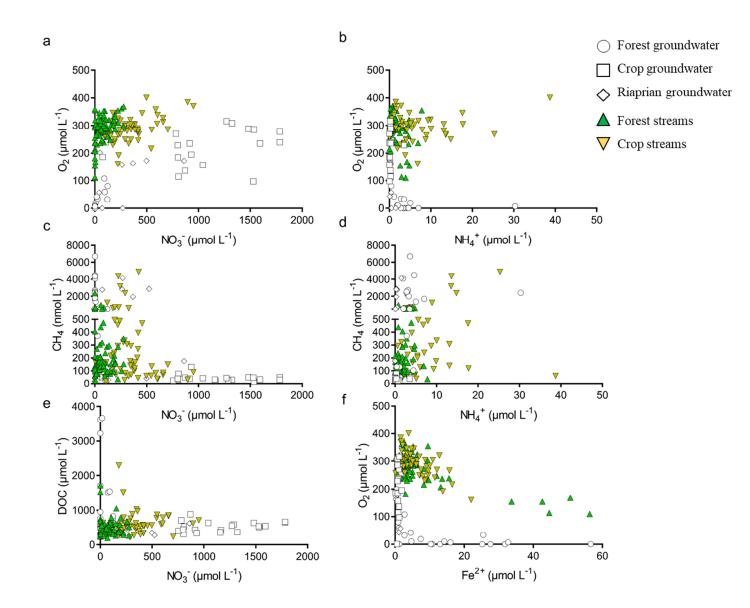


Figure 7: Scatter plots of (a)  $O_2(\mu mol\ L^{-1})$  vs.  $NO_3^-(\mu mol\ L^{-1})$ , (b)  $O_2(\mu mol\ L^{-1})$  vs.  $NH_4^+(\mu mol\ L^{-1})$ , (c)  $CH_4(\mu mol\ L^{-1})$  vs.  $NO_3^-(\mu mol\ L^{-1})$ , (d)  $CH_4(\mu mol\ L^{-1})$  vs.  $DOC(\mu mol\ L^{-1})$ , (e)  $DOC(\mu mol\ L^{-1})$  vs.  $NO_3^-(\mu mol\ L^{-1})$ , and (f)  $O_2(\mu mol\ L^{-1})$  vs.  $Fe^{2+}(\mu mol\ L^{-1})$ , in all sampled groundwater and streams.



## Supplementary Material

Seasons	Date	(	Groundwater	Streams	
		P5	P1, P2, P3	P4	
HF	29/01/2014				X
HF	12/02/2014	X			
HF	07/03/2014				X
HF	17/03/2014	X			
BF	24/04/2014				X
BF	16/05/2014	X			
BF	21/05/2014				X
BF	25/06/2014				X
BF	17/072014	X			X
BF	27/08/2014	X	X		X
BF	24/09/2014	X	X		X
BF	31/10/2014	X	X		X
BF	21/11/2014	X	X		X
BF	16/12/2014	X	X		X
HF	27/01/2015	X	X	X	X
HF	04/03/2015	X	X	X	X
BF	10/04/2015	X	X	X	X
BF	07/05/2015	X	X	X	X
BF	03/06/2015	X	X	X	X
BF	09/07/2015	X	X	X	X

Table S1: Sampling dates of groundwaters and streams. X corresponds to a sampling. HF and BF are corresponding to high flow and base flow periods, respectively.

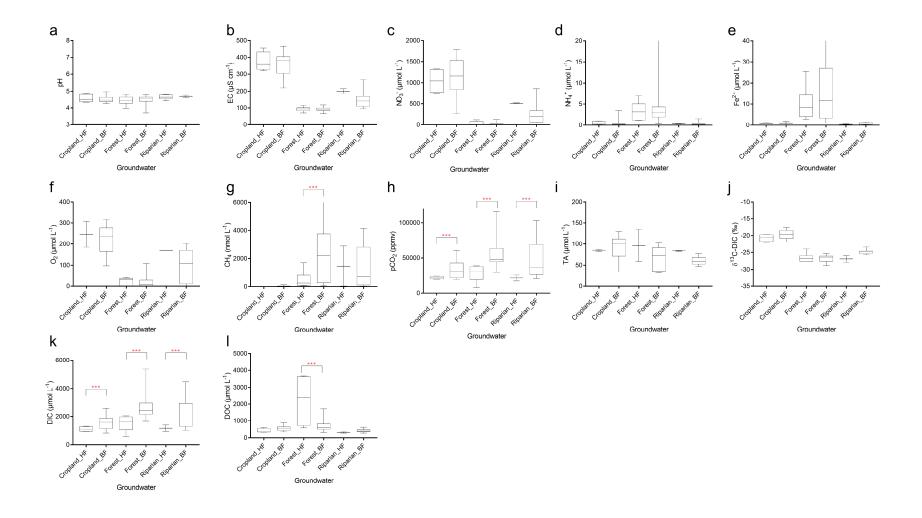


Fig. S2. Values of carbon and ancillary parameters throughout the sampling period (Jan. 2014-Jul. 2015) in groundwaters across hydrological seasons and land use. Box-plots represent the median (black bar) and the extreme (min-max) values. We defined six groups that are Cropland\_HF/Cropland\_BF, Forest\_HF/Forest\_BF and Riparian\_HF/Riparian\_BF corresponding to groundwater during high or base flow; in either cropland, forest or riparian forest. Then, based on Mann-Whitney statistical analysis, we compared Cropland\_HF vs. Cropland\_BF,

Forest\_HF vs. Forest\_BF and Riparian\_HF vs. Riparian\_BF. Three red stars (\*\*\*) indicate that data were significantly different with p<0.001. One blue star (\*) indicates that data were significantly different with p<0.05. No stars indicate that data were not significantly different (p>0.05)

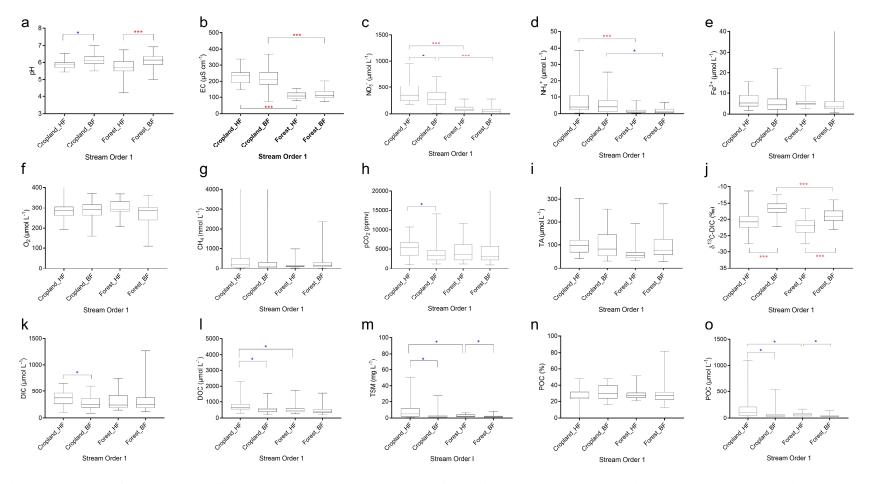


Figure S3. Values of carbon and ancillary parameters throughout the sampling period (Jan. 2014-Jul. 2015) in streams across hydrological seasons and land use Box-plots represent the median (black bar) and the extreme (min-max) values. We defined four groups that are Cropland\_HF/Cropland\_BF, Forest\_HF/Forest\_BF corresponding to streams during high or base flow periods; in either cropland-affected or forest-dominated land use. Then, based on Mann-Whitney statistical analysis, we compared Cropland\_HF vs. Cropland\_BF, Forest\_HF vs. Forest\_BF, Cropland\_BF vs. Forest\_HF and Cropland\_BF vs. Forest\_BF. Three red stars (\*\*\*) indicate that data were significantly different with p<0.001. One blue star (\*) indicates that data were significantly different with p<0.05)

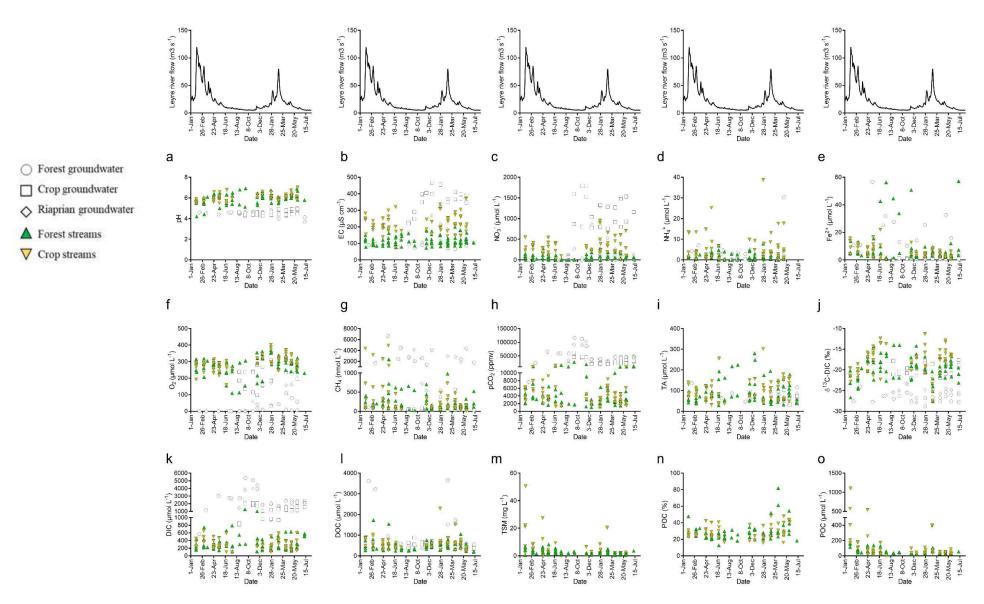


Figure S4: Temporal variations of carbon and ancillary parameters throughout the sampling period (Jan. 2014-Jul. 2015) in all sampled

groundwater and streams. Top panels represent the Leyre River flow (main stem).

