

Renal, Hepatic and Splenic Biotoxicity of Cadmium Sulphate in the Wistar Rats

Vandjiguiba Diaby, Francis Adou Yapo, Adon Arsène Mousan, Mireille Dosso, Joseph Allico Djama

▶ To cite this version:

Vandjiguiba Diaby, Francis Adou Yapo, Adon Arsène Mousan, Mireille Dosso, Joseph Allico Djama. Renal, Hepatic and Splenic Biotoxicity of Cadmium Sulphate in the Wistar Rats. International Journal of Environmental Science and Toxicology Research, 2016, 4 (6), pp.103-110. hal-02092756

HAL Id: hal-02092756

https://hal.science/hal-02092756

Submitted on 8 Apr 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Full Length Research Paper

Renal, Hepatic and Splenic Biotoxicity of Cadmium Sulphate in the Wistar Rats

DIABY Vandjiguiba ^{1,2*}, YAPO Adou Francis², ADON Arsène Mousan¹, DOSSO Mireille^{1,3}, DJAMA Allico Joseph^{2,3}

¹Cellular unit of biology, Institute Pasteur de Cote d'Ivoire, 01 BP 490 Abidjan 01, Côte.d'ivoire ²Laboratory of Pharmacodynamics-Biochemical, UFR Biosciences, University of FELIX Houphouet Boigny-Abidjan (Côte.d'ivoire), 22 BP 582 Abidjan 22.

Received 8 July, 2016; Accepted 18 July, 2016

Heavy metals (lead, mercury and cadmium) cause problems of public health. Studies undertaken in Côte d'Ivoire within the framework of the environmental monitoring made it possible to apprehend the state of the cadmium pollution of the sediments, of the market garden grounds. Six (6) batches of rats Wistar males and females were made up. These rats were contaminated with cadmium sulfate by daily during 30 days with various proportion respectively 4; 5; 6.66; 10 and 20 mg/kg body weight (bw). The rats were weighed each week. At the end of the experiment, the serum was used for proportioning of the enzymes and of the substrates using *Cobas C311 ROCHE HITACHI*. Work completed made it possible to determine the LD₅₀ of cadmium sulfate which is of 200 mg/kg bw. The weight of the rats of the batches treated with cadmium decreased significantly compared to the batch untreated (control). Transaminases, creatinine and the urea were increased significantly (p<0.05) when the bilirubin and cholesterol were decreased at the two sexes. However, the effect on cholesterol is not significant. Cadmium caused liver and kidney dysfunction and it decreased the weight of rats of both sexes.

Keywords: Cadmium, Biochemical markers of kidneys, Biochemical markers of liver and spleen.

INTRODUCTION

Heavy metals (lead, mercury, arsenic, cadmium...) are pollutants generated for most of the time by the human activity. They have a high toxicological impact on the plants, the products for current human consumption and on man (Amir *et al.*, 2014).

These heavy metals remain a serious problem of environment more and more worrying especially because of the affections noted on the populations which are exposed (Oumar *et al.*, 2014). With regard to cadmium, it is present in a significant way in certain food, like the seafood, meat offal, certain cereals (rice, corn...), mushrooms, vegetables and to a lesser extent,

in fish, the fruits and meat (Pascal et al., 2010).

In Côte d'Ivoire, the Cadmium concentrations are 5 to 127 times more significant in the discharges than in the grounds according to Traoré et al. (2014). Environmental pollution with cadmium relates to especially the rubbish dump garbage of Akouedo (Abidjan, Côte d'Ivoire) where place of many studies had. Indeed, this discharge of Akouedo is known for its heavy metal threat in particular with cadmium. It is classified with the row of the wild discharges whose natural drainage is done towards the lagoon Ebrié (bay of Me Badon), to less than 2.1 km. This liquid rejection downstream from the discharge generates a significant flow (474 m3 /days). Indeed, the lixiviat resulting from waste drains heavy metals (cadmium) and constitutes a potential source of underground pollution waters and surface (Adjiriet et al., 2008). The populations generally

³Department of Medical and Fundamental Biochemistry Institute Pasteur de Cote d'Ivoire, 01 BP 490 Abidjan 01.

^{*}Corresponding Author Email: vandjiguiba@yahoo.fr

complain about the bad smells, of diseases (respiratory, digestive, cutaneous) and the ground of the discharge contains a high heavy metal concentration (9.87 cadmium ppm) (Adjiriet et al., 2008). In addition, metals and metalloids associated with the aqueous phase with the grounds can be transported through Unsaturated Zone (ZNS) either towards the plants, or towards the underground sheet of water (Adjiriet et al., 2008). It is thus to fear as for the exposure of the populations to the consumption of water of well, of the market-gardening and food products cultivated on the site of the discharge. For the simple reason that this accumulation of heavy metals in the environment can be reflected on the health of the human beings and the animals (Yao and Kouassi, 2015). Several studies undertaken on the mammals showed that it causes genetic damage, the change of genes, the break of the ADN (IARC, 1993). Different experimental studies also showed that cadmium causes testicular attacks and decreases the fertility in males (Pascal et al., 2010).

This is the reason why our study is focused on cadmium sulfate biotoxicity in Wistar rats.

MATERIAL AND METHODS

Material

Biological material

The biological material is the serum of rats male and female of Wistar weighing $106 \pm 6g$ and old from 8 to 12 weeks. The rats were placed in cages furnished with litters of shaving of which revival was done each week. They were acclimatized before the beginning of handling.

Chemical Products

The cadmium sulfate used for the tests was dehydrated salt (MERCK), and of serial number 1.02027.0100.

Methods

Preparation of the cadmium sulfate solution

The rats were weighed by batch and the average (P) for each batch was thus determined. For the preparation of the cadmium sulfate solution, the formula below was used (Michel, 2011):

V: Volume to be managed or with gaver (ml)

D: Proportion to manage (mg/Kg body weight or 0. 001 mg/g)

C: Concentration of the solution (mg/ml)

The quantity of cadmium sulfate to be weighed was thus given. This quantity was deposited in a tube falcon and was supplemented to 10 ml with water distilled to constitute the cadmium sulfate solution. For an amount of 300 mg/Kg or 0.3 mg/g of body weight and for an average weight of 78g,

$$C = \frac{0.3\text{mg/g x 78g}}{1\text{ml}} = 23.4 \text{ mg/ml i.e. } 23.4 \text{ powder mg of}$$

$$\frac{1}{1}$$

$$\frac{1}{1}$$

$$\frac{1}{1}$$

Thus, for a final cadmium sulfate solution of 10 ml, it will be necessary to weigh 234 mg.

Study of acute toxicity by method 423 OECD

Method 423 OECD allowed to determine the lethal amount 50% (DL50) of cadmium sulfate managed by way oral examination with the rats.

Study of the subacute toxicity of cadmium sulfate

Identification of the animals

Each batch consists of the same animal's sex and appreciably equal weights. The animals were marked with the tail by batch and inside each batch in order to identify them during the experimentation.

Methodologies of cramming

Twelve (12) Batches of five (5) rats were made up including 6 male groups and 6 groups of females.

The–Groups 1 of males and females constituted the control groups where the animals received distilled water (1ml/day). Groups 2, 3, 4, 5 and 6 of each sex received respectively 1/50th, 1/40th, 1/30th, 1/20th and 1/10th of the DL50 of the cadmium sulfate solution. They are the treated groups.

The duration of treatment of the rats with cadmium sulfate was 30 Days (Layachi, 2012) and managed volume was of 1ml per day each morning throughout treatment.

Weighing of the rats, taking away of blood and experimental techniques

The rats were weighed the first day (D0) of the experimentation before the force-feeding and were weighed each week (D7, D14, D21, D30). For each batch and each week of weighing, the percentage P (see formula below) is calculated where Po represents the weight at the day 0 (D0) of the treatment and Pn weight

After the period of exposure (30 Days) to cadmium sulfate, the rats were sacrificed. The blood of each rat was collected immediately in the sample dry tube which was used to obtain the serum after centrifugation with 3000 rpm during 5min for the biochemical parameters with the automat *Cobas C311 ROCHE HITACHI* at the Pasteur Institute of Abidjan (Côte d' Ivoire).

Statistical analyses of the data

Statistical analysis and layout of the curves were undertaken with *GraphPad Prism V5.01* software (Washington, USA). Groups of data were compared oneway analysis of variance (ANOVA). Due to the small population size, the non-parametric Dunnett test was performed to compare assess difference between the control group and the other groups. Differences were considered statistically significant at p < 0.05.

RESULTS

Determination of the DL50 by Method 423 OECD

According to the system of classification harmonized overall (SGH), cadmium is classified of category 3 whose DL50 is located between 50 and 300 mg/kg body weight (bw). Thus, the limiting DL50 is fixed at 200 mg/kg bw.

Subacute Toxicity of Cadmium Sulfate

Symptomatology, Evaluation of Mortality and Determination of Maximum Tolerated Dose

The behavior of the rats presented the following characters: loss of hair as of the first week, calm, deceleration of the more visible growth for the rats exposed to 10 mg/Kg bw of cadmium and those exposed 20 mg/Kg in the males as in the females. The animals treated with cadmium ate and drank little proportionally with the amount of exposure. The animals did not make a hematuria but they urinated little. The control animals (Male and Female) ate and drank abundantly.

In the females, no mortality was noted. On the other hand at the males, in batch 6 (20 mg/Kg bw), two mortalities were noted. Therefore, the maximum tolerated dose to the males was 10 mg/Kg bw and for the females, it was 20 mg/Kg bw, according to the method of Galtier, (1974) modified.

Effect of Cadmium on the Body Weight of the Rats

Generally, the cadmium sulfate reduced the body weight of the males rats tested compared to the control males rats that weighed D7 = $115.244 \pm 4.96g$; D14 = 130.598 \pm 8.27g; D21 = 142.088 \pm 12.11g; D30 = 148.03 \pm 11.93g. Thus, according to figure 1, this reduction was significant for the rats of batch 2 in D21(128.106 \pm 5.84g; p<0.05) and D30 (129. 922 ± 7.37g; p<0.05), of batch 3 in D21 (127.116 \pm 7.13g; p<0.05), of batch 4 in D14 $(115.034 \pm 2.95g; p<0.05), D21 (122.084 \pm 3.56g; p0,05)$ and J30 (129. 922 ± 4.05g; p<0.05), of batch 5 since D14 (110. 470 \pm 11.17g; p<0.001) until D30 (116.306 \pm 7.68g; p<0.0001) and of batch 6 since D7 (102.634 ± 8.61g; p<0.001) until D30 (110.577±6.16g; p<0.0001). The rate in body weight reduction of the male animals is dose-dependent and this reduction is high to the animals of batch 6 in D30 (25.30%; p<0.0001) (Figure 1).

The females rats also knew a reduction in their body weight compared to the control batch that weighted D7 = 117.446 ± 2.32 g; D14 = 130.986 ± 5.10 g; D21= 135.108 ± 4.74 g; D30= 140.822 ± 5.49 g. According to figure 2, this reduction was significant for batch 2 in D7 (104.104 ± 2.58 g; p<0.0001) and D14 (113.016 ± 5.71 g; p<0,001), batch 3 in D7 (106.848 ± 3.02 g; p<0.001), batch 5 in D7 (103.312 ± 3.95 g; p<0.0001) and D14 (108.846 ± 9.78 g; p<0.001) and batch 6 since D7 (95.472 ± 6.64 g; p<0.0001) until D30 (103.924 ± 5.43 g; p<0.0001). In the females, this reduction in the body weight is not dosedependent, but remains very high in female animals from D14 to D30 at about 25% (figure 2).

Cadmium sulfate induced a slowdown in the growth of animals

Action of Cadmium on the Hepatic Function

Table I as well shows an increase in the activity of the ALT in the males as in the females compared to the control batches (male: 42.66 ± 9.796 UI/L and female: 40.17 ± 1.88 UI/L). This increase is significant in the males rats for batches 2 (70.62 ± 20.09 UI/L; p<0.001), batch 5 (74.47 ± 9.05 UI/L; p<0.0001) and batch 6 (161.4 ± 1.13 UI/L; p<0.0001). In the females, the increase in the activity of the ALT is significant for the batches 5 (60.18 ± 12.34 UI/L; p<0.0001) and batch 6 (77.64 ± 17.80 UI; p<0.0001).

As for the AST, the concentration of 5mg/kg bw induced a significant reduction in the activity as well in the males (223.2 \pm 35.18UI/L; p<0.05) that in the females (228.75 \pm 15.20 UI/L; p<0.001) compared to the activity at the control rats (male: 316.76 \pm 60.03 UI/L; females: 307.82 \pm 34.77 UI/L). The high concentrations did not induce a significant variation of the AST activity in female rats, whereas in the males, this high concentration (batch 6: 20mg/kg bw) induced a

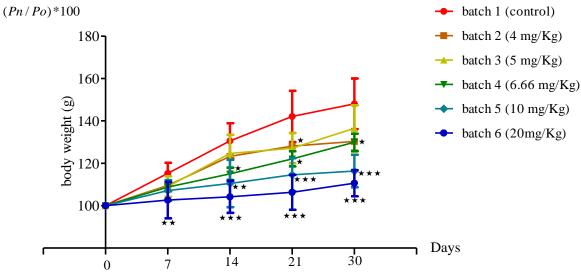


Figure 1. Effect of cadmium on the body weight of male rats Significance level:* = p<0.05;** = p<0.001;*** = p<0.0001 with n = 5

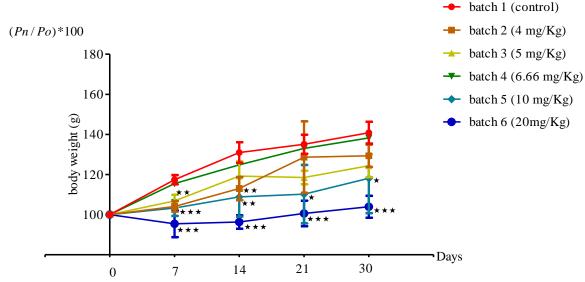


Figure 2. Effect of cadmium on the body weight of female rats Significance level:* = p<0.05;*** = p<0.001;**** = p<0.0001 with n = 5

Table 1. Effect of cadmium sulfate on the liver enzymes in the rats $% \left(1\right) =\left(1\right) \left(1\right)$

| Sex | Ma | le | Female | |
|--------------------|-------------------|-----------------|-----------------|------------------|
| Parameter | ASTL (UI/L) | ALTL (UI/L) | ASTL (UI/L) | ALTL (UI/L) |
| LOT 1 (Témoin) | 316.76 ± 60.03 | 42.66 ± 9.796 | 307.82± 34.77 | 40.17± 1.88 |
| LOT 2 (4mg/Kg) | 352 ± 48.74 | 70.62 ± 20.09** | 334.40± 3.81 | 54.00 ± 12.44 |
| LOT3 (5mg/Kg) | 223.2± 35.18* | 37.60± 10.37 | 228.75± 15.20** | 39.75 ± 2.05 |
| LOT 4 (6.66mg /Kg) | 339.7±54.76 | 58.6 ±0.61 | 315.53.± 7.87 | 54.72±9.05 |
| LOT 5 (10mg/Kg) | 374.4 ± 57.23 | 74.47±9.05*** | 324.02± 33.30 | 60.18 ± 12.34* |
| LOT 6 (20mg /Kg) | 1025.5± 66.04*** | 161.4 ± 1.13*** | 333.16± 51.83 | 77.64± 17.80*** |

Significance level: $\P^* = p < 0.05$; $\P^{**} = p < 0.001$; $\P^{***} = p < 0.0001$ with n = 5

significant increase (1025.5 \pm 66.04 UI/L; p<0.0001) (Table I) The AST and ALT activities were dose-dependent in male rats, but on the other hand, the ALT activity was not significantly varied for the female rats.

Action of Cadmium on the Renal Function

The tested concentrations induced a non-significant increase in creatinin in the rats regardless of sex from the values in the control groups (Table 2). However, according to table II, the high concentrations (6.66; 10 and 20 mg/kg bw) induced respectively a significant increase in urea in the males rats $(0.50 \pm 0.06 \text{ g/L}, \text{ p<0,001}; 0.51 \pm 0.06 \text{ g/L}, \text{ p<0.001}$ and $0.498 \pm 0.025 \text{ g/L}, \text{ p<0.0001}$). However, only the high concentration (20mg/kg bw) induced a significant increase in the females $(0.524 \pm 0.08 \text{ g/L}; \text{ p<0.05})$ compared to the control groups values (male: $0.38 \pm 0.06 \text{ g/L}$ and female: $0.3914 \pm 0.01 \text{ g/L}$). The change of renal function both in male and female rats, is correlated to the variation of the value of the urea

Action of Cadmium on the Splenic Function ¶

The rate of total bilirubin was decreased significantly for all the test concentrations (4; 5; 6.66;; 10 and 20 lg/kg bw) both in the male rats (1.35 \pm 0.7g/L; p<0.0001; 1.60 \pm 0.31 g/L; p<0.001; 1.76 \pm 0.27 g/L; p<0.0001 and 0.21 \pm 0.09 g/L; p<0.0001) than in females compared to the control rats groups (male: 2.43 \pm 0.68 g/L and female:1.62 \pm 0.26 g/L) (table 3). This reduction is dosedependent in the male rats.

For total cholesterol, table 3 shows non-significant variation both in the two sex. The disturbance of the splenic function in the rat would be due to the variation of the rate of the total bilirubin.

DISCUSSION

For the population, the principal ways of contamination to cadmium are food, water and cigarette smoke. According to conditions of exposure to metal, 5% of introduced cadmium is absorbed by the gastro-intestine tract in the form of salt, while 90% of inhaled cadmium is absorbed by pulmonary way (Jarup, 2002). The experimental study showed that the cadmium sulfate cause losses of weight and a deceleration of the growth. That can be explained by the fact why the rats treated with cadmium ate little compared to the control animals noted at the time of the study of Wang et al. (2014) according to the dose of exposure. Consequently, for Sajjadet et al. (2014), cadmium would have caused a loss of appetite in the rats (Sajjadet et al., 2014). Indeed, there is a correlation between the loss of body weight

and the cadmium concentration in the hypothalamus and the pituitary gland in the animals treated with cadmium (Sajjadet et al., 2014). For several authors, other factors can explain this loss of weight. In the guinea-pigs, the loss noted during the exposure to cadmium 5 mg/Kg is explained by damage of the system glucocorticoïd, essential hormone in the regulation of glucose and the metabolism of the lipids and proteins (Mouhamed and Azab, 2014) necessary to the loss and the profit of weight. These results of loss of weights noted at the time of the study are in agreements with those of Sandrita et al., (2003) where the mice were watered with water enriched with cadmium chloride (10 mg/L) during 8 weeks. Losses of weight, weaknesses and mortalities were noted. Indeed, Already with 0.15mg/kg cadmium sulfate considered as proportions pathological at the time of the study of Berroukche et al. (2014) showed a similar loss of weight where the weakest concentration was 4 mg/kg body weight.

The biochemical parameter makes it possible to evaluate possible toxic effects of an agent on the physiological functions of the organism responsible for many diseases (Smaoui *et al.*, 2000). The aspartate aminotransferase (AST) and alanine aminotransferase (ALT) are used in animal toxicology like markers of the dysfunction of liver (Hannah *et al.*, 2002). The quantification of the activity of these enzymes in the animals is a marker of the exposure to pollutants (Hannah *et al.*, 2002) observed in fish.

The results of the study showed a significant increase in the enzymatic activity of AST and ALT in the serum of the contaminated rats with cadmium sulfate compared to the control group in the males as in the females. Increased in these activities of enzymes in the blood, represented their leakage of tissue into plasma following a hepatic lesion responsible for the deterioration of the membrane permeability (Layachi and Kechrid, 2013). This damage and the relaxation of the enzymes in blood is due to an accumulation of cadmium sulfate in the liver. In his study, Nashwa (2013) put in obviousness two ways characterizing the hepathotoxicity of cadmium. On a side it causes an inflammation leading to damage of the liver and kidneys (Fahim et al., 2012) characterized by the high rate of white globules (Mouhamed and Azab, 2014), and other, it destroys the cells of the liver directly.

In their studies (Egwurugwu et al., 2007), the Wistar rats exposed to 200 mg/L showed a similar rise in transaminases compared to the control group rats like in fish (Jisha et al., 2013) and in chickens with a concentration of 100 ppm cadmium (Bharavi et al., 2011). The rate of accumulation of heavy metals in the animals varies according to the species and on the individuals in the population, but, also depends on the sex, the age, the size and the food (Bedii and Kenan, 2005; Haki et al., 2005).

Cadmium is biotoxic environmental pollution; it

Table 2. Effect of cadmium on kidney markers in rat

| Sexs | Male | | Female | |
|--------------------|---------------------|------------------|------------------|--------------------|
| Paramètre | Urea (g/L) | Creatinin (mg/L) | Urea (g/L) | Creatinin (mg/L) ¶ |
| LOT 1 (Témoin) | 0.38± 0.06 | 5.4± 0.54 | 0.391± 0.01 | 4.75± 0.96 |
| LOT 2 (4mg/Kg) | 0.42 ± 0.06 | 6 ± 1 | 0.395 ± 0.06 | 6.5 ± 2.38 |
| LOT3 (5mg/Kg) | 0.47±0.04 | 5.1± 0.57 | 0.393± 0.05 | 5± 1.41 |
| LOT 4 (6,66mg /Kg) | 0.50± 0.06** | 6 ± 1 | 0.4768±0.06 | 6.2 ± 0.44 |
| LOT 5 (10mg/Kg) | $0.51 \pm 0.06**$ | 5.5 ± 0.58 | 0.482± 0.09 | 5.4 ± 0.55 |
| LOT 6 (20mg /Kg) | $0.498 \pm 0.025^*$ | 6.33± 1.15 | 0.524± 0.08* | 5.2 ± 0.84 |

Significance level: $\P^* = p < 0.05$; $\P^{**} = p < 0.001$; \P with n = 5

Table 3. Effect of cadmium on spleen markers in rat

| Sexe | Male | | Female | |
|--------------------|-----------------|-------------------|-----------------|-------------------|
| Parameters | Bilirubin (g/L) | Cholesterol (g/L) | Bilirubin (g/L) | Cholesterol (g/L) |
| LOT 1 (Témoin) | 2.43±0.68 | 0.946± 0.06 | 1.62± 0.26 | 0.66± 0.10 |
| LOT 2 (4mg/Kg) | 1.35± 0.7*** | 0.96 ± 0.22 | 0.99±0.13*** | 0.72± 0.13 |
| LOT3 (5mg/Kg) | 1.60± 0.31** | 0.72 ± 0.09 | 1.08±0.08** | 0.85 ± 0.33 |
| LOT 4 (6.66mg /Kg) | 1.76±0.27* | 0.88±0.06 | 1.09.± 0.09** | 0.63±0.09 |
| LOT 5 (10mg/Kg) | 1.46 ± 0.29*** | 0.91±0.22 | 1.14± 0.01** | 0.73± 0.17 |
| LOT 6 (20mg /Kg) | 0.21± 0.09*** | 0.97 ± 0.38 | 1.16± 0.41* | 0.54± 0.11 |

Significance level: $\P^* = p < 0.05$; $\P^{**} = p < 0.001$; $\P^{***} = p < 0.0001$ with n = 5

accumulates in tissues mainly in the liver, the kidneys, osseous marrow, the reproductives bodies and in the immune system (Egwurugwu *et al.*, 2007). In addition, the major body implied in the toxicity of cadmium is liver (Singh *et al.* 2013). What is confirmed by Sandrita *et al.* (2003) where the mice contaminated with 10 mg/L cadmium recorded 0.032µg/g and 0,12 cadmium µg/g respectively in the liver and the kidneys.

The results of the study showed in animals of both sexes, a significant increase in levels of creatinine and urea in the blood compared to control group. This increase in these two parameters testifies to a renal insufficiency (Boujelbeneet et al., 2002). Indeed, a significant rate of the blood volume passes by the kidneys, them making thus extremely vulnerable to the cytotoxic effect of many pollutants such as cadmium, lead and mercury (Boujelbeneet et al., 2002). During their experiment on the evaluation of the toxicity of cadmium sulfate, the control group of Wistar rats recorded the presence of cadmium in their kidneys (Asagba and Obi, 2004), reflecting food and water pollution. Kidneys are the main targets after ingestion of cadmium (Wang et al., 2014) where 50 % of metal accumulate with a half-life from 10 to 35 years and create kidney damage (Poontaweeet et al., 2016). Cadmium causes a tubular kidney dysfunction and a decrease in glomerular filtration rate thus leading to kidney failure reported by Fahim et al. (2012). This renal failure is evaluated by creatinine and urea determination (Fahimet et al., 2012). The results showed that the increase of creatinin is not significant (p > 0.05) in the females rats. Our results corroborate those of Fahim *et al.* (2012) and Wang *et al.* (2014) who have not observed the changes of creatinine level in the blood after 28 days of cadmium treatment.

The increase of uremia translated some kidney damage and is an acute exposure marker. As for creatinine, its increase is a sign of the kidney function failure (Abdel-Moneim and Ghafee, 2007). Therefore, the exposure of the rats to cadmium in this study causes a disturbance of the renal function in the males.

The bilirubin is the breakdown product of red blood corpuscles out-of-date or damage. These red blood corpuscles will release heme and the globine (Basso *et al.*, 1991). In the liver, bilirubin binds to other molecules before being excreted in the bile. The results obtained showed a significant reduction in the bilirubin compared to the control group in both rats sexes. These results confirmed the total correlation between bilirubin and cadmium where Zeneli *et al.*, (2009) noted that the high cadmium concentration in blood is due to cigarette smoke. In fact, this cadmium inhibits the synthesis of bilirubin. The bilirubin rate reduction in blood may be caused by anemia (Zeneli *et al.*, 2009) due to cadmium.

CONCLUSION

Cadmium cause a weight decrease noted by a deceleration of the growth Cadmium causes slower

growth through decreased weight, anemia, renal failure and liver damage. These observed pathologies are the risks to people living in a polluted environment to cadmium. This is the place to draw the attention of clinicians so that incorporation metal settings to blood checkups including cadmium for better management of heavy metal poisoning.

REFERENCES

- Abdel-Moneim WM, Ghafee H (2007). The Potential Protective Effect of Natural Honey Against Cadmium-Induced Hepatotoxicity And Nephrotoxicity. Mansoura J. Forensic Med. Clin.Toxicol. XV(2): 75-98
- Adjiri AO, Lanciné GU, Kouame Innocent K, Bamory K, Biemi J (2008). Caractérisation de la pollution chimique et microbiologique del'environnement de la décharge d'Akouédo, Abidjan-Côte d'Ivoire. *Int. J. biol. Chem. Sci.* 2(4): 401-410.
- Albasha MO, El-Saied Azab A (2014). Effect of Cadmium on the Liver and Amelioration by Aqueous Extracts of Fenugreek Seeds, Rosemary, and Cinnamon in Guinea pigs: Histological and Biochemical Study. Cell Biology. 2 (2):7-17.
- Amir W, Jahanzaib A, Farhat I, Ashif S, Zahid M, Ghulam M (2014. Pollution Status of Pakistan: A Retrospective Review on Heavy Metal Contamination of Water, Soil, and Vegetables. BioMed Research International., 29 pages
- Asagba SO, Obi FO (2004). Effects of Oral Cadmium Exposure on Renal Glomerular and Tubular Functions in the Rat. J. Appl. Sci. Environ. 8 (1): 29 32
- Barbier O, Jacquillet G, Tauc M, Poujeol P, Cougnon M (2004). Acute study of interaction among cadmium, calcium, and zinc transport along the rat nephron in vivo. Am J Physiol Renal Physiol, 287, F1067-1075.
- Basso T, Fabris C, Plebani M, Del Favero G, Muraca M (1991). Alterations in bilirubin metabolism during extra-and intrahepatic cholestasis. J. Mol. Med., 70: 1432-1440.
- Berroukche A, Miloud S, Khaled K, Hafsa K, Cheikh A (2014). Evaluation de l'activité du cadmium, en présence duzinc, sur les structures des tissus régulateurs du métabolisme chez le rat Wistar. *Int. J. Biol. Chem. Sci. 8*(4): 1796-1807.
- Bharavi K, Reddy AG, Rao GS, Kumar PR, Kumar DS, Prasadini PP (2011). Prevention of cadmium bioaccumulation by herbal adaptogens. Indian Journal of pharmacology. 43(1): 45-49
- Boujelbene M, Gorbel F, AYADI F, Makni-Ayadi F, Makni-Guermazi F, Kammoun A, Croute F, Soleilhavoup JP, El eki A (2002). Impact d'un
- Cicik B, Engin K (2005). The effects of cadmium on levels of glucose in serum and glycogen reserves in the liver and muscle tissues of *Cyprinus carpio* (L. 1758). *Turk. J. Vet. Anim. Sci.* 29: 113-117.
- Deepti G, Shabad P, Dua KK (2010). Chronic Cadmium Toxicity in Rats: Treatment with Combined Administration of Vitamins, Amino Acids, Antioxidants and Essential Metals. Journal of Food and Drug Analysis, 18(6): 464-470
- Egwurugwu JN, Úfearo CS, Abanobi OC, Nwokocha CR, Duruibe JO, Adeleye GS, Ebunlomo AO, Odetola AO, Onwufuji O (2007). African Journal of Biotechnology. 6 (18): 2078-2082.
- El-Said El-Boshy M, Gadalla HA, Abd El-Hamied FM (2014). mmunological, hematological and biochemical changes induced by short term exposure to cadmium in catfish (Clariasgariepinus). Journal of Coastal Life Medicine. 2(3): 175-180
- Fahim MA, Nemmar A, Dhanasekaran S, Singh S, Shafiullah Yasin YM, Zia S, Hasan MY (2012). Acute Cadmium Exposure Causes Systemic and Thromboembolic Events in Mice. Physiol. Res. 61: 73-80.
- Galtier P, MORE J, BODIN G, BRUNEL-DUBECH N, ALVINERIE M (1974). TOXINES D'ASPERGILLUS OCHRACEUS WILHELM. III. {TOXICITE AIGUE DE L'OCHRATOXINE A CHEZ LE RAT ET LA

- SOURIS ADULTES. Annales de Recherches Vétérinaires, 1974, 5 (2), pp.233-247.
- Gonick HC (2008). Nephrotoxicity of cadmium & lead. *Indian J Med Res.* 128(4):335-52.128: 335-352
- Haki K, Fikret K, Halit C (2005). Effect of Single Dose Cadmium Chloride Administration on Oxidative Stress in Male and Female Rats. Turk J Vet Anim Sci. 29: 37-42
- Harley CB, Rajeshwari Menon C, Rachubinski RA, Nieboer E (1989). Metallothionein mRNA and protein induction by cadmium in peripheral-blood leucocytes.Biochem. J. 262: 873-879
- IARC (1993). Cadmium and cadmium compounds. IARC MonogrEvalCarcinog Risks Hum 58:119–238
- Jarup L (2002). Cadmium overload and toxicity.Nephrol Dial Transplant, 17 Suppl 2, 35-39.
- Jisha J, Hari Sankar HS, Smitha Bhanu V, Aniladevikunjamma KP, Remya V, Babu P (2013). Cadmium ion induced changes in the protein catabolism of *Oreochromis mossambicus*. Int. J. Sci. Res. Publications, 3(8): 1-8
- Layachi N, Kechrid Z (2012). Combined protective effect of vitamins C and E on cadmium induced oxidative liver injury in rats. African Journal of Biotechnology 11(93): 16013-16020
- Nashwa KI (2013). Possible Protective Effect of Kombucha Tea Ferment on Cadmium Chloride Induced Liver and Kidney Damage in Irradiated Rats. International Journal of Biological and Life Sciences. 9: 7-12
- NGUIAN Pouofo M (2011). Evaluation comparative des proprieties aphrodisiaque et androgénique de l'extrait aqueux des écorces de racinessde Uapacaguineensis et du Viagra chez les rats mâles. Université de Yaoundé. Mémoire. 60 pages
- OUMAR B, EKENGELE NL, BALLA OAD (2014). Évaluation du niveau de pollution par les métaux lourds des lacs Bini et Dang, Région de l'Adamaoua, Cameroun. *Afrique SCIENCE*, 10(2):184 198.
- Pascal Andujar, Lynda Bensefa-Colas, Alexis Descatha, 2010.Acute and chronic cadmium poisoning. Rev Med Interne, 31 (2), pp.107-15
- Poontawee W, Surapol N, Orawan W (2016). Protective Effect of Cleistocalyxnervosum var. paniala Fruit Extract against Oxidative Renal Damage Caused by Cadmium. Molecules 21(133): 1-13.
- Radhakrishnan MV (2010). Immunological effect of cadmium in *Heteropneustes fossilis* Bloch. Global veterinaria. Global veterinaria. 4(6): 544-547
- Radhouane C, Faycal F (2013). Effets des expositions simultanées aux métaux et aux polluants organiques. Anses, Bulletin de veille scientifique, 22: 26-29
- Sajjad S, Malik H, Farooq U, Rashid F, Nasim H, Tariq S, Rehman S (2014). Cadmium Chloride Toxicity Revisited :Effect on Certain Andrological, Endocrinological and Biochemical Parameters of Adult Male Rabbits. Physiol. Res. 63: 505-512
- Sampath H, Ramesh M, Manavalaramanujam (2002). Responses of plasma transaminase activity in cyprinuscarpio var. communis to mercury toxicity. Journal of the Indian Fisheries Association. 29: 7-13
- Šimonytė S, Čerkašin G, Plančiūnienė R, Naginienė R, Ryselis S, Ivanov L (2003). Influence of cadmium and zinc on the mice resistance to Listeria monocytogenes infection. MEDICINA. 39: 767-772
- Singh N, Rani P, Gupta M, Goel N, Tandan N (2013). Effects of aqueous extract of Camellia sinensis (I.) O. kuntze on liver markers of cadmium treated rats. Journal of Biotechnology and Pharmaceutical Research. 4(5):89-93.
- Smaoui M, Fatma G, Manel B, Fatma MA, Abdel fattah E (2000). Impact de l'exposition chronique aux gaz d'échappement d'origine automobile sur certains biomarqueurs touchant la fonction hormonale sexuelle mâle, la fonction rénale et l'hémogramme chez le rat. POLLUTION ATMOSPHÉRIQUE. 167: 439-449
- traitement chronique au cadmium sur la fonction renale chez le rat. Détermination d'un biomarqueurd'exposition = Kidney's deficiency induced by the cadmium. *L' Eurobiologiste*, 35(258): 3-17.
- Traore A, Soro G, Ahoussi KE, Bamba BS, Soro N, Biemi J (2014).

 Niveau de contamination en métaux lourds des sédiments d'une

- lagune tropicale : la lagune Aghien (Sud-Est de la Côte d'Ivoire). Afrique Science 10(3): 73 88.
- TRAORE A, SORO G, AHOUSSI KE, BAMBA BS, SORO N, BIEMI J (2014). Niveau de contamination en métaux lourds des sédiments d'une lagune tropicale : la lagune Aghien (Sud-Est de la Côte d'Ivoire). Afrique SCIENCE 10(3):73 88.
- WANG HJ, LIU ZP, JIA XD, CHEN H, TAN Y (2014). Endocrine Disruption of Cadmium in Rats Using the OECD Enhanced TG 407 Test System. Biomed Environ Sci, 27(12): 950-959.
- Yao MK, Kouassi N'g LB (2015). Etude des propriétés d'adsorption et de désorption du Plomb (Pb) et duCadmium (Cd) par les sédiments d'une lagune tropicale en présenced'Allylthiourée. *Int. J. boil. Chem. Sci.* 9(1): 483-491.
- Zeneli L, Paçarizi H, Daci NM, Daci-Ajvazi M, Prenaj A (2009). The Effects of Air Pollution and Smoking on CadmiumConcentration in Human Blood and Correlation with Biochemical Parameters. Am. J. Biochem. & Biotech. 5 (2): 59-62

Zienab Mohamed Y, Heba El-Gharieb H (2009). Effect of Cadmium Chloride On Function Of Thyroid Gland In Rats. Egypt. J. Comp. Path. Clinic. Path. 22(3): 10 – 23

How to cite this article: DIABY V, YAPO AF, ADON AM, DOSSO M, DJAMA AJ (2016). Renal, Hepatic and Splenic Biotoxicity of Cadmium Sulphate in the Wistar Rats. Int. J. Environ. Sci. Toxic. Res. Vol. 4(6): 103-110