

Comprehensive review on electrospinning techniques as versatile approaches toward antimicrobial biopolymeric composite fibers

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1	Comprehensive review on electrospinning techniques as versatile
2	approaches toward antimicrobial biopolymeric composite fibers
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1 Graphical Abstract

2



12 Abstract

2 Electrospun (bio)polymeric fibers have attracted widespread interest as functional materials with suitable morphology and properties for their use as tissue engineering scaffolds and/or 3 wound dressings. The fibrous/porous morphology of this type of materials promotes the 4 5 adhesion and proliferation of tissue cells, but on the other hand, pathogenic microorganisms 6 unfortunately can also be attached to the fibers, thus leading to serious infections and 7 consequently to the immediate removal of the scaffolds or wound dressings, which may imply greater tissue damage. In this context, this review addresses the more recent 8 approaches based on electrospinning and related techniques for developing composite 9 10 (bio)polymeric fibers with tailored antimicrobial properties either by using mere electrospinning for the incorporation of well-defined antimicrobial nanoparticles (silver, 11 gold, titanium dioxide, zinc oxide, copper oxide, etc.) or by resorting to the combination of 12 electrospraying and electrospinning for the generation of nanoparticle-coated fibers, as well 13 as coaxial electrospinning for obtaining fibers with nanoparticle-rich surface. 14

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Keywords: electrospinning; electrospraying; coaxial electrospinning, antimicrobial
 particles; biopolymers; fibrous composites

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1 1. Introduction

2

Polymeric fibers derived from electrospinning and related techniques have recently been 3 4 exploited for obtaining new materials with potential application in medicine, energy, and environmental issues [1-7]. The most attractive features of these materials are those related 5 6 to the morphology, such as fiber diameter in the sub-micrometric regime, high porosity 7 with interconnected voids, large surface area-to-volume ratio, etc. In this context, the ability 8 of fibrous polymeric materials to mimic the extracellular matrix is particularly important 9 for their use in tissue engineering, as scaffolds and/or wound dressings, since it promotes 10 good adhesion and proliferation of tissue cells [1, 8]. Nevertheless, the fibrous morphology is prone not only to tissue cell ingrowth but also to pathogenic microorganisms; therefore, it 11 12 is necessary to develop polymeric fibers with biocide properties but without cytotoxic effects on the receiving tissue. 13

14

In this regard, a vast diversity of organic compounds with recognized antimicrobial activity 15 have been incorporated into polymers and subsequently subjected to electrospinning 16 techniques in order to develop antimicrobial fibers. The reported organic compounds 17 18 exhibit a relevant drawback related to their thermal and chemical instability, which could lead to difficulties throughout the electrospinning process and also produce fibrous mats 19 20 with limited antimicrobial activity due to partial decomposition of the antimicrobial agent. 21 Moreover, the massive use of antibiotics has provoked that a wide range of microorganisms developed a strong resistance against the majority of drugs; therefore, there is a need to 22 explore novel antimicrobial agents [9-11]. 23

Nanostructures with well-known antimicrobial properties have emerged as promising 1 2 compounds for developing fibrous mats with better antimicrobial performance than those containing typical organic compounds, such as antibiotics and biocides. Likewise, 3 antimicrobial nanoparticles, including zinc oxide, titanium dioxide, silver, and gold 4 analogues, have demonstrated some advantages like their higher chemical and thermal 5 stability that may lead to a new generation of biomedical devices with improved 6 7 antibacterial activity as well as enhanced mechanical, thermal and other physical properties 8 [12].

9

In this context, the most recent reports related to approaches for obtaining antimicrobial polymeric fibers by means of electrospinning and related techniques are addressed in the subsequent sections of this review. Some significant concepts of electro-hydrodynamic techniques (*i.e.*, electrospraying and electrospinning) and typical polymers meant for the design of biomedical devices are also discussed. Lastly, the main mechanism of microbial adhesion to the (bio)polymeric fibers as well as that related to the antimicrobial activity associated with nanoparticle-containing composite fibers are detailed.

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18 2. Overview of electrospinning and related techniques

19

In this section, basic principles of electrospraying and electrospinning are addressed as well
as the most important process and solution parameters that influence fiber morphology.
Furthermore, some recent approaches based on these techniques for producing composite
fibrous materials are analyzed.

24

Electrospraying is basically the atomization of a liquid (passing through a capillary) by 1 2 means of the application of a high voltage (typically 1-30 kV). This electro-hydrodynamic technique can be used for obtaining either inorganic or polymeric nano- or micro-particles 3 derived from the corresponding solution/dispersion, as depicted in Figure 1. The process 4 begins at the tip of the capillary, where a droplet of the solution/dispersion is subjected to 5 6 high voltage, whose electric charges generate an electrostatic force inside the droplet, 7 which is eventually deformed into a cone (known as Taylor cone) from which smaller droplets will be ejected as soon as the surface tension is overcome. These generated 8 droplets are accelerated towards the grounded collector, and the solvent is simultaneously 9 10 evaporated throughout that occurrence [3, 13-15].



11

Figure 1. Scheme of the electrospraying process, more specifically the section between needle and collector, in which the formation of the Taylor cone, the Coulomb fission and the solvent evaporation take place.

15

16 The versatility of the electrospraying technique is attributed to several advantageous17 characteristics, including:

2 Droplet size is smaller than that available from conventional mechanical atomizers, and 3 can be smaller than 1 µm. Therefore, the resulting particles can be in the nanoscopic regime. 4 The size distribution of the droplets is usually narrow, with small standard deviation 5 • 6 that allows for production of particles of nearly uniform size. 7 Charged droplets are self-dispersing in space (due to their mutual repulsion), resulting 8 in the absence of droplet coagulation, and consequently lower agglomeration. 9 The motion of charged droplets can be easily controlled (including deflection or • 10 focusing) by varying electric fields. The deposition efficiency of a charged spray on an object is one order of magnitude 11 • 12 higher than that for uncharged droplets. 13 The morphological characteristics of the particles derived from the electrospraying 14 15 technique are affected by different parameters related to both the solution/dispersion properties and process conditions, which will be described below. 16 17 18 Electrical conductivity is a fundamental solution/dispersion property since it allows for the charge transfer inside the liquid, so in turn, this property is governed by the nature of 19 solvent and solute. Generally, an increase in electrical conductivity results in a smaller 20

their Coulomb fission (breakup). Solution/dispersion viscosity is another property that
strongly influences the electrospraying behavior and final morphology, and it is

21

8

particle diameter due to a higher number of charges around the droplets, which promotes

proportional to solute concentration and molecular weight (especially for polymers). As a
rule, the solution/dispersion viscosity should be sufficiently low to allow for the formation
of droplets; moreover, a highly viscous solution requires higher voltage in order to
overcome the surface tension which is not practical [3, 13-15].

5

Regarding the process conditions, solution/dispersion flow rate is a crucial parameter for 6 7 controlling the final morphology of electrosprayed nano- or micro-particles along with the 8 previously discussed solution parameters. The flow rate should be in a sufficiently low 9 range so that complete solvent evaporation might occur, which is not possible when a 10 higher flow rate is used. In case of incomplete solvent evaporation, a distorted morphology is commonly obtained due to fusion of wet and partially solvated particles after deposition 11 12 on the collector. The relationship between the flow rate, solution/dispersion properties and particle diameter is established by Equation 1 [16]. 13

14
$$d_p = \Phi^{1/3} d \sim G(\varepsilon) \left(\frac{Q\varepsilon\varepsilon_0}{\kappa}\right)^{1/3}$$
(1)

15

where d_p and d are the particle and droplet diameter, respectively, Φ is the solute volume fraction, Q is the flow rate, ε and ε_0 are the permittivity of solution/dispersion and vacuum, respectively, and κ is the solution/dispersion conductivity.

19

Other process parameters, more specifically those related to the electrospraying equipment, are the capillary diameter and capillary-to-collector distance. The former determines the size of the Taylor cone, which in turn influences the droplet and particle diameter. Concerning the capillary-to-collector distance, it was demonstrated that a shorter distance is beneficial for generating a higher electrical field strength, thus producing smaller particles
but also has the drawback of possible electric discharge and insufficient time for solvent
evaporation, which may lead to coalescence/agglomeration of wet particles on the collector.
Similarly, the applied voltage provokes a strong effect on particle morphology: the higher
the applied voltage, the lower the particle size, which can be explained in terms of higher
charge density in the solution/dispersion and breakup of ejected droplets [3, 13-15, 17].

7

8 Electrospinning is based on the same electro-hydrodynamic phenomena previously 9 discussed for the case of electrospraying. Therefore, the morphology of resulting materials 10 depends on similar parameters: electrical conductivity and viscosity of solution/dispersion, 11 flow rate, capillary diameter, capillary-to-collector distance and applied voltage. The main 12 difference between electrospinning and electrospraying is that the former technique 13 generates a continuous jet from the Taylor cone, thus collecting a mat consisting of 14 polymeric fibers [1, 18-20].

15

It is important to mention that fibers are typically engineered from a solution containing 16 high molar mass compounds, *i.e.* polymers, since the stability of the jet is a result of chain 17 18 entanglements. In this regard, a critical polymer concentration (i.e., viscous polymeric solution) is required for the occurrence of chain entanglements, thus producing fibers. 19 20 Below this critical polymer concentration (*i.e.*, dilute regime) only droplets are collected, 21 *i.e.* electrospraying takes place, since polymer chains are separated by the solvent. With an intermediate concentration (i.e., semi-dilute regime), the polymer chains overlap each 22 23 other, but this is not enough for significant entanglement formation, therefore, particle-like morphology is still dominant. Fibers with high number of beads can be obtained up to the 24

entanglement concentration (C_e), *i.e.* the transition between semi-dilute unentangled and 1 2 semi-dilute entangled regime. Above the entanglement concentration, fibers with welldefined morphology can be produced, which generally is 2-2.5 times C_e . The polymeric 3 solution regimes and their repercussions on morphology of the obtained materials are 4 5 illustrated in Figure 2. It is worth mentioning that the solution/dispersion and process 6 parameter described in previous sections for the case of electrospraying process also affect 7 the final morphological parameters of electrospun fibers, namely average fiber diameter, fiber diameter distribution and porosity ratio [21-23]. These solution and process factors as 8 well as their corresponding effect on morphology are summarized in Table 1. 9



10

11 Figure 2. Influence of the concentration of the polymeric solution on chain entanglements,

12 formation of fibers and their final morphology derived from the electrospinning process.

- **Table 1.** Effect of different parameters on the morphology of polymeric fibers derived from
- 2 the electrospinning process

Parameter	Effect on fiber morphology			
Increase in applied voltage	Fiber diameter initially decreases, then increases (not monotonic)			
Increase in flow rate	Fiber diameter increases (beaded morphologies occur if the flow rate is too high)			
Increase in needle-to- collector distance	Fiber diameter decreases, (beaded morphologies occur if the needle-to-collector distance is too short)			
Increase in polymer concentration (viscosity)	Fiber diameter increases (within optimal range)			
Increase in solution conductivity	Fiber diameter decreases (broad diameter distribution)			
Increase in solvent volatility	Fibers exhibit surface microtexture (pores/rugosity on their surfaces, which may increase surface area)			
Source: Sill and von Recum 2008 [11].				
3. Design of antimicrobial composite fibers based on electrospun polymers and inorganic nanoparticles				

As previously mentioned in the Introduction, polymeric fibers derived from electrospinning 1 2 techniques are suitable for applications related to tissue engineering (scaffolds and/or wound healing materials) due to the sub-micrometric fibrous structure with interconnected 3 pores that could mimic the extracellular matrix, thus promoting the adhesion and 4 proliferation of distinct tissue cells. Nonetheless, that particular morphology is also prone 5 to adhesion of pathogenic microorganisms, which can originate a protecting assembly 6 7 known as biofilm (composed of extracellular DNA, proteins and polysaccharides). For this 8 reason, there is a trend to develop new electrospun composite materials with antimicrobial properties. Although there are several organic compounds with antimicrobial activity, 9 10 nanoparticles have recently been preferred since they could confer new properties and/or enhance others. Hence, the subsequent sections will firstly focus on the design of 11 12 composites fibers by electrospinning and related techniques and, secondly, on the most studied antimicrobial nanoparticles and their effect on the generation and final properties of 13 14 (bio)polymeric fibers [24-34].

15

16 17

3.1. Approaches for designing fibrous composite materials by electrospinning and related techniques

18

Several approaches for fabricating composite materials with tailored morphology have been studied, whose basic methodology and some advantages/disadvantages will be revised in this section. To get a better understanding of the electrospinning and related techniques, Figure 3 shows a scheme of the different steps for obtaining composite fibrous materials and the resulting morphology.

24

Mere electrospinning is the most reported approach for obtaining composite fibers, and it 1 2 consists in preparing a polymeric solution containing the corresponding antimicrobial nanoparticles or their precursory compounds, which eventually is subjected to 3 electrospinning process. The main advantage of this approach is its relatively 4 straightforwardness, thus producing polymeric fibers with nanoparticles mostly embedded 5 6 (some particles could however migrate to the fibers surface during the electrospinning 7 process). On the other hand, the main disadvantage is related to the low antimicrobial efficiency due precisely to the embedding of nanoparticles, thus reducing the surface area 8 9 prone to interact with pathogen microorganisms [12,35-37].

10

Nanoparticles-coated polymeric fibers can be engineered by two techniques. One of them 11 12 involves a two-step process, *i.e.* mere electrospinning to obtain neat polymeric fibers from the corresponding solution, then followed by the deposition of the antimicrobial 13 nanoparticles by several well-known techniques, such as sputtering or impregnation. The 14 second technique is a tandem process denominated electrospinning/electrospraying, which 15 consists of a simultaneous electrospinning of a polymeric solution and the electrospraying 16 of the corresponding antimicrobial nanoparticles dispersion. Both techniques have been 17 18 utilized for enhancing the properties dependent on the nanoparticles surface area, such as antimicrobial activity; however, an uninvestigated concern is related to the adhesion and 19 20 loss of nanoparticles. Comparing the techniques, electrospinning/electrospraying tandem 21 process has the advantage of producing nanoparticle-coated polymeric fibers in one-step. Unfortunately, complementary equipment (e. g. electrospraying) is needed for carrying out 22 23 the fabrication of the composite fibers, thus increasing the initial technological investment 24 [38-46].

Another outstanding technique is coaxial electrospinning, which can be performed in a similar apparatus than that used for conventional electrospinning, but the ejection device is formed by concentric needles, thus when solutions are ejected from them, core-sheath fibers are collected. Combination with other electrospinning-related techniques could be used for generating a variety of morphologies with potential applications as antimicrobial devices for medical and environmental concerns (see Figure 3) [4,47-51].

7

The wide range of electrospinning and related techniques for designing composite fibrous 8 materials has permitted to tune the antimicrobial activity. As a matter of example, mere 9 10 electrospinning could produce antimicrobial activity based on migration of ionic species form the fibers, thus producing wound dressings or scaffolds for controlled release during 11 12 long time. In contrast, the use of electrospinning/electrospraying tandem process could produce antimicrobial materials for rapid antimicrobial activity when needed. Further 13 insights related to the effect of electrospinning techniques, type of antimicrobial 14 nanoparticles and others relevant factors on composite fibrous materials properties, 15 especially antimicrobial activity, will be discussed in subsequent sections. 16



Figure 3. Scheme of the approaches for designing composite fibrous materials engineered
by electrospinning and related techniques and the corresponding outer and inner
morphologies: (1,2) mere electrospinning, (3-5) electrospinning/electrospraying and (6-8)
coaxial electrospinning.

3.2. Polymers used as fibrous matrices for tissue engineering

9 An appropriate polymer for tissue engineering applications should be biocompatible,10 biodegradable, nontoxic, non-mutagenic, and non-immunogenic. Polymers are generally

classified into two types, namely synthetic polymers and natural polymers (*i.e.* biopolymers) [52].

3

Natural polymers are produced by biological systems (i.e., microorganisms, plants, and 4 animals) whose main advantages are their bioactivity, *i.e.* the ability to interact with the 5 surrounding host tissue and to induce cell deposition/proliferation. Moreover, this type of 6 7 polymers provokes low inflammatory effect, they can be degraded by enzymes, and their 8 degradation products can be consumed throughout the cellular metabolism. However, 9 natural polymers are highly temperature sensitive, and their complex structure makes them 10 difficult to be processed by electrospinning techniques. Another well-known disadvantage of natural polymers is the highly divergent properties as a function of the bio-source or 11 12 batch. Two types of natural polymers are generally utilized for tissue engineering: (i) polysaccharides, such as chitin, chitosan, and alginate; (ii) proteins, such as collagen and 13 14 gelatin.

15

Synthetic polymers on the other hand, are highly useful in tissue engineering as their 16 properties (e.g., porosity, degradation time, and mechanical characteristics) can be easily17 18 tailored for specific requirements of medical devices and host tissue. Furthermore, these polymers are often less expensive compared with natural analogues, and they can be 19 20 produced in larger quantities. Since natural polymers exhibit certain difficulties for processing them in electrospinning techniques (*i.e.*, mainly solubility), synthetic polymers 21 are typically blended with low amounts of natural polymers, which enhances their 22 23 solubility [48, 53-58]. Several reviews have hitherto been published in which the mechanical and thermal properties as well as biocompatibility, solubility and 24

biodegradation of both natural and synthetic polymers are compiled and thoroughly
analyzed. Therefore, further information can be consulted for specific polymeric systems in
these feature publications [59-63].

4

5 3.3. Antimicrobial nanomaterials and their incorporation into biopolymeric fibrous 6 materials

7

8 Several inorganic nanostructures have been incorporated into polymeric fibers by using electrospinning and related techniques. The inspection of recognized scientific data bases 9 10 generates interesting statistical results (see Figure 4). Approximately 4000 reports related to nanofibrous composites have been published over the last decade. About 30% of these 11 12 publications have focused on the use of well-known antimicrobial metallic (silver and gold) and metal oxide (mainly those based on zinc, titanium, copper and iron) nanoparticles. 13 Researchers have mainly focused on the analysis of process and solution properties 14 associated with electrospinning techniques, and on final fiber morphology and properties, 15 such as optical, mechanical, photo-catalytic, among others. Nevertheless, it is noteworthy 16 that only one third out of the resulting materials has been used for antimicrobial 17 18 applications, whose more relevant results are discussed below.



Figure 4. (a) Comparison of the number of publications per year related to electrospinning and related techniques used for developing composite polymeric fibers. (b) Percentage of publications which are focused on antimicrobial electrospun fibers with different nanoparticles. Source: ISI Web of Knowledge, May 2018.

6

7 **3.3.1.** Metallic nanoparticles

8

9 Silver nanoparticles (nano-Ag) have been extensively studied as antimicrobial agents in
10 (bio)polymeric fibers derived from electrospinning and related techniques, due to their
11 exceptional ability to kill a wide range of pathogenic microorganisms. Regarding the
12 polymeric matrices used, several aliphatic polyesters, such as polylactide (PLA) or its co13 polymers, poly(ε-caprolactone) (PCL), and poly(3-hydroxyalkanoate)s (PHAs), have been
14 preferred, due to their well-known biodegradation behavior and good mechanical properties
15 for tissue engineering applications [29, 64].

16

17 The (bio)polymer/nano-Ag composite fibers can be obtained from polymeric solutions 18 containing the as-synthesized nanoparticles (Ag^0) or some silver salt. Silver-based 19 compounds can enhance the spinability of the solution, leading to well-defined

(bio)polymeric fibers, which has been attributed to the increase in electrical conductivity by 1 the presence of Ag^+ ions released from Ag^0 or originated from the silver salt dissociation. 2 Currently there is a trend to add AgNO₃ as a precursor of nano-Ag since this salt is easily 3 solubilized in polar solvents typically used for electrospinning, thus increasing the electrical 4 conductivity. As expected, the presence of Ag⁺ ions strongly affects the final morphology 5 of fibers: at low concentration (< 8%) average fiber diameter is generally reduced, and on 6 7 the contrary, at high concentration (> 8%) the agglomeration of Ag species promotes phase 8 segregation, thus increasing the average fiber diameter [65-68].

9

10 The use of AgNO₃ as precursory species for obtaining (bio)polymeric fibers with nano-Ag involves a reduction step, which can be induced by heat [69], chemical reducing agents [36, 11 12 70] or photo-irradiation [32, 33, 69, 71, 72]. Recently, UV-induced photochemistry has been taken advantage because of its simplicity, low cost, and sustainability. Interestingly, 13 14 several researchers have claimed about the fabrication of polymer/nano-Ag composite 15 fibers with controlled antimicrobial activity. This can be achieved by electrospinning the corresponding solution containing AgNO₃, followed by a photo-reduction. The size, 16 content, and dispersion/distribution of the embedded Ag nanoparticles are controlled by 17 18 AgNO₃ concentration and time of UV irradiation. Long irradiation time promotes the migration of Ag^+ ions through the outer layer of the fibers as well as the production of Ag^0 19 20 and its increase in size, thus tailoring the antimicrobial performance reaching up to 99.9% of growth inhibition at relatively low AgNO₃ concentration (from 0.1 to 1 wt-% in relation 21 to polymer) [32, 33, 69, 71, 72]. Thus, poly(vinyl alcohol) (PVA) [66, 69], cellulose 22 derivatives [32, 71-73], poly(lactic acid) [68], gelatin [33], chitosan, poly(ethylene oxide) 23 or polymer blend-based fibers [55, 57, 58, 74] have been engineered with high 24

antimicrobial *in vitro* performance against Gram-positive (mainly *Staphylococcus aureus*)
and Gram-negative bacteria (*Escherichia coli, Klebsiella pneumonia, Pseudomonas aeruginosa, Proteus mirabilis, Bacillus cereus*), as well as fungal culture (*Candida albicans*).

5

The ideal composite polymeric fibers must possess a nanoparticle-rich surface in order to 6 7 exhibit suitable antimicrobial activity for using in medical devices, which is very difficult 8 to achieve by mere electrospinning, since nanoparticles are buried in the inner part of the 9 fibers. However, a more accurate procedure is to create fibers with core-sheath 10 morphology, where the sheath contains Ag nanoparticles. These particular fibers can 11 exclusively be obtained by an alternative technique, the so-called coaxial electrospinning, 12 which is based on the same principles of simple electrospinning, but the system that ejects the polymer solution consists of two coaxial capillaries with different diameters by which 13 14 two fluids converge forming a core of one polymer covered by the other polymeric system. 15 The second polymeric solution could contain specific nanoparticles in order to produce a 16 composite sheath with enhanced antimicrobial properties.

17

Coaxial electrospinning has scarcely been exploited for the design of polymer/nano-Ag composite fibers, and the selected matrix has not been necessarily biocompatible or biodegradable. Yu *et al.*[51] have deposited Ag nanoparticles preferentially on the surface of polyacrylonitrile (PAN) fibers by coaxial electrospinning of the corresponding polymeric solution (core) and N,N-dimethylacetamide/AgNO₃ solution (sheath). An additional device for UV irradiation was adapted near to the capillary tip in order to reduce the Ag⁺ ions during the ejection process. Interestingly, the authors found that the solution used for sheath

generation provoked the formation of thinner fibers than those derived from neat PAN 1 2 solution, which was attributed to the increase in electrical conductivity, *i.e.* more induced charges over the polymer solution surface, thus reducing the average fiber diameter [49]. 3 4 Moreover, it was demonstrated that the feed rate ratio of sheath/core precursory solutions played a significant role for accomplishing good nanoparticle dispersion, since it regulated 5 the deposition of Ag⁺ ions on the core jet during the process of coaxial electrospinning. The 6 7 obtained composite fibers showed a high bacterial growth inhibition (>99.9%) against Escherichia coli and Bacillus subtilis with only 4% Ag nanoparticles. Unfortunately, the 8 authors did not compare with fibers obtained by mere electrospinning to corroborate the 9 10 effect of the specific technique used.

11

12 Castro-Mayorga *et al.* have recently reported an electrospinning-based approach to develop antibacterial multilayer materials consisting of a poly(3-hydroxybutyrate-co-3-13 hydroxyvalerate) (PHBHV) film coated with submicron fibers of PHAs and Ag 14 nanoparticles. The obtained films exhibited high toxicity against Listeria monocytogenes 15 and Salmonella enterica. More interestingly, the antimicrobial activity was achieved with 16 only 0.002 wt-%, while preserving thermal, mechanical and optical performances for their 17 18 use as food packaging [75].

19

There are some biocompatible and biodegradable polymers with good mechanical performance for wound dressing materials and/or tissue engineering scaffolds, such as PCL. Nevertheless, its hydrophobicity limits a suitable cell attachment and proliferation, thus limiting its use in the aforementioned biomedical devices. Coaxial electrospinning offers the possibility to adapt the surface properties by forming a polymeric sheath with

different chemical nature. In this regard, Kalwar et al. [48] designed co-electrospun fibers 1 2 based on PCL/chitosan (core/sheath) with average diameters ranging from 200 to 450 nm. The presence of chitosan promoted an increase in the hydrophilicity of co-electrospun 3 4 fibers, and on the other hand, well-dispersed Ag nanoparticles into the sheath due to interactions between nanoparticles and amino and hydroxyl groups of chitosan. 5 6 Furthermore, the obtained fibrous materials exhibited good inhibition growth of E. coli and 7 S. aureus when subjected to in vitro tests (disc diffusion method) and insignificant 8 difference was observed as a function of the bacterium type.

9

10 Further studies related to the use of co-electrospun fibrous materials as wound dressings and tissue engineering scaffolds have been published by Duan and Wei [47, 50]. The 11 12 authors described the production of co-electrospun fibers based on multi-components: PCL core containing vitamin A and collagen sheath with embedded Ag nanoparticles. The 13 obtained materials showed good swelling ratio (190%) and antibacterial activity against S. 14 15 *aureus (in vitro)*. On the other hand, the fibers exhibited a persistent release of vitamin A within 72 h, in conjunction with good cell attachment (L929 cells), thus indicating that 16 these fibrous mats could be used as biocompatible dressings with dual function, namely 17 18 antibacterial and wound healing ability. Nonetheless, the authors did not give any information about final Ag nanoparticles content, which could be useful for determine the 19 20 minimal amount at which nanoparticles do not cause cytotoxicity.

21

Even though polymer/nano-silver fibrous composites derived from electrospinning techniques have showed promising antimicrobial properties, one of the most important concerns is the potential toxicity of nanoparticles when exposed to tissue cells, which could

limit their massive production and application. Currently there are controversial results 1 2 associated with the viability of different tissue cells when exposed to polymeric mats 3 containing the aforementioned metallic nanoparticles. As a matter of example, Sumitha's 4 work evidenced that Ag-embedded PCL fibers (200-260 nm in diameter) could efficiently kill S. aureus bacteria (disc diffusion tests) at concentration as low as ~0.002 wt-% without 5 compromising the adhesion of human mesenchymal stem cells (viability ~99%). On the 6 7 contrary, such PCL fibers with ~0.06 wt-% Ag exhibited 50% cell viability, implying that 8 nanoparticles induced toxicity to cells at very low concentration [76].

9

10 In 2014, Lin et al. evaluated the viability of the PA317 cell line (mouse fibroblasts with herpes gene) in the presence of PVA/Ag composite mats derived from electrospinning and 11 subsequent chemical reduction (induced by the matrix, UV and heat). Some relevant 12 findings were reported, such as the effect of reduction methodology on the amount and size 13 14 of formed nano-Ag, and consequently on the antibacterial properties against E. coli and S. aureus (all materials showed antibacterial activity >99%). The authors concluded that the 15 critical factor to avoid cell toxicity was to select an appropriate reduction process by which 16 no Ag⁺ ions remained into PVA fibers, since these ionic species hampered the proliferation 17 of PA317 cells. The Ag⁰ nanoparticles did not induce toxicity even at high concentration 18 (10 and 12 wt-%), thus increasing the potential of these mats as antibacterial wound 19 20 dressings and/or scaffolds [77].

21

Additional reports have corroborated the good biocompatibility of electrospun biopolymeric fibers with embedded Ag nanoparticles [78-81]. In spite of these encouraging results, most of the evaluation approaches have been carried out by *in vitro* tests, so that it

is necessary to perform more realistic studies, *i.e. in vivo* tests, which are limited. On this 1 2 topic, antibacterial electrospun fibers (130-190 nm in diameter) based on poly(vinyl alcohol) and chitosan oligosaccharides (2:1 ratio) containing silver nanoparticles (15-22 nm 3 in diameter) or AgNO₃ (both silver species at 3 wt-% compared to polymer) were subjected 4 to *in vivo* tests with the aim of determine their biocompatibility. Initially, the obtained mats 5 6 were placed on dorsal skin of rabbits, and the irritation skin levels were monitored. It was 7 observed that mats caused no irritation to normal skin after 72 h, indicating the 8 acceptability of these fibrous materials for transdermal drug delivery. Furthermore, the 9 wound healing performance of PVA-based mats was inferred by wound healing and 10 histological examination in rat back during 14 days. It was observed that PVA-based nanofibers containing Ag nanoparticles did not interfere with the healing process and the 11 12 wound closure was accomplished in the analyzed period. Moreover, PVA/chitosan/nano-Ag mats showed histologically superior wound healing than commercial materials (wound 13 plast and gauze) and PVA/chitosan/AgNO₃ fibers since wounds with these mats displayed 14 ulcerated surfaces, formation of granulation tissue, and infiltration of inflammatory cells 15 after 7 days. On day 14, newly synthesized fibrous tissue and sparse inflammatory cells in 16 the dermis and subcutis were covered by completely re-epithelialized epidermis in each 17 18 group, thus confirming that the PVA/chitosan electrospun fibers with Ag nanoparticles could provide a suitable scaffold for wound healing [53]. 19

20

Dong *et al.* have also provided information about the *in vivo* biocompatibility of PCL fibers containing silver-decorated mesoporous silica (Ag-MSN, 0.05 wt-% to polymer). As shown in Figure 5, skin injuries were purposely made on the specimens (male Wistar rats). It was found that the wounds healed better and faster after being treated with Ag-MSN/PCL fibrous membranes than with other materials (gauze). On week 5, a rough and thin skin layer was formed in the presence of PCL/Ag-MSN fibers and without signals of inflammation by histological analysis. In contrast, the wounds without one such dressing, with neat PCL or gauze, were still in the late phase of inflammation. These results demonstrated that the Ag-MSN/PCL nanofibrous membranes could strongly help the healing process [82].



Figure 5. Skin wound healing accelerated by Ag-MSN/PCL nanofibrous membranes. (A)
Exhibition of the entire skin injury model and treatment process in a Wistar rat. (B) Gross
observation of skin wounds treated with gauze, virgin PCL, Ag-MSN/PCL respectively at
1, 2, 3, 4, and 5 weeks after injury. Untreated wound was considered as the control. Bars
represent 10 mm [82].

1

2 Other researchers have exploited collagen as the matrix for electrospinning, as this natural polymer is the most abundant protein in the human body, a key element of the extracellular 3 matrix and imparts structural integrity and tensile strength to tissues. Tissue disruption 4 following injury requires collagen for the repair and restoration of structure and function. 5 Rath et al. designed cross-linked collagen/silver nanoparticles through electrospinning. 6 7 The obtained mats inhibited the proliferation of S. aureus and P. aeruginosa (minimum inhibitory concentration of 5.8±0.3 mg/mL and 7.4±0.2 mg/mL, respectively). It was also 8 9 demonstrated an increase of wound healing by wound examination, histological study, and 10 hydroxyproline determination. Complementary studies, such as swelling index and quantification of silver ions released helped to suggest that the presence of collagen formed 11 hydrophilic gels that maintained a moist environment, thus promoting faster re-12 epithelization and wound contraction. Besides, anti-inflammatory property of nano-Ag also 13 promoted wound healing by reducing the level of pro-inflammatory cytokine or decreasing 14 15 mast cell infiltration. Finally, controlled release profile of silver nanoparticles supported the wound-healing process by maintaining the drug concentration in the therapeutic range over 16 an extended period of time [83]. 17

18

19 Recently, the addition of natural additives has emerged as a sustainable trend and for 20 imparting multifunctionality to polymer fibers containing Ag nanoparticles. As a matter of 21 example, rice flour-based fibers containing Ag and PVA/ β -cyclodextrin as binders have 22 been studied as sustainable filters. The authors demonstrated the multifunctionality of the 23 obtained mats through antibacterial tests, air permeability and adsorption of volatile organic 24 compounds. Fibrous materials exhibited high inhibition growth of *E. coli* and *S. aureus*, due to the presence of nano-Ag as well as good removal of toluene attributed to βcyclodextrin without affectation on air permeation and mechanical performance [84].
Similarly, soy protein has been used in conjunction with Ag to confer toxic gases removal
and antibacterial activity in fibrous materials based on polyamide 6 [85].

5

Gold nanoparticles have been also incorporated into electrospun biopolymeric fibers for 6 7 improving antimicrobial properties. Likewise, these metallic nanoparticles have attracted a 8 particular attention mainly due to their anti-proliferative activity on carcinogenic cells that 9 could lead to mats with anticancer properties. In this regard, Manjumeena et al. developed 10 PVA-based fibers containing a low content of gold nanoparticles (0.1 wt-%), which resulted in a suitable environment for proliferation of Vero cells, thus leading to a viability 11 12 of ca. 90% after 72 h of incubation. The good cell proliferation was attributed to the hydrophilicity of the mats as well as the electrostatic attraction between cell membrane and 13 PVA fibers, regardless of the presence of gold nanoparticles. More interestingly, the 14 15 presence of Au nanoparticles in the electrospun fibers provoked antiproliferative effects in MCF-7 and HeLa cell lines, showing 8 and 9% proliferation, respectively. Additionally, the 16 obtained mats showed excellent antimicrobial performance against a wide range of 17 18 microorganisms, such as B. subtilis, A. coli, K. pneumonia, M. luteus, P. aeruginosa, S. aureus, C. albicans and C. krusei [86]. 19

20

Similarly, McKeon-Fischer *et al.* demonstrated that PLLA/Au nanoparticles could successfully be engineered by the electrospinning technique. However, the resulting fibers did not stimulate the proliferation of rat muscle cells, which was not caused by the presence of gold nanoparticles since parallel studies corroborated the non-toxicity of isolated

nanoparticles at high concentrations (7-21 wt-%). The authors claimed that the cell seeding
on the obtained mats was insufficient to induce proliferation; however, no clear
experimental evidence was provided [87].

4

5 **3.3.2.** Metal oxides

6

7 Nanostructures based on metal oxides are other extensively utilized antimicrobial compounds. The increasing interest in metal oxide nanoparticles mainly arises from their 8 simple and inexpensive synthesis, which is commonly carried out by solution-phase 9 10 methods (sol-gel, co-precipitation, hydrothermal, etc.), affording excellent control over particle morphology. Besides, such nanoparticles are relatively environmentally friendly. 11 12 Regarding the applications, several optical, electrical, magnetic and catalytic properties of metal oxides can be exploited for designing sensors, catalytic systems, (opto)-electronic 13 materials, among others [88-92]. Furthermore, metal oxides have been studied as 14 antimicrobial species when incorporated into (bio)polymeric fibers, especially those based 15 16 on zinc, iron, titanium, and copper.

17

Titanium dioxide (TiO₂) is a well-known multifunctional material with significant UV absorption, antimicrobial, and catalytic properties; therefore, the nanoparticles derived from this metal oxide have been applied in medicine and environment-related areas as UVblocking agents, sensors, photo-catalytic systems, etc. [93]. Currently, there is a trend to produce TiO₂ nanostructures with controlled morphology by electrospinning a polymeric solution containing a Ti-containing source (such as titanium isopropoxide), and subsequently thermally degrade the polymeric matrix [94]. 1

2 Regarding electrospinning for the fabrication of biopolymeric fibers, some researchers have demonstrated the multi-functionality of the corresponding mats with nano-TiO₂ and have 3 also proved the antibacterial activity and potential application in tissue engineering by in 4 vitro approaches. In this sense, Lee et al. took advantage of the well-known 5 6 biocompatibility of poly(vinyl alcohol) for preparing PVA/nano-TiO₂ (2-3 wt-%) 7 composite fibers with excellent antibacterial activity against S. aureus and K. pneumoniae, 8 UV protection, and the ability to degrade formaldehyde, thus evidencing the possibility to 9 develop advanced textile materials [95]. Similarly, PVA-based composite mats with 10 enhanced antibacterial properties were obtained by incorporation of low amounts of chitosan (3-15 wt-%), which is an inherent antibacterial natural polymer, and TiO₂ 11 12 nanoparticles. When comparing fibers with embedded TiO_2 and Ag nanoparticles at the same concentration, the latter nanoparticles exhibited superior antibacterial performance 13 (10% higher with 0.04 wt-% Ag nanoparticles), which could be explained in terms of 14 release feasibility [57]. 15

16

Poly(3-hydroxybutyrate) (PHB) is another interesting natural polymer which strongly 17 18 draws attention for tissue engineering, due to its excellent biocompatibility in the presence of a wide range of medically important cells [96, 97]. Nano-TiO₂ has been incorporated 19 20 into electrospun PHB fibers in order to prevent bacteria adhesion, specifically against E. 21 coli. Korina et al. studied PHB/TiO₂ fibrous materials obtained by mere electrospinning, its combination with electrospraying or impregnation with TiO_2 nanoparticles, thus leading to 22 23 TiO₂-embedded mats for the mere electrospinning technique and TiO₂-coated mats for the electrospinning/electrospraying electrospinning/impregnation 24 or approaches. The

comparative study revealed that all composite fibers completely killed the bacterium, but 1 2 those with TiO₂ the surface (electrospraying/electrospinning on and electrospinning/impregnation) achieved the maximum antibacterial effect in only 30 min, 3 while it took 60 min for fibers containing embedded TiO₂. The main concern when 4 5 preparing PHB or other PHAs fibrous nanocomposites is the preservation of its inherent 6 cell adhesion and proliferation. In this regard, the presence of relatively high content of 7 TiO_2 nanoparticles (5-10%) did not interfere with the proliferation of different cell lines 8 [37, 98].

9

10 Poly(lactic acid) (PLA) and its copolymers with poly(glycolic acid) (PLGA) are biodegradable polyesters with high potential as fibrous materials for medical applications, 11 12 due to their mechanical and thermal properties comparable with other commercially available polyesters, and especially approval by the US Food and Drug Administration [38, 13 52, 99-102]. Concerning PLA-derived electrospun fibers with TiO₂ nanoparticles, it was 14 demonstrated that the combination of electrospinning of PLA and electrospraying of a Ti 15 precursor, followed by subsequent hydrothermal treatment, resulted in an excellent 16 approach to develop PLA-based mats with strong antibacterial activity against E. coli and 17 18 S. aureus at high TiO₂ content (ca. 35 wt-%) [103]. Further studies revealed that PLA/TiO_2 fibers preserved their antibacterial efficiency against S. aureus at relatively low 19 nanoparticle content (1-5 wt-%) without evidence of in vitro cytotoxicity (24-168 h, 20 21 fibroblast cell line) [104].

22

Advanced biomedical devices consisting of two layers, the first one of chitosan/TiO₂
electrospun fibers and the second one of adipose-derived extracellular matrix were recently

reported. These new materials exhibited low percentage of bacterial penetration (*E. coli* and
 S. aureus), due to the synergistic effect of chitosan and TiO₂. Additionally, *in vivo* analysis
 in conjunction with the corresponding histological and immunofluorescence staining
 proved the ability of the obtained mats to heal wounds in only 21 days [105].

5

Variation and/or combination of electrospinning techniques have barely been used for 6 7 designing polymeric fibers with TiO₂, and the few reports are focused on non-medical applications, with the exception of the previously cited works by Korina and Toniatto [45, 8 9 106]. For instance, in 2010 the production of chitosan fibers decorated with TiO_2 was 10 reported. The fibers were obtained by the coaxial electrospinning technique, and the engineered mats were meant for self-cleaning textiles since TiO₂ assisted the degradation of 11 12 pigments [107]. The polymer/TiO₂ fibers derived from coaxial electrospinning have mainly been used in applications associated to energy, photo-voltaic and photo-catalysis, in which 13 14 the polymeric matrices are typically non-biodegradable, and in most of the cases the 15 matrices are further sacrificed by thermal treatment [5, 108-111].

16

Another important inorganic nanomaterial with strong inhibitory and bactericidal effect is 17 18 zinc oxide, which has been utilized for decades. Furthermore, this metal oxide has demonstrated a higher ability to kill various types of bacteria than TiO_2 nanoparticles [28, 19 20 112, 113]. For instance, methicillin-resistant *Staphylococcus aureus* is one of the major nosocomial pathogens, whose proliferation has been inhibited by ZnO nanoparticles with 21 diameter size ca. 100 nm in a major extent than with TiO₂. This behavior resulted even 22 23 more interesting due to the fact that TiO_2 exhibited smaller size (ca. 50 nm), *i.e.* higher surface area, which should potentially lead to higher antibacterial activity; however this 24

behavior was not observed [114]. Moreover, ZnO nanoparticles can open the possibility of
developing materials not only with antibacterial properties, but also with the power to exert
a toxic effect against cancer cells, which has recently been demonstrated by Akhtar *et al.*[115].

5

ZnO nanoparticles have also been incorporated as antimicrobial agents for textiles with 6 7 fiber diameters ranging from 5-10 µm [31, 112]; however, the interest in developing sub-8 micrometric fibers has increased significantly, due to the peculiar features of these fibers, 9 as mentioned in previous sections. With respect to the incorporation of ZnO nanoparticles 10 into biopolymeric fibers for obtaining antibacterial mats, several papers have been published, in which mere electrospinning or combined techniques were implemented, thus 11 12 leading to new insights into the effect of nanoparticles dispersion/distribution on final properties, especially bactericidal activity. Concerning the ZnO-incorporated mats derived 13 14 from mere electrospinning, the matrices have mainly been limited to poly(vinyl alcohol), some natural polymers, namely chitosan, cellulose or gelatin, and aliphatic (bio)polyesters, 15 such as PLA, PCL, and PHAs. 16

17

PVA/ZnO fibers were fabricated by electrospinning of the corresponding PVA solution containing zinc acetate as ZnO precursor (diameter ~240 nm), and they were evaluated as antibacterial materials against both Gram-positive (Methicillin-resistant *Staphylococcus aureus*) and Gram-negative (*Salmonella typhi, Proteus mirabilis Pseudomonas aeruginosa* and *Klebsiella pneumonia*) bacteria. The *in vitro* results indicated good inhibition for *S. aureus*, *S. typhi*, and *P. mirabilis*. However, no antibacterial activity was shown against *P. aeruginosa* and *K. pneumonia*, which was justified in terms of affinity with the bacterium cells and structure of cell wall. The ZnO nanoparticles were positively charged and consequently could interact with the negatively charged cell wall surface of bacteria. Grampositive bacteria have a cytoplasmic membrane and a thick wall composed of a peptidoglycan multilayer. On the other hand, Gram-negative bacteria have complex cell wall structure with a layer of peptidoglycan between the outer membrane and cytoplasmic membrane. The nanoparticles could then more easily attach to the Gram-positive bacteria and cause higher damage compared to the Gram-negative bacteria [116].

8

9 Lubasova et al. also designed PVA/ZnO fibers by conventional incorporation of 10 nanoparticles into the corresponding polymeric solution and subsequent electrospinning. Interestingly, the authors compared the antibacterial performance of ZnO with other metal 11 12 oxides, namely TiO₂, ZrO₂, and SiO₂. A high content of nanoparticles (10 wt-%) was necessary in order to expose them on the PVA-based fiber surface, which was theoretically 13 a pre-requisite for good antibacterial activity. The results confirmed that the incorporation 14 15 of metal oxides conferred antibacterial properties to PVA fibers, and the growth inhibition of the tested bacteria (S. aureus and E. coli) was increased when the nanoparticle content 16 was augmented for all metal oxide nanoparticles. Nonetheless, the antibacterial behavior of 17 18 PVA-based mats depended on the type of nanoparticles, and the ability to inhibit bacteria growth was in the decreasing order as follows: $ZnO > ZrO_2 > TiO_2 > SnO_2$. It is important to 19 20 mention that ZnO inhibited the growth of S. aureus and E. coli in 90% and 100%, respectively, while SnO₂ reduced exclusively the proliferation of S. aureus in 82% [117]. 21 Similar studies were recently carried out by the same authors but using poly(vinyl butyral) 22 as a matrix, and the resulting fibers with ZnO showed superior antibacterial performance 23
than those containing other metal oxide-based nanoparticles (CuO, ZrO₂, TiO₂, SnO₂) and
even AgNO₃ [118].

3

4 PVA-based fibers have also been used as templates for the design of advanced mesoporous materials with combined osteoconduction and antibacterial properties. These materials were 5 obtained by electrospinning of the corresponding polymeric solution with $Ca(NO_3)_2 \cdot 4H_2O$ 6 7 and a stoichiometric amount of $NH_4H_2PO_4$ as hydroxyapatite precursor. On the other hand, 8 zinc acetate was selected as ZnO precursor. After calcination of PVA fibers, novel mesoporous materials based on hydroxyapatite/ZnO were obtained, and they inhibited 9 10 bacterial growth from 20 to 80%, which was directly proportional to the ZnO/hydroxyapatite molar ratio (1/3, 1/5 and 1/10) and depended on the type of bacteria, 11 being S. aureus more susceptible to the presence of ZnO [119]. 12

13

14 Natural polymers have also been exploited for developing electrospun fibers with potential 15 use as antibacterial wound dressings. Chitosan is a well-known polysaccharide which itself provides bacteriostatic and fungistatic activities (i.e. momentarily stops the proliferation of 16 certain microorganisms). In this regard, Wang et al. demonstrated that the addition of ZnO 17 18 nanoparticles into chitosan-based fibers conducted to mats with synergistic antibacterial effect against E. coli and C. albicans, since neat chitosan fibers exhibited a minimum 19 inhibitory concentration (MIC) of 130 and 190 µm/mL for E. coli and C. albicans, 20 respectively, while the composite fibers displayed lower values (110 and 160 μ m/mL for E. 21 coli and C. albicans, respectively) [120]. Cellulose acetate was also used in the form of 22 sub-micrometric fibers with ZnO nanoparticles. The resulting mats were tested in vitro 23 against S. aureus, E. coli, C. freundii and K. pneumonia, and no antibacterial effect was 24

observed for the latter bacterium, which was attributed to differences in cell walls, as
 mentioned above [121].

3

Regarding the biodegradable polyesters, PLA and PHB electrospun fibers with 4 incorporated ZnO nanoparticles have also showed moderate-to-good antibacterial 5 6 performance. Virovska et al. demonstrated the antibacterial activity of PLA fibers with high 7 content of surface-modified ZnO nanostructures (23 wt-%, rod-like with diameter 10-30 nm 8 and 100 nm in length) against E. coli and S. aureus. Moreover, the obtained PLA/ZnO mats 9 showed excellent photo-catalytic properties which were exploited for the degradation of 10 pollutant dyes (methylene blue and reactive red). Interestingly, the mats demonstrated high degradation efficiency after several cycles, thus showing their reusability [41]. 11

12

A parallel study carried out by Rodríguez-Tobías et al. evidenced that electrospun 13 polymeric fibers based on PLA or PHB with low content of smaller ZnO nanoparticles (1-5 14 15 wt-%, 12 nm in diameter) successfully inhibited the proliferation of the same bacterium strains as those examined by Virovska [39, 42, 43, 122]. The incorporation of low ZnO 16 content is an advantageous feature, since it ensures the biocompatibility of this type of 17 18 materials as reported by other authors [123, 124]. Additionally, these research groups have corroborated that the combination of electrospinning and electrospraying techniques 19 20 enhance the antibacterial properties of the resulting mats (the tandem system and the obtained materials are depicted in Figure 6), which was explained by the fact that a larger 21 22 ZnO surface area was directly exposed to bacteria. Such ZnO nanoparticles deposited on polymeric nanofibers were then more prone to act as efficient antibacterial agents, thus 23 potentiating the use of ZnO-coated mats as antibacterial wound dressings. 24



2

Figure 6. Scheme of the equipment used for the electrospinning/electrospraying technique.
The inserted micrographs are the obtained biopolyester fibers coated with (a) 1, (b) 3, and
(c) 5 wt-% of ZnO nanoparticles.

PCL is another important biodegradable polyester whose electrospun composite mats have been exhaustively studied by Augustine *et al.* The authors evidenced that decreasing the ZnO nanoparticle content (60 nm in size) in the polymeric solution up to 1 wt-% reduced the diameter of the resulting PCL fibers (neat PCL fibers exhibited an average-diameter of 2.5 μ m, meanwhile PCL/ZnO fibers was 1.3 μ m). On the contrary, higher nanoparticle contents induced the formation of thicker fibers, which was attributed to changes in

conductivity and viscosity. Regarding the antibacterial properties, the authors discovered 1 2 that significant antibacterial effect could be achieved only when ZnO concentration was higher than 4 wt-%. Furthermore, PCL/ZnO fibers resulted more effective against S. aureus 3 than against E. coli, which was explained by cell wall differences, as discussed in previous 4 sections. Relevant insights into the cytocompatibility of PCL/ZnO fibers have also been 5 provided by Augustine *et al.* who proved that ZnO concentrations lower than or equal to 2 6 7 wt-% led to enhanced fibroblast proliferation compared with neat PCL fibers. The authors 8 carried out further analyses in order to elucidate the effect of PCL/ZnO fibers on the 9 healing of skin wounds. Interestingly, in vivo tests corroborated that PCL/ZnO fibrous mats 10 promoted the proliferation of cells, thus inducing faster healing process in Guinea pig specimens within only 20 days. The reduction of biocompatibility of PCL fibers with 11 12 relatively high concentration of ZnO nanoparticles has been ascribed to higher production of reactive oxygen species which could damage cells, thus inducing inflammation and 13 14 lowering wound healing index [122, 124].

15

Electrospun fibrous coatings consisting of PCL/ZnO (nanoparticles content equal to 1 and 3 16 wt-%) with enhanced biocompatibility for osteoblasts have recently been reported by Kim 17 18 et al. who deposited the mentioned composite fibers on magnesium alloy implants, which showed corrosion resistance and antibacterial properties [125]. Contrasting results about 19 20 biocompatibility of PCL-based fibers with ZnO nanoparticles (irregular morphology with 21 size ranging from 20-150 nm) were reported in 2015 by Münchow et al.. The authors demonstrated that relatively high contents of ZnO nanoparticles (5-30 wt-%) moderately 22 23 reduced the viability of human dental pulp stem cells, and the antibacterial properties

against important periodontal pathogens (*Porphyromonas gingivalis* and *Fusobacterium nucleatum*) were preserved [126].

3

4 It is thus noteworthy that the use of electrospinning in combination with electrospraying for developing antimicrobial (bio)polymeric fibers with ZnO has only been explored in two 5 series of works. Rodríguez-Tobías et al. and Virovska et al. have implemented the 6 7 combination of electrospinning of a polyester solution with simultaneous electrospraying of ZnO suspensions. The former authors have additionally provided relevant information 8 about the influence of ZnO concentration, surface modification, nanoparticle 9 10 distribution/dispersion on mechanical, phase transitions, thermal stability, and photodegradation of the resulting composite polyester mats [39-43, 46]. 11

12

Last, copper is easily available and one of the most used metals for a wide range of 13 applications. Moreover, scientists have well documented the synthesis and properties of 14 CuO nanostructures. Nonetheless, up to date there are few reports related to the 15 incorporation of CuO nanostructures into polymeric fibers by means of electrospinning 16 techniques, and most of them have only addressed the effect of nanoparticles on the final 17 18 fiber morphology, while the ability to confer antimicrobial properties to electrospun (bio)polymeric fibers has scarcely been considered. The limited use of CuO nanostructures 19 20 might be due to their propensity to oxidation reactions, thus changing their overall features 21 [35, 127-129].

22

Among the few research works related to the fabrication of antimicrobial polymer/CuO fibers, that belonging to Haider *et al.* could be particularly mentioned. The authors

evidenced that the presence of a low amount of CuO nanoparticles (0.5 wt-%, 40-100 nm in 1 2 diameter) did not affect the average fiber diameter (ca. 550 nm) of poly(lactic acid-coglycolic acid) (PLGA) fibers. The relatively large size of CuO nanoparticles was enough 3 for their partial exposition on the polymeric fiber surface, thus facilitating the interactions 4 between the studied inorganic antimicrobial nanoparticles with the analyzed bacteria (E. 5 6 *coli* and *S. aureus* strains), and consequently inducing the growth inhibition (>60% for both 7 bacteria). The research work also demonstrated that the antibacterial mechanism was governed by the release of Cu^{2+} ions. With regard to biocompatibility and cell viability, 8 various tests revealed that both PLGA/CuO and PLA/CuO fibers possess suitable 9 10 characteristics for adhesion and proliferation of fibroblastic cell lines, so that the obtained electrospun mats could be envisioned as scaffolds and/or wound dressings with strong 11 12 antibacterial activity [35].

13

14 In another recent report, PLGA was once again preferred for obtaining electrospun 15 scaffolds with embedded CuO nanostructures combined with hydroxyapatite. Composite PLGA ultrafine fibers were engineered via a two-step methodology. First, copper oxide 16 nanostructures were synthesized using a wet chemical method resulting in quasi-spherical 17 18 particles with average diameter of ~7 nm. Then, hydroxyapatite and the as-synthesized CuO nanoparticles were incorporated by mixing with the PLGA solution, and the mixed 19 20 solution was subsequently electrospun. The resulting fibers showed an average diameter (1-21 1.2 µm) larger than that for pristine PLGA, which was attributed to the high content of hydroxyapatite (high solution viscosity). The added nanostructures were well-dispersed and 22 exposed on PLGA fiber surface since electron probe microanalysis showed CuO- and 23 hydroxyapatite-rich maps. A fact revealed for the first time by the authors was the 24

synergistic antibacterial effect (against *E. coli*) of hydroxyapatite and CuO nanostructures;
 however, the governing mechanism was partially elucidated [128].

PHA-based multilayer materials have been extensively studied by Castro-Mayorga et al. 3 The authors have engineered PHBHV copolymer films with electrospun PHAs/CuO fibers 4 5 on their surface, which conferred exceptional antibacterial (Salmonella enterica and 6 *Listeria monocytogenes*) and antiviral (*Murine norovirus*) activity with only 0.05 wt-% of 7 CuO nanoparticles. It was further demonstrated that the obtained materials exhibited good 8 oxygen permeability, and mechanical performance. They were completely disintegrated 9 under composting after 27-30 days, thus making them as promising multilayer films for 10 food packaging [130].

11

12 Dimensional stability in conjunction with good elasticity performance are required features for fibrous materials meant for tissue engineering scaffolds or similar applications, since 13 the receiving tissue could be subjected to constant stresses and/or movement. PVA is a 14 biodegradable and biocompatible polymer which can be mechanically stabilized by 15 crosslinking. For this reason, it was selected by Rezaee et al. for developing novel CuO-16 coated PVA fibers by a three-step process consisting in electrospinning the PVA solution, 17 18 followed by a glutaraldehyde-catalyzed crosslinking reaction, and finally the electrospraying of a colloidal CuO dispersion. The authors optimized the fabrication of 19 20 PVA fibers by varying some important parameters of the electrospinning process, namely polymer molecular weight (viscosity) and flow rate. The effect of crosslinking degree on 21 final morphology of mats was also investigated, and it was revealed an optimal 22 23 concentration of glutaraldehyde in which the fibrous morphology was preserved. Even though the antibacterial efficiency of isolated nanoparticles was excellent, the same 24

property was not determined for the PVA/CuO mats, thus limiting their potential
 application as antibacterial scaffolds [129].

PVA has also been used as sacrificial matrix for electrospun multifunctional composite 3 4 filters based on CuO and ZnO nanostructures, whose elaboration was carried out by electrospinning of a solution containing PVA/copper acetate monohydrate/zinc acetate 5 dehydrate, and a subsequent calcination step. The resulting inorganic CuO/ZnO mixed 6 7 fibers showed smaller diameter (ca. 180 nm) than those composed by PVA/CuO/ZnO (ca. 8 300 nm), which was attributed to PVA removal during the calcination step. Regarding the 9 antibacterial activity, it was demonstrated that S. aureus was more sensitive to CuO-ZnO 10 fibers than E. coli. Significant bactericidal effect was only achieved with 200 µg/mL of composite fibers in the presence of S. aureus, while E. coli exhibited higher resistance since 11 bactericidal effect was observed when 350 µg/mL of composite fibers were used. On the 12 other hand, the composite fibers were highly effective for removal of Congo red dye, thus 13 14 validating the potential use as filters for water purification [131].

15

At this stage, taking into consideration the previous information, it could be inferred that metal oxides based on titanium, zinc, and copper are the main nanoparticles used as antimicrobial agents. However, there is a great interest in employing naturally available metal oxides, and iron oxides are among these types of materials, however their antimicrobial activity is the property least investigated.

21

Iron oxide nanoparticles have mainly been utilized for conferring magnetic properties to electrospun fibers. For instance, a contribution from Rashkov's group was related to the design of PHB fibers embedded or coated with a combination of TiO_2 and Fe_2O_3

nanoparticles. The nanoparticle-embedded PHB fibers were derived from the 1 2 electrospinning of a solution consisting of dissolved PHB and dispersed nanoparticles (2.5 and 10 wt-% for Fe₂O₃ and TiO₂, respectively). The dispersion of the nanoparticles was 3 aided by a low amount of chitosan oligosaccharides (COS, 0.5 wt-%). On the other hand, 4 the nanoparticle-coated PHB fibers were designed by the combination of electrospinning of 5 6 PHB solution and electrospraying of TiO₂/Fe₂O₃/COS dispersion. Regarding the 7 morphology, the composite PHB fibers derived from electrospinning exhibited a significant 8 number of beads, while the materials produced by the combined techniques of 9 electrospinning/electrospraying showed aggregates on the surface. Furthermore, a smaller 10 amount of nanoparticles than initial feed was determined by X-ray photoelectron spectroscopy (XPS), which was ascribed to the formation of a dense layer of the stabilizing 11 12 agent used (chitosan or oligomers). Since the antibacterial activity of identical materials had already been explored [44], the new series of electrospun PHB fibrous mats were used 13 14 as membranes for the degradation of pollutants pigments. The degradation activity was 15 higher for the fibrous materials obtained by the conjunction of techniques, due to higher 16 nanoparticle surface area exposed to the solution containing the pigment [45].

17

Another report related to the design of polymeric fibers with iron oxide nano- and microparticles was recently reported by Moon *et al.* From a solution containing poly(ethylene oxide), sodium alginate and hydrated iron oxide, fibers were elaborated with a diameter ranging from 160 to 475 nm. The authors carried out an exhaustive study of the effect of hydrated iron oxide and sodium alginate on fiber diameter and morphology of the obtained fibers, and the results were related to the increase of electrical conductivity. It was demonstrated that increasing either the content of sodium alginate or iron-based nanoparticles the fiber diameter was reduced and the fibers exhibited a smother surface.
Furthermore, a blend consisted of sodium alginate with hydrated iron oxide showed
excellent ability to removal of *Vibrio vulnificus* (a strong human pathogen) and the authors
claimed the potential use as filters. However, the removal property of the engineered
fibrous materials was not determined, thus misleading the authors conclusions related to
final applications [132].

7

8 **3.4.** Electrospun antimicrobial polymers combined with antimicrobial nanomaterials

9

10 A vast number of researchers have taken advantage of inherently antimicrobial activity of 11 natural or synthetic polymers to develop new materials with stronger biocide effect than the 12 typical polymers used in packaging, filters, medical devices, etc. [133-135]. Nevertheless, 13 the use of electrospinning techniques to generate fibrous materials based on antimicrobial 14 polymers is limited, and the fabrication of the corresponding nanoparticle-containing 15 materials is even more so, therefore, the more relevant reports will be addressed below.

16

Regarding natural polymers, chitosan is well-known as an antimicrobial polymer able to 17 18 kill a broad spectrum of pathogen microorganisms; consequently, it is undoubtedly the most studied. In the case of composite fibrous chitosan containing antimicrobial 19 20 nanomaterials, they have been generally fabricated by blending with other polymers since 21 chitosan possess poor solubility in common solvents. Thus, layered, core-sheath, nanoparticles-decorated and nanoparticles-embedded chitosan-based fibrous materials have 22 been developed by combining techniques. Regarding the nanomaterials, the most 23 representative metallic (silver) and metal oxides (TiO₂, Fe₂O₃ and ZnO) have been used as 24

- antimicrobial additives, whose combination with chitosan exhibited superior antimicrobial
 performance against three microorganisms, *i.e. Escherichia coli*, *Staphylococcus aureus*,
 and *Candida albicans* [10,48,53-55,57,58,74,98,105,120].
- 4

Other naturally occurring or bio-sourced polymers, such as cellulose derivatives, poly(L-5 lysine), poly(hexamethylene guanidine), among others, have also been investigated as 6 7 antimicrobial materials, but only the last two have demonstrated inherent antimicrobial activity and good biocompatibility. They have also been processed by electrospinning 8 9 techniques, however, the corresponding reports have been focused on the preparation of 10 fibrous materials based on blends of these polymers [136-139]. In the case of electrospun cellulose derivatives, antimicrobial activity was conferred by the incorporation of silver or 11 12 zinc oxide nanoparticles, resulting in biocide materials for *Staphylococcus aureus*, Escherichia coli, Spectromyces arenus, Aspergillus flavus, and Citrobacter [32,71-13 14 73,121,140].

15

Electrospun materials based on antibacterial synthetic polymers have been more 16 investigated, which could be attributed to the wide range of raw materials for their 17 18 production, and the versatility in terms of chemical modifications. The main feature of this type of polymers is the presence of ionic groups, by which a destructive interaction occurs 19 20 with the outer cell structure, thus conducing to the microbicidal effect. Quaternized poly[2-(dimethylamino) ethyl methacrylate] (PDMAEMA) is a well-known antimicrobial polymer, 21 and it has been transformed into submicron fibers by the electrospinning technique. 22 23 PDMAEMA with a large variety of macromolecular structures (homo- or copolymers) or compositions (blends) resulted suitable for killing *Escherichia coli*, *Staphylococcus aureus*, 24

and *Pseudomonas aeruginosa* [141-143]. Furthermore, nanoparticles-coated PDMAEMA 1 2 fibers showed higher antibacterial activity than that of neat fibers. It is noteworthy that the coating process has been achieved by a two-step approach: mere electrospinning of 3 PDMAEMA followed by a sputtering process (ZnO and Ag nanoparticles were deposited 4 5 onto the fibers) [144] or immersion in a dispersion containing the nanoparticles (Ag 6 immobilized onto fibers surface by sodium citrate) [142]. The latter approach produced 7 mats with no cytotoxic effect, so that these fibrous materials are promising for tissue 8 engineering.

9

4. Mechanical properties of antimicrobial fibrous composite materials derived from electrospinning techniques

12

Antimicrobial fibrous composite materials engineered by electrospinning techniques are generally utilized as wound dressings and scaffolds in tissue engineering, and as filters for water and/or air decontamination. These devices are subjected to deformation throughout their life-time, so that the evaluation of mechanical properties is of paramount importance [8,50,70,85,131]. In this regard, several authors have investigated the effect of antimicrobial nanomaterial (nanoparticles) content and the electrospinning technique on mechanical properties, mainly modulus and elongation.

20

Our research consortium has recently investigated the mechanical properties of ZnOembedded and ZnO-coated PLA fibers obtained by mere electrospinning and electrospinning/electrospraying tandem process, respectively. The mechanical assessment revealed an optimal ZnO nanoparticles concentration in which the higher mechanical

properties (tensile strength and Young's modulus) were achieved. This optimal value was 1 2 independent of the electrospinning technique; however, ZnO-coated PLA fibrous materials exhibited the highest Young's modulus (38 MPa) and tensile strength (18 N). This 3 mechanical performance was attributed to hydrogen bonds between hydroxyl groups onto 4 de ZnO surface and the carbonyl groups of PLA fibers [43]. The effect of ZnO surface 5 modification (PLA grafts by ring-opening polymerization of D.L-lactide) on mechanical 6 7 performance of electrospun PLA fibers was further investigated, and it was evidenced that the presence of PLA grafts significantly improved the dispersion and interaction with the 8 PLA-based fibers, thus improving not only the aforementioned mechanical properties but 9 10 also the antibacterial efficiency [42].

11

12 Mechanical properties enhancement has also been observed for other polymeric submicron fibers, but containing silver nanoparticles. An et al. demonstrated that wet and dry 13 chitosan/poly(ethylene oxide)-based mats exhibited higher Young' modulus when Ag 14 nanoparticles were embedded in the fibers (49.2 MPa and 322 MPa for neat and composites 15 fibers, respectively). The main difference was observed in wet mats, which suffered more 16 elongation by the presence of water [55]. Other fibrous mats containing silver or its 17 18 precursory species, specifically PVA/chitosan with AgNO₃ or Ag nanoparticles, exhibited tensile strength (9.9 and 15.2 MPa, respectively) suitable for handling related to wound 19 dressings [53]. This increase in mechanical properties was achieved with low silver 20 21 nanoparticles content, typically ≤ 5 wt-%.

22

In the case of composite fibrous antimicrobial materials derived from coaxialelectrospinning, their mechanical performance has scarcely been investigated. In tissue

engineering, coaxial electrospinning has been used only to create fibers with higher
mechanical stability, for example, a biocompatible sheath with a more mechanically stable
polymer. It was demonstrated that the main advantage of this techniques is the relatively
ease to control the morphology of the core-sheath fibers by adjusting the processing
parameters, thus tailoring their mechanical performance [145].

6

7 5. Antimicrobial mechanism of electrospun fibers based on biopolymers and 8 nanoparticles

9

10 The majority of current polymeric medical devices are prone to microorganism colonization, and in the case of electrospun materials, this propensity could be increased 11 12 due to larger surface area available to interact with microorganisms. Adhesion of microorganisms is a multi-stage process which eventually leads to biofilm formation, and 13 this is the predominant mode of microbial life (both in Nature and in disease), due its 14 inherent resistance to antimicrobials. Therefore, when developing antimicrobial electrospun 15 fibers, it is crucial to understand how the cells initially interact with fibrous mats, and more 16 importantly, how this interaction could be avoided, being the incorporation of antimicrobial 17 18 nanoparticles one of the most developed approaches.

19

Currently, there are few studies related to the microbial adhesion on electrospun fibers.
However, from the existing reports, it can be established that the mechanism occurs by
several stages [146-150], which are summarized below:

23

(i) Initially, bacteria are transported near to the mat/fibers surface by long range
 physical forces, such as gravitational and Brownian forces. In addition to this
 passive transport, controlled-like motion is involved in the transport of bacteria
 species having proteinic organelles called flagella.

5

Then, the attraction between microorganism and electrospun fibers surface takes 6 (ii)7 place by means of physico-chemical interactions, which can conduce to reversible and irreversible adhesion, and they are influenced by the surface 8 9 chemistry and the fibers/mat morphology. Physico-chemical interactions are 10 basically electrostatic forces such as van der Waals and hydrogen bond formation. The former leads to a reversible microbial adhesion and the latter 11 leads to irreversible microbial adhesion. Moreover, reaction between 12 microorganism surface and fiber surface might occur, thus provoking 13 irreversible adhesion. 14

15

(iii) After anchoring on the material surface (electrospun fibers), the microorganism
 proliferate and synthesize the biofilm matrix (composed by exopolysaccharides,
 proteins, lipids, among others compounds depending on the microorganism).
 Proliferation is accomplished by cloning the cells which lead to formation of
 colonies and eventually enhance the spreading of the surface.

21

(*iv*) Finally, biofilm maturation can be achieved and subsequent detachment occurs.
 Mature biofilms are complex microorganism communities which possess a tri dimensional structure that allows the access to nutrients and oxygen also

2

protecting against hostile surroundings. It is noteworthy that despite common characteristics, biofilms differ considerably between species and strains.

3

4 Considering the microbial adhesion mechanism explained above, either stage from I to IV should be circumvented. The previously described metallic or metal oxide nanoparticles 5 can meet this aim, through a well-known mechanism based on the attack of microorganisms 6 7 by the metallic ions released from nanoparticles and/or the formation of reactive oxygen species (ROS). Another mechanism based on the absorption of nanoparticles has also been 8 9 suggested, however, the experimental evidences have concluded that the ions released 10 and/or ROS formation are behind the biocide activity; therefore, this mechanism is explained below. 11

12

Once the microorganism approximated to polymeric fiber surface, the presence of moisture 13 from different potential sources (environment, growth medium, adsorbed on polymer 14 and/or nanoparticle surfaces, etc.) promotes the corrosion and/or dissolution of 15 nanoparticles present in the fiber surface region, and eventually the production, release, and 16 diffusion of ions/ROS out of the polymeric matrix. Taking into account this stage of the 17 18 antimicrobial mechanism, the electrospinning techniques should be used for developing polymeric nanofibers with nanoparticle-rich surface, since that is a critical concern in order 19 20 to enhance the production of the ionic or radical species. As largely illustrated in previous 21 sections, coaxial electrospinning or the combination of electrospinning and electrospraying 22 could be used to achieve this characteristic fiber morphology. It is important to consider 23 that the corrosion/dissolution process, ions/ROS release, and the attack to microorganism can be changed by several factors, such as pH variation, radiation, porosity of fiber surface, 24

- variation of the microorganism-polymer affinity, among others; for each specific polymer microorganism system, which is out of the scope of this review.
- 3

4 Another important factor that potentially influences the antimicrobial properties of fibers derived from electrospinning techniques is their morphology, mainly the fiber average 5 diameter and the mat porosity. Abrigo et al. have reported a comprehensive study to 6 7 evaluate the growth behavior of E. coli, P. aeruginosa, and S. aureus on polystyrene micro-8 and nano-fibers [151]. It was demonstrated that the bacteria adhered to fibers regardless the 9 average fiber diameter, but they preferentially adhered to fibers with diameter similar to 10 bacteria size. Furthermore, the bacteria shape also influenced the adhesion to electrospun 11 fibers, since round bacterium (S. aureus) evidenced higher adhesion and growth that rod-12 like bacteria (E. coli and P. aeruginosa) [151]. These findings could be exploited to increase the antibacterial activity of neat or composite fibrous materials and control the 13 14 adhesion and proliferation of tissue cells [152].

15

Regarding the porous morphology typically exhibited in materials engineered by 16 electrospinning techniques, it has been evidenced that fibrous wound dressings with 17 18 porosity higher than 60 % were suitable for wound healing, since they allowed for permeation and diffusion of oxygen from air to skin [1]. Additionally, one such porous 19 morphology acted as a barrier to some microorganisms as long as the interfiber average size 20 was smaller than the dimensions of the microorganism involved, this antimicrobial 21 22 mechanism being well-recognized as sieve effect. It is noteworthy that sieve effect by itself is not highly effective, while the incorporation of nanoparticles has proven to be a 23 24 coadjutant approach [153].

Fiber porosity is another factor with potential repercussions in the antimicrobial activity of 1 2 materials obtained by electrospinning techniques. It has been demonstrated that the more heterogeneous the surface of polymeric substrates, the higher the surface area, thus 3 increasing the sites for cell attachments [154]. Therefore, it is important to control the 4 smoothness of fibers to avoid microorganism adhesion and proliferation, and it is well-5 6 reported that this morphological feature can be minimized by the selection of solvents with 7 relatively low volatility [155,156]. In the case of composite fibrous materials meant for 8 antimicrobial devices, it is of paramount importance to achieve excellent distribution of antimicrobial nanomaterials, since agglomeration of them could promote the formation of 9 10 fiber defects, which could be considered as sites of adhesion for microorganisms 11 [39,42,43].

12

13 6. Conclusions and future prospects

14

Electrospinning and related techniques, namely tandem electrospinning/electrospraying or 15 coaxial electrospinning, have demonstrated versatility for developing polymeric fibrous 16 materials with strong antimicrobial properties conferred by means of the presence of 17 18 inorganic nanostructures. The incorporation of metallic silver and gold nanoparticles as well as metal oxides based on titanium, zinc, copper and iron, is the most developed 19 20 approach. These inorganic compounds in the nano-metric regime have demonstrated strong 21 antimicrobial properties, and additionally, other relevant properties can be modified or conferred, such as mechanical, magnetic, catalytic, and optical properties, thus increasing 22 23 the possibility to develop multi-functional fibrous materials with wider application areas.

24

Researchers have focused their efforts on the investigation related to the variation or 1 2 combination of electrospinning-related techniques, which have yielded important findings 3 related to the influence of nanoparticle distribution/dispersion in electrospun 4 (bio)polymeric fibers. It was demonstrated that electrospinning/electrospraying and coaxial electrospinning are suitable approaches for designing (bio)polymeric fibers with higher 5 antibacterial activity than that of mats fabricated with mere electrospinning, which is due to 6 7 higher surface area prone to interact with pathogenic microorganisms. However, in the case of polymer/Ag composite fibers, mere electrospinning of the corresponding polymeric 8 9 solution containing AgNO₃ could also lead to fibers with Ag-rich surface, because of ion migration, which can subsequently be reduced to Ag^0 . 10

11

Regarding the biocompatibility of the reported (bio)polymer/nanoparticles mats, *in vitro* investigation has demonstrated that low contents (typically lower than 5 wt-%) of metallic and metal oxide nanostructures, especially Ag, TiO₂, and ZnO, do not exhibit toxicity against medically important cell strains, such as mesenchymal human cells, fibroblasts, and osteoblasts. On the other hand, *in vivo* tests have been limited to skin healing, showing that the fibrous materials can promote a faster wound healing in the same range of nanostructure concentration than that for *in vitro* tests.

19

Despite the major findings about antimicrobial electrospun composite fibers, there is currently a need to evaluate the toxicity against more microorganisms, since most of the analyses have been restricted to two model bacteria, *i.e. Escherichia coli* and *Staphylococcus aureus*. In sharp contrast, few or no information related to fungi, viruses and other dangerous bacteria has been published. It could also be inferred that further *in* *vivo* studies have to be carried out in order to provide more realistic findings and medical
 prospections.

3

Another challenging issue is to carry out further studies related to the use of coaxial 4 electrospinning and the combination electrospinning/electrospraying techniques for the 5 6 fabrication of a large variety of antimicrobial composite polymeric fibers. It is also 7 necessary to develop novel or adapted equipment that enable the fabrication of electrospun 8 sub-micrometric fibers in a larger scale in order to achieve cheaper, sustainable, and massive commercialization of obtained mats. In this regard, alternative techniques also 9 10 based on electro-hydrodynamic principles have been developed, such as needle-less electrospinning [157, 158], multi-jet electrospinning [159, 160] and portable 11 12 electrospinning [161]. In addition, sub-micrometric polymeric fibers have also been obtained by an interesting process denominated Force-spinning[®], which uses centrifugal 13 14 force for promoting the flow of the polymeric solution or melted polymer [162-164].

15

The aforementioned contributions about the incorporation of antimicrobial nanomaterials, their dispersion, the control of fiber morphology, the evaluation of antimicrobial performance, *in vitro* and *in vivo* studies, and so on, could foster faster development and application of composite (bio)polymeric electrospun fibers as tissue engineering scaffolds and wound dressings.

21

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