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Julie Schmitt, Sébastien Jenni, Angélique Sour, Valérie Heitz, Frédéric Bolze, et al.. A Porphyrin Dimer–GdDOTA Conjugate as a Theranostic Agent for One- and Two-Photon Photodynamic Therapy and MRI. *Bioconjugate Chemistry*, 2018, 29 (11), pp.3726-3738. 10.1021/acs.bioconjchem.8b00634 . hal-02071234

HAL Id: hal-02071234

<https://hal.science/hal-02071234>

Submitted on 24 Nov 2021

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A Porphyrin Dimer-GdDOTA Conjugate as a Theranostic Agent for One- and Two-Photon Photodynamic Therapy and MRI

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Abstract A molecular theranostic agent designed for photodynamic therapy (PDT) treatment in the near-infrared and for imaging tissue tumors with magnetic resonance imaging (MRI) is reported. It consists of a linear π -conjugated Zn(II)-porphyrin dimer linked at each extremity to a GdDOTA-type complex. This agent has shown very promising potential for PDT applications with good singlet oxygen generation in DMSO and high linear absorption in the near-infrared ($\lambda_{\max} = 746$ nm, $\varepsilon \approx 10^5$ M⁻¹cm⁻¹). Moreover, this molecule has a propensity for two-photon excited PDT with high two-photon cross-sections (≈ 8000 GM in 880-930 nm range), which should allow for deeper tumor treatments and higher spatial precision as compared to conventional one-photon PDT. Regarding the MRI contrast agent properties, the molecule has shown superior relaxivity (14.4 mM⁻¹s⁻¹ at 40 MHz, 298 K) in comparison to clinical contrast agents and ability to be internalized in cells, thanks to its amphiphilic character. Irradiation of HeLa cells using either one-photon (740 nm) or two-photon excitation (910 nm) has led in both cases to important cell death.

■ Introduction

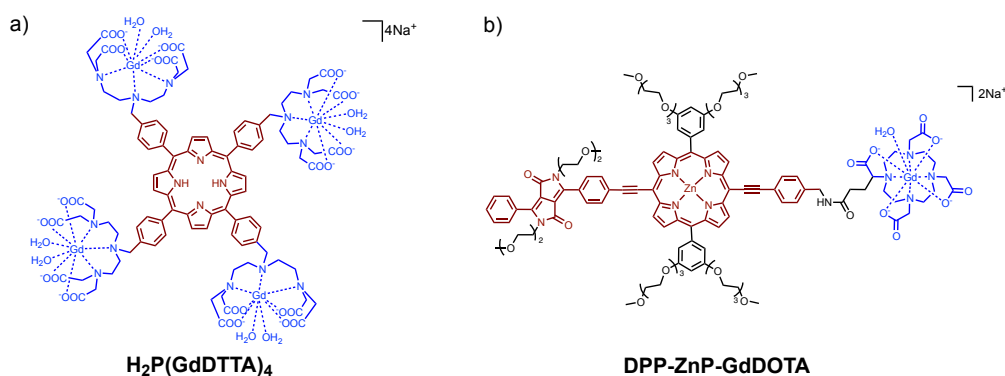
Photodynamic therapy is an appealing therapy classified as non-invasive and intrinsically selective since a light source targeting the disease tissue is used to activate a photosensitizing drug.^{1,2} Following light absorption, the drug will interact from its triplet excited state with surrounding oxygen *via* energy transfer to generate singlet oxygen or *via* electron transfer to generate reactive oxygen species that react with biological substrates, both processes leading to

cell death. PDT is particularly suited to cure early-stage cancers and localized tumors. It has no long-term side effects and can be operated repeatedly without development of treatment resistance. It is therefore also used as an adjuvant therapy to chemotherapy or radiotherapy. The efficiency of PDT is strongly dependent on (i) the accessibility of the photosensitizer to light exposure and (ii) the concentration of the photosensitizer inside tumor tissues and cells. Light excitation is currently performed in the visible region where the clinically used porphyrin-based photosensitizers mainly absorb. However, irradiation performed below 700 nm is far from being optimal for PDT, since these photons are absorbed and scattered by living tissues and cause photodamage to healthy tissue. In addition, these phenomena induce limited tissue penetration, restricting the use of PDT to treat superficial and interstitial areas that are directly accessible by a light source. To overcome these limitations, various chromophores related to porphyrins, chlorins or expanded porphyrins with long-wavelength absorption bands in the optical therapeutic window (700-1000 nm) were developed.³⁻⁶ Lutrin®, a lutetium texaphyrin ($\lambda_{\text{max}} = 732$ nm) has reached clinical trials⁴ and Redaporfin®, a halogenated bacteriochlorin⁶ ($\lambda_{\text{max}} = 749$ nm) and Tookad®, a Pd(II) bacteriopheophorbide^{7, 8} ($\lambda_{\text{max}} = 746$ nm) have recently been approved. An alternative and emerging strategy is to take advantage of two-photon absorption (TPA), a non-linear optical process displayed by some specific π -conjugated molecules capable of absorbing simultaneously two low energy photons. With respect to one-photon absorption, TPA occurs at the focal point of the laser thus increasing the spatial confinement of the excited photosensitizer and the 3D spatial resolution of the treatment.⁹ Several reports have shown promising results with TP-PDT based on molecular systems¹⁰ or on nanomaterials.¹¹⁻¹⁴

The association of an imaging modality to PDT should optimize the PDT protocol by imaging the drug accumulation over time and assessing the outcome of the therapy. The improvement of the therapy by combination of an imaging and a therapeutic agent within one molecule corresponds to the theranostic approach and it represents a major challenge in the development of personalized medicine.¹⁵⁻¹⁷ The fluorescence of the PDT photosensitizer has been used to track its localization¹⁸⁻²⁰ but good PSs are not expected to be strong emitters since singlet emission competes with intersystem crossing and thus with singlet oxygen production. Otherwise, photobleaching, autofluorescence from the tissue and scattered photons from the light excitation beam can also lead to misleading interpretation of the detected emission and prevent quantitative evaluation of the drug concentration in the diseased tissues.²¹ An alternative is to associate a fluorescent agent^{22, 23} or probes detectable in other imaging modalities to the PS.^{24, 25} MRI is particularly well suited to follow the PDT at every stage.²⁶⁻³¹ It is minimally

invasive, uses non-ionizing radiation, provides three-dimensional anatomical images with resolution on the millimeter scale and also provides *in vivo* physico-chemical information of soft tissues.³²⁻³⁴ MR image contrast relies on the water density and on the relaxation time of water protons in tissues, which is strongly affected by their environment. It is often enhanced by administration of contrast agents based on stable paramagnetic gadolinium complexes that reduce the relaxation rates of water protons in their vicinity. The chemical association of an MRI contrast agent with a PDT drug is expected to bring synergetic effects for imaging the drug localization as (i) the lipophilicity of the PDT sensitizer will favor contrast agent internalization in tumor cells and allow the uptake of 10^8 - 10^9 gadolinium ions per cell which is detectable on MR images,³⁵ (ii) the increased size of the conjugate should enhance relaxivity, and (iii) the contrast enhancement in the area to be treated will be proportional to the concentration of the contrast agent-drug conjugate allowing for drug quantification.

Until now, only a few molecular systems for one-photon PDT and MRI have been evaluated as theranostic agents.³⁶⁻⁴⁷ In this context, we have reported a water-soluble molecular system consisting of a porphyrin linked to four gadolinium complexes [GdDTTA] (DTTA: diethylenetriamine-*N, N, N''*, *N'''*- tetraacetate), **H₂P(GdDTTA)₄** (Scheme 1).⁴⁸ Thanks to its high rigidity, this Gd(III)-based contrast agent displays the highest relaxivity among medium-size molecular systems. Additionally, good phototoxicity results were also obtained upon visible light excitation showing the potential of this molecule as theranostic agent. To access to deeper PDT treatment while imaging with MRI, we have designed a π -delocalized porphyrin-diketopyrrolopyrrole conjugate attached to a [GdDOTA] complex (DOTA: 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetate), **DPP-ZnP-GdDOTA** (Scheme 1). We have demonstrated the ability of **DPP-ZnP-GdDOTA** to behave as a two-photon photosensitizer for PDT with excitation in the near infrared (910 nm) and as an MRI agent, even at low micromolar incubation concentration.⁴⁹



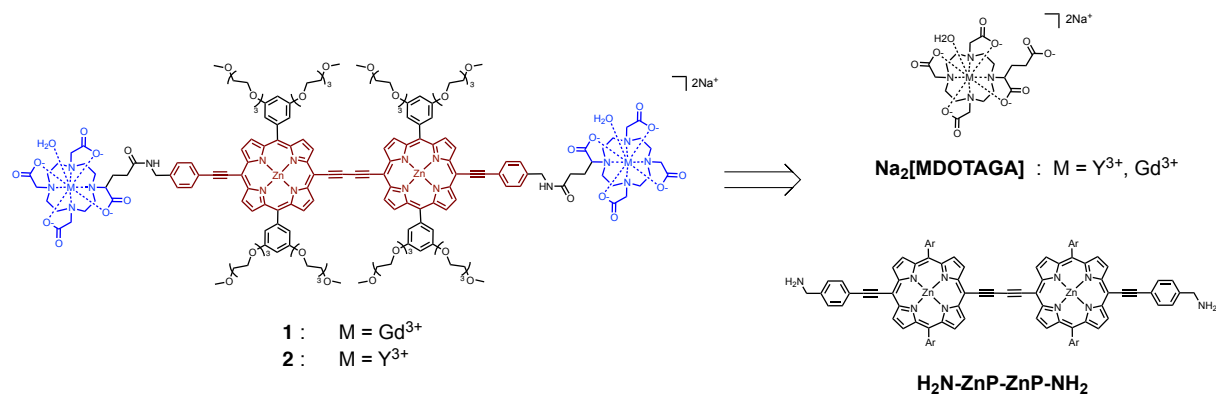
Scheme 1. Structures of two previously reported theranostic agents (a) **H₂P(GdDTTA)₂**⁴⁸ and (b) **DPP-ZnP-GdDOTA**⁴⁹.

Herein, we report a novel molecular theranostic system, **1**, consisting of a linear π -conjugated Zn-porphyrin dimer linked at each extremity to a [GdDOTA]-type complex. The two Gd(III) complexes endow the conjugate with MRI detection capabilities whereas the Zn(II) porphyrin dimer is designed to perform efficient one- and two-photon PDT in the optical therapeutic window. The physicochemical and photophysical properties of this bifunctional agent are evaluated regarding its MR imaging and photosensitizing potential. Cellular internalization of the molecule is assessed as well as its dark toxicity and phototoxicity following one- or two-photon excitation.

■ Results and discussion

Design, synthesis and characterization of the compounds.

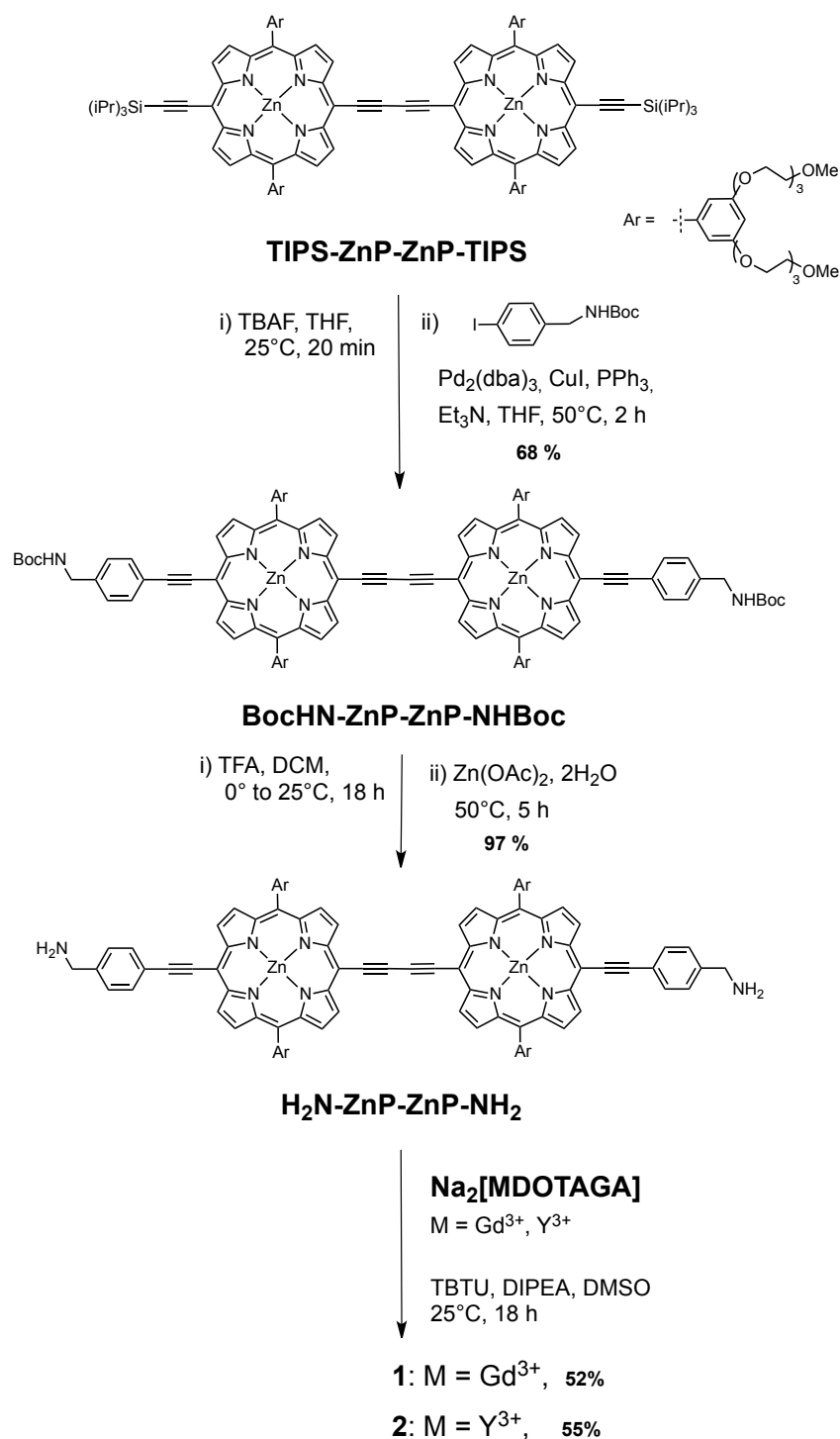
The chemical structures of the targeted bifunctional conjugate **1** and of the related diamagnetic conjugate **2**, which incorporates [YDOTA] complexes are shown in Scheme 2. Compound **2** was prepared to optimize the final coupling step, since **2** unlike **1**, can be characterized by proton NMR. The target molecule **1** consists of a central π -conjugated porphyrin dimer designed as a two-photon PDT photosensitizer with two peripheral gadolinium(III) complexes [GdDOTA] which provide the imaging capability. Hydrophilic anionic or cationic butadiyne-linked Zn-porphyrin dimers developed in the group of Anderson have shown high efficiency as two-photon sensitizers for *in vivo* PDT.^{50,51} The highly hydrophobic character of the butadiyne-linked porphyrin dimer core represents a challenge for water solubilization. Therefore, two *meso* phenyl groups of the porphyrins were decorated with hydrophilic triethylene glycol chains. In addition, the two hydrophilic Gd(III) complexes in **1** also enhance water solubility of the final conjugate, indispensable for biological applications. The synthesis of both conjugates is based on a convergent approach and involves the coupling of the benzylic amine groups of the zinc(II)porphyrin dimer precursor **H₂N-ZnP-ZnP-NH₂** with the non-coordinating carboxylate arm of the DOTA-based complex, **Na₃[MDOTAGA]** (DOTAGA : 1,4,7,10-tetraazacyclododecane-1-glutaric-4,7,10-triacetate).



Scheme 2. Synthetic strategy used to prepare the theranostic system **1** and the related diamagnetic complex **2**.

The synthesis of the precursor **H₂N-ZnP-ZnP-NH₂** (see Scheme 3) starts from the zinc(II)-porphyrin dimer **TIPS-ZnP-ZnP-TIPS**⁵² whose synthesis was inspired by a related compound reported by Anderson *et al.*⁵³ After classical cleavage of the *triisopropyl* silyl groups using tetrabutylammonium fluoride (TBAF), the air sensitive intermediate **H-ZnP-ZnP-H** was submitted to Sonogashira coupling reaction with a *tert*-butoxycarbonyl (Boc)-protected 3-iodobenzylamine.⁵⁴ **BocHN-ZnP-ZnP-NHBoc** was obtained in 68% yield after purification by silica gel column chromatography. Both Boc-protecting groups were then removed using trifluoroacetic acid. This reaction led to partial demetalation of the porphyrin, which was remetalated with Zn(OAc)₂·2H₂O affording **H₂N-ZnP-ZnP-NH₂** in almost quantitative yield.

Na₂[GdDOTAGA] and its diamagnetic analogue **Na₂[YDOTAGA]** were readily obtained by complexation of GdCl₃ and YCl₃, respectively, with DOTAGA-anhydride as reported in the literature.⁵⁵



Scheme 3. Synthetic route to compounds **1** and **2**.

The final hetero-tetranuclear complexes **1** and **2** were obtained by a careful choice of the amide coupling reaction conditions in order to avoid demetalation reactions. The activated ester was obtained by dissolving **Na₂[MDOTAGA]** in dry dimethylsulfoxide (DMSO) at 60°C followed by addition of the activating agent *O*-(benzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium tetrafluoroborate (TBTU) and a base *N,N*-diisopropylethylamine (DIPEA). A solution of **H₂N-**

ZnP-ZnP-NH₂ in DMSO was then added and the reaction mixture was stirred overnight at room temperature. The desired compounds **1** and **2** were isolated by precipitation after addition of dichloromethane (DCM) and thorough washing with DCM, acetone and water. Both compounds were characterized by high-resolution mass spectrometry. The isotopic profile of the molecular ionic species $[M-2H_2O]^{2+}/2$ was detected for both compounds and was in accordance with the calculated profiles (see Figures S9 and S10). Inductively coupled plasma-atomic emission spectroscopy (ICP-AES) confirmed the 1:1 ratio of Gd(III) and Zn(II) ions in compound **1**. Compound **2** was further characterized with 1D and 2D ¹H NMR spectroscopy in DMSO-*d*₆ (see Figures S7 and S8).

Relaxivity measurements.

The capacity of a Gd(III) complex to reduce the water proton relaxation times is expressed by its relaxivity. Relaxivity is defined as the paramagnetic enhancement of the longitudinal water proton relaxation rates and it is a direct measure of the MRI efficiency of the agent. MRI agents are typically characterized by nuclear magnetic relaxation dispersion (NMRD) profiles, which represent the magnetic field dependence of the relaxivity and therefore are useful to assess the underlying relaxation mechanisms. Relaxivity is affected by a number of microscopic parameters of the Gd(III) chelate, the most important being the hydration number (*q*), the exchange rate of the coordinated water with the bulk (*k_{ex}*), the rotational dynamics of the chelate and the parameters describing electron spin relaxation (*τ_v* and *Δ²*). The relation between these parameters and relaxivity are defined by the Solomon-Bloembergen-Morgan (SBM) theory of paramagnetic relaxation.³³

NMRD profiles have been recorded for **1** in water containing 2 % pyridine at pH 6.6 (the presence of pyridine ensures the solubility in the mM range) and at two different temperatures (25 and 37 °C) in the range of 10 kHz-600 MHz proton Larmor frequency (Figure 1 and Table S1). They have been interpreted in the frame of the SBM theory (see Supporting Information for equations and details of the fit) and the best-fit parameters are given and compared to those for analogous systems in Table 1. The hydration number was considered to be 1, and the water exchange parameters have been fixed to the values reported for [GdDOTA].⁵⁶ Data were analyzed only above 6 MHz where the validity of the SBM theory to describe electronic relaxation and rotational dynamics of slowly rotating objects is justified. The most important information that we can obtain from this analysis concerns indeed the rotational dynamics of the conjugate. It was described in the fit with the Lipari-Szabo formalism that allows separating

fast local and slow global motions based on the field dependency of longitudinal proton relaxation.⁵⁷ This analysis yields a local rotational correlation time, τ_l characterizing fast local motions of the Gd(III)-water proton axis, and a global rotational correlation time, τ_g related to the slow global motion of the system. The degree of spatial restriction of the local with respect to the global motion is expressed by the generalized, model independent order parameter S^2 . Indeed, the high field bump of the profiles indicate slow rotational dynamics, which is an evident consequence of the large size of the molecule. The photophysical measurements pointed to some aggregation in a 1% pyridine/water mixture. Therefore, in order to check the possible presence of aggregation phenomena, the relaxivities at 40 MHz (and 25°C) were also measured at six different concentrations covering a range of 0.1-1 mM, and they were found identical. At this frequency, the relaxivities are particularly sensitive to rotational dynamics, thus the identical values suggest that aggregation is either not important or at least does not change within this concentration range. The global rotational correlation time, $\tau_g^{298} = 1360$ ps, is only slightly higher than that of the metallostar $\{\text{Fe}[\text{Gd}_2\text{bpy}(\text{DTTA})_2(\text{H}_2\text{O})_4]_3\}^{4+}$ compound ($\tau_g^{298} = 930$ ps) which is a discrete molecular MRI agent of similar molecular weight, bearing six [GdDTTA] complexes around an iron(II)-tris(bipyridine) core (Scheme S1). This difference in τ_g^{298} of **1** can be indeed ascribed to the very different topology of the two molecules (the metallostar is more spherical while **1** is more elongated with a rigid planar core). Therefore, based on the value of the global rotational correlation time, aggregation seems to remain limited under our experimental conditions.

The parameters governing the relaxivity of **1**, of the metallostar and of two theranostic agents reported by us are gathered in Table 1. In comparison to the other porphyrin-conjugated Gd(III) complexes and considering the large and rigid bis-porphyrin core of the molecule, we could expect a higher relaxivity for **1**. Nevertheless, for **DPP-ZnP-GdDOTA**, the high relaxivity was related to a strong aggregation in aqueous solution, while for the tetramer **H₂P(GdDTTA)₄**, the bishydrated nature of the Gd(III) chelate and a remarkable rigidity of the whole conjugate were responsible for the high relaxivity. Despite the rigid bis-porphyrin core, the two peripheral Gd(III) chelates in compound **1** experience important flexibility, as it is indicated by the low value of the generalized order parameter, $S^2 = 0.3$. This flexibility can then account for the limited increase in relaxivity, which is nevertheless around 4-fold with respect to clinically used MRI agents. Moreover, in the presence of bovine serum albumin (42.7 g.L⁻¹), the relaxivity measured at 40 MHz increases considerably from 14.4 to 29.7 mM⁻¹s⁻¹ (298 K) or from 11.8 to 26.6 mM⁻¹s⁻¹ (310 K), indicating strong albumin binding of the conjugate. The ability of our

theranostic agent for non-covalent binding to this blood pool protein should increase its circulation time in the blood and favour its tumoral accumulation.

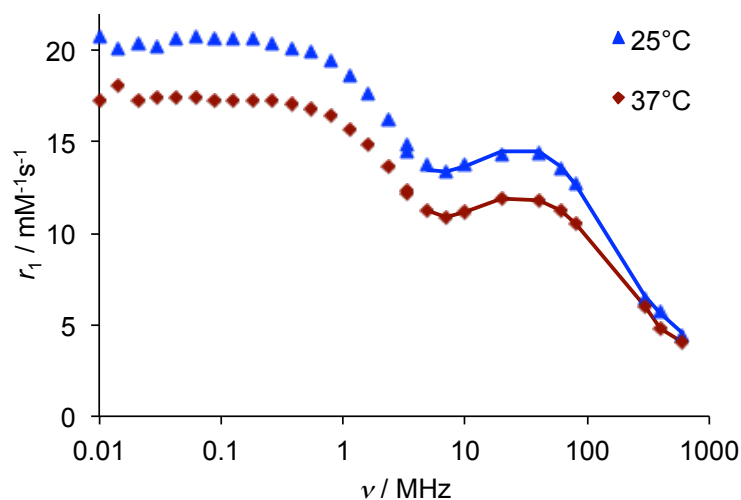


Figure 1. ^1H Nuclear Magnetic Relaxation Dispersion (NMRD) profiles for **1** (0.81 mM) at 25°C (\blacktriangle) and 37°C (\blacklozenge) in H_2O containing 2 % pyridine, at pH 6.6. The continuous lines represent the fitted curves according to the SBM theory.

Table 1. Best-fit parameters obtained from the fitting of the ^1H NMRD profiles to the SBM theory, including the Lipari-Szabo approach to describe internal flexibility.

Parameters	1 in H_2O + 2% pyridine	$\text{H}_2\text{P}(\text{GdDTTA})_4$ in H_2O^b	[DPP-ZnP- GdDOTA] in H_2O^c	$\{\text{Fe}[\text{Gd},\text{bpy}(\text{DTT})_2(\text{H}_2\text{O})_2]\}_n$ in H_2O^d
M_w (g.mol $^{-1}$)	3913	2761	2471	3744
r_1 (mM $^{-1}$.s $^{-1}$; 40 MHz, 25°C)	14.42	48.59	18.61	33.6
q^a	1	2	1	2
k_{ex}^{298} (10 6 s $^{-1}$) ^a	4.1	8.3	4.1	7.4
ΔH^\ddagger (kJ.mol $^{-1}$) ^a	49.8	40	49.8	41.3
E_l (kJ.mol $^{-1}$)	26 \pm 5	40	40	-
τ_l^{298} (ps)	207 \pm 14	94	245	190
E_g (kJ.mol $^{-1}$)	25 \pm 3	29	14	-
τ_g^{298} (ps)	1360 \pm 140	1950	2640	930
S^2	0.30 \pm 0.03	0.60	0.26	0.6

^a fixed in the fit; ^b ref ⁴⁸; ^c ref ⁴⁹; ^d ref ⁵⁸

One-photon photophysical characterization.

Linear absorption, emission and singlet oxygen generation properties of compound **1** were evaluated in order to derive important parameters for both one- and two-photon PDT applications. The characterization has been performed in two polar solvents, dimethyl sulfoxide (DMSO) and H₂O, the latter added with 1% either of pyridine or DMSO to enhance solubility. The absorption spectra are shown in Figure 2 and relevant parameters are collected in Table 2. The spectrum in DMSO is dominated by the absorption of the porphyrin units and is red-shifted, by 20 nm for the Soret band and by *ca.* 10 nm for the Q-bands, with respect to that of a simpler Zn-porphyrin precursor.⁵⁹ A marked splitting of the Soret band is also observed. These features can be ascribed to the extended conjugation *via* acetylene bridges and to excitonic interactions between the two units.⁶⁰⁻⁶² The spectra in aqueous solutions are broader and less intense than in DMSO, indicating moderate aggregation phenomena.

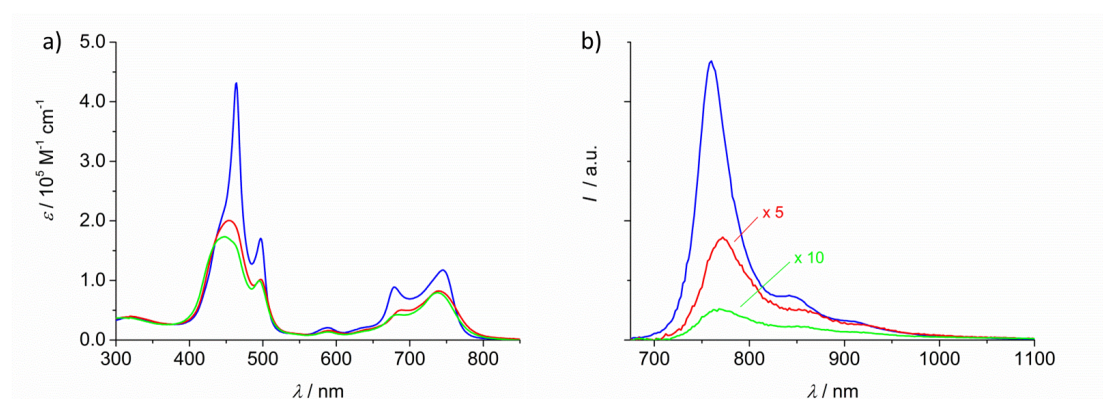


Figure 2. a) Absorption spectra of compound **1** in DMSO (blue), H₂O containing 1% pyridine (red) and H₂O containing 1% DMSO (green). b) Corrected emission spectra of isoabsorbing solutions of **1** in DMSO (blue), H₂O containing 1% pyridine (red) and H₂O containing 1% DMSO (green) at 502 nm ($A_{502} = 0.122$). The red and green spectra are multiplied by a factor 5 and 10, respectively.

Table 2. Absorption and luminescence data of compound **1** at room temperature.

	$\lambda_{\text{max,abs}} / \text{nm}$ ($\epsilon / 10^5 \text{ M}^{-1} \text{ cm}^{-1}$)	$\lambda_{\text{max,em}} / \text{nm}^a$	ϕ_n^b	τ / ns^c	ϕ_{Δ}^d
DMSO	464 (4.32), 497 (1.71), 679 (0.89), 746 (1.17)	760, 842, 912 sh	0.21	0.98	0.36
H ₂ O + 1% pyridine	454 (2.01), 497 (1.02), 689 (0.50), 740 (0.82)	772, 856, 918 sh	1.9×10^{-2}	0.99	0.29
H ₂ O + 1% DMSO	447 (1.73), 495 (0.99), 683 (0.42), 738 (0.80)	768, 850, 912 sh	3.2×10^{-3}	0.82	0.21

^a From corrected emission spectra. ^b Fluorescence quantum yields, measured with reference to TPP (tetra-phenyl-porphyrin) in aerated toluene as a standard. ^c Fluorescence lifetimes,

excitation at 465 nm.⁴ Singlet oxygen production quantum yields, see the Experimental Section for details.

Fluorescence spectra of **1** in DMSO and aqueous solutions are displayed in Figure 2b and the luminescence data are summarized in Table 2. The spectra extend in the near infrared and show a red shift and broadening when moving from DMSO to H₂O. The notable fluorescence quantum yield in DMSO (0.21) decreases from one to two orders of magnitude in aqueous solutions, to be ascribed to the existence of aggregation phenomena in aqueous environment. An estimation of the size of the aggregates was obtained from dynamic light scattering (DLS) experiments (Table S2).

Excitation spectra collected in the different solvents are in good agreement with the absorption spectra (Figure S11), confirming the nature of the observed emission. It can be noticed that the fluorescence decay of **1** in H₂O is mono-exponential, with a lifetime similar to that measured in DMSO (Table 2).

Singlet oxygen production quantum yields have been measured in DMSO and aqueous solutions by means of indirect methods (see the Experimental Section for details). [Note1] A notable value of 0.36 is obtained in DMSO (Figure 3), that is very close to the ϕ_{Δ} value of 0.38 obtained for the parent PS **TIPS-ZnP-ZnP-TIPS**⁵² and near values of 0.29 and 0.21 are measured in H₂O containing 1% of pyridine and 1% of DMSO, respectively (Table 2), in accordance with moderate aggregation of **1** in the latter environments. The indirect measurement of ϕ_{Δ} also evidenced the photostability of compound **1** upon irradiation in the visible range up to 21 min irradiation at 680 nm in DMSO (Figure 3) and up to 120 min irradiation at 516 nm in aqueous environments (Figure S13). The bifunctional conjugate **1** can be thus considered a promising photosensitizer for PDT applications *in vivo*.

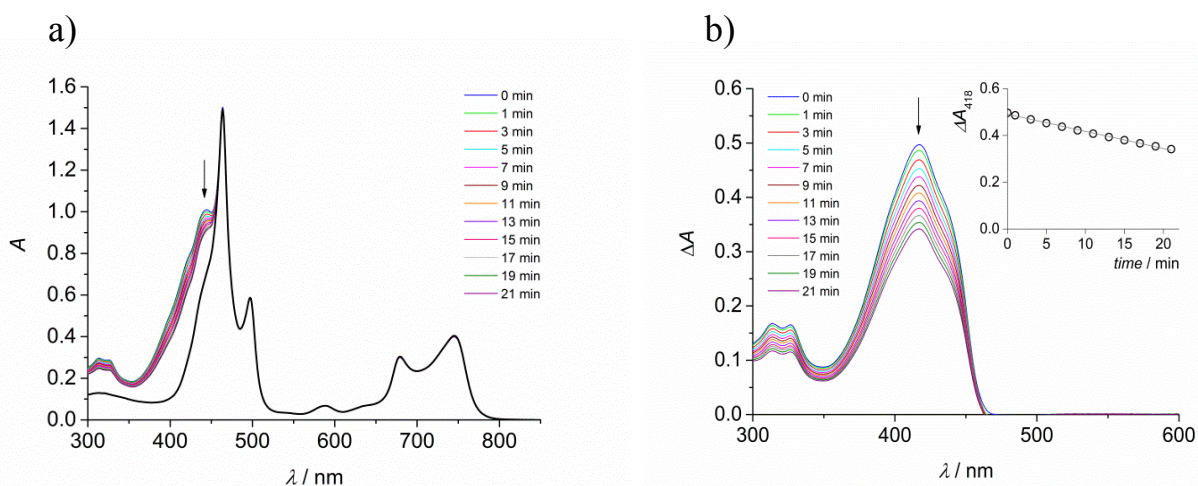


Figure 3. Singlet oxygen quantum yield determination in DMSO using 1,3-diphenylisobenzofuran (DPBF) as a singlet oxygen trap. a) Absorption spectra of a DMSO solution containing **1** (3.4×10^{-6} M, black thick) and DPBF (2.2×10^{-5} M) upon irradiation at 680 nm (0-20 min). b) The same spectra as (a) subtracted by the contribution of compound **1**. The inset shows the value of DPBF absorbance at 418 nm as a function of irradiation time. The same experiment for the standard ZnPc is reported in Figure S12.

Two-photon absorption properties.

The two-photon absorption properties of **1** were studied by the two-photon induced fluorescence method.⁶³ The low fluorescence quantum yield in water (even in the presence of pyridine or DMSO) precluded the determination of TPA properties in water by the fluorescence method. Therefore the two-photon excitation spectrum was obtained in DMSO at a concentration of 10^{-5} M (Figure 4). The compound displays a large band in the 870-940 nm range with intense TPA cross-section above 7000 GM and a maximum observed at 920 nm with a σ_2 value of 9400 GM. The values of σ_2 below 860 nm were discarded due to the progressive loss of the quadratic dependency of the fluorescence intensity vs the power of the laser excitation beam ascribed to the contribution of the one-photon lowest energy Q band absorption.

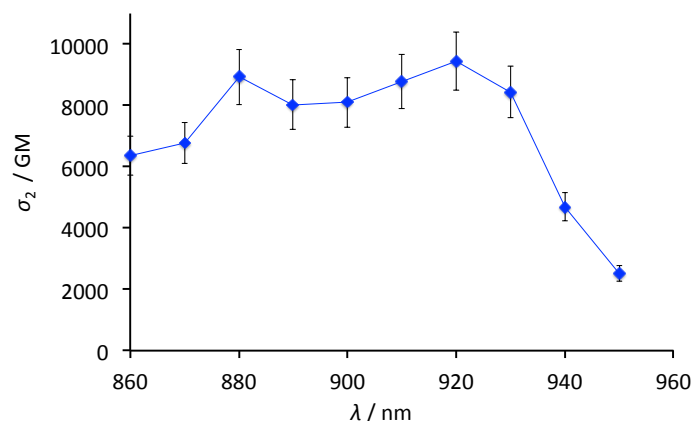


Figure 4. Two-photon excitation spectrum of **1** in DMSO.

Ethynylene-conjugated porphyrin dimers have indeed attracted attention since the highlight of their very efficient two-photon absorption properties. The electronic structure of **1** is similar to the one of butadiyne-bridged porphyrins described by Drobizhev *et al.*⁶⁴ and Anderson *et al.*⁵¹ and the two-photon absorption cross-sections were found to be in the same range. For two-photon PDT applications, a relevant parameter for the PS is expressed by the two-photon singlet oxygen sensitization value ($\sigma_2 \cdot \phi_\Delta$), which is around 3400 GM for **1** in DMSO at 930 nm. This high value, obtained for an excitation within the optical therapeutic window, highlights the potential of this compound to behave as a promising photosensitizer for two-photon excited PDT.

Cell penetration, subcellular localization and cytotoxicity

Cell penetration capacity and subcellular localization of the PS influence strongly the efficiency of the PDT treatment. The cellular behavior of **1** was studied on HeLa cell cultures. As for the anionic Gd(III) containing sensitizer **DPP-ZnP-GdDOTA**, the cell penetration is rather slow at 1 μM compared to the sensitizer without gadolinium complex.⁴⁹ After 24 h, the fluorescent signal was found to be stable in cells and further studies were performed after 24 h incubation in the culture medium (DMEM) at 1 μM . The uptake of **1** in HeLa cells was quantitatively assessed by measuring the Gd content in cells by ICP-MS. This uptake was compared with the cellular uptake of a commercial contrast agent $[\text{GdDTPA}]_2^3$ at an equivalent Gd incubation concentration (2 μM and 4 μM). As expected, the extracellular contrast agent $[\text{GdDTPA}]_2^3$ hardly penetrates in the cells (Figure S14). By contrast, compound **1** was able to enter in the cells and the cellular uptake strongly increased with the incubation concentration. The cellular uptake was 26 times higher than the one obtained with $[\text{GdDTPA}]_2^3$ at the Gd incubation concentration of 4 μM . This result shows the benefit of the design of an amphiphilic PS with a

large lipophilic core and hydrophilic complexes on the periphery to increase the concentration of the contrast agents in tumor cells.

Subcellular localization was detected by colocalization studies using fluorescent dyes for nuclei (Hoechst 33258), mitochondria (Mitotracker green) and lysosomes (Lysotracker blue). None of these cell constituents were targeted by the theranostic agent as shown by the absence of a clear colocalization with these three reference probes (Figure 5). Compound **1** shows a diffuse localization in the cytoplasm with higher accumulation in bright spots, probably endosomes.⁴⁸

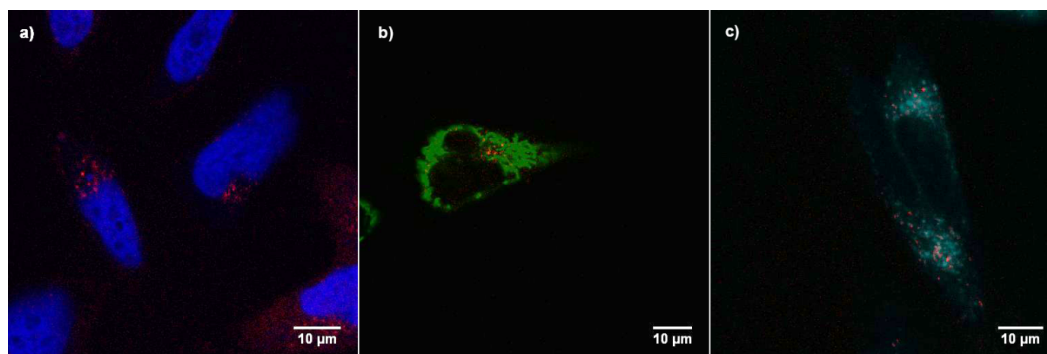


Figure 5. Confocal fluorescence images of HeLa cells incubated with **1** (1 μ M, 24 h) detected by its red fluorescence (λ_{ex} : 488 nm, λ_{em} : 700-800 nm) ; the cells are co-stained with a) Hoechst 33258 (λ_{ex} : 405 nm, λ_{em} : 425-500 nm), b) Mitotracker green (λ_{ex} : 488 nm, λ_{em} : 500-600 nm) and c) Lysotracker blue (λ_{ex} : 405 nm, λ_{em} : 425-500 nm). Scale bar: 10 μ m

We then examined the toxicity of the compound in the absence of light. HeLa cells were incubated with compound **1** at concentrations ranging from 0.5 to 16 μ M during 24 h. Compound **1** showed low cytotoxicity for concentrations lower than 2.5 μ M, with an IC_{50} value of 10 μ M (Figure 6). This cytotoxicity is comparable to many other sensitizing agents containing porphyrins, being in the 1-30 μ M range.^{65,66}

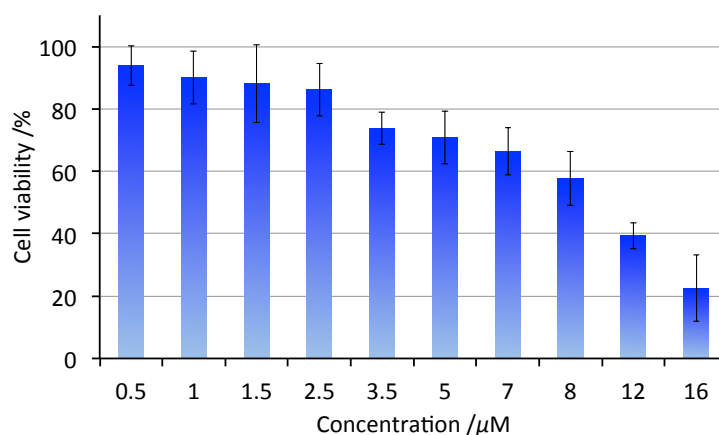


Figure 6. Cytotoxicity evaluation of **1** on HeLa cell culture after 24 h incubation

Phototoxicity evaluation following one-photon excitation.

The red-extended intense absorption of this new molecule ($\epsilon = 1.17 \times 10^5 \text{ M}^{-1}\text{cm}^{-1}$ at 746 nm in DMSO, Table 2) makes it appealing as one-photon PS for PDT with irradiation in the near-infrared (Figure 7). One-photon phototoxicity was evaluated as previously reported,^{48, 49} by irradiation of HeLa cells incubated for 24 h with **1** ($1 \mu\text{M}$) at 740 nm, with a fluence of 23 mW/cm², followed by an MTT cell toxicity test.

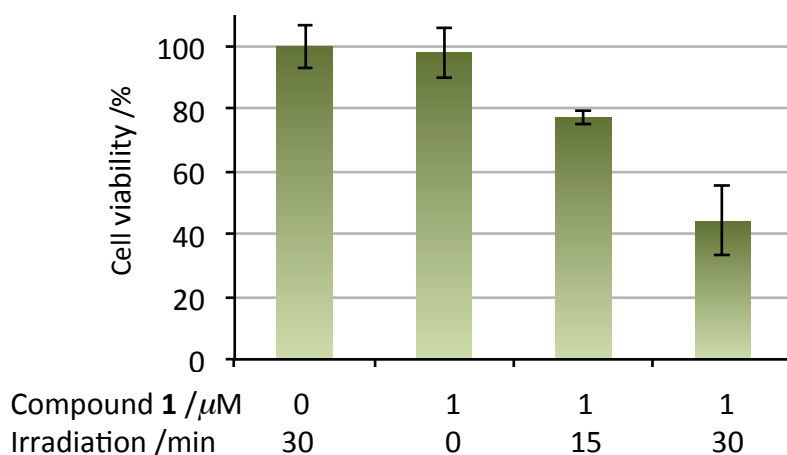


Figure 7. Viability of HeLa cells upon incubation of **1** ($1 \mu\text{M}$) for 24 h followed by 0, 15 or 30 min of one-photon irradiation at 740 nm. Two control experiments corresponding to irradiated/non-irradiated cells in absence/presence of compound **1** are also shown.

Neither irradiation alone at 740 nm for 30 min nor incubation of the photosensitizer at $1 \mu\text{M}$ without irradiation induced significant cell death (Figure 7), as expected by the use of near IR light and by the low dark toxicity of **1** at $1 \mu\text{M}$. Conversely, a significant phototoxicity was already observed after 15 min irradiation, whereas 50% of cell death was obtained after 30 min of irradiation. In these conditions, the light dose to induce 50% cell death LD_{50} was 40 J/cm².

Phototoxicity evaluation following two-photon excitation. Two-photon phototoxicity of **1** on HeLa cell cultures was performed using a femtosecond laser and evaluated in two ways, by following the cell morphology modifications just after two-photon irradiation and by evaluating the cell viability, 4 hours after two-photon irradiation. Directly after the excitation with an average power of 77 mW at the pupil of the objective, HeLa cells showed only small cell shrinkage but did not evidence any sign of cell death. After 10 min, formation of bubbles was detected on all cells, indicating an efficient TP induced phototoxicity (Figure 8). Control

experiments performed at the same excitation power without the PS did not exhibit any variation in the cell morphology (Figure S15).

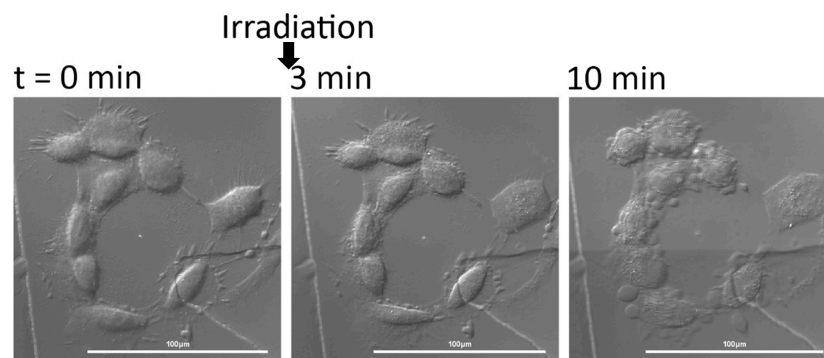


Figure 8. DIC images of HeLa cells incubated with **1** ($2\ \mu\text{M}$, 24 h) before ($t = 0\ \text{min}$) and after TP irradiation ($t = 3$ and $10\ \text{min}$) performed at $910\ \text{nm}$ with 300 scans ($77\ \text{mW}$ at the back pupil of the objective). Scale bar: $100\ \mu\text{m}$

For a quantitative assessment of this phototoxicity, cells were irradiated in the same way as before and then incubated for 4 hours before performing a cell viability test using Hoechst and Sytox orange dyes as described previously.^{48, 49} Figure 9 shows the images of the cell viability tests using excitation powers up to about $200\ \text{mW}$. Control experiments with irradiation but without the PS were performed and no phototoxicity was detected below an average power of $200\ \text{mW}$ measured at the back pupil of the objective. Cell death after incubation with **1** ($2\ \mu\text{M}$, 24 hours) and two-photon irradiation was not significant below $30\ \text{mW}$. Phototoxicity started to be more important after $77\ \text{mW}$. Remarkably, with a light power of $108\ \text{mW}$, 100 % of cell death was observed. (Figure 9 and Table S3).

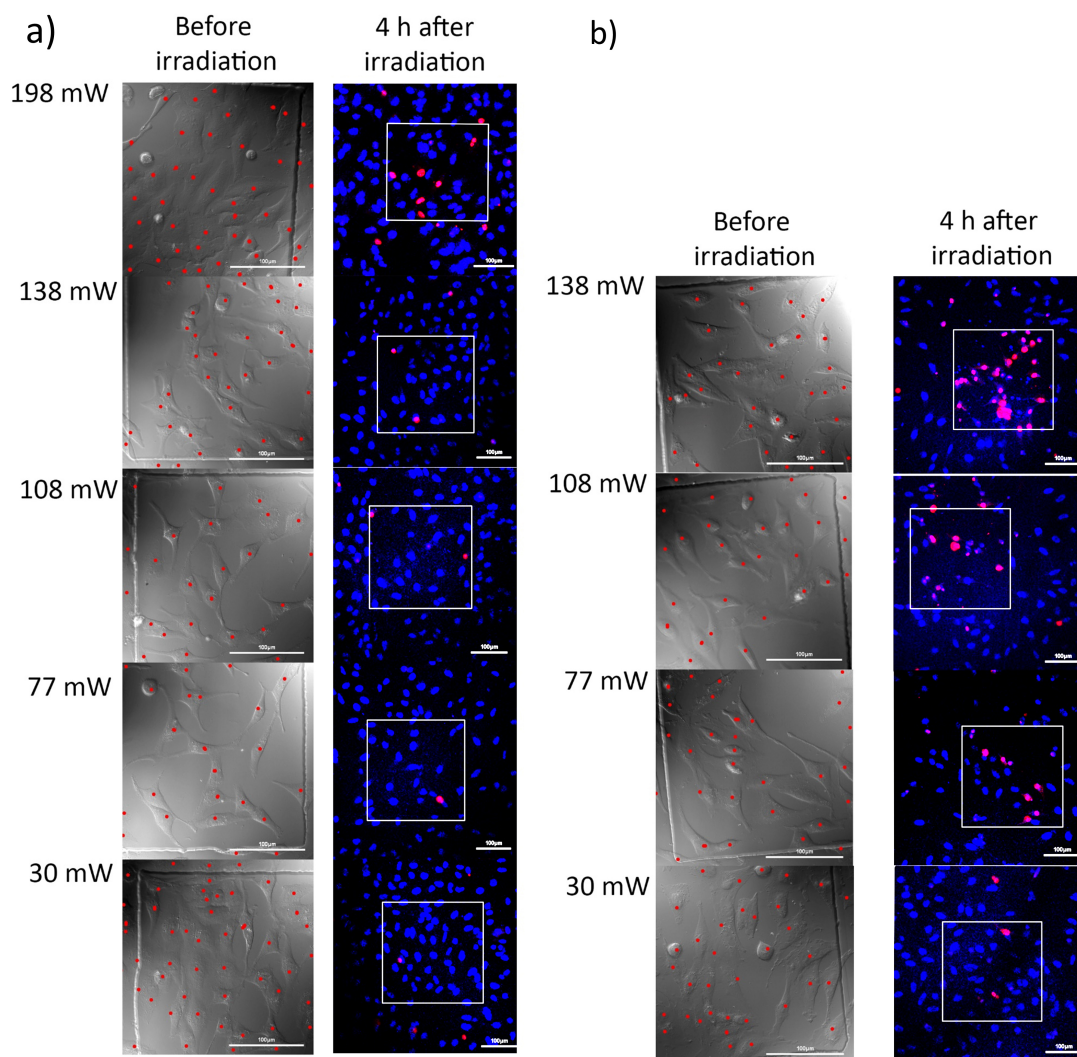


Figure 9. Images of HeLa cells obtained before irradiation (DIC images) and 4 h after two-photon irradiation at 910 nm (confocal images); a) Control experiments in the absence of PS; b) experiments in the presence of the PS 1 (2 μM , incubation 24 h). Cells counted before TP irradiation in the white square are highlighted with a red dot. All nuclei are stained with Hoescht (blue) and damaged cells are co-stained with Sytox orange (red). TP irradiation has been performed at different laser powers going from 30 to 198 mW. Scale bar: 100 μm .

■ Conclusion

A new theranostic agent designed to perform two functions, PDT in the optical therapeutic window and MRI detection was synthesized and characterized. This new molecule, consisting of a Zn(II) porphyrin dimer endowed with two GdDOTA-type complexes, behaves as a promising medium-size contrast agent, with a four time higher relaxivity (14.4 $\text{mM}^{-1}\cdot\text{s}^{-1}$ at 40 MHz per gadolinium at 25°C) than the commercial contrast agents and thanks to its

amphiphilicity, it is internalized in cancer cells. Moreover, with two gadolinium complexes, the value of relaxivity per molecule is twice as high, which is in favor of imaging at lower concentration. This agent has also shown appealing properties in solution as PDT photosensitizer regarding its solubility, its good ability to generate singlet oxygen and its high absorption in the near-infrared, above 700 nm. It also offers another spectral window between 870 – 930 nm, for deeper and highly selective treatment based on two-photon PDT, since it displays high two-photon cross sections above 7000 GM in this spectral range. These features were confirmed with PDT experiments performed on cancer cells incubated at low concentration with the theranostic agent. Both one-photon light irradiation at 740 nm and two-photon light irradiation at 910 nm applied to the cell cultures led to important cell death in conditions where both the theranostic agent or the light excitation alone were non-toxic. Such behavior opens new perspectives for PDT with a PS than can be activated at different tissue depths with a classical protocol or a more selective one, depending on the size and localization of the tumor. With the association of a new kind of PDT drug and Gd(III) complexes for MRI, this theranostic agent offers new features regarding both its imaging capacity and its applications in fine-tuned therapy, in line with the current research of non-invasive and personalized treatments.

■ Experimental procedures

All chemicals were of the best commercially available grade and used without further purification. Tetrahydrofuran was dried using a dry solvent station GT S100. Dry chloroform was obtained by distillation over CaH_2 under nitrogen. Triethylamine (Et_3N , 99%), and anhydrous dimethyl sulfoxide (DMSO, 99.7%) were purchased from Aldrich Chemicals (France). Analytical thin layer chromatography (TLC) was carried out on Merck aluminium backed silica gel 60 F254 plates and visualization when required was achieved using UV light. Precoated plates (PLC Silica gel 60 F₂₅₄, 2 mm) were used for preparative thin-layer chromatography. Column chromatographies were carried out on silica (Fluka 60, 70-230 mesh). Size-exclusion chromatography was carried out using Bio-Beads S-X1, 200-400 mesh (Bio-Rad). NMR spectra were recorded at 298 K using Bruker AVANCE 300, 400 or 500 spectrometers. Chemical shifts are quoted as parts per million (ppm) relative to the residual peak of solvent and coupling constants (J) are quoted in Hertz (Hz). The 2D-NMR techniques COSY and ROESY have been used to achieve full assignment of the signals. In the assignments, the chemical shift (in ppm) is given first, followed, in brackets, by the multiplicity

of the signal (s : singlet, d : doublet, t : triplet, m : multiplet, br s : broad signal), the value of the coupling constants in hertz if applicable, the number of protons implied, and finally the assignment. UV-vis spectra were recorded on a UVIKON XL spectrophotometer. Mass spectra were obtained by using a Bruker MicroTOF spectrometer (ES-MS). ICP-MS was performed with an Agilent 7500ce spectrometer. ICP-AES was performed by emission spectrometry with a Varian 720-ES spectrophotometer. The compounds **TIPS-ZnP**⁵⁹, **TIPS-ZnP-ZnP-TIPS**⁵² *tert*-butoxycarbonyl (Boc)-protected 3-iodobenzylamine (I-Bn-NHBoc)⁵⁴, Na₂[GdDOTAGA]⁵⁵ and Na₂[YDOTAGA]⁵⁵ were prepared according to the literature.

Synthesis of BocHN-ZnP-ZnP-NHBoc. To a degassed solution of **TIPS-ZnP-ZnP-TIPS** (113 mg, 41 μ mol) in dry THF (10 mL) was added, under argon, a solution of tetrabutylammonium fluoride (26 mg, 82 μ mol) in dry THF (1 mL). The reaction mixture was stirred at room temperature for 20 minutes and then, anhydrous calcium chloride (93 mg, 0.82 mmol) was added. The solvent was evaporated and the crude was submitted to a silica gel column chromatography (DCM/MeOH 96:4). Due to its instability, the deprotected compound **H-ZnP-ZnP-H** obtained as a brown-green solid was rapidly used for the next reaction. **H-ZnP-ZnP-H**, Pd₂(dba)₃ (3.8 mg, 4 μ mol), CuI (2 mg, 11 μ mol), PPh₃ (5.4 mg, 20 μ mol) and Boc-protected 3-iodobenzylamine (36 mg, 107 μ mol) were dried under vacuum for 1 hour at 45°C. A solution of dry THF (10 mL) and Et₃N (5 mL) degassed by four freeze-thaw cycles was transferred *via cannula* to the solids placed under argon. The resulting mixture was stirred at 50°C for 2 hours. The solvents were evaporated and the product was purified by silica gel column chromatography (DCM / pyridine 1%/ MeOH 1.5 to 2.5%). **BocHN-ZnP-ZnP-NHBoc** was obtained as a brown-green solid in 68% yield (80 mg). ¹H NMR (500 MHz, CDCl₃+1% pyridine) : δ = 1.50 (s, 18H, H₁₀), 3.29 (s, 24H, H₁), 3.47-3.49 (m, 16H, H₂), 3.60-3.62 (m, 16H, H₃), 3.67-3.69 (m, 16H, H₄), 3.76-3.78 (m, 16H, H₅), 3.93-3.94 (m, 16H, H₆), 4.31-4.33 (m, 16H, H₇), 4.43 (d, *J* = 5.2 Hz, 4H, H₈), 5.30 (br s, 2H, H₉), 6.94 (t, *J* = 2.0 Hz, 4H, p), 7.38 (d, *J* = 2.3 Hz, 8H, o), 7.56 (d, *J* = 7.9 Hz, 4H, m'), 7.97 (d, *J* = 7.9 Hz, 4H, o'), 8.91 (d, *J* = 4.6 Hz, 4H, py₃), 9.00 (d, *J* = 4.6 Hz, 4H, py₂), 9.67 (d, *J* = 4.6 Hz, 4H, py₄), 9.84 ppm (d, *J* = 4.6 Hz, 4H, py₁). ¹³C NMR (125 MHz, CDCl₃+1% pyridine) : δ = 28.59, 44.73, 59.11, 67.80, 70.68, 70.78, 71.00, 72.01, 82.35, 88.45, 93.42, 96.42, 99.38, 101.07, 101.91, 114.86, 122.71, 127.91, 130.85, 130.89, 131.92, 132.61, 133.09, 139.59, 144.55, 149.74, 150.10, 152.02, 153.35, 156.20, 157.98 ppm. **ES-MS** : *m/z* (%) 1449.53 (100) [M+2Na]²⁺/2 (calcd 1449.55 for [C₁₅₂H₁₈₀N₁₀O₃₆Zn₂Na₂]²⁺/2).

Synthesis of H₂N-ZnP-ZnP-NH₂. Compound **BocHN-ZnP-ZnP-NHBoc** (80 mg, 28 μ mol) was dissolved in dry DCM (20 mL) and the solution was cooled to 0°C. Trifluoroacetic acid (0.5 mL) was added dropwise and the solution was stirred overnight at room temperature. The organic phase was washed with water (10 mL) and aqueous saturated NaHCO₃ (10 mL). The organic layer was dried over Na₂SO₄, filtered and evaporated. The crude product was dissolved in dry CHCl₃ (20 mL) and a solution of Zn(OAc)₂·2H₂O (62 mg, 0.28 mmol) in MeOH (2 mL) was added dropwise. The resulting mixture was stirred at 50°C for 5 hours. After evaporation of the solvents, DCM (10 mL) was added and the solution was washed twice with brine (2 x 10 mL) then with water (10 mL). The organic layer was dried over Na₂SO₄, filtered and evaporated to dryness. **H₂N-ZnP-ZnP-NH₂** was obtained as a brown-green solid in 97% yield (72 mg). **¹H NMR** (500 MHz, CDCl₃+1% pyridine) : δ = 3.30 (s, 24H, H₁), 3.48-3.50 (m, 16H, H₂), 3.61-3.63 (m, 16H, H₃), 3.67 (br s, 4H, H₄), 3.68-3.70 (m, 16H, H₅), 3.77-3.79 (m, 16H, H₆), 3.94-3.96 (m, 16H, H₇), 4.32-4.34 (m, 16H, H₈), 6.95 (t, *J* = 2.1 Hz, 4H, p), 7.31 (d, *J* = 7.8 Hz, 4H, m'), 7.40 (d, *J* = 2.1 Hz, 8H, o), 7.95 (d, *J* = 7.9 Hz, 4H, o'), 8.92 (d, *J* = 4.6 Hz, 4H, py₁), 9.01 (d, *J* = 4.6 Hz, 4H, py₂), 9.68 (d, *J* = 4.6 Hz, 4H, py₃), 9.85 ppm (d, *J* = 4.6 Hz, 4H, py₄). **¹³C NMR** (125 MHz, CDCl₃+1% pyridine) : δ = 29.85, 46.17, 59.15, 70.01, 70.71, 70.81, 71.03, 72.04, 82.37, 88.47, 93.21, 96.63, 99.36, 101.11, 102.13, 114.89, 122.71, 122.82, 127.38, 130.86, 130.96, 131.88, 133.11, 143.19, 144.60, 149.75, 150.11, 152.06, 158.01 ppm. **HR ES-MS** : *m/z* (%) 1338.9996 (100) [M+2Na]²⁺/2 (calcd 1338.9986 for [C₁₄₀H₁₆₅N₁₀O₃₆Zn₂Na₂]²⁺/2).

Synthesis of compounds 1 and 2. A solution of the complex **Na₂[MDOTAGA]** (19.7 mmol.L⁻¹, 89 μ mol) in dry DMSO under argon was obtained after 5 hours stirring at 60°C. TBTU (31 mg, 98 μ mol) and DIPEA (30 μ L, 178 μ mol) were added to the resulting solution. A solution of **H₂N-ZnP-ZnP-NH₂** (72 mg, 28 μ mol) in dry DMSO (4 mL) was then added dropwise. The reaction mixture was stirred at room temperature overnight. The compound was isolated by precipitation with addition of DCM (4 mL). After filtration, the solid was washed with DCM, acetone and water and dried under vacuum.

Characterization data of the gadolinium(III) complex **1**: dark-green solid (58 mg, 52%) **UV-vis.** (DCM): λ_{max} (log ϵ) = 464 (5.64), 498 (5.21) 587 (4.31), 680 (4.94), 745 nm (5.04). **HR ES-MS** : *m/z* (%) 1938.5846 (100) [M-2H₂O]²⁺/2 (calcd 1938.5999 for [C₁₈₀H₂₁₆Gd₂N₁₈O₃₆Zn₂]²⁺/2, 100%). **ICP-AES** : [Gd] = 13.1 \pm 0.<1 mg/kg = 83.3 \pm 0.6 mmol/kg, [Zn] = 5.64 \pm 0.04 mg/kg = 86.2 \pm 0.6 mmol/kg.

Characterization data of the yttrium(III) complex **2**: dark-green solid (40 mg, 55%). **¹H NMR** (500 MHz, DMSO) : δ = 1.70-1.88 (m, 4H, H₁₀), 2.21-2.84 (m, 50H, DOTA), 3.19 (s, 24H, H₁),

3.39-3.41 (m, 16H, H₂), 3.51-3.53 (m, 16H, H₃), 3.57-3.58 (m, 16H, H₄), 3.64-3.65 (m, 16H, H₅), 3.86 (m, 16H, H₆), 4.35 (m, 16H, H₇), 4.42-4.49 (m, 4H, H₈), 7.05 (m, 4H, p), 7.37 (m, 8H, o), 7.52 (d, 4H, $J = 7.3$ Hz, m'), 8.10 (d, $J = 7.5$ Hz, 4H, o'), 8.55 (br s, 2H, H₉), 8.91 (d, $J = 4.1$ Hz, 4H, py₃), 9.00 (d, $J = 4.0$ Hz, 4H, py₂), 9.71 (d, $J = 4.3$ Hz, 4H, py₄), 9.83 ppm (d, $J = 4.3$ Hz, 4H, py₁). **HR ES-MS**: m/z 1870.0761 (100) [M-2H₂O]²⁻/2 (calcd 1870.0816 for [C₁₈₀H₂₁₆Y₂N₁₈O₃₀Zn₂]²⁻/2, 100%).

Relaxometric measurements. ¹H NMRD profiles ([**1**] = 0.81 mM, pH = 6.6 in 2% pyridine/H₂O) were recorded on a Stellar SMARTracer Fast Field Cycling relaxometer (0.01-10 MHz) and a Bruker WP80 NMR electromagnet adapted to variable field measurements (20-80 MHz) and controlled by a SMARTracer PC-NMR console. Higher field relaxivities were measured on Bruker AVANCE NMR spectrometers at 300 MHz, 400 MHz, and 600 MHz. The temperature was monitored by a VTC91 temperature control unit and maintained by a gas flow and was determined by previous calibration with a Pt resistance temperature probe. The least-square fit of the ¹H NMRD data was performed using Visualiseur/Optimiseur⁶⁷ running on a MATLAB 8.3.0 (R2014a) platform. The relaxivity was also measured in the presence of BSA (42.7 g.L⁻¹) for [**1**] = 0.60 mM at 298 and 310 K and 40 MHz. The exact Gd(III) ion concentration was determined by ICP-MS analysis after microwave-assisted mineralization.

One-photon spectroscopy and photophysics. Spectroscopic grade DMSO was purchased from Carlo Erba. Pyridine, 1,3-diphenylisobenzofuran (DPBF), Zn-phthalocyanine (ZnPc) and 5,10,15,20-tetrakis(4-sulfonatophenyl)porphyrin (TPPS₂) were from Aldrich. Tri-distilled (Millipore Milli-Q) H₂O was used and the solutions prepared with 1% of pyridine were adjusted to pH = 7 with microadditions of diluted HCl aqueous solutions. Absorption spectra were recorded with a Perkin-Elmer Lambda 650 UV-Vis spectrophotometer. Molar absorption coefficients in H₂O added with 1 % pyridine and 1% DMSO were derived from that in DMSO by vacuum drying a DMSO solution of known concentration and adding the aqueous solvent. Emission spectra were collected with an FLS920 fluorimeter (Edinburgh) equipped with an Hamamatsu R5509-72 InP/InGaAs photomultiplier tube supercooled at 193 K in a liquid nitrogen cooled housing and a TM300 emission monochromator with a NIR grating blazed at 1000 nm (sensitivity range: 300-1700 nm). The spectra have been corrected for the wavelength dependent phototube response. The fluorescence quantum yields have been determined with reference to TPP (tetra-phenyl-porphyrin) in aerated toluene ($\phi_f = 0.11$)⁶⁸. Fluorescence lifetimes have been measured with an IBH Time Correlated Single Photon Counting apparatus with nanoLED excitation at 465 nm. The analysis of the luminescence decay profiles against time

was accomplished with the DAS6 Decay Analysis Software provided by the manufacturer. Estimated errors are 10% on exponential lifetimes, 20% on quantum yields, 20% on molar absorption coefficients and 3 nm on emission and absorption peaks. Singlet oxygen production quantum yields in DMSO have been measured with a comparative method using DPBF as a singlet oxygen trap.⁶⁹ ZnPc has been used as a standard ($\phi_{\Delta} = 0.67$).⁷⁰ Solutions of the standard or of compound **1** containing DPBF 2.2×10^{-5} M, prepared in the dark, have been irradiated at 680 nm by using an irradiation set-up composed by a 150 W Xenon lamp (LOT) and a Omni- λ 150 monochromator (Zolix) with a 16 nm slit and completed by a 635 nm cutoff filter, under continuous stirring. The light intensity was 3.5 mW/cm². The decrease of DPBF absorption, derived by subtracting the constant absorption contribution of **1** or the standard from the spectrum of the mixture, has been followed at 418 nm (see Figures 3 and S12). Singlet oxygen quantum yields in H₂O added with 1 % pyridine and 1% DMSO have been derived with the same indirect method by using anthracene-9,10-dipropionic acid (ADPA) disodium salt as a singlet oxygen trap.⁷¹ TPPS, has been used as a standard ($\phi_{\Delta} = 0.51$).⁷² Solutions of the standard or **1** containing 3.2×10^{-5} M ADPA, prepared in the dark, have been irradiated at 516 nm by using the same irradiation setup with a 16 nm slit and completed by a 500 nm cutoff filter, under continuous stirring. The light intensity was 2 mW/cm². The decrease of ADPA absorption has been followed at 378 nm (see Figure S13). The singlet oxygen quantum yield of the sample (ϕ_{Δ}) has been determined by using equation (1):

$$(1) \quad \Phi_{\Delta} = \Phi_{\Delta}^{Std} \frac{R_{abs}^{Std}}{R_{abs}^{Std} I_{abs}}$$

where R and R^{std} are the trap (DPBF or ADPA) degradation rates for the sample and the standard, respectively, and I_{abs} is the absorption at the excitation wavelength calculated as described previously.⁴⁹

DLS measurements. The particle hydrodynamic diameter was measured using a NanoBrook Omni Analyzer from Brookhaven Instruments Corporation. Average particle size distributions were measured by means of dynamic light scattering at a fixed scattering angle of 90° and at 25°C. The hydrodynamic diameter was calculated applying an autocorrelation function to the scattered light intensity profile considering the particles as spheres. Red laser light (640 nm) was used in the scattering experiments. From four to five measurements were carried out for reproducibility in each solvent. Disposable polystyrene cuvettes were used. The concentration of **1** in both solvents was 2×10^{-6} M.

Two-photon spectroscopy. The two-photon excitation spectra was obtained by upconverted fluorescence measurements using a Ti:sapphire femtosecond laser Insight DS with pulse width <120 fs and a repetition rate of 80 MHz (Spectra-Physics) as described previously.⁶³ The excitation beam was collimated over the cell length (10 mm) and the fluorescence, collected at 90° of the excitation beam, was focused into an optical fiber connected to a spectrometer. The incident beam intensity was adjusted to ensure an intensity-squared dependence of the fluorescence over the whole spectral range investigated. Calibration of the spectra was performed by comparison with the published rhodamine B TPA spectrum.⁷³

Cell culture and MTT cell viability test. HeLa cells were cultured in DMEM complete culture medium containing phenol red at 37°C with 5% CO₂. They were seeded and maintained in 25 mL Falcon culture flask or multi well LabTek (Lab-Tek® II) culture flasks. MTT toxicity tests were performed in pentaplicate with extemporaneous solutions of 3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (50 μL of a 5 mg/mL solution in DMEM by well). After an incubation period of 45 minutes, the media was removed and replaced by 150 μL of DMSO. The absorbance was measured in 96 well plates with a Safas Xenius spectrofluorimeter at 550 nm.

HeLa cell preparation for Gd quantification by ICP-AES. HeLa cells were seeded in 6 wells LabTek culture plate in complete DMEM medium as described in the cell culture paragraph. The cultures were incubated with **1** (1 μM) and [GdDTPA]²⁻ for 24 hours. They were then washed three times with PBS (3 x 2 mL). Cells were suspended using trypsin (0.5 mL) and after 5 min DMEM (9.5 mL) was added and transferred in 10 mL Falcon tubes. The cells were counted using a Malassez cell. The culture medium was removed by gentle centrifugation at 60 × g. A 60% nitric acid solution was added to the centrifugation pellet and the cells were digested for 24 hours at room temperature. The Gd content in aqueous samples was determined by ICP-AES after mineralization in an Anton Paar microwave oven (multiwave ECO 3000) with a nitric acid solution (HNO₃, 20% for trace analysis ≥69.0%, TraceSELECT®, from Fluka).

Dark cytotoxicity. HeLa cells were seeded in 96 wells culture plates and incubated with different concentrations of **1**. After 24 hours the cell viability was estimated using previously described MTT test.

One-photon phototoxicity tests. One-photon phototoxicity tests were performed as described in the literature using a homemade apparatus^{88,99} using 800 mA high power LED Far Red / Infra Red (740-745 nm) (FutureEden™) adapted for Corning® 96 well special optic plates. Cells were cultured 1 day in these 96 well plates and incubated with 1 μM solution of

photosensitizer in complete DMEM medium for 24 hours. The medium was replaced by new DMEM free of sensitizer and the plates were irradiated for 15 or 30 min. The LEDs power was measured with a Thorlabs PM100D power-meter. The cell viability was then estimated 24 hours after irradiation using previously described MTT test.

Two-photon phototoxicity tests. HeLa cells were plated at 25% confluence in IBIDI μ -Dish 35 mm, grid-500. After 24 h growth the culture medium was replaced by a fresh one containing **1** (2 μ M). Two-photon irradiation was performed after 24 h incubation, with a cell confluence of about 80 % using a Leica SP5 inverted microscope with a HCX PL APO CS 40x oil Leica objective and a Coherent Chameleon Ultra 2 laser as fs laser source (150 fs pulses with a repetition rate of 80 MHz set at 910 nm). A region of 260 x 260 μ m (zoom set to 1,5) was irradiated. The laser power was measured at the back pupil of the objective and set between 30 and 200 mW. The focus plane was selected by DIC (differential interference contrast) microscopy (λ_{ex} = 513 nm) to be at the bottom part of the cells. They were irradiated using a series of 10 slices (10 up the selected focal plane corresponding to the coverslip), with 1 μ m z increments, with 30 scans per slice (300 scans in total). After irradiation the cells were imaged during 10 min in wild field DIC to detect morphological changes. For the cell viability tests, the irradiated cells were placed for 3h30 in an incubator. Hoescht 33342 (5 μ g/mL) and Sytox orange (5 μ M) were added and the cells placed in the incubator for another 30 min before imaging. Confocal images were obtained with a Leica TSP SPE microscope using two channels, one for Hoescht (ex. 405 nm) and one for Sytox orange (ex. 561 nm).

A control experiment was performed for each light power using the same conditions but without **1**.

■ Associated content

Supporting information

The Supporting Information is available free of charge on the ACS Publications website at DOI XX

¹H, ¹³C NMR and HR ES-MS of the synthesized compounds, excitation spectra of **1**, quantum yield determination, Gd content per cell, cell DIC images after TP excitation, longitudinal relaxivity of **1** and estimation of cell viability after TP excitation.

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Notes

The authors declare no competing financial interest

■ Acknowledgements

The icFRC (<http://www.icfrc.fr>), LabEx CSC, and Region Alsace are gratefully acknowledged for a fellowship to J.S, and the Ministry of Education and Research is acknowledged for a Ph.D. fellowship to S.J. The Italian CNR (Project "PHEEL") and MIUR-CNR project Nanomax N-CHEM are gratefully acknowledged. C.S.B. and E.T. thank the French Ligue contre le cancer for financial support. Dr. F. Manoli (ISOF-CNR) is acknowledged for a loan of ADPA and for support in DLS measurements. The authors are also grateful to Anne Boos and Pascale Ronot for ICP-AES measurements.

■ References

NOTE 1 : Direct singlet oxygen phosphorescence analysis was precluded both in DMSO, due to the poor luminescence properties of singlet oxygen in this solvent, and in H₂O, due the required pH control which is less straightforward in D₂O, the solvent usually employed for direct phosphorescence measurement in aqueous environments because of the longer lifetime of ¹O₂ in the deuterated solvent.

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