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► **To cite this version:**

Cécile Breyton, Waqas Javed, Anne Vermot, Charles-Adrien Arnaud, Christine Hajjar, et al.. Assemblies of lauryl maltose neopentyl glycol (LMNG) and LMNG-solubilized membrane proteins. *Biochimica et Biophysica Acta: Biomembranes*, 2019, 1861 (5), pp.939-957. 10.1016/j.bbamem.2019.02.003 . hal-02057161

HAL Id: hal-02057161

<https://hal.science/hal-02057161v1>

Submitted on 13 Mar 2020

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Assemblies of lauryl maltose neopentyl glycol (LMNG) and LMNG-solubilized membrane proteins

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Keywords

Membrane proteins – Detergent – LMNG – homogeneity – crystallization – rods

Footnotes

AUC: Analytical Ultracentrifugation ; DSF: Differential Scanning Fluorimetry; SEC-LS: Size Exclusion Chromatography coupled to Light Scattering; SAXS: Small Angle X-Ray Scattering; SANS: Small Angle Neutron Scattering; TLC: Thin Layer Chromatography; CMC: critical micelle concentration ; DDM: dodecyl- β -D-maltopyranoside; DDAO: decyldimethylamine-N-oxide; LDAO: lauryldimethylamine-N-oxide; LMNG: lauryl maltose neopentyl glycol; OTG: octyl- β -D-thioglucopyranoside; bR: bacteriorhodopsin from *Halobacterium*; BmrA: Bacillus multidrug resistance ATP; SpNOX: *S. pneumoniae* NOX protein; FhuA: *E. coli* outer membrane ferrichrome-iron transporter. Vi: orthovanadate;

1 Abstract

2
3 Laurylmaltose neopentylglycol (LMNG) bears two linked hydrophobic chains of equal length
4 and two hydrophilic maltoside groups. It arouses a strong interest in the field of membrane
5 protein biochemistry, since it was shown to efficiently solubilize and stabilize membrane
6 proteins often better than the commonly used dodecylmaltopyranoside (DDM), and to allow
7 structure determination of some challenging membrane proteins. However, LMNG was
8 described to form large micelles, which could be unfavorable for structural purposes. We thus
9 investigated its auto-assemblies and the association state of different membrane proteins
10 solubilized in LMNG by analytical ultracentrifugation, size exclusion chromatography
11 coupled to light scattering, centrifugation on sucrose gradient and/or small angle scattering.
12 At high concentrations (in the mM range) LMNG forms long rods, and it stabilized the
13 membrane proteins investigated herein, *i.e.* a bacterial multidrug transporter, BmrA; a
14 prokaryotic analogous of the eukaryotic NADPH oxidases, SpNOX; an *E. coli* outer
15 membrane transporter, FhuA; and the halobacterial bacteriorhodopsin, bR. BmrA, in the Apo
16 and the vanadate-inhibited forms showed reduced kinetics of limited proteolysis in LMNG
17 compared to DDM. Both SpNOX and BmrA display an increased specific activity in LMNG
18 compared to DDM. The four proteins form LMNG complexes with their usual quaternary
19 structure and with usual amount of bound detergent. No heterogeneous complexes related to
20 the large micelle size of LMNG alone were observed. In conditions where LMNG form
21 assemblies of large size, FhuA crystals diffracting to 4.0 Å were obtained by vapor diffusion.
22 LMNG large micelle size thus does not preclude membrane protein homogeneity and
23 crystallization.

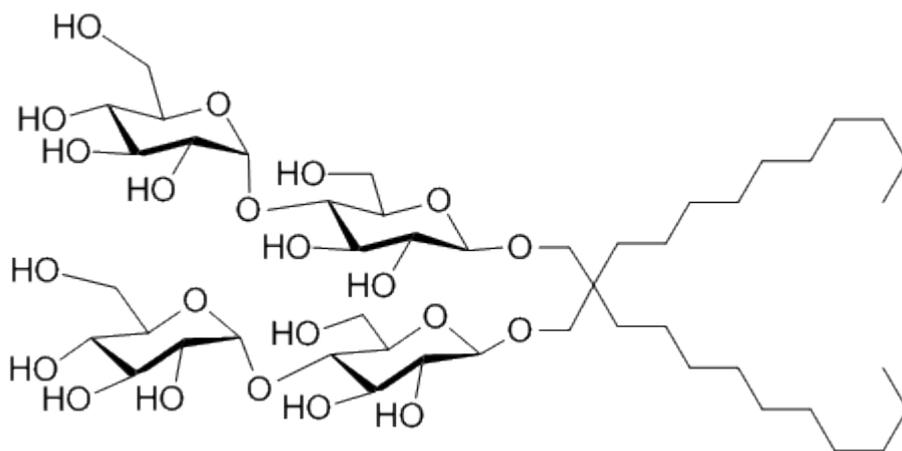
24 Introduction

25
26
27 Detergents are amphipatic molecules that auto-assemble into micelles above the critical
28 micelle concentration (CMC). Above the CMC, they are able to solubilize lipids and
29 membrane proteins, forming protein-detergent complexes of small and well-defined size [1].
30 Detergents are essential for membrane protein solubilization and purification steps, as well as
31 functional and structural studies. However, they often lead to membrane protein inactivation.
32 One of the mechanisms leading to inactivation (for a recent review, see [2]) is the dissociation
33 of subunits, loss of essential lipids or other hydrophobic co-factors. As an example,
34 cytochrome *b₆f* inactivation and monomerization was related to lipid loss [3]. Detergent
35 dynamics, *i.e.* detergent exchange between the free monomers, or between micelles, and
36 bound detergent at the surface of the transmembrane protein surface, could trigger transient
37 exposure of this hydrophobic surface leading to irreversible protein aggregation. Detergent
38 binding can also alter protein structure: molecular dynamics simulation indeed showed, for a
39 thermostable mutant of the adenosine receptor, that the harsh octylglucoside detergent
40 molecules intercalate between trans-membrane helices, moving them apart in the 200 ns
41 simulation [4]. The structure and dynamics of different α -helical membrane proteins
42 solubilized in alkyl phosphocholines appear deeply altered [5]. For example, NMR, molecular
43 dynamics simulations, and functional studies, amongst other techniques, show that
44 mitochondrial carriers are incorrectly folded in these detergents. Some parts of the
45 transmembrane segments are disordered, with molecules of detergent penetrating between the
46 helices; the proteins are in a highly dynamic state with unstable tertiary contacts; and weak
47 ligand binding results from non-specific interactions [5]. In the challenging aim to obtain high
48 resolution structure of membrane proteins by crystallography, a usual strategy is to select

49 detergents that not only preserve membrane protein stability, function and solubility, but also
50 form small micelles (*e.g.* by adding compounds decreasing the micelle size), in order to favor
51 protein-protein contacts [6,7].

52
53 The propensity of detergents to inactivate membrane proteins drove the search for new
54 amphiphilic environments (for a review, see [6,8,9]). Peptides that form an α -helix or a β -
55 sheet [10–13], amphipatic polymers with multiple hydrophobic tails [14,15], fluorinated
56 surfactants (reviewed in [8,16,17]), Calixarene based detergents (*e.g.* [18]) or detergents with
57 varying architectures: tripod amphiphiles (*e.g.* [19,20]), facial amphiphiles [21–23],
58 polycyclic based [24], bi-tails [25–29], tri-tails [30], and other compounds [31–33] have been
59 synthesized to decrease their exchange kinetics, and/or reinforce interactions with membrane
60 protein surfaces. This is done noticeably by increasing the rigidity as well as the surface of
61 their hydrophobic moieties. Among these compounds, neopentyl glycols (MNG) constitute a
62 class of detergents with notable interest [25,26]. Members with different chain lengths or with
63 different sugar head groups have been described and are commercialized (Anatrace). We will
64 focus here on the most commonly used one, lauryl maltose neopentyl glycol (LMNG), also
65 named MNG3 (MNG3, however, designates the related family of compounds in [26]). Its
66 structure (Scheme 1) is formed by two molecules of n-dodecyl- β -D-maltopyranoside (DDM)
67 linked at the junction between the hydrophilic and hydrophobic moieties of the molecule. The
68 presence of the two hydrophobic chains of equal length mimics better the structure of lipids
69 than classical detergents having a single hydrophobic tail. According to the authors who
70 designed this detergent in 2010, the central quaternary carbon, was “intended to place subtle
71 restraints on conformational flexibility” [26]. In this work, LMNG was shown to display a set
72 of very interesting features: to extract integral membrane proteins from membranes; to
73 improve substantially the stability of various membrane proteins, including G protein-coupled
74 receptors and respiratory complexes. It also allowed diffraction up to 3.4 Å resolution of
75 LMNG-solubilized cytochrome *b₆f* crystals obtained by vapor diffusion [26]. Shortly after this
76 first publication, high resolution structures of the challenging β 2 adrenergic receptor, in
77 complex with G-proteins [34] or with covalently bound agonist [35], were obtained. The
78 proteins were solubilized and purified in LMNG followed by crystallization in mesophase.
79 LMNG was then described as successfully used for various applications, including membrane
80 solubilization and membrane protein stabilization (*e.g.* [26,36–40]), excited state
81 intermediates [37,41–44], NMR studies [44], H/D exchange coupled to mass spectroscopy
82 (*e.g.* [45,46]), negative stain electron microscopy (*e.g.* [43,46,47]), or reconstitution in lipid
83 vesicles [48]. Impressively, LMNG has now been used to solve the structure of more than ten
84 G-protein coupled receptors (*e.g.* refs in [49]; in all cases, LMNG was used for protein
85 purification and crystallization was performed in mesophase). Other proteins, *e.g.* ABC
86 transporters [41,50,51] or channels [52], were purified and cystallized in LMNG by vapor
87 diffusion. Furthermore, LMNG was also successfully used to solve the high resolution
88 structures of membrane proteins by single particle cryo-electron microscopy (*e.g.* [47,53–
89 55]).

90
91



92
93 **Scheme 1: Chemical structure of LMNG**
94

95 Regarding the β_2 adrenergic receptor in complex with G-proteins, not only does the complex
96 display long term stabilization in LMNG, but surprisingly, its ligand binding activity is
97 preserved even after diluting LMNG 1000 fold below its CMC [34]. The kinetics of
98 interconversion between the different β_2 adrenergic receptor conformations are significantly
99 slower for the protein solubilized in LMNG *vs.* DDM, as shown by F-NMR on labelled
100 proteins [44]. Notably, it was proposed that this was related to the very low CMC of LMNG.
101 From simple considerations, assuming that the on-rate for a monomer forming a micelle is
102 diffusion-limited (thus related only to the detergent monomer size, similar for LMNG and
103 DDM), the detergent off-rates are expected to be lower for LMNG than that for DDM, by a
104 factor being the ratio of their CMCs [44] (the value of the CMC of LMNG will be discussed
105 below). The large off-rate of LMNG may be an explanation for the slower protein dynamics.
106 The fact that individual monomers dissociate much slower from the protein than other
107 standard detergents would also explain the exceptional stabilization of protein-LMNG
108 complexes, and the presence of residual activity measured after dilution below the CMC:
109 LMNG “sticks” to the protein surface, and protein-detergent complexes are preserved. A
110 protocol (GraDeR) was indeed proposed to prepare samples at 1 CMC of LMNG, thus devoid
111 of excess LMNG, for obtaining cryo-electron microscopy images of significantly enhanced
112 quality [47].
113

114 However, probably due to the particular geometry of this molecule, the LMNG detergent
115 assemblies formed in solution may be rather large: a hydrodynamic radius of 7.2 nm was
116 determined for a 0.5 % solution of LMNG [49]. The questions to be addressed are therefore:
117 how are the large LMNG assemblies/micelles organized in solution? Does the large size of
118 LMNG micelles lead to large protein-detergent complexes? What is the homogeneity and the
119 quaternary structure of the solubilized membrane proteins, and how do they compare to those
120 in more usual detergents? Is the crystallization process affected by the large size of the
121 detergent micelles and/or the possible large size of the solubilized protein-detergent
122 complexes?
123

124 To answer these questions, and having in mind that the effects of detergents are protein
125 dependent, we characterized the assemblies in solution of LMNG alone and in complex with
126 four different membrane proteins, FhuA, SpNOX, BmrA and bacteriorhodopsin (bR). We
127 confirmed the stabilization propensity of LMNG compared to DDM, by measuring thermal
128 denaturation by Differential Scanning Fluorimetry (DSF) of the four proteins, limited
129 proteolysis of BmrA, and specific activities of BmrA and SpNOX. We also used
130 complementary techniques such as small angle X-ray and Neutron scattering (SAXS and

131 SANS), Analytical Ultracentrifugation Sedimentation Velocity (AUC-SV) and Size exclusion
132 chromatography coupled to light scattering (SEC-LS) and dynamic light scattering (DLS). We
133 showed that, although LMNG alone forms increasingly large assemblies with increasing
134 concentration, the protein-LMNG complexes are of comparable size to those formed with
135 other detergents, with overall the same weight amount of bound detergent, and the oligomeric
136 state of each protein remains unchanged. Furthermore, FhuA solubilized in LMNG showed an
137 overall increased number of hits when screened for crystallization, suggesting that the large
138 size of LMNG micelles does not prevent protein crystallization.

139

140 **Materials and method**

141

142 **Chemicals and buffers**

143 Detergents are from Anatrace, except octyl- β -D-thioglucopyranoside (OTG) from Acros,
144 other chemicals are typically from Sigma-Aldrich.

145 Buffer A : 50 mM Tris-HCl pH 7.0, 300 mM NaCl

146

147 **FhuA purification**

148 FhuA was purified from the *E. coli* strain AW740 transformed with a plasmid encoding the
149 *fhuA* gene in which a 6xHis.tag has been inserted in the extracellular loop L5 [56]. Cells were
150 grown in LB medium at 37 °C in the presence of 100 mM of the iron-chelating agent
151 dipyrindine, and broken with a microfluidizer (10 passages at 15 kpsi) in a 20 mM Tris-HCl
152 pH 8, 150 mM NaCl buffer with a pinch of DNase I. Unbroken cells were removed by a first
153 centrifugation (15 min, 6000 rpm, rotor SX4250), and the membrane fraction was recovered
154 by ultracentrifugation (20 min, 35 000 rpm, 45Ti). A first solubilization with 2 % OPOE
155 (octylpolyoxyethylene), 20 mM Tris-HCl pH 8.0, 30 min at 37 °C under gentle shaking
156 solubilized the inner membranes, and the outer membrane pellet (20 min, 35 000 rpm, 45Ti)
157 was solubilized in 1 % lauryldimethylamine-N-oxide (LDAO), 20 mM Tris-HCl pH 8, 1 mM
158 EDTA for 1 h at 37 °C. Insoluble material was removed (20 min, 35 000 rpm, 45Ti). The
159 protein was purified by Nickel affinity chromatography (HiTrap Chelating, 5 mL, GE
160 Healthcare). The LDAO supernatant was supplemented with 5 mM MgCl₂ and 5 mM
161 Imidazole, loaded onto the column and thoroughly washed (20 mM Tris-HCl pH 8, 150 mM
162 NaCl, 0.1 % LDAO). A delipidation step was achieved by washing with 10 mL of the same
163 buffer containing 1% LDAO. The protein was eluted with 20 mM Tris-HCl pH 8.0, 200 mM
164 Imidazole, 0.1 % LDAO. Fractions containing the protein were pooled and loaded onto an
165 anion-exchange chromatography (HiTrap Q, 1 mL, GE Healthcare) equilibrated with 20 mM
166 Tris-HCl pH 8.0, 0.05 % LDAO and the protein was eluted by a linear gradient of NaCl in 20
167 mM Tris-HCl pH 8, 0.05 % (2.2 mM) LDAO (final NaCl concentration ~150 mM). The
168 concentration of FhuA was determined using $\epsilon_{280\text{ nm}} = 103,690\text{ M}^{-1}\text{ cm}^{-1}$.

169

170 **FhuA detergent exchange**

171 For LDAO - LMNG exchange, the protein sample was diluted 50 times with water, to reach a
172 detergent concentrations far below the LDAO CMC. Protein aggregation was checked by
173 spectrometry (scattering), and the protein was recovered as a pellet by ultracentrifugation (20
174 min, 35 000 rpm, 45 Ti or 20 min, 80 000 rpm TLA 100.3 depending on the volume). The
175 protein pellet was rinsed with water, and resuspended in 11.9 mM LMNG, 20 mM Tris-HCl
176 pH 8.0 to reach a final protein concentration of 7-10 mg mL⁻¹. A different protocol is
177 described below for the thermal denaturation assays.

178

179 **FhuA crystallization**

180 Crystallization screening was carried out using commercial screens (Qiagen and Hampton
181 Research). Sitting drops, consisting of 100 nL protein and 100 nL crystallization buffer, were

182 dispensed in 96-well plates (Greiner Crystal Quick plates) using a Cartesian PIXSYS 4200
183 robot (Genomic Solutions) and equilibrated at 20 °C against 100 mL of crystallization buffer.
184 Hits were then manually reproduced and improved using the vapour diffusion hanging drop
185 technique. The drops, consisting of 0.8 mL protein and 0.8 mL crystallization buffer, were
186 equilibrated against 250 mL of crystallization buffer at 20 °C in 48-well plates (Hampton
187 Research). Crystals were transferred to the crystallization buffer supplemented with 20% (v/v)
188 glycerol for 30 s, flash-cooled and stored in liquid nitrogen.

189

190 **FhuA LMNG data processing**

191 The final data set was collected at 4 Å on beamline FIP-BM30A at the European Synchrotron
192 Radiation Facility, Grenoble, France [57]. Data reduction was performed using the XDS
193 program [58]. The space group was C222₁ (a = 149.53 Å, b = 210.96 Å, c = 188.70 Å) with
194 two molecules per asymmetric unit (solvent content of 73 %). The structure was solved by
195 molecular replacement performed with the Phaser program [59] using the model of the *E. coli*
196 protein available in the Protein Data Bank (code 2FCP). A refinement was carried out with
197 the *phenix.refine* program [60,61] in the 48-4 Å resolution range.

198

199 **SpNOX production**

200 SpNOX was overexpressed and purified in typically 0.025 mM LMNG, as previously
201 described [62]. When using DDM as the detergent, the same purification protocol was used
202 but with adjustments for DDM concentrations in the purification buffers corresponding to
203 each step. Indeed, the solubilization step was performed using 5.1 mM DDM, the loaded Ni-
204 HisTrap column was washed in the corresponding buffer containing 2 mM DDM, and eluted
205 with 0.3 mM DDM in the elution buffer. Final size exclusion was performed in 50 mM Tris-
206 HCl pH 7.0, 300 mM NaCl, 0.3 mM DDM.

207

208 **BmrA production**

209 Typical bacterial culture protocol: pET23b-*bmrA* [63] was used to transform C41(DE3) strain
210 of *E. coli*. For preculture, 200 mL LB medium containing 100 µg mL⁻¹ ampicillin was
211 inoculated with a freshly transformed colony and growth allowed overnight at 37 °C and 180
212 rpm. The next morning, enough preculture was added into 1 L 2X YT medium, containing 100
213 µg mL⁻¹ ampicillin, to get an optical density of 0.1, and growth then allowed at 37 °C 180
214 rpm. When the optical density reached 0.6, the culture was induced for the overexpression of
215 BmrA for 4 h at 25 °C and 180 rpm by the addition of 0.7 mM IPTG (final concentration).
216 Thereafter, the cells were harvested by centrifugation at 4,000 g at 4 °C for 20 min and the
217 bacterial pellet was frozen at -80 °C until further use. Inverted Membrane Vesicles
218 preparation : The bacterial pellet was resuspended in 50 mM Tris-HCl pH 8, 5 mM MgCl₂, 1
219 mM dithiothreitol, with an anti-protease cocktail from Roche. The bacterial cells were lysed by
220 passing them thrice through MicrofluidizerTM at 18,000 psi. Next, the cellular debris was
221 removed by centrifugation at 15,000 g for 30 min at 4 °C. The membrane fraction was
222 subsequently obtained by centrifugation at 150,000 g for 1 h at 4 °C. The membrane fraction
223 was resuspended and centrifuged again, with the same conditions as before, after resuspension
224 in 50 mM Tris-HCl pH 8, 1.5 mM EDTA, 1 mM dithiothreitol. Finally, the obtained membrane
225 vesicles were resuspended in 20 mM Tris-HCl pH 8, 300 mM sucrose, 1 mM EDTA and flash
226 frozen in liquid nitrogen before storage at -80 °C. The total protein concentration in
227 membrane was analysed by BCA assay. BmrA purification: BmrA enriched Inverted
228 Membrane Vesicles were solubilized for 1 h at 4 °C in 50 mM Tris-HCl pH 8, 10% glycerol,
229 100 mM NaCl, with 1% detergent (LMNG or DDM). The soluble fraction obtained after
230 ultra-centrifugation, at 150,000 g for 1 h at 4 °C, was injected into a 1 mL HisTrap HP
231 column (GE Healthcare) which was pre-equilibrated with the same buffer with 20 mM
232 imidazole, and 0.1% (2 mM) DDM or 0.01% (0.1 mM) LMNG. The column was washed with

233 20 column volumes of the same buffer. Gradient elution was eventually performed with the
234 same buffer with imidazole gradient from 20 mM to 500 mM. The protein fractions were
235 dialyzed overnight at 4 °C against 50 mM Hepes pH 8, 10% glycerol, 50 mM NaCl, 2 mM
236 DDM or 0.1 mM LMNG. The protein concentration, 2.2 mg mL⁻¹ for both BmrA in 2 mM
237 DDM and BmrA in 0.1 mM LMNG, was determined from UV absorbance at 280 nm by using
238 Nanodrop spectrophotometer, and a $\epsilon_{280\text{ nm}} = 38850\text{ M}^{-1}\text{ cm}^{-1}$. Reconstitution of BmrA in
239 proteoliposomes was performed as previously described [64].

240

241 **Limited Proteolysis of BmrA**

242 Apo form: Purified BmrA in detergent was added to buffer 50 mM Hepes-KOH pH 8, 50 mM
243 NaCl, 10% glycerol and the specified detergent, in case of trypsin, or 100 mM ammonium
244 bicarbonate pH 7.8 with the specified detergent, in case of protease V8. After 15 min of
245 incubation, either trypsin (1 µg/250 µg of protein) or protease V8 (1 µg/20 µg of protein) was
246 added. Samples of 10 µL (5 µg BmrA) were withdrawn at 0, 2, 5, 15, 30, 60, 120, 180 and
247 300 minutes. 2.5 µL of TFA 5% was added immediately to each sample to stop the reaction. 3
248 µL of Laemmli 5x was then added and the samples were placed in ice before resolving them
249 on SDS-PAGE. Vanadate-inhibited form: For the Vi-inhibited form, before the addition of
250 purified BmrA, the buffer was supplemented with 3 mM MgCl₂, 2 mM ATP and 1 mM Vi.
251 The samples were then incubated for 15 min before the addition of protease. Samples of 10
252 µL were withdrawn and treated thereafter in the same way as the apo form.

253

254 **bR solubilization and detergent exchange by sucrose gradient**

255 Purified purple membrane was solubilized for 40 h at 4 °C with 89 mM OTG (CMC = 9 mM)
256 at a membrane concentration of 1.5 g L⁻¹ in 20 mM sodium phosphate buffer, pH 6.8.
257 Samples were diluted to reach a final OTG concentration of 15 mM, supplemented with
258 2 mM of the surfactant to be tested, and incubated 15 min prior to being loaded onto a 10–30
259 % (w/w) sucrose gradient containing 20 mM sodium phosphate buffer pH 6.8 and 0.2, 2 or
260 20 mM LMNG. A control gradient contained 1 mM DDM. Gradients were centrifuged for
261 15 h at 55,000 rpm (200,000 g) in the TLS55 rotor of a TL100 ultracentrifuge (Beckman).
262 Bands containing the colored protein were collected with a syringe, and protein samples were
263 kept at 4 °C in the dark for UV-visible spectrophotometry. To check for detergent exchange,
264 10 µl of each recovered bR band was deposited onto a Thin Layer Chromatography (TLC)
265 silica plate (Macherey-Nagel, ref. 818423), and migration performed in a
266 Chloroform/Methanol/Water solvent (65/35/5). After drying, the plate was first stained with
267 iodine vapour and then with orcinol (0.1 % orcinol in 3 % H₂SO₄ and 72.5% ethanol) and
268 charring at 120 °C.

269

270 **Thermal denaturation and activity assays**

271 Thermal unfolding experiments and analysis were performed by differential scanning
272 fluorimetry coupled to back scattering using a Prometheus NT.48 instrument (Nanotemper
273 Technologies, Munich, DE), and the provided software PR.thermocontrol v2.0.4. Up to 48
274 capillary containing 10 µL of sample are sequentially illuminated at 280 nm, and fluorescence
275 intensity at 350 (F350) and 330 (F330) nm, and back scattering measured as a function of
276 temperature. The derivatives of F350/F330 and of the back scattering were used to estimate
277 the melting temperature, T_m , and the onset of aggregation, T_{agg} , respectively. For FhuA :
278 samples were prepared by diluting 48 times a stock sample of 1.92 mg mL⁻¹ FhuA, in 2.2 mM
279 LDAO, 20 mM Tris pH 8.0, ~150 mM NaCl, into 20 mM Tris pH 8.0 with the appropriate
280 detergent, leading to a final residual LDAO concentration of ≈ 0.05 mM (0.05 CMC), with
281 FhuA at 0.04 mg mL⁻¹. For SpNOX : samples were prepared after purification in LMNG, and
282 eventual detergent exchange in DDM, and concentration. Buffer is 50mM Tris HCl pH 7.0,
283 300 mM NaCl, 10 µM FAD, with 0.025 mM LMNG or 0.2 mM DDM initial concentrations,

284 0.21 LMNG and 1.27 mM DDM final estimated concentrations, considering co-concentration
285 of protein and detergent, and SpNOX at 8.3 mg mL⁻¹, in the T_m assays. For activity assays,
286 SpNOX was diluted to 1 µg mL⁻¹ in 50 mM Tris HCl pH 7.0, 300 mM NaCl, 10 µM FAD ,
287 with 0.025 mM LMNG or 0.2 mM DDM. Activity assays were performed in the presence of
288 100 µM Cytochrome C, 10 µM FAD, 200 µM NADPH, following Cytochrome C absorbance
289 at 550 nm. For BmrA : for T_m measurements, samples were prepared at a final protein
290 concentration of 1 mg mL⁻¹, from dilution of the purified protein in 50 mM Hepes pH 8, 10%
291 glycerol, 50 mM NaCl, with 2 mM DDM or 0.1 mM LMNG. For Vi-inhibited form of BmrA,
292 the dilution buffer also contained 10 mM ATP, 10 mM MgCl₂ and 1 mM Vi (all final
293 concentrations). BmrA samples were incubated for 15 min at room temperature before
294 allowing them to be filled in the capillaries for Nano DSF analysis. The activity assays of
295 BmrA were done in a quartz cuvette with a final volume of 700 µL. The buffer, containing 50
296 mM Hepes/KOH pH 8, 10 mM MgCl₂, 4 mM phosphoenolpyruvate, 0.3 mM NADH, 32 µg
297 mL⁻¹ lactate dehydrogenase, 60 µg mL⁻¹ pyruvate kinase, 10 mM ATP and either 2 mM DDM
298 or 0.1 mM LMNG, was added into the cuvette and was allowed to attain the desired
299 temperature i.e. 37 °C for 5 min, before adding 3 µg protein, and measuring the absorbance at
300 340 nm for 20 min at 37 °C. For bR : to perform T_m measurements, after the solubilization
301 step (see above), solubilized bR was incubated 40 mins at 4 °C in the presence of biobeads
302 (10 g/g OTG), that reduced the concentration of OTG to 70 mM estimated from TLC, from
303 the comparison with OTG aqueous solution at 10, 25, 50, 75 and 100 mM. TLC was
304 performed as described above in bR preparation, but the mobile phase was
305 Chloroform/Methanol 2/1, and the volume deposited was 1 µL. Imaging and quantification
306 were done with the Gel Doc XR system and Image lab software (Biorad). The bR was then
307 diluted 18 times in 0.1 M NaCl, 0.02 M Na phosphate buffer supplemented with the
308 appropriate detergent, residual OTG concentration being estimated as 0.4 its CMC, and final
309 bR concentration to 0.14 mg mL⁻¹. In the thermal denaturation assays, the temperature was
310 increased by 1 (FhuA, bR, SpNOX) or 0.5 (BmrA) °C/min from 15 °C (FhuA, bR, SpNOX)
311 or 20 °C (BmrA) up to 95 °C.

312

313 **Density measurements**

314 31.8 mg of dried LMNG was dissolved in 1.99948 g of water. From the density of water
315 (0.998205 +/- 0.000001 g mL⁻¹) and sample (1.001416 +/- 0.000002 g mL⁻¹) determined from
316 triplicate measurements at 20 °C using a DMA 58 density meter (Anton Paar, Graz, Austria),
317 we derived [65] a LMNG concentration of 15.68 mg mL⁻¹ and a partial specific volume $\bar{v} =$
318 0.797 mL g⁻¹.

319

320 **AUC-SV**

321 AUC-SV experiments were conducted in an XLI analytical ultracentrifuge (Beckman, Palo
322 Alto, CA) using an ANTi-50 rotor, using double channel Ti centre pieces (Nanolytics,
323 Germany) of 12, 3, or 1.5 mm optical path length equipped with sapphire windows, with the
324 reference channel being typically filled with solvent without detergent. For BmrA, LMNG
325 was in the buffer in the reference channel, and we used double sector capillary type cells
326 (Beckman, Palo Alto, CA) allowing perfect matching of the channel heights upon
327 centrifugation 90 min at 7,000 rpm (3,600 g). Acquisition was done at 42,000 rpm (130,000
328 g), overnight, using absorbance (280 nm, and additionally for SpNOX, 412nm) and
329 interference detection. Data processing and analysis was done using the program SEDFIT
330 [66] from P. Schuck (NIH, USA), REDATE [67] and GUSSE [68] from C. Brautigam (USA),
331 and using standard equations and protocols described in [65,69–71]. LMNG samples in water
332 were prepared from precise weight dilution from a stock solution at 9.88 mM. The
333 sedimentation velocity profiles for LMNG at 23.2 and 35 mM in Buffer FhuA showed an

334 optical artefact, and s -values were determined from $(r_b/r_m)=s\omega^2t$, with r_b the radial position at
335 half the plateau signal, r_m the meniscus position, ω the angular velocity, and t the time.
336 Corrected s -values for solvent density and viscosity, s_{20w} , are calculated, for membrane
337 proteins, considering for their partial specific volume, the mean value between protein and
338 detergent.

339

340 **Dynamic light scattering (DLS) of LMNG**

341 LMNG at 20 mM in water was measured using the DynaPro NANOSTAR (Wyatt, Santa-
342 Barbara, USA) instrument, at room temperature, and analyzed with the associated software
343 DYNAMICS.

344

345 **SEC-LS**

346 SEC-LS experiments were conducted at 4 or 6 °C on a HPLC consisting of a degasser DGU-
347 20AD, a LC-20AD pump, an autosampler SIL20-AC_{HT}, a communication interface CBM-
348 20A and a UV-Vis detector SPD-M20A (Schimadzu, Kyoto, Japan), a column oven XL-
349 Therm (WynSep, Sainte Foy d'Aigrefeuille, France), a static light scattering miniDawn Treos,
350 a dynamic light scattering DynaPro NANOSTAR, and a refractive index Optilab rEX
351 detectors. The analysis was made with the software ASTRA, v5.4.3.20 (Wyatt, Santa-
352 Barbara, USA). Samples of 10 to 60 μL were injected at 0.5 mL min^{-1} on a Superdex 200
353 10/300 GL (GE Healthcare), equilibrated with the elution buffer. Bovine serum albumin at
354 2 mg mL^{-1} in PBS buffer was injected as a control in each experiment. The extinction
355 coefficient and refractive index increments for the proteins were calculated from the amino-
356 acid sequences using the program SEDFIT described above. For LMNG, we used
357 $\partial n/\partial c=0.146 \text{ mL g}^{-1}$ determined by AUC.

358

359 **SAXS and SANS samples**

360 A first series of SAXS measurements was carried out with 14 samples of LMNG in H_2O , at
361 concentrations between 0.31 and 20 mM, and a second series, measured by SAXS and SANS,
362 with four samples of LMNG in H_2O and in D_2O , at 20 and 10 mM. SANS contrast variation
363 determination was achieved for LMNG at 20 mM in 25 mM Tris pH 8, 150 mM NaCl, 10%
364 glycerol, with different D_2O %s. LMNG concentrations were precisely determined using
365 weighing-controlled solubilization and dilutions.

366

367 **SAXS experiment**

368 SAXS experiments were conducted on the BM29 beamline at the European Synchrotron
369 Radiation Facility (Grenoble, France). The data were recorded for $0.004 < Q < 0.5 \text{ \AA}^{-1}$
370 ($Q=(4\pi/\lambda)\sin\Theta$ is the modulus of the scattering vector, with 2Θ being the scattering angle,
371 and λ the wavelength), using a two-dimensional 1M Pilatus detector, at 20 °C, with a
372 monochromatic X-ray beam with $\lambda = 0.9919 \text{ \AA}$ and a sample to detector distance of 2.864 m.
373 Measurements were performed with 50 μL loaded sample, in a quartz capillary, with a
374 continuous flow. In the first series, 10 acquisitions with 1 s irradiation per acquisition (flow of
375 2.7 $\mu\text{L/s}$), in the second series, 10 and 30 acquisitions with 0.5 s irradiation (flows of 5 and 1
376 $\mu\text{L/s}$), were recorded for the samples and the solvents. The scattering curves were
377 indistinguishable with the two flows. Data reduction was performed using the automated
378 standard beamline software (BSxCuBE) [72], and data processing, including the elimination
379 of data suffering from radiation damage, averaging, buffer subtraction, Guinier plots, using
380 PRIMUS of the software suite ATSAS [73]. Absolute scales were obtained using the
381 scattering of water.

382

383 **SANS experiment**

384 SANS experiments were carried out on the small-angle diffractometer D22 at the Institut
 385 Laue Langevin (Grenoble, France), at 20 °C, using neutron wavelengths λ of 6 and 12 Å, with
 386 samples measured in Hellma quartz cells 100QS with 1-mm optical path length. Scattering
 387 data for LMNG at 10 and 20 mM in H₂O and D₂O were recorded at 20 °C for $0.003 < Q <$
 388 0.45 \AA^{-1} , using a neutron wavelength λ of 6 Å, and sample-detector/collimation distances (SD
 389 / coll) of 17.6 m / 17.6 m, 5.6 m / 5.6 m, and 1.4 m / 2.8 m for 15, 5 and 5 mins, respectively.
 390 For LMNG at 20 mM in D₂O, additional data were recorded at (SD/coll) of 17.6 m / 17.6 m at
 391 12 Å, leading to a total Q -range of 0.0012-0.45 Å⁻¹. Transmissions were measured at 17 m for
 392 1 min. For the determination of the contrast match point, samples were measured at 10 °C,
 393 with SD / coll of 5.6 m / 5.6 m, 1.4 m / 2.8 m, for 5 and 1 min. respectively, and transmissions
 394 were measured at 5.6 m for 1 min. LMNG at its match point, in 21.4 % D₂O, was measured at
 395 15 °C, with SD / coll of 8 m / 8 m and 1.4 m / 2 m for 60 and 15 min, respectively. Data
 396 reduction to obtain scattering curves in absolute scale, and manipulation -merging, buffer
 397 subtraction- were done in the usual way (see *e.g.*[74]) using Grasp (a MatlabTM script
 398 application produced by ILL) and SANS reduction macros provided by NIST Center running
 399 on IGOR (Wavemetrics)[75]. Absolute intensities are obtained using the direct beam
 400 measurement.

401

402 **SAXS and SANS Guinier analysis**

403 The radii of gyration (R_g) and the intensities scattered in the forward direction ($I(0)$) were
 404 extracted by the Guinier approximation, with $R_g Q \leq 1.3$. Aggregation number were derived
 405 from the molar mass of LMNG micelle, M (g mol⁻¹) obtained from the absolute forward
 406 intensity, $I(0)$ (cm⁻¹), using $M = (I(0)/c)N_A/(\partial\rho_{el}/\partial c)^2$, in SAXS, and $M = (I(0)/c)N_A/(\partial\rho_N/\partial c)^2$,
 407 in SANS. N_A is Avogadro's number, c the concentration (g mL⁻¹), and $\partial\rho_{el}/\partial c$ and $\partial\rho_N/\partial c$ (cm
 408 g⁻¹) the increment of electron and neutron, respectively, scattering length density per g of
 409 LMNG. $\partial\rho_{el}/\partial c = (\rho_{el} - \rho_{el}^o) r_{el} \bar{v}$, with $\rho_{el} = 4.105 \cdot 10^{23}$ (from composition and \bar{v}) and $\rho_{el}^o =$
 410 $3.297 \cdot 10^{23}$ the numbers of electron per mL of LMNG and solvent, respectively, and $\rho_{el} =$
 411 $2.818 \cdot 10^{-13}$ cm the scattering length of an electron. $\partial\rho_N/\partial c$ was calculated to be $1.108 \cdot 10^{10}$ cm
 412 g⁻¹ in H₂O, and $-3.509 \cdot 10^{10}$ cm g⁻¹ in D₂O.

413

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415

416 **SAXS and SANS combined analysis for LMNG shape**

417 SAXS and SANS data were analyzed using shape-dependent models in SASview [76]. The
 418 scattered intensity from an homogeneous dispersion of particles is written as:

$$419 \quad I(Q) = N V^2 (\Delta\rho)^2 P(Q) S(Q)$$

420 Where N is the number density of particles, V their volume, $\Delta\rho$ (cm⁻²) is the scattering
 421 contrast, $P(Q)$ is the inter particle form factor and $S(Q)$ is the interparticle structure factor.

422 In the present case, SAXS and SANS data have been fitted simultaneously with a core shell
 423 cylinder form factor and hard sphere potential with beta approximation using SASview
 424 software. The cylinder form factor included size polydispersity on radius (fixed at 5 %) using
 425 a gaussian distribution. The scale factor was fixed by the surfactant concentration. The head
 426 groups SLD or ED were fitted using the same hydration level of the anhydrous polar head.
 427 Radius, length and shell thickness of the cylinder were constrained and kept equal for X-ray
 428 and neutrons. Fitting was performed using the optimizer DREAM which allows to set
 429 simultaneous fits as well as proper error determination.

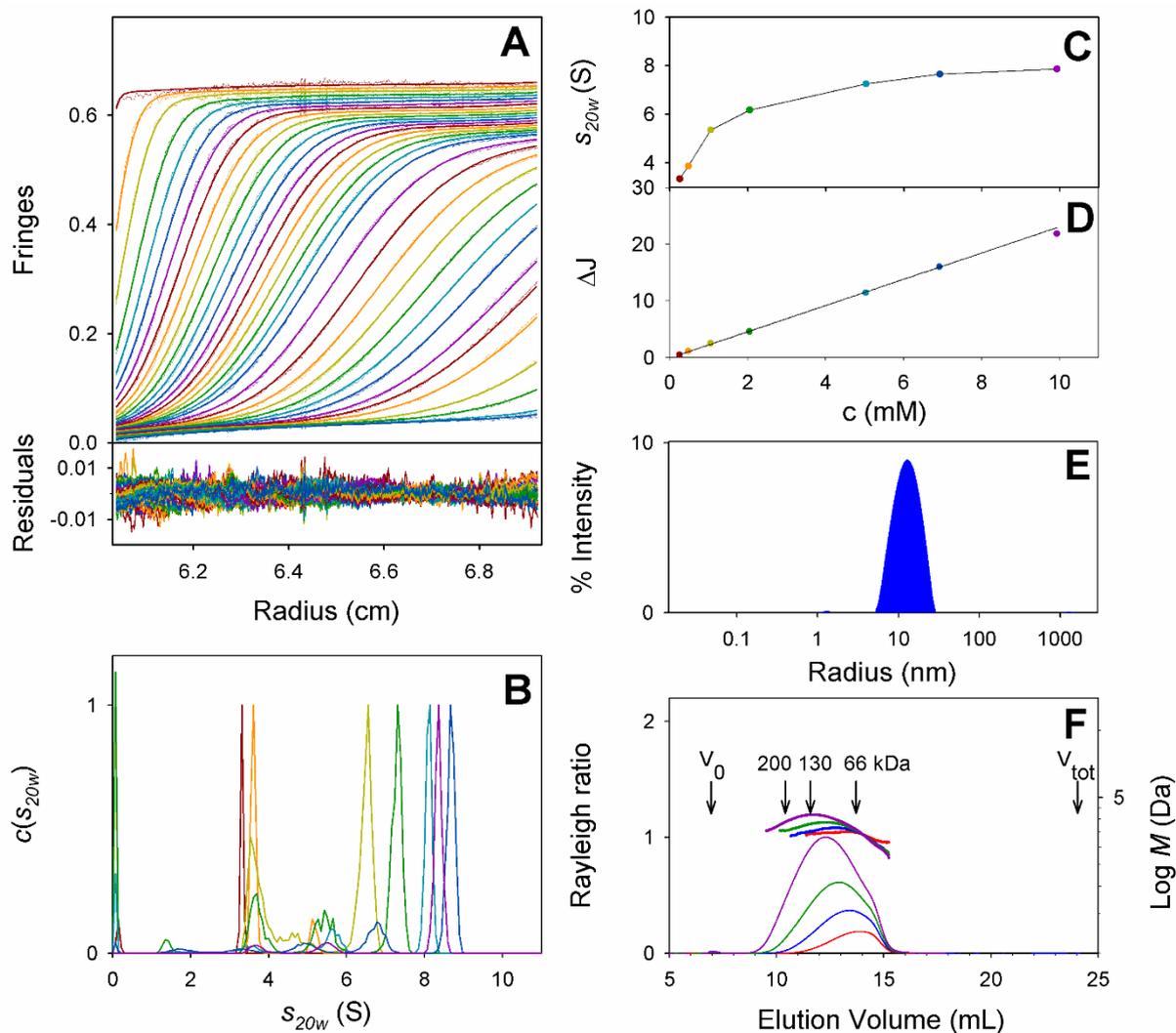
430

431 **Results**

432

433 ***Hydrodynamic characterization of LMNG***

434
435 Our first aim was to characterize the assembly properties of LMNG molecules by itself.
436 AUC-SV experiments were done for LMNG solubilized in water at different concentrations.
437 The SV profiles and analysis in terms of continuous distributions $c(s_{20w})$ of sedimentation
438 coefficients, s_{20w} , showed a unique or main boundary. The mean s_{20w} - value increased with
439 concentration, from 3.3 S at 0.25 mM to 7.9 S at 9.8 mM (Fig 1A-C). This indicates that
440 LMNG micelle aggregation number increases with concentration, an atypical behaviour
441 compared to usual detergents. Complementary AUC-SV in a dilute buffer (20 mM Tris-HCl
442 pH 8.0, 150 mM NaCl), at larger LMNG concentrations of 23 and 35 mM, gave estimates for
443 s_{20w} of 7.7 and 8.7S, respectively. By integrating the signal for the micelles at the different
444 LMNG concentrations (Fig 1D), we determine, in water, a refractive index increment for
445 LMNG $\partial n/\partial c=0.146 \text{ mL g}^{-1}$ (close to that of DDM, 0.143 mL g^{-1} [65]), and a CMC of 40 μM .
446 The knowledge of the partial specific volume, \bar{v} , of LMNG is required to correlate s_{20w} and
447 molar mass, or aggregation numbers N_{agg} . We measured, with density measurement, $\bar{v} = 0.797$
448 mL g^{-1} , a value close to that of DDM: 0.82 mL g^{-1} from [65] and $0.81\text{-}0.837 \text{ mL g}^{-1}$ from [1].
449
450 Minimum aggregation numbers N_{agg} of 63 and 230 are calculated at 0.25 and 10 mM
451 considering globular compact micelles (frictional ratio of 1.25), but N_{agg} are larger, at least for
452 the larger micelles, since their shape is anisotropic (see below). Dynamic light scattering
453 (DLS) of 20 mM LMNG in water showed essentially one population with a polydispersity
454 index of 35% and a hydrodynamic radius, R_H , of 14 nm (Fig 1E), in line with the reported
455 value of 7.2 nm at 5 mM [49]. Combining s_{20w} and R_H values at 5 and 20 mM provides N_{agg}
456 estimates of 300 and 620, corresponding to frictional ratios of 1.65 and 2.4, indicating
457 moderately and strongly elongated shapes, respectively.
458
459 In SEC-LS experiments with 0.05 mM LMNG in the equilibration and elution buffer
460 (bufferA), LMNG injected at 2, 5, 10 and 20 mM eluted as a very large peak covering the
461 elution volumes of BSA monomer (65 kDa) to BSA trimer (Fig 1F). When LMNG
462 concentration increased, the elution volume decreased. However, the molar masses
463 determined from coupled refractive index and light scattering detections did not change
464 significantly, neither along the elution, nor with the injected LMNG concentration: from 67
465 kDa ($N_{\text{agg}} = 67$) at 2 mM to 76 kDa ($N_{\text{agg}} = 76$) at 20 mM (Fig 1F). This suggests the
466 dissociation of the largest LMNG complexes upon sample dilution on the column.
467



468
 469 **Fig 1. LMNG in AUC-SV, DLS, and SEC-LS**
 470 AUC-SV: **A:** Superposition of experimental and fitted sedimentation velocity profiles (top) and residuals
 471 (bottom), obtained at 130,000 g and 20 °C during \approx 5 hours using interference optics and 12 mm optical path
 472 centerpiece, for LMNG at 0.25 mM in water. **B:** Superposition of the normalized $c(s)$ distributions for LMNG at
 473 0.25 (brown), 0.5 (orange), 1 (yellow-green), 2 (green), 5 (cyan), 7 (blue), and 10 (purple) mM. **C:** plot of the
 474 mean sedimentation coefficient s_{20w} versus LMNG concentration. Note that the mean s_{20w} is obtained from the
 475 integration of the $c(s_{20w})$ over the whole s_{20w} -range; At 10 mM compared to 7 mM, as seen in Panel B, s_{20w}
 476 for the main species is decreased, while, as seen in Panel C, the mean s_{20w} is slightly increased. **D:** Fringe
 477 number ΔJ , normalized to 1 cm optical path-length, for the micelle contributions vs total LMNG concentration,
 478 and linear fit. DLS: **E:** Hydrodynamic radius distribution of LMNG at 20 mM in water. SEC-LS: **F:** Rayleigh
 479 ratio measured along the elution profiles, for 30 μ L LMNG injected at 2, 5, 10, and 20 mM (red, blue, green, and
 480 purple thin lines, respectively) on a Superdex 200 10/300 GL (GE Healthcare) column eluted with 50 mM Tris-
 481 HCl pH 7.0, 300 mM NaCl, 0.05 mM LMNG, and molar masses (thick lines, right axis) derived from the light
 482 scattering and refractive index detections. V_0 is the dead volume, V_{tot} the total volume of the column, and the
 483 arrows indicate the elution volumes of BSA as a monomer, dimer, and trimer (66, 130, and 200 kDa) from a
 484 different injection.

485
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487 **Structural characterization of LMNG by small angle scattering**

488 SAXS and SANS experiments were performed to get additional information on the size and
 489 shape of LMNG micelles. A first set of SAXS experiments were done with LMNG between
 490 0.3 and 20 mM (0.03 and 2%) (Fig 2A). From the Guinier analysis, the size of the micelle
 491 increases with R_g and aggregation numbers increasing from 3.4 nm to 10 nm, and from 55 to
 492 240, respectively, when LMNG concentration is raised from 0.3 to 10 mM. (Fig 2B and 2C).

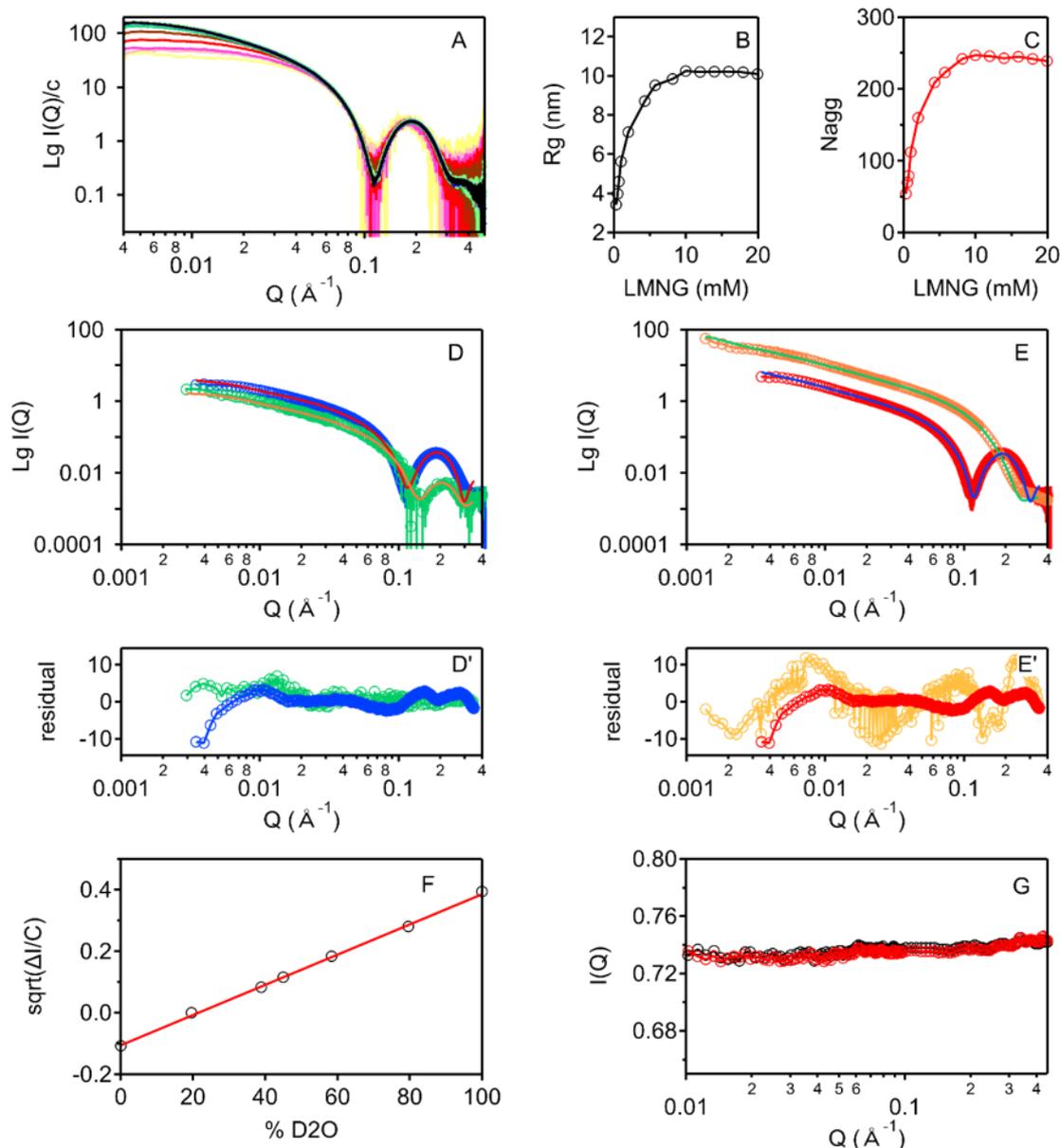
493 These values remain constant for LMNG concentrations increasing from 10 to 20 mM, and
494 indeed the scattering curves are superimposed (Fig 2A). All curves are superimposed at the
495 largest angles, representing the smallest distances in the real space, compatible with a circular
496 cross section. To ascertain the shape of the LMNG micelles, 10 and 20 mM LMNG samples
497 were measured by SAXS and SANS, in H₂O and D₂O. In the same buffer, the scattering
498 curves normalized by concentration were superimposable for the 10 and 20 mM LMNG
499 samples. From Guinier plots, the radii of gyration from SAXS were 11.5 and 11.6 nm at 10
500 and 20 mM in H₂O, and 17.7 and 17.2 nm at 10 and 20 mM in D₂O. The aggregation numbers
501 were 285 in H₂O, and 528 in D₂O. The R_g and aggregation numbers from SAXS, in H₂O were
502 slightly larger but similar to those of the first set of experiments. The size of the micelles
503 appears significantly larger here for LMNG in D₂O than in H₂O, but, as will be discussed
504 below, the significance of this difference is not ascertained.

505
506 The SAXS and SANS scattering curves of 20 mM LMNG in H₂O and D₂O were analyzed
507 globally. We used a model of a core shell cylinder with hard sphere interaction, defined by a
508 core representing the hydrophobic tails (*i.e.* the hydrocarbon chains) covered by a shell
509 representing the hydrated polar heads. The scattering length densities (SLD) of the core and of
510 the solvent were fixed to the theoretical values. The SLD of the polar head (different in SAXS
511 and SANS), and the dimensions of the micelles (radius of the cylinder core, thickness of the
512 shell, and length, common in SAXS and SANS) were globally fitted keeping the same
513 hydration level in X-ray and neutron. The results are presented in Table 1. The fitted values
514 for the SLD of the anhydrous shell values are in the same order of magnitude as those
515 calculated for the anhydrous polar head. We extracted reasonable hydration of 30 % in H₂O
516 and 44 % in D₂O. The radius of the hydrophobic core, 14 Å, and that of the shell thickness, 7-
517 9 Å, are in line with those expected from the LMNG size.

518
519 Finally, we investigated whether LMNG could be totally matched out in SANS. This is
520 required to be able to focus on protein contribution. The differential distribution of detergent
521 hydrophobic tails and hydrophilic heads in the detergent micelles and bound detergent indeed
522 corresponds to ordered scattering density fluctuations, which makes the scattering curve of
523 the micelles and bound detergent possibly not flat even at the match point [77]. We thus
524 measured for LMNG a set of contrast variation curves, using a solvent with different %s of
525 D₂O. We determined a contrast match point of 21.4 % D₂O (Fig 2F). The scattering curve of
526 the 2 mM LMNG sample in 21.4 % D₂O perfectly superimposes that of the solvent (Fig 2G).
527 Thus LMNG can be homogeneously masked. The analysis of the neutron scattering curves of
528 membrane proteins solubilized in LMNG, in 21.4 % D₂O will not require the fastidious step
529 of detergent modelling to access membrane protein envelope structure. Because the contrast
530 match point of hydrogenated proteins is \approx 44 % D₂O, membrane proteins should be deuterated
531 to improve their scattering signal, but this is not problematic, since deuterating proteins, in
532 general, is not an issue.

533
534 To summarise, SAXS and SANS showed that LMNG forms large elongated rod-like micelles
535 above the mM concentration. Given this observation, the question of the homogeneity of
536 different membrane proteins at various LMNG concentrations was addressed below.

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Fig 2. SAXS and SANS analysis of LMNG

A: Superposition of 13 SAXS profiles obtained at 0.3 (yellow), 0.5 (pale pink), 0.7 (pink), 1 (red), 2 (brown), 6 (green), 8 (light green), 10 (light blue), 12 (blue), 14 (navy), 16 (purple), 18 (grey), and 20 (black) mM LMNG, in H₂O, at 10 °C. B: corresponding R_g , and, C: aggregation numbers, versus LMNG concentrations. D,E: Superposition, for LMNG at 20 mM in H₂O (D), and D₂O (E) of the experimental scattering curves obtained by SAXS (blue and red dots, respectively) and SANS (green and orange dots, respectively), and of the fitted curves, in the model of a core-shell cylinder, in SAXS (red and blue lines in H₂O and D₂O, respectively), and in SANS (orange and green lines lines in H₂O and D₂O, respectively); D',E': respective normalized by data error residuals. SAXS and SANS data in each solvent were globally fitted (Table 1). F: Match point determination from SANS scattering curves obtained for 20 mM LMNG in 0, 20, 40, 45, 60, 80, and 100 % D₂O. G: Superposition of raw scattering curves obtained in a solvent with 21.4 % D₂O for LMNG at 2 mM (black) and of the solvent alone (red).

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Table 1. Rod analysis of SAXS and SANS data for 20 mM LMNG

	LMNG 20 mM in H ₂ O		LMNG 20 mM in D ₂ O	
	SAXS	SANS	SAXS	SANS
Q range (\AA^{-1})	0.0035-0.35	0.0030-0.35	0.0035-0.35	0.0015-0.35
χ^2_{R}	3.2	3.2	2.3	26.7
Background, fixed (cm^{-1})	0.0008	0.001	0.0012	0.0015
SLD solvent, fixed (10^{-6}\AA^{-2})	9.4	-0.56	9.4	6.4
SLD core, fixed (10^{-6}\AA^{-2})	8.3	-0.3	8.3	-0.3
SLD shell, fixed (10^{-6}\AA^{-2})	14.75	1.88	14.75	3.99
SLD shell, fitted (10^{-6}\AA^{-2})	13.12 \pm 0.02	1.134*	12.42 \pm 0.02	5.048*
Fitted shell hydration (%)	30		44	
Radius (\AA)**	14.51 \pm 0.02		13.29 \pm 0.01	
Thickness (\AA)	7.33 \pm 0.04		9.25 \pm 0.03	
Length (\AA)	603 \pm 7		5376 \pm 16	
Vol fraction	0.02 \pm 0.00001		0.01951 \pm 0.00006	

χ^2_{R} : reduced χ^2 . *: linked parameter. **: a polydispersity on radius width was fixed to 5%.

FhuA-LMNG assemblies

FhuA is an *E. coli* outer membrane ferrichrome-iron transporter involved in bacteriophage infection [78,79]. The structure of this robust β -barrel protein was solved in 1998 [80]. We investigated FhuA stability, homogeneity and association state at different LMNG concentrations, and whether the protein can crystallize in LMNG.

We first compared the thermal stability of FhuA in LDAO and in LMNG by DSF, allowing to measure the melting temperature (T_m) of the protein by probing the fluorescence emission ($F_{350\text{nm}}/F_{330\text{nm}}$ ratio) of the aromatic residues upon increasing temperature. Simultaneous light back-reflection measurements permit to detect qualitatively aggregation events. Detergent was exchanged by a 48 times dilution, leading to a final residual concentration of LDAO that was much below its CMC (20 times). The melting curves are similar whatever the final detergent concentration, corresponding to CMC + 0.2, 0.5 mM and 1-2 mM, *i.e.* LDAO at 1.2, 1.5, and 2.2 mM and LMNG at 0.21, 0.51 and 2.01 mM. They show (Fig 3A) two unfolding events, attributed to, first, the unfolding of the cork, and then of the barrel [81] (both domains contain tryptophans). From light back-reflection, the onset of protein aggregation coincides with the first T_m , and significant aggregation to the second T_m . In LDAO, the T_m at 63 and 69 °C are in line with the published values of 60-65 and 74-75 °C, from scanning calorimetry [81], or synchrotron radiation circular dichroism [78] in slightly different buffers with 1.3 or 4.4 mM LDAO. In LMNG, unfolding is shifted to higher temperatures. The first T_m is at 67 °C, showing a moderate stabilization (+ 4 °C) of the cork, the second one at 80 °C indicating a significant stabilization (+ 11°C) of the barrel. T_m values are reported in Table 2. For comparison, binding of ferrichrome, the natural ligand, was described to have a strong stabilizing effect on FhuA cork, shifting the first transition by 6 °C [81]; binding of phage T5 Receptor Binding Protein pb5 leads to one unique unfolding event at 89 °C (+ 15 °C vs T_m of barrel for FhuA alone) [78]. Thus, LMNG significantly stabilizes both the flexible parts and the rigid domain of this β -barrel protein.

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Table 2. Melting temperatures of FhuA, SpNOX, BmrA, and bR

Protein	Detergent type	detergent conc. (mM)	Micelle conc. (mM)	T_m (°C)	T_{agg} (°C)
FhuA	LDAO	1.2, 1.5, 2.2	0.2, 0.5, 1.2	63.0 + 69.0	65
FhuA	LMNG	0.21, 0.51, 2.0	0.2, 0.5, 1.0	67.0 + 80.0	74
SpNOX	DDM	1,27	1.1	40.5	56
SpNOX	LMNG	0.21	0.2	53.9	60
BmrA Apo	DDM	2	1.8	41.2	54
BmrA Apo	LMNG	0.1	0.1	45.7	70
BmrA Vi-inh	DDM	2	1.8	46.4	45
BmrA Vi-inh	LMNG	0.1	0.1	59.3	50
bR	OTG	11	2.0	53.6	43
bR	DDM	2.01	1.8	59.9	52
bR	LMNG	2.17	2.2	65.9	63

632 T_m : melting temperature measured by differential scanning fluorimetry; T_{agg} : onset temperature for aggregation from light
633 back reflexion. BmrA Vi-inh, Vi-inhibited forms were incubated with 10 mM ATP, 10 mM MgCl₂, and 1 mM vanadate.
634 Standard deviations of typically 0.2 were determined from triplicates experiments in BmrA, otherwise the precision on T_m is
635 estimated at 1°C. Micelle concentrations (*i.e.* above the CMC concentrations) were calculated considering CMC-values of 1,
636 0.01, 0.17 and 9 mM for LDAO, LMNG, DDM, and OTG, respectively.

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For FhuA, detergent exchange can easily be done, by dilution of the purified protein solution until FhuA precipitates and re-solubilization of the pellet in the desired detergent. The resulting sample in LMNG, also used for crystallization assays, contained $\approx 7.5 \text{ mg mL}^{-1}$ FhuA and 12 mM (total concentration) LMNG. SV-AUC analysis shows a main contribution ($\approx 75\%$ of the absorbance) at $s_{20w} = 7.8 \text{ S}$, *i.e.* the same value as that published for FhuA in DDM [82]. Additional contributions are detected at $s_{20w} = 10.7 \text{ S}$ ($\approx 5\%$) and 6.2 S (15-20%), which correspond reasonably to FhuA dimer and LMNG free micelles, respectively (Fig 4AB). Analysis of the absorbance of interference fringe signals provides an estimate of 1 g g^{-1} (80 mol mol^{-1}) of bound LMNG, and a free micelle concentration of 4.5 mM. In order to investigate the effect of LMNG concentration on FhuA homogeneity, complementary SV-AUC experiments were done on a similarly prepared sample with nominal concentrations of 5.4 mg mL^{-1} FhuA and 13 mM LMNG, and two derived samples, with FhuA diluted twice, and increased LMNG nominal concentrations to 18 and 24 mM. For these three samples too, only one main boundary is observed ($> 80\%$) at about 7 S (Table 3). Probably, due to the large total concentrations of LMNG and FhuA, the fit is poor and the s_{20w} values imprecise (data not shown). The data can be interpreted by the non-ideal co-sedimentation of FhuA complexes and of LMNG free micelles, sedimenting at nearly the same s_{20w} value (Table 3). Nevertheless the membrane protein appears to remain monomeric in the presence of even very large concentrations of LMNG, where the latter forms large micelles. SEC-LS performed on a similar sample injected after 10 times dilution, showed traces of aggregates, FhuA dimer as a shoulder, FhuA monomer as the main contribution at 12.3 mL, and LMNG micelles at 14.5 mL (Fig 4C). The molar mass analysis gives for the main peak 79.3 kDa for FhuA molar mass contribution, in agreement with the protein sequence (79.9 kDa), and 98.5 kDa for LMNG contribution, corresponding to 1.2 g g^{-1} (95 mol mol^{-1}) bound detergent.

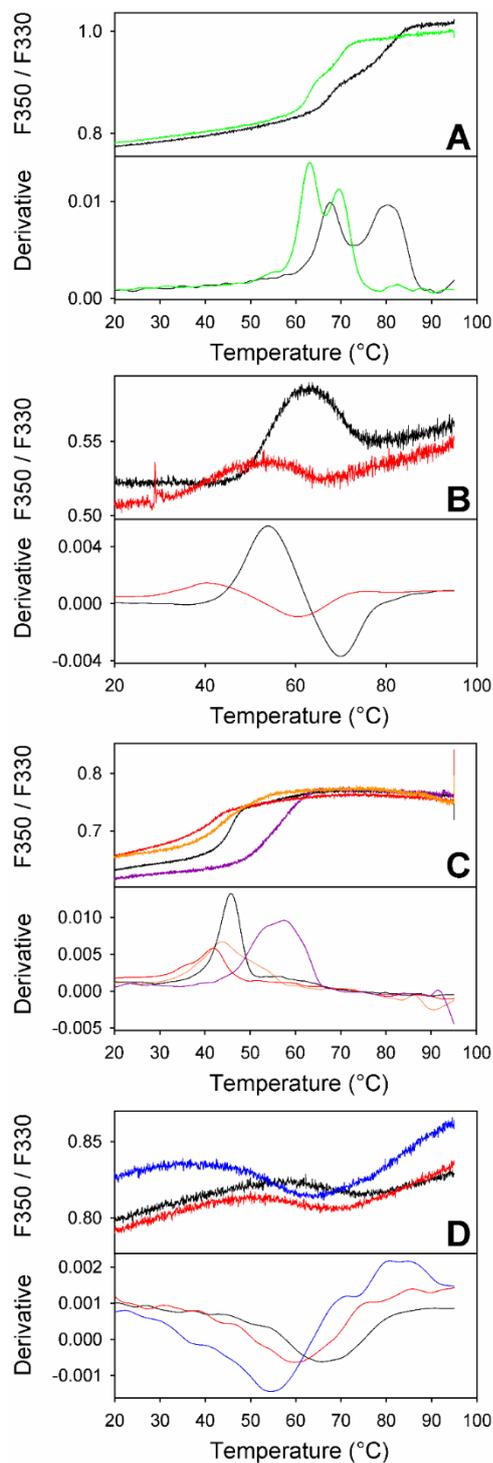


Fig 3. Thermal denaturation of FhuA, SpNOX, BmrA, and bR, by differential scanning fluorimetry

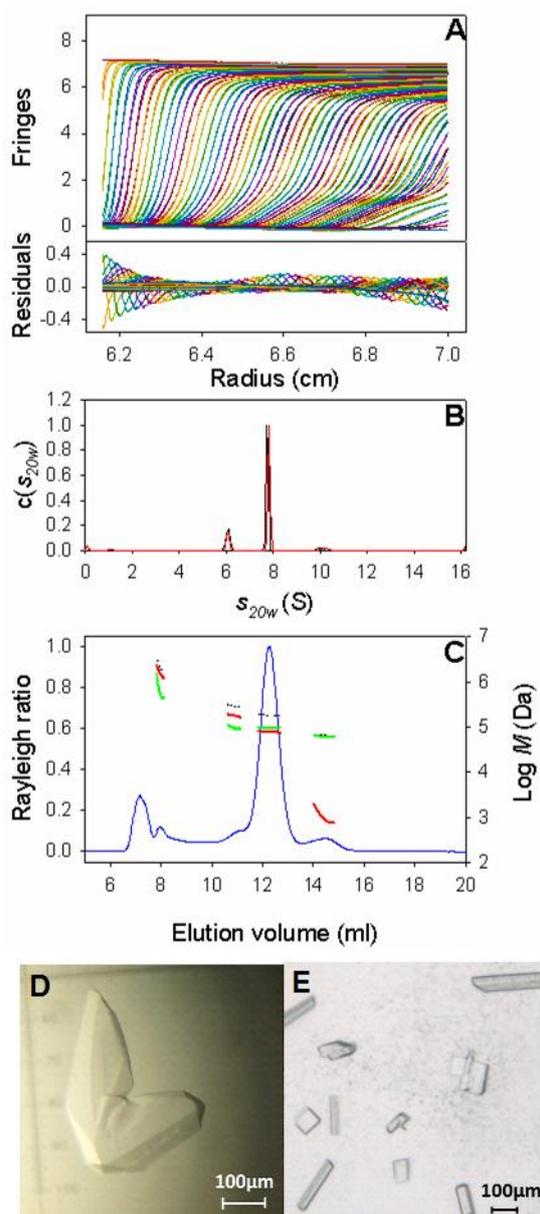
Ratio of the fluorescence emitted at 350 and 330 nm (top panels), and derivative (bottom panels) for **A:** FhuA at 0.04 mg mL^{-1} in the presence of LDAO 1.2 mM (green) or LMNG 0.21 mM (black); **B:** SpNOX at 8.3 mg mL^{-1} in the presence of DDM 1.27 mM (red) or LMNG 0.21 mM (black); **C:** BmrA at 1 mg mL^{-1} , in DDM 2 mM , in the Apo (red), or Vi-inhibited (orange) forms, and in LMNG 0.1 mM , in the Apo (black), or Vi-inhibited (purple) forms. The Vi-inhibited forms were incubated with 1 mM Vi, 10 mM ATP and 10 mM MgCl_2 for 15 minutes. **D:** bR at 0.14 mg mL^{-1} in OTG 11 mM (blue), DDM 2.17 mM (red), or LMNG 0.21 mM (black).

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Table 3. LMNG solubilized FhuA in AUC-SV

exp	FhuA (mg mL^{-1})	total LMNG (mM)	free LMNG ^a (mM)	Main s_{20w} (S) from A_{280} ^b	Main s_{20w} (S) from ΔJ ^b	s_{20w} (S) for LMNG ^c
1	7.5	12	4.5	7.8	7.8	6.8
2	5.4	13	7.6	6.8	6.8	7.6
3	2.7	18	15.3	6.4	7.1	7.6
4	2.7	24	21.3	7.0	7.5	7.7

LMNG free micelles and FhuA complexes sediment separately (Fig 4) in exp 1, but are not resolved in exp 2-4. ^acalculated with the hypothesis of 1 g g^{-1} LMNG bound to FhuA, except for exp. 1, where free LMNG micelle concentration was



measured in AUC-SV. ^bfor FhuA complex in exp. 1, and unresolved LMNG free micelles and FhuA complex in exp. 2-4. ^cfrom AUC-SV experiments of pure LMNG (Fig 1) for the LMNG concentrations reported on the fourth column.

Fig 4. Characterisation and crystallization of LMNG solubilized FhuA

AUC-SV: **A:** Superposition of experimental and fitted sedimentation velocity profiles (top) and residuals (bottom), obtained at 130 000 g and 4 °C during \approx 4.5 hours using interference optics and 1.5 mm optical path centerpiece, for 7.5 mg mL⁻¹ FhuA solubilized with 12 mM LMNG in 20 mM Tris-HCl pH 8.0, 150 mM NaCl. Reference channel was filled with solvent without LMNG. **B:** Superposition of the normalized $c(s_{20w})$ distributions obtained at 280 nm (red line) and with interference optics (black line). SEC-LS: **C:** Rayleigh ratio measured along the elution profiles, for 35 μ L of FhuA at 1 mg mL⁻¹ and LMNG at 1.2 mM injected on a Superdex 200 10/300 GL (GE Healthcare) column equilibrated at 6 °C with 100 mM Tris-HCl pH 8.0, 150 mM NaCl, 1 mM EDTA, 0.5 mM LMNG, and molar masses (thick lines, right axis) for the whole complex (black), and LMNG (green) and FhuA (red) contributions, derived from combining light scattering, refractive index, and absorbance at 280 nm detections. **D.** Typical DDAO-solubilized FhuA crystals grown in 0.1 M MES pH 6.0, 10% PEG 6000, 2% DDAO. **E.** Typical LMNG-solubilized FhuA crystals grown in 0.1 M ADA pH 6.5, 12% PEG 6000, 1.2% LMNG.

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753

754

755 **FhuA crystallization**

756 480 conditions were screened for the crystallization of FhuA solubilized in 1.2% (12 mM)
 757 LMNG or, for comparison, 1.6% (79.4 mM) decyldimethylamide-N-oxide (DDAO). DDAO
 758 is the detergent in which the protein was historically crystallized [56], and in our hands,
 759 FhuA-DDAO crystals diffracting to 2.2 Å could be obtained (Arnaud and Breyton,
 760 unpublished data).

761

762 The FhuA-LMNG complex is clearly more susceptible to precipitating conditions: only
 763 53.7% of the drops remained clear (*vs.* 86% for FhuA-DDAO). In addition, 13.3% of the
 764 conditions with LMNG yielded unambiguous crystal formation *vs.* only 3% in DDAO, and 10
 765 *vs.* 8 conditions, respectively showed single crystals. Whereas conditions allowing single
 766 crystal growth in DDAO were closely related (high molar mass PEG, pH 6-7 and MgCl₂),
 767 those in LMNG displayed more differences: different precipitants, including wide ranges of
 768 PEG molar mass (400-8000) and pH (4-9) and a variety of co-crystallization salts. The shapes
 769 of the crystals were also very different, with feather-like shapes for FhuA-DDAO and more

770 rod for FhuA-LMNG crystals (Fig 4D). X-ray diffraction data were collected on FhuA-
 771 LMNG crystals (Table 4). Crystals grown in the screening assay, with little optimization,
 772 diffracted up to 4 Å, which allowed to get a model at low resolution and to determine the
 773 crystal packing. Interestingly, these crystals have a different space groups (C222₁) than
 774 crystals obtained with DDAO (P6₁).

775
 776 Thus, LMNG largely expanded the number of potential conditions in which FhuA crystals are
 777 grown, possibly by allowing different crystal packing. These data seemingly support the idea
 778 that LMNG enhances successful crystallization of membrane proteins [26].

779

Table 4. Characteristics of LMNG solubilized FhuA crystals

780

Crystal data collection statistics for FhuA – LMNG

781		
782	Crystal data collection statistics for FhuA – LMNG	
783	Space group	C 2 2 2 ₁
784	Unit cell dimension (Å)	149.53 210.96 188.7 90 90 90
785	Number of reflections	371461 (37282)
786	Number of unique reflections	25522 (2481)
787	Resolution limits (Å)	40.61 - 4.0
788	Higher resolution shell (Å)	4.143 - 4.0
789	Completeness	98.13 % (98.06 %)
790	Redundancy	14.6 (14.8)
791	Rmerged	0.1771 (7.384)
792	Rmeasured	0.1838 (7.648)
793	I/(I)	9.51 (0.50)

794

Molecular replacement with Phaser

795 LLG = 3698.6

796 TFZ = 16.4

797

Crystallographic refinement statistics

798		
799	Crystallographic refinement statistics	
800	Resolution range (Å)	40.61 - 4.0
801	Number of reflections	25098
802	R / FreeR	0.3141 / 0.3546
803	Number of atoms (total)	11024
804	Mean B value (Å ²)	209.52
805		
806	rms deviation from ideal values:	
807	bond length (Å)	0.007
808	bond angle (degree)	1.52

809

810

SpNOX-LMNG assemblies

811 We then investigated the prokaryotic enzyme SpNOX [62]. This protein is a recently
 812 identified analog to the eukaryotic NOX membrane proteins. They are the central catalytic
 813 component, involved in transmembrane electrons transfer, within NADPH oxidase
 814 complexes, which play essential roles in *e.g.* immunity, cardiovascular physiology, etc.
 815 Despite the progress done in the characterization of the soluble subunits of the NADPH
 816 oxidase complexes [83,84], there were very few structural information on the NOX α-helical
 817 membranous component up to the recent composite structure of a NOX5 isoform from a
 818 cyanobacteria [85]. Apart from its fundamental biological interest, this protein displays
 819 several features that make it a convenient model to test and develop new methods or
 820 processes in membrane protein biochemistry. Indeed, the red color brought by the presence of
 821 two hemes *b* within the transmembrane protein constitutes a good reporter in all steps of
 822

823 protein manipulation as well as a precious indicator of the correct folding of the protein.
 824 Moreover, SpNOX has an enzymatic activity that can be measured spectrophotometrically in
 825 a fast and convenient way.

826
 827 For SpNOX solubilization and purification, maltoside detergents were identified as essential
 828 in order to preserve protein activity [62]. However, several lines of improvement have been
 829 observed by using LMNG instead of DDM. First, from the elution profile from size exclusion
 830 chromatography of the second purification step (Fig 5A), the peak of SpNOX aggregates, at
 831 45 mL, decreases drastically from DDM to LMNG while, in addition to minor amounts of
 832 dimers detected in the two conditions, that of monomeric SpNOX, at 70 mL, increases.
 833 SpNOX is obtained with an improved homogeneity in LMNG compared to DDM. Moreover,
 834 SpNOX shows a significantly increased specific activity in LMNG compared to DDM (Table
 835 5). We evaluate the relative thermal stability of SpNOX in LMNG compared to DDM. T_m in
 836 LMNG is drastically increased (+13°C) (Table 2).

837
 838

839 **Table 5. Compared activities, in LMNG and DDM, of SpNOX and BmrA**

Protein	Environment	Activity
841 SpNOX	DDM 1.27 mM	5.9 +/- 0.5 mol Cyt c reduced s ⁻¹ mol ⁻¹ SpNOX
	LMNG 0.21 mM	9.3 +/- 0.2 mol Cyt c reduced s ⁻¹ mol ⁻¹ SpNOX
BmrA	DDM 2 mM	0.8 +/- 0.1 μmol ATP hydrolyzed min ⁻¹ mg ⁻¹ BmrA
BmrA	LMNG 0.1 mM	1.4 +/- 0.2 μmol ATP hydrolyzed min ⁻¹ mg ⁻¹ BmrA
BmrA	liposome	5.0 +/- 0.6 μmol ATP hydrolyzed min ⁻¹ mg ⁻¹ BmrA

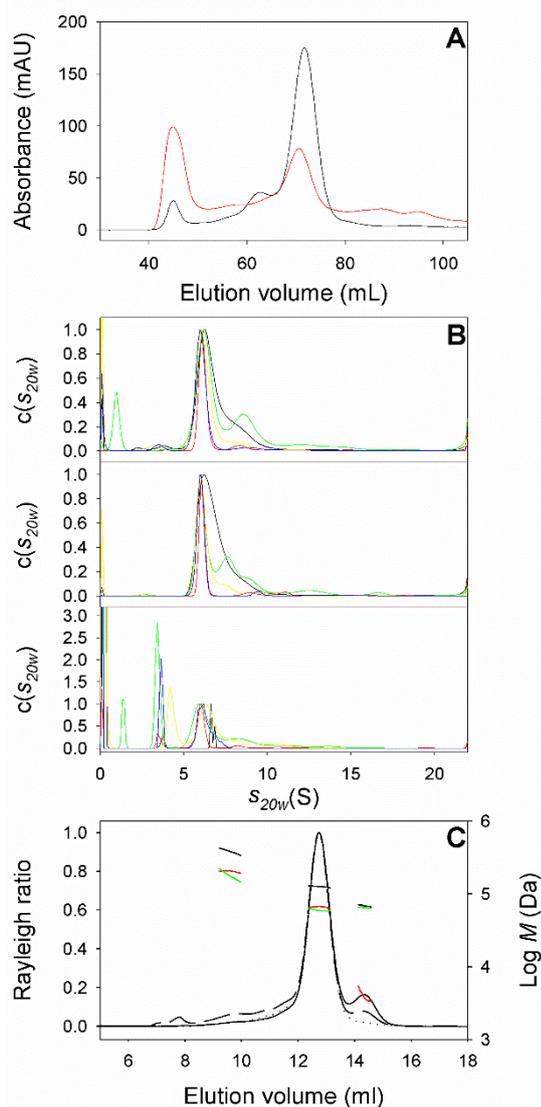


Fig 5. LMNG solubilized SpNOX: preparative SEC, AUC-SV and SEC-LS

A: Superdex 200 16/60 elution profiles, in the second step of SpNOX purification, performed in 50 mM Tris-HCl pH 7, 300 mM NaCl, with either 0.3 mM DDM (red line) or 0.03 mM LMNG (black line). Aggregates elute at 45 mL, while monomeric SpNOX elutes at 70 mL. **B:** Superposition of normalized $c(s_{20w})$ distributions obtained from AUC-SV at 280 nm (top), 412 nm (middle) and with interference optics (bottom), for SpNOX samples *c0.1*, *c3*, *c0.25d*, *c0.1d*, *c0.25dLMNG*, described in the text (black, red, green, yellow, and blue lines, respectively). The vertical dashed line at 6 S indicates the main SpNOX complex. **C:** SEC-LS normalized elution profiles obtained at 280 nm (dotted line), by refractive index (continuous line), and light scattering (dashed line), for 30 μL of SpNOX at 2.9 mg mL⁻¹ injected on a Superdex 200 10/300 GL (GE Healthcare) column equilibrated at 4 °C with 50 mM Tris-HCl pH 7.0, 300 mM NaCl, 0.05 mM LMNG (buffer A) and molar masses (thick lines, right axis) for the whole complex (black), and LMNG (green) and SpNOX (red) contributions, derived from combining static light scattering, refractive index, and absorbance

877 at 280 nm detections. A hydrodynamic radius of 4.0 nm was measured by DLS at the maximum of the main
878 peak.

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886 We then investigated, using SV-AUC, homogeneity and association state of SpNOX
887 solubilized in LMNG, at different concentrations of SpNOX and LMNG. After a final SEC
888 purification step with 0.03 mM LMNG in the elution buffer (buffer A, see legend to Fig 5),
889 SpNOX was obtained at 0.1 mg mL⁻¹ (sample *c0.1*). Then the protein was concentrated 30
890 times by ultrafiltration with a 30-kDa cut off membrane (sample *c3*). To further investigate
891 the effects of protein and LMNG concentrations on the size and composition of the
892 LMNG/SpNOX complexes, part of *c3* was rediluted 12 or 30 times in buffer A (containing
893 0.03 mM LMNG) and 12 times with the same buffer but containing LMNG at 1.5 mM
894 (samples *c0.25d*, *c0.1d*, *c0.25dLMNG*, respectively). AUC-SV of these samples was followed
895 at 412 nm, in addition to the usual 280 nm and interference detection, which allows following
896 the heme b cofactors. The normalized $c(s)$ curves (Fig 5B) at 280 and 412 nm superimpose
897 showing that the different SpNOX complexes in all samples have the same heme content. The
898 measured ratio $A_{412}/A_{280}=1.75 \pm 0.05$ is that expected for pure SpNOX [62]. The main
899 SpNOX species (typically 85%) sediments at $s_{20w} = 6.0 \pm 0.1$ S. A minor contribution
900 (typically 8%) is detected at ≈ 8.2 S, in addition to poorly-defined aggregates (typically 7 %).
901 *c0.25dLMNG* appears to be more homogeneous, and *c0.25d* more heterogeneous, but we
902 cannot ascertain the relevance of these subtle variations. We conclude that SpNOX main
903 assemblies have clearly the same composition in all samples, and increasing LMNG or
904 protein concentration does not lead to larger complexes. Interference optics allows the
905 additional detection of LMNG micelles below 4 S, in the expected s -range, since we
906 measured s at 2.5 and 5 S for pure LMNG at 0.05 and 1.5 mM in buffer A. From the
907 integration of the signal, we derived a concentration of 1.2 mM free micelles in *c3*, which
908 indicated an almost stoichiometrical co-concentration of LMNG and SpNOX. For the main
909 SpNOX complex, the ratio of the interference fringes and A_{280} signals is the same for *c0.1*, *c3*,
910 *c0.25d*, suggesting that no or few LMNG micelles co-sediment (the ratio is however larger for
911 *c0.25dLMNG* and *c0.1d*, but it may be related to the uncertainty of the analysis), and provides
912 a rough estimate of 1 ± 0.1 g g⁻¹ LMNG bound to SpNOX (this value is imprecise, as
913 discussed below).

914

915 Complementary SEC-LS-DLS in buffer A with 0.05 mM LMNG was performed on four
916 samples of SpNOX from a different purification. One at 0.16 mg mL⁻¹ is from SEC, the three
917 others are from subsequent concentration to 1.0, 2.9 and 8.7 mg mL⁻¹ (concentration by a
918 factor of 8, 18 and 54, respectively) by ultrafiltration. The elution profiles of the four SpNOX
919 samples are very similar, and one example is shown on Fig 5C. The main SpNOX complex
920 elutes at 12.8 mL with a $R_H = 4.0$ nm measured from DLS. The analysis provides molar
921 masses for the main complex of 68 ± 3 kDa and 59 ± 3 kDa for SpNOX and LMNG-
922 components, remarkably constant for the four samples. These values correspond to a
923 detergent/protein ratio similar to that estimated from AUC. The protein molar mass, however
924 is intermediate between the theoretical ones for a monomer (48 kDa) and a dimer. This
925 suggests a systematical error in the input value, used in AUC and SEC-LS, of the SpNOX
926 extinction coefficient. We thus rather consider the total molar mass of the complex of 126
927 kDa, derived from LS and RI detection to derive, in the hypothesis of a monomeric protein,

928 an amount of bound detergent of 1.6 g/g. Combining s from AUC and R_H from DLS, a
929 buoyant molar mass of 26.2 kDa is derived, which corresponds to a globular compact
930 monomer of SpNOX binding 1.6 g.g⁻¹ of LMNG, with a frictional ratio of 1.2, corresponding
931 to a globular compact shape.

932

933 *BmrA-LMNG assemblies*

934

935 BmrA (for ‘Bacillus multidrug resistance ATP’) is a member of the ABC (“ATP-Binding
936 Cassette”) transporters family. It is capable to export multiple drugs with no chemical
937 relationship [86]. In human, its closest homologue MDR1 confers resistance to
938 chemotherapeutic treatments [63]. Crystal structures of homologs stabilized in different states
939 [87,88] suggest that, to permit drug translocation, large conformational changes must occur in
940 an alternating access model. BmrA could be purified in a functional homodimeric form in
941 0.05% DDM [89,90], and, indeed it was shown to be flexible and adopt different
942 conformations [91–94].

943

944 Since then, the protein was purified in LMNG [95]. It was noticed that LMNG, in contrast to
945 DDM, significantly reduced BmrA aggregation with time. Moreover, the ATPase activity of
946 BmrA was significantly increased, by a factor of about 1.5 to 2, in LMNG compared to DDM
947 (Table 5). The activity in LMNG is thus getting closer to that of the protein reconstituted into
948 liposomes: 1.4 vs. 5, respectively, $\mu\text{mol ATP hydrolyzed /min/mg protein}$ (Table 5).

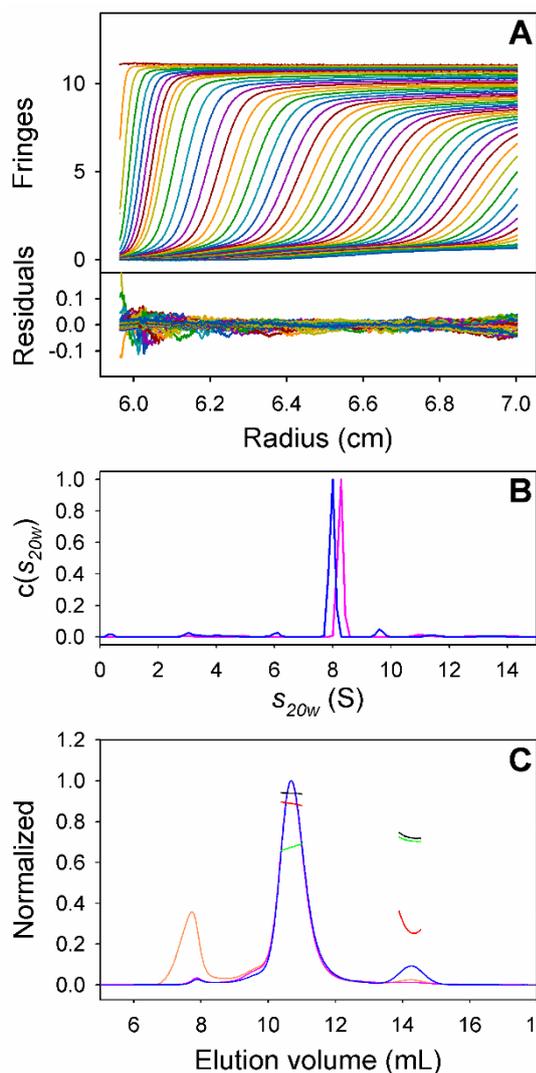
949

950 We evaluated by DSF the protein’s thermal stability for two freshly purified -in either DDM
951 or LMNG- batches of BmrA. Measurements were done on the Apo and the vanadate (Vi)
952 inhibited forms. The Apo-form is expected to be in an open conformation, the Vi-inhibited
953 form would exist in a closed conformation ([64,93] and unpublished results). Fig 3 shows the
954 measurements for BmrA Apo and Vi-inhibited forms, and Table 2 reports the melting
955 temperatures T_m . In DDM and in LMNG, the T_m increases steadily from the less stable Apo to
956 the more stable Vi-inhibited form. For the two forms, BmrA unfolds at higher temperature
957 when purified in LMNG compared to DDM: by 4.5 °C for BmrA Apo, and ≈ 13 °C for the
958 Vi-inhibited BmrA (Table 2).

959

960 Limited proteolysis experiments were performed on the Apo- and Vi-inhibited BmrA forms,
961 to evaluate differences in protein flexibility between the two forms, in DDM and in LMNG.
962 After incubation for different times with a protease, BmrA samples were run on SDS-PAGE.
963 Fig 6A-D show clearly, for BmrA purified in DDM and LMNG, respectively, and incubated
964 with trypsin, the increased robustness of Vi-inhibited BmrA compared to the Apo form. In
965 DDM, the 55 kDa band representing the intact protein essentially disappeared after 30 min of
966 incubation for the Apo form, and more than 300 min for the Vi-inhibited form, in agreement
967 with previously published results [93]. In LMNG, 60 min are required for an almost full
968 disappearance of the intact Apo BmrA, and the Vi-inhibited form is essentially preserved for
969 the entire time period tested. The reduced kinetics of proteolysis in LMNG compared to DDM
970 is also observed with protease V8 (Fig 6E-F). We checked that the detergent type had no
971 effect on protease activity by itself.

972



983 adding the proteases. The arrows indicate intact BmrA.

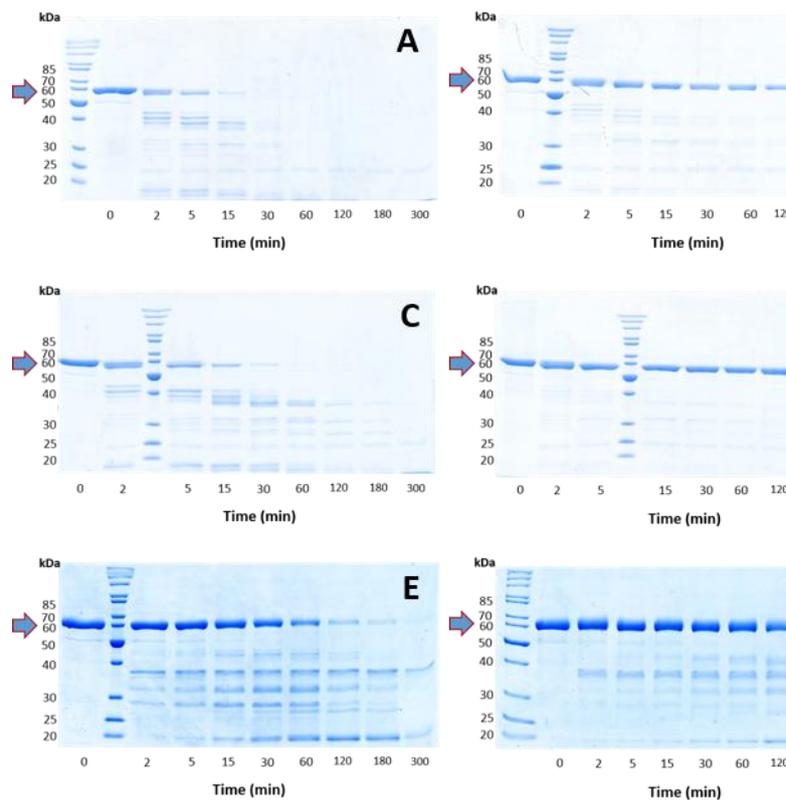


Fig 6. BmrA limited proteolysis experiments

A-D: incubation with trypsin at $1 \mu\text{g}/250 \mu\text{g}$ of protein, for A: Apo BmrA in DDM, B: Vi-inhibited BmrA in DDM, C: Apo BmrA in LMNG, D: Vi-inhibited BmrA in LMNG. E-F: incubation with protease V8 at $1 \mu\text{g}/20 \mu\text{g}$ of protein, for Apo BmrA, in E: DDM, and F: LMNG. DDM and LMNG concentrations are 2 and 0.1 mM, respectively. Time zero is taken after 15 min of pre-incubation before

983
 984
 985
 986 In order to evaluate protein homogeneity in LMNG, samples of BmrA were purified with a
 987 last step of SEC in a buffer containing LMNG at 0.1 mM, concentrated using a centrifugal
 988 filter device by a factor of 4.4 for a final BmrA concentration of 1.3 mg mL^{-1} . SV-AUC
 989 revealed a very homogeneous preparation, with one protein complex at $s_{20w} = 8.2 \text{ S}$
 990 (experimental value of 4.2S, at 10°C , in the buffer containing glycerol) (Fig 7A). SEC-LS
 991 experiments, done with a BmrA sample at 2.7 mg mL^{-1} , prepared in a similar way, stored
 992 frozen at -80°C and injected on a column equilibrated with buffer A, showed minor (total \approx
 993 5%) contributions of large and small aggregates eluting at 7.8 mL and as a shoulder at 9.7
 994 mL, in addition to the main complex, eluting at 10.7 mL, with a $R_H = 5.5 \text{ nm}$ measured from
 995 DLS, and a total molar mass of 240 kDa, corresponding to a dimer with 0.8 g/g bound
 996 detergent (Fig 7B). Combining R_H and the s -values, we derived a buoyant molar mass of 45.9
 997 kDa, corresponding to a dimer with 0.6 g/g bound LMNG, and a frictional ratio of 1.35.
 998 Excess LMNG micelles were undetected in the AUC experiments; in SEC-LS their amount
 999 corresponds to an injected LMNG concentration of 0.2 mM. In conclusion, very
 1000 homogeneous BmrA samples can be prepared in LMNG with protein concentration in the mg
 1001 mL^{-1} range, which remains essentially homogeneous after storage at -80°C .

Fig 7. LMNG solubilized BmrA in AUC-SV and SEC-MALS

1005 AUC-SV: **A:** Superposition of experimental and fitted sedimentation velocity profiles (top) and residuals
1006 (bottom), obtained at 130,000 g and 10 °C during \approx 8 hours using interference optics and 12 mm optical path
1007 centerpiece, for BmrA at 2.3 mg mL⁻¹ and free LMNG at 0.44 mM (putative concentration) in 25 mM Tris-HCl
1008 pH 8.0, 150 mM NaCl, 10% glycerol. Reference channel was filled with solvent without LMNG. **B:**
1009 Superposition of the normalized $c(s_{20w})$ distributions obtained at 280 nm (pink line) and with interference optics
1010 (blue line). SEC-LS: **C:** Rayleigh ratio, refractive index, and absorbance at 280 nm (orange, blue, and pink lines,
1011 respectively) measured along the elution profiles, for 50 μ L of BmrA at 2.7 mg mL⁻¹ and free LMNG at 0.06
1012 mM, this concentration being experimentally determined in SEC-LS, injected on a Superdex 200 10/300 GL (GE
1013 Healthcare) column equilibrated at 4 °C with 50 mM Tris-HCl pH 7.0, 300 mM NaCl, 0.05 mM LMNG (buffer
1014 A), and molar masses (thick lines, right axis) for the whole complex (black), and LMNG (green) and BmrA (red)
1015 contributions, derived from combining the three detections. The protein elutes as a main species at 10.7 mL,
1016 together with large and small aggregates, in minor amounts, in the void volume (7.8 mL) and as a shoulder at 9.7
1017 mL, respectively; free LMNG elutes at 14.2 mL.

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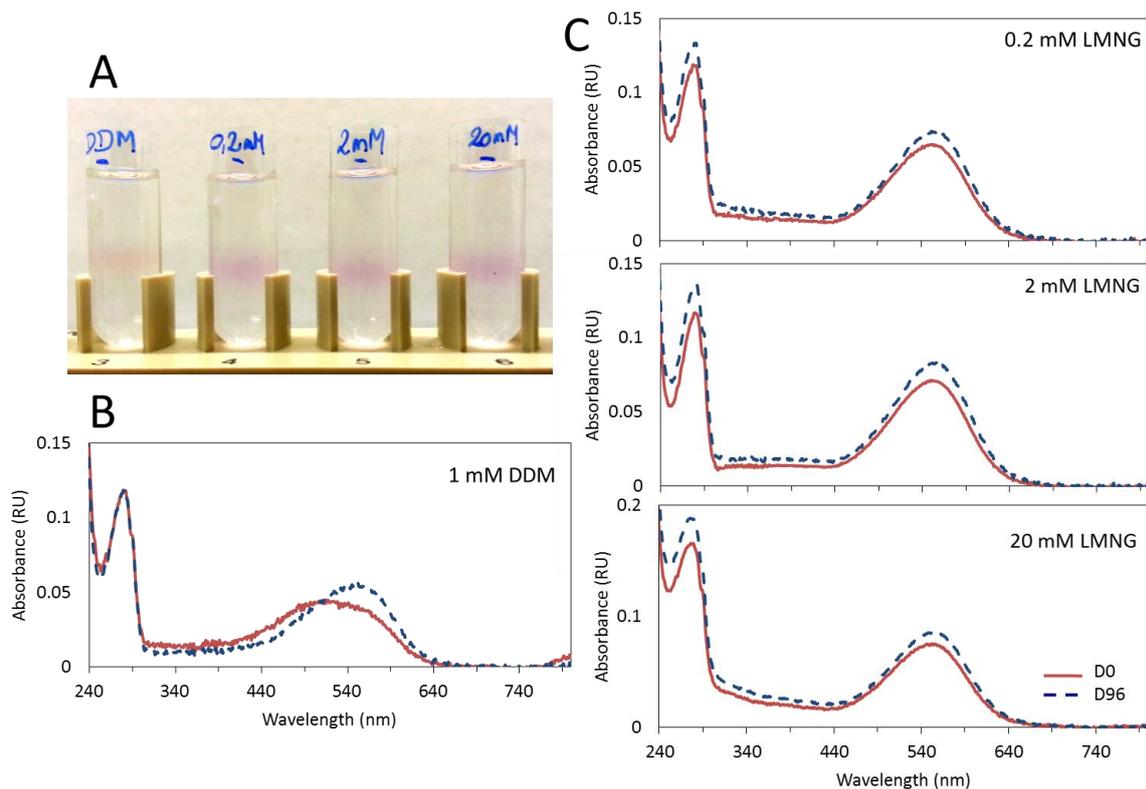
1024 ***bR homogeneity, and time and thermal stability***

1025 Lastly, we investigated the homogeneity and time stability of the bR as a model protein. bR is
1026 a light-driven proton pump purified from the archaea *Halobacterium*. It is composed of seven
1027 transmembrane α -helices and binds a covalent cofactor, a retinal molecule that confers purple
1028 color to the protein.

1029

1030 Sucrose gradients are a convenient means to perform both detergent exchange and evaluate
1031 the colloidal homogeneity of the protein-detergent complex. In the case of colored proteins,
1032 this latter information is directly visible after centrifugation, by the broadness of the protein-
1033 detergent band. We routinely use this method to evaluate the potentialities of fluorinated
1034 surfactants in the biochemistry of membrane proteins, using the bR. Fig 8A shows the results
1035 of sucrose density gradient experiments in the presence of 1 mM DDM, or 0.2, 2 and 20 mM
1036 LMNG. After 15 h centrifugation, the protein migrated past the middle of the gradient,
1037 insuring total detergent exchange, as checked by TLC. In all LMNG conditions, the protein
1038 band migrates at the same depth and appears to have the same width/broadness as that in
1039 DDM, suggesting that the bR-LMNG complexes are homogeneous monomers as the bR-
1040 DDM complexes. The retinal molecule, whose visible absorption spectrum is very sensitive to
1041 its local environment, is a convenient reporter of the state of the protein: the trimeric protein
1042 in its native membrane reveals a visible absorption spectrum with a maximum at $\lambda_{\max} =$
1043 570 nm; when solubilized in detergent, the protein monomerises and displays $\lambda_{\max} = 554$ nm;
1044 in both cases, the protein appears purple. When the protein denatures, the retinal is released,
1045 and λ_{\max} shifts to 400–380 nm: the protein solution turns yellow. Fig 8B and 8C report the
1046 absorption spectra of the protein recovered just after centrifugation and three months after,
1047 stored on ice in the dark. The DDM sample, which appears pinkish in the gradient, displays a
1048 broad absorption peak, with a $\lambda_{\max} \sim 530$ nm (Fig 8B), suggesting a poorly folded protein.
1049 After three months however, the spectrum shows a usual peak, with $\lambda_{\max} = 555$ nm. The
1050 protein probably started to denature in OTG, the detergent used for its solubilization, but
1051 returned to its native fold when transferred to DDM. In all LMNG samples, the absorption
1052 peak displays a $\lambda_{\max} = 554$ nm (Fig 8C), suggesting that the dynamics of the bR monomer in
1053 LMNG is different than in classical detergents. After three months incubation, the absorption
1054 spectra are unchanged, indicating that the protein is very stable.

1055



1056
1057 **Fig 8. bR sucrose gradient**

1058 **A:** Migration of bR in 10-30% sucrose gradients in the presence of either 1 mM DDM, 0.2, 2 or 20 mM LMNG.
1059 Gradients were centrifuged 15 h at 200,000 x g. bR was recovered from the gradients and its UV-visible
1060 spectrum measured immediately (D0, solid line) or three months after incubation at 4°C in the dark (D96, dashed
1061 line). **B:** bR in 1 mM DDM, **C:** bR in 0.2, 2 and 20 mM LMNG.

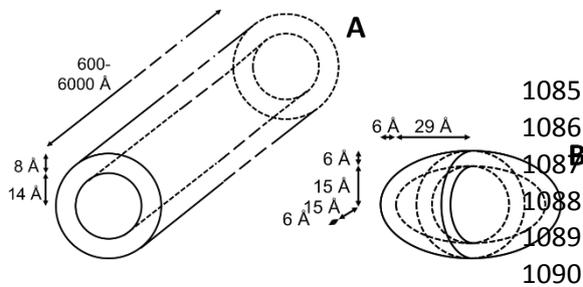
1062
1063 At last, bR thermal denaturation assays were performed. Solubilized bR samples were
1064 incubated with biobeads to decrease the OTG concentration, and diluted 18 times (leading to
1065 a residual OTG concentration of 0.4 times the CMC) in OTG, DDM, or LMNG, at a
1066 concentration of 2 mM above the CMC of each detergent, for thermal denaturation assays.
1067 Result are presented on Fig 3 and Table 2. bR has an increased thermostability in DDM
1068 compared to OTG, and LMNG is even more stabilizing, with T_m increased by 6 and 12 °C,
1069 respectively.

1070
1071
1072 **Discussion and conclusion**

1073
1074 **On the self-assembly of LMNG**

1075 From AUC experiments, we determined the CMC of LMNG to be 40 μM. This is coherent
1076 with previously published value from hydrophobic dye solubilization measurements (10
1077 μM) [26], while isothermal titration calorimetry measurements yielded a significantly lower
1078 concentration (11.3 nM) [44]. The CMC is not an absolute value, rather a range of
1079 concentrations, and different techniques used to determine the CMC probe different properties
1080 of micelle formation. Thus, it is not unusual to find different values depending on the
1081 technique used, and discrepancies become exacerbated when the CMC is very low.

1082
1083 Regarding the size of the micelles, the s_{20w} -value of LMNG at 0.25 mM (0.025 %) is 3.3 S.
1084 This value is in the same range than that of DDM (3.12 S) that forms globular compact



1085 micelles of about 60 kDa in a large range
 1086 of detergent concentration [65]. If LMNG
 1087 forms quite small micelles at 0.25 mM,
 1088 AUC, DLS, SAXS, and SANS, clearly
 1089 indicate it forms larger micelles, rods of
 1090 increasing length, when increasing the

1091 concentration above 0.25 mM. The lateral dimension of the hydrophobic core radius is 14 Å,
 1092 and that of the shell thickness is 7 Å in H₂O (9 Å in D₂O). These values correspond to that of
 1093 DDM micelles, described from SAXS to form oblate ellipsoids with, in the minor axis, a
 1094 hydrophobic core of ≈ 15 Å surrounded by a hydrophilic layer of ≈ 6 Å thickness [96,97]
 1095 (Scheme 2). The differences of 1 Å between our LMNG and the published DDM dimensions
 1096 are not significant since different techniques and buffers, thus contrasts (in terms of electron
 1097 density or neutron scattering length densities), were used. To this date, this is only one
 1098 membrane protein pdb file (4b4a) containing LMNG as a ligand (pdf identifier “LMN”). The
 1099 maximum C-O, C-C and O-O distances in the whole molecule, in the two tails, and in the two
 1100 heads, respectively, are 21.9, 13 and 11, 9.8 and 9.7 Å, respectively (we note that in this
 1101 unique structure, the hydrophilic head is quite open: we measure a maximal distance between
 1102 the O of the two sugars of LMNG of 19 Å). For DDM as a ligand (pdf identifier “LMT”), in
 1103 an arbitrary selection of membrane protein pdb files (6haw, 6hqb, 4kfm, 6cnn), we measure
 1104 maximal distances for the whole molecule, the tail, and the head, of 20.5 +/- 1, 12 +/- 1, and
 1105 9.2 +/- 0.8 Å, respectively. These values are logically similar to that of LMNG. The
 1106 maximum length of the tail is logically lower than 16.7 Å corresponding to the dimension of
 1107 an extended alkyl chain with 12 carbons [97].
 1108

1109 If DDM is described to form slightly elongated micelles, for LMNG at 20 mM however, we
 1110 measured lengths of 600 Å in H₂O and more than 5000 Å in D₂O. It corresponds to LMNG
 1111 rods with ratios of the total length over the total diameter, of 14 and 120. For LMNG in H₂O,
 1112 the rod length value coincides with the maximum distances estimated between 10 and 20 mM
 1113 LMNG, from the pair distribution analysis of the – measured independently – SAXS data of
 1114 Fig 2A. In a preliminary independent SANS experiment, however, rods of only 1200 Å long
 1115 were modelled for LMNG micelles at 20 mM in D₂O. It corresponds to a length over diameter
 1116 ratio of 28, 4-5 times smaller than that presented above. It is likely that the precise length of
 1117 LMNG rods depends on uncontrolled parameters during sample preparation. It is possible that
 1118 LMNG forms longer rods in D₂O, compared to H₂O, but we cannot ascertain it. For both
 1119 SAXS and SANS, the shape and size of the micelles does not change between 10 and 20 mM.
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1127 **Scheme 2. Schematic representation of LMNG and DDM micelles**

1128 **A:** LMNG micelles at 20 mM in H₂O or D₂O, from modelling of combined SANS and SAXS data (this work) **B:**
 1129 DDM micelles from modelling of SAXS data [96,97].
 1130
 1131

1132 Israelachvili et al. [98,99] have provided a rationale to explain how the molecular geometry of
 1133 individual surfactant molecules affects and controls the shape and size of the assemblies they
 1134 form. The geometry of the molecule can be described by three parameters: the area of the
 1135 hydrated polar head, a_0 , the volume, v , and the extension, l_c , of the hydrophobic tail. The

1136 value of the related critical packing parameter, $v/(a_0 l_c)$, representing the ratio of the mean area
1137 of the hydrophobic tail on that of the hydrophilic head, determines the geometry of the
1138 assemblies: for $v/(a_0 l_c) < 1/3$ (large hydrophilic surface), spherical micelles are formed; for
1139 $1/3 < v/(a_0 l_c) < 1/2$, rods are favored; then, for $v/(a_0 l_c) < 1$, ~ 1 , and > 1 (with increasingly
1140 large hydrophobic lateral extensions), flexible or planar bilayers, and inverted micelles,
1141 respectively, are stabilized. The LMNG molecule corresponds to two linked DDM molecules.
1142 The linkage is done through a quaternary carbon, localized on the lauryl chain, at the second
1143 carbon following the ether group belonging to the maltose moiety. It is thus at the junction
1144 between the hydrophilic head and the hydrophobic tail of the detergent. The constraints of this
1145 link on each moiety dimensions are hardly intuitively predictable. The fact that DDM forms
1146 globular micelle while LMNG forms rod means that the packing parameter $v/(a_0 l_c)$ increases
1147 from DDM to LMNG, which can be related to an increase, in LMNG compared to DDM of
1148 the hydrophilic surface per maltose group, and/or of the mean length of the hydrophobic
1149 chain, corresponding to changes in their landscape of the molecular conformational stability
1150 and/or dynamics.

1151
1152 Rod formation has noticeable consequences for biochemists. When using ultrafiltration
1153 centrifugal devices to concentrate proteins, LMNG co-concentrates with the membrane
1154 protein by the same factor. Depending on the initial detergent concentration, typically from
1155 0.02 to 0.1 mM, LMNG in the concentrated sample will form small to very large assemblies.
1156 Concentrated LMNG dissociates upon SEC, but elutes as a very broad peak, preventing
1157 membrane protein in concentrated LMNG to be recovered with a well-defined LMNG
1158 concentration.

1160 **On the relative stabilization of proteins in LMNG compared to DDM**

1161 Our work corroborates the stabilizing propensity of LMNG towards membrane proteins,
1162 already described in the literature (see the introduction). All four proteins investigated here
1163 are thermally stabilized in LMNG compared to DDM. The temperature shift is very large,
1164 both for FhuA (+11 °C), a very stable β -barrel protein, and for SpNOX (+13 °C), a fragile α -
1165 helical protein. For the flexible BmrA, temperature shifts between 4 and 13 °C are observed
1166 depending on the presence or absence of ligands. For bR, the gain is 6 °C. SpNOX solubilized
1167 and partially purified in LMNG shows a significantly decreased content of aggregates,
1168 corresponding to an increased stability. The reduced kinetics of limited proteolysis in LMNG
1169 compared to DDM for the Apo and the Vi-inhibited forms of BmrA, expected to represent the
1170 open and closed conformations, could have suggested some restricted flexibility of the
1171 protein. However, the protein is more active in LMNG as compared to DDM thereby
1172 indicating that the flexibility required for ATPase activity is not impaired in LMNG. Indeed,
1173 in LMNG, BmrA seems to be more prone to reach or stabilize the closed conformation which
1174 is required to get a high ATPase activity. Hence, BmrA is in a more stable and perhaps, more
1175 physiologically relevant conformations in LMNG as compared to DDM. Restricted protein
1176 flexibility is also suggested for bR, which UV-visible spectrum displays a maximum
1177 absorption peak close to that of the constrained trimer in the purple membrane, far from that
1178 of the more flexible monomer in DDM. These results are in the line with the restraint of
1179 protein inter-conversion dynamics measured for GPCRs. The reduced dynamics of proteins
1180 may be related to the strong binding of the LMNG correlated to its low CMC, deriving from
1181 the connection of the two hydrophobic tails in its structure. As for BmrA, SpNOX displays an
1182 increased specific activity in LMNG compared to DDM, which may reflect a significantly
1183 higher proportion of inactive protein in DDM compared to LMNG, or be related to a
1184 difference in SpNOX flexibility, or to the stabilization of some conformations.

1185

1186 **On the assembly of LMNG-membrane proteins**

1187 The solubilized membrane proteins investigated in this work have the same quaternary
1188 structure in LMNG as in their usual detergent: FhuA, SpNOX, and bR, are monomeric and
1189 BmrA is dimeric. In the case of SpNOX, dimers of the main species –probably inactivated
1190 forms– are detected in LMNG, for typically 10%, the proportion varying slightly depending
1191 on the purification. These species are also detected in DDM. In the AUC experiments where
1192 LMNG concentration was voluntarily raised to 24 mM, FhuA and SpNOX association states
1193 were not affected. The amount of bound detergent did not vary too, despite rod micelles of
1194 LMNG are expected at these large concentrations. This behavior was not anticipated. We
1195 previously investigated fluorinated surfactants (for reviews, see [8,16,17]), and, in a series of
1196 surfactants whose polar head size was modulated by the presence of one, two, or three
1197 glucose moieties, homogeneous complexes with bR and cytochrome *b₆f* were obtained only
1198 with the compounds that form homogeneous micelles, *i.e.* the surfactants with the more
1199 voluminous head. In the surfactant forming long rods, with only one glucose, the membrane
1200 proteins were soluble and active, but formed heterogeneous detergent protein complexes. We
1201 anticipated it could be related to the heterogeneity in the amount of bound detergent [100].
1202 Other fluorinated surfactants with relatively small head groups (lactoside, maltoside,
1203 octaethylene glycol ether) formed large and polydisperse, probably rod-like assemblies, and
1204 indeed formed also large and heterogeneous complexes with membrane proteins (refs in
1205 [100]). By contrast, the fluorinated surfactant F-TAC (now commercialized by Calixar),
1206 which has a voluminous polymeric hydrophilic head, was shown to assemble into small
1207 micelles and form well defined complexes with membrane proteins. The present work on
1208 LMNG shows that large size of surfactant micelle does not necessarily translate into protein
1209 detergent complex heterogeneity.

1210

1211 **On the assembly of LMNG-membrane proteins: bound detergent**

1212 In the case of FhuA, previous measurements determined 1.2 g of bound DDM per g of
1213 protein, using SEC in the presence of radiolabeled DDM [81]. This value is close to 1 and 1.2
1214 g g⁻¹ determined in the present work for the protein solubilized in LMNG by AUC and SEC-
1215 LS. For BmrA, the amount of bound LMNG (0.6 g g⁻¹) we determined by combining *s*- and
1216 *R_H*-values, is lower but in the same order of magnitude than that determined for bound DDM
1217 by the same technique (0.9 g g⁻¹)[101]. A similar trend, in the comparison of DDM and
1218 LMNG bound to BmrA, was reported using MALDI/TOF for the evaluation of bound
1219 detergent [7]. These estimates of bound DDM to BmrA are much lower than that determined
1220 by SEC in the presence of radiolabeled DDM (1.5 g g⁻¹)[89], which emphasizes the difficulty
1221 to determine precisely bound detergent. Nevertheless, these comparisons presented in Table 6,
1222 show that the amounts of bound detergent are similar whether the protein is solubilized in
1223 DDM or LMNG. It means that the amount in mol/mol is nearly twice for DDM than for
1224 LMNG, and it is possible that the amount (in g g⁻¹) of bound LMNG is slightly less compared
1225 to that of DDM.

1226

1227

1228 **Table 6. Bound LMNG and DDM to FhuA, BmrA, and SpNOX**

1229

1230 Protein	detergent	Bound detergent	technique	reference
1231		g/g	mol/mol*	
1232 FhuA	DDM	1.2	188	SEC/[¹⁴ C]DDM [81]
1233 FhuA	LMNG	1.0	80	AUC present work
1234 FhuA	LMNG	1.2	95	SEC-LS [81]

1237

1238	BmrA	DDM	0.9	234	AUC+ SEC-LS	[101]
1239	BmrA	DDM	1.3-1.6	388-416	MALDI/TOF	[7]
1240	BmrA	DDM	1.5	390	SEC/[¹⁴ C]DDM	[89]
1241						
1242	BmrA	LMNG	0.6	79	AUC+ SEC-LS	present work
1243	BmrA	LMNG	1.1-1.3	145-171	MALDI/TOF	[7]
1244						
1245	SpNOX	LMNG	1.6	77	AUC+ SEC-LS	present work
1246	* considering FhuA and SpNOX as monomers, and BmrA as a dimer.					

1247
1248

On the crystallization of LMNG-membrane protein

1250 The high resolution structures of several GPCRs were obtained thanks to purification in
1251 LMNG (cf Introduction) followed by crystallization in mesophase, a process in which the
1252 membrane protein in the crystal is stripped off from its bound detergent [102]. Crystallization
1253 in LMNG using vapor diffusion was described for *e.g.* the cytochrome *b₆f* complex [26], the
1254 TatC core of the twin-arginine protein transporter [50], an ABC transporter lipid-linked
1255 oligosaccharide flippase [41], a borate efflux transporter [51], and a plant voltage-gated two-
1256 pore channel [52]. FhuA is yet another one. One of our motivations, in crystallizing this well
1257 described protein, was to check whether the ability to obtain crystals was prevented by the
1258 presence of large LMNG assemblies in the membrane protein sample. It is clearly not the
1259 case, and on the contrary, comparing the behavior of the protein in crystallization screens, we
1260 observed that the number of initial hits was larger when the protein was solubilized in LMNG
1261 when compared to the classically used DDAO. The space group of crystals of FhuA grown in
1262 LMNG is not the same as that of the protein grown in DDAO. However, this is not
1263 remarkable, as the same space group is found when the protein is crystallized in LDAO.

1264

1265 To summarize, the interest of structural biologists for LMNG is very strong, since it stabilizes
1266 noticeably most membrane proteins compared to other detergents. Due to its low desorption
1267 kinetics, it would stabilize membrane proteins even in sub-CMC concentrations. It forms
1268 large rod assemblies at concentration above the sub mM range, but, for the examples we
1269 investigated, the solubilized proteins remain as homogeneous as in usual detergents, with
1270 similar amounts of bound detergent, and the presence of the large LMNG micelles does not
1271 prohibit membrane protein crystallization using vapor diffusion or mesophase.

1272

Acknowledgment

1274 This work used the platforms of the Grenoble Instruct centre (ISBG; UMS 3518 CNRS-CEA-
1275 UJF-EMBL) with support from FRISBI (ANR-10-INSB-05-02) and GRAL (ANR-10-LABX-
1276 49-01) within the Grenoble Partnership for Structural Biology (PSB). Gina Reyes-Mejia,
1277 Marie-Ange Marrel, Séraphine Crassac, and Emma Lundell (UGA) performed BmrA and
1278 FhuA preparations, FhuA and bR thermal denaturation assays, respectively, during their
1279 master internships at IBS; Marie-Pierre Candusso for advices with BmrA limited proteolysis;
1280 Adam Round and Martha Brennich (ESRF), and Frank Gabel (IBS), for help in SAXS data
1281 acquisition. This work benefited from the use of the SasView application, originally
1282 developed under NSF Award DMR-0520547. SasView also contains code developed with
1283 funding from the EU Horizon 2020 programme under the SINE2020 project Grant No
1284 654000. This work was supported by the French Agence Nationale de la Recherche (ANR-16-
1285 CE92-0001) to CE and CB, (ANR-14-CE09-0024B) to JMJ and (ANR-17-CE11-0013) to JD,
1286 CH, FF, AV, IPH, MT. A. Vermot thanks for her support through the Emergence program
1287 from the Univ. Grenoble Alpes.

1288

1289 **References :**

1290

1291 [1] M. le Maire, P. Champeil, J.V. Moller, Interaction of membrane proteins and lipids with
1292 solubilizing detergents, *Biochim Biophys Acta.* 1508 (2000) 86–111.

1293 [2] J.-L. Popot, Extracting Membrane Proteins from Their Native Environment, in: *Membr.*
1294 *Proteins Aqueous Solut. Deterg. Amphipols*, Springer International Publishing, 2018: pp. 59–95.

1295 [3] C. Breyton, C. Tribet, J. Olive, J.-P. Dubacq, J.-L. Popot, Dimer to monomer conversion of
1296 the cytochrome b6f complex. Causes and consequences, *J Biol Chem.* 272 (1997) 21892–900.

1297 [4] S. Lee, A. Mao, S. Bhattacharya, N. Robertson, R. Grisshammer, C.G. Tate, N. Vaidehi, How
1298 Do Short Chain Nonionic Detergents Destabilize G-Protein-Coupled Receptors?, *J. Am. Chem. Soc.*
1299 138 (2016) 15425–15433. doi:10.1021/jacs.6b08742.

1300 [5] C. Chipot, F. Dehez, J.R. Schnell, N. Zitzmann, E. Pebay-Peyroula, L.J. Catoire, B. Miroux,
1301 E.R.S. Kunji, G. Veglia, T.A. Cross, P. Schanda, Perturbations of Native Membrane Protein Structure
1302 in Alkyl Phosphocholine Detergents: A Critical Assessment of NMR and Biophysical Studies, *Chem.*
1303 *Rev.* 118 (2018) 3559–3607. doi:10.1021/acs.chemrev.7b00570.

1304 [6] Q. Zhang, H. Tao, W.-X. Hong, New amphiphiles for membrane protein structural biology,
1305 *Methods San Diego Calif.* 55 (2011) 318–323. doi:10.1016/j.ymeth.2011.09.015.

1306 [7] V. Chaptal, F. Delolme, A. Kilburg, S. Magnard, C. Montigny, M. Picard, C. Prier, L.
1307 Monticelli, O. Bornert, M. Agez, S. Ravaud, C. Orelle, R. Wagner, A. Jawhari, I. Broutin, E. Pebay-
1308 Peyroula, J.-M. Jault, H.R. Kaback, M. le Maire, P. Falson, Quantification of Detergents Complexed
1309 with Membrane Proteins, *Sci. Rep.* 7 (2017) 41751. doi:10.1038/srep41751.

1310 [8] G. Durand, M. Abla, C. Ebel, C. Breyton, New amphiphiles to handle membrane proteins:
1311 “ménage à trois” between chemistry, physical-chemistry and biochemistry, in: *Membr. Proteins Prod.*
1312 *Struct. Anal.*, Springer, I. Muss-Veteau, 2014.

1313 [9] J.-L. Popot, Alternatives to Detergents for Handling Membrane Proteins in Aqueous
1314 Solutions, in: *Membr. Proteins Aqueous Solut. Deterg. Amphipols*, Springer International Publishing,
1315 2018: pp. 97–149.

1316 [10] C.E. Schafmeister, L.J.W. Miercke, R.A. Stroud, Structure at 2.5 Å of a designed peptide that
1317 maintains solubility of membrane proteins, *Science.* 262 (1993) 734–738.

1318 [11] X. Zhao, Y. Nagai, P.J. Reeves, P. Kiley, H.G. Khorana, S. Zhang, Designer short peptide
1319 surfactants stabilize G protein-coupled receptor bovine rhodopsin, *Proc. Natl. Acad. Sci. U. S. A.* 103
1320 (2006) 17707–17712. doi:10.1073/pnas.0607167103.

1321 [12] K. Corin, P. Baaske, D.B. Ravel, J. Song, E. Brown, X. Wang, C.J. Wienken, M. Jerabek-
1322 Willemsen, S. Duhr, Y. Luo, D. Braun, S. Zhang, Designer lipid-like peptides: a class of detergents for
1323 studying functional olfactory receptors using commercial cell-free systems, *PloS One.* 6 (2011)
1324 e25067. doi:10.1371/journal.pone.0025067.

1325 [13] H. Tao, S.C. Lee, A. Moeller, R.S. Roy, F.Y. Siu, J. Zimmermann, R.C. Stevens, C.S. Potter,
1326 B. Carragher, Q. Zhang, Engineered nanostructured β -sheet peptides protect membrane proteins, *Nat.*
1327 *Methods.* 10 (2013) 759–761. doi:10.1038/nmeth.2533.

1328 [14] C. Tribet, R. Audebert, J.-L. Popot, Amphipols : polymers that keep membrane proteins
1329 soluble in detergent-free aqueous solutions, *Proc Nat Acad Sci USA.* 93 (1996) 15047–15050.

1330 [15] J.-L. Popot, *Membrane Proteins in Aqueous Solutions: From Detergents to Amphipols*,
1331 Springer International Publishing, 2018. //www.springer.com/de/book/9783319731469 (accessed June

- 1332 13, 2018).
- 1333 [16] C. Breyton, B. Pucci, J.-L. Popot, Amphipols and fluorinated surfactants: Two alternatives to
 1334 detergents for studying membrane proteins in vitro, *Methods Mol. Biol.* 601 (2010) 219–245.
 1335 doi:10.1007/978-1-60761-344-2_14.
- 1336 [17] J.-L. Popot, Amphipols, nanodiscs, and fluorinated surfactants: three nonconventional
 1337 approaches to studying membrane proteins in aqueous solutions, *Annu. Rev. Biochem.* 79 (2010) 737–
 1338 775. doi:10.1146/annurev.biochem.052208.114057.
- 1339 [18] R. Matar-Merheb, M. Rhimi, A. Leydier, F. Huché, C. Galián, E. Desuzinges-Mandon, D.
 1340 Ficheux, D. Flot, N. Aghajari, R. Kahn, A. Di Pietro, J.-M. Jault, A.W. Coleman, P. Falson,
 1341 Structuring detergents for extracting and stabilizing functional membrane proteins, *PloS One.* 6 (2011)
 1342 e18036. doi:10.1371/journal.pone.0018036.
- 1343 [19] P.S. Chae, P.D. Laible, S.H. Gellman, Tripod Amphiphiles for Membrane Protein
 1344 Manipulation, *Mol. Biosyst.* 6 (2010) 89–94.
- 1345 [20] P.S. Chae, K.H. Cho, M.J. Wander, H.E. Bae, S.H. Gellman, P.D. Laible, Hydrophobic
 1346 variants of ganglio-tripod amphiphiles for membrane protein manipulation, *Biochim. Biophys. Acta.*
 1347 1838 (2014) 278–286. doi:10.1016/j.bbamem.2013.09.011.
- 1348 [21] Q. Zhang, X. Ma, A. Ward, W.-X. Hong, V.-P. Jaakola, R.C. Stevens, M.G. Finn, G. Chang,
 1349 Designing facial amphiphiles for the stabilization of integral membrane proteins, *Angew. Chem. Int.*
 1350 *Ed Engl.* 46 (2007) 7023–7025. doi:10.1002/anie.200701556.
- 1351 [22] P.S. Chae, K. Gotfryd, J. Pacyna, L.J.W. Miercke, S.G.F. Rasmussen, R.A. Robbins, R.R.
 1352 Rana, C.J. Loland, B. Kobilka, R. Stroud, B. Byrne, U. Gether, S.H. Gellman, Tandem facial
 1353 amphiphiles for membrane protein stabilization, *J. Am. Chem. Soc.* 132 (2010) 16750–16752.
 1354 doi:10.1021/ja1072959.
- 1355 [23] S.C. Lee, B.C. Bennett, W.-X. Hong, Y. Fu, K.A. Baker, J. Marcoux, C.V. Robinson, A.B.
 1356 Ward, J.R. Halpert, R.C. Stevens, C.D. Stout, M.J. Yeager, Q. Zhang, Steroid-based facial
 1357 amphiphiles for stabilization and crystallization of membrane proteins, *Proc. Natl. Acad. Sci. U. S. A.*
 1358 110 (2013) E1203-1211. doi:10.1073/pnas.1221442110.
- 1359 [24] P.S. Chae, S.G.F. Rasmussen, R.R. Rana, K. Gotfryd, A.C. Kruse, A. Manglik, K.H. Cho, S.
 1360 Nurva, U. Gether, L. Guan, C.J. Loland, B. Byrne, B.K. Kobilka, S.H. Gellman, A new class of
 1361 amphiphiles bearing rigid hydrophobic groups for solubilization and stabilization of membrane
 1362 proteins, *Chem. Weinh. Bergstr. Ger.* 18 (2012) 9485–9490. doi:10.1002/chem.201200069.
- 1363 [25] P.S. Chae, R.R. Rana, K. Gotfryd, S.G.F. Rasmussen, A.C. Kruse, K.H. Cho, S. Capaldi, E.
 1364 Carlsson, B. Kobilka, C.J. Loland, U. Gether, S. Banerjee, B. Byrne, J.K. Lee, S.H. Gellman, Glucose-
 1365 neopentyl glycol (GNG) amphiphiles for membrane protein study, *Chem. Commun. Camb. Engl.* 49
 1366 (2013) 2287–2289. doi:10.1039/c2cc36844g.
- 1367 [26] P.S. Chae, S.G.F. Rasmussen, R.R. Rana, K. Gotfryd, R. Chandra, M.A. Goren, A.C. Kruse,
 1368 S. Nurva, C.J. Loland, Y. Pierre, D. Drew, J.-L. Popot, D. Picot, B.G. Fox, L. Guan, U. Gether, B.
 1369 Byrne, B. Kobilka, S.H. Gellman, Maltose-neopentyl glycol (MNG) amphiphiles for solubilization,
 1370 stabilization and crystallization of membrane proteins, *Nat. Methods.* 7 (2010) 1003–1008.
 1371 doi:10.1038/nmeth.1526.
- 1372 [27] A. Sadaf, J.S. Mortensen, S. Capaldi, E. Tikhonova, P. Hariharan, O. de Castro Ribeiro, C.J.
 1373 Loland, L. Guan, B. Byrne, P.S. Chae, A Class of Rigid Linker-bearing Glucosides for Membrane
 1374 Protein Structural Study, *Chem. Sci.* 7 (2016) 1933–1939. doi:10.1039/C5SC02900G.
- 1375 [28] M. Ehsan, Y. Du, N.J. Scull, E. Tikhonova, J. Tarrasch, J.S. Mortensen, C.J. Loland, G.

- 1376 Skiniotis, L. Guan, B. Byrne, B.K. Kobilka, P.S. Chae, Highly Branched Pentasaccharide-Bearing
1377 Amphiphiles for Membrane Protein Studies, *J. Am. Chem. Soc.* 138 (2016) 3789–3796.
1378 doi:10.1021/jacs.5b13233.
- 1379 [29] H. Hussain, J.S. Mortensen, Y. Du, C. Santillan, O. Ribeiro, J. Go, P. Hariharan, C.J. Loland,
1380 L. Guan, B.K. Kobilka, B. Byrne, P.S. Chae, Tandem malonate-based glucosides (TMGs) for
1381 membrane protein structural studies, *Sci. Rep.* 7 (2017) 3963. doi:10.1038/s41598-017-03809-3.
- 1382 [30] K.H. Cho, O. Ribeiro, Y. Du, E. Tikhonova, J.S. Mortensen, K. Markham, P. Hariharan, C.J.
1383 Loland, L. Guan, B.K. Kobilka, B. Byrne, P.S. Chae, Mesitylene-Cored Glucoside Amphiphiles
1384 (MGAs) for Membrane Protein Studies: Importance of Alkyl Chain Density in Detergent Efficacy,
1385 *Chem. Weinh. Bergstr. Ger.* 22 (2016) 18833–18839. doi:10.1002/chem.201603338.
- 1386 [31] M. Das, Y. Du, J.S. Mortensen, O. Ribeiro, P. Hariharan, L. Guan, C.J. Loland, B.K. Kobilka,
1387 B. Byrne, P.S. Chae, Butane-1,2,3,4-tetraol-based amphiphilic stereoisomers for membrane protein
1388 study: importance of chirality in the linker region, *Chem. Sci.* 8 (2017) 1169–1177.
1389 doi:10.1039/c6sc02981g.
- 1390 [32] P.S. Chae, M.J. Wander, K.H. Cho, P.D. Laible, S.H. Gellman, Carbohydrate-containing
1391 Triton X-100 analogues for membrane protein solubilization and stabilization, *Mol. Biosyst.* 9 (2013)
1392 626–629. doi:10.1039/c3mb25584k.
- 1393 [33] P.S. Chae, H.E. Bae, M. Das, Adamantane-based amphiphiles (ADAs) for membrane protein
1394 study: importance of a detergent hydrophobic group in membrane protein solubilisation, *Chem.*
1395 *Commun. Camb. Engl.* 50 (2014) 12300–12303. doi:10.1039/c4cc05746e.
- 1396 [34] S.G.F. Rasmussen, H.-J. Choi, J.J. Fung, E. Pardon, P. Casarosa, P.S. Chae, B.T. Devree,
1397 D.M. Rosenbaum, F.S. Thian, T.S. Kobilka, A. Schnapp, I. Konetzki, R.K. Sunahara, S.H. Gellman,
1398 A. Pautsch, J. Steyaert, W.I. Weis, B.K. Kobilka, Structure of a nanobody-stabilized active state of the
1399 $\beta(2)$ adrenoceptor, *Nature.* 469 (2011) 175–180. doi:10.1038/nature09648.
- 1400 [35] D.M. Rosenbaum, C. Zhang, J.A. Lyons, R. Holl, D. Aragao, D.H. Arlow, S.G.F. Rasmussen,
1401 H.-J. Choi, B.T. Devree, R.K. Sunahara, P.S. Chae, S.H. Gellman, R.O. Dror, D.E. Shaw, W.I. Weis,
1402 M. Caffrey, P. Gmeiner, B.K. Kobilka, Structure and function of an irreversible agonist- $\beta(2)$
1403 adrenoceptor complex, *Nature.* 469 (2011) 236–240. doi:10.1038/nature09665.
- 1404 [36] T.T. Selao, R. Branca, P.S. Chae, J. Lehtiö, S.H. Gellman, S.G.F. Rasmussen, S. Nordlund, A.
1405 Norén, Identification of chromatophore membrane protein complexes formed under different nitrogen
1406 availability conditions in *Rhodospirillum rubrum*, *J. Proteome Res.* 10 (2011) 2703–2714.
1407 doi:10.1021/pr100838x.
- 1408 [37] X. Jiang, L. Guan, Y. Zhou, W.-X. Hong, Q. Zhang, H.R. Kaback, Evidence for an
1409 intermediate conformational state of LacY, *Proc. Natl. Acad. Sci. U. S. A.* 109 (2012) E698-704.
1410 doi:10.1073/pnas.1201107109.
- 1411 [38] A. Amin, P. Hariharan, P.S. Chae, L. Guan, Effect of Detergents on Galactoside Binding by
1412 Melibiose Permeases, *Biochemistry.* 54 (2015) 5849–5855. doi:10.1021/acs.biochem.5b00660.
- 1413 [39] C. Nasrallah, K. Rottier, R. Marcellin, V. Compan, J. Font, A. Llebaria, J.-P. Pin, J.-L.
1414 Banères, G. Lebon, Direct coupling of detergent purified human mGlu5 receptor to the heterotrimeric
1415 G proteins Gq and Gs, *Sci. Rep.* 8 (2018) 4407. doi:10.1038/s41598-018-22729-4.
- 1416 [40] T.S. Owen, D. Salom, W. Sun, K. Palczewski, Increasing the Stability of Recombinant Human
1417 Green Cone Pigment, *Biochemistry.* 57 (2018) 1022–1030. doi:10.1021/acs.biochem.7b01118.
- 1418 [41] C. Perez, S. Gerber, J. Boilevin, M. Bucher, T. Darbre, M. Aebi, J.-L. Reymond, K.P. Locher,
1419 Structure and mechanism of an active lipid-linked oligosaccharide flippase, *Nature.* 524 (2015) 433–

1420 438. doi:10.1038/nature14953.

1421 [42] E.M. Quistgaard, M. Martinez Molledo, C. Löw, Structure determination of a major facilitator
1422 peptide transporter: Inward facing PepTSt from *Streptococcus thermophilus* crystallized in space
1423 group P3121, *PloS One*. 12 (2017) e0173126. doi:10.1371/journal.pone.0173126.

1424 [43] Y. Gao, G. Westfield, J.W. Erickson, R.A. Cerione, G. Skinotis, S. Ramachandran, Isolation
1425 and structure-function characterization of a signaling-active rhodopsin-G protein complex, *J. Biol.*
1426 *Chem.* 292 (2017) 14280–14289. doi:10.1074/jbc.M117.797100.

1427 [44] K.Y. Chung, T.H. Kim, A. Manglik, R. Alvares, B.K. Kobilka, R.S. Prosser, Role of
1428 detergents in conformational exchange of a G protein-coupled receptor, *J. Biol. Chem.* 287 (2012)
1429 36305–36311. doi:10.1074/jbc.M112.406371.

1430 [45] K.Y. Chung, S.G.F. Rasmussen, T. Liu, S. Li, B.T. DeVree, P.S. Chae, D. Calinski, B.K.
1431 Kobilka, V.L. Woods Jr, R.K. Sunahara, Conformational changes in the G protein Gs induced by the
1432 β_2 adrenergic receptor, *Nature*. 477 (2011) 611–615. doi:10.1038/nature10488.

1433 [46] G.H. Westfield, S.G.F. Rasmussen, M. Su, S. Dutta, B.T. DeVree, K.Y. Chung, D. Calinski,
1434 G. Velez-Ruiz, A.N. Oleskie, E. Pardon, P.S. Chae, T. Liu, S. Li, V.L. Woods Jr, J. Steyaert, B.K.
1435 Kobilka, R.K. Sunahara, G. Skinotis, Structural flexibility of the G alpha s alpha-helical domain in
1436 the beta2-adrenoceptor Gs complex, *Proc. Natl. Acad. Sci. U. S. A.* 108 (2011) 16086–16091.
1437 doi:10.1073/pnas.1113645108.

1438 [47] F. Hauer, C. Gerle, N. Fischer, A. Oshima, K. Shinzawa-Itoh, S. Shimada, K. Yokoyama, Y.
1439 Fujiyoshi, H. Stark, GraDeR: Membrane Protein Complex Preparation for Single-Particle Cryo-EM,
1440 *Struct. Lond. Engl.* 1993. 23 (2015) 1769–1775. doi:10.1016/j.str.2015.06.029.

1441 [48] T.E. Kraft, R.C. Hresko, P.W. Hruz, Expression, purification, and functional characterization
1442 of the insulin-responsive facilitative glucose transporter GLUT4, *Protein Sci. Publ. Protein Soc.* 24
1443 (2015) 2008–2019. doi:10.1002/pro.2812.

1444 [49] K.H. Cho, M. Husri, A. Amin, K. Gotfryd, H.J. Lee, J. Go, J.W. Kim, C.J. Loland, L. Guan,
1445 B. Byrne, P.S. Chae, Maltose neopentyl glycol-3 (MNG-3) analogues for membrane protein study,
1446 *The Analyst*. 140 (2015) 3157–3163. doi:10.1039/c5an00240k.

1447 [50] S.E. Rollauer, M.J. Tarry, J.E. Graham, M. Jääskeläinen, F. Jäger, S. Johnson, M.
1448 Krehenbrink, S.-M. Liu, M.J. Lukey, J. Marcoux, M.A. McDowell, F. Rodriguez, P. Roversi, P.J.
1449 Stansfeld, C.V. Robinson, M.S.P. Sansom, T. Palmer, M. Högbom, B.C. Berks, S.M. Lea, Structure of
1450 the TatC core of the twin-arginine protein transport system, *Nature*. 492 (2012) 210–214.
1451 doi:10.1038/nature11683.

1452 [51] B.H. Thurtle-Schmidt, R.M. Stroud, Structure of Bor1 supports an elevator transport
1453 mechanism for SLC4 anion exchangers, *Proc. Natl. Acad. Sci. U. S. A.* 113 (2016) 10542–10546.
1454 doi:10.1073/pnas.1612603113.

1455 [52] J. Guo, W. Zeng, Q. Chen, C. Lee, L. Chen, Y. Yang, C. Cang, D. Ren, Y. Jiang, Structure of
1456 the voltage-gated two-pore channel TPC1 from *Arabidopsis thaliana*, *Nature*. 531 (2016) 196–201.
1457 doi:10.1038/nature16446.

1458 [53] K.W. Huynh, M.R. Cohen, J. Jiang, A. Samanta, D.T. Lodowski, Z.H. Zhou, V.Y.
1459 Moiseenkova-Bell, Structure of the full-length TRPV2 channel by cryo-EM, *Nat. Commun.* 7 (2016)
1460 11130. doi:10.1038/ncomms11130.

1461 [54] A. Oshima, K. Tani, Y. Fujiyoshi, Atomic structure of the innexin-6 gap junction channel
1462 determined by cryo-EM, *Nat. Commun.* 7 (2016) 13681. doi:10.1038/ncomms13681.

- 1463 [55] S. Dang, S. Feng, J. Tien, C.J. Peters, D. Bulkley, M. Lolicato, J. Zhao, K. Zuberbühler, W.
1464 Ye, L. Qi, T. Chen, C.S. Craik, Y.N. Jan, D.L. Minor, Y. Cheng, L.Y. Jan, Cryo-EM structures of the
1465 TMEM16A calcium-activated chloride channel, *Nature*. 552 (2017) 426–429.
1466 doi:10.1038/nature25024.
- 1467 [56] A.D. Ferguson, J. Breed, K. Diederichs, W. Welte, J.W. Coulton, An internal affinity-tag for
1468 purification and crystallization of the siderophore receptor FhuA, integral outer membrane protein
1469 from *Escherichia coli* K-12, *Protein Sci.* 7 (1998) 1636–8.
- 1470 [57] M. Roth, P. Carpentier, O. Kaikati, J. Joly, P. Charrault, M. Pirocchi, R. Kahn, E. Fanchon, L.
1471 Jacquamet, F. Borel, A. Bertoni, P. Israel-Gouy, J.L. Ferrer, FIP: a highly automated beamline for
1472 multiwavelength anomalous diffraction experiments, *Acta Crystallogr. D Biol. Crystallogr.* 58 (2002)
1473 805–814.
- 1474 [58] W. Kabsch, XDS, *Acta Crystallogr. D Biol. Crystallogr.* 66 (2010) 125–132.
1475 doi:10.1107/S0907444909047337.
- 1476 [59] A.J. McCoy, R.W. Grosse-Kunstleve, P.D. Adams, M.D. Winn, L.C. Storoni, R.J. Read,
1477 Phaser crystallographic software, *J. Appl. Crystallogr.* 40 (2007) 658–674.
1478 doi:10.1107/S0021889807021206.
- 1479 [60] P.V. Afonine, R.W. Grosse-Kunstleve, N. Echols, J.J. Headd, N.W. Moriarty, M.
1480 Mustyakimov, T.C. Terwilliger, A. Urzhumtsev, P.H. Zwart, P.D. Adams, Towards automated
1481 crystallographic structure refinement with phenix.refine, *Acta Crystallogr. D Biol. Crystallogr.* 68
1482 (2012) 352–367. doi:10.1107/S0907444912001308.
- 1483 [61] J.J. Headd, N. Echols, P.V. Afonine, R.W. Grosse-Kunstleve, V.B. Chen, N.W. Moriarty,
1484 D.C. Richardson, J.S. Richardson, P.D. Adams, Use of knowledge-based restraints in phenix.refine to
1485 improve macromolecular refinement at low resolution, *Acta Crystallogr. D Biol. Crystallogr.* 68
1486 (2012) 381–390. doi:10.1107/S0907444911047834.
- 1487 [62] C. Hajjar, M.V. Cherrier, G. Dias Mirandela, I. Petit-Hartlein, M.J. Stasia, J.C. Fontecilla-
1488 Camps, F. Fieschi, J. Dupuy, The NOX Family of Proteins Is Also Present in Bacteria, *MBio*. 8
1489 (2017). doi:10.1128/mBio.01487-17.
- 1490 [63] E. Steinfels, C. Orelle, O. Dalmas, F. Penin, B. Miroux, A. Di Pietro, J.-M. Jault, Highly
1491 efficient over-production in *E. coli* of YvcC, a multidrug-like ATP-binding cassette transporter from
1492 *Bacillus subtilis*, *Biochim. Biophys. Acta*. 1565 (2002) 1–5.
- 1493 [64] C. Orelle, O. Dalmas, P. Gros, A. Di Pietro, J.-M. Jault, The conserved glutamate residue
1494 adjacent to the Walker-B motif is the catalytic base for ATP hydrolysis in the ATP-binding cassette
1495 transporter BmrA, *J. Biol. Chem.* 278 (2003) 47002–47008. doi:10.1074/jbc.M308268200.
- 1496 [65] A. Salvay, C. Ebel, Analytical Ultracentrifuge for the Characterization of Detergent in
1497 Solution, in: *Prog. Colloid Polym. Sci.*, 2006: pp. 74–82. doi:10.1007/2882_006.
- 1498 [66] P. Schuck, Size-distribution analysis of macromolecules by sedimentation velocity
1499 ultracentrifugation and lamm equation modeling, *Biophys. J.* 78 (2000) 1606–1619.
1500 doi:10.1016/S0006-3495(00)76713-0.
- 1501 [67] H. Zhao, R. Ghirlando, C. Alfonso, F. Arisaka, I. Attali, D.L. Bain, M.M. Bakhtina, D.F.
1502 Becker, G.J. Bedwell, A. Bekdemir, T.M.D. Besong, C. Birck, C.A. Brautigam, W. Brennerman, O.
1503 Byron, A. Bzowska, J.B. Chaires, C.T. Chaton, H. Cölfen, K.D. Connaghan, K.A. Crowley, U. Curth,
1504 T. Daviter, W.L. Dean, A.I. Díez, C. Ebel, D.M. Eckert, L.E. Eisele, E. Eisenstein, P. England, C.
1505 Escalante, J.A. Fagan, R. Fairman, R.M. Finn, W. Fischle, J.G. de la Torre, J. Gor, H. Gustafsson, D.
1506 Hall, S.E. Harding, J.G.H. Cifre, A.B. Herr, E.E. Howell, R.S. Isaac, S.-C. Jao, D. Jose, S.-J. Kim, B.

1507 Kokona, J.A. Kornblatt, D. Kosek, E. Krayukhina, D. Krzizike, E.A. Kuszniir, H. Kwon, A. Larson,
1508 T.M. Laue, A. Le Roy, A.P. Leech, H. Lilie, K. Luger, J.R. Luque-Ortega, J. Ma, C.A. May, E.L.
1509 Maynard, A. Modrak-Wojcik, Y.-F. Mok, N. Mücke, L. Nagel-Steger, G.J. Narlikar, M. Noda, A.
1510 Nourse, T. Obsil, C.K. Park, J.-K. Park, P.D. Pawelek, E.E. Perdue, S.J. Perkins, M.A. Perugini, C.L.
1511 Peterson, M.G. Peverelli, G. Piszczek, G. Prag, P.E. Prevelige, B.D.E. Raynal, L. Rezabkova, K.
1512 Richter, A.E. Ringel, R. Rosenberg, A.J. Rowe, A.C. Rufer, D.J. Scott, J.G. Seravalli, A.S. Solovyova,
1513 R. Song, D. Staunton, C. Stoddard, K. Stott, H.M. Strauss, W.W. Streicher, J.P. Sumida, S.G.
1514 Swygert, R.H. Szczepanowski, I. Tessmer, R.T. Toth, A. Tripathy, S. Uchiyama, S.F.W. Uebel, S.
1515 Unzai, A.V. Gruber, P.H. von Hippel, C. Wandrey, S.-H. Wang, S.E. Weitzel, B. Wielgus-Kutrowska,
1516 C. Wolberger, M. Wolff, E. Wright, Y.-S. Wu, J.M. Wubben, P. Schuck, A multilaboratory
1517 comparison of calibration accuracy and the performance of external references in analytical
1518 ultracentrifugation, *PLoS One*. 10 (2015) e0126420. doi:10.1371/journal.pone.0126420.

1519 [68] C.A. Brautigam, Calculations and Publication-Quality Illustrations for Analytical
1520 Ultracentrifugation Data, *Methods Enzymol.* 562 (2015) 109–133. doi:10.1016/bs.mie.2015.05.001.

1521 [69] A. Le Roy, H. Nury, B. Wiseman, J. Sarwan, J.-M. Jault, C. Ebel, Sedimentation velocity
1522 analytical ultracentrifugation in hydrogenated and deuterated solvents for the characterization of
1523 membrane proteins, *Methods Mol. Biol.* Clifton NJ. 1033 (2013) 219–251. doi:10.1007/978-1-62703-
1524 487-6_15.

1525 [70] A. Le Roy, K. Wang, B. Schaack, P. Schuck, C. Breyton, C. Ebel, AUC and Small-Angle
1526 Scattering for Membrane Proteins, *Methods Enzymol.* 562 (2015) 257–286.
1527 doi:10.1016/bs.mie.2015.06.010.

1528 [71] A.G. Salvay, M. Santamaria, M. le Maire, C. Ebel, Analytical ultracentrifugation
1529 sedimentation velocity for the characterization of detergent-solubilized membrane proteins Ca⁺⁺-
1530 ATPase and ExbB, *J. Biol. Phys.* 33 (2007) 399–419. doi:10.1007/s10867-008-9058-3.

1531 [72] P. Pernot, A. Round, R. Barrett, A. De Maria Antolinos, A. Gobbo, E. Gordon, J. Huet, J.
1532 Kieffer, M. Lentini, M. Mattenet, C. Morawe, C. Mueller-Dieckmann, S. Ohlsson, W. Schmid, J. Surr,
1533 P. Theveneau, L. Zerrad, S. McSweeney, Upgraded ESRF BM29 beamline for SAXS on
1534 macromolecules in solution, *J. Synchrotron Radiat.* 20 (2013) 660–664.
1535 doi:10.1107/S0909049513010431.

1536 [73] M.V. Petoukhov, D. Franke, A.V. Shkumatov, G. Tria, A.G. Kikhney, M. Gajda, C. Gorba,
1537 H.D.T. Mertens, P.V. Konarev, D.I. Svergun, New developments in the ATSAS program package for
1538 small-angle scattering data analysis, *J. Appl. Crystallogr.* 45 (2012) 342–350.
1539 doi:10.1107/S0021889812007662.

1540 [74] C. Ebel, C. Breyton, A. Martel, Examining membrane proteins by neutron scattering, in:
1541 *Biophys. Membr. Proteins Methods Protoc.*, Springer, Vincent Postis and Adrian Goldmann, 2018.

1542 [75] S.R. Kline, Reduction and analysis of SANS and USANS data using IGOR Pro, *J. Appl.*
1543 *Crystallogr.* 39 (2006) 895–900. doi:10.1107/S0021889806035059.

1544 [76] M. Doucet, J.H. Cho, G. Alina, J. Bakker, W. Bouwman, P. Butler, K. Campbell, M.
1545 Gonzales, R. Heenan, A. Jackson, P. Juhas, S. King, P. Kienzle, J. Krzywon, A. Markvardsen, T.
1546 Nielsen, L. O’Driscoll, W. Potrzebowski, R. Ferraz Leal, T. Richter, P. Rozycko, T. Snow, A.
1547 Washington, SasView version 4.2, Zenodo, 2018. doi:10.5281/zenodo.1412041.

1548 [77] C. Breyton, F. Gabel, M. Lethier, A. Flayhan, G. Durand, J.-M. Jault, C. Juillan-Binard, L.
1549 Imbert, M. Moulin, S. Ravaud, M. Härtlein, C. Ebel, Small angle neutron scattering for the study of
1550 solubilised membrane proteins, *Eur. Phys. J. E Soft Matter.* 36 (2013) 9889. doi:10.1140/epje/i2013-
1551 13071-6.

- 1552 [78] A. Flayhan, F. Wien, M. Paternostre, P. Boulanger, C. Breyton, New insights into pb5, the
1553 receptor binding protein of bacteriophage T5, and its interaction with its Escherichia coli receptor
1554 FhuA, *Biochimie*. 94 (2012) 1982–1989. doi:10.1016/j.biochi.2012.05.021.
- 1555 [79] C. Breyton, A. Flayhan, F. Gabel, M. Lethier, G. Durand, P. Boulanger, M. Chami, C. Ebel,
1556 Assessing the Conformational Changes of pb5, the Receptor-binding Protein of Phage T5, upon
1557 Binding to Its Escherichia coli Receptor FhuA, *J. Biol. Chem.* 288 (2013) 30763–30772.
1558 doi:10.1074/jbc.M113.501536.
- 1559 [80] A.D. Ferguson, E. Hofmann, J.W. Coulton, K. Diederichs, W. Welte, Siderophore-mediated
1560 iron transport: crystal structure of FhuA with bound lipopolysaccharide, *Science*. 282 (1998) 2215–20.
- 1561 [81] M. Bonhivers, M. Desmadril, G.S. Moeck, P. Boulanger, A. Colomer-Pallas, L. Letellier,
1562 Stability studies of FhuA, a two-domain outer membrane protein from Escherichia coli, *Biochemistry*.
1563 40 (2001) 2606–13.
- 1564 [82] P. Boulanger, M. le Maire, M. Bonhivers, S. Dubois, M. Desmadril, L. Letellier, Purification
1565 and structural and functional characterization of FhuA, a transporter of the Escherichia coli outer
1566 membrane, *Biochemistry*. 35 (1996) 14216–14224. doi:10.1021/bi9608673.
- 1567 [83] D. Durand, C. Vivès, D. Cannella, J. Pérez, E. Pebay-Peyroula, P. Vachette, F. Fieschi,
1568 NADPH oxidase activator p67(phox) behaves in solution as a multidomain protein with semi-flexible
1569 linkers, *J. Struct. Biol.* 169 (2010) 45–53. doi:10.1016/j.jsb.2009.08.009.
- 1570 [84] J. Marcoux, P. Man, I. Petit-Haertlein, C. Vivès, E. Forest, F. Fieschi, p47phox molecular
1571 activation for assembly of the neutrophil NADPH oxidase complex, *J. Biol. Chem.* 285 (2010) 28980–
1572 28990. doi:10.1074/jbc.M110.139824.
- 1573 [85] F. Magnani, S. Nenci, E. Millana Fananas, M. Ceccon, E. Romero, M.W. Fraaije, A. Mattevi,
1574 Crystal structures and atomic model of NADPH oxidase, *Proc. Natl. Acad. Sci. U. S. A.* 114 (2017)
1575 6764–6769. doi:10.1073/pnas.1702293114.
- 1576 [86] E. Steinfels, C. Orelle, J.-R. Fantino, O. Dalmas, J.-L. Rigaud, F. Denizot, A. Di Pietro, J.-M.
1577 Jault, Characterization of YvcC (BmrA), a multidrug ABC transporter constitutively expressed in
1578 *Bacillus subtilis*, *Biochemistry*. 43 (2004) 7491–7502. doi:10.1021/bi0362018.
- 1579 [87] A. Ward, C.L. Reyes, J. Yu, C.B. Roth, G. Chang, Flexibility in the ABC transporter MsbA:
1580 Alternating access with a twist, *Proc. Natl. Acad. Sci. U. S. A.* 104 (2007) 19005–19010.
1581 doi:10.1073/pnas.0709388104.
- 1582 [88] M.S. Jin, M.L. Oldham, Q. Zhang, J. Chen, Crystal structure of the multidrug transporter P-
1583 glycoprotein from *Caenorhabditis elegans*, *Nature*. 490 (2012) 566–569. doi:10.1038/nature11448.
- 1584 [89] S. Ravaud, M.-A. Do Cao, M. Jidenko, C. Ebel, M. Le Maire, J.-M. Jault, A. Di Pietro, R.
1585 Haser, N. Aghajari, The ABC transporter BmrA from *Bacillus subtilis* is a functional dimer when in a
1586 detergent-solubilized state, *Biochem. J.* 395 (2006) 345–53.
- 1587 [90] O. Dalmas, M.-A. Do Cao, M.R. Lugo, F.J. Sharom, A. Di Pietro, J.-M. Jault, Time-resolved
1588 fluorescence resonance energy transfer shows that the bacterial multidrug ABC half-transporter BmrA
1589 functions as a homodimer, *Biochemistry*. 44 (2005) 4312–4321. doi:10.1021/bi0482809.
- 1590 [91] C. Orelle, F. Gubellini, A. Durand, S. Marco, D. Lévy, P. Gros, A. Di Pietro, J.-M. Jault,
1591 Conformational change induced by ATP binding in the multidrug ATP-binding cassette transporter
1592 BmrA, *Biochemistry*. 47 (2008) 2404–2412. doi:10.1021/bi702303s.
- 1593 [92] M.-A. Do Cao, S. Crouzy, M. Kim, M. Becchi, D.S. Cafiso, A. Di Pietro, J.-M. Jault, Probing
1594 the conformation of the resting state of a bacterial multidrug ABC transporter, BmrA, by a site-

1595 directed spin labeling approach, *Protein Sci. Publ. Protein Soc.* 18 (2009) 1507–1520.
1596 doi:10.1002/pro.141.

1597 [93] S. Mehmood, C. Domene, E. Forest, J.-M. Jault, Dynamics of a bacterial multidrug ABC
1598 transporter in the inward- and outward-facing conformations, *Proc. Natl. Acad. Sci. U. S. A.* 109
1599 (2012) 10832–10836. doi:10.1073/pnas.1204067109.

1600 [94] P.F. Fribourg, M. Chami, C.O.S. Sorzano, F. Gubellini, R. Marabini, S. Marco, J.-M. Jault, D.
1601 Lévy, 3D cryo-electron reconstruction of BmrA, a bacterial multidrug ABC transporter in an inward-
1602 facing conformation and in a lipidic environment, *J. Mol. Biol.* 426 (2014) 2059–2069.
1603 doi:10.1016/j.jmb.2014.03.002.

1604 [95] B. Wiseman, A. Kilburg, V. Chaptal, G.C. Reyes-Mejia, J. Sarwan, P. Falson, J.-M. Jault,
1605 Stubborn contaminants: influence of detergents on the purity of the multidrug ABC transporter BmrA,
1606 *PloS One.* 9 (2014) e114864. doi:10.1371/journal.pone.0114864.

1607 [96] J. Lipfert, L. Columbus, V.B. Chu, S.A. Lesley, S. Doniach, Size and shape of detergent
1608 micelles determined by small-angle X-ray scattering, *J. Phys. Chem. B.* 111 (2007) 12427–12438.
1609 doi:10.1021/jp073016l.

1610 [97] R.C. Oliver, S.V. Pingali, V.S. Urban, Designing Mixed Detergent Micelles for Uniform
1611 Neutron Contrast, *J. Phys. Chem. Lett.* 8 (2017) 5041–5046. doi:10.1021/acs.jpcclett.7b02149.

1612 [98] J.N. Israelachvili, D.J. Mitchell, B.W. Barry W. Ninham, Theory of self-assembly of lipid
1613 bilayers and vesicles, *Biochim Biophys Acta - Biomembr.* 470 (1977) 185–201.

1614 [99] J.N. Israelachvili, *Intermolecular and Surface Forces - 3rd Edition*, London: Academic Press,
1615 1992. [https://www.elsevier.com/books/intermolecular-and-surface-forces/israelachvili/978-0-12-](https://www.elsevier.com/books/intermolecular-and-surface-forces/israelachvili/978-0-12-391927-4)
1616 [391927-4](https://www.elsevier.com/books/intermolecular-and-surface-forces/israelachvili/978-0-12-391927-4) (accessed July 24, 2018).

1617 [100] C. Breyton, F. Gabel, M. Abla, Y. Pierre, F. Lebaupain, G. Durand, J.-L. Popot, C. Ebel, B.
1618 Pucci, Micellar and biochemical properties of (hemi)fluorinated surfactants are controlled by the size
1619 of the polar head, *Biophys. J.* 97 (2009) 1077–1086. doi:10.1016/j.bpj.2009.05.053.

1620 [101] E. Boncoeur, C. Durmort, B. Bernay, C. Ebel, A.M. Di Guilmi, J. Croizé, T. Vernet, J.-M.
1621 Jault, PatA and PatB form a functional heterodimeric ABC multidrug efflux transporter responsible for
1622 the resistance of *Streptococcus pneumoniae* to fluoroquinolones, *Biochemistry.* 51 (2012) 7755–7765.
1623 doi:10.1021/bi300762p.

1624 [102] E. Pebay-Peyroula, G. Rummel, J.P. Rosenbusch, E. Landau, X-ray structure of
1625 bacteriorhodopsin at 2.5 Å from microcrystals grown in lipidic cubic phases, *Science.* 277 (1997)
1626 1676–1881.

1627