



HAL
open science

The Complex Fine-Tuning of K⁺ Fluxes in Plants in Relation to Osmotic and Ionic Abiotic Stresses

Isabelle Chérel, Isabelle Gaillard

► To cite this version:

Isabelle Chérel, Isabelle Gaillard. The Complex Fine-Tuning of K⁺ Fluxes in Plants in Relation to Osmotic and Ionic Abiotic Stresses. *International Journal of Molecular Sciences*, 2019, 20 (3), pp.715. <10.3390/ijms20030715>. <hal-02054481>

HAL Id: hal-02054481

<https://hal.science/hal-02054481v1>

Submitted on 20 Sep 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons CC BY 4.0 - Attribution - International License



Review

The Complex Fine-Tuning of K⁺ Fluxes in Plants in Relation to Osmotic and Ionic Abiotic Stresses

Isabelle Chérel * and Isabelle Gaillard

BPMP, Univ Montpellier, CNRS, INRA, SupAgro, Montpellier, France; isabelle.gaillard@inra.fr

* Correspondence: isabelle.cherel@inra.fr; Tel.: +33-4-99612565

Received: 29 November 2018; Accepted: 29 January 2019; Published: 7 February 2019



Abstract: As the main cation in plant cells, potassium plays an essential role in adaptive responses, especially through its involvement in osmotic pressure and membrane potential adjustments. K⁺ homeostasis must, therefore, be finely controlled. As a result of different abiotic stresses, especially those resulting from global warming, K⁺ fluxes and plant distribution of this ion are disturbed. The hormone abscisic acid (ABA) is a key player in responses to these climate stresses. It triggers signaling cascades that ultimately lead to modulation of the activities of K⁺ channels and transporters. After a brief overview of transcriptional changes induced by abiotic stresses, this review deals with the post-translational molecular mechanisms in different plant organs, in *Arabidopsis* and species of agronomical interest, triggering changes in K⁺ uptake from the soil, K⁺ transport and accumulation throughout the plant, and stomatal regulation. These modifications involve phosphorylation/dephosphorylation mechanisms, modifications of targeting, and interactions with regulatory partner proteins. Interestingly, many signaling pathways are common to K⁺ and Cl⁻/NO₃⁻ counter-ion transport systems. These cross-talks are also addressed.

Keywords: plants; abiotic stress; abscisic acid; potassium transport regulation

1. Introduction

Global warming is impacting crop yield and quality. Increases in atmospheric CO₂ and light intensity, erratic rainfalls, periods of drought stress, and heat waves are the climatic components most affecting terrestrial plants. Also, an increase in irrigation use leads to soil salinization. Drought and heat stresses, especially when combined, are responsible for significant losses in agriculture [1–3]. Different strategies can be applied to lessen their effects, relying on modifications of agricultural practices, isolation of the best genotypes, and transgenesis [3]. This requires knowledge of the mechanisms that govern plant adaptation to these adverse conditions. The different forms of signal transduction in response to drought, salt, and heat stresses from membrane sensors to the terminal cell responses in plant cells were documented [4]. Abscisic acid (ABA) plays an essential role in responses to drought and salt/osmotic stresses [4–7] and data obtained with the plant model *Arabidopsis thaliana* also indicate that ABA plays a role in the adaptation to high temperatures [6,8,9].

In plants, the synthesis of the phytohormone ABA, which probably occurs mainly in vascular tissues [10], is induced by abiotic stresses [5]. ABA is then transported to target tissues via both xylem and phloem, which allows transport in both directions between roots and shoots [10,11]. Also, drought stress leads to a transient pH increase in xylem sap and apoplast, favoring the dissociation of ABA to its ionic form that would be accumulated in the apoplast of all plant tissues [12]. This is thought to function as a root-to-shoot signal leading to leaf transpiration decrease. It is interesting to note that the transport of ABA to different sites leads to organ-specific and cell-type-specific responses. Finally, coordinated fluxes of K⁺ and anions across cellular membranes typically occur at the end of the biological responses to environmental stimuli.

Modulation of K^+ fluxes is a well-known response to ABA drought and salt stress. K^+ is the main cation in plant cells, essential for plant growth and adaptation to the environment [13,14]. Due to its abundance in plant cells, it is involved in neutralization of negative charges, pH homeostasis, and control of the electrical membrane potential [15]. Its concentration in the cytosol is maintained around 100–200 mM, and, in this compartment, it cannot be replaced by another cation such as Na^+ because the latter affects the water H-bonding at the protein surface more than K^+ [16]. In contrast, vacuolar K^+ , which is involved in cell turgor, can to some extent be replaced by other osmotica [17,18]. As a consequence of its function in cell turgor, K^+ is involved in nasties, including stomatal movements [19]. K^+ also displays biochemical properties. It is involved in the direct or indirect activation of enzymes involved in metabolic processes, such as carbon metabolism and protein synthesis [19]. In K^+ -deficient plants, photosynthesis is also impaired. This is due to the prominent role of K^+ in mesophyll CO_2 conductance, chloroplast organization, improvement of Rubisco activity, and translocation of photoassimilates in the phloem [20]. Under high light, K^+ limits reactive oxygen species (ROS) production that leads to photoinhibition [20]. Maintaining K^+ homeostasis even under stressful conditions is, therefore, a necessity for plant growth and adaptation [21]. Drought stress impairs K^+ uptake and, thus, exacerbates the effects of K^+ deficiency [21]. It also decreases K^+ translocation to the xylem [22], thus maintaining root growth [23], and triggers K^+ -mediated stomatal closure for preventing dehydration. Salinity also results in root and shoot decline in K^+ , salt tolerance being correlated to the ability to save K^+ , and to the cytosolic K^+/Na^+ ratio [21,24]. Indeed, it was documented that K^+ homeostasis is particularly critical for plant tolerance to drought and salt stresses [21,23] and high light intensity [25].

K^+ transport systems are targets of signaling pathways, leading to environmental responses such as K^+ uptake, translocation, and stomatal movements. Multiple channels and transporters were identified in plants [26]. In this review, we focus on regulations of K^+ transport systems which drive K^+ fluxes as essential end responses to different abiotic stresses and ABA signaling cascades, and on their relationships with those regarding mineral anion (Cl^- , NO_3^-) transports.

2. K^+ Transport in Plants

2.1. Structure and Function of K^+ Transport Systems

2.1.1. K^+ -Selective Channels

The Shaker channels: K^+ channels of the Shaker family mediate the major K^+ fluxes at the plasma membrane [27], and are the best characterized. They are tetramers of four subunits arranged around a central pore, selective for K^+ ions. Each subunit is made up of a short cytosolic N-terminal sequence, a membrane part containing the pore-forming domain with the K^+ selectivity filter, and a large C-terminal intracytoplasmic part including a cyclic-nucleotide-binding domain, an ankyrin repeat protein–protein interaction domain, and a conserved hydrophobic and acidic (KHA) C-terminal end. In *Arabidopsis thaliana*, Shaker channels are encoded by a family of nine genes, which are grouped into five subfamilies (for reviews, see References [15,27]). The preferential assembly of these subunits as heterotetrameric structures enables a multiplicity of combinations increasing functional diversity [28–30]. Shaker channels are voltage-gated. Depending on the subfamily, Shaker subunits form inwardly rectifying (Kin) or outwardly rectifying (Kout) channels (activated by membrane hyperpolarization or depolarization, respectively). In *Arabidopsis*, AKT1, AKT5, AKT6 (*Arabidopsis* K^+ transporters 1, 5 and 6), KAT1, KAT2 (K^+ *Arabidopsis* transporters 1 and 2), and AtKC1 (*Arabidopsis thaliana* K^+ channel 1) subunits assemble as Kin channels, whereas SKOR (stelar K^+ outward rectifier) and GORK (guard cell outward rectifying channel) subunits form Kout channels [27]. No assembly could be detected between Kin and Kout channel subunits [31]. A notable exception to the strict voltage dependence is AKT2, a “weakly rectifying” channel, that switches from an inwardly rectifying to a non-rectifying state [31]. At the macroscopic level, this results in the superposition of two current components; a “leak-like” component of AKT2 channels open in

the whole range of physiological membrane potentials, and an inward rectifying component that closes upon depolarization. Furthermore, AtKC1 is thought to be a regulatory subunit. Indeed, AtKC1 subunits do not form functional homotetrameric channels on their own but they assemble with inward-channel-forming subunits to give rise to heteromeric channels that activate at much more hyperpolarized potentials compared to homotetramers [32–34]. Patch-clamp analyses on root hair protoplasts reveal that inward K^+ currents are likely to correspond to AKT1/AtKC1 heteromers rather than AKT1 homotetramers [35]. Shaker channels are present in all plant organs [36]. They are involved in main functions such as uptake of K^+ from the soil (even from K^+ solutions as dilute as 10 μ M) and its secretion into the xylem, transport of solutes in the phloem, stomatal movements, and cell elongation (Table 1). Members of the different Shaker gene subfamilies were identified in all studied plant species such as rice, tomato, maize, poplar, grapevine, etc. [27].

TPKs (Two-pore K^+ channels) constitute the other group of K^+ -selective channels. This family comprises six members in *Arabidopsis*. Except for KCO3 (Ca^{2+} activated outward rectifying K^+ channel 3), which could be an inactive channel, they are dimers of two-pore domain-subunits, composed of four transmembrane segments with two pore domains including the K^+ -selectivity motif GYGD. Of these, only TPK4 (two-pore K^+ transporter 1) is targeted to the plasma membrane [15,37], with the four other TPKs (TPK1, 2, 3, and 5) located on endocellular membranes. The roles of TPK channels are much less well understood than those of Shakers. TPK4, which is blocked by extracellular Ca^{2+} and cytoplasmic acidification, is an open rectifier channel. This channel is mainly expressed in pollen and is thought to contribute to the K^+ conductance of the pollen tube plasma membrane [38]. The subunits of TPK1, 2, 3 and 5 channels also contain binding sites for 14-3-3 proteins in the cytosolic N-terminal part and two Ca^{2+} -binding EF hands in the cytosolic C-terminal part. TPK3 is expressed in the stromal lamellae of thylakoid membranes where it controls the transmembrane Δ pH and, thus, the light-induced proton motive force [39]. The tonoplast TPK1 channel is ubiquitously expressed. It mediates the VK voltage-independent, K^+ -selective currents observed in guard cells and mesophyll cells [40,41]. In guard cells, it is important for vacuolar K^+ release during stomatal closure [40,41]. This channel is involved in intracellular K^+ homeostasis [40]. The biophysical features and physiological functions of TPK 2 and 5 remain unknown.

Table 1. Plant K⁺ transport systems described in this study and their regulations in response to ionic and osmotic stresses.

Gene Name ¹	ID	Protein Family	Function	Type of Stress	Transcriptional Regulation (Drought, ABA, Heat Stress, NaCl)	Protein Level/Activity
<i>AKT1</i>	At2g26650	Shaker	K ⁺ uptake by roots [42], inhibition of stomatal closure in response to drought [43]	ABA	No significant change in roots, downregulation in shoots [44] and seedlings [45]	Inhibition of currents by intracellular Na ⁺ [43]
				NaCl	No significant change in roots and shoots [44]	
<i>OsAKT1</i>	LOC4326245	Shaker	K ⁺ uptake by roots [46,47]	NaCl	Downregulation [48], downregulation in the exodermis in salt-tolerant lines [49]	
<i>VvK1.1</i>	VIT_11s0016g04750	Shaker	K ⁺ uptake in roots and phloem [50]	Drought	Induction in leaves and berries, downregulation in roots [50]	
				ABA	Induction in leaves, not in roots [50]	
<i>VvK1.2</i>	VIT_04s0008g04990	Shaker	K ⁺ uptake by flesh cells [51]	Drought	Strong induction in berries from véraison [51]	
				ABA	Induction in leaves [44,54], not in guard cells [55]	
<i>AKT2</i>	At4g22200	Shaker	Regulation of phloem membrane potential [52,53]	Mannitol, NaCl, drought	Moderate induction in leaves [56]	
				NaCl	Induction [57]	
				Heat	Induction [58]	
<i>VvK3.1</i>	VIT_12s0034g02240	Shaker	Phloem transport, leaf movements [59]	Drought	Induction in leaves and (moderately) in berries, not in roots [59]	
<i>KAT1</i>	At5g46240	Shaker	Stomatal opening [60,61]	ABA	Down-regulation in guard cells [55]	
				Drought	No change in guard cells [62]	
				NaCl	No significant change [44]	
<i>KAT2</i>	At4g18290	Shaker	Stomatal opening [60,61]	ABA	Down-regulation in guard cells [55]	
<i>AtKC1</i>	At4g32650	Shaker	K ⁺ uptake in root hairs together with AKT1 [35]	ABA	Transient down-regulation [44]	
				NaCl	High induction in leaves, not in roots [44]	
<i>SKOR</i>	At3g02850	Shaker	K ⁺ loading of the xylem in roots [63]	ABA	Downregulation [63]	
				NaCl	No significant change [44], strong induction [57]	
<i>GORK</i>	At5g37500	Shaker	Stomatal closure [64]	ABA	Induction in seedlings, cultured cells, root hair protoplasts, not in guard cells [65]	
				Heat	Induction in roots [56]	
				Water stress	Induction in leaves [65]	
				NaCl	Induction in roots [56]	

Table 1. Cont.

Gene Name ¹	ID	Protein Family	Function	Type of Stress	Transcriptional Regulation (Drought, ABA, Heat Stress, NaCl)	Protein Level/Activity
<i>HAK5</i>	At4g13420	KUP	High-affinity K ⁺ uptake from the soil [66]	NaCl	Strong downregulation [67]	
				NaCl, drought, mannitol	Unclear [56]	
<i>KUP6</i>	At1g70300	KUP	K ⁺ efflux from roots and stomata, ABA- and auxin-dependent inhibition of lateral root formation [68]	NaCl	Induction [57]	
				Heat	Induction (Genevestigator, https://genevestigator.com , [58])	
				Mannitol	Strong induction [56]	
<i>KUP8</i>	At5g14880	KUP	K ⁺ efflux from roots and stomata, ABA- and auxin-dependent inhibition of lateral root formation [68]	ABA, NaCl, mannitol	Slight downregulation [56], especially in roots (NaCl, mannitol)	
<i>OsHAK1</i>	LOC4335729	KUP	K ⁺ acquisition [69]	Water stress	Transient induction in roots and shoots [69]	
<i>OsHAK5</i>	LOC4326945	KUP	K ⁺ acquisition, root to shoot transport [70]	NaCl	Induction in roots and shoots [70]	
<i>OsHAK21</i>	LOC9269115	KUP	K ⁺ uptake and distribution [71]	NaCl	Induction in roots and shoots [71]	
<i>TPK1</i>	At1g02880	Two-pore channels	K ⁺ homeostasis, guard cell vacuolar K ⁺ release [41]	NaCl	No significant change [72]	Induction of protein phosphorylation [72]
				ABA	No change [45,73]	
<i>SLAC1</i>	At1g12480	C4 dicarboxylate transporters	Stomatal closure [74,75]	Drought	Short treatment: no change in roots and shoots [66], strong induction in guard cell protoplasts after a few days [62]	
				ABA	No change [45,73,76]	
				NaCl	Moderate induction in shoots [56]	
				Mannitol	No significant change in roots and shoots [56]	
<i>SLAH3</i>	At5g24030	C4 dicarboxylate transporters	Cl ⁻ root to shoot translocation [77], NO ₃ ⁻ -mediated stomatal closure [78], pollen tube growth [79]	Drought	Short treatment: downregulation in roots [56]; strong induction in guard cell protoplasts after a few days [62]	
				ABA	No change [45]	
				NaCl	Downregulation in roots [56,77]	
				Mannitol	Downregulation in roots [56]	
				PEG	Strong downregulation in roots [77]	
<i>CHL1/NRT1.1/NPF6.3</i>	At1g12110	NPF	Nitrate uptake [80], stomatal opening [81], nitrate sensing, auxin transport [82]	ABA	Unclear, depends on tissue/experimental conditions [45]	
				NaCl, mannitol	Unclear, transient down-regulation in roots? [56]	

¹ Gene names: AKT: *Arabidopsis* K⁺ transporter, KAT: K⁺ *Arabidopsis* transporter, AtKC: *Arabidopsis thaliana* K⁺ channel, SKOR: stelar K⁺ outward rectifier, GORK: guard cell outward rectifying K⁺ channel, HAK: high affinity K⁺ transporter, KUP: K⁺ uptake permease, TPK: two-pore (or tandem pore) K⁺ channel, SLAC: slow anion channel, SLAH: SLAC1 Homologue, CHL: chlorate-resistant, NRT: Nitrate Transporter, NPF NRT1/PTR Family. Genes originate from *Arabidopsis thaliana* unless otherwise stated. *Os*: *Oryza sativa*, *Vv*: *Vitis vinifera*.

2.1.2. K⁺-Selective KUP/HAK/KT Transporters (K⁺ Uptake Permease, High Affinity K⁺, K⁺ Transporters)

This family comprises 13 members in *Arabidopsis*. The most popular are AtHAK5 from *Arabidopsis* and its homologs in other species, which mediate high-affinity K⁺ uptake in root from very dilute soil solutions, below 10 μM (reviewed by References [83–85]). The energy for K⁺ uptake would be provided by co-transport with protons [83]. KUP transporters display multiple functions in plants. In roots, AtKUP1 is a dual-affinity transporter mediating K⁺ uptake [86], whereas KUP 2, 4, and 6 negatively impact K⁺ (Rb⁺) uptake, and are supposed to mediate K⁺ efflux [68]. KUPs are found at the plasma membrane and in endomembranes, and, in addition to K⁺ uptake from the soil, they are also involved in K⁺ homeostasis, long-distance K⁺ transport, cell elongation, response to osmotic stress, and other unexpected functions such as adenylate cyclase activity and regulation of auxin transport [84].

2.1.3. Cation Transporters

The HKT transporter family is divided into two subfamilies, “Na⁺-selective” and “K⁺-permeable”, which can also transport Na⁺. The latter subfamily, which seems to be specific to monocots, is characterized by the presence of a conserved glycine in the selectivity filter [27]. HKT transporters mediate Na⁺ or K⁺ influx, or Na⁺/K⁺ co-transport [27,83]. NHX (Na⁺/H⁺ exchangers) and CHX (cation/H⁺ exchangers) are cation transporters found on endomembranes including the tonoplast, and CHX transporters are also present at the plasma membrane. Some of them are involved in the control of K⁺ homeostasis [83,87]. These families, poorly characterized with regard to post-translational regulatory networks, are not included in the present review.

2.2. Role of Potassium Transport Systems in Adaptation to Stress

Genes encoding membrane transporters and their transcriptional regulations by climate-related stresses (drought, osmotic stress, salt, heat) are presented in Table 1.

2.2.1. K⁺ Uptake and Transfer to the Xylem in Roots

As reported above, maintaining sufficient K⁺ uptake is critical to reduce the effects of drought stress. K⁺ is taken up from a soil solution that is highly variable in composition, with K⁺ concentration that can range from a few μM to several mM. In *Arabidopsis*, below 10 μM K⁺ and in the absence of NH₄⁺, only HAK5 is able to mediate K⁺ uptake, whereas AKT1 intervenes at higher, but still low, K⁺ concentrations, due to the high hyperpolarization of root cell membranes in K⁺-deficient soils [88,89]. Salinity strongly represses HAK5 expression in roots [67]. However, HAK5 is critically required for plant growth under low-K and saline conditions, because Na⁺ absorption results in depolarization of membranes of external root cells, which promotes an unusual K⁺ efflux via AKT1 [67]. In these experiments, the involvement of the outwardly rectifying GORK channel, also known to mediate K⁺ efflux under salt stress [90] (see below), was not addressed. Closest homologs of HAK5 belong to clade 1a of the KUP family [66]. They are involved (or thought to be involved) in K⁺ uptake from the soil at low K⁺ concentrations [27]. Like HAK5, homologous genes in other plant species (rice, barley, pepper, tomato, *Thellungiella halophila*) are highly upregulated by K⁺ shortage; however, for at least three of them, this upregulation is suppressed by Na⁺ [27]. Although AKT1 and HAK5 seem to be the main players for K⁺ uptake from the soil even in the low-K⁺ range [88], AtKUP7 in *Arabidopsis* (clade V [66]) was recently found to also contribute to K⁺ uptake under low-K⁺ conditions [91]. In parallel, the AtKC1 subunit, which is expressed in the root cortex, epidermis, and root hairs [44], and interacts with AKT1 to form a functional heterotetrameric channel, prevents K⁺ loss through AKT1 under low-K⁺ conditions [33]. Neither AKT1, HAK5, AtKUP7, nor AtKC1 appear to display strong and durable up- or downregulations at the transcriptional level in response to water stress, heat, or ABA in roots [44,45]. In rice, OsAKT1 also contributes to a significant extent to K⁺ uptake in roots, in the absence of regulation by an AtKC1-like subunit [46]. It improves resistance to drought stress

triggered by polyethylene glycol (PEG) or reduction of soil watering, by increasing K^+ content in roots. Conversely, it has no effect on resistance to salinity [47].

The Kout channel GORK is also expressed in roots [65]. In the case of salt stress, loss of K^+ must be avoided to maintain the K^+/Na^+ ratio. However, Na^+ entry depolarizes the plasma membrane and evokes massive loss of K^+ mainly through GORK [90,92,93]. Non-selective cation channels [93] are also significant players of K^+ efflux. In pea, they assume this function in place of GORK [94]. In *Arabidopsis*, KUP6 and KUP8 (clade II [66]) are also thought to mediate K^+ efflux from roots [68]. KUP8 is downregulated under salt stress, but not the GORK gene [56]. Conversely, GORK is upregulated by heat stress in roots [56]. The reduction of net K^+ influx observed in olive tree roots exposed to heat [95] might, thus, be attributed to an increase in K^+ efflux, but this has to be confirmed. Although the role of GORK-mediated K^+ efflux is much less understood than in guard cells, it is thought to be involved in repolarization of the root-hair plasma membrane, for instance, after elicitor-induced depolarization [96] or in response to different stresses [93]. GORK is activated by ROS (more specifically hydroxide radicals) that are generated following different stresses including salt stress [90]. Depending on stress intensity, GORK-mediated K^+ efflux could either trigger programmed cell death or prevent activation of anabolic enzymes by K^+ , thus releasing energy for adaptation to stress and reparation [93]. Repolarization of the plasma membrane could have a role in action potentials and propagation of stress and hormone electrical signaling [93].

Once taken up by epidermal and cortical cells at the root periphery, K^+ is transported radially (via an apoplastic or symplastic pathway) and then delivered to the xylem. KUP6 and KUP8 are expressed in the pericycle, and they enhance ABA response to inhibit lateral root formation [68]. The SKOR channel drives about 50% of K^+ transported in the xylem sap to the shoot [63]. The SKOR gene transcript is considerably reduced in response to ABA treatment [63], in accordance with the previously observed strong decrease of outward K^+ currents in the maize stele [22]. This would ensure that roots can keep growing under stress, and is also expected to provide a signal for drought stress to the shoots [97], since the decrease in K^+ content in the xylem sap is accompanied by an increase in Ca^{2+} [63] that serves as an intermediate for signaling cascades. Alongside its role in K^+ uptake from roots, KUP7 was identified as a transporter allowing K^+ efflux to the xylem especially under K^+ -deficient conditions (accounting for about one-third of xylem K^+ [91]). Systematic studies on whole plants do not reveal transcriptional regulation by drought, ABA, or heat for the KUP7 gene [56]. KUP7 could, therefore, ensure a minimal transfer to the xylem when SKOR expression is repressed by ABA. In rice, at least two transporters of the KUP family (OsHAK1 and OsHAK5), belonging to the same clade Ia as HAK5, mediate not only K^+ uptake from the soil but also translocation to the xylem, possibly (at least for OsHAK5) by allowing K^+ loading of xylem parenchyma cells prior to K^+ efflux via SKOR-like channels [85,98]. A recent report [69] highlighted the role of OsHAK1 from rice in drought stress tolerance. For its part, OsHAK5 promotes salt resistance by increasing the K^+/Na^+ ratio [70]. OsHAK21 (clade Ia, [66]), mainly expressed in xylem parenchyma cells, is involved in K^+ uptake and control of K^+ homeostasis under salt stress [71].

2.2.2. K^+ Loading and Unloading in the Phloem

Due to the difficulty to access phloem sap and phloem membranes, little is known on the behaviors of phloem K^+ transport systems during adaptation to stress. The first transport system identified in the phloem was the Shaker channel AKT2 (also referred to as AKT3) from *Arabidopsis* [54,99]. KAT1 and KCO6 (*AtTPK3*) transcripts were also detected in isolated phloem companion cell protoplasts [100]. The KAT2 [60], OsHAK1 [98], and OsHAK5 [70] promoters are active (in fusion with the GUS reporter gene) in phloem tissues. Due to its unique gating properties, AKT2 was extensively studied with the aim of understanding its role in the phloem transport of solutes. The AKT2 channel from *Arabidopsis* and its counterparts in other species substantially contribute to the phloem K^+ conductance in both sieve elements and companion cells, where they have a major role in phloem K^+ loading and unloading [52,54,99–104]. Surprisingly, the phloem sap of the *akt2* mutant plant is not depleted in

K⁺. Instead, it contains only half the wild-type content of sucrose [52]. This is expected to result from the effect of a “leak” channel on membrane polarization, preventing membrane depolarization during sucrose uptake by the sucrose/proton symporter. The *AKT2* transcript is increased in leaves in response to ABA, thus possibly favoring K⁺ recirculation to the root under drought stress [54].

2.2.3. K⁺ Fluxes in Guard Cells

Most of the literature on drought stress adaptation deals with stomatal regulation. In shoots, water-saving is tightly controlled by ion-driven guard-cell movements. Due to their importance in plant hydric status preservation and their accessibility to (electro)physiological studies, guard cells became a model for the regulation of ion (notably K⁺) fluxes in response to drought stress and ABA. Also, the molecular mechanisms of light-induced stomatal opening and closure were extensively studied (reviewed by References [105–109]). Comparatively to drought stress-induced stomatal closing and light-induced stomatal opening, little is known about the effect of high temperatures on stomatal movements. Under high temperatures and drought conditions, guard cells have to face the dilemma of saving water while controlling leaf temperature through transpiration [110].

ABA perception by PYR/PYL/RCAR (pyrabactin resistance, pyrabactin resistance-like, regulatory component of ABA receptor) ABA receptors triggers an increase in cytoplasmic calcium (due to release from internal stores and Ca²⁺ uptake, regulated by ROS and nitric oxide [42,111–113]) leading to activation of anion channels (slow- and quick-activating SLAC1 (slow anion channel 1), SLAH3 (SLAC1 homolog), and QUAC1 (quick anion channel 1)) [114] that depolarize the plasma membrane. The plasma membrane depolarization, in turn, triggers the opening of the K⁺ voltage-gated GORK channel that mediates K⁺ efflux [64,115]. This release of ions drives water out of guard cells, which causes guard-cell deflating and stomatal closure [108,109]. In addition to GORK, KUP6 and KUP8 are also involved in the control of stomatal closure. *KUP6* is highly responsive to osmotic stress, especially in shoots [56,68]. *KUP6* over-expressors exhibit a clear drought resistance phenotype, thus highlighting the importance of the *KUP6* gene in stress adaptation, and close their stomata more efficiently than wild-type plants in the presence of ABA [68]. Shaker genes *KAT1/KAT2*, *AKT1*, *GORK*, and, to a much lesser extent, *AKT2* and *AtKC1* are expressed in guard-cell protoplasts [55,96,116]. *KAT1* and *KAT2* encode inward-rectifying-forming subunits that, after tetramerization, form the *KAT1* and *KAT2* channels, which are the key actors of stomatal opening. Stomatal opening mechanism is an inversion of stomatal closure in terms of osmolyte accumulation and direction of fluxes. *KAT1* and *KAT2* mediate K⁺ influx that drives water influx and opens stomata [61]. Upon ABA exposure, *KAT1* and *KAT2* genes are downregulated at the transcriptional level, as expected for genes involved in stomatal opening [55,117]. The *GORK* gene transcription, which is clearly upregulated by ABA in most tissues [65] was unexpectedly found insensitive (or weakly sensitive) to ABA in guard-cell protoplasts [65,73], thus revealing a specific type of regulation for these cells. *AKT1*, previously found to be essential for root K⁺ uptake [118], was only recently identified as a player of stomatal movements. In the *akt1* loss-of-function mutant, the stomatal conductance is lower under drought stress and ABA has a much stronger effect on stomatal closing [43].

Although most data rely on plasma membrane channel regulation, it should be emphasized that vacuoles, which serve as solute reservoirs, play a significant role in stomatal movements. The voltage-independent, K⁺-selective TPK1 channel from *Arabidopsis* is essential for the control of K⁺ homeostasis and vacuolar K⁺ release during ABA-mediated stomatal closure [41,119].

2.2.4. K⁺ Fluxes in Reproductive Organs

Pollen germination on the stigma requires cell elongation maintained by K⁺ uptake through the *Arabidopsis* SPIK channel [120]. Data are also emerging regarding the contribution of different K⁺ transport systems to the mechanisms of response to stress and their consequences on fruit quality. Most of them were obtained on grape berry, considered as a model for fleshy fruits [50]. Whereas K⁺ is often limiting in plants, especially under hot and dry climates, it is sometimes accumulated in

excess. In grape berries, potassium accumulates after the véraison stage that marks the beginning of maturation with change in berry color and firmness, dramatic increase in sugar content, loss of acidity, and development of aromas. This K^+ accumulation increased during the past decades and this was attributed to global warming, since high temperatures favor K^+ accumulation in berries [121]. Although K^+ seems to be beneficial for enhancing berry resistance to drought [122], it can be deleterious for wine quality. Indeed, due to the neutralization of organic acids by K^+ ions, high amounts of K^+ in the must are associated with high pH [122], resulting in wines with loss of aromas and poor aging potential [122,123]. After véraison, due to the loss of xylem functionality for solute transport [124], K^+ enters in the berry mainly via the phloem. The weakly rectifying Shaker channel VvK3.1 which belongs to the AKT2 channel phylogenetic branch, would be involved in the massive K^+ fluxes from the phloem cell cytosol to the berry apoplast during berry K^+ loading [59]. Once delivered to the apoplastic space, K^+ is taken up by flesh and skin cells [122] to be stored in vacuoles. Several transport systems related to these functions were identified. Two Kup transporters are involved in K^+ uptake into the skin cells at pre-véraison stages [125]. VvK1.1, the ortholog of AKT1, is expressed in the phloem and in seeds and is strongly upregulated by water stress and ABA in berries not only before but also after véraison [50]. However, its expression level in berries is far below that of the other AKT1-type gene, VvK1.2, which bursts after véraison and is highly induced by water stress [51]. In light of its expression pattern, VvK1.2 is expected to mediate K^+ retrieval by perivascular and flesh cells from the apoplast.

3. Post-Translational Regulations Linked to Osmotic, Salt, or Water Stress and Heat

3.1. Regulation by Kinases, Phosphatases, and Associated Proteins

Three main kinase families share the regulation of K^+ membrane transport system activity: the SnRK2 (SNF1-related protein kinase 2) family, represented notably by OST1/SnRK2.6/SnRK2E [126], the SnRK3 family (SNF1-related protein kinase 3 or CIPK; calcineurin B-like-interacting protein kinase [127]), and the CDPK/CPK family (calcium-dependent protein kinase) [128] (Table 2). All are involved in signaling pathways related to different stresses such as K^+ deficiency, drought, osmotic, and salt stresses [14,129,130]. The *Arabidopsis* genome encodes 10 SnRK2s, of which OST1/SnRK2.6 is a positive regulator of ABA-dependent stomatal movements [126]. The SnRK2 family is involved in ABA-dependent, Ca^{2+} -independent signaling, whereas CIPK and most CPK kinases depend on calcium for their activity [127,131]. CIPKs are activated by interaction with calcineurin B-like (CBL) proteins that bind calcium, which is released in cells in response to different stimuli. They are composed of a kinase domain, an auto-inhibitory (NAF/FISL) domain, and a phosphatase-binding (protein phosphatase interaction, PPI) domain. The calcium-dependent CBL interaction with the NAF domain of the CIPK suppresses the auto-inhibition [132]. CBLs also require to be phosphorylated by their CIPKs to be fully active [133]. There are 26 CIPKs and 10 CBLs in *Arabidopsis thaliana*, which are combined to regulate ion homeostasis and response to ABA in a highly specific manner [134]. In contrast to CIPKs, CPKs (34 members in *A. thaliana*) contain an intrinsic C-terminal calmodulin-like domain comprising EF-hand motives that directly bind calcium. When calcium levels are low, kinases are auto-inhibited, but calcium elevation triggers binding to EF-hands and conformational changes that relieve the auto-inhibition [131]. Calcium sensitivity appears to be highly variable in the CPK family, since some CPKs such as CPK13 and CPK23 seem to be insensitive or much less sensitive to calcium than the others [131]. It is important to notice that members of these SnRK2, CIPK, and CPK families interact with clade A protein phosphatases 2C, known to be involved in ABA signaling [78,135–141]. These phosphatases are trapped and inactivated by PYR/PYL ABA receptors in the presence of ABA. This interaction releases kinases that are bound to and inactivated by protein phosphatases 2C (PP2Cs), such as OST1 (open stomata 1) [136,142–145]. A similar mechanism seems to be effective for some CPKs [78,139,141], and the effects of CIPKs are also regulated by PP2Cs [140]. Some of these kinase/phosphatase pairs play critical roles in K^+ channel regulation, as described below.

Table 2. Regulatory partners of plant K⁺ transport systems, their targets among K⁺ and mineral anion transport systems, and their regulations in response to ionic and osmotic stresses.

Gene Name ¹	ID	Protein Family	Plasma Membrane Targets	Type of Stress	Transcriptional Regulation (Drought, ABA, Heat Stress, NaCl)	Protein Level/Activity
<i>OST1/SnRK2.6/SRK2E</i>	At4g33950	SnRK2 kinases	KAT1 [146,147], SLAC1 [136,144], KUP6 [68]	ABA	No change in transcript level in whole plants [126], induction in guard cells [55]	Induction of kinase activity in roots and guard cell protoplasts [126]
				Low air humidity		Induction of kinase activity in leaves [148]
				NaCl, sorbitol		Induction of kinase activity in roots [148]
				NaCl	Slow induction [56]	
				Drought	Moderate induction [149]	
<i>CIPK6</i>	At4g30960	SnRK3 kinases	AKT1 [135,140], AKT2 [150]	ABA, mannitol	Induction in seedlings (including roots) [151]	
				NaCl	Induction in shoots [56] and in seedlings (including roots) [151]	
<i>CIPK16</i>	At2g25090	SnRK3 kinases	AKT1 [135]	NaCl	Induction in root stele [152]	
<i>CIPK23</i>	At1g30270	SnRK3 kinases	AKT1 [135,153,154], HAK5 [155], SLAC1 [156], CHL1 [157]	ABA, drought, mannitol, NaCl	No change [45]	
<i>CBL1</i>	At4g17615	CBL	AKT1 [158]	Drought	Short air treatment: transient induction in roots only [56]; strong and transient induction after 1-3h [159,160]; long-term induction in soil [149]	
				ABA	No significant change in mesophyll and guard cells [73] and in seedlings [160,161]	Protein accumulation in leaves [162]
				NaCl	Transient induction in roots [163]	Protein accumulation in leaves [162]
<i>CBL4/SOS3</i>	At5g24270	CBL	AKT1 [158]	NaCl, mannitol	Induction in roots [56], no significant change in seedlings [164]	
				ABA	Down-regulation in guard cells [73]	
<i>CBL9</i>	At5g47100	CBL	AKT1 [158]	ABA, NaCl, mannitol, drought	No significant change [45]	
<i>CBL10</i>	At4g33000	CBL	AKT1 [165]	Mannitol	Down-regulation in shoots [56]	
				NaCl	Transient and moderate induction [72]	
<i>CPK3/CDPK6</i>	At4g23650	CPK/CDPK	KAT1/KAT2, GORK [163], TPK1 [72]	NaCl	No change in seedlings [72]	
				ABA	Slight down-regulation in guard cells [73,76]	
<i>CPK6/CDPK3</i>	At2g17290	CPK/CDPK	GORK [163], SLAC1 [139,166-168]	ABA, mannitol, NaCl	No change [45,73]	
				NaCl, PEG	Transient induction [169]	
<i>CPK13</i>	At3g51850	CPK/CDPK	KAT1/KAT2 [170], GORK [163]	NaCl, drought	No significant change [56]	
				Mannitol	Downregulation [56]	
				ABA	No change in guard cells [73,76]	

Table 2. Cont.

Gene Name ¹	ID	Protein Family	Plasma Membrane Targets	Type of Stress	Transcriptional Regulation (Drought, ABA, Heat Stress, NaCl)	Protein Level/Activity
CPK21	At4g04720	CPK/CDPK	GORK [171], SLAC1 [139], SLAH3 [78]	ABA, drought, mannitol, NaCl	No significant change, except perhaps a slight down-regulation in response to ABA in guard cells [45]	Enhancement of kinase activity [172]
				Mannitol		
CPK33	At1g50700	CPK/CDPK	GORK [163], slow anion channels? [173]	ABA, drought	Moderate up-regulation [173]	
ABI1	At4g26080	Clade A PP2C phosphatases	SLAC1 [167]	ABA	Induction in guard cells [55,73,76] and seedlings [45]	
				NaCl, mannitol		
				Heat		
ABI2	At5g57050	Clade A PP2Cs	GORK [174]	ABA	Induction in guard cells [55,73,76] and seedlings [45]	
				NaCl, mannitol		
AtPP2CA	At3g11410	Clade A PP2Cs	AKT2 [175,176], GORK [174], SLAC1 [136]	ABA	Strong induction in guard cells [55,73,76] and whole plants [56,176–178]	
				NaCl, mannitol		
				Heat		
				ABA		
AIP1	At1g07430	Clade A PP2Cs	AKT1 [135,140]	NaCl, mannitol	Strong induction in roots (NaCl) or shoots (mannitol) after 3 hours [56]	
				Heat		
				NaCl		
NtSYR1	LOC107768839	Syntaxins	Nicotiana K and Cl channels [179]	NaCl	Induction of transcripts in leaves [180]	Protein accumulation in leaves [180]
				ABA		
SYP121	At3g11820	Syntaxins	AtKC1, AKT1-AtKC1 [181], KAT1 [182]	NaCl	Induction in roots, not in leaves [56]	
				ABA		
VAMP721	At1g04750	Synaptobrevin-like	KAT1, AtKC1 [183]	ABA, NaCl, drought, mannitol	No significant change [45]	
VAMP722	At2g33120	Synaptobrevin-like	KAT1, AtKC1 [183]	ABA, NaCl, drought, mannitol	No significant change [45]	
GRF6	At2g06200	14-3-3	TPK1 [184]		Low expression	

¹ Gene names: OST: open stomata, CIPK: CBL-interacting protein kinase, SnRK/SRK: SNF1-related protein kinase, CBL: calcineurin B-like, SOS: salt overly sensitive, CPK or CDPK: calcium dependent protein kinase, ABI: ABA-insensitive, PP2C: protein phosphatase 2C, AIP: AKT1-interacting protein, SYR: syntaxin-related protein, SYP: syntaxin-related protein, VAMP: vesicle-associated membrane protein. All genes originate from *Arabidopsis thaliana* except *NtSYR1* (from *Nicotiana tabacum*).

3.1.1. Regulation of K⁺ Uptake and Release at the Root/Soil Interface

The first kinase to be clearly identified as a partner of a K⁺ transport system was CIPK23, interacting with the AKT1 channel in *Arabidopsis thaliana* [153,154]. Mutants in the *CIPK23* gene displayed reduced K⁺ uptake and leaf chlorosis on low-K⁺ medium [153]. CIPK23 also appears to be involved in drought resistance [185]. To activate AKT1, CIPK23 interacts with CBL1 and CBL9. Whereas *CIPK23* transcripts do not appear to display marked fluctuations in response to stress or ABA [56], with the exception of K⁺ deficiency [153], it is noticeable that the *CBL1* gene can be transiently but strongly induced by drought [159,160], and that the protein level increases in response to ABA, mannitol, and NaCl [162]. Root-specific transcript accumulation is observed in response to salt [56]. The *CBL9* gene is highly expressed in radicles of germinated seedlings; however, afterward, its expression in roots is restricted to growing zones [186]. CIPK23 phosphorylates AKT1 [153,154] and it interacts via its kinase domain with the ankyrin domain of the channel [135]. Conversely, the regulatory subunit AtKC1, closely associated with AKT1, does not interact with CIPK23 [153]. AKT1 also directly interacts with CIPK6 [135,140], whose gene expression is induced by NaCl, ABA, and mannitol, especially in roots [151], and with CIPK16 [135,140]. The CIPK6/CBL1 couple activates AKT1 in *Xenopus* oocytes in a similar way as CIPK23/CBL1 [140]. Attempts to transpose the CIPK23/CBL1 regulation to species of agronomical interest led to the characterization of the OsCIPK23/OsCBL1 complex from rice [187], which accounts for a significant part of K⁺ uptake [46], and TaCIPK23/TaCBL1 from wheat, both involved in drought tolerance [187,188]. Transcriptional responses to water-related abiotic stresses can differ between *AtCIPK23* (which seems to be insensitive to these stresses in roots and shoots) and its counterparts in other species. In whole rice seedlings, *OsCIPK23* is induced by ABA [187], and, in wheat mesophyll protoplasts, *TaCIPK23* is induced by drought, salt, and ABA [188]. It would be informative to get root-specific data. In soybean, *GmCIPK20* (closest homolog of *AtCIPK23*) is induced by drought stress in leaves and stems but not in roots [189]. Homologs of *CIPK6* were also identified in other species. *BnCIPK6* and *BnCBL1* genes (from *Brassica napus*) are both upregulated by NaCl, mannitol, and ABA in roots, in accordance with the increased salt tolerance of transgenic *Arabidopsis* plants overexpressing these genes [190].

Interestingly, HAK5 is the target of the same CIPK23/CBL1 complex as AKT1. CIPK23 phosphorylates HAK5, resulting in activation of the transporter, required for K⁺ uptake in low-K medium [155]. This regulation is conserved among plant species, with tomato and pepper HAK5 counterparts being activated by the *Arabidopsis* CIPK23/CBL1 complex in yeast [155]. Figure 1 summarizes the regulation of K⁺ uptake by CIPK/CBL complexes at the root/soil interface.

Recently, CPK21 was identified as a kinase activated by 14-3-3 proteins, phosphorylating the GORK C-terminal cytoplasmic domain. Following a sudden salt stress, a transient K⁺ efflux is mediated by GORK at the plasma membrane. This efflux is reduced in 14-3-3 mutants. Indeed, salt stress is expected to trigger membrane depolarization and a rise in cytosolic calcium that activates CPKs together with 14-3-3s, to promote GORK phosphorylation leading to its activation [171].

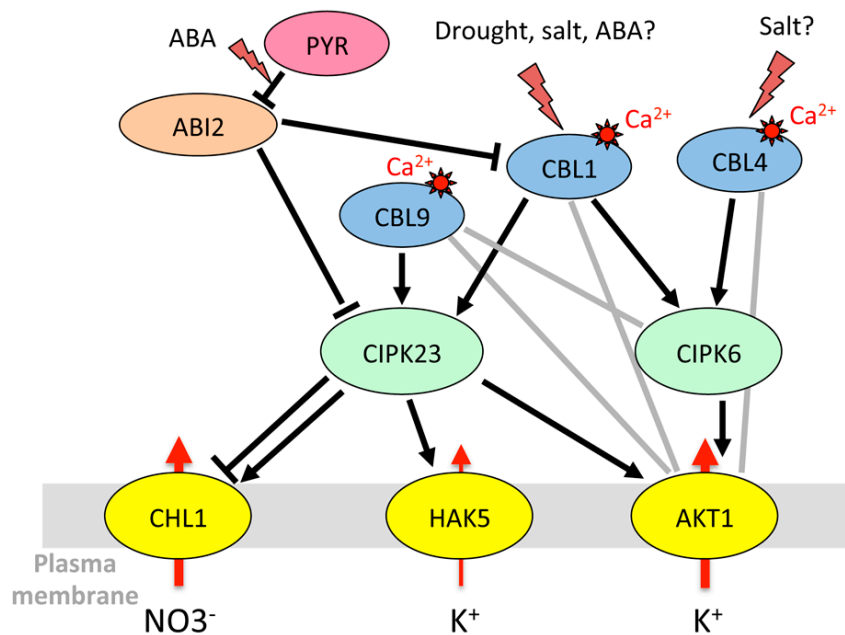


Figure 1. Model for cross-talks between CIPK (CBL-interacting protein kinase)/CBL (Calcineurin B-like protein) regulations of K^+ and NO_3^- uptake by *Arabidopsis* roots, obtained by compilations of expression and interaction data. Positive and negative regulations are depicted in black (arrows and T-bars, respectively). Indications for physical interactions (two-hybrid, bimolecular fluorescence complementation (BiFC)) in the absence of further evidence for functional effect are represented by a gray line. Red arrows symbolize the ion fluxes. The lightning symbol is used when a stress or abscisic acid (ABA) results in a change in protein amount, protein activity or protein-protein interaction. The symbol “?” stresses that the effects of the treatments on CBL1 protein accumulation [162] were only addressed in whole leaves. Also, myristoylation of CBL4 is required for its function in salt resistance [191], but was not demonstrated to occur in response to salt. References: “drought, salt, ABA?” to CBL1 [160,162]; “salt” to CBL4 [191]; ABI2 to CIPK23 and CBL4 [192]; CBL1 to CIPK6 [140,150], CBL1 and CBL9 to CIPK23 [153,154,185]; CBL4 and CBL9 to CIPK6 [150]; CIPK6 to AKT1 [135,140]; CIPK23 to AKT1 [153,154]; CBL1, CBL4 and CBL9 to AKT1 [193]; CIPK23 to HAK5 [155]; CIPK23 to CHL1 [157]. The phosphatase AtPP2CA, mainly expressed in phloem tissues [176], and not characterized for its role in root ion uptake, was not included in this figure.

3.1.2. AKT2 and Its Orthologs: Regulation of Phloem K^+ Potential

From the beginning of its characterization, AKT2 received most of the attention due its unique gating properties, which allow it to combine a time-dependent voltage-activated current and an instantaneous ohmic current with the ability to switch between these two gating modes [31]. The mechanism via which AKT2 becomes a non-rectifying channel was deciphered progressively. The AtPP2CA protein phosphatase, which belongs to the clade A of PP2Cs involved in ABA signaling, interacts with the AKT2 channel [175,176] (Figure 2).

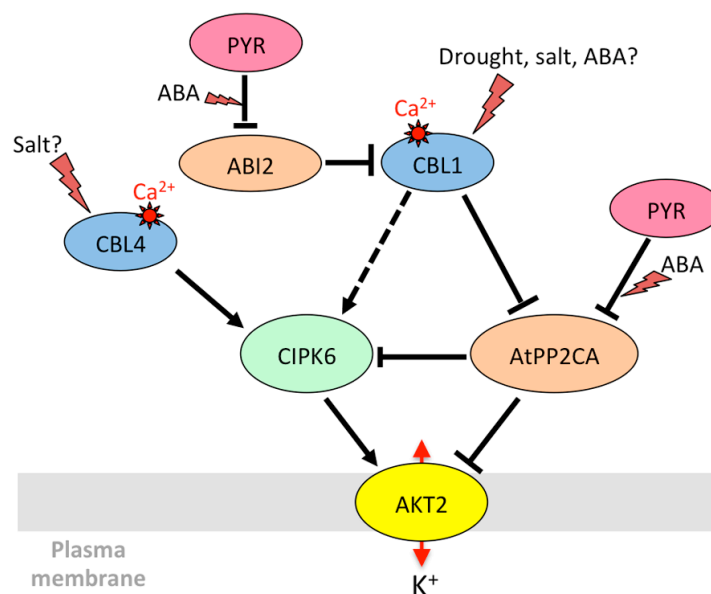


Figure 2. Model for the regulation of K^+ transport through AKT2 (Arabidopsis K^+ transporter 2) by CIPK6 and associated proteins in *A. thaliana*. See Figure 1 for details of the legend. The dotted arrow indicates that CBL1, though activating CIPK6 for AKT1 opening [131], has no functional effect on AKT2/CIPK6 [189]. References: “Drought, salt, ABA?” to CBL1 [160,162]; “Salt?” to CBL4 [191]; ABI2 to CBL1 [192]; CBL1 to CIPK6 and AtPP2CA, AtPP2CA to CIPK6 [140]; CBL4 to CIPK6, CIPK6 to AKT2 [150]; AtPP2CA to AKT2 [176].

AtPP2CA is a negative regulator of ABA signaling [177,178] selectively interacting with some PYR/PYL/RCAR receptors [194]. Interestingly, the phosphatase changed the rectifying properties of the channel by preferentially inhibiting the leak component [176]. Later on, Michard et al. [195] revealed that phosphorylation is essential to the functioning of AKT2, by activating silenced AKT2 channels and promoting the switch to the non-rectifying, leaky mode. To date, the kinase(s) that phosphorylate AKT2 remain unidentified. Instead, the CIPK6/CBL4 couple was found to strongly activate AKT2 in a calcium-dependent but phosphorylation-independent manner, by promoting the targeting of the AKT2/CIPK/CBL complex to the plasma membrane through myristoylation of CBL4 [150]. Plants overexpressing CIPK6 are more sensitive to ABA and more tolerant to salt stress [151]. CBL4, also known as SOS3 (salt overly sensitive 3), is essential for salt resistance and potassium homeostasis [196]. Thus, regulation of AKT2 by CIPK6 and CBL4 could take part in adaptation to salt stress. A receptor-like pseudokinase, MRH1 (morphology of root hair 1), also interacts with AKT2, but has no direct functional effect. It is proposed to be involved in the recruitment of other, still unknown, partners [197].

3.1.3. Stomatal Aperture Control in Case of Abiotic Stress

During ABA-mediated stomatal closure, mechanisms leading to blue-light-induced stomatal opening are inhibited. This includes inactivation of H^+ -ATPase and Kin channels (see Reference [198] and references therein). In *A. thaliana* guard cells, the KAT1 and KAT2 channels and, probably to a lesser extent, the AKT1 channel are the major actors of K^+ influx. Early studies revealed that the KAT1 channel was phosphorylated in vitro by 57-kD calcium-dependent and 48-kD ABA-regulated (ABR) protein kinases from *Vicia faba* [199,200]. The ABR kinase shared characteristics with a kinase named AAPK (ABA-activated protein kinase) [201], strongly suggesting that AAPK and ABR kinase are the same protein [200]. AAPK is a serine/threonine kinase specifically activated (autophosphorylated) by ABA, involved in stomatal closure [201]. Later on, the kinase OST1/SnRK2.6/SRK2E from *A. thaliana*, homologous to AAPK, was identified as a key player of ABA-mediated stomatal closure, acting upstream of ROS production [126]. The ABA-mediated inhibition of Kin channels was found reduced

in the *ost1* mutants, suggesting that OST1 inhibits KAT1 [146]. OST1 phosphorylates the C-terminal cytosolic region of KAT1 [147], and direct interaction between KAT1 and OST1 can be visualized using bimolecular fluorescence complementation (BiFC) [146] (Figure 3). However, it was recently reported that KAT1 expressed in *Xenopus* oocytes is not inactivated by OST1 [62]. The mechanism of KAT1 inactivation by OST1 observed in guard cells and the effect of KAT1 phosphorylation on its K⁺ transport activity, thus, await further investigation.

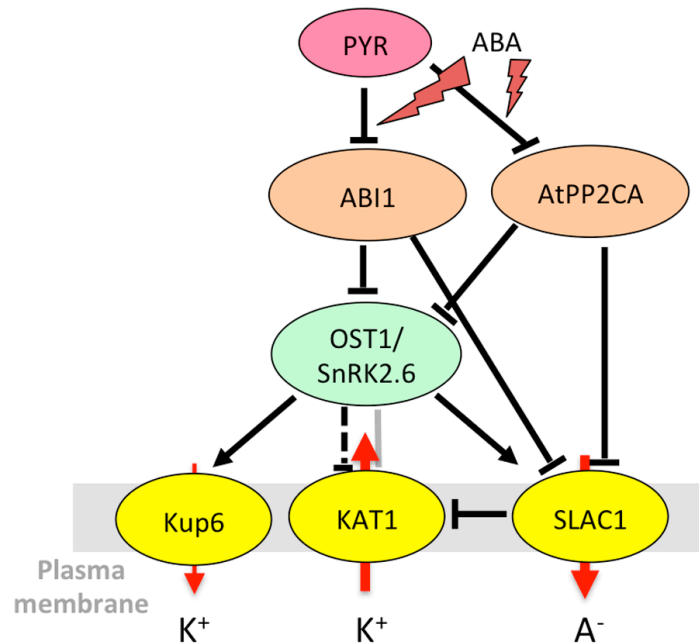


Figure 3. Model for the OST1 network leading to K⁺ and anion fluxes in *Arabidopsis* guard cells. The dotted T-bar indicates that the effect might be indirect. See Figure 1 for details of the legend. References: ABI1 to OST1 [144,148]; AtPP2CA to OST1 and SLAC1 [136]; ABI1 to SLAC1 [167]; OST1 to SLAC1 [136,144]; OST1 to KAT1 (in planta physiological effect) [146]; OST1 to KAT1 (interaction) [146,147]; OST1 to KUP6 [68]; SLAC1 to KAT1 [62].

Regarding calcium-dependent protein kinases (CPKs), CPK3, 4, 6, 7, 11, 13, and 33 were pinpointed as being expressed in guard cells [163]. A systematic functional screening performed in *Xenopus* oocytes led to the identification of a few of them that activate or inhibit Shaker Kin currents [163] (Figure 4). CPK13, which belongs to the small subgroup of CPKs insensitive to calcium ions, inhibits KAT2. It impairs light-induced stomatal opening but has no effect on ABA-induced stomatal closure [170], despite its inhibitory effect on GORK [163]. CPK21, which phosphorylates GORK [171], is also present in guard cells [73].

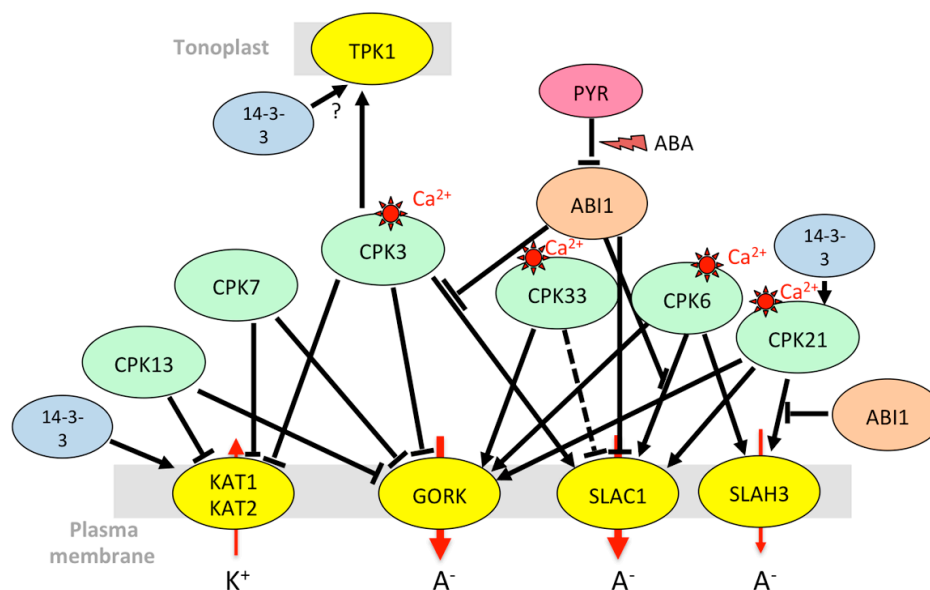


Figure 4. Model for integrated regulation of Shaker and anion channels by CPKs (calcium-dependent protein kinases) in *Arabidopsis* guard cells. The dotted T-bar indicates that the effect might be indirect, and the symbol “?” that the activation of TPK1 by 14-3-3 proteins was not studied in guard cells. See Figure 1 for details of the legend. References: 14-3-3 to CPK21 [171]; 14-3-3 to TPK1 [184], 14-3-3 to KAT1/KAT2 [202], ABI1 to CPK3-SLAC1 and CPK6-SLAC1 [141]; ABI1 to CPK21-SLAH3 [78]; ABI1 to SLAC1 [167]; CPK3 to TPK1 [72], CPK13 to KAT1/KAT2 [163,170], CPK13 to GORK [163]; CPK3 and CPK7 to KAT1/KAT2 and GORK [163]; CPK6 and CPK33 to GORK [163]; CPK21 to GORK [171]; CPK3 to SLAC1 [168]; CPK6 to SLAC1 [139,167,168]; CPK33 to SLAC1 [173]; CPK6 to SLAH3 [141]; CPK21 to SLAC1 [139]; CPK21 to SLAH3 [78]; CPK33 to SLAC1 [173];

AKT1 is activated by the CIPK23/CBL1 complex in guard cells as it is in roots [43] (Figure 5). It is thought to take part in the control of stomatal aperture under drought stress (but not in well-watered conditions), by limiting the extent of ABA-dependent stomatal closure. Since *akt1* plants are more resistant to drought, the exact role of AKT1 in guard cells (slowing stomatal closure for leaf cooling or maintaining photosynthesis?) remains to be clarified. Based on mutant phenotypes, it appears that its activating partners CIPK23 and CBL1/CBL9 also counteract stomatal closure in the presence of ABA and increase sensitivity to drought [185]. AKT1 also interacts with AIP1/HAI2 (AKT1-interacting protein phosphatase 1/highly ABA-induced 2) [51], an ABA-inducible clade A PP2C present in guard cells but almost absent in root epidermis and cortex [73], which itself binds CIPK23 [135,140]. AIP1, previously thought to be a positive regulator, is rather a negative regulator of the ABA signaling pathway like other clade A PP2Cs [203], interacting specifically with some PYR/PYL/RCAR receptors [204]. Together with CIPK23, the CIPK6 gene, encoding another activating partner of AKT1 [140], is highly expressed in guard cells [73]. Its overexpression confers tolerance to salt stress and hypersensitivity to ABA [151]. The CIPK23 and CIPK6 kinases, thus, might have different roles in stomatal regulation. Unlike CIPK23, CIPK6 interacts with AtPP2CA, and AtPP2CA itself interacts with many CBLs including CBL1 [140]. This study also revealed the existence of many other possible channel/CIPK/CBL/PP2C complexes. A model is proposed in which AKT1, activated by a CIPK/CBL complex, is deactivated by a PP2C interacting with the kinase. Other CBLs that specifically interact with the PP2C reactivate the complex, promoting K⁺ influx [140]. It should also be noted that CBLs can directly interact with the channel and modulate its activity independent of CIPKs [59,165,193]. To our knowledge, the role of CIPK6 in guard cell movements is yet to be studied.

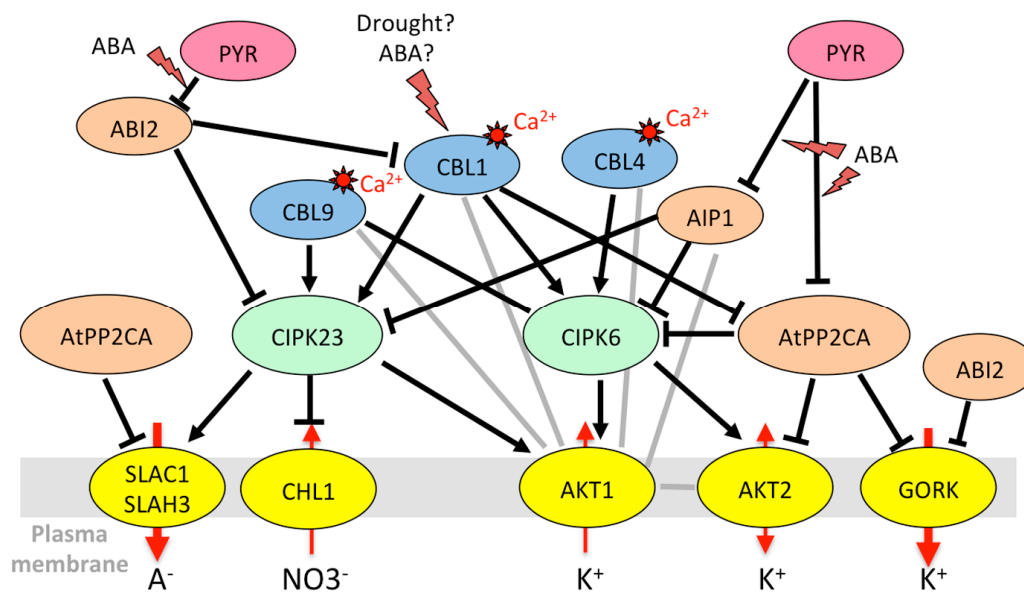


Figure 5. Model for the CIPK/CBL/PP2C network regulating both Shaker and anion channels in *Arabidopsis* guard cells. See Figure 1 for details of the legend. The symbol “?” indicates that the CBL1 protein, involved in stomatal movements together with CBL9 [185], is accumulated in whole leaf tissues after drought and ABA treatments [162], but this remains to be confirmed specifically in guard cells. References: “Drought, ABA?” to CBL1 [162,185]; ABI2 to CIPK23 and CBL1 [192]; AIP1 to CIPK6 and CIPK23 [140]; AtPP2CA to CIPK6 [140]; CBL1 and CBL9 to CIPK23 [153,154,185]; CBL1 to CIPK6 [140,150]; CBL4 and CBL9 to CIPK6 [150]; CBL1 to AtPP2CA [140]; CIPK6 to AKT1 [135,140]; CIPK23 to AKT1 [153,154]; AIP1 to AKT1 [135]; CIPK6 to AKT2 [150]; CBL1, CBL4 and CBL9 to AKT1 [193]; AtPP2CA to AKT2 [176]; AtPP2CA to GORK [174]; ABI2 to GORK [174]; AtPP2CA to SLAC1/SLAH3 [136]; CIPK23 to SLAC1/SLAH3 [156]; CIPK23 to CHL1 [157]; formation of AKT1-AKT2 heterotetramers [44].

As the main channel mediating K^{out} currents at the plasma membrane of guard cells, GORK is expected to be a target of regulatory mechanisms to control stomatal closure including the closing speed. However, its regulatory network is much less characterized than that of AKT1 or KAT1. Unlike other cell types, guard cells do not induce *GORK* transcripts in response to ABA [65,73] and, therefore, need to control the channel activity via specific post-translational mechanisms. GORK activity in *Xenopus* oocytes is inhibited by CPK3, CPK7, and CPK13, and it is activated by CPK6 and CPK33 [163]. Accordingly, the *cpk33* mutants do not close their stomata in the presence of calcium. However, they are also more resistant to drought and their stomata close more efficiently in the presence of ABA [173]. GORK activity is partially inhibited by AtPP2CA [174], thus providing a fine-tuning regulation during ABA-evoked stomatal closure. Finally, the ability of GORK to change its spatial distribution at the guard-cell plasma membrane, at least in response to high K⁺, suggests the existence of additional post-translational regulatory mechanisms [205]. KUP6, also mediating K⁺ efflux from guard cells, is phosphorylated by OST1 [68], while this kinase has no effect on GORK activity in *Xenopus* oocytes [174].

Although much less studied than K⁺ fluxes at the plasma membrane, K⁺ loading and unloading of vacuoles play a central role in the control of guard-cell turgor. The TPK1 tonoplast channel seems to be the main K⁺-selective channel involved in that process. It is phosphorylated by CPK3 that is highly expressed in guard cells [73] and responds to calcium elevation in the cytosol [72]. In response to ABA, TPK1 is also phosphorylated by a receptor-like kinase, named KIN7 (kinase 7), resulting in channel activation and stomatal closure [119].

3.1.4. Regulation of K⁺ Transport in Grape Berry by CIPK/CBL Complexes and SnRK2s

The AKT1-related VvK1.1, VvK1.2, and AKT2-related VvK3.1 genes are strongly upregulated by drought stress (VvK1.1 also by ABA treatment), and these channels are activated by CIPK/CBL couples [50,51,59]. Grapevine CIPK and CBL genes display differential responses to various abiotic stresses [206]. The *VvCIPK04* gene, encoding a kinase strongly activating VvK1.2, is upregulated by drought stress in the ripening berry [51]. In the ABA signaling cascade, SnRK2 kinases and clade A PP2Cs, which are likely to modulate the activities of Shaker K⁺ channels, interact with each other in a specific manner and corresponding genes are differentially regulated [207]. Among the kinases, OST1 counterparts SnRK2.1 and SnRK2.4 display a positive signal in two-hybrid tests with the Shaker channel VvK2.1, the ortholog of KAT1 [207]. In the grape berry, ABA induces berry ripening [208] and ABA content increases under heat [209,210]. It is, therefore, likely that the increase in K⁺ berry content correlated to temperature increases can be attributed at least in part to Shaker channels in relation to SnRK2 and CIPK kinases.

3.1.5. Vacuolar K⁺ Uptake and Release in Response to Salt Stress

As highlighted above, maintaining K⁺ homeostasis is crucial. In the case of salt stress, Na⁺ is accumulated in vacuoles, and K⁺ is released from the vacuole to maintain the cytosolic K/Na ratio. For efficient sequestration of Na⁺ in the vacuoles, slow and fast vacuolar currents allowing Na⁺ leakage from the vacuole to the cytosol must be inactivated [211]. Under salt stress, they are suppressed by polyamines and an increase in vacuolar calcium. Conversely, TPK1 remains active. It mediates K⁺ flux back to the cytosol to counterbalance Na⁺ entry into the vacuole and compensates for K⁺ efflux through GORK at the plasma membrane [211]. In poplar, overexpression of *PeTPK1* results in improvement of salt tolerance [212]. Leaf extracts from *Arabidopsis* salt-stressed plants, but not those of control plants, in vitro phosphorylate the TPK1 N-terminus [72]. Looking for kinases that would phosphorylate TPK1, Latz et al. [72] identified several CPKs including CPK29, induced by salt stress at the transcriptional level, and CPK3, known to be involved in stress responses. CPK3 responds to cytosolic calcium elevation triggered by salt stress, and phosphorylates a serine residue in the N-terminus of TPK1 [72] (Figure 4). Phosphorylation at this site enables GRF6 (growth-regulating factor 6), a 14-3-3 protein, to bind to TPK1 and activate the channel [184].

3.2. Regulation by Vesicle-Associated Proteins

As discussed above for AKT2, an increase in channel activity at the plasma membrane can result from a change in channel gating properties (e.g., due to phosphorylation) or from a regulation of membrane targeting. Screening for guard-cell tobacco RNAs able to induce ABA-dependent Cl⁻ efflux in *Xenopus* oocytes, Leyman et al. [179] isolated Nt-SYR1 (*Nicotiana tabacum* syntaxin-related protein), a syntaxin (Qa SNARE, soluble N-ethylmaleimide sensitive factor attachment receptor) associated with the plasma membrane. This protein accumulates in leaves after ABA treatment [180]. A cleavage fragment of this syntaxin (Sp2) not only blocked ABA-dependent regulation of chloride channels, but also that of inward and outward K⁺ channels [179]. Visualization of KAT1 fused to GFP revealed that SP2 and SYP121 (*Arabidopsis* homolog of Nt-SYR1) modify KAT1 distribution and mobility at the plasma membrane [213]. During stomatal closure (or after ABA treatment [214]), KAT1 is internalized, and SYP121 favors KAT1 recycling to the plasma membrane for stomatal reopening [182]. SYP121 also binds to AtKC1, and this interaction leads to the activation of the AKT1/AtKC1 complex [181]. The binding site of SYP121 to K⁺ channels is the voltage sensor, and binding is favored by hyperpolarization, thereby allowing coordination of vesicle trafficking for cell volume adjustment and K⁺ channel voltage-dependent activation [158]. Other SNAREs associated with trafficking vesicles, which belong to the VAMP (vesicle-associated membrane protein) family (R-SNAREs), also interact with KAT1 and AtKC1. Contrary to SYP121, VAMP721 suppresses the KAT1

and AKT1/AtKC1 currents in *Xenopus* oocytes. Altogether, SYP121 and VAMP have complementary actions on K⁺ channel activities [183] (Table 2).

4. Cross-Talks with Anion Transport

As the major cation in plant cells, K⁺ has to be neutralized by organic acids and mineral anions. To maintain electro-neutrality, membrane K⁺ transport is accompanied by the transport of anions. In guard cells, efflux of K⁺ is driven by an efflux of anions. From the beginning of the characterization of channel regulatory proteins, it is striking that elements of the calcium and ABA signaling pathways (PP2C and their associated kinases) can be involved in the regulation of both K⁺ and mineral anion transport. The main mineral anions in plants are Cl⁻ and NO₃⁻. Anion channels of the SLAC1/SLAH (slow anion channel 1/ SLAC1 homolog) family (five members in *Arabidopsis*) mediate voltage-independent slow and massive chloride and nitrate fluxes at the plasma membrane, for root anion uptake, transfer of anions to the shoot, and stomatal closure [215]. Recent findings indicate that the anion channels SLAC1 and SLAH3 interact with Shaker channels. KAT1 (but not its closest homolog KAT2) is inhibited by both SLAC1 and SLAH3 via direct physical interaction [62].

In guard cells, SLAC1 and SLAH3 mediate nitrate and chloride transport. SLAC1 was found inhibited by AtPP2CA [136], like AKT2 [176] and GORK [174]. This mechanism would permit a fine coordination of K⁺ and anion effluxes and modulation of the speed of ABA-triggered stomatal closure. SLAC1 interacts with and is activated by the OST1 kinase, whose activity is inhibited by AtPP2CA and its close relative ABI1 (ABA-insensitive 1) in an ABA-dependent manner [136,144].

Like OST1, kinases of the CPK family, activated by calcium, also interact with SLAC1 and SLAH3 to modulate anion efflux in guard cells. CPK21 and CPK23 both activate SLAC1 and SLAH3 in an ABA- and ABI1-dependent manner, but with contrasting effects on channel currents and calcium sensitivity [78,139,141]. In the *cpk3 cpk6* double knock-out mutant, Ca²⁺- and ABA-activated S-type anion channel currents are abolished, and stomatal closure is partially impaired [166]. Accordingly, CPK6 was found to activate SLAC1 in *Xenopus* oocytes and to phosphorylate the SLAC1 N-terminus at a much higher speed than CPK23 and OST1 [167]. CPK21 [139] and CPK3 [141] also activate SLAC1 in oocytes, but only in their truncated, constitutively active form, suggesting that they are weakly sensitive to calcium and operate only in response to an increase in cytosolic calcium. Contrary to CPK3, CPK6, CPK21 and CPK23, which activate SLAC1 and SLAH3, CPK33 counteracts ABA-dependent slow anion channel activation in guard-cell protoplasts [173]. Finally, it is interesting to note that the CIPK23/CBL1 complex, already known to regulate the K⁺ channel AKT1 and the high-affinity transporter HAK5, also interacts with and activates SLAC1 and SLAH3. Like CPK23, it phosphorylates the N-terminus of SLAC1 in a calcium-dependent manner, but there exist specificities for target phosphorylation sites [156].

In the nitrate transporter family, CHL1 (NRT1.1/NPF6.3) (chlorate-resistant 1, nitrate transporter 1, NRT1/PTR family 6.3), involved in root NO₃⁻ uptake [80], is also targeted by the same kinases and phosphatases. Under low-nitrate conditions, CIPK23/CBL9 phosphorylates CHL1, and converts it into a high-affinity nitrate transporter. However, when nitrate level is high, the transporter returns to the low-affinity state and the CIPK/CBL complex inhibits nitrate transport through CHL1 [157]. CBL1 also associates with CIPK23 to decrease NO₃⁻ uptake under high external nitrate in *Xenopus* oocytes [192]. This inhibitory effect is reversed by the clade A PP2Cs ABI1 and ABI2 [192]. In addition to its role in NO₃⁻ uptake, CHL1 is also involved in stomatal opening and plant drought susceptibility [81]. CIPK23 and CBL1 are both expressed in guard cells [73]. Since the CBL1 protein level is enhanced in response to ABA or osmotic stress (Table 2), the inhibition of CHL1 by the CIPK23/CBL1 complex could be a means to control anion influx that would counteract stomatal closure.

Clade A PP2C phosphatases are at the center of ABA signaling pathways. Their incorporation into kinase/channel complexes most often results in complete abolition of the effect of the kinases. Indeed, AtPP2CA, ABI1, and/or ABI2 drastically inhibit the effect of OST1 [136,144,167], CPK23 [139], CPK6 [167], CPK21 [78], and CIPK23 [156] on SLAC1 or SLAH3 in *Xenopus* oocytes. AtPP2CA [136,138]

and ABI1 [138,144] directly inactivate OST1, but not CPK6 [168], despite the existence of an ABI1–CPK6 interaction [141]. In parallel, AtPP2CA directly inhibits AKT2 [176], GORK [174], and SLAC1 [136] activities, and ABI1 inhibits SLAC1 [167]. ABI1 and ABI2 also suppress the inhibitory effect of CIPK23/CBL1 on CHL1-mediated nitrate uptake [192]. Thus, PP2C phosphatases act at several levels on channel activities: inhibition of upstream kinases [78,136,144] and/or direct inactivation of channels [136,174] via binding [174] or dephosphorylation [167,168].

In summary, common regulatory pathways can control the transport of cations and anions through diverse transport systems to maintain ion homeostasis. Kinases play a central role in these regulations. For instance, OST1 causes stomatal closure both by leading to the inactivation of Kin channels [146] and by activating anion efflux through SLAC1 [33,136]. It also phosphorylates AtRBOHF (respiratory burst oxidase homolog plasma membrane NADPH oxidase) that produces ROS for calcium signaling, in accordance with its role in stomatal closure [42]. However, things can be more complex since some regulations seem to go in opposite directions. A big node in the regulatory network is the kinase CIPK23 that interacts with and phosphorylates multiple membrane targets including AKT1, HAK5, SLAC1, and CHL1 [127]. It activates both AKT1, which prevents stomatal closing, and SLAC1/SLAH3 channels, which are involved in stomatal closure. The phenotype of the *cipk23* mutants (enhanced tolerance to drought stress [43]) rather fits with its role in AKT1 regulation. In the CPK family, CPK13 inhibits Kin channels KAT1 and KAT2 [170], but also the K⁺ efflux channel GORK [163]. CPK33 activates GORK and favors calcium-dependent stomatal closure [163] but downregulates slow anion channels and slows down ABA-mediated stomatal closure [173]. The *cpk21* mutant also displays resistance to mannitol [172], while CPK21 activates SLAH3 [172], which would result in enhanced stomatal closure and drought resistance. A multiplicity of targets (Figures 1–5) and variations in spatiotemporal gene expression might explain why mutant phenotypes are not always consistent with what is expected when considering only one mechanism. Furthermore, knocking down one gene can result in the deregulation of other genes in stress signaling processes, as shown for CPK23 [139] and CPK21 [172]. Clade A PP2Cs such as AtPP2CA, ABI1, and ABI2 are also key nodes in ABA and calcium-dependent signaling. They target multiple proteins in the cytosol and nucleus to allow plant adaptation to multiple stimuli [216]. Depending on environmental conditions, they trap or release kinases that modulate the activity of ion channels and transporters, and they also directly bind to ion membrane transporters and CBL proteins. Among CBLs, CBL1 is a big hub. The gene is upregulated by salt stress, while ABA and abiotic stresses (salt, osmotic) result in the accumulation of the protein. The protein is under control of PP2Cs, and it has multiple downstream targets that can interact with each other (Figures 1, 2 and 5). This shows the inter-connection of different signaling pathways associating ion transporters (cations, anions), kinases (CIPKs, CPKs, SnRK2), CBLs, and PP2C phosphatases, and suggests the existence of multi-protein complexes that can be recruited according to the plant's status and environment.

Funding: This work was supported by the Sweetkaligrappe ANR (Agence Nationale pour la Recherche) contract ANR-14-CE20-0002.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. De Boeck, H.J.; Dreesen, F.E.; Janssens, I.A.; Nijs, I. Whole-system responses of experimental plant communities to climate extremes imposed in different seasons. *New Phytol.* **2011**, *189*, 806–817. [[CrossRef](#)]
2. Fahad, S.; Bajwa, A.A.; Nazir, U.; Anjum, S.A.; Farooq, A.; Zohalb, A.; Sadia, S.; Nasim, W.; Adkins, S.; Saud, S.; et al. Crop production under drought and heat stress: Plant responses and management options. *Front. Plant Sci.* **2017**, *8*, 1147. [[CrossRef](#)]
3. Lamaoui, M.; Jemo, M.; Datla, R.; Bekkaoui, F. Heat and Drought Stresses in Crops and Approaches for Their Mitigation. *Front. Chem.* **2018**, *6*, 26. [[CrossRef](#)]
4. Zhu, J.K. Abiotic Stress Signaling and Responses in Plants. *Cell* **2016**, *167*, 313–324. [[CrossRef](#)]
5. Tuteja, N. Abscisic Acid and abiotic stress signaling. *Plant Signal. Behav.* **2007**, *2*, 135–138. [[CrossRef](#)]

6. Hong, J.H.; Seah, S.W.; Xu, J. The root of ABA action in environmental stress response. *Plant Cell Rep.* **2013**, *32*, 971–983. [[CrossRef](#)]
7. Sah, S.K.; Reddy, K.R.; Li, J. Abscisic Acid and Abiotic Stress Tolerance in Crop Plants. *Front. Plant Sci.* **2016**, *7*, 571. [[CrossRef](#)]
8. Larkindale, J.; Knight, M.R. Protection against heat stress-induced oxidative damage in Arabidopsis involves calcium, abscisic acid, ethylene, and salicylic acid. *Plant Physiol.* **2002**, *128*, 682–695. [[CrossRef](#)]
9. Larkindale, J.; Hall, J.D.; Knight, M.R.; Vierling, E. Heat stress phenotypes of Arabidopsis mutants implicate multiple signaling pathways in the acquisition of thermotolerance. *Plant Physiol.* **2005**, *138*, 882–897. [[CrossRef](#)]
10. Kuromori, T.; Seo, M.; Shinozaki, K. ABA Transport and Plant Water Stress Responses. *Trends Plant Sci.* **2018**, *23*, 513–522. [[CrossRef](#)]
11. Lacombe, B.; Achard, P. Long-distance transport of phytohormones through the plant vascular system. *Curr. Opin. Plant Biol.* **2016**, *34*, 1–8. [[CrossRef](#)]
12. Geilfus, C.M. The pH of the Apoplast: Dynamic Factor with Functional Impact Under Stress. *Mol. Plant* **2017**, *10*, 1371–1386. [[CrossRef](#)]
13. Luan, M.; Tang, R.J.; Tang, Y.; Tian, W.; Hou, C.; Zhao, F.; Lan, W.; Luan, S. Transport and homeostasis of potassium and phosphate: Limiting factors for sustainable crop production. *J. Exp. Bot.* **2017**, *68*, 3091–3105. [[CrossRef](#)]
14. Zhang, Y.; Lv, Y.; Jahan, N.; Chen, G.; Ren, D.; Guo, L. Sensing of Abiotic Stress and Ionic Stress Responses in Plants. *Int. J. Mol. Sci.* **2018**, *19*, 3298. [[CrossRef](#)]
15. Sharma, T.; Dreyer, I.; Riedelsberger, J. The role of K(+) channels in uptake and redistribution of potassium in the model plant Arabidopsis thaliana. *Front. Plant Sci.* **2013**, *4*, 224. [[CrossRef](#)]
16. Collins, K.D. Charge density-dependent strength of hydration and biological structure. *Biophys. J.* **1997**, *72*, 65–76. [[CrossRef](#)]
17. Leigh, R.A.; Wyn Jones, R.G. A hypothesis relating critical potassium concentrations for growth to the distribution of this ion in the plant cell. *New Phytol.* **1984**, *97*, 1–13. [[CrossRef](#)]
18. Benito, B.; Haro, R.; Amtmann, A.; Cuin, T.A.; Dreyer, I. The twins K+ and Na+ in plants. *J. Plant Physiol.* **2014**, *171*, 723–731. [[CrossRef](#)]
19. Maathuis, F.J. Physiological functions of mineral macronutrients. *Curr. Opin. Plant Biol.* **2009**, *12*, 250–258. [[CrossRef](#)]
20. Trankner, M.; Tavakol, E.; Jakli, B. Functioning of potassium and magnesium in photosynthesis, photosynthate translocation and photoprotection. *Physiol. Plant.* **2018**. [[CrossRef](#)]
21. Shabala, S.; Pottosin, I. Regulation of potassium transport in plants under hostile conditions: Implications for abiotic and biotic stress tolerance. *Physiol. Plant.* **2014**, *151*, 257–279. [[CrossRef](#)] [[PubMed](#)]
22. Roberts, S.K. Regulation of K+ channels in maize roots by water stress and abscisic acid. *Plant Physiol.* **1998**, *116*, 145–153. [[CrossRef](#)]
23. Anschutz, U.; Becker, D.; Shabala, S. Going beyond nutrition: Regulation of potassium homeostasis as a common denominator of plant adaptive responses to environment. *J. Plant Physiol.* **2014**, *171*, 670–687. [[CrossRef](#)] [[PubMed](#)]
24. Shabala, S.; Cuin, T.A. Potassium transport and plant salt tolerance. *Physiol. Plant.* **2008**, *133*, 651–669. [[CrossRef](#)] [[PubMed](#)]
25. Cakmak, I. The role of potassium in alleviating detrimental effects of abiotic stresses in plants. *J. Plant Nutr. Soil Sci.* **2005**, *168*, 521–530. [[CrossRef](#)]
26. Maser, P.; Thomine, S.; Schroeder, J.I.; Ward, J.M.; Hirschi, K.; Sze, H.; Talke, I.N.; Amtmann, A.; Maathuis, F.J.; Sanders, D.; et al. Phylogenetic relationships within cation transporter families of Arabidopsis. *Plant Physiol.* **2001**, *126*, 1646–1667. [[CrossRef](#)] [[PubMed](#)]
27. Very, A.A.; Nieves-Cordones, M.; Daly, M.; Khan, I.; Fizames, C.; Sentenac, H. Molecular biology of K+ transport across the plant cell membrane: What do we learn from comparison between plant species? *J. Plant Physiol.* **2014**, *171*, 748–769. [[CrossRef](#)] [[PubMed](#)]
28. Dreyer, I.; Antunes, S.; Hoshi, T.; Muller-Rober, B.; Palme, K.; Pongs, O.; Reintanz, B.; Hedrich, R. Plant K+ channel alpha-subunits assemble indiscriminately. *Biophys. J.* **1997**, *72*, 2143–2150. [[CrossRef](#)]

29. Xicluna, J.; Lacombe, B.; Dreyer, I.; Alcon, C.; Jeanguenin, L.; Sentenac, H.; Thibaud, J.B.; Cherel, I. Increased functional diversity of plant K⁺ channels by preferential heteromerization of the shaker-like subunits AKT2 and KAT2. *J. Biol. Chem.* **2007**, *282*, 486–494. [[CrossRef](#)] [[PubMed](#)]
30. Lebaudy, A.; Pascaud, F.; Very, A.A.; Alcon, C.; Dreyer, I.; Thibaud, J.B.; Lacombe, B. Preferential KAT1-KAT2 heteromerization determines inward K⁺ current properties in Arabidopsis guard cells. *J. Biol. Chem.* **2010**, *285*, 6265–6274. [[CrossRef](#)] [[PubMed](#)]
31. Dreyer, I.; Michard, E.; Lacombe, B.; Thibaud, J.B. A plant Shaker-like K⁺ channel switches between two distinct gating modes resulting in either inward-rectifying or “leak” current. *FEBS Lett.* **2001**, *505*, 233–239. [[CrossRef](#)]
32. Duby, G.; Hosy, E.; Fizames, C.; Alcon, C.; Costa, A.; Sentenac, H.; Thibaud, J.B. AtKC1, a conditionally targeted Shaker-type subunit, regulates the activity of plant K⁺ channels. *Plant J.* **2008**, *53*, 115–123. [[CrossRef](#)] [[PubMed](#)]
33. Geiger, D.; Becker, D.; Vosloh, D.; Gambale, F.; Palme, K.; Rehers, M.; Anschuetz, U.; Dreyer, I.; Kudla, J.; Hedrich, R. Heteromeric AtKC1/AKT1 channels in Arabidopsis roots facilitate growth under K⁺-limiting conditions. *J. Biol. Chem.* **2009**, *284*, 21288–21295. [[CrossRef](#)] [[PubMed](#)]
34. Jeanguenin, L.; Alcon, C.; Duby, G.; Boeglin, M.; Cherel, I.; Gaillard, I.; Zimmermann, S.; Sentenac, H.; Very, A.A. AtKC1 is a general modulator of Arabidopsis inward Shaker channel activity. *Plant J.* **2011**, *67*, 570–582. [[CrossRef](#)] [[PubMed](#)]
35. Reintanz, B.; Szyroki, A.; Ivashikina, N.; Ache, P.; Godde, M.; Becker, D.; Palme, K.; Hedrich, R. AtKC1, a silent Arabidopsis potassium channel alpha -subunit modulates root hair K⁺ influx. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 4079–4084. [[CrossRef](#)] [[PubMed](#)]
36. Lebaudy, A.; Very, A.A.; Sentenac, H. K⁺ channel activity in plants: Genes, regulations and functions. *FEBS Lett.* **2007**, *581*, 2357–2366. [[CrossRef](#)] [[PubMed](#)]
37. Voelker, C.; Gomez-Porrás, J.L.; Becker, D.; Hamamoto, S.; Uozumi, N.; Gambale, F.; Mueller-Roeber, B.; Czempinski, K.; Dreyer, I. Roles of tandem-pore K⁺ channels in plants - a puzzle still to be solved. *Plant Biol.* **2010**, *12* (Suppl. 1), 56–63. [[CrossRef](#)] [[PubMed](#)]
38. Becker, D.; Geiger, D.; Dunkel, M.; Roller, A.; Bertl, A.; Latz, A.; Carpaneto, A.; Dietrich, P.; Roelfsema, M.R.; Voelker, C.; et al. AtTPK4, an Arabidopsis tandem-pore K⁺ channel, poised to control the pollen membrane voltage in a pH- and Ca²⁺-dependent manner. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 15621–15626. [[CrossRef](#)] [[PubMed](#)]
39. Carraretto, L.; Formentin, E.; Teardo, E.; Checchetto, V.; Tomizioli, M.; Morosinotto, T.; Giacometti, G.M.; Finazzi, G.; Szabo, I. A thylakoid-located two-pore K⁺ channel controls photosynthetic light utilization in plants. *Science* **2013**, *342*, 114–118. [[CrossRef](#)] [[PubMed](#)]
40. Bihler, H.; Eing, C.; Hebeisen, S.; Roller, A.; Czempinski, K.; Bertl, A. TPK1 is a vacuolar ion channel different from the slow-vacuolar cation channel. *Plant Physiol.* **2005**, *139*, 417–424. [[CrossRef](#)] [[PubMed](#)]
41. Gobert, A.; Isayenkov, S.; Voelker, C.; Czempinski, K.; Maathuis, F.J. The two-pore channel TPK1 gene encodes the vacuolar K⁺ conductance and plays a role in K⁺ homeostasis. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 10726–10731. [[CrossRef](#)] [[PubMed](#)]
42. Sierla, M.; Waszczak, C.; Vahisalu, T.; Kangasjarvi, J. Reactive Oxygen Species in the Regulation of Stomatal Movements. *Plant Physiol.* **2016**, *171*, 1569–1580. [[CrossRef](#)] [[PubMed](#)]
43. Nieves-Cordones, M.; Caballero, F.; Martinez, V.; Rubio, F. Disruption of the Arabidopsis thaliana inward-rectifier K⁺ channel AKT1 improves plant responses to water stress. *Plant Cell Physiol.* **2012**, *53*, 423–432. [[CrossRef](#)] [[PubMed](#)]
44. Pilot, G.; Gaymard, F.; Mouline, K.; Cherel, I.; Sentenac, H. Regulated expression of Arabidopsis shaker K⁺ channel genes involved in K⁺ uptake and distribution in the plant. *Plant Mol. Biol.* **2003**, *51*, 773–787. [[CrossRef](#)] [[PubMed](#)]
45. Winter, D.; Vinegar, B.; Nahal, H.; Ammar, R.; Wilson, G.V.; Provar, N.J. An “Electronic Fluorescent Pictograph” browser for exploring and analyzing large-scale biological data sets. *PLoS ONE* **2007**, *2*, e718. [[CrossRef](#)] [[PubMed](#)]
46. Li, J.; Long, Y.; Qi, G.N.; Li, J.; Xu, Z.J.; Wu, W.H.; Wang, Y. The Os-AKT1 channel is critical for K⁺ uptake in rice roots and is modulated by the rice CBL1-CIPK23 complex. *Plant Cell* **2014**, *26*, 3387–3402. [[CrossRef](#)] [[PubMed](#)]

47. Ahmad, I.; Mian, A.; Maathuis, F.J. Overexpression of the rice AKT1 potassium channel affects potassium nutrition and rice drought tolerance. *J. Exp. Bot.* **2016**, *67*, 2689–2698. [[CrossRef](#)] [[PubMed](#)]
48. Fuchs, I.; Stolzle, S.; Ivashikina, N.; Hedrich, R. Rice K⁺ uptake channel OsAKT1 is sensitive to salt stress. *Planta* **2005**, *221*, 212–221. [[CrossRef](#)]
49. Golladack, D.; Quigley, F.; Michalowski, C.B.; Kamasani, U.R.; Bohnert, H.J. Salinity stress-tolerant and -sensitive rice (*Oryza sativa* L.) regulate AKT1-type potassium channel transcripts differently. *Plant Mol. Biol.* **2003**, *51*, 71–81. [[CrossRef](#)]
50. Cuellar, T.; Pascaud, F.; Verdeil, J.L.; Torregrosa, L.; Adam-Blondon, A.F.; Thibaud, J.B.; Sentenac, H.; Gaillard, I. A grapevine Shaker inward K(+) channel activated by the calcineurin B-like calcium sensor 1-protein kinase CIPK23 network is expressed in grape berries under drought stress conditions. *Plant J.* **2010**, *61*, 58–69. [[CrossRef](#)]
51. Cuellar, T.; Azeem, F.; Andrianteranagna, M.; Pascaud, F.; Verdeil, J.L.; Sentenac, H.; Zimmermann, S.; Gaillard, I. Potassium transport in developing fleshy fruits: The grapevine inward K(+) channel VvK1.2 is activated by CIPK-CBL complexes and induced in ripening berry flesh cells. *Plant J.* **2013**, *73*, 1006–1018. [[CrossRef](#)] [[PubMed](#)]
52. Deeken, R.; Geiger, D.; Fromm, J.; Koroleva, O.; Ache, P.; Langenfeld-Heyser, R.; Sauer, N.; May, S.T.; Hedrich, R. Loss of the AKT2/3 potassium channel affects sugar loading into the phloem of Arabidopsis. *Planta* **2002**, *216*, 334–344. [[CrossRef](#)] [[PubMed](#)]
53. Dreyer, I.; Gomez-Porras, J.L.; Riedelsberger, J. The potassium battery: A mobile energy source for transport processes in plant vascular tissues. *New Phytol.* **2017**, *216*, 1049–1053. [[CrossRef](#)] [[PubMed](#)]
54. Lacombe, B.; Pilot, G.; Michard, E.; Gaymard, F.; Sentenac, H.; Thibaud, J.B. A shaker-like K(+) channel with weak rectification is expressed in both source and sink phloem tissues of Arabidopsis. *Plant Cell* **2000**, *12*, 837–851. [[PubMed](#)]
55. Leonhardt, N.; Kwak, J.M.; Robert, N.; Waner, D.; Leonhardt, G.; Schroeder, J.I. Microarray expression analyses of Arabidopsis guard cells and isolation of a recessive abscisic acid hypersensitive protein phosphatase 2C mutant. *Plant Cell* **2004**, *16*, 596–615. [[CrossRef](#)] [[PubMed](#)]
56. Kilian, J.; Whitehead, D.; Horak, J.; Wanke, D.; Weinl, S.; Batistic, O.; D'Angelo, C.; Bornberg-Bauer, E.; Kudla, J.; Harter, K. The AtGenExpress global stress expression data set: Protocols, evaluation and model data analysis of UV-B light, drought and cold stress responses. *Plant J.* **2007**, *50*, 347–363. [[CrossRef](#)] [[PubMed](#)]
57. Maathuis, F.J. The role of monovalent cation transporters in plant responses to salinity. *J. Exp. Bot.* **2006**, *57*, 1137–1147. [[CrossRef](#)]
58. Suzuki, N.; Bassil, E.; Hamilton, J.S.; Inupakutika, M.A.; Zandalinas, S.I.; Tripathy, D.; Luo, Y.; Dion, E.; Fukui, G.; Kumazaki, A.; et al. ABA Is Required for Plant Acclimation to a Combination of Salt and Heat Stress. *PLoS ONE* **2016**, *11*, e0147625. [[CrossRef](#)]
59. Nieves-Cordones, M.; Andrianteranagna, M.; Cuéllar, T.; Chérel, I.; Gibrat, R.; Boeglin, M.; Moreau, B.; Paris, N.; Verdeil, J.L.; Zimmermann, S.; et al. Characterization of the grapevine Shaker K⁺ channel VvK3.1 supports its function in massive potassium fluxes necessary for berry potassium loading and pulvinus-actuated leaf movements. *New Phytol.* **2019**, in press. [[CrossRef](#)]
60. Pilot, G.; Lacombe, B.; Gaymard, F.; Cherel, I.; Boucherez, J.; Thibaud, J.B.; Sentenac, H. Guard cell inward K⁺ channel activity in Arabidopsis involves expression of the twin channel subunits KAT1 and KAT2. *J. Biol. Chem.* **2001**, *276*, 3215–3221. [[CrossRef](#)]
61. Kwak, J.M.; Murata, Y.; Baizabal-Aguirre, V.M.; Merrill, J.; Wang, M.; Kemper, A.; Hawke, S.D.; Tallman, G.; Schroeder, J.I. Dominant negative guard cell K⁺ channel mutants reduce inward-rectifying K⁺ currents and light-induced stomatal opening in Arabidopsis. *Plant Physiol.* **2001**, *127*, 473–485. [[CrossRef](#)] [[PubMed](#)]
62. Zhang, A.; Ren, H.M.; Tan, Y.Q.; Qi, G.N.; Yao, F.Y.; Wu, G.L.; Yang, L.W.; Hussain, J.; Sun, S.J.; Wang, Y.F. S-type Anion Channels SLAC1 and SLAH3 Function as Essential Negative Regulators of Inward K⁺ Channels and Stomatal Opening in Arabidopsis. *Plant Cell* **2016**, *28*, 949–955. [[CrossRef](#)] [[PubMed](#)]
63. Gaymard, F.; Pilot, G.; Lacombe, B.; Bouchez, D.; Bruneau, D.; Boucherez, J.; Michaux-Ferriere, N.; Thibaud, J.B.; Sentenac, H. Identification and disruption of a plant shaker-like outward channel involved in K⁺ release into the xylem sap. *Cell* **1998**, *94*, 647–655. [[CrossRef](#)]

64. Hosy, E.; Vavasseur, A.; Mouline, K.; Dreyer, I.; Gaymard, F.; Poree, F.; Boucherez, J.; Lebaudy, A.; Bouchez, D.; Very, A.A.; et al. The Arabidopsis outward K⁺ channel GORK is involved in regulation of stomatal movements and plant transpiration. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 5549–5554. [[CrossRef](#)]
65. Becker, D.; Hoth, S.; Ache, P.; Wenkel, S.; Roelfsema, M.R.; Meyerhoff, O.; Hartung, W.; Hedrich, R. Regulation of the ABA-sensitive Arabidopsis potassium channel gene GORK in response to water stress. *FEBS Lett.* **2003**, *554*, 119–126. [[CrossRef](#)]
66. Nieves-Cordones, M.; Rodenas, R.; Chavanieu, A.; Rivero, R.M.; Martinez, V.; Gaillard, I.; Rubio, F. Uneven HAK/KUP/KT Protein Diversity Among Angiosperms: Species Distribution and Perspectives. *Front. Plant Sci.* **2016**, *7*, 127. [[CrossRef](#)]
67. Nieves-Cordones, M.; Aleman, F.; Martinez, V.; Rubio, F. The Arabidopsis thaliana HAK5 K⁺ transporter is required for plant growth and K⁺ acquisition from low K⁺ solutions under saline conditions. *Mol. Plant* **2010**, *3*, 326–333. [[CrossRef](#)]
68. Osakabe, Y.; Arinaga, N.; Umezawa, T.; Katsura, S.; Nagamachi, K.; Tanaka, H.; Ohiraki, H.; Yamada, K.; Seo, S.U.; Abo, M.; et al. Osmotic stress responses and plant growth controlled by potassium transporters in Arabidopsis. *Plant Cell* **2013**, *25*, 609–624. [[CrossRef](#)]
69. Chen, G.; Liu, C.; Gao, Z.; Zhang, Y.; Jiang, H.; Zhu, L.; Ren, D.; Yu, L.; Xu, G.; Qian, Q. OsHAK1, a High-Affinity Potassium Transporter, Positively Regulates Responses to Drought Stress in Rice. *Front. Plant Sci.* **2017**, *8*, 1885. [[CrossRef](#)]
70. Yang, T.; Zhang, S.; Hu, Y.; Wu, F.; Hu, Q.; Chen, G.; Cai, J.; Wu, T.; Moran, N.; Yu, L.; et al. The role of a potassium transporter OsHAK5 in potassium acquisition and transport from roots to shoots in rice at low potassium supply levels. *Plant Physiol.* **2014**, *166*, 945–959. [[CrossRef](#)]
71. Shen, Y.; Shen, L.; Shen, Z.; Jing, W.; Ge, H.; Zhao, J.; Zhang, W. The potassium transporter OsHAK21 functions in the maintenance of ion homeostasis and tolerance to salt stress in rice. *Plant Cell Environ.* **2015**, *38*, 2766–2779. [[CrossRef](#)] [[PubMed](#)]
72. Latz, A.; Mehlmer, N.; Zapf, S.; Mueller, T.D.; Wurzing, B.; Pfister, B.; Csaszar, E.; Hedrich, R.; Teige, M.; Becker, D. Salt stress triggers phosphorylation of the Arabidopsis vacuolar K⁺ channel TPK1 by calcium-dependent protein kinases (CDPKs). *Mol. Plant* **2013**, *6*, 1274–1289. [[CrossRef](#)] [[PubMed](#)]
73. Yang, Y.; Costa, A.; Leonhardt, N.; Siegel, R.S.; Schroeder, J.I. Isolation of a strong Arabidopsis guard cell promoter and its potential as a research tool. *Plant Methods* **2008**, *4*, 6. [[CrossRef](#)] [[PubMed](#)]
74. Vahisalu, T.; Kollist, H.; Wang, Y.F.; Nishimura, N.; Chan, W.Y.; Valerio, G.; Lamminmaki, A.; Brosche, M.; Moldau, H.; Desikan, R.; et al. SLAC1 is required for plant guard cell S-type anion channel function in stomatal signalling. *Nature* **2008**, *452*, 487–491. [[CrossRef](#)] [[PubMed](#)]
75. Negi, J.; Matsuda, O.; Nagasawa, T.; Oba, Y.; Takahashi, H.; Kawai-Yamada, M.; Uchimiya, H.; Hashimoto, M.; Iba, K. CO₂ regulator SLAC1 and its homologues are essential for anion homeostasis in plant cells. *Nature* **2008**, *452*, 483–486. [[CrossRef](#)]
76. Pandey, S.; Wang, R.S.; Wilson, L.; Li, S.; Zhao, Z.; Gookin, T.E.; Assmann, S.M.; Albert, R. Boolean modeling of transcriptome data reveals novel modes of heterotrimeric G-protein action. *Mol. Syst. Biol.* **2010**, *6*, 372. [[CrossRef](#)] [[PubMed](#)]
77. Cubero-Font, P.; Maierhofer, T.; Jaslan, J.; Rosales, M.A.; Espartero, J.; Diaz-Rueda, P.; Muller, H.M.; Hurter, A.L.; Al-Rasheid, K.A.; Marten, I.; et al. Silent S-Type Anion Channel Subunit SLAH1 Gates SLAH3 Open for Chloride Root-to-Shoot Translocation. *Curr. Biol.* **2016**, *26*, 2213–2220. [[CrossRef](#)] [[PubMed](#)]
78. Geiger, D.; Maierhofer, T.; Al-Rasheid, K.A.; Scherzer, S.; Mumm, P.; Liese, A.; Ache, P.; Wellmann, C.; Marten, I.; Grill, E.; et al. Stomatal closure by fast abscisic acid signaling is mediated by the guard cell anion channel SLAH3 and the receptor RCAR1. *Sci. Signal.* **2011**, *4*, ra32. [[CrossRef](#)]
79. Gutermuth, T.; Lassig, R.; Portes, M.T.; Maierhofer, T.; Romeis, T.; Borst, J.W.; Hedrich, R.; Feijo, J.A.; Konrad, K.R. Pollen tube growth regulation by free anions depends on the interaction between the anion channel SLAH3 and calcium-dependent protein kinases CPK2 and CPK20. *Plant Cell* **2013**, *25*, 4525–4543. [[CrossRef](#)] [[PubMed](#)]
80. Tsay, Y.F.; Schroeder, J.I.; Feldmann, K.A.; Crawford, N.M. The herbicide sensitivity gene CHL1 of Arabidopsis encodes a nitrate-inducible nitrate transporter. *Cell* **1993**, *72*, 705–713. [[CrossRef](#)]
81. Guo, F.Q.; Young, J.; Crawford, N.M. The nitrate transporter AtNRT1.1 (CHL1) functions in stomatal opening and contributes to drought susceptibility in Arabidopsis. *Plant Cell* **2003**, *15*, 107–117. [[CrossRef](#)] [[PubMed](#)]

82. Krouk, G.; Lacombe, B.; Bielach, A.; Perrine-Walker, F.; Malinska, K.; Mounier, E.; Hoyerova, K.; Tillard, P.; Leon, S.; Ljung, K.; et al. Nitrate-regulated auxin transport by NRT1.1 defines a mechanism for nutrient sensing in plants. *Dev. Cell* **2010**, *18*, 927–937. [[CrossRef](#)] [[PubMed](#)]
83. Nieves-Cordones, M.; Al Shiblawi, F.R.; Sentenac, H. Roles and Transport of Sodium and Potassium in Plants. *Met. Ions Life Sci.* **2016**, *16*, 291–324. [[PubMed](#)]
84. Santa-Maria, G.E.; Olfieruk, S.; Moriconi, J.I. KT-HAK-KUP transporters in major terrestrial photosynthetic organisms: A twenty years tale. *J. Plant Physiol.* **2018**, *226*, 77–90. [[CrossRef](#)] [[PubMed](#)]
85. Li, W.; Xu, G.; Alli, A.; Yu, L. Plant HAK/KUP/KT K(+) transporters: Function and regulation. *Semin. Cell Dev. Biol.* **2018**, *74*, 133–141. [[CrossRef](#)] [[PubMed](#)]
86. Fu, H.H.; Luan, S. AtKup1: A dual-affinity K⁺ transporter from Arabidopsis. *Plant Cell* **1998**, *10*, 63–73. [[PubMed](#)]
87. Sze, H.; Chanroj, S. Plant Endomembrane Dynamics: Studies of K(+)/H(+) Antiporters Provide Insights on the Effects of pH and Ion Homeostasis. *Plant Physiol.* **2018**, *177*, 875–895. [[CrossRef](#)] [[PubMed](#)]
88. Rubio, F.; Nieves-Cordones, M.; Aleman, F.; Martinez, V. Relative contribution of AtHAK5 and AtAKT1 to K⁺ uptake in the high-affinity range of concentrations. *Physiol. Plant.* **2008**, *134*, 598–608. [[CrossRef](#)] [[PubMed](#)]
89. Aleman, F.; Nieves-Cordones, M.; Martinez, V.; Rubio, F. Root K(+) acquisition in plants: The Arabidopsis thaliana model. *Plant Cell Physiol.* **2011**, *52*, 1603–1612. [[CrossRef](#)] [[PubMed](#)]
90. Demidchik, V.; Cuin, T.A.; Svistunenko, D.; Smith, S.J.; Miller, A.J.; Shabala, S.; Sokolik, A.; Yurin, V. Arabidopsis root K⁺-efflux conductance activated by hydroxyl radicals: Single-channel properties, genetic basis and involvement in stress-induced cell death. *J. Cell Sci.* **2010**, *123 Pt 9*, 1468–1479. [[CrossRef](#)]
91. Han, M.; Wu, W.; Wu, W.H.; Wang, Y. Potassium Transporter KUP7 Is Involved in K(+) Acquisition and Translocation in Arabidopsis Root under K(+)-Limited Conditions. *Mol. Plant* **2016**, *9*, 437–446. [[CrossRef](#)] [[PubMed](#)]
92. Jayakannan, M.; Bose, J.; Babourina, O.; Rengel, Z.; Shabala, S. Salicylic acid improves salinity tolerance in Arabidopsis by restoring membrane potential and preventing salt-induced K⁺ loss via a GORK channel. *J. Exp. Bot.* **2013**, *64*, 2255–2268. [[CrossRef](#)] [[PubMed](#)]
93. Demidchik, V. Mechanisms and physiological roles of K⁺ efflux from root cells. *J. Plant Physiol.* **2014**, *171*, 696–707. [[CrossRef](#)] [[PubMed](#)]
94. Bose, J.; Shabala, L.; Pottosin, I.; Zeng, F.; Velarde-Buendia, A.M.; Massart, A.; Poschenrieder, C.; Hariadi, Y.; Shabala, S. Kinetics of xylem loading, membrane potential maintenance, and sensitivity of K(+) -permeable channels to reactive oxygen species: Physiological traits that differentiate salinity tolerance between pea and barley. *Plant Cell Environ.* **2014**, *37*, 589–600. [[CrossRef](#)] [[PubMed](#)]
95. Benlloch-Gonzalez, M.; Sanchez-Lucas, R.; Benlloch, M. Effects of olive root warming on potassium transport and plant growth. *J. Plant Physiol.* **2017**, *218*, 182–188. [[CrossRef](#)] [[PubMed](#)]
96. Ivashikina, N.; Becker, D.; Ache, P.; Meyerhoff, O.; Felle, H.H.; Hedrich, R. K(+) channel profile and electrical properties of Arabidopsis root hairs. *FEBS Lett.* **2001**, *508*, 463–469. [[CrossRef](#)]
97. Hetherington, A.M. Plant physiology: Spreading a drought warning. *Curr. Biol.* **1998**, *8*, R911–R913. [[CrossRef](#)]
98. Chen, G.; Hu, Q.; Luo, L.; Yang, T.; Zhang, S.; Hu, Y.; Yu, L.; Xu, G. Rice potassium transporter OsHAK1 is essential for maintaining potassium-mediated growth and functions in salt tolerance over low and high potassium concentration ranges. *Plant Cell Environ.* **2015**, *38*, 2747–2765. [[CrossRef](#)]
99. Marten, I.; Hoth, S.; Deeken, R.; Ache, P.; Ketchum, K.A.; Hoshi, T.; Hedrich, R. AKT3, a phloem-localized K⁺ channel, is blocked by protons. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 7581–7586. [[CrossRef](#)]
100. Ivashikina, N.; Deeken, R.; Ache, P.; Kranz, E.; Pommerrenig, B.; Sauer, N.; Hedrich, R. Isolation of AtSUC2 promoter-GFP-marked companion cells for patch-clamp studies and expression profiling. *Plant J.* **2003**, *36*, 931–945. [[CrossRef](#)]
101. Deeken, R.; Sanders, C.; Ache, P.; Hedrich, R. Developmental and light-dependent regulation of a phloem-localised K⁺ channel of Arabidopsis thaliana. *Plant J.* **2000**, *23*, 285–290. [[CrossRef](#)] [[PubMed](#)]
102. Ache, P.; Becker, D.; Deeken, R.; Dreyer, I.; Weber, H.; Fromm, J.; Hedrich, R. VFK1, a Vicia faba K(+) channel involved in phloem unloading. *Plant J.* **2001**, *27*, 571–580. [[CrossRef](#)] [[PubMed](#)]

103. Philippar, K.; Buchsenschutz, K.; Abshagen, M.; Fuchs, I.; Geiger, D.; Lacombe, B.; Hedrich, R. The K⁺ channel KZM1 mediates potassium uptake into the phloem and guard cells of the C4 grass *Zea mays*. *J. Biol. Chem.* **2003**, *278*, 16973–16981. [[CrossRef](#)] [[PubMed](#)]
104. Hafke, J.B.; Furch, A.C.; Reitz, M.U.; van Bel, A.J. Functional sieve element protoplasts. *Plant Physiol.* **2007**, *145*, 703–711. [[CrossRef](#)] [[PubMed](#)]
105. Hubbard, K.E.; Nishimura, N.; Hitomi, K.; Getzoff, E.D.; Schroeder, J.I. Early abscisic acid signal transduction mechanisms: Newly discovered components and newly emerging questions. *Genes Dev.* **2010**, *24*, 1695–1708. [[CrossRef](#)] [[PubMed](#)]
106. Joshi-Saha, A.; Valon, C.; Leung, J. A brand new START: Abscisic acid perception and transduction in the guard cell. *Sci. Signal.* **2011**, *4*, re4. [[CrossRef](#)] [[PubMed](#)]
107. Lim, C.W.; Baek, W.; Jung, J.; Kim, J.H.; Lee, S.C. Function of ABA in Stomatal Defense against Biotic and Drought Stresses. *Int. J. Mol. Sci.* **2015**, *16*, 15251–15270. [[CrossRef](#)] [[PubMed](#)]
108. Assmann, S.M.; Jegla, T. Guard cell sensory systems: Recent insights on stomatal responses to light, abscisic acid, and CO₂. *Curr. Opin. Plant Biol.* **2016**, *33*, 157–167. [[CrossRef](#)] [[PubMed](#)]
109. Jezek, M.; Blatt, M.R. The Membrane Transport System of the Guard Cell and Its Integration for Stomatal Dynamics. *Plant Physiol.* **2017**, *174*, 487–519. [[CrossRef](#)] [[PubMed](#)]
110. Chaves, M.M.; Costa, J.M.; Zarrouk, O.; Pinheiro, C.; Lopes, C.M.; Pereira, J.S. Controlling stomatal aperture in semi-arid regions-The dilemma of saving water or being cool? *Plant Sci.* **2016**, *251*, 54–64. [[CrossRef](#)] [[PubMed](#)]
111. Gayatri, G.; Agurla, S.; Raghavendra, A.S. Nitric oxide in guard cells as an important secondary messenger during stomatal closure. *Front. Plant Sci.* **2013**, *4*, 425. [[CrossRef](#)] [[PubMed](#)]
112. Murata, Y.; Mori, I.C.; Munemasa, S. Diverse stomatal signaling and the signal integration mechanism. *Annu. Rev. Plant Biol.* **2015**, *66*, 369–392. [[CrossRef](#)] [[PubMed](#)]
113. Farnese, F.S.; Menezes-Silva, P.E.; Gusman, G.S.; Oliveira, J.A. When Bad Guys Become Good Ones: The Key Role of Reactive Oxygen Species and Nitric Oxide in the Plant Responses to Abiotic Stress. *Front. Plant Sci.* **2016**, *7*, 471. [[CrossRef](#)] [[PubMed](#)]
114. Kollist, H.; Jossier, M.; Laanemets, K.; Thomine, S. Anion channels in plant cells. *FEBS J.* **2011**, *278*, 4277–4292. [[CrossRef](#)] [[PubMed](#)]
115. Ache, P.; Becker, D.; Ivashikina, N.; Dietrich, P.; Roelfsema, M.R.; Hedrich, R. GORK, a delayed outward rectifier expressed in guard cells of *Arabidopsis thaliana*, is a K⁽⁺⁾-selective, K⁽⁺⁾-sensing ion channel. *FEBS Lett.* **2000**, *486*, 93–98. [[CrossRef](#)]
116. Szyroki, A.; Ivashikina, N.; Dietrich, P.; Roelfsema, M.R.; Ache, P.; Reintanz, B.; Deeken, R.; Godde, M.; Felle, H.; Steinmeyer, R.; et al. KAT1 is not essential for stomatal opening. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 2917–2921. [[CrossRef](#)] [[PubMed](#)]
117. Takahashi, Y.; Ebisu, Y.; Kinoshita, T.; Doi, M.; Okuma, E.; Murata, Y.; Shimazaki, K. bHLH transcription factors that facilitate K⁽⁺⁾ uptake during stomatal opening are repressed by abscisic acid through phosphorylation. *Sci. Signal.* **2013**, *6*, ra48. [[CrossRef](#)] [[PubMed](#)]
118. Hirsch, R.E.; Lewis, B.D.; Spalding, E.P.; Sussman, M.R. A role for the AKT1 potassium channel in plant nutrition. *Science* **1998**, *280*, 918–921. [[CrossRef](#)] [[PubMed](#)]
119. Isner, J.C.; Begum, A.; Nuehse, T.; Hetherington, A.M.; Maathuis, F.J.M. KIN7 Kinase Regulates the Vacuolar TPK1 K⁽⁺⁾ Channel during Stomatal Closure. *Curr. Biol.* **2018**, *28*, 466–472.e4. [[CrossRef](#)]
120. Mouline, K.; Very, A.A.; Gaymard, F.; Boucherez, J.; Pilot, G.; Devic, M.; Bouchez, D.; Thibaud, J.B.; Sentenac, H. Pollen tube development and competitive ability are impaired by disruption of a Shaker K⁽⁺⁾ channel in *Arabidopsis*. *Genes Dev.* **2002**, *16*, 339–350. [[CrossRef](#)]
121. Mira de Orduna, R. Climate change associated effects on grape and wine quality and production. *Food Res. Int.* **2010**, *43*, 1844–1855. [[CrossRef](#)]
122. Rogiers, S.Y.; Coetzee, Z.A.; Walker, R.R.; Deloire, A.; Tyerman, S.D. Potassium in the Grape (*Vitis vinifera* L.) Berry: Transport and Function. *Front. Plant Sci.* **2017**, *8*, 1629. [[CrossRef](#)]
123. Kodur, S. Effects of juice pH and potassium on juice and wine quality, and regulation of potassium in grapevines through rootstocks (*Vitis*): A short review. *Vitis* **2011**, *50*, 1–6.
124. Bondada, B.R.; Matthews, M.A.; Shackel, K.A. Functional xylem in the post-veraison grape berry. *J. Exp. Bot.* **2005**, *56*, 2949–2957. [[CrossRef](#)]

125. Davies, C.; Shin, R.; Liu, W.; Thomas, M.R.; Schachtman, D.P. Transporters expressed during grape berry (*Vitis vinifera* L.) development are associated with an increase in berry size and berry potassium accumulation. *J. Exp. Bot.* **2006**, *57*, 3209–3216. [[CrossRef](#)]
126. Mustilli, A.C.; Merlot, S.; Vavasseur, A.; Fenzi, F.; Giraudat, G. Arabidopsis OST1 protein kinase mediates the regulation of stomatal aperture by abscisic acid and acts upstream of reactive oxygen species production. *Plant Cell* **2002**, *14*, 3089–3099. [[CrossRef](#)]
127. Kudla, J.; Becker, D.; Grill, E.; Hedrich, R.; Hippler, M.; Kummer, U.; Parniske, M.; Romeis, T.; Schumacher, K. Advances and current challenges in calcium signaling. *New Phytol.* **2018**, *218*, 414–431. [[CrossRef](#)]
128. Kleist, T.J.; Luan, S. Constant change: Dynamic regulation of membrane transport by calcium signalling networks keeps plants in tune with their environment. *Plant Cell Environ.* **2016**, *39*, 467–481. [[CrossRef](#)]
129. Halford, N.G.; Hey, S.J. Snf1-related protein kinases (SnRKs) act within an intricate network that links metabolic and stress signalling in plants. *Biochem. J.* **2009**, *419*, 247–259. [[CrossRef](#)]
130. Coello, P.; Hey, S.J.; Halford, N.G. The sucrose non-fermenting-1-related (SnRK) family of protein kinases: Potential for manipulation to improve stress tolerance and increase yield. *J. Exp. Bot.* **2011**, *62*, 883–893. [[CrossRef](#)]
131. Boudsocq, M.; Sheen, J. CDPKs in immune and stress signaling. *Trends Plant. Sci.* **2013**, *18*, 30–40. [[CrossRef](#)]
132. Chaves-Sanjuan, A.; Sanchez-Barrena, M.J.; Gonzalez-Rubio, J.M.; Moreno, M.; Ragel, P.; Jimenez, M.; Pardo, J.M.; Martinez-Ripoll, M.; Quintero, F.J.; Albert, A. Structural basis of the regulatory mechanism of the plant CIPK family of protein kinases controlling ion homeostasis and abiotic stress. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, E4532–E4541. [[CrossRef](#)]
133. Hashimoto, K.; Eckert, C.; Anschutz, U.; Scholz, M.; Held, K.; Waadt, R.; Reyer, A.; Hippler, M.; Becker, D.; Kudla, J. Phosphorylation of calcineurin B-like (CBL) calcium sensor proteins by their CBL-interacting protein kinases (CIPKs) is required for full activity of CBL-CIPK complexes toward their target proteins. *J. Biol. Chem.* **2012**, *287*, 7956–7968. [[CrossRef](#)]
134. Mao, J.; Manik, S.M.; Shi, S.; Chao, J.; Jin, Y.; Wang, Q.; Liu, H. Mechanisms and Physiological Roles of the CBL-CIPK Networking System in *Arabidopsis thaliana*. *Genes* **2016**, *7*, 62. [[CrossRef](#)]
135. Lee, S.C.; Lan, W.Z.; Kim, B.G.; Li, L.; Cheong, Y.H.; Pandey, G.K.; Lu, G.; Buchanan, B.B.; Luan, S. A protein phosphorylation/dephosphorylation network regulates a plant potassium channel. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 15959–15964. [[CrossRef](#)]
136. Lee, S.C.; Lan, W.; Buchanan, B.B.; Luan, S. A protein kinase-phosphatase pair interacts with an ion channel to regulate ABA signaling in plant guard cells. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 21419–21424. [[CrossRef](#)]
137. Lee, S.C.; Luan, S. ABA signal transduction at the crossroad of biotic and abiotic stress responses. *Plant Cell Environ.* **2012**, *35*, 53–60. [[CrossRef](#)]
138. Umezawa, T.; Sugiyama, N.; Mizoguchi, M.; Hayashi, S.; Myouga, F.; Yamaguchi-Shinozaki, K.; Ishihama, Y.; Hirayama, T.; Shinozaki, K. Type 2C protein phosphatases directly regulate abscisic acid-activated protein kinases in *Arabidopsis*. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 17588–17593. [[CrossRef](#)]
139. Geiger, D.; Scherzer, S.; Mumm, P.; Marten, I.; Ache, P.; Matschi, S.; Liese, A.; Wellmann, C.; Al-Rasheid, K.A.; Grill, E.; et al. Guard cell anion channel SLAC1 is regulated by CDPK protein kinases with distinct Ca²⁺ affinities. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 8023–8028. [[CrossRef](#)]
140. Lan, W.Z.; Lee, S.C.; Che, Y.F.; Jiang, Y.Q.; Luan, S. Mechanistic analysis of AKT1 regulation by the CBL-CIPK-PP2CA interactions. *Mol. Plant* **2011**, *4*, 527–536. [[CrossRef](#)]
141. Scherzer, S.; Maierhofer, T.; Al-Rasheid, K.A.; Geiger, D.; Hedrich, R. Multiple calcium-dependent kinases modulate ABA-activated guard cell anion channels. *Mol. Plant* **2012**, *5*, 1409–1412. [[CrossRef](#)]
142. Ma, Y.; Szostkiewicz, I.; Korte, A.; Moes, D.; Yang, Y.; Christmann, A.; Grill, E. Regulators of PP2C phosphatase activity function as abscisic acid sensors. *Science* **2009**, *324*, 1064–1068. [[CrossRef](#)]
143. Park, S.Y.; Fung, P.; Nishimura, N.; Jensen, D.R.; Fujii, H.; Zhao, Y.; Lumba, S.; Santiago, J.; Rodrigues, A.; Chow, T.F.; et al. Abscisic acid inhibits type 2C protein phosphatases via the PYR/PYL family of START proteins. *Science* **2009**, *324*, 1068–1071. [[CrossRef](#)]
144. Geiger, D.; Scherzer, S.; Mumm, P.; Stange, A.; Marten, I.; Bauer, H.; Ache, P.; Matschi, S.; Liese, A.; Al-Rasheid, K.A.; et al. Activity of guard cell anion channel SLAC1 is controlled by drought-stress signaling kinase-phosphatase pair. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 21425–21430. [[CrossRef](#)]

145. Soon, F.F.; Ng, L.M.; Zhou, X.E.; West, G.M.; Kovach, A.; Tan, M.H.; Suino-Powell, K.M.; He, Y.; Xu, Y.; Chalmers, M.J.; et al. Molecular mimicry regulates ABA signaling by SnRK2 kinases and PP2C phosphatases. *Science* **2012**, *335*, 85–88. [[CrossRef](#)]
146. Acharya, B.R.; Jeon, B.W.; Zhang, W.; Assmann, S.M. Open Stomata 1 (OST1) is limiting in abscisic acid responses of Arabidopsis guard cells. *New Phytol.* **2013**, *200*, 1049–1063. [[CrossRef](#)]
147. Sato, A.; Sato, Y.; Fukao, Y.; Fujiwara, M.; Umezawa, T.; Shinozaki, K.; Hibi, T.; Taniguchi, M.; Miyake, H.; Goto, D.B.; et al. Threonine at position 306 of the KAT1 potassium channel is essential for channel activity and is a target site for ABA-activated SnRK2/OST1/SnRK2.6 protein kinase. *Biochem. J.* **2009**, *424*, 439–448. [[CrossRef](#)]
148. Yoshida, R.; Umezawa, T.; Mizoguchi, T.; Takahashi, S.; Takahashi, F.; Shinozaki, K. The regulatory domain of SRK2E/OST1/SnRK2.6 interacts with ABI1 and integrates abscisic acid (ABA) and osmotic stress signals controlling stomatal closure in Arabidopsis. *J. Biol. Chem.* **2006**, *281*, 5310–5318. [[CrossRef](#)]
149. Liu, S.; Lv, Z.; Liu, Y.; Li, L.; Zhang, L. Network analysis of ABA-dependent and ABA-independent drought responsive genes in Arabidopsis thaliana. *Genet. Mol. Biol.* **2018**, *41*, 624–637. [[CrossRef](#)]
150. Held, K.; Pascaud, F.; Eckert, C.; Gajdanowicz, P.; Hashimoto, K.; Corratge-Faillie, C.; Offenborn, J.N.; Lacombe, B.; Dreyer, I.; Thibaud, J.B.; et al. Calcium-dependent modulation and plasma membrane targeting of the AKT2 potassium channel by the CBL4/CIPK6 calcium sensor/protein kinase complex. *Cell Res.* **2011**, *21*, 1116–1130. [[CrossRef](#)]
151. Chen, L.; Wang, Q.Q.; Zhou, L.; Ren, F.; Li, D.D.; Li, X.B. Arabidopsis CBL-interacting protein kinase (CIPK6) is involved in plant response to salt/osmotic stress and ABA. *Mol. Biol. Rep.* **2013**, *40*, 4759–4767. [[CrossRef](#)]
152. Roy, S.J.; Huang, W.; Wang, X.J.; Evrard, A.; Schmockel, S.M.; Zafar, Z.U.; Tester, M. A novel protein kinase involved in Na(+) exclusion revealed from positional cloning. *Plant Cell Environ.* **2013**, *36*, 553–568. [[CrossRef](#)]
153. Xu, J.; Li, H.D.; Chen, L.Q.; Wang, Y.; Liu, L.L.; He, L.; Wu, W.H. A protein kinase, interacting with two calcineurin B-like proteins, regulates K⁺ transporter AKT1 in Arabidopsis. *Cell* **2006**, *125*, 1347–1360. [[CrossRef](#)]
154. Li, L.; Kim, B.G.; Cheong, Y.H.; Pandey, G.K.; Luan, S. A Ca²⁺ signaling pathway regulates a K⁺ channel for low-K response in Arabidopsis. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 12625–12630. [[CrossRef](#)]
155. Ragel, P.; Rodenas, R.; Garcia-Martin, E.; Andres, Z.; Villalta, I.; Nieves-Cordones, M.; Rivero, R.M.; Martinez, V.; Pardo, J.M.; Quintero, F.J.; et al. The CBL-Interacting Protein Kinase CIPK23 Regulates HAK5-Mediated High-Affinity K⁺ Uptake in Arabidopsis Roots. *Plant Physiol.* **2015**, *169*, 2863–2873.
156. Maierhofer, T.; Diekmann, M.; Offenborn, J.N.; Lind, C.; Bauer, H.; Hashimoto, K.; S Al-Rasheid, K.A.; Luan, S.; Kudla, J.; Geiger, D.; et al. Site- and kinase-specific phosphorylation-mediated activation of SLAC1, a guard cell anion channel stimulated by abscisic acid. *Sci. Signal.* **2014**, *7*, ra86. [[CrossRef](#)]
157. Ho, C.H.; Lin, S.H.; Hu, H.C.; Tsay, Y.F. CHL1 functions as a nitrate sensor in plants. *Cell* **2009**, *138*, 1184–1194. [[CrossRef](#)]
158. Grefen, C.; Karnik, R.; Larson, E.; Lefoulon, C.; Wang, Y.; Waghmare, S.; Zhang, B.; Hills, A.; Blatt, M.R. A vesicle-trafficking protein commandeers Kv channel voltage sensors for voltage-dependent secretion. *Nat. Plants* **2015**, *1*, 15108. [[CrossRef](#)]
159. Kudla, J.; Xu, Q.; Harter, K.; Gruissem, W.; Luan, S. Genes for calcineurin B-like proteins in Arabidopsis are differentially regulated by stress signals. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 4718–4723. [[CrossRef](#)]
160. Albrecht, V.; Weinl, S.; Blazevic, D.; D'Angelo, C.; Batistic, O.; Kolukisaoglu, U.; Bock, R.; Schulz, B.; Harter, K.; Kudla, J. The calcium sensor CBL1 integrates plant responses to abiotic stresses. *Plant J.* **2003**, *36*, 457–470. [[CrossRef](#)]
161. Guo, Y.; Xiong, L.; Song, C.P.; Gong, D.; Halfter, U.; Zhu, J.K. A calcium sensor and its interacting protein kinase are global regulators of abscisic acid signaling in Arabidopsis. *Dev. Cell* **2002**, *3*, 233–244. [[CrossRef](#)]
162. Cheong, Y.H.; Kim, K.N.; Pandey, G.K.; Gupta, R.; Grant, J.J.; Luan, S. CBL1, a calcium sensor that differentially regulates salt, drought, and cold responses in Arabidopsis. *Plant Cell* **2003**, *15*, 1833–1845. [[CrossRef](#)]
163. Corratge-Faillie, C.; Ronzier, E.; Sanchez, F.; Prado, K.; Kim, J.H.; Lanciano, S.; Leonhardt, N.; Lacombe, B.; Xiong, T.C. The Arabidopsis guard cell outward potassium channel GORK is regulated by CPK33. *FEBS Lett.* **2017**, *591*, 1982–1992. [[CrossRef](#)]

164. Quan, R.; Lin, H.; Mendoza, I.; Zhang, Y.; Cao, W.; Yang, Y.; Shang, M.; Chen, S.; Pardo, J.M.; Guo, Y. SCABP8/CBL10, a putative calcium sensor, interacts with the protein kinase SOS2 to protect Arabidopsis shoots from salt stress. *Plant Cell* **2007**, *19*, 1415–1431. [[CrossRef](#)]
165. Ren, X.L.; Qi, G.N.; Feng, H.Q.; Zhao, S.; Zhao, S.S.; Wang, Y.; Wu, W.H. Calcineurin B-like protein CBL10 directly interacts with AKT1 and modulates K⁺ homeostasis in Arabidopsis. *Plant J.* **2013**, *74*, 258–266. [[CrossRef](#)]
166. Mori, I.C.; Murata, Y.; Yang, Y.; Munemasa, S.; Wang, Y.F.; Andreoli, S.; Tiriach, H.; Alonso, J.M.; Harper, J.F.; Ecker, J.R.; et al. CDPKs CPK6 and CPK3 function in ABA regulation of guard cell S-type anion- and Ca(2+)-permeable channels and stomatal closure. *PLoS Biol.* **2006**, *4*, e327. [[CrossRef](#)]
167. Brandt, B.; Brodsky, D.E.; Xue, S.; Negi, J.; Iba, K.; Kangasjarvi, J.; Ghassemian, M.; Stephan, A.B.; Hu, H.; Schroeder, J.I. Reconstitution of abscisic acid activation of SLAC1 anion channel by CPK6 and OST1 kinases and branched ABI1 PP2C phosphatase action. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 10593–10598. [[CrossRef](#)]
168. Brandt, B.; Munemasa, S.; Wang, C.; Nguyen, D.; Yong, T.; Yang, P.G.; Poretsky, E.; Belknap, T.F.; Waadt, R.; Aleman, F.; et al. Calcium specificity signaling mechanisms in abscisic acid signal transduction in Arabidopsis guard cells. *eLife* **2015**, *4*. [[CrossRef](#)]
169. Xu, J.; Tian, Y.S.; Peng, R.H.; Xiong, A.S.; Zhu, B.; Jin, X.F.; Gao, F.; Fu, X.Y.; Hou, X.L.; Yao, Q.H. AtCPK6, a functionally redundant and positive regulator involved in salt/drought stress tolerance in Arabidopsis. *Planta* **2010**, *231*, 1251–1260. [[CrossRef](#)]
170. Ronzier, E.; Corratge-Faillie, C.; Sanchez, F.; Prado, K.; Briere, C.; Leonhardt, N.; Thibaud, J.B.; Xiong, T.C. CPK13, a noncanonical Ca²⁺-dependent protein kinase, specifically inhibits KAT2 and KAT1 shaker K⁺ channels and reduces stomatal opening. *Plant Physiol.* **2014**, *166*, 314–326. [[CrossRef](#)]
171. Van Kleeff, P.J.M.; Gao, J.; Mol, S.; Zwart, N.; Zhang, H.; Li, K.W.; de Boer, A.H. The Arabidopsis GORK K(+) channel is phosphorylated by calcium-dependent protein kinase 21 (CPK21), which in turn is activated by 14-3-3 proteins. *Plant Physiol. Biochem.* **2018**, *125*, 219–231. [[CrossRef](#)]
172. Franz, S.; Ehlert, B.; Liese, A.; Kurth, J.; Cazale, A.C.; Romeis, T. Calcium-dependent protein kinase CPK21 functions in abiotic stress response in Arabidopsis thaliana. *Mol. Plant* **2011**, *4*, 83–96. [[CrossRef](#)]
173. Li, C.L.; Wang, M.; Wu, X.M.; Chen, D.H.; Lv, H.J.; Shen, J.L.; Qiao, Z.; Zhang, W. THI1, a Thiamine Thiazole Synthase, Interacts with Ca²⁺-Dependent Protein Kinase CPK33 and Modulates the S-Type Anion Channels and Stomatal Closure in Arabidopsis. *Plant Physiol.* **2016**, *170*, 1090–1104. [[CrossRef](#)]
174. Lefoulon, C.; Boeglin, M.; Moreau, B.; Very, A.A.; Szponarski, W.; Dauzat, M.; Michard, E.; Gaillard, I.; Cherel, I. The Arabidopsis AtPP2CA Protein Phosphatase Inhibits the GORK K⁺ Efflux Channel and Exerts a Dominant Suppressive Effect on Phosphomimetic-activating Mutations. *J. Biol. Chem.* **2016**, *291*, 6521–6533. [[CrossRef](#)]
175. Vranova, E.; Tahtiharju, S.; Sriprang, R.; Willekens, H.; Heino, P.; Palva, E.T.; Inze, D.; Van Camp, W. The AKT3 potassium channel protein interacts with the AtPP2CA protein phosphatase 2C. *J. Exp. Bot.* **2001**, *52*, 181–182. [[CrossRef](#)]
176. Cherel, I.; Michard, E.; Platet, N.; Mouline, K.; Alcon, C.; Sentenac, H.; Thibaud, J.B. Physical and functional interaction of the Arabidopsis K(+) channel AKT2 and phosphatase AtPP2CA. *Plant Cell* **2002**, *14*, 1133–1146. [[CrossRef](#)]
177. Kuhn, J.M.; Boisson-Dernier, A.; Dizon, M.B.; Maktabi, M.H.; Schroeder, J.I. The protein phosphatase AtPP2CA negatively regulates abscisic acid signal transduction in Arabidopsis, and effects of abh1 on AtPP2CA mRNA. *Plant Physiol.* **2006**, *140*, 127–139. [[CrossRef](#)]
178. Yoshida, T.; Nishimura, N.; Kitahata, N.; Kuromori, T.; Ito, T.; Asami, T.; Shinozaki, K.; Hirayama, T. ABA-hypersensitive germination3 encodes a protein phosphatase 2C (AtPP2CA) that strongly regulates abscisic acid signaling during germination among Arabidopsis protein phosphatase 2Cs. *Plant Physiol.* **2006**, *140*, 115–126. [[CrossRef](#)]
179. Leyman, B.; Geelen, D.; Quintero, F.J.; Blatt, M.R. A tobacco syntaxin with a role in hormonal control of guard cell ion channels. *Science* **1999**, *283*, 537–540. [[CrossRef](#)]
180. Leyman, B.; Geelen, D.; Blatt, M.R. Localization and control of expression of Nt-Syr1, a tobacco SNARE protein. *Plant J.* **2000**, *24*, 369–381. [[CrossRef](#)]
181. Honsbein, A.; Sokolovski, S.; Grefen, C.; Campanoni, P.; Pratelli, R.; Paneque, M.; Chen, Z.; Johansson, I.; Blatt, M.R. A tripartite SNARE-K⁺ channel complex mediates in channel-dependent K⁺ nutrition in Arabidopsis. *Plant Cell* **2009**, *21*, 2859–2877. [[CrossRef](#)]

182. Eisenach, C.; Chen, Z.H.; Grefen, C.; Blatt, M.R. The trafficking protein SYP121 of Arabidopsis connects programmed stomatal closure and K(+) channel activity with vegetative growth. *Plant J.* **2012**, *69*, 241–251. [[CrossRef](#)]
183. Zhang, B.; Karnik, R.; Wang, Y.; Wallmeroth, N.; Blatt, M.R.; Grefen, C. The Arabidopsis R-SNARE VAMP721 Interacts with KAT1 and KC1 K+ Channels to Moderate K+ Current at the Plasma Membrane. *Plant Cell* **2015**, *27*, 1697–1717. [[CrossRef](#)]
184. Latz, A.; Becker, D.; Hekman, M.; Muller, T.; Beyhl, D.; Marten, I.; Eing, C.; Fischer, A.; Dunkel, M.; Bertl, A.; et al. TPK1, a Ca(2+)-regulated Arabidopsis vacuole two-pore K(+) channel is activated by 14-3-3 proteins. *Plant J.* **2007**, *52*, 449–459. [[CrossRef](#)]
185. Cheong, Y.H.; Pandey, G.K.; Grant, J.J.; Batistic, O.; Li, L.; Kim, B.G.; Lee, S.C.; Kudla, J.; Luan, S. Two calcineurin B-like calcium sensors, interacting with protein kinase CIPK23, regulate leaf transpiration and root potassium uptake in Arabidopsis. *Plant J.* **2007**, *52*, 223–239. [[CrossRef](#)]
186. Pandey, G.K.; Cheong, Y.H.; Kim, K.N.; Grant, J.J.; Li, L.; Hung, W.; D'Angelo, C.; Weinl, S.; Kudla, J.; Luan, S. The calcium sensor calcineurin B-like 9 modulates abscisic acid sensitivity and biosynthesis in Arabidopsis. *Plant Cell* **2004**, *16*, 1912–1924. [[CrossRef](#)]
187. Yang, W.; Kong, Z.; Omo-Ikerodah, E.; Xu, W.; Li, Q.; Xue, Y. Calcineurin B-like interacting protein kinase OsCIPK23 functions in pollination and drought stress responses in rice (*Oryza sativa* L.). *J. Genet. Genom.* **2008**, *35*, 531–543. [[CrossRef](#)]
188. Cui, X.Y.; Du, Y.T.; Fu, J.D.; Yu, T.F.; Wang, C.T.; Chen, M.; Chen, J.; Ma, Y.Z.; Xu, Z.S. Wheat CBL-interacting protein kinase 23 positively regulates drought stress and ABA responses. *BMC Plant Biol.* **2018**, *18*, 93. [[CrossRef](#)]
189. Zhu, K.; Chen, F.; Liu, J.; Chen, X.; Hewezi, T.; Cheng, Z.M. Evolution of an intron-poor cluster of the CIPK gene family and expression in response to drought stress in soybean. *Sci. Rep.* **2016**, *6*, 28225. [[CrossRef](#)]
190. Chen, L.; Ren, F.; Zhou, L.; Wang, Q.Q.; Zhong, H.; Li, X.B. The Brassica napus calcineurin B-Like 1/CBL-interacting protein kinase 6 (CBL1/CIPK6) component is involved in the plant response to abiotic stress and ABA signalling. *J. Exp. Bot.* **2012**, *63*, 6211–6222. [[CrossRef](#)]
191. Ishitani, M.; Liu, J.; Halfter, U.; Kim, C.S.; Shi, W.; Zhu, J.K. SOS3 function in plant salt tolerance requires N-myristoylation and calcium binding. *Plant Cell* **2000**, *12*, 1667–1678. [[CrossRef](#)]
192. Leran, S.; Edel, K.H.; Pervent, M.; Hashimoto, K.; Corratge-Faillie, C.; Offenborn, J.N.; Tillard, P.; Gojon, A.; Kudla, J.; Lacombe, B. Nitrate sensing and uptake in Arabidopsis are enhanced by ABI2, a phosphatase inactivated by the stress hormone abscisic acid. *Sci. Signal.* **2015**, *8*, ra43. [[CrossRef](#)]
193. Grefen, C.; Blatt, M.R. Do calcineurin B-like proteins interact independently of the serine threonine kinase CIPK23 with the K+ channel AKT1? Lessons learned from a menage a trois. *Plant Physiol.* **2012**, *159*, 915–919. [[CrossRef](#)]
194. Antoni, R.; Gonzalez-Guzman, M.; Rodriguez, L.; Rodrigues, A.; Pizzio, G.A.; Rodriguez, P.L. Selective inhibition of clade A phosphatases type 2C by PYR/PYL/RCAR abscisic acid receptors. *Plant Physiol.* **2012**, *158*, 970–980. [[CrossRef](#)]
195. Michard, E.; Dreyer, I.; Lacombe, B.; Sentenac, H.; Thibaud, J.B. Inward rectification of the AKT2 channel abolished by voltage-dependent phosphorylation. *Plant J.* **2005**, *44*, 783–797. [[CrossRef](#)]
196. Liu, J.; Zhu, J.K. An Arabidopsis mutant that requires increased calcium for potassium nutrition and salt tolerance. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 14960–14964. [[CrossRef](#)]
197. Sklodowski, K.; Riedelsberger, J.; Raddatz, N.; Riadi, G.; Caballero, J.; Cherel, I.; Schulze, W.; Graf, A.; Dreyer, I. The receptor-like pseudokinase MRH1 interacts with the voltage-gated potassium channel AKT2. *Sci. Rep.* **2017**, *7*, 44611. [[CrossRef](#)]
198. Yin, Y.; Adachi, Y.; Ye, W.; Hayashi, M.; Nakamura, Y.; Kinoshita, T.; Mori, I.C.; Murata, Y. Difference in abscisic acid perception mechanisms between closure induction and opening inhibition of stomata. *Plant Physiol.* **2013**, *163*, 600–610. [[CrossRef](#)]
199. Li, J.; Lee, Y.R.; Assmann, S.M. Guard cells possess a calcium-dependent protein kinase that phosphorylates the KAT1 potassium channel. *Plant Physiol.* **1998**, *116*, 785–795. [[CrossRef](#)]
200. Mori, I.C.; Uozumi, N.; Muto, S. Phosphorylation of the inward-rectifying potassium channel KAT1 by ABR kinase in *Vicia* guard cells. *Plant Cell Physiol.* **2000**, *41*, 850–856. [[CrossRef](#)]
201. Li, J.; Wang, X.Q.; Watson, M.B.; Assmann, S.M. Regulation of abscisic acid-induced stomatal closure and anion channels by guard cell AAPK kinase. *Science* **2000**, *287*, 300–303. [[CrossRef](#)]

202. Sottocornola, B.; Visconti, S.; Orsi, S.; Gazzarrini, S.; Giacometti, S.; Olivari, C.; Camoni, L.; Aducci, P.; Marra, M.; Abenavoli, A.; et al. The potassium channel KAT1 is activated by plant and animal 14-3-3 proteins. *J. Biol. Chem.* **2006**, *281*, 35735–35741. [[CrossRef](#)]
203. Kim, W.; Lee, Y.; Park, J.; Lee, N.; Choi, G. HONSU, a protein phosphatase 2C, regulates seed dormancy by inhibiting ABA signaling in Arabidopsis. *Plant Cell Physiol.* **2013**, *54*, 555–572. [[CrossRef](#)]
204. Lim, C.W.; Kim, J.H.; Baek, W.; Kim, B.S.; Lee, S.C. Functional roles of the protein phosphatase 2C, AtAIP1, in abscisic acid signaling and sugar tolerance in Arabidopsis. *Plant Sci.* **2012**, *187*, 83–88. [[CrossRef](#)]
205. Eisenach, C.; Papanatsiou, M.; Hillert, E.K.; Blatt, M.R. Clustering of the K⁺ channel GORK of Arabidopsis parallels its gating by extracellular K⁺. *Plant J.* **2014**, *78*, 203–214. [[CrossRef](#)]
206. Xi, Y.; Liu, J.; Dong, C.; Cheng, Z.M. The CBL and CIPK Gene Family in Grapevine (*Vitis vinifera*): Genome-Wide Analysis and Expression Profiles in Response to Various Abiotic Stresses. *Front. Plant Sci.* **2017**, *8*, 978. [[CrossRef](#)]
207. Boneh, U.; Biton, I.; Schwartz, A.; Ben-Ari, G. Characterization of the ABA signal transduction pathway in *Vitis vinifera*. *Plant Sci.* **2012**, *187*, 89–96. [[CrossRef](#)]
208. Wheeler, S.; Loveys, B.; Ford, C.; Davies, C. The relationship between the expression of abscisic acid biosynthesis genes, accumulation of abscisic acid and the promotion of *Vitis vinifera* L. berry ripening by abscisic acid. *Aust. J. Grape Wine Res.* **2009**, *15*, 195–204. [[CrossRef](#)]
209. Carbonell-Bejerano, P.; Santa Maria, E.; Torres-Perez, R.; Royo, C.; Lijavetzky, D.; Bravo, G.; Aguirreolea, J.; Sanchez-Diaz, M.; Antolin, M.C.; Martinez-Zapater, J.M. Thermotolerance responses in ripening berries of *Vitis vinifera* L. cv Muscat Hamburg. *Plant Cell Physiol.* **2013**, *54*, 1200–1216. [[CrossRef](#)]
210. Rienth, M.; Torregrosa, L.; Luchaire, N.; Chatbanyong, R.; Lecourieux, D.; Kelly, M.T.; Romieu, C. Day and night heat stress trigger different transcriptomic responses in green and ripening grapevine (*Vitis vinifera*) fruit. *BMC Plant Biol.* **2014**, *14*, 108. [[CrossRef](#)]
211. Pottosin, I.; Shabala, S. Polyamines control of cation transport across plant membranes: Implications for ion homeostasis and abiotic stress signaling. *Front. Plant Sci.* **2014**, *5*, 154. [[CrossRef](#)]
212. Wang, F.; Deng, S.; Ding, M.; Sun, J.; Wang, M.; Zhu, H.; Han, Y.; Shen, Z.; Jing, X.; Zhang, F.; et al. Overexpression of a poplar two-pore K⁺ channel enhances salinity tolerance in tobacco cells. *Plant Cell Tissue Organ Cult.* **2013**, *112*, 19–31. [[CrossRef](#)]
213. Sutter, J.U.; Campanoni, P.; Tyrrell, M.; Blatt, M.R. Selective mobility and sensitivity to SNAREs is exhibited by the Arabidopsis KAT1 K⁺ channel at the plasma membrane. *Plant Cell* **2006**, *18*, 935–954. [[CrossRef](#)]
214. Sutter, J.U.; Sieben, C.; Hartel, A.; Eisenach, C.; Thiel, G.; Blatt, M.R. Abscisic acid triggers the endocytosis of the Arabidopsis KAT1 K⁺ channel and its recycling to the plasma membrane. *Curr. Biol.* **2007**, *17*, 1396–1402. [[CrossRef](#)]
215. Hedrich, R.; Geiger, D. Biology of SLAC1-type anion channels - from nutrient uptake to stomatal closure. *New Phytol.* **2017**, *216*, 46–61. [[CrossRef](#)]
216. Singh, A.; Pandey, A.; Srivastava, A.K.; Tran, L.S.; Pandey, G.K. Plant protein phosphatases 2C: From genomic diversity to functional multiplicity and importance in stress management. *Crit. Rev. Biotechnol.* **2016**, *36*, 1023–1035. [[CrossRef](#)]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).