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Molecular Phylogeny, Classification and Evolution of Conopeptides

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Molecular phylogeny, classification and evolution of conopeptides

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Abstract

Conopeptides are toxins expressed in the venom duct of cone snails (*Conoidea*, *Conus*). These are mostly well-structured peptides and mini-proteins with high potency and selectivity for a broad range of cellular targets. In view of these properties, they are widely used as pharmacological tools and many are candidates for innovative drugs. The conopeptides are primarily classified into superfamilies according to their peptide signal sequence, a classification that is thought to reflect the evolution of the multigenic system. However, this hypothesis has never been thoroughly tested. Here we present a phylogenetic analysis of 1,364 conopeptide signal sequences extracted from GenBank. The results validate the current conopeptide superfamily classification, but also reveal several important new features. The so-called “cysteine-poor” conopeptides are revealed to be closely related to “cysteine-rich” conopeptides; with some of them sharing very similar signal sequences, suggesting that a distinction based on cysteine content and configuration is not phylogenetically relevant and does not reflect the evolutionary history of conopeptides. A given cysteine pattern or pharmacological activity can be found across different superfamilies. Furthermore, a few conopeptides from GenBank do not cluster in any of the known superfamilies, and could represent yet-undefined superfamilies. A clear phylogenetically-based classification should help to disentangle the diversity of conopeptides, and could also serve as a rationale to understand the evolution of the toxins in the numerous other species of conoideans and venomous animals at large.

Introduction

Cone snails of the genus *Conus* are predatory venomous marine mollusks feeding on fish, worm or snails. After decades of biological prospecting, conopeptides expressed in their venom duct have emerged as one of the richest and most promising marine sources of natural products (Blunt et al. 2012). The analysis of cone snail venoms has revealed a complex exogenome that is characterized by an extremely high level of diversity. With more than 600 described *Conus* species, each producing an estimated 100-200 venom components, the ensemble of cone snails were, until recently, estimated to produce between 50,000 and 100,000 different toxins (Menez et al. 2006; Olivera 2006). Recent studies, however, clearly demonstrate that this figure is an underestimation, probably by a factor of ten or so, with several new species described every year, more venom components detected in each sample using evolving technologies such as mass spectrometry (Biass et al. 2009; Ueberheide et al. 2009; unpublished results) and NextGen sequencing (Hu et al. 2011; Terrat et al. 2011) or combinations thereof (Violette et al. 2012), and marked intra-species and even intra-specimen variations in venom composition (Davis et al. 2009; Dutertre et al. 2010; Jakubowski et al. 2005). It is now estimated that the number of cone snail venom components exceeds one million.

An important characteristic of conopeptides, which makes them attractive for drug development is their high selectivity for molecular targets that span a broad range of therapeutic applications (Gayler et al. 2005; Leary et al. 2009; Molinski et al. 2009). So far, the conopeptide MVIIA (SNX-111, Prialt, or Ziconotide) from *Conus magus* (the magician cone) that selectively blocks Cav2.2 N-type voltage-gated calcium channels has been approved for the treatment of severe chronic pain (McGivern 2007; Miljanich 2004) and there are more promising drug candidates in the pipeline (*e.g.*, see Favreau et al. 2012; Han et al. 2008a; Lewis 2012). The potential of this rich source of pharmacological products has stimulated a race for the discovery of new toxins. From the traditional bioactivity-guided identification, lead discovery efforts have evolved towards modern

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3 structure-driven characterization (venom peptidomics and proteomics, venom gland transcriptomics,
4 targeted genomics, structure-function studies) and biocomputing-assisted analyses (proprietary
5 databases and bioinformatic tools) (Daly and Craik 2009; Favreau and Stöcklin 2009; Koua et al.
6 2012; Laht et al. 2011). In addition, phylogenetic approaches have recently emerged as an effective
7 way to quickly identify divergent lineages that are likely to have evolved with different functional
8 characteristics. This approach to identify these previously uncharacterized conopeptides is referred to
9 as concerted discovery (Conticello et al. 2001; Duda and Remigio 2008; Olivera 2006; Puillandre and
10 Holford 2010).

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22 However, despite the effectiveness of phylogenetic approaches in concerted discovery, the
23 technique is rarely used for the classification of conopeptides (but see Aguilar et al. 2009; Conticello
24 et al. 2001; Wang et al. 2008; Zhangsun et al. 2006). Several statistical methods for conopeptide
25 classification, such as Mahalanobis (Lin and Li 2007) or BLAST and Euclidian distances among
26 others (Mondal et al. 2006) have been described; however, most of these approaches are primarily
27 designed for classification of new sequences rather than for testing the current classification (*i.e.*,
28 checking the validity of each known group by a blind-exploratory approach). Conopeptide precursors
29 are characterized by a typical structural organization consisting of a highly conserved signal region,
30 followed by a more variable pro-region and a hyper-variable mature toxin containing a few
31 conserved amino acids such as the cysteine residues required for disulfide bonds. Conopeptides are
32 mainly named and classified according to three properties: first, they are characterized by their signal
33 sequence, this short sequence (~20 amino-acids) is highly conserved, and has been used to define
34 superfamilies; second, mature toxins structural families are characterized depending on their pattern
35 of cysteines (the Cys-pattern), for example, the mature toxin can include a variable number of
36 cysteines (most commonly 4 or 6), and their respective position can vary (4 cysteines can be
37 organized as C-C-C-C or CC-C-C where “-” represents a variable number of amino-acids); finally,
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3 several conopeptides have also been characterized according to their molecular targets, referred to
4 hereafter as “functional families”, and also previously termed “pharmacological families”.

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8 In a recent paper, Kaas *et al.* (2010) reviewed the structure, function and diversity of
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10 conopeptides on the ConoServer database (www.conoserver.org). In particular, they proposed that
11 “the ‘gene superfamily’ classification scheme focuses on evolutionary relationships between
12 conopeptides”, while the two other classification schemes (cysteine framework and function) do not.
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14 Their underlying hypothesis was that similarities in the Cys-pattern or function might have arisen by
15 convergence. While we fully agree with this statement, we also argue that it could serve as a rationale
16 to assess the congruence between the current gene superfamily classification and the evolution of the
17 corresponding multigenic system, and to accurately demonstrate that convergence phenomena are
18 common in conopeptide structure and function.
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29 Here we review the current superfamily classification of conopeptides by analysing all the
30 signal sequences available in GenBank using a phylogenetic approach to check: (i) if all the defined
31 superfamilies correspond to homogeneous groups; and (ii) if all the GenBank signal sequences
32 belong to a known superfamily. This study seeks to provide a "rationale" for a phylogenetic
33 classification of conopeptides and to clarify their current classification, thus complementing the work
34 initiated by Kaas *et al.* (2010).
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46 **Materials and Methods**

47 *1. Sequences from GenBank*

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49 Since the signal sequences used for phylogenetic analyses (see below), are only found on
50 complete nucleotide precursors and are not known for conopeptide discovered using proteomic
51 approaches, all the nucleotide sequences associated with the genus *Conus* were downloaded from
52 GenBank (www.ncbi.nlm.nih.gov). The sequences corresponding to non-coding regions, ribosomal
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3 genes, mitochondrial genes, and genes with a function that did not relate to toxin activity were
4 removed from the dataset, thus keeping only coding genes with a potential toxin activity. Only
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6 sequences obtained from *Conus* species belonging to the large major clade (Duda and Kohn 2005)
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8 were conserved, as a large number of the conopeptides found in species from other clades (*e.g.*, *C.*
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10 *californicus*) are highly divergent and do not match with any of the currently known superfamilies
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12 (Biggs et al. 2010; www.conoserver.org). Consequently, the classification in the present analysis is
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14 relevant only for conopeptides of the large major clade species. Conopeptide superfamilies are
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16 defined by a conserved signal sequence, thus we used the Signalp 3.0 server (Bendtsen et al. 2004) to
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18 identify the signal sequence; all sequences that did not include at least 50% of the signal region were
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20 removed, together with sequences including a stop codon. Only the signal region was used for
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22 phylogenetic analyses, as only this part of the conopeptides can be aligned within and, to some
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24 extent, between superfamilies.
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34 2. Phylogenetic analysis

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36 Aligning signal sequences between highly divergent conopeptides (*i.e.*, belonging to different
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38 superfamilies) is arduous, and homology hypotheses are doubtful. Thus, sequences were translated to
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40 amino acids and automatically aligned using two different algorithms: Muscle (Edgar 2004
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42 www.ebi.ac.uk/Tools/msa/muscle) and ClustalW (<http://clustalw.ddbj.nig.ac.jp/top-e.html>). Best
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44 model of evolution for these two datasets was selected using Modelgenerator V.85 (Keane et al.
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46 2006) following the corrected Akaike Information Criterion (with four discrete gamma categories)
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48 and used to reconstruct phylogenetic trees. The best model of evolution identified by Modelgenerator
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50 was JTT+G (Jones Taylor Thornton model, implemented under the name “Jones model” in MrBayes
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52 – Jones et al. 1992) for both datasets. Bayesian analyses were performed by running two parallel
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54 analyses in MrBayes (Huelsenbeck et al. 2001), each consisting of eight Markov chains of
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3 30,000,000 generations each with a sampling frequency of one tree every ten thousand generations.
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5 The number of swaps was set to 5, and the chain temperature at 0.02. A Neighbor-Joining tree
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7 obtained with MEGA5 (Tamura et al. 2011) was used as starting tree. Convergence of the parameters
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9 was evaluated using Tracer 1.4.1 (Rambaut and Drummond 2007), and analyses were terminated
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11 when ESS values were all superior to 200. A consensus tree was then calculated after omitting the
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13 first 25% trees as burn-in.
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17 As is the case for most multigenic families, the identification of an outgroup was highly
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19 problematic. No gene phylogenetically related to, and proven to be an outgroup for, conopeptides has
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21 been described. Furthermore, the use of toxins from other conoidean species was not possible, as it
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23 would require that the toxins from cone snails all arose from duplication events that took place after
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25 the divergence between the cone snails and other conoideans. Consequently, no outgroup was
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27 included in the analysis. This absence of an outgroup did not allow us to infer ancestor/descendant
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29 relationships.
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33 34 35 36 **Results**

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38 A total of 1,364 sequences potentially corresponding to conopeptides and with a signal
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40 sequence were downloaded from GenBank (performed on 1st of July, 2011). Alignments were 34 and
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42 30 amino-acids long with Muscle and Clustal W, respectively. To limit the time of calculation for
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44 phylogenetic analysis, only one sequence per amino-acid haplotype was kept; finally, 585 sequences
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46 were retained. Overall, the phylogenetic trees obtained from the Muscle and Clustal alignments were
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48 congruent; discrepancies were not supported (Posterior Probabilities < 0.90) and concerned
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50 phylogenetic relationships between the main clades and the position of a few highly divergent
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52 sequences (see details below). For clarity, only the phylogenetic tree based on the Clustal alignment
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54 is presented (Fig. 1), but results obtained from the Muscle alignment, when different, are discussed.
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3 Using information from GenBank and the literature, it was possible to link the clades defined
4 with the bayesian analysis to known superfamilies. Most of the defined superfamilies (A, D, I1, I2,
5 I3, J, L, O1, O3, P, S, T, V) corresponded to monophyletic groups, with some highly supported (Fig.
6 1). With the Muscle alignment, the O2 superfamily was included within the O1 superfamily; the
7 superfamily Y was represented by a single sequence, and corresponded to a unique lineage in the
8 tree. However, some superfamilies did not correspond to a monophyletic group, as they included
9 other conopeptides (*e.g.*, O2 included sequences of contryphans, and M included conomarphin – a
10 result already discussed by Han *et al.* 2008b). Several conopeptides from GenBank did not cluster in
11 any of the known superfamilies. These corresponded to known cysteine-poor conopeptides,
12 contulakin and conantokin, shown in Fig. 1 as the B and C superfamilies, respectively (the C
13 superfamily has been previously defined by Jimenez *et al.* (2007)); two conoCAP sequences
14 (FN868446.1 and FN868447.1 – named X1 in the Fig. 1 and appendix 1) described by Möller *et al.*
15 (2010); and sequences putatively annotated (FJ237364.1, named X2) or without annotation in
16 GenBank (DQ359922.1, EF493183.1/EF493184.1 and DQ359921.1, named respectively X3, X4 and
17 X5). In the Clustal alignment, two other groups of sequences, FJ375238.1/FJ375239.1/FJ375240.1
18 and EF208033.1 clustered in the superfamily A and O1, respectively with long branches, but
19 corresponded to independent lineages in the Muscle alignment (X6 and X7, respectively).

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Function and cysteine pattern were not clade-specific; conopeptides with the same function or
cysteine pattern were found in different clades. Additionally, sixteen new (*i.e.*, not numbered with
Roman numbers) cysteine patterns were identified; however, most of them certainly correspond to
anecdotic mutations of the canonical framework in a given family (*i.e.*, C-CC-C-C, C-C-CC-C-CC,
and C-CC-C-C, found in the O1 superfamily, differ from the pattern VI/VII by only one mutation),
while others may represent a new Cys-pattern number (*e.g.*, the Cys-pattern C-C-C-CC-C, found in

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3 the three members of the X6 group). The results are summarized in Table 1 (full details are provided
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5 in Appendix 1).
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8 Table 2 lists the number of conopeptides found in each superfamily and their distribution
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10 among the 71 *Conus* species. The superfamilies A, M and O1 were the largest, each containing at
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12 least 39 species, followed by the superfamilies T and I2. *Conus characteristicus*, *C. imperialis* and *C.*
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14 *litteratus* each express conopeptides belonging to more than 10 different superfamilies in their
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16 venom; however, it was difficult to know if this result reflects a higher conopeptide diversity in
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18 comparison to other species, or is due to a greater sampling effort in these species. All the
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20 superfamilies present in more than 10 *Conus* species (A, B, I2, M, O1, O2 and T) were found in
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22 mollusk, worm and fish-hunting species.
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29 Discussion

30 31 32 1. An updated classification of conopeptides

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34 Overall, the molecular phylogeny, based on more than 1,300 conopeptides signal sequences
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36 extracted from GenBank, strongly supports the current superfamily classification based on phenetic
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38 resemblances, as established in ConoServer. But, this relative congruency between phylogenetic and
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40 phenetic classifications is not surprising given the relative conservation of the signal sequence within
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42 superfamilies compared with between superfamilies, and the phylogenetic tree reflects these
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44 differences. However, the phylogenetic approach also revealed several new features, the most
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46 striking of which is the presence of deeply divergent lineages that, until now, were not included in the
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48 conotoxin superfamily classification. There are two main explanations for this result. First, the
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50 conopeptide superfamily classification reviewed by Kaas *et al.* (2010) includes only what is
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52 traditionally referred to as “cysteine-rich” conotoxins (*i.e.*, conopeptides with at least two disulfide
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54 bridges in the mature sequence as defined by Norton and Olivera in 2006), thus excluding the
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3 conopeptides with two cysteines and linear conopeptides also broadly present in the venom
4 (unpublished results). However, although the authors noted that “in future, all disulfide-poor
5 conopeptides will probably have to be attributed to a superfamily”, they refrained from doing so
6 because of the low number of cysteine-poor conopeptides with precursor sequences in ConoServer
7 (21). In GenBank we identified more than 50 such sequences and included them in the current
8 analysis. The signal sequences of cysteine-poor conopeptides do not cluster separately from the
9 conotoxins; some of them share highly similar signals with known superfamilies (contryphan with O2
10 and conomarphin with M), therefore, their exclusion from the superfamily classification is not
11 phylogenetically justified. We identified two additional superfamilies, B and C, for conantokins and
12 contulakins, respectively, one of which (C) has been proposed previously (Jimenez et al. 2007).
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14 Second, including non-annotated sequences from GenBank in the dataset helped to identify several
15 independent lineages in the tree (X1–X7). The level of divergence of their respective signal
16 sequences with the signals of other superfamilies was equivalent to the level of divergence between
17 known superfamilies, and they thus deserve recognition as new superfamilies. However, as these
18 independent lineages are represented by only one, two or three sequences, and because some of them
19 may not exhibit toxin activity (even if they were all found in venom ducts of cone snails), we
20 refrained from proposing new superfamily names, and only provided temporary names (X1-X7). It
21 should also be borne in mind that many other conopeptides have been described in the literature,
22 some of which have been given formal names (conkunitzin, conolysin, conomap, conophysin,
23 conopressin, conorfamide and conorphan). Because their signal sequences are not represented as
24 nucleotides in GenBank, they were not included in the analysis. However, a search in the protein
25 database of GenBank retrieved two complete precursors of Conkunitzin, with highly similar signal
26 sequences (P0C1X2.1 and P0CY85.1) and a local BLAST search (performed using BioEdit – Hall
27 1999) of the dataset used for the phylogenetic analyses revealed that the conkunitzin signals were
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3 unique, and probably represent a new superfamily. Finally, if most of the superfamily-level clades are
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5 highly supported, most of the inter-superfamily nodes are not, preventing any reliable conclusion
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7 concerning the phylogenetic relationships at this level.
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10 The original results presented herein raise several issues concerning the classification and
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12 nomenclature of the conopeptides and, more generally, of the genes that belong to multigenic
13
14 systems. The updated classification system we propose is based on a phylogenetic reconstruction that
15
16 guarantees the identification of sequences clusters that share a common ancestor. However, such
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18 phylogenetic trees cannot help in deciding which clades deserve a superfamily-level ranking and
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20 which ones do not. One common solution is to rely on a threshold of genetic distances, but the
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22 analyses of the genetic distances (calculated as the number of differences) between all the
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24 conopeptide signal sequences revealed that the distribution of genetic distances within superfamilies
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26 of conopeptides largely overlaps with the distribution of genetic distances between superfamilies
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28 (Fig. 2). This overlap can be linked to the high level of homoplasy found in conopeptides, making
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30 two conopeptides from different clades having, by chance, a relatively low genetic distance, or to the
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32 fact that two previously defined superfamilies would actually correspond to only one. This is the case
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34 of the L and I3 superfamilies, separated by genetic distances comprised between 0.38 and 0.69 that
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36 would, in most cases, correspond to within superfamily genetic distances.
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43 Consequently, it is not possible to rely only on a genetic threshold to define superfamilies for
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45 conotoxins. A threshold of 0.6, roughly corresponding to the gap between the two distributions of
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47 genetic distances (Fig. 2), would lead to the division of the M-superfamily into numerous
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49 superfamilies (indeed, Wang *et al.* 2008 proposed to divide the M superfamily in M1 and M2), and to
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51 the grouping of the superfamilies I1, I3 and L in a single one. However, our approach is aimed at
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53 offering a complementary guidance to help, in the future, deciding if a conotoxin or a group of
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55 conotoxins deserve a superfamily name: (i) since the minimum genetic distance between conotoxins
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3 is 0.32, this distance should be the minimum distance between the potential new conotoxin(s) and all
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5 the others; (ii) the new conotoxin(s) should correspond to an independent lineage, *i.e.* it should not
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7 cluster in any of the superfamily clades previously defined; (iii) the molecular target of the new
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9 conotoxin(s) should ideally be identified, to avoid naming conopeptides that would not be functional;
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11 (iv) the structure (cysteine pattern) and/or function should be different from the most closely related
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13 conotoxins in terms of genetic distances and/or phylogenetic relationships. All these criteria apply to
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15 the B and C superfamilies (genetic distances with other superfamilies > 0.3, these two lineages are
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17 independent and monophyletic, their molecular targets are identified – Mena et al. 1990, Craig et al.
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19 1999 –, and their cysteine framework are different from their respective sister-groups), justifying the
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21 attribution of new superfamily names. We followed the traditional nomenclature of conopeptide
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23 superfamilies, *i.e.* a Roman capital letter. As the number of Roman letter is limited, some
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25 superfamilies have been named with a Roman letter followed by an Arabic number (*e.g.* I1, I2, I3,
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27 O1, O2, O3) when several superfamilies share a common cysteine framework or molecular target.
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29 Because of the potentially high number of unknown superfamilies of conopeptides, we have no doubt
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31 that the nomenclature based on both Roman letters and Arabic numbers will become the reference
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33 rule.
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41 The first and fourth criteria also apply to the seven “X” lineages (Fig. 1), but the second
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43 applies to only 5 of them (two clustered within the A and O1 superfamilies with the muscle
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45 alignment) and the third to none of them. We propose to name such potential superfamilies of
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47 conopeptides that currently do not meet all the criteria but could in the future with the X Roman
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49 letter, followed by an Arabic number, waiting for either to be fully recognized as a separate
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51 superfamily or as belonging to an existing one.
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57 2. Evolution of the conopeptides

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3 The phylogenetic analysis clearly confirms that most of the defined superfamilies include
4 conopeptides with different cysteine frameworks and functions. Conversely, similar cysteine
5 frameworks and functions are found in different superfamilies, suggesting that a given cysteine
6 framework or function can appear several times independently, probably as a result of convergent
7 evolution. The multiple apparitions of the same framework and function during conotoxin evolution
8 are probably linked to the extremely rapid diversification of the genes. Several molecular
9 mechanisms have been proposed as being responsible for this high rate of diversification. Pi *et al.*
10 (2006) suggested that alternative splicing, unequal crossing-over or exon shuffling could explain this
11 diversity. Olivera *et al.* (1999) proposed two other mechanisms: the lack of a mismatch repair
12 system, at least in the hypervariable part of the sequence (the mature toxin); and recombination
13 mechanisms. Several other hypotheses, such as a high rate of duplication, followed by a strong
14 diversifying selection on the newly created gene copies that could lead to the rapid appearance of
15 several structurally and functionally highly divergent genes, have been also proposed and tested by
16 different authors (Duda and Palumbi 1999; Conticello *et al.* 2000; Duda and Palumbi 2000;
17 Conticello *et al.* 2001; Espiritu *et al.* 2001; Duda and Remigio 2008; Chang and Duda 2012). All
18 these molecular mechanisms, together with observed differences in the expression pattern between
19 species, maybe linked to episodes of gene silencing and reactivation (“Lazarotoxins”, Conticello *et al.*
20 2001; Duda and Palumbi 2004; Duda 2008), could favor the rapid diversification of *Conus*
21 species, by allowing them to envenomate and feed on new prey and thus colonize new niches (Duda
22 and Lee 2009).

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51 A phylogenetic approach could be very useful to identify divergent conopeptides with
52 potentially different functions, even if they share a common structural framework. For example, the
53 cysteine framework IV, found in the A-superfamily, is already linked to two different functions (α A -
54 Hopkins *et al.* 1995 and κ A - Craig *et al.* 1998). However, conotoxins, described by Conticello *et al.*
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3 in 2001, with the same framework, belong to the M superfamily, suggesting that these IV-conotoxins
4 that are structurally convergent with the IV-conotoxins in a different superfamily, could exhibit a
5 completely different function. A similar strategy could also apply within each superfamily, where not
6 only the signal sequence, but also the propeptide and mature regions can be aligned, and could reveal
7 divergent lineages with as yet uncharacterized functions (*e.g.*, see Aguilar et al. 2009; Puillandre et
8 al. 2010; Wang et al. 2008; Zhangsun et al. 2006).

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18 Furthermore, our identification of numerous new cysteine frameworks among the GenBank
19 sequences was also surprising. Even if some of them may be non-functional genes (pseudogenes),
20 others could correspond to novel protein structures. A few publications demonstrated that even toxins
21 with odd numbers of cysteines can be functional, for example with two 5-Cys toxins forming a
22 functional dimer or bioactive polymers of the 13-Cys “Con-ikot-ikot” peptide from *Conus striatus*
23 (Quinton et al. 2009, Walker et al. 2009). Our findings challenge the traditional view where
24 conotoxins are characterized by a limited number of cysteine frameworks: by exploring new
25 evolutionary pathways, the apparition of novel cysteine frameworks may also participate in the
26 hyper-diversification of the conotoxins. Additionally, this raises the question of the total number of
27 cysteine patterns one could expect to find among cone snail toxins. It is possible to predict the
28 theoretic number of cysteine patterns that could exist. If we limit the exercise to the 2, 4 and 6
29 cysteine patterns and exclude those with more than two consecutive cysteines, 20 different
30 frameworks can be proposed (C-C*, CC, CC-C-C*, CC-CC*, C-CC-C, C-C-CC*, C-C-C-C*, CC-
31 CC-CC, CC-CC-C-C, CC-C-CC-C, CC-C-C-CC*, CC-C-C-C-C*, C-CC-CC-C, C-CC-C-CC, C-CC-
32 C-C-C*, C-C-CC-CC, C-C-CC-C-C*, C-C-C-CC-C*, C-C-C-C-CC, C-C-C-C-C-C*). Ten of these
33 frameworks (marked with an *) can be found in GenBank. Given the extreme capacity of the
34 conopeptides to evolve and the apparent lack of evolutionary constraints (as illustrated by the
35 multiple apparitions of identical frameworks during their evolution), there is no reason that all these
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3 theoretical patterns will not be found in the future. It could be argued that mechanical constraints
4 would prevent the existence of some cysteine patterns; for example, it could be unfavorable to have a
5 disulfide bridge between two adjacent cysteines. However, despite this we found a short mature toxin
6 in the venom of one cone snail with a disulfide bridge between adjacent cysteines (unpublished
7 results). The peptide has been reproduced by protein synthesis, confirming this finding.
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18 3. *Conus* and *Conoidea* toxin diversity

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20 The diversity of conotoxins in the venom of several *Conus* species (Table 2) confirms that most
21 species are able to express a variety of conotoxins, as widely reported in literature (e.g., Olivera
22 2002). Furthermore, our results also suggest that *Conus* diet (fish, mollusk and worm) is not
23 correlated with differences in venom composition at the superfamily level. If differences exist, as
24 suggested in the literature (e.g., Conticello et al. 2001; Kaas et al. 2010), they most likely occur at the
25 species and intra-superfamily levels. Furthermore, phylogenetic analyses suggest that, at least, the
26 worm- and fish-hunting species are not monophyletic, as these two diets appeared independently
27 several times during the *Conus* evolution (Duda and Palumbi 2004; Espiritu et al. 2001; Kraus et al.
28 2011). Thus, differences in the venom composition should not be sought between the three diet
29 groups, but between the monophyletic clades defined within these three groups (Duda and Palumbi
30 2004).
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46 Diversity of the marine snail toxins is not limited to species included in the large major clade
47 of *Conus*. Recent analyses in other conoidean taxa suggest that toxin hyperdiversity is not the
48 privilege of the *Conus* large major clade. *C. californicus*, which is highly divergent from all the other
49 *Conus* species (Duda and Kohn 2005), showed a high diversity of toxins in its venom and several of
50 them were thought to correspond to new superfamilies (Biggs et al. 2010;
51 www.conoserver.org/?page=classification&type=genesuperfamilies). To a lesser extent, species in
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3 the small major clade of *Conus*, may also contain several novel conotoxins, as suggested by an
4 original Cys-pattern (XIII) found in the species *C. delessertii* (Aguilar et al. 2005). In addition to the
5 family Conidae, original toxins have already been reported in several other species of Conoidea, such
6 as *Polystira albida* (Lopez-Vera et al. 2004; Rojas et al. 2008), *Gemmula periscelida* (Lopez-Vera et
7 al. 2004), *G. speciosa*, *G. sogodensis*, *G. diomedea*, *G. kieneri* (Heralde et al. 2008), *Lophiotoma*
8 *olangoensis* (Watkins et al. 2006), *Terebra subulata* (Imperial et al. 2003), *Hastula hectica* (Imperial
9 et al. 2007) and *Crassispira cerithina* (Cabang et al. 2011). Furthermore, taxonomic surveys
10 (Bouchet et al. 2009) and phylogenetic analyses (Puillandre et al. 2011) suggest that the superfamily
11 Conoidea actually comprises a number of deeply divergent clades, whose species diversity is
12 currently largely underestimated. Presently, around 4,500 species have been described, but the group
13 is believed to include more than 10,000 species (Bouchet et al. 2009). Even if the venom apparatus
14 has been lost in several lineages of Conoidea (e.g., Fedosov 2007, Fedosov and Kantor 2008; Holford
15 et al. 2009; Medinskaya and Sysoev 2003), these findings suggest that the conotoxin diversity
16 characterized so far represents only a small part. If the level of diversity across all conoidean species
17 is similar to that found in those already investigated, the number of toxins produced by this single
18 superfamily could be as high as ten millions.

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Conflict of interest

The authors declare that they have no conflict of interest.

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Table 1: Number of sequences found in each superfamily, with list of cysteine patterns identified and known function in each superfamily.

Superfamily ID	# of sequences	ID	Cysteine pattern	# of sequences	Known function
A	153	I	CC-C-C	119	α , κ , ρ
		II	CCC-C-C-C	3	
		IV	CC-C-C-C-C	25	
		VI/VII	C-C-CC-C-C	1	
		XIV	C-C-C-C	3	
			C	1	
B	41		0	38	conantokin
			C-C	3	
C	4		0	1	contulakin
			C-C	3	
D	13	XX	C-CC-C-CC-C-C-C-C	5	α
			C-CC-C-CC-C-C-C	1	
			C-C-C-CC-C-C-C-C	7	
I1	6	XI	C-C-CC-CC-C-C	6	ι
I2	45	XI	C-C-CC-CC-C-C	35	κ
		XII	C-C-C-C-CC-C-C	9	
			C-C-CC-CC-C	1	
I3	7	XI	C-C-CC-CC-C-C	7	
J	12	XIV	C-C-C-C	12	$\alpha + \kappa$
L	4	XIV	C-C-C-C	3	α
			C-C-C	1	
M	193		0	3	α , κ , μ , conomarphin
		II	CCC-C-C-C	1	
		III	CC-C-C-CC	172	
		IV	CC-C-C-C-C	4	
		IX	C-C-C-C-C-C	1	
		XVI	C-C-CC	1	
		XIX	C-C-C-CCC-C-C-C-C	1	
			C	1	
			C-C	2	
			CC-C-C-C	1	
			CC-C-C-CC-C	2	
	C-CC-C-C-C	4			
O1	625	VI/VII	0	4	δ , κ , μ , ω
			C-C-CC-C-C	613	
			C-C-C	1	
			C-C-CC-C	1	
			C-CC-C-C	4	
			C-C-C-C-C	1	
	C-C-CC-C-CC	1			
O2	67	VI/VII	C-C-CC-C-C	51	γ , contryphan
			C-C	7	
		XV	C-C-CC-C-C-C-C	9	
O3	25	VI/VII	C-C-CC-C-C	25	bromosleeper
P	7	XIV	C-C-C-C	2	
			IX	C-C-C-C-C-C	
S	7	VIII	C-C-C-C-C-C-C-C-C	7	σ , α
T	140	X	0	12	ε , χ , τ
			CC-CXPC	4	

		V		CC-CC	121	
				C-C	2	
				CC-CCC	1	
	V	2	XV	C-C-CC-C-C-C-C	2	
	X1	2		C-C-C-C-C-C-C	2	conoCAP
	X2		III	CC-C-C-CC	1	
	X3	1		0	1	
	X4	2		C-C-C-C-C-C-C-C-C-C-C-C-C-C-C-C-C-C-C-C	2	
	X5	1	VIII	C-C-C-C-C-C-C-C-C	1	
	X6	3		C-C-C-CC-C	3	
	X7	1	VIII	C-C-C-C-C-C-C-C-C-C	1	
	Y	1	XVII	C-C-CC-C-CC-C	1	

Table 2: Number of conopeptides in each superfamily and species. Feeding types: F: Fish hunting species; M: Mollusc-hunting species; W: Worm-hunting species

Species	Prey	A	B	C	D	I1	I2	I3	J	L	M	O1	O2	O3	P	S	T	V	X1	X2	X3	X4	X5	X6	X7	Y	Occurrence
<i>achatinus</i>	F	4														1											2
<i>aurisiacus</i>	F	2									5	1															3
<i>bullatus</i>	F	4									10	8															3
<i>catus</i>	F	4									29																2
<i>circumciscus</i>	F	4									1	4															3
<i>consors</i>	F	8	1								3	10															4
<i>ermineus</i>	F	4									1	4															3
<i>geographus</i>	F	3	5	1							2	5					1	2									7
<i>lynceus</i>	F										1																1
<i>magus</i>	F	5									5	13	1														4
<i>monachus</i>	F	4										3															2
<i>obscurus</i>	F	3	5																								2
<i>ochroleucus</i>	F		2																								1
<i>parius</i>	F			1																							1
<i>purpurascens</i>	F	4									1	9						1									4
<i>radiatus</i>	F	6	3			2					4	3		1		1											7
<i>stercusmuscarum</i>	F	6									3	6															3
<i>striatus</i>	F	10					1				3	50					1										5
<i>striolatus</i>	F	1										8															2
<i>sulcatus</i>	F	8	2																								2
<i>tulipa</i>	F	3									2	2															3
<i>ammiralis</i>	M	2									5	3															3
<i>aulicus</i>	M	2									3	3															3
<i>aureus</i>	M										2																1
<i>bandanus</i>	M	2																									1
<i>dalli</i>	M										3	8															2
<i>episcopatus</i>	M	1				1					3	2															4
<i>gloriamaris</i>	M		2								3	6	1		1		4										6
<i>marmoreus</i>	M	4					2				14	15	5				12					2					7
<i>omaria</i>	M										4	6															2
<i>pennaceus</i>	M	4									6	13	3	2			16										6
<i>textile</i>	M	5					2				15	29	24		1	1	18										8
<i>victoriae</i>	M	3																									1
<i>abbreviatus</i>	W											98															1
<i>arenatus</i>	W		1								2	31		8			8										5
<i>aristophanes</i>	W											8															1
<i>betulinus</i>	W	8				1	1				10	1	2				1										7
<i>capitaneus</i>	W						1				1	2	2														4
<i>characteristicus</i>	W	2	5					3			6	3	1	1		3	5								1	1	11
<i>coronatus</i>	W										1	7															2
<i>distans</i>	W	1									2	5															3
<i>ebraeus</i>	W											29															1
<i>eburneus</i>	W	2	1	2			4		1	1	6	2		1													9

Figure caption

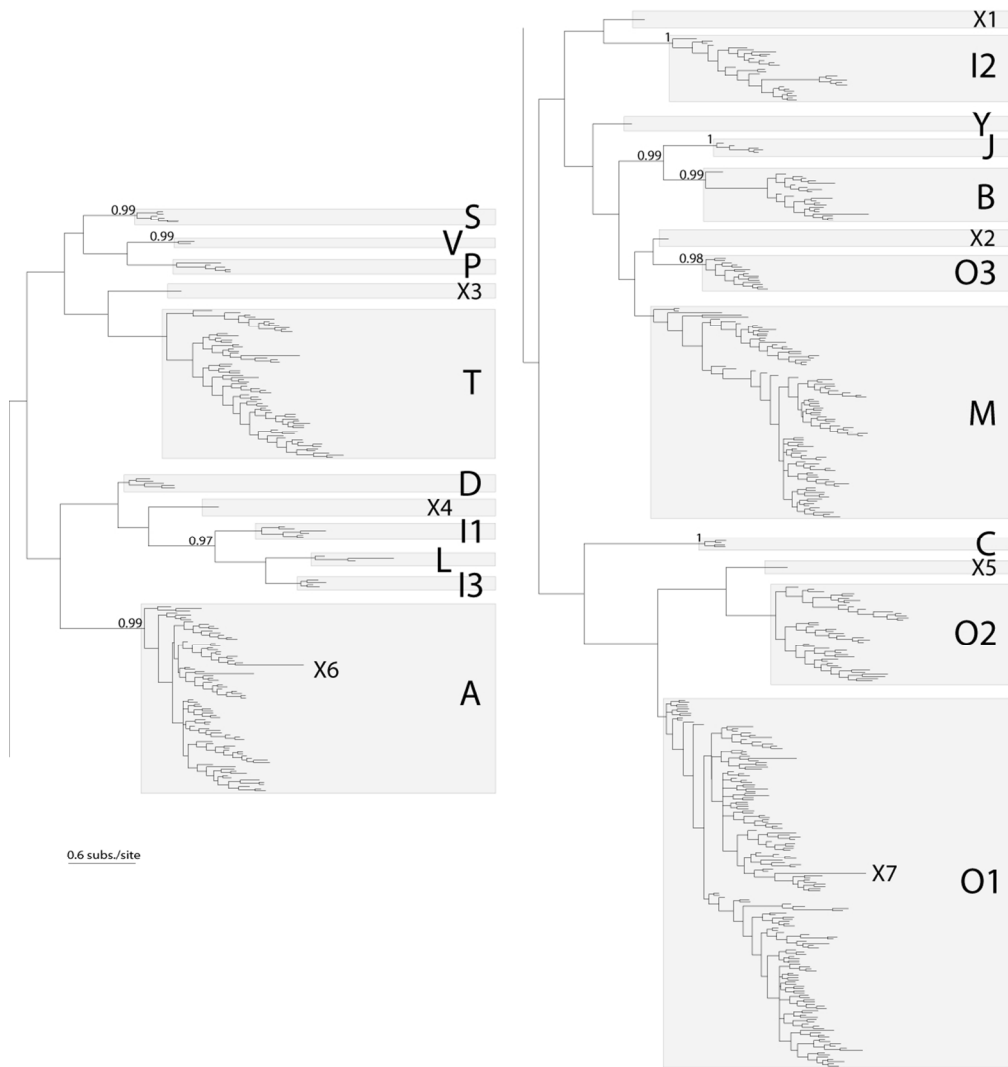
Figure 1: Bayesian phylogenetic tree (midpoint rooting) obtained from the Clustal alignment of the signal sequences of conopeptides from GenBank. Posterior Probabilities (when > 0.9) are provided for each node. Grey boxes are used to visualize the superfamilies. The B and C superfamilies respectively correspond to the contulakins and conantokins. The lineages X1-X7 potentially correspond to previously unrecognized superfamilies (see details in the text).

Figure 2: Pairwise distribution of genetic distances (p-distances) calculated with MEGA5 using the Clust alignment. Genetic distances between sequences from the same superfamily are shown in grey, genetic distances between sequences from different superfamily in black.

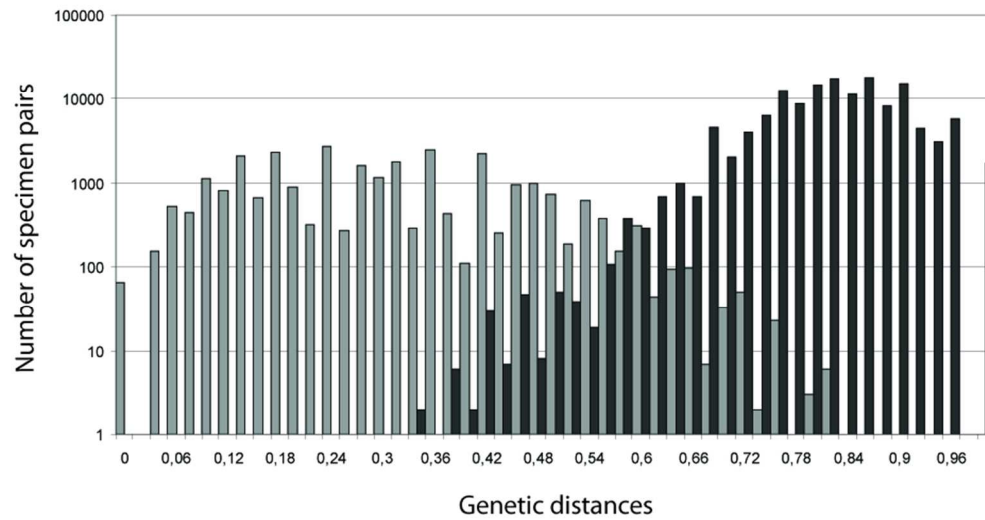
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3 Appendix 1: List of analysed sequences with superfamily assignation, GenBank numbers, Cys-
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5 pattern, species from which the sequence originated and corresponding feeding type F: Fish
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7 hunting species; M: Mollusc-hunting species; W: Worm-hunting species).
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For Peer Review

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205x218mm (150 x 150 DPI)



159x85mm (150 x 150 DPI)

Peer Review

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