

When behavioural geographic differentiation matters: inter-populational comparison of aggressiveness and group structure in the European perch

Lola Toomey, Martin Bláha, Emilie Mauduit, Tatyana Vanina, Margot Baratçabal, Yannick Ledore, Sami Vesala, Pascal Fontaine, Alain Pasquet,
Thomas Lecocq

▶ To cite this version:

Lola Toomey, Martin Bláha, Emilie Mauduit, Tatyana Vanina, Margot Baratçabal, et al.. When behavioural geographic differentiation matters: inter-populational comparison of aggressiveness and group structure in the European perch. Aquaculture International, 2019, 27 (5), pp.1177-1191. 10.1007/s10499-019-00343-z. hal-01989468

HAL Id: hal-01989468 https://hal.science/hal-01989468v1

Submitted on 25 Feb 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	When behavioural geographic differentiation matters: inter-
2	populational comparison of aggressiveness and group structure in
3	the European perch
4	
5	Original article can be found here:
6	https://doi.org/10.1007/s10499-019-00343-z
7	
8	
9	
10	
11	
12	
13	
14	
15	

When behavioural geographic differentiation matters: inter-

16

17

18

NANCY CEDEX FRANCE

34

populational comparison of aggressiveness and group structure in

the European perch

19 Short title: Behavioural geographic differentiation in Perca fluviatilis 20 Lola Toomey^{1*}, Martin Bláha², Emilie Mauduit¹, Tatyana Vanina², Margot Baratçabal¹, 21 Yannick Ledoré¹, Sami Vesala³, Pascal Fontaine¹, Alain Pasquet^{1, 4}, Thomas Lecocq^{1*} 22 ¹ Université de Lorraine, Inra, URAFPA, F-54000 Nancy, France 23 ² Faculty of Fisheries and Protection of Waters, South Bohemian Research Centre of 24 Aquaculture and Biodiversity of Hydrocenoses, University of South Bohemia in Ceske 25 Budejovice, Zátiší 728/II, CZ-389 25 Vodňany, Czech Republic 26 ³ Natural Resources Institute Finland, Helsinki, Finland 27 ⁴ CNRS (Centre national de la Recherche Scientifique) 28 29 * Corresponding Authors 30 31 lola.toomey@univ-lorraine.fr +33372745200 thomas.lecocq@univ-lorraine.fr +33372745696 32 University of Lorraine, Boulevard des Aiguillettes, BP 70239, 54506 VANDOEUVRE-LES-33

ABSTRACT

Domestication is still a long and difficult process and it is particularly impacted by species behavioural traits. Indeed, tolerance to high densities in intensive cultures and sociability are major features which facilitate domestication and influence the effectiveness of aquaculture production. Moreover, behavioural domestication predispositions could change at the intraspecific level. Here, we investigate three essential behavioural traits: aggressive interactions, group structure, and activity between three allopatric populations of *Perca fluviatilis*, a fish species at its nascent stage of production. We highlight inter-populational differences in group structure and aggressive interactions but not in activity. A more cohesive and homogeneous group structure was demonstrated for Finnish populations compared to Lake Geneva at 45-46 days post-hatching. In addition, Lake Geneva presented a higher aggressiveness. These inter-populational differences could be used in European perch aquaculture in order to improve production as well as welfare of individuals.

KEYWORDS

- 50 Aggression aquaculture behaviour cannibalism intraspecific differentiation larvae -
- *Perca fluviatilis* social structure

INTRODUCTION

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

Increasing the agriculture sustainability, including aquaculture, relies partly on the production and domestication of new species (Gepts et al. 2012). Domestication is considered as the process in which populations are bred in man-controlled environment and modified across successive generations from their wild ancestors in ways making them more useful to humans who control, increasingly during the process, their reproduction and food supply (Lecocq, 2018). However, domestication remains a difficult, long, and expensive process ridden by unfruitful outcomes, mostly due to zootechnical issues or taxon intrinsic features (Liao and Huang 2000; Diamond 2002; Teletchea and Fontaine 2014). This is particularly acute in intensive aquaculture (i.e. nowadays, intensive monoculture is the primary aquaculture) in which many new species domestication trials are hampered by several bottlenecks and end up being abandoned (Teletchea and Fontaine 2014). For instance, some fish species display low resistance to diseases or low food conversion efficiency, which impede or slow down their domestication (Liao and Huang 2000; Otton 2004). Conversely, other traits are favouring domestication such as fast growth rate and acceptance of artificial feeds, and consequently make taxon production an economically viable initiative (Liao and Huang 2000; Le François et al. 2010). Among these features, some behavioural traits are particularly essential since they can deeply facilitate domestication (Liao and Huang 2000; Jobling 2010; Le François et al. 2010) and subsequent aquaculture production (Huntingford et al. 2012).

Among behavioural traits, inter-individual relationships, group structure, and activity can affect directly the ability of a species to be domesticated and efficiently produced in intensive monoculture conditions. Tolerance to conspecifics in a limited area is an essential

parameter for production (Kristiansen et al. 2004) since it affects individual welfare (Huntingford 2004; Ashley 2007). Selecting populations which present an aggregative and cohesive group structure, therefore limiting stress, would be favouring welfare. However, living in group is not costless as it can trigger for instance competition for resources (Pitcher and Parrish 1993; Martins et al. 2012; Ward and Webster 2016). In culture conditions, this can lead to the emergence of aggressive behaviours (Damsgård and Huntingford 2012), such as attacks or bites, leading in some cases to cannibalism (Baras 2013). These aggressive behaviours have several potentially negative consequences in fish culture such as mortalities, stress, immune-suppression, or uneven competition for food (Damsgård and Huntingford 2012 and references therein). Cannibalism (type I: prey is caught tail first and ingested partially; type II: prey is caught by the head or tail and fully ingested; Baras et al. 2003; Baras 2013) is a major bottleneck in finfish aquaculture (Naumowicz et al. 2017) since it can lead to important losses (Baras et al. 2003; Huntingford et al. 2012). For example, cannibalism can cause up to 50% losses in Perca fluviatilis (Baras et al. 2003; Kestemont et al. 2003). Finally, activity is also an important factor in aquaculture as it contributes to the total energetic budget (e.g. up to 40% of Perca flavescens budget; Boisclair and Leggett 1989). Moreover, less active taxa could contribute to lower potential contacts and subsequent potential aggressive interactions. Therefore, in domestication processes, it is necessary to take into account the ability for taxa to present the most suitable group structure, low aggressive interaction rate, as well as lower activity. Yet, there is an intraspecific differentiation (differentiation between allopatric populations of conspecific individuals; Mayr 1963) which could further help to improve domestication processes.

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

Behavioural intraspecific differentiation and its potential for selection of founder populations have been poorly investigated to date. However, intraspecific differences in aggressive behaviour (Magurran and Seghers 1991; Mandiki et al. 2004; Bell 2004), time spent foraging in an open habitat (Bell 2004; Magnhagen 2006), schooling (Magurran and Seghers 1991), or boldness (Wright et al. 2003) have been already assessed for a few species (see also Foster 1999). Abiotic factors can influence social behaviours (e.g. temperature, light, population density; Baras et al. 2003; Kestemont et al. 2003), yet a genetic basis was also suggested since allopatric populations or geographically distinct strain differentiations were demonstrated for a few species (Amundsen et al. 1999; Damsgård and Huntingford 2012; Magnhagen et al. 2015). Therefore, considering such behavioural intraspecific differentiation could allow improving aquaculture for species for which production is still limited by behavioural bottlenecks.

The European perch, *Perca fluviatilis* L., is one of the fish species involved in the European aquaculture diversification (Kestemont et al 2015). Its long standing socio-economic interest (high market value and recreational interest) led to the development of its aquaculture in the 90's (Kestemont and Mélard 2000; Kestemont et al. 2015). However, its production is still limited due to several bottlenecks including some aspects related to fish behaviour such as aggressiveness and high cannibalism and subsequent mortalities (Kestemont et al. 2015). However, geographic differentiation has been previously observed for some of problematic behavioural traits (e.g. cannibalism rate, Mandiki et al. 2004). Therefore, we aim in this study at (i) assessing if European perch allopatric populations present differentiation for group structure and activity, as well as for aggressive interactions during first-life stages, and (ii) identifying populations presenting behavioural advantages for production.

MATERIAL AND METHODS

Rearing conditions

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

Rearing parameters were chosen according to trade-offs between abiotic culture conditions used in literature (e.g. Vlavonou 1996; Kestemont et al. 2003; Kestemont et al. 2015), our practices, and fish farming practices. The rearing protocol was tested and validated with a domesticated population from the fish farm "Lucas Perches" (Hampont, France) comparing growth and survival results to literature (e.g. Vlavonou 1996; Fiogbé and Kestemont 2003). Egg ribbons were obtained during the 2018 spawning season (May 2018) from lakes Geneva (GEN; Switzerland; 46°26'N, 6°33'E), Valkea-Müstajärvi (VAL; Finland; 61°13'08"N, 25°07'05"E), and Iso-Valkjärvi (ISO; Finland; 60°57'21"N, 26°13'3"E). After transport, 19 egg ribbons per lake were incubated at the Experimental Platform of Aquaculture (Unit of Animal Research and Functionality of Animal Products, University of Lorraine, Vandœuvrelès-Nancy, France) in incubators (110x64x186cm; one incubator per population), containing nine racks each (45x7x12cm), at 13°C. Each incubator had its own temperature control and recirculated water (flow rate of 4m³.h⁻¹) system and water was UV sterilised. Oxygen rate $(10.5 \pm 0.2 \text{mg.L}^{-1})$ and temperature $(13.0 \pm 0.3 ^{\circ}\text{C})$ were checked daily while pH was measured three times a week (8.0 \pm 0.1). Ammonium (lower than 0.05mg.L⁻¹) and nitrite concentrations were monitored three times a week until hatching (lower than 0.01mg.L⁻¹). Photoperiod was 12L:12D and light intensity was 400 lx at the water surface. Two independent experiments were performed in order to ensure availability of larvae

across the rearing period: experiment I from hatching until the end of weaning (26 days

post-hatching, dph) and experiment II from 27 dph until 60 dph. All populations were reared in independent structures.

Concerning experiment I, after hatching, larvae from the different egg ribbons were mixed and transferred to three green internal-wall 71L cylindro-conical tanks (three replicates per population; recirculated aquaculture system (RAS)) at a density of 50 larvae.L⁻¹. Temperature was gradually increased during two weeks to 20°C, photoperiod was 12L:12D and light intensity was 400lx. Larvae were fed with newly hatched *Artemia* naupli (Sep-Art, INVE) every 1h30 from 3 dph until weaning. At 16 dph, *Artemia* ration was decreased by 25% every three days and dry feed ration (BioMar, 300µm until 21 dph, then 500µm) was increased by the same ratio. After 25 dph, larvae were only fed with dry feed *ad libitum* (BioMar 500µm, then 700µm at 44 dph until the end of the experiment). At 26 dph, the larvae in cylindroconical tanks were removed to start experiment II.

For experiment II, larvae not used for experiment I were held after hatching in 2m³ tanks (RAS) under the same temperature, feeding, light intensity, and photoperiod regimes as individuals of the experiment I. At 27 dph, these larvae were transferred at a density of 19 larvae.L⁻¹ to the three cylindro-conical tanks in order to start experiment II. Light intensity was 80 lx at water surface, all else remaining equal to experiment I (except for density).

During the two experiments, oxygen concentration (8.7 \pm 2.3mg.L⁻¹) and temperature (20.0 \pm 0.6 °C) were checked daily for all tanks. Ammonium (0.14 \pm 0.1 mg.L⁻¹), pH (7.2 \pm 0.9 mg.L⁻¹), and nitrite concentrations (0.08 \pm 0.08 mg.L⁻¹) were monitored three times a week. Tanks were cleaned daily after first feeding and dead individuals were removed every morning. Survival rate for ISO, VAL and GEN were respectively 40.1 % (\pm 12.0), 29.4 % (\pm 14.5) and 6.6 % (\pm 3.4) for experiment I (26 dph; statistical difference between Geneva Lake and the two

Finnish populations; F=7.2, df=2, P<0.05) and 31.5 % (± 4.3), 28.0 % (± 11.1) and 37.4 (± 8.1) for experiment II (60 dph; no statistical difference between the three populations; F=0.98, df=2, P=0.42)). These ranges of survival rates were comparable to what is found in literature (e.g. Tamazouzt et al. 2000; Baras et al. 2003; Fiogbé and Kestemont 2003).

Group structure and activity

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

For each population, three replicates for each cylindro-conical tank were performed over two days (25 and 26 dph). At 24 and 25 dph, a total of 90 individuals (n=30 for each cylindroconical tank, 10 individuals per replicate) were sampled for each population and transferred to three aquaria (58 L; one aquarium per cylindro-conical tank; order of cylindro-conical replicates randomly assessed over two days; see appendix 1) with an 80lx light intensity and a temperature of 20.0°C (± 0.5). Individuals were not fed from the moment they were transferred to the beginning of the experiment the following day in order to have individuals in the same energetic state. After one night of acclimatisation, individuals were tested by groups of ten in circular arenas. Groups of ten individuals might not reflect faithfully what occurs in cylindro-conical tanks. However, evaluation cannot be performed directly in the tanks and this method was previously validated (e.g. Colchen et al. 2016). Three circular arenas (30 cm diameter with 1.5cm of water depth) were used to investigate group structure and activity (Colchen et al. 2016). Water in the arena was the same as in the aquaria, room temperature was maintained at 20.0 °C (± 0.6) and arenas were lit at 10lx from underneath in order to avoid shadows during recording. For each replicate, individuals were transferred from the aquarium to the arena with a beaker and a siphon. After 30min acclimatisation, individuals were filmed for 30min using camcorders (Sony, Handycam, DCR-SR72E) located 50cm above the arena. The three arenas were filmed simultaneously and the order of replicates tested was randomly assessed. After 1h, individuals were euthanised with an overdose of MS-222 following European rules and kept in formalin 4% for later length measurements. Larvae tested from ISO, VAL and GEN were respectively 14.05 ± 0.55 mm, 12.90 ± 0.62 mm, and 13.87 ± 0.26 mm. This full experiment was performed again during experiment II with fish sampled from cylindro-conical tanks at 44 and 45 dph. For this second test, individuals from ISO, VAL and GEN were respectively 26.74 ± 1.67 mm, 26.28 ± 1.99 mm, and 22.97 ± 1.08 mm (no statistical difference between populations for the two experiments; experiment I: F=0.712, df=2, P=0.528; experiment II: F=1.68, df=2, P=0.263).

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

Group structure analysis was performed using the ImageJ software. Images were extracted from videos at 3-min interval (11 images per video). From each image, exact coordinates of each individual were noted using the middle point between the eyes. Three parameters were evaluated to assess the group structure: the nearest neighbour distance, the mean of inter-individual distances, and the variance of these inter-individual distances (Buske and Gerlai 2011a). Nearest neighbour distance represents the distance between a focal fish and its closest neighbour and is an indicator of the group aggregation. The mean of interindividual distances corresponds to the mean of distances between a focal fish and all the other fish of the group and the average of values from all group members is an indicator of the group cohesion. Finally, the average of variances of inter-individual distances from each fish represents the homogeneity of distribution (Buske and Gerlai 2011a). Activity was also calculated in ImageJ. One image per second was extracted for six consecutive seconds every five minutes. Coordinates of each individual were noted for each image and then distance swam was calculated every second during the five seconds then averaged to obtain the mean distance swam for each individual per second.

All statistical analyses were performed in R 3.0.3 (R Core Team 2017). To test the normality of distributions, a Shapiro-Wilk test (R Core Team 2017) was used and homogeneity of variances was tested using the Levene test (Gastwirth et al. 2015). Then, linear mixed models were used with distances and activity as fixed factors and cylindro-conical tanks as random factor (Bates et al. 2004). There was no influence of the cylindro-conical tank on all models. Therefore, one-way analyses of variance (ANOVA F test) followed by Tukey post hoc tests were used to evaluate differences between populations (R Core Team 2017).

Quantification of aggressive interactions

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

Daily observations were carried out at different moments of the day previous to this experiment but did now allow identifying a cannibalism peak during the photophase. Therefore, we hypothesised that the beginning of the photophase would correspond to the cannibalism peak since individuals were not fed between 5.30pm and the next morning, that they were used to be fed during the photophase and that European perch is a visual predator (Graeb et al. 2005; Kestemont et al. 2008). Therefore, observations were performed after first feeding with a five-minute acclimatisation to the presence of the observer and five minutes of focal sampling (Colchen et al., 2019). Daily observations were carried out between 8.30am and 10am from 10 dph until the end of experiment I (26 dph). For experiment II, observations were carried out every three days (see Appendix 1). One replicate per population was observed per day and the same person performed all observations. Since hatching times were asynchronous between populations, the order of populations or tanks observed were randomly assigned. Several aggressive behaviours were noted: (i) Pursuit: an individual heads towards a conspecific, gets close and follows it when the conspecific moves; this involves a change of direction of the two individuals, (ii) Attack:

when an individual heads towards a congener and gets rapidly close to it without necessarily contact between the two individuals, (iii) Bite: when an individual catches with its mouth a part of a conspecific's body and then releases it, and (iv) Capture: when an individual ingests a part or the whole conspecific (type I and II cannibalisms). Taking into account all these aggressive interactions, a global daily aggressive interaction rate was calculated relatively to the initial number of individuals in the tank. Enucleation, being a specific indicator of aggressiveness in perch (Jourdan et al. 2000), was also evaluated by counting daily the number of dead individuals enucleated. Enucleation rate was calculated relatively to the initial number of individuals in the tank. For phase II, type II cannibalism rate was estimated by subtracting from the initial number of individuals the number of survivors and dead individuals over phase II and calculating a rate relatively to the initial number of individuals in the tank. Cannibalism rate could not be evaluated for experiment I as a precise monitoring of mortality was not possible the first week due to fast degradation of dead larvae. Finally, mortality rates attributed to cannibalism and enucleation were also calculated. In order to meet assumptions of normality (Shapiro-Wilk test, R Core Team 2017) and homogeneity of variances (Levene test, Gastwirth et al. 2015), data for all aggressive parameters was transformed (i.e. log(x+1)). One-way analyses of variance (ANOVA F test) followed by Tukey post hoc tests were used to evaluate differences between populations (R Core Team 2017). When assumptions were not respected (only for the aggressiveness rate during experiment I), Kruskal-Wallis H tests (R Core Team 2017) were used followed by Dunn post-hoc tests (Pohlert 2015).

Compliance with ethical standards

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

All along experimental procedures, individuals were handled as little as possible. All procedures were in accordance with the national and international guidelines for protection of animal welfare (Directive 2010/63/EU). This study was conducted with the approval Animal Care Committee of Lorraine (CELMA n°66) and the Ministry of Higher Education, Research, and Innovation (APAFIS13368-2018020511226118).

RESULTS

Group structure and activity

Experiment I (25-26 dph)

Inter-individual distances (F=7.8, df=2, P<0.05), variance of inter-individual distances (F=9.9, df=2, P<0.05), and activity (F=8.2, df=2, P<0.05) are significantly lower for GEN compared to VAL and ISO (Fig. 1). There is no statistical difference between VAL and ISO. There is no statistical difference between populations for the nearest neighbour distance (F=1.4, df=2, P=0.2; Fig. 1).

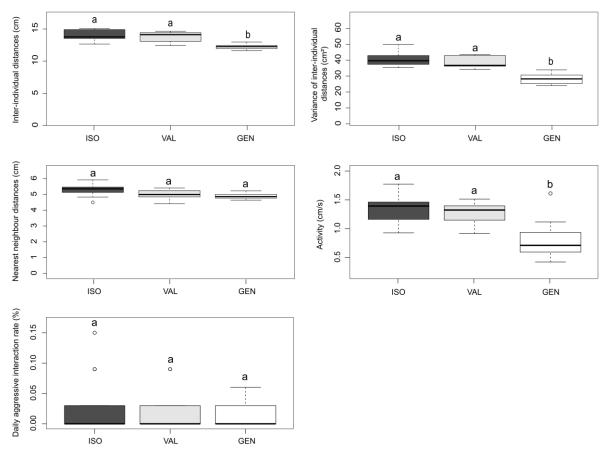


Fig. 1: Boxplots representing group structure and aggressive interaction results for experiment I (n=3330). Four measures of group structure are presented: inter-individual distances, variance of inter-individual distances, nearest neighbour distances, and activity. The black line represents the median, the outsider box corresponds to lower and upper quartile values and white dots correspond to most extreme values within 1.5 times the interquartile range from the ends of the box. Different letters indicate significant differences between populations (P < 0.05) using post hoc tests.

Experiment II (45-46 dph)

Inter-individual distances (F=7.8, df=2, P<0.05) and variance of inter-individual distances (F=9.9, df=2, P<0.05) are significantly higher for GEN compared to VAL and ISO (Fig. 2). There is no statistical difference between ISO and VAL. There is no statistical difference between populations for the nearest neighbour distance (F=1.4, df=2, P=0.3) and activity (F=1.2, df=2, P=0.3; Fig. 2).

Quantification of aggressive interactions

The first cases of cannibalism (type II only) were observed at 26 dph for ISO and at 51 dph for GEN. No cannibalism case was observed for VAL. There is no difference of daily aggressive interaction rate in experiment I (H=0.60, df=2, P=0.74; Fig. 1) but in experiment II these behaviours were significantly higher in GEN than ISO and VAL (F=7.21, df=2, P<0.05; Fig. 2). There is no statistical difference between ISO and VAL. Enucleation rate, which is null in experiment I for all populations, is significantly higher in GEN compared to VAL and ISO (F=70.74, df=2, P<0.05; Fig. 2) in experiment II. Cannibalism rate is not statistically different between populations (F=0.018, df=2, P=0.98; Fig. 2). Mortality rate attributed to cannibalism and enucleation is not statistically different between the three populations (F=0.69, df=2, P=0.59; Fig. 2).

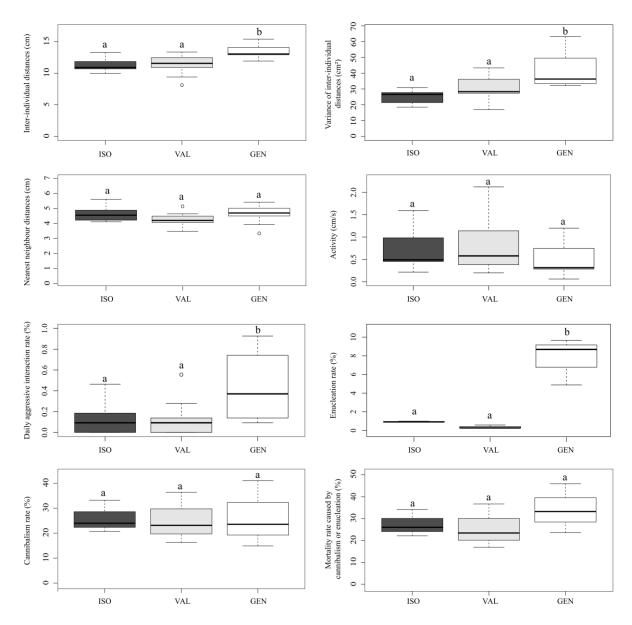


Fig.2: Boxplots representing group structure and aggressive interaction results for experiment II (n=1080). Four measures of group structure are presented: inter-individual distances, variance of inter-individual distances, nearest neighbour distances and activity. Aggressive interactions results include aggressive interaction rate, enucleation rate, cannibalism rate, and mortality rate caused by aggressive interactions. The black line represents the median, the outsider box corresponds to lower and upper quartile values and white dots correspond to most extreme values within 1.5 times the interquartile range from the ends of the box. Different letters indicate significant differences between populations (P < 0.05) using post hoc tests.

DISCUSSION

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

Inter-populational differentiation in behavioural traits and its potential causes

In this study, we highlight intraspecific differentiation between the two Finnish populations and GEN for group structure (experiments I and II) and aggressive interactions (only experiment II) while activity does not differ. We cannot exclude some potential biases in our experiments. For instance, (i) aggressive behaviour observations have been made after first feeding but cannibalism peaks (and aggressions) might occur at different moments of the day between populations and (ii) calculated cannibalism rate might also include dead individuals eaten by conspecifics which can blurry differences in cannibalism rates. However, since (i) temporal differentiation in cannibalism peaks has not been reported to date and (ii) the cannibalism estimation method, widely used across literature (e.g. Kestemont et al. 2003; Mandiki et al. 2004), allows to compare populations, we argue that bias related to observations of inter-populational behavioural differentiations at two ages are limited. Intraspecific differentiation in group structure and aggressive behaviours has been already highlighted for several fish species (Rosenau and McPhail 1987; Magurran and Seghers 1991; Amundsen et al. 1999; Lahti et al. 2001; Huizinga et al. 2009; Wark et al. 2011; Song et al. 2011). Here, a more cohesive and homogeneous structure is demonstrated for Finnish populations compared to GEN at 45-46 dph (Fig. 2). These results, associated with a similar nearest neighbour distance, indicate a structure in sub-groups in all populations but with a distance between these groups higher for GEN at 45 dph. The less homogeneous group structure of GEN at 45 dph is quite congruent with the higher aggressiveness highlighted for this population. Indeed, although daily aggressive interaction rate seems low (0.1-0.9 %; Fig. 2), the congruence between aggressive interaction patterns and enucleation rate supports

the higher aggressiveness of GEN compared to the two Finnish populations. The absence of difference in cannibalism rate indicates that aggressive interactions are not necessarily followed by type II cannibalism. Therefore, our study is not congruent with Mandiki et al. (2004) who showed a difference in intra-cohort cannibalism rate between different European perch allopatric populations (but with different populations than the ones investigated here). Here, we highlight differences in aggressive interactions aside from cannibalism rate. Inter-populational behavioural differences can be shaped by genetic differentiation, by phenotypic plasticity, or by their combination.

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

On the one hand, the observed inter-populational behavioural differences could be shaped by genetic differentiation. Indeed, population-specific demographic histories and potential local adaptations fostered by particular selective pressures can lead to the acquisition of distinct behavioural phenotypic traits or development rates between allopatric conspecific populations (Foster and Endler 1999; Foster 1999). For instance, it was shown a link between aggressiveness and the level of predation of the natural living site (Huntingford 1982; Magurran and Seghers 1991) as well as other environmental factors such as food availability and water current velocity (Lahti et al. 2001 and references therein). The occurrence of inherited differences in aggressive interactions was assessed for several species (Huntingford et al. 2012; Damsgård and Huntingford 2012). Similarly, inter-populational differences in activity were found to be connected to prey size distribution, total prey biomass, and water transparency (Boisclair and Leggett 1989). Unfortunately, we do not have enough information on the different lakes abiotic and biotic parameters to make any assessment. Another explanation of inter-populational differences could be divergences in development rates potentially triggered by genetic specificities. Indeed, we compare the populations at

the same age but we do not know if the compared fishes are at the same developmental stage (i.e. the lack of development table for larval and juvenile stages of *P. fluviatilis* prevent us to assess if the development is synchronous between populations). The populations might have divergent development rates, which can trigger inter-populational differences in parameters investigated. For instance, the higher aggressiveness in experiment II might be related to the development of muscular and nutritional structures through the larval stage (Kestemont et al. 1996; Vlavonou 1996). It can also be related to the development of visual structures since the visual acuity, essential for capture of prey, increases until metamorphosis (Guma'a 1982). In addition, the aggregation in sub-groups might be due to several factors such as kinship (Behrmann-Godel et al. 2006), the nature of interactions (e.g. aggressive interactions), spatial distribution, or differential sizes (Hinde 1976). Since these two last factors are sensitive to developmental stage, group structure is also influenced by development rate.

On the other hand, phenotypic plasticity (i.e. the ability of a genotype to produce more than one phenotype when exposed to different environments; Pigliucci et al. 2006; Kelly et al. 2012) is an alternative explanation of the observed behavioural differentiations between populations (DeWitt and Scheiner, 2004) with behaviour reflecting the strategy adopted under the influence of environmental factors. This was suggested as the driving factor for cannibalism in *P. fluviatilis* (Krol et al. 2015 and references therein) as well as in other species (e.g. Svenning and Borgstrøm 2005). Since we have used an experimental transplant approach (common environment), we speculate that we have minimized the effect of the environment (West-Eberhard 2003). Nevertheless, influences of past environmental conditions (i.e. before the beginning of our experiment) cannot be ruled out. On the one

hand, we have collected individuals at the egg stage in the wild and phenotypic response to environmental conditions could have occurred during development (Swain and Lindsey 1986). On the other hand, environmental conditions experienced by the parents might have influenced offspring phenotype (Mousseau and Fox 1998; Youngson and Whitelaw 2008). At last, it was also shown an influence of maternal size on larvae performance (Olin et al. 2012). Since we have no information on the parents of egg sampled, we cannot exclude the influence of maternal effects which were demonstrated in *P. fluviatilis* for other traits (Babiak et al. 2004; Krol et al. 2015).

Overall, we cannot assess the importance of genetic differentiation, phenotypic plasticity, and specific development rates in population-specific behaviour. Behavioural differentiation might be the result of the interaction of all factors (see for instance for cannibalism, Baras and Jobling 2002; Yang et al. 2015). Moreover, effects of experience on behaviour cannot be ruled out as it was previously suggested as an important factor for the behavioural variation (Hellström and Magnhagen 2011; Magnhagen 2015). Further analyses over several generations with populations under identical rearing conditions as well as the establishment of a development table for larval and juvenile stages of *P. fluviatilis* are needed to assess the importance of each factor in the geographic differentiation of behaviour.

Differences in group structure and aggressive interactions between the two different ages

For each population, we observe a behavioural differentiation between the two studied ages. Such a differentiation has been previously observed in other species. For instance, development of shoaling (increasing protection against predators, foraging efficiency, and mate encounters) with age through a decrease of inter-individual and nearest neighbour distances has already been observed in zebrafish (Buske and Gerlai 2011b; Buske and Gerlai

2012). We observe similar development for ISO and VAL populations. In contrast, the opposite pattern observed for GEN population is unexpected and corresponds to the establishment of a less homogeneous group with age. Mechanisms underlying the age-dependant changes in group structure are so far unknown. Differential ontogenies of sensory development might play a role into differences at the different ages (Buske and Gerlai 2012). Several neuroanatomical, physiological or biochemical factors have been suggested to be involved (Buske and Gerlai 2012). There might also be some group regulation mechanisms subsequent to weaning, which can potentially increase competition for resources linked for instance to bioenergetic needs or physiological shifts. This competition could explain the higher occurrence of aggressive interactions in experiment II.

Integrating inter-populational behaviour differentiation: a way to improve aquaculture production?

The variability occurring in behaviour at the intraspecific level offers the opportunity to select fishes whose behaviours make them more suitable for aquaculture production (Huntingford et al. 2012). In the European perch, the lack of population-specific activity tends to make this trait useless to select best population for aquaculture purpose. In contrast, difference in aggressiveness (i.e. and its consequences: losses due to aggressive interactions ranged from about 20% to 40% depending on the population; experiment II, Fig. 2) is a potential selection criterion for farmers since such interactions are highly detrimental for fish production. Similarly, the population-specific group structure should be considered as highly important information to highlight most suitable populations for intensive aquaculture. Based on our result, the more cohesive group structure and less aggressive interactions of Finnish *P. fluviatilis* make them the most suitable populations for

aquaculture. However, more populations need to be compared in order to identify populations of interest across the species range. Moreover, we cannot exclude the future potential impact of domestication since behavioural traits are modified by this process (Kohane and Parsons 1988). Yet, taking into account behavioural intraspecific differentiation would allow starting domestication program on populations presenting the best behavioural pre-disposition.

Selecting best populations for aquaculture production cannot be made through only behavioural trait comparisons. Indeed, selective breeding for low stress responsiveness has for instance been applied in several fish species but these low-stress response fish were also the ones which were more aggressive (Huntingford et al. 2012). Intraspecific differentiation has been already assessed for several other traits of interest such as growth (e.g. Mandiki et al. 2004; Leithner and Wanzenböck 2015), feed conversion efficiency (e.g. Imsland et al. 2000; Jonassen et al. 2000), or disease resistance (e.g. Imsland et al. 2002; Overturf et al. 2003). Therefore, the choice of the founder population must then be based on a multifunction and multi-traits approach rather than a single-trait decision framework.

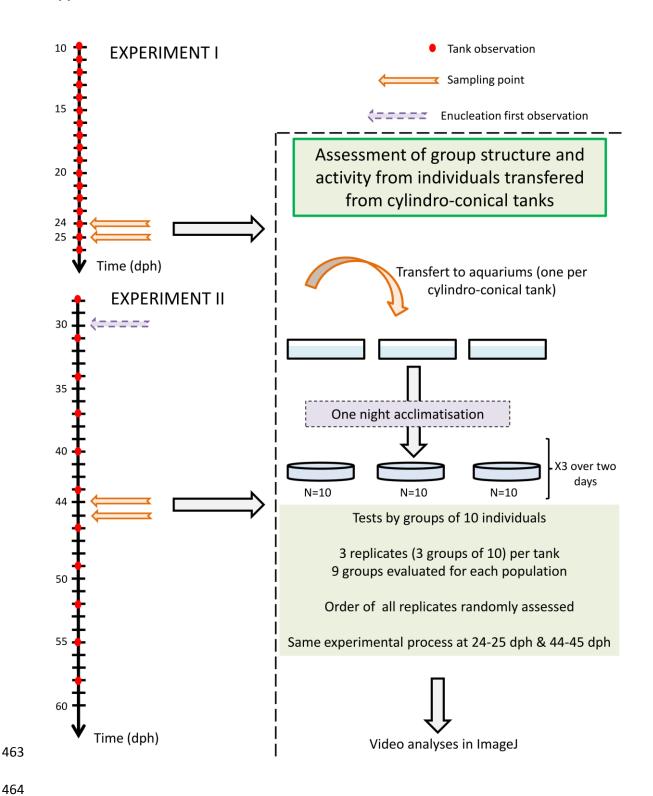
ACKNOWLEDGEMENTS

This project has received funding from the European Union's Horizon 2020 research and innovation programme under grant agreement No. 652831 (AQUAEXCEL²⁰²⁰). This output reflects only the author's view and the European Union cannot be held responsible for any use that may be made of the information contained therein. LT is supported by a grant from the French "Ministère de l'Enseignement Supérieur et de la Recherche". TV is supported by the Ministry of Education, Youth and Sports of the Czech Republic - projects CENAKVA (No.

443	CZ.1.05/2.1.00/01.0024), CENAKVA II (No. LO1205 under the NPU I program), GAJU project
444	(No. 060/2016/Z) and NAZV project (No. QJ 1510117). Authors acknowledge Laurent Espinat
445	and Chloé Goulon (INRA, Thonon-Les-Bains, France) for their help with egg ribbon collection
446	from Lake Geneva.
447	CONFLICT OF INTEREST AND ETHICAL STATEMENT
448	Conflict of Interest: The authors declare that they have no conflict of interest.
449	Ethical approval: "All applicable international, national, and/or institutional guidelines for
450	the care and use of animals were followed by the authors."
451	ABBREVIATIONS
452	GEN: Lake Geneva
453	VAL: Lake Valkea-Müstajärvi
454	ISO: Lake Iso-Valkjärvi
455	RAS: Recirculated Aquaculture System
456	
457	
458	
459	
460	

APPENDIX

Appendix 1: Material and methods workflow.



REFERENCES

466	Amundsen P-A, Svenning M-A, Slikavuoplo SI (1999) An experimental comparison of cannibalistic response in
467	different Arctic charr (Salvelinus alpinus (L.)) stocks. Ecol Freshw Fish 8:43–48. doi: 10.1111/j.1600-
468	0633.1999.tb00051.x
469	Ashley PJ (2007) Fish welfare: Current issues in aquaculture. Appl Anim Behav Sci 104:199–235. doi:
470	10.1016/j.applanim.2006.09.001
471	Babiak I, Mandiki SNM, Ratsinjomanana K, Kestemont P (2004) Initial weight and its variation in post-larval
472	Eurasian perch affect quantitative characteristics of juvenile cohorts under controlled conditions.
473	Aquaculture 234:263–276. doi: 10.1016/j.aquaculture.2003.12.010
474	Baras E (2013) Cannibalism in fish larvae: What have we learned? In: Qin JG (ed) Larval Fish Aquaculture. pp
475	167–199
476	Baras E, Jobling M (2002) Dynamics of intracohort cannibalism in cultured fish. Aquac Res 33:461–479. doi:
477	10.1046/j.1365-2109.2002.00732.x
478	Baras E, Kestemont P, Mélard C, Mélard C (2003) Effect of stocking density on the dynamics of cannibalism in
479	sibling larvae of <i>Perca fluviatilis</i> under controlled conditions. Aquaculture 219:241–255. doi:
480	10.1016/S0044-8486(02)00349-6
481	Bates D, Maechler M, Bolker B, Walker S (2004) lme4: Linear mixed-effects models using Eigen and S4
482	Behrmann-Godel J, Gerlach G, Eckmann R (2006) Kin and population recognition in sympatric Lake Constance
483	perch (Perca fluviatilis L.): can assortative shoaling drive population divergence? Behav Ecol Sociobiol
484	59:461–468. doi: 10.1007/s00265-005-0070-3
485	Bell AM (2004) Behavioural differences between individuals and two populations of stickleback (Gasterosteus
486	aculeatus). J Evol Biol 18:464–473. doi: 10.1111/j.1420-9101.2004.00817.x
487	Boisclair D, Leggett WC (1989) The Importance of Activity in Bioenergetics Models Applied to Actively Foraging
488	Fishes. Can J Fish Aquat Sci 46:1859–1867. doi: 10.1139/f89-234
489	Buske C, Gerlai R (2011a) Early embryonic ethanol exposure impairs shoaling and the dopaminergic and
490	serotoninergic systems in adult zebrafish. Neurotoxicol Teratol 33:698–707. doi:
491	10.1016/j.ntt.2011.05.009
492	Buske C, Gerlai R (2011b) Shoaling develops with age in Zebrafish (Danio rerio). Prog Neuropsychopharmacol

493	Biol Psychiatry 35:1409–1415. doi: 10.1016/j.pnpbp.2010.09.003
494	Buske C, Gerlai R (2012) Maturation of shoaling behavior is accompanied by changes in the dopaminergic and
495	serotoninergic systems in zebrafish. Dev Psychobiol 54:28–35. doi: 10.1002/dev.20571
496	Colchen T, Teletchea F, Fontaine P, Pasquet A (2016) Temperature modifies activity, inter-individual
497	relationships and group structure in fish. Curr Zool 1–9. doi: 10.1093/cz/zow048
498	Colchen T, Fontaine P, Ledoré Y, Teletchea F, Pasquet A (2019) Intra-cohort cannibalism in early life stages of
499	pikeperch. Aquac Res. In press. https://doi.org/10.1111/are.13966
500	Damsgård B, Huntingford F (2012) Fighting and Aggression. In: Aquaculture and Behavior. Wiley-Blackwell,
501	Oxford, UK, pp 248–285
502	DeWitt TJ, Scheiner SM (2004) Phenotypic variation from single genotypes: a primer. In Phenotypic Plasticity:
503	Functional and Conceptual Approaches. Oxford University Press, New York, NY
504	Diamond J (2002) Evolution, consequences and future of plant and animal domestication. Nature 418:700–707
505	doi: 10.1038/nature01019
506	Fiogbé ED, Kestemont P (2003) Optimum daily ration for Eurasian perch <i>Perca fluviatilis</i> L. reared at its
507	optimum growing temperature. Aquaculture 216:243–252. doi: 10.1016/S0044-8486(02)00409-X
508	Foster SA (1999) The geography of behaviour: an evolutionary perspective. Trends Ecol Evol 14:190–195.
509	Foster SA, Endler JA (1999) Geographic variation in behavior : perspectives on evolutionary mechanisms.
510	Oxford University Press
511	Gastwirth JL, Gel YR, Hui W, Lyubchich V, Miao W, Nogushi K (2015) Lawstat - Tools for Biostatistics, Public
512	Policy, and Law.
513	Gepts P, Famula TR, Bettinger RL, Brush SB, Damania AB, McGuire PE, Qualset CO (2012) Biodiversity in
514	Agriculture Domestication, Evolution, and Sustainability. Cambridge University Press, Davis, USA
515	Graeb BD, Galarowicz T, Wahl DH, Dettmers JM, Simpson MJ (2005) Foraging behavior, morphology, and life
516	history variation determine the ontogeny of piscivory in two closely related predators. Can J Fish Aquat
517	Sci 62:2010–2020. doi: 10.1139/f05-112
518	Guma'a SA (1982) Retinal development and retinomotor responses in perch, Perca fluviatilis L. J Fish Biol
519	20:611–618. doi: 10.1111/j.1095-8649.1982.tb03960.x
520	Hellström G, Magnhagen C (2011) The influence of experience on risk taking: results from a common-garden
521	experiment on populations of Eurasian perch. Behav Ecol Sociobiol 65:1917–1926. doi: 10.1007/s00265-

522	011-1201-7
523	Hinde RA (1976) Interactions, Relationships and Social Structure. Man 11:1–17. doi: 10.2307/2800384
524	Huizinga M, Ghalambor CK, Reznick DN (2009) The genetic and environmental basis of adaptive differences in
525	shoaling behaviour among populations of Trinidadian guppies, <i>Poecilia reticulata</i> . J Evol Biol 22:1860–
526	1866. doi: 10.1111/j.1420-9101.2009.01799.x
527	Huntingford FA (2004) Implications of domestication and rearing conditions for the behaviour of cultivated
528	fishes. J Fish Biol 65:122–142. doi: 10.1111/j.0022-1112.2004.00562.x
529	Huntingford FA (1982) Do inter- and intraspecific aggression vary in relation to predation pressure in
530	sticklebacks? Anim Behav 30:909–916. doi: 10.1016/S0003-3472(82)80165-6
531	Huntingford F, Jobling M, Kadri S, Wiley InterScience (Online service) (2012) Aquaculture and Behavior. John
532	Wiley & Sons
533	Imsland AK, Foss A, Naevdal G, Cross T, Bonga SW, Ham EV, Stefansson SO (2000) Countergradient variation in
534	growth and food conversion efficiency of juvenile turbot. J Fish Biol 57:1213–1226. doi:
535	10.1006/jfbi.2000.1384
536	Imsland AK, Jonassen TM, Langston A, Hoare R, Wergeland H, Fitzgerald R, Mulcahy M, Stefansson SO (2002)
537	The interrelation of growth and disease resistance of different populations of juvenile Atlantic halibut
538	(Hippoglossus hippoglossus L.). Aquaculture 204:167–177. doi: 10.1016/S0044-8486(01)00656-1
539	Jobling M (2010) Farmed species and their characteristics. In: Le François N, Jobling M, Carter C, Blier P (eds)
540	Finfish aquaculture diversification, CABI. pp 88–99
541	Jonassen TM, Imsland AK, Fitzgerald R, Bonga SW, Ham EV, Naevdal G, Stefánsson MO, Stefansson SO (2000)
542	Geographic variation in growth and food conversion efficiency of juvenile Atlantic halibut related to
543	latitude. J Fish Biol 56:279–294. doi: 10.1006/jfbi.1999.1159
544	Jourdan S, Fontaine P, Boujard T, Vandeloise E, Gardeur JN, Anthouard M, Kestemont P, De La Paix ND (2000)
545	Influence of daylength on growth, heterogeneity, gonad development, sexual steroid and thyroid levels,
546	and N and P budgets in <i>Perca fluviatilis</i> . Aquaculture 186:253–265.
547	Kelly SA, Panhuis TM, Stoehr AM (2012) Phenotypic Plasticity: Molecular Mechanisms and Adaptive
548	Significance. In: Comprehensive Physiology. John Wiley & Sons, Inc., Hoboken, NJ, USA, pp 1417–1439
549	Kestemont P, Dabrowski K, Summerfelt RC (2015) Biology and culture of percid fishes: Principles and practices.
550	Kestemont P, Jourdan S, Houbart M, Mélard C, Paspatis M, Fontaine P, Cuvier A, Kentouri M, Baras E (2003)

551	Size heterogeneity, cannibalism and competition in cultured predatory fish larvae: Biotic and abiotic
552	influences. Aquaculture 227:333–356. doi: 10.1016/S0044-8486(03)00513-1
553	Kestemont P, Mélard C (2000) Chap. 11 - Aquaculture. In: Craig JF (ed) Percid fishes. Systematics, ecology and
554	exploitation. Blackwell Science, Oxford, pp 191–224
555	Kestemont P, Mélard C, Fiogbe E, Vlavonou R, Masson G (1996) Nutritional and animal husbandry aspects of
556	rearing early life stages of Eurasian perch Perca fluviatilis. J Appl Ichthyol 12:157–165. doi:
557	10.1111/j.1439-0426.1996.tb00082.x
558	Kestemont P, Rougeot, Musil CJ, Toner D (2008) Larval and juvenile production. Farming Eurasian Perch Spec
559	Publ BIM 24:30–41.
560	Kohane MJ, Parsons PA (1988) Domestication: evolutionary change under stress. Evol Biol 23:31–48.
561	Kristiansen TS, Fernö A, Holm JC, Privitera L, Bakke S, Fosseidengen JE (2004) Swimming behaviour as an
562	indicator of low growth rate and impaired welfare in Atlantic halibut (Hippoglossus hippoglossus L.)
563	reared at three stocking densities. Aquaculture 230:137–151. doi: 10.1016/S0044-8486(03)00436-8
564	Krol J, Dauchot N, Mandiki SNM, Van Cutsem P, Kestemont P (2015) Cannibalism in cultured Eurasian perch,
565	Perca fluviatilis (Actinopterygii: Perciformes: Percidae) - Implication of maternal influence, kinship, and
566	sex ratio of progenies. Acta Ichthyol Piscat 45:65–73.
567	Lahti K, Laurila A, Enberg K, Piironen J (2001) Variation in aggressive behaviour and growth rate between
568	populations and migratory forms in the brown trout, Salmo trutta. Anim Behav 62:935–944. doi:
569	10.1006/anbe.2001.1821
570	Le François NR, Otton D, Werstink G (2010) Considerations for the Selection and Commercialization of New or
571	Alternate Species. In: Le François N, Jobling M, Carter C, Blier P (eds) Finfish Aquaculture Diversification.
572	CABI, pp 100–114
573	Lecocq T (2018) Insects: the disregarded domestication histories. In: Teletchea F (ed) Animal Domestication.
574	IntechOpen, https://doi.org/10.5772/intechopen.81834
575	Leithner S, Wanzenböck J (2015) Rearing Larvae of Different Strains of <i>Coregonus lavaretus</i> under Cold Water
576	Conditions: Comparison of a Special Cold-Water Line with a Standard Agglomerated Microdiet. J Agric Sc
577	7:28–36. doi: 10.5539/jas.v7n5p28
578	Liao IC, Huang YS (2000) Methodological approach used for the domestication of potential candidates for
579	aquaculture. Recent Adv Mediterr Aquac finfish species Diversif - Cah Options Méditerranéennes 47:97–

580	107.
581	Magnhagen C (2006) Risk-taking behaviour in foraging young-of-the-year perch varies with population size
582	structure. Oecologia. doi: 10.1007/s00442-005-0302-2
583	Magnhagen C (2015) Behaviour of Percid Fishes in the Wild and Its Relevance for Culture. In: Biology and
584	Culture of Percid Fishes. Springer Netherlands, Dordrecht, pp 399–416
585	Magnhagen C, Backström T, Øverli Ø, Winberg S, Nilsson J, Vindas MA, Brännäs E (2015) Behavioural responses
586	in a net restraint test predict interrenal reactivity in Arctic charr Salvelinus alpinus. J Fish Biol 87:88–99.
587	doi: 10.1111/jfb.12691
588	Magurran AE, Seghers BH (1991) Variation in Schooling and Aggression Amongst Guppy (Poecilia reticulata)
589	Populations in Trinidad. Behaviour 118:214–234. doi: 10.1163/156853991X00292
590	Mandiki SNM, Blanchard G, Mélard C, Koskela J, Kucharczyk D, Fontaine P, Kestemont P (2004) Effects of
591	geographic origin on growth and food intake in Eurasian perch (Perca fluviatilis L.) juveniles under
592	intensive culture conditions. Aquaculture 229:117–128. doi: 10.1016/S0044-8486(03)00359-4
593	Martins CIM, Galhardo L, Noble C, Damsgård B, Spedicato MT, Zupa W, Beauchaud M, Kulczykowska E,
594	Massabuau JC, Carter T, Planellas SR, Kristiansen T (2012) Behavioural indicators of welfare in farmed
595	fish. Fish Physiol Biochem 38:17–41. doi: 10.1007/s10695-011-9518-8
596	Mayr E (1963) Animal species and evolution. In: Animal species and their evolution, Belknap Pr. Harvard
597	University Press, Cambridge, Massachusetts, p 797
598	Mousseau TA, Fox CW (1998) The adaptive significance of maternal effects. Trends Ecol Evol 13:403–407. doi:
599	10.1016/S0169-5347(98)01472-4
600	Naumowicz K, Pajdak J, Terech-Majewska E, Szarek J (2017) Intracohort cannibalism and methods for its
601	mitigation in cultured freshwater fish. Rev Fish Biol Fish 27:193–208. doi: 10.1007/s11160-017-9465-2
602	Olin M, Jutila J, Lehtonen H, Vinni M, Ruuhijärvi J, Estlander S, Rask M, Kuparinen A, Lappalainen J (2012)
603	Importance of maternal size on the reproductive success of perch, Perca fluviatilis, in small forest lakes:
604	Implications for fisheries management. Fish Manag Ecol 19:363–374. doi: 10.1111/j.1365-
605	2400.2012.00845.x
606	Otton DD (2004) Establishment of an agribusiness model for assessing the commercial viability of new species
607	for aquaculture. PhD Thesis. University of Tasmania
608	Overturf K, Casten MT, LaPatra SL, Rexroad C, Hardy RW (2003) Comparison of growth performance,

609	immunological response and genetic diversity of five strains of rainbow trout (<i>Uncornynchus mykiss</i>).
610	Aquaculture 217:93–106. doi: 10.1016/S0044-8486(02)00014-5
611	Pigliucci M, Murren CJ, Schlichting CD (2006) Phenotypic plasticity and evolution by genetic assimilation. J Exp
612	Biol 209:2362-7. doi: 10.1242/jeb.02070
613	Pitcher TJ, Parrish JK (1993) Functions of Shoaling Behaviour in Teleosts. In: Pitcher TJ (ed) Behaviour of Teleost
614	Fishes. Chapman & Hall, London, pp 363–439
615	Pohlert T (2015) PMCMR: Calculate Pairwise Multiple Comparisons of Mean Rank Sums.
616	R Core Team (2017) R: A language and environment for statistical computing.
617	Rosenau ML, McPhail JD (1987) Inherited Differences in Agonistic Behavior between Two Populations of Coho
618	Salmon. Trans Am Fish Soc 116:646–654. doi: 10.1577/1548-8659(1987)116<646:IDIABB>2.0.CO;2
619	Song Z, Boenke MC, Rodd FH (2011) Interpopulation Differences in Shoaling Behaviour in Guppies (Poecilia
620	reticulata): Roles of Social Environment and Population Origin. Ethology 117:1009–1018. doi:
621	10.1111/j.1439-0310.2011.01952.x
622	Svenning M-A, Borgstrøm DR (2005) Cannibalism in Arctic charr: do all individuals have the same propensity to
623	be cannibals? J Fish Biol 66:957–965. doi: 10.1111/j.1095-8649.2005.00646.x
624	Swain DP, Lindsey CC (1986) Meristic variation in a clone of the cyprinodont fish Rivulus marmoratus related to
625	temperature history of the parents and of the embryos. Can J Zool 64:1444–1455. doi: 10.1139/z86-216
626	Tamazouzt L, Chatain B, Fontaine P (2000) Tank wall colour and light level affect growth and survival of
627	Eurasian perch larvae (<i>Perca fluviatilis</i> L.). Aquaculture 182:85–90. doi: 10.1016/S0044-8486(99)00244-6
628	Teletchea F, Fontaine P (2014) Levels of domestication in fish: Implications for the sustainable future of
629	aquaculture. Fish Fish 15:181–195. doi: 10.1111/faf.12006
630	Vlavonou RS (1996) Elevage expérimental de la perche Perca fluviatilis L. : développement larvaire et
631	croissance. PhD thesis, University of Lorraine
632	Ward A, Webster M (2016) Sociality: The Behaviour of Group-Living Animals. Springer International Publishing,
633	Cham
634	Wark AR, Greenwood AK, Taylor EM, Yoshida K, Peichel CL (2011) Heritable Differences in Schooling Behavior
635	among Threespine Stickleback Populations Revealed by a Novel Assay. PLoS One 6:e18316. doi:
636	10.1371/journal.pone.0018316
637	West-Eberhard MJ (2003) Developmental plasticity and evolution. Oxford University Press, New-York

638	Wright D, Rimmer LB, Pritchard VL, Krause J, Butlin RK (2003) Inter and intra-population variation in shoaling
639	and boldness in the zebrafish (Danio rerio). Naturwissenschaften 90:374–377. doi: 10.1007/s00114-003-
640	0443-2
641	Yang S, Yang K, Liu C, Sun J, Zhang F, Zhang X, Song Z (2015) To what extent is cannibalism genetically
642	controlled in fish? A case study in juvenile hybrid catfish Silurus meridionalis—asotus and the progenitors
643	Aquaculture 437:208–214. doi: 10.1016/J.AQUACULTURE.2014.12.005
644	Youngson NA, Whitelaw E (2008) Transgenerational Epigenetic Effects. Annu Rev Genomics Hum Genet 9:233-
645	257. doi: 10.1146/annurev.genom.9.081307.164445