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The behavior of nickel isotopes at the biogeochemical interface between ultramafic soils and Ni accumulator species

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Abstract

Ultramafic derived soils are characterized by low nutrient soils, a low Ca:Mg ratio, and high metal contents such as Ni, Co and Cr. The vegetation growing on these soils is highly adapted and includes both Ni hyperaccumulator and accumulator species. Today, approximately 530 Ni hyperaccumulator species are listed worldwide and the Ni concentration can be extremely high, e.g. up to 25% in latex from *Pycnanandra acuminata* (Sapotaceae), a tree found in New Caledonia. The aim of this study is to identify the potential role of Ni hyperaccumulator plants in the Ni biogeochemical cycle at the soil surface by using Ni isotopes. A set of Ni hyperaccumulator and Ni accumulator plants as well as topsoils were sampled on the Barro Alto and Niquelândia ultramafic complexes (Goiás State, Brazil). Three Ni hyperaccumulator plants were collected: *Justicia lanstyiakii*, *Heliotropium aff. salicoides*, *Cnidocolus aff. urens*, as well as one Ni accumulator plant, *Manihot* sp. The isotopic compositions of the whole plants were determined and compared to those of the bulk topsoils and DTPA-extractable Ni. The topsoils exhibited $\delta^{60}\text{Ni}$ values ranging from $-0.30 \pm 0.06\text{‰}$ to $0.16 \pm 0.05\text{‰}$. The DTPA-extractable Ni in the topsoils ranged from 94 to 623 mg kg^{-1} , i.e. 0.9–4.9% of the total soil Ni and was found to be isotopically heavier than the corresponding topsoil (from $-0.30 \pm 0.05\text{‰}$ to $0.34 \pm 0.08\text{‰}$). The $\delta^{60}\text{Ni}$ values for the Ni accumulator plants showed an enrichment in heavy Ni isotopes in the aerial parts of the plant compared to the roots, whereas similar $\delta^{60}\text{Ni}$ values for the roots, stems and aerial parts suggested that no significant fractionation results from Ni uptake and translocation in Ni hyperaccumulator plants. Moreover, the aerial parts (i.e. leaves and flowers) from all of the plants analyzed showed the highest Ni concentrations and the heaviest $\delta^{60}\text{Ni}$ values up to $1.21 \pm 0.05\text{‰}$. The enrichment in heavy Ni isotopes in the leaves ($0.09 \pm 0.06\text{‰} < \Delta^{60}\text{Ni}_{\text{leaves-soil}} < 1.06 \pm 0.03\text{‰}$) may result in a heavy Ni input in the litter during organic matter restitution. There is a non-negligible amount of Ni uptake by Ni accumulator and Ni hyperaccumulator plants and this may modify both the Ni isotope composition at the soil-plant interface and the overall cycle of Ni in surface soils

Highlights

► DTPA extractable Ni in topsoil is isotopically heavier than the total Ni pool. ► Ni-accumulators exhibit enrichment in Ni heavy isotopes in aerial parts. ► Similar $\delta^{60}\text{Ni}$ values for each plant compartment are observed for Ni-hyperaccumulators. ► Enrichment in heavy Ni isotopes in leaves may lead to a heavy Ni input in the litter.

Keywords : Nickel, Isotope, Ni hyperaccumulator species, Ultramafic environment

1. Introduction

Ultramafic (UM) soils are characterized by strong chemical peculiarities: low nutrient contents (N, P, K), a high Mg to Ca imbalance, and elevated contents of metals such as Fe, Mn, Ni, Co and Cr (e.g. Walker, 1954; Whittaker, 1954; Proctor and Woodell, 1975; Becquer et al., 2010; Echevarria, 2018). Therefore, the natural vegetation growing on UM soils is adapted to the “serpentinic syndrome” (Whittaker, 1954; Proctor and Woodell, 1975). Plants growing on UM soils can be either adapted to nutrient deficiencies and/or high metal contents or be (hyper)accumulators of metals. In their dry weight, Ni hyperaccumulator plants contain foliar tissue $> 1000 \mu\text{g g}^{-1}$ of Ni when growing in their natural habitat (Baker and Brooks, 1989; van der Ent et al., 2013; Reeves et al., 2017). This Ni concentration is 100 to 1000 times larger than the Ni concentration of plants grown on soil developed on non-UM rocks (Welch, 1981; Reeves et al., 2003; van der Ent et al., 2013).

Studies carried out over the past 40 years have focused on Ni (hyper)accumulators (e.g. Jaffré et al., 1976; Lee et al., 1977; Brooks et al., 1979; Reeves et al., 1983) to better understand the Ni accumulation phenomenon resulting from the stimulated uptake of Ni by the roots and an efficient transfer of Ni to the leaves through efficient xylem transport and phloem distribution (Centofanti et al., 2013; Tang et al., 2016; Deng et al., 2018), and to evaluate their phytomining and phytoremediation potential (e.g. Minguzzi and Vergnano, 1948; Reeves et al., 1996, 1999; Puschenreiter et al., 2005; Centofanti et al., 2012; van der Ent et al., 2015). In July 2017, the database listed 532 Ni hyperaccumulator species (52 families and 130 genera) (Reeves et al. 2018). To date, these species are found worldwide and most of them grow on tropical and subtropical ultramafic soils (van der Ent et al., 2013). For example, foliar Ni concentrations up to 7.6 wt% are found in the Ni hyperaccumulator species *Berkheya coddii* from South Africa (Mesjasz-Przybylowicz et al., 2004), whereas 25% Ni is found in latex from *Pycnandra acuminata*, a tree found in New Caledonia (Jaffré et al., 1976).

Therefore, the phenomenon of high Ni accumulation by plants on UM sites as reported by Reeves et al. (2018) raises the question of how the Ni biogeochemical cycle is modified or regulated by this biological process. The biogeochemical cycling of metals in both natural and contaminated systems

can be accessed via isotope fractionation studies. These studies may be able to clarify the contribution of the different processes such as sorption, dissolution, lattice incorporation, complexation or biological uptake (Weiss et al., 2008; Bullen, 2014; Wiederhold, 2015). In order to understand the fractionation mechanisms occurring in Ni cycling at the soil-plant interface, Ni isotope systematics have been introduced in studies focusing on both Ni (hyper)accumulator plants in hydroponic conditions and on plants collected in the field (Deng et al., 2014; Estrade et al., 2015; Tang et al., 2016). Deng et al. (2014) described the Ni and Zn isotopic fractionation during hydroponic plant growth. Ni hyperaccumulator plant growth (*Alyssum murale* and *Noccaea caerulescens*) is characterized by an enrichment in light Ni isotopes in the aerial parts of the plant, whereas heavy isotope enrichment is observed from the roots to the aerial parts for Ni non-accumulator plants (*Thlaspi arvense*). A field study on UM soils from Albania carried out by Estrade et al. (2015) reported the composition of the Ni isotopes for the subterranean and aerial organs of Ni hyperaccumulator plants (*Alyssum markgrafii*, *Alyssum murale* and *Alyssum bertolonii* subsp. *scutarinum*) and a non-hyperaccumulator plant (*Euphorbia spinosa*) collected in a Mediterranean climate. These authors suggest that the very efficient Ni uptake by Ni hyperaccumulators inhibited Ni isotope fractionation and uptake via the roots in soil solution enriched in heavy Ni isotopes and seems to control the Ni isotopic composition in the Ni hyperaccumulator plant. Conversely, an enrichment in light Ni isotopes was observed during Ni translocation from the roots to the leaves for the Ni non-accumulator plant (*Euphorbia spinosa*). Recently, Zelano et al. (2018) investigated the replenishment of the available Ni pools in soils by simulating litter decomposition using leaves from *Alyssum murale* and *Rinorea bengalensis* (hyperaccumulator plants). During the first days of leaching, the leaves released more than 80% of the total Ni content and the leached solution clearly displayed an enrichment in heavy isotopes for *Rinorea bengalensis*, while no isotope fractionation is observed for *Alyssum murale*. Therefore, the remaining litter could contain lighter Ni contents compared to the fresh leaves as also observed by Estrade et al. (2015). Consequently, these studies suggest that, in the upper soil horizons, the Ni cycle may be strongly affected by biological activity, particularly through the uptake of Ni in the roots of Ni hyperaccumulators, litterfall and litter recycling/biodegradation (Echevarria 2018).

The goal of this article is to determine the Ni isotope composition of Ni hyperaccumulator and Ni accumulator plants in two UM massifs located in Brazil in order to document the Ni isotope fractionation occurring in situ in plant-soil systems and to evaluate the impact of Ni uptake by Ni (hyper)accumulator plants on the Ni biogeochemical surface cycle. Within this framework, the Ni isotopic composition of the roots and aerial organs from several specimens of three Ni hyperaccumulator plants and one Ni accumulator plant collected in the UM massifs of Barro Alto and Niquelândia (Goiás state, Brazil) is completed with 19 topsoils taken from the same locations (Ratié et al., 2016).

2. Materials and Methods

2.1. Sampling and digestion

A set of Ni hyperaccumulator and Ni accumulator plants was sampled on the UM complex of Barro Alto and Niquelândia in 2013 and 2014 at different vegetative stages. The plant selection was based on the works of Reeves et al. (2007) and Raous et al. (2008), who identified close to 30 species of Ni (hyper)accumulator plants in Niquelândia and Barro Alto (Goiás State, Brazil). Three Ni hyperaccumulator species were sampled according to their abundance in the massif and they were identified by their flowering: *Justicia lanstykii* (four specimens), *Heliotropium aff. Salicoide* (six specimens), *Cnidocolus aff. urens* (three specimens), as well as one Ni accumulator species: *Manihot sp.* (four specimens) (Fig. 1). At each plant location, the bulk topsoil (0-5 cm) was also sampled, dried and sieved at 2 mm (Ratié et al., 2016).

The plant samples were carefully dry washed and rinsed with MilliQ® water in order to avoid soil contamination. They were then dried, finely cut and homogenized. Acid digestion was performed in a clean lab at the University of Brasilia with a microwave (Speedwave 4, Berghof). All of the reagents used in the digestion procedure were of analytical grade and bi-distilled. Approximately 300 mg of the samples was digested in Teflon bombs with 9 mL of concentrated HNO₃, 3 mL of HF, 2 mL of HCl and 1 mL of H₂O₂ under three successive conditions (150°C, 30 bars; 190°C, 35 bars; 50°C, 25 bars). The leachates were subsequently evaporated to dryness then taken up with 6 M HCl for the

chromatographic separation of Ni. The same acid digestion was also performed on two international standards. The measured Ni concentrations were $2.5 \pm 0.2 \text{ mg kg}^{-1}$ and $0.9 \pm 0.1 \text{ mg kg}^{-1}$ in NIST SRM 1570a Spinach Leaves and NIST SRM 1515 Apple Leaves, respectively, in agreement with the NIST publication.

The soil samples were homogenized and finely ground before acid digestion. Approximately 100 mg of the samples was digested in Teflon vessels with 5 mL of concentrated HF and 1.5 mL of HClO_4 at 180°C and then evaporated to dryness. They were subsequently digested in a mixture of concentrated HNO_3 -HCl (3.75 mL of HCl and 1.25 mL of HNO_3) at 150°C and evaporated to dryness. The samples were then taken up with 6 M HCl for chromatography separation of Ni (Ratié et al., 2015) and an aliquot were evaporated to dryness and taken up with 0.5 M HNO_3 to perform the chemical measurements.

2.2. Geochemical analysis

Measurements of the Ca, Fe, Mg, Ni, Ca, K, Mn and Na concentrations were performed by AAS (Varian, AA240FS, Fast Sequential Atomic Absorption Spectrometer, GEOPS-Université Paris Sud) The accuracy of the AAS measurements was controlled using standard solutions (EPL3, EPH3 and ESH2, SCP Science) and were always higher than 8% relative standard deviation (RSD) with respect to the certified values for the reference materials. N_{tot} and organic content (OC) measurements and the determination of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ were performed at GEOPS by EA-IRMS (VG SIRA 10).

Nickel availability was determined by extraction with 0.005 M DTPA + 0.01 M CaCl_2 buffered at pH 5.3 (L'Huillier and Edighoffer, 1996). Suspensions with 1:5 soil:solution ratio (w/v) were mixed end-over-end for 2 h, and subsequently centrifuged, filtered at $0.45 \mu\text{m}$ (Millipore®), and the extracts were analyzed with AAS to determine the Ni content. The extracts were evaporated and digested with concentrated acids using the same protocol used for the soil samples and analyzed for isotopic measurements after appropriate Ni purification.

2.3. Nickel isotopes

The Ni chemical purification procedure for the samples is based on a two-step chromatography separation that is fully described in Ratié et al. (2015). A first set of ion-exchange chromatography columns is filled with 2 mL (wet volume) of anionic resin AG1-X8 in 6 M HCl (BioRad 100-200 mesh). This resin retains Fe, Zn and a high amount of Co and Cu (Moynier et al., 2007) while Ni remains in the solution. Before the second chromatography column, a Ni double spike (^{61}Ni and ^{62}Ni) is added to the samples with a spike/natural ratio of 1 (Gueguen et al., 2013). The second set of ion-exchange chromatography columns use a specific Ni-resin (Eichrom) composed of polymethacrylate containing a dimethylglyoxime (DMG) molecule that retains Ni onto the resin as an insoluble Ni-DMG complex at pH 8-9. The eluted Ni solution is evaporated and taken up in HNO_3 2%.

The Ni isotope ratios were measured with a Neptune (Thermo-Electron) MC-ICP-MS at the Pôle Spectrométrie Océan (PSO) at IFREMER (Centre de Brest, France). The samples and standards were introduced via an ApexQ (50-75 V per $\mu\text{g/mL}$) in HNO_3 2%. A single “run” consisted of one block of 40 measurements. During the measurement, the Ni concentration (spike + natural) in the sample is 200 $\mu\text{g/L}$ ($^{61}\text{Ni}/^{62}\text{Ni} = 1.1004$ with a total Ni concentration of 100 mg/L ; Gueguen et al. (2013). The double-spike calculation procedure was based upon the method described by Siebert et al. (2001) for the determination of Mo isotopes. This method consists of determining the corrected isotopic ratio and instrumental mass bias through iterative calculations (Albarede and Beard, 2004; Quitté and Oberli, 2006; Cameron et al., 2009). In addition, each sample analysis was bracketed by the measurements of the spiked standard Ni NIST SRM 986 solutions at the same concentration and same spike/standard ratio as the samples. The $\delta^{60}\text{Ni}$ ratios were expressed in per mil (‰) and normalized to the average value of the bracketing standard SRM-986 (Eq. 1) (Gramlich et al., 1989).

$$\delta^{60}\text{Ni} = \left(\frac{\left(\frac{\delta^{60}\text{Ni}}{\delta^{58}\text{Ni}} \right)_{\text{sple}}}{\left(\frac{\delta^{60}\text{Ni}}{\delta^{58}\text{Ni}} \right)_{\text{SRM-986}}} - 1 \right) \times 1000 \text{ (Eq. 1)}$$

The long-term analytical sample reproducibility of the Ni standard NIST SRM 986 was $\pm 0.05\text{‰}$ (2 SD, $n = 320$). The average $\delta^{60}\text{Ni}$ reproducibility of the sample after the full chemical procedure

(purification and measurement) was determined by measuring two samples four times each. This reproducibility was better than 0.03%. Although each sample was measured three to four times, we only report the average values and their 2 SD values calculated from the replicate measurements (in the figures and tables). Note that a 2 SD value of 0.05‰ (external reproducibility) was reported when the calculated 2 SD was less than 0.05‰. A procedural blank sample was included within each batch of samples. The average amount of Ni for the blanks was 17 ng ($n = 8$), which is negligible relative to the amount of Ni that was processed for each sample (usually 8-10 μg of Ni).

3. Results & Discussion

3.1. Soil chemistry

3.1.1. Bulk composition

The main characteristics of the UM soils of Barro Alto and Niquelândia are detailed in Ratié et al. (2015) and (2016) and the chemical and Ni isotope compositions are summarized in Table 1. The soil samples are rich in metals such as Fe ($378 \pm 77 \text{ g kg}^{-1}$), Ni ($10.2 \pm 3.3 \text{ g kg}^{-1}$) and Mg ($16.4 \pm 8.1 \text{ g kg}^{-1}$) compared to Ca ($2.9 \pm 2.9 \text{ g kg}^{-1}$), and very poor in K ($0.1 \pm 0.1 \text{ g kg}^{-1}$). The soil chemistry is in accordance with other soils analyzed in tropical UM environments, such as in Brazil, Cuba and New Caledonia (Reeves et al., 1999, 2007; Becquer et al., 2001; Massoura et al., 2006; Garnier et al., 2009). In comparison with other studies on Ni isotopes in soils and plants, the Ni contents in soils are significantly higher than those of the UM soils studied by Estrade et al. (2015) ($1.4\text{-}3.8 \text{ g kg}^{-1}$) and Pędziwiatr et al. (2018) ($0.82\text{-}2.1 \text{ g kg}^{-1}$). The soil mineralogy is dominated by iron oxide (goethite, hematite), spinel (i.e., chromite and/or magnetite) and quartz (Garnier et al., 2006, 2009; Ratié et al., 2015). In non-serpentinized UM soils developed in humid tropical conditions, the main Ni-bearing phases are goethite and Mn-oxides (Echevarria et al., 2006). Nickel is mainly hosted by goethite in the soils of Barro Alto and Niquelândia. Moreover, the soils present high OC and N_{tot} contents ranging from 1.61 to 5.40% and from 0.08 to 0.33%, respectively. Nickel may also be hosted by organic matter (OM) as demonstrated by its affinity to bind to OM (e.g. Saar and Weber, 1982; Buffle, 1988; McCarthy and Zachara, 1989).

The soil samples were acidic ($\text{pH}_w = 5.2\text{-}6.8$), and the ΔpH ($\text{pH}_{\text{KCl}} - \text{pH}_w$), ranging from -1.84 to -0.42 , indicating that the acidity reserve was very high and that negative charges prevailed in these soil samples, in relation to the significant organic content (up to 5.4 wt%). The isotope composition for $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ ranges from -25.65 to -15.63‰ and from 4.30 to 7.60‰ , respectively. The $\delta^{13}\text{C}$ values of soils from Niquelândia are clearly lighter than for soils from Barro Alto and therefore it appears that the origin of the OC is different between the two ultramafic sites.

3.1.2. Nickel availability in the soil samples

The DTPA-extractable Ni ranged from 14 to 623 mg kg^{-1} ($n = 18$), representing 0.3 to 4.9% of the total Ni. These DTPA-extractable Ni concentrations were in the same range as those reported for UM soils developed in a tropical environment on Mt Kinabalu, Malaysia (van der Ent et al., 2018) and in New Caledonia (Perrier et al., 2006). However, in temperate climates, values ranging from 1.48 to 10.5% in the A horizons of Albanian and Czech UM soils and 3.4 to 8.4% in rhizospheric soils were reported (Quantin et al., 2008, Bani et al., 2014; Estrade et al. 2015). More recently, Pędziwiatr et al. (2018) reported a comparable proportion of DTPA-extractable Ni in A horizons of UM soils ($4.0\text{--}6.8\%$) in Lower Silesia (Poland).

The DTPA-extractable Ni pool of topsoils corresponds to the Ni pool from which hyperaccumulators and non-accumulators take up Ni (Massoura et al., 2005; Chardot et al., 2005); this makes it a suitable method to assess the chemical pool accessible to plants. Ni availability may be more strongly influenced by the biogeochemical recycling of litter, and therefore, soil organic matter may host a significant fraction of available Ni after the decay of Ni-rich litter, demonstrating the strong correlation between Ni-DTPA with total organic carbon (Bani et al., 2014). However, even if DTPA extraction has been shown to be able to efficiently determine the amount of Ni available for plants (Echevarria et al., 1998, 2006), in some cases it could underestimate the actual phytoavailability of Ni because the extraction period was too short or because of Ni oversaturation (Kukier and Chaney, 2001; Echevarria et al., 2006). This may have an influence on the isotopic composition of the phytoavailable Ni determined by DTPA extraction (Estrade et al., 2015).

The DTPA-extractable Ni was within the same range as the values reported for UM soils developed in a tropical environment (Perrier et al., 2006; van der Ent et al., 2018), ranging from 14 to 623 mg kg⁻¹ (n = 18), representing 0.3 to 4.9% of the total Ni.

3.1.3. Ni isotope compositions of the soil samples

The $\delta^{60}\text{Ni}$ values for the soil samples collected in Barro Alto and Niquelândia range from $-0.30 \pm 0.06\text{‰}$ to $0.16 \pm 0.05\text{‰}$ (Tab. 1, Ratié et al., 2015, 2016). This Ni isotope composition is consistent with the values found for the UM soils studied by Estrade et al. (2015) in Albania, but lighter than those reported by Pędziwiatr et al. (2018). In Barro Alto, Ratié et al. (2015) have also reported $\delta^{60}\text{Ni}$ values from three horizons of one soil profile in the same area ranging from -0.19 to -0.02‰ . The range of $\delta^{60}\text{Ni}$ values of soils appear to be primarily isotopically light compared to UM rocks and slightly weathered materials from Barro Alto and Niquelândia (from 0.10 to $0.32 \pm 0.05\text{‰}$; Ratié et al., 2015, 2018) as well as Bulk Silicate Earth, the isotopic composition of which was first estimated as $0.05 \pm 0.05\text{‰}$ (Gueguen et al., 2013) and recently reevaluated to be $0.23 \pm 0.08\text{‰}$ (Gall et al., 2017). Although Ni appears to be associated to goethite in the mineral horizons or to OM in the topsoils, Figure 2 shows that no correlation is observed between the $\delta^{60}\text{Ni}$ values and the Fe/Ni ratio (Fig. 2a), while the heaviest Ni isotope signature corresponds to the largest OC/Ni ratio (Fig. 2b) suggesting that $\delta^{60}\text{Ni}$ could be controlled by OM mainly coming from litter decomposition.

The $\delta^{60}\text{Ni}$ values for the extracted Ni-DTPA present a wide variability from $-0.30 \pm 0.05\text{‰}$ to $0.34 \pm 0.05\text{‰}$ (Tab. 1). A trend towards heavy Ni isotope enrichment in the DTPA solution relative to the soil is observed (Fig. 2c). These results are in line with the work of Estrade et al. (2015), who showed that the Ni isotope composition of the Ni-DTPA fraction was systematically heavier than that of the soil and thus strongly influenced by the water-soluble fraction of Ni. As discussed previously, the possible incomplete extraction of bioavailable Ni in the DTPA solution may induce a bias in the Ni isotopic measurement of this pool, as $\delta^{60}\text{Ni}_{\text{DTPA}}$ depends on the yield of the extraction and the isotopic fractionation range during the chemical reaction (Estrade et al., 2015). Moreover, the significant variability in the $\delta^{60}\text{Ni}$ values makes it complicated to assert that the isotopic composition of the DTPA extraction represents the isotopic signature of the Ni pool bioavailability for plants. However, it

is not excluded that this variability is natural and related to the properties of the soil as discussed above. Moreover, when tracing the available Ni with radioactive ^{63}Ni , Echevarria et al. (1998) were able to demonstrate that the DTPA-extractable pool of Ni in one soil had the same isotopic composition as Ni in the soil solution, thus showing that DTPA-extractable Ni can be in chemical equilibrium with the Ni in the soil solution after a long period of tracing. This heavy Ni isotope enrichment in the available Ni pool shows that the Ni that is accessible to plants comes from another source than the Ni pool associated with iron oxides (i.e. characterized by a light Ni isotope composition) and thus potentially from the Ni released during litter decay. Therefore, Ni-bearing phases play an important role in determining the isotopic composition of the available Ni. Given that goethite (i.e. the dominant Ni-bearing phase in tropical UM soils) acts as a stable sink for metal in Ni-rich soils (Echevarria et al., 2006), the availability of the Ni contained in the sink was extremely low and thus another Ni-bearing phase (i.e. organic matter) must be responsible for this heavy isotope enrichment of the DTPA-extractable pool.

3.2. Plant chemistry

The chemical composition of the plant organs (root, stem, leaf and flower) is described in the supplementary information (Tab. 2). For each specimen, the translocation factors (TF) showed that the aerial parts were enriched in Mg, Fe and Ni compared to the roots and bulk soil, and that they were also enriched in Ca and K despite their low concentrations in these UM environments. Figure 3 shows the evolution of the Ni concentration from the soil to the aerial part with a Ni content exceeding 1000 mg kg^{-1} in the leaves of the *Justicia lanstyiakii* ($1591 \pm 817 \text{ mg kg}^{-1}$) and *Heliotropium aff. Salicoides* specimens ($1446 \pm 716 \text{ mg kg}^{-1}$) and in *Cnidocolus aff. Urens* flowers ($978 \pm 754 \text{ mg kg}^{-1}$). In contrast with Estrade et al. (2015), the Ni concentrations in hyperaccumulator plants were somewhat different from the Ni content in the Ni-DTPA fraction. This lower accumulation could be explained by lower amount of DTPA-extractable Ni or by Ni uptake at a greater depth by the roots.

The leaves from Ni accumulator plants (i.e. *Manihot sp.* specimens) contained Ni concentrations of $331 \pm 241 \text{ mg kg}^{-1}$, i.e. four to five times less Ni than the leaves of Ni hyperaccumulator plants. In the

literature, plants growing on uncontaminated and non-UM soils have Ni contents ranging from 0.05 to 5 mg kg⁻¹ (Welch, 1981). Ni (hyper)accumulator plants possess up to 300 times the maximum concentration of non-accumulator plants.

Figure 3 also shows highly diverse chemical compositions for the various specimens of the same species collected in the same area and between the two UM sites (Barro Alto and Niquelândia). Reeves et al. (2007) also showed this high variability in the Ni contents in plants growing on the ultramafic soils of Goiás, where the Ni contents varied from 274 to 2690 mg kg⁻¹ for *Justicia lanstykii*, from 96 to 2015 mg kg⁻¹ for *Heliotropium aff. Salicoides* and from 156 to 1019 mg kg⁻¹ for *Cnidocolus aff. urens*. Up until now, it is unclear why this variability exists; however, it could be related to the different soil conditions and, in particular, to Ni solid speciation and its subsequent bioavailability, as well as the age, growth stage and genetic internal physiological differences among the specimens and species (Reeves et al., 2007). As previously described, the $\delta^{13}\text{C}$ value for soils from Niquelândia and Barro Alto clearly have two distinct origins for the OC which could be explained by the different chemical composition of the various specimens. Centofanti et al. (2012) demonstrated under controlled conditions that the Ni uptake by *Alyssum corsicum* (Ni hyperaccumulator species) is related to the solubility of the mineral species present in the soil. These factors must be taken into account when explaining this spatial variability in the elemental content in plants, as well as the intra-specimen genetic differences and growth stages.

3.3. Overall Ni isotopic trend during Ni uptake by plants

The Ni isotopic compositions of the various plant parts are presented in Figure 3 and all of the $\delta^{60}\text{Ni}$ values are given in Table 2. The $\Delta^{60}\text{Ni}$ (‰) values calculated with the $\delta^{60}\text{Ni}$ mean value for each part of the plant are provided in Table 3. There is a high intra-specimen variability giving large error ranges (Fig. 3).

The $\delta^{60}\text{Ni}$ values in *Justicia lanstykii* roots vary from $-0.47 \pm 0.08\text{‰}$ to $0.03 \pm 0.11\text{‰}$ ($n = 4$) and are close to those of the Ni-DTPA extracts. *Justicia lanstykii* leaves are enriched in heavy Ni isotopes relative to the stems ($\Delta^{60}\text{Ni}_{\text{leaves-stems}} = 0.24 \pm 0.17\text{‰}$). In contrast with *Justicia lanstykii*, the $\delta^{60}\text{Ni}$

values in *Heliotropium aff. Salicoide* and *Cnidoscopus aff. urens* roots range from -0.12 ± 0.05 to $0.43 \pm 0.06\text{‰}$ and from $0.52 \pm 0.05\text{‰}$ to $0.55 \pm 0.07\text{‰}$, respectively, showing a heavier Ni isotope composition than that for the corresponding Ni-DTPA extracts ($-0.21 \pm 0.05\text{‰}$ to $0.07 \pm 0.06\text{‰}$). Similarly to *Justicia lanstykii*, *Heliotropium aff. salicoides* and *Cnidoscopus aff. urens* leaves are enriched in heavy Ni isotopes compared to the stems ($\Delta^{60}\text{Ni}_{\text{leaves-stems}} = 0.11 \pm 0.12\text{‰}$ and $\Delta^{60}\text{Ni}_{\text{leaves-stems}} = 0.85 \pm 0.15\text{‰}$, respectively). The $\delta^{60}\text{Ni}$ values for *Manihot sp.* follow the trends observed for the three Ni hyperaccumulator plants. The stems present a range of $\delta^{60}\text{Ni}$ values from $0.01 \pm 0.10\text{‰}$ to $0.45 \pm 0.08\text{‰}$ ($n = 4$), while the leaves exhibit heavier $\delta^{60}\text{Ni}$ values from $0.37 \pm 0.11\text{‰}$ to $0.92 \pm 0.06\text{‰}$ ($n = 4$).

No linear trend from the roots to the leaves is observed for the Ni hyperaccumulator plants. However, two trends are observed (Tab. 3): (1) the stems are more enriched in light Ni isotopes than the roots ($-0.22 \pm 0.19\text{‰} < \Delta^{60}\text{Ni}_{\text{stems-roots}} < -0.02 \pm 0.19\text{‰}$) and (2) the leaves are more enriched in heavy Ni isotopes than the stems ($0.11 \pm 0.12\text{‰} < \Delta^{60}\text{Ni}_{\text{leaves-stems}} < 0.85 \pm 0.15\text{‰}$). In the case of the Ni accumulator plant, all of the *Manihot sp.* specimens showed a trend of heavy Ni isotope enrichment during the translocation process from the roots to the leaves ($\Delta^{60}\text{Ni}_{\text{leaves-roots}} = 0.41 \pm 0.17\text{‰}$) (Tab. 3).

Estrade et al. (2015) showed that Ni hyperaccumulator plants collected within a UM context (*Alyssum murale* and *bertolonii*) in a Mediterranean climate (Albania) did not present any significant changes in terms of the isotopic composition among the various organs of the plant. These authors argued that the large amount of Ni transferred to the plant, which was redistributed through phloem flow in the flowering stage, could suppress Ni isotope fractionation (Estrade et al., 2015). Conversely, the results obtained for Ni hyperaccumulator plants in a hydroponic solution (i.e. under controlled conditions) showed an enrichment in light isotopes in the aerial parts (Deng et al., 2014). Accordingly, in the present study, all of the stems from Ni hyperaccumulator plants are enriched in light Ni isotopes compared with the roots ($-0.22 \pm 0.19\text{‰} < \Delta^{60}\text{Ni}_{\text{stems-roots}} < -0.02 \pm 0.19\text{‰}$).

Later, it was confirmed for Ni hyperaccumulator plants that an enrichment of Ni was found in the leaf phloem exudates (Tang et al., 2016). Nickel is primarily complexed by citrate in *P. balgooyi* (van der

Ent et al., 2017), while malate is the main chelator for phloem Ni in *N. caerulea* (Tang et al., 2016). Unlike xylem flow (Centofanti et al., 2013), Nickel translocation in phloem is therefore regulated by carboxylic acid complexation. For *Alyssum* species, phloem redistribution is at its maximum in the flowering stage, which explains why the fractionation between the roots, stems, leaves and flowers was null at this stage (Estrade et al., 2015). Therefore, the redistribution of Ni via the phloem could be responsible for the Ni isotope composition observed for the Ni hyperaccumulator species studied in this present work.

3.4. Implications for the Ni cycle in surface soils

Several studies have shown that the metal solubilization and accumulation mechanism mediated by plants does not cause soil acidification (Bernal et al., 1994; McGrath et al., 1997; Zhao et al., 2001). However, it is evident that plants secrete organic acids or amino acids intended for the chelation of metallic elements (Briat and Lebrun, 1999; Puschenreiter et al., 2003; Wenzel et al., 2003). Some hyperaccumulators may also enhance mineral dissolution to improve their Ni uptake (Chardot-Jacques et al., 2013). This contribution modified the soil edaphic conditions and may increase the bioavailable Ni content and consequently may potentially modify its Ni isotopic signature. Moreover, Zelano et al. (2018) observed that, under their experimental conditions, Ni binding by carboxylic acids such as citric and oxalic acids produced a very small Ni fractionation compared to free Ni in the solution ($\Delta^{60}\text{Ni}_{\text{bound-free}} < 0.2 \text{ ‰}$), while complexation with purified humic acid did not show any fractionation.

The leaves, which are the main constituents of the Brazilian “Cerrado” litter, are the plant compartments with the highest Ni concentrations. The heavy isotope enrichment of Ni from the leaves relative to the soils ($0.09 \pm 0.06 \text{ ‰} < \Delta^{60}\text{Ni}_{\text{leaves-soil}} < 1.06 \pm 0.03 \text{ ‰}$, Tab. 3) suggests that the restitution and decomposition of this organic matter into topsoil will be accompanied by a supply of heavy labile Ni isotopes in the topsoil as measured in the DTPA solution. Zelano et al. (2018) observed that, during the degradation of leaves from hyperaccumulator plants, most of the Ni released had the same isotopic composition as the original leaves from *Alyssum murale*; for *Rinorea bengalensis*, during the first days of leaching, the Ni released in solution appeared to be isotopically heavy. However, this may

contradict the results provided by Estrade et al. (2015) who reported that hyperaccumulators probably take up the lighter Ni fraction of the available heavy pool determined by DTPA, but that this fraction is still heavier than the total Ni in the soil.

The released Ni can then be sorbed and/or incorporated in the mineral or organic phases, as well as be taken up by plants or topsoil microorganisms. Assuming that Ni bound to organic matter in UM soil is more readily available than Ni bound to iron oxides, the heavy Ni isotope composition measured in the aerial parts of the plants may therefore be related to the heavy isotope signature of the DTPA extraction. The Ni contained in the DTPA solution would originate from the Ni initially complexed with the organic matter, resulting in a rapid cycling of heavy Ni isotopes at the soil surface. This remobilized Ni can also be leached down the soil profile to be sorbed deeper or discharged to surface or groundwaters. This progressive loss of heavy Ni in the surface horizons may explain the relatively light Ni isotope composition of Ni in the soils (Estrade et al., 2015; Ratié et al., 2015, 2016).

4. Conclusions

Ni hyperaccumulator plants (*Justicia lanstykii*, *Heliotropium aff. salicoides*, *Cnidocolus aff. urens*) and Ni accumulator plants (*Manihot sp.*) and their respective rhizospheric soils were collected at two UM sites in the Brazilian state of Goiás. The bioavailability of Ni for plants in rhizospheric soils, obtained by DTPA extraction, exhibits $\delta^{60}\text{Ni}$ values ranging from -0.30 ± 0.05 to $0.34 \pm 0.05\text{‰}$ which can be used to identify an enrichment in heavy Ni isotopes in the DTPA-extractable Ni in topsoils ($-0.31 \pm 0.10\text{‰} < \Delta^{60}\text{Ni}_{\text{soils-DTPA}} < -0.01 \pm 0.12\text{‰}$).

In both Ni hyperaccumulator and accumulator plants, the Ni concentrations exhibit a high content and large variability (up to 2.3 g kg^{-1}). Likewise, their Ni isotope compositions show large variability (-0.47 ± 0.08 to $1.21 \pm 0.05\text{‰}$). Ni accumulator plants exhibit an enrichment in heavy Ni isotopes in the aerial parts, while the similar $\delta^{60}\text{Ni}$ values for the roots, stems and aerial parts suggests that no significant fractionation occurs for the Ni hyperaccumulator plants during Ni uptake. However, two trends are identified between the various compartments induced by redistribution and transfer of Ni in

plants: (1) the stems are more enriched in light Ni isotopes than the roots (*Justicia lanstyakii*: $\Delta^{60}\text{Ni}_{\text{stems-roots}} = -0.02 \pm 0.19\text{‰}$; *Heliotropium aff. Salicoides*: $\Delta^{60}\text{Ni}_{\text{stems-roots}} = -0.22 \pm 0.19\text{‰}$; *Cnidoscopus aff. Urens*: $\Delta^{60}\text{Ni}_{\text{stems-roots}} = -0.17 \pm 0.13\text{‰}$), while (2) the leaves are more enriched in heavy Ni isotopes than the stems (*Justicia lanstyakii*: $\Delta^{60}\text{Ni}_{\text{leaves-stems}} = 0.24 \pm 0.17\text{‰}$; *Heliotropium aff. Salicoides*: $\Delta^{60}\text{Ni}_{\text{leaves-stems}} = 0.11 \pm 0.12\text{‰}$; *Cnidoscopus aff. Urens*: $\Delta^{60}\text{Ni}_{\text{leaves-stems}} = 0.85 \pm 0.15\text{‰}$).

In the laterite profile, the fractionation of Ni performed by plants and more especially the heavier Ni isotope composition in the leaves for all of the plants compared to the rhizospheric soil ($0.09 \pm 0.06\text{‰} < \Delta^{60}\text{Ni}_{\text{leaves-soil}} < 1.06 \pm 0.03\text{‰}$) may play a role in the overall isotopic Ni cycle. The input of heavy Ni litter falling onto the soil may gradually increase the heavy Ni isotope pool in the topsoil. Moreover, this pool is the first output of Ni in soils through lixiviation due to Ni recycling via litter degradation in the UM area.

All of these features highlight how complex it is to understand the role of plants in Ni biogeochemical cycling and indicates that the determination of the Ni isotope composition in these types of studies can help to identify and better quantify the biogeochemical cycle of metals in the soil-plant continuum. Our results also highlight the need to couple field studies with a laboratory approach with experiments under controlled conditions; hydroponic experiments may allow for the quantification of masses at the whole plant scale, on a larger number of specimens, and at different growth stages.

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Table captions

Table 1: Chemical (K, Ca, Mg, Al, Fe, Mn, Ni, N_{org} and OC) and isotopic composition (N, C and Ni) of topsoils collected in Barro Alto and Niquelândia. Sample color was determined according to the Munsell chart. Localization (UM BA: in the UM Barro Alto complex, UM Niq: in the UM Niquelândia complex). DTPA extraction was performed with measurement of Ni content extracted by DTPA and its Ni isotope composition. * published names used in Ratié et al. 2016. n.d.: not determined.

Table 2: Chemical and Ni isotopic composition of plant samples. For each specimen, chemical composition of topsoil and its Ni DTPA extract were given. Jus : *Justicia Lanstykii*, Hel : *Heliotropium aff. Salicoide*, Cni : *Cnidocolus aff. Urens*, Man : *Manihot sp.*, * sampled in UM site of Barro Alto, ** sampled in UM site of Niquelândia.

Table 3: Table 2: $\Delta^{60}\text{Ni}$ (‰) calculated with the $\delta^{60}\text{Ni}$ mean value for each part of plant. Jus: *Justicia Lanstykii*, Hel: *Heliotropium aff. salicoide*, Cni: *Cnidocolus aff. urens*, Man: *Manihot sp.* Mean SD values have been calculated: $\frac{1}{n} \sum 2\sigma$. Hyp: Ni-hyperaccumulator, Acc: Ni-accumulator.

Sample name	Sample color	Localization	Total														DTPA extraction			
			K	Ca	Mg	Al	Fe	Mn	Ni	N _{tot}	OC	pH	Δ pH	$\delta^{15}\text{N}$	$\delta^{13}\text{C}$	$\delta^{60}\text{Ni}$	2SD	Ni DTPA	$\delta^{60}\text{Ni}$	2SD
			g/kg							%			$\text{pH}_{\text{KCl}}-\text{pH}_{\text{w}}$	‰			mg/kg	‰		
BA_S1*	2,5YR 2,5/3	UM BA	0.1	4.2	24.9	24.6	290.8	5.2	12.8	0.26	3.82	5.9	-0.76	7.52	-17.93	-0.09	0.05	623	n.d.	n.d.
BA_S2*	2,5YR 2,5/3	UM BA	0.1	3.7	16.9	26.2	298.5	5.1	10.8	0.26	3.58	6.1	-0.72	7.60	-18.64	-0.19	0.05	444	n.d.	n.d.
BA_S3*	2,5YR 2,5/2	UM BA	0.2	6.4	19.0	42.4	318.3	5.2	9.8	n.d.	n.d.	5.9	-0.42	n.d.	n.d.	-0.15	0.05	n.d.	n.d.	n.d.
BA_S4*	2,5YR 2,5/3	UM BA	0.2	4.5	20.0	44.0	307.4	5.0	9.9	0.19	2.63	6.2	-0.74	5.95	-16.30	-0.16	0.05	220	-0.13	0.14
BA_S5*	2,5YR 3/2	UM BA	0.2	0.5	6.4	35.3	336.2	5.2	7.5	0.33	5.40	6.3	-1.28	4.95	-18.99	-0.10	0.08	206	-0.06	0.05
BA_S6*	2,5YR 3/3	UM BA	0.1	0.3	4.3	34.6	357.3	6.3	7.6	0.24	3.82	6.2	-1.23	6.41	-16.75	-0.14	0.05	117	-0.21	0.05
BA_S7*	2,5YR 2,5/3	UM BA	0.1	0.1	1.5	35.2	390.3	5.2	4.3	0.21	3.61	6	-1.54	5.44	-17.65	0.10	0.05	14	-0.08	0.08
BA_S8*	10R 3/2	UM BA	0.2	0.1	1.2	32.9	278.3	3.8	3.3	0.21	3.59	6.3	-1.84	4.98	-19.20	0.11	0.10	16	-0.06	0.06
BA_S9*	2,5YR 2,5/3	UM BA	0.2	5.8	16.6	37.5	347.2	5.1	10.4	0.18	2.41	6.8	-1.56	5.47	-15.92	-0.20	0.13	181	-0.05	0.06
BA_S10*	2,5YR 3/4	UM BA	0.2	2.8	14.6	36.8	314.5	4.3	9.2	0.22	3.16	6.5	-1.23	5.63	-17.86	-0.09	0.08	260	-0.03	0.05
BA_S11*	5YR 2,5/3	UM BA	0.2	8.2	20.4	33.5	356.0	5.8	10.7	0.18	2.54	n.d.	n.d.	4.30	-17.04	-0.26	0.09	191	-0.30	0.05
BA_S12*	2,5YR 2,5/3	UM BA	0.1	4.6	22.9	36.3	509.6	7.0	14.2	0.16	2.28	n.d.	n.d.	5.78	-16.04	-0.17	0.05	124	-0.02	0.05
BA_S13*	2,5YR 3/4	UM BA	0.2	8.4	24.7	30.2	410.6	7.1	13.7	0.17	2.28	n.d.	n.d.	6.16	-15.63	-0.30	0.06	198	-0.27	0.05
NQ_S1*	10R 3/3	UM Niq	0.1	0.7	22.8	23.9	364.8	6.8	9.2	0.17	2.73	6	-0.88	6.01	-23.32	-0.07	0.05	301	n.d.	n.d.
NQ_S2*	10R 3/4	UM Niq	0.1	0.4	14.9	22.3	459.7	7.9	10.1	0.16	2.47	6.3	-1.22	5.08	-25.65	-0.06	0.05	269	0.05	0.05
NQ_S3*	10R 3/3	UM Niq	0.1	0.4	16.1	19.2	457.4	7.7	9.4	0.08	1.61	6.5	-1.08	5.22	-24.09	-0.09	0.05	94	0.23	0.08
NQ_S4*	10R 2,5/2	UM Niq	0.1	0.3	16.5	21.9	517.4	9.5	12.2	0.17	2.70	6.1	-0.9	5.21	-25.07	-0.05	0.14	186	0.07	0.06
NQ_S9	10R 3/4	UM Niq	0.1	0.7	24.3	13.9	513.2	8.1	16.9	n.d.	n.d.	6.5	-0.95	n.d.	n.d.	0.15	0.05	293	0.34	0.05
NQ_S10	5YR 5/6	UM Niq	0.1	1.4	24.8	16.4	343.5	6.5	11.8	n.d.	n.d.	6.6	-0.68	n.d.	n.d.	0.16	0.05	412	0.34	0.06

Table 1

Name	Type	Ca	K	Mg	Na	Fe	Ni	$\delta^{60}\text{Ni}$	2 S.D.	Ni DTPA	$\delta^{60}\text{Ni}$	2 S.D.	Translocation Factor							
		mg kg^{-1}						‰		mg kg^{-1}		‰		Ca	K	Mg	Na	Fe	Ni	
Jus1*	BA_S11	8231	180	20423	392	356000	10700	-0.26	0.09											
	Roots	1535	1339	2199	162	422	368	-0.47	0.08	192	-0.30	0.05	8.8	13.7	11.1	1.3	6.2	3.4		
	Stems	3840	11327	2197	224	953	666	-0.44	0.07											
	Leaves	13465	18300	24500	214	2604	1239	-0.25	0.07											
Jus2*	BA_S12	4599	145	22942	293	509597	14159	-0.17	0.05	124	-0.03	0.07	6.6	3.4	13.2	0.7	5.1	0.8		
	Roots	2727	2893	2524	222	524	723	-0.21	0.09											
	Stems	1984	14196	2327	171	533	369	-0.15	0.09											
	Leaves	17999	9766	33434	166	2661	602	0.03	0.07											
Jus3*	BA_S13	8424	158	24689	411	410590	13719	-0.30	0.06	198	-0.27	0.05	12.5	22.5	3.7	1.4	1.6	5.5		
	Roots	919	1201	2087	182	663	418	-0.45	0.06											
	Stems	2163	5998	2357	267	1563	691	-0.40	0.05											
	Leaves	11495	27080	7745	260	1051	2309	-0.16	0.05											
Jus4*	BA_S1	4201	125	24923	273	290819	12798	-0.09	0.05	623	n.d.	n.d.	8.5	2.2	6.7	1.0	0.2	2.7		
	Roots	1630	4998	4058	430	3760	814	0.03	0.11											
	Stems	2149	7988	2675	452	377	505	-0.19	0.05											
	Leaves	13935	10766	27080	441	898	2213	0.16	0.05											
Hel1*	BA_S6	289	99	4308	40	357324	7589	-0.14	0.05	117	-0.21	0.05	18.0	4.6	1.5	0.9	1.8	2.3		
	Roots	3356	1493	4997	182	264	542	0.01	0.07											
	Stems	5095	3799	2871	156	214	459	-0.18	0.15											
	Leaves	60561	6872	7425	157	474	1272	-0.02	0.12											
Hel2*	BA_S9	5796	146	16631	321	347163	10430	-0.20	0.13	181	-0.05	0.06	17.2	1.1	2.3	0.7	5.3	8.1		
	Roots	2566	4117	4909	244	294	295	-0.01	0.05											
	Stems	15494	5600	5031	155	2147	841	-0.26	0.05											
	Leaves	44108	4409	11396	161	1544	2381	0.02	0.05											
Hel3*	BA_S1	4201	125	24923	273	290819	12798	-0.09	0.05	623	n.d.	n.d.	18.9	1.4	2.7	0.9	0.8	2.2		
	Roots	2744	7343	2612	84	1995	371	-0.12	0.05											
	Stems	9798	6595	2953	79	2389	337	-0.13	0.05											
	Leaves	51871	10327	7099	79	1522	800	-0.08	0.05											
Hel4*	BA_S4	4532	196	20029	193	307363	9848	-0.16	0.05	217	-0.13	0.14	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.		
	Roots	12960	7087	3544	75	494	432	-0.36	0.06											
	Stems	55006	11559	11875	90	1446	946	-0.13	0.05											
	Leaves	53990	8629	10518	77	752	455	-0.17	0.05											
Hel5**	NQ_S2	457	127	14939	36	459661	10074	-0.06	0.05	269	0.05	0.05	18.5	1.9	5.0	1.1	7.0	1.7		
	Roots	1728	1771	3113	142	257	1375	0.43	0.06											
	Stems	4897	2865	6639	160	399	1626	0.14	0.08											
	Leaves	31968	3296	15445	162	1791	2316	0.02	0.1											
Hel6**	NQ_S4	324	116	16470	28	517380	12242	-0.05	0.14	186	0.07	0.06	24.9	1.5		1.3	6.1	8.1		
	Roots	1518	2170	n.d.	120	664	118	0.06	0.07											
	Stems	6277	2865	4295	152	1153	164	-0.06	0.1											
	Leaves	37795	3229	17980	160	4038	959	0.01	0.07											
Cni1*	NQ_S9	719	87	24309	43	513204	16873	0.15	0.05	292	0.22	0.12	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.		
	Roots	2896	6890	4816	154	221	325	0.55	0.07											
	Stems	11725	4419	15069	135	133	429	0.63	0.1											
	Leaves	11842	11106	4892	137	2195	1511	0.47	0.05											
Cni2*	NQ_S9	719	87	24309	43	513204	16873	0.15	0.05	292	0.34	0.05	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.		
	Roots	1593	7503	2125	145	136	135	0.52	0.05											
	Stems	2716	5266	6114	136	302	41	0.14	0.05											
	Leaves	11464	9899	8633	152	964	445	0.32	0.09											
Cni3*	NQ_S10	1382	89	24831	/	343511	11775	0.16	0.06	412	0.34	0.06	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.		
	Stems	6019	17405	6736	188	77	239	0.32	0.05											
Man1*	BA_S1	4201	125	24923	273	290819	12798	-0.09	0.05	623	n.d.	n.d.	1.7	3.1	1.0	0.6	9.6	0.6		
	Roots	3322	3798	4272	125	147	295	0.16	0.06											
	Stems	9791	6816	3337	77	180	338	0.17	0.08											
	Leaves	5560	11625	4085	72	1415	178	0.37	0.11											
Man2**	NQ_S3	416	84	16123	30	457404	9392	-0.09	0.05	95	0.23	0.08	23.8	50.6	13.3	1.1	2.9	13.9		
	Roots	396	146	378	155	123	45	0.23	0.05											
	Stems	2327	4716	1828	141	3	119	0.01	0.1											
	Leaves	9421	7386	5029	168	356	625	0.41	0.06											
Man3**	NQ_S9	719	87	24309	43	513204	16873	0.15	0.05	292	0.34	0.05	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.		
	Stems	13689	2622	0	154	45	148	0.45	0.08											
	Leaves	8158	4023	7902	129	406	96	0.92	0.06											
	Flowers	18136	6689	8195	146	23	33	0.23	0.05											
Man4**	NQ_S1	709	109	22749	n.d.	364841	9207	-0.07	0.05	302	n.d.	n.d.	2.9	1.4	2.1	0.5	3.1	2.1		
	Roots	2663	7465	2402	386	304	199	0.04	0.05											
	Stems	6304	11954	2286	457	75	187	0.12	0.05											
Leaves	7748	10471	5056	199	933	425	0.53	0.05												

Table 2

Plant	$\Delta^{60}\text{Ni}_{\text{soils-DTPA}}$	$\Delta^{60}\text{Ni}_{\text{roots-DTPA}}$	$\Delta^{60}\text{Ni}_{\text{stems-roots}}$	$\Delta^{60}\text{Ni}_{\text{leaves-roots}}$	$\Delta^{60}\text{Ni}_{\text{leaves-stems}}$	$\Delta^{60}\text{Ni}_{\text{leaves-soil}}$
$\Delta^{60}\text{Ni}_{x-z}$ in ‰ ($\delta^{60}\text{Ni}_x - \delta^{60}\text{Ni}_z$)						
Jus	-0.01 ± 0.12	-0.08 ± 0.19	-0.02 ± 0.19	0.22 ± 0.21	0.24 ± 0.17	0.15 ± 0.14
Hel Hyp.	-0.06 ± 0.09	0.13 ± 0.16	-0.22 ± 0.19	-0.1 ± 0.14	0.11 ± 0.12	0.09 ± 0.06
Cni	-0.15 ± 0.04	0.24 ± 0.05	-0.17 ± 0.13	0.68 ± 0.04	0.85 ± 0.15	1.06 ± 0.03
Man Acc.	-0.31 ± 0.10	-0.14 ± 0.09	0.04 ± 0.14	0.41 ± 0.17	0.37 ± 0.22	0.58 ± 0.18

Table 3

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Figure captions

Figure 1: Photographies of the sampled species: (a) *Justicia Lanstykii*, (b) *Heliotropium aff. salicoides*, (c) *Cnidocolus aff. urens*, d) *Manihot sp.* (Raous et al., 2007 for a, b and c, and personal photography for d).

Figure 2: $\delta^{60}\text{Ni}$ values as a function of Fe/Ni ratio(a), OC/Ni ratio(b) and $\delta^{60}\text{Ni}$ values of DTPA-Ni fraction. Linear regression of all data (Barro Alto and Niquelândia) is represented in (a) and (b) with correlation factor R^2 . Black line in (c) represents a linear regression 1:1.

Figure 3: Ni isotope composition for each specimen of Ni-hyperaccumulator and Ni-accumulator species: (a) *Justicia Lanstykii*. (4 specimens), (b) *Heliotropium aff. salicoide*. (6 specimens), (c) *Cnidocolus aff. urens* (3 specimens) and (d) *Manihot sp.* (4 specimens). Error bars are 2 s.d. of NIST SRM 986 Ni for all of the measurements except when the uncertainty from the replicated measurements is higher than 0.05 ‰. Ni contents in the different parts of each specimen are also represented. Mean value of Ni content in topsoils is annotated for each species and error bars are included in the mark size. *Collected at Barro Alto, **Collected at Niquelândia.

Highlights

DTPA extractable Ni in topsoil is isotopically heavier than the total Ni pool

Ni-accumulators exhibit enrichment in Ni heavy isotopes in aerial parts

Similar $\delta^{60}\text{Ni}$ values for each plant compartment are observed for Ni-hyperaccumulators

Enrichment in heavy Ni isotopes in leaves may lead to a heavy Ni input in the litter

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Figure 1

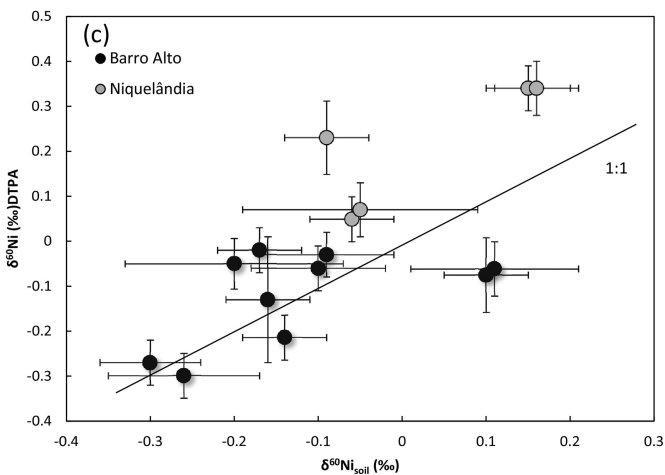
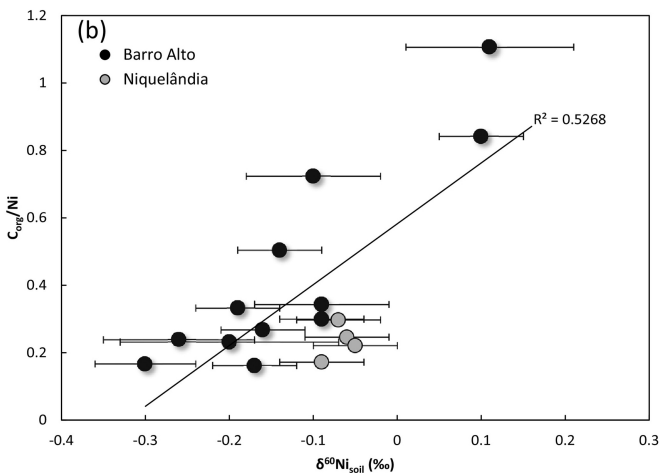
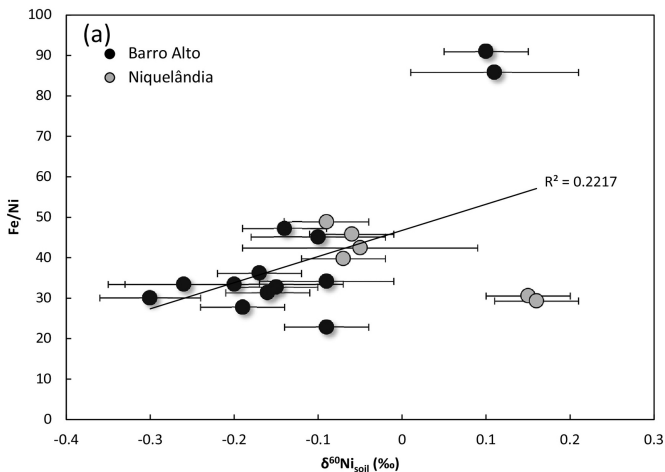


Figure 2

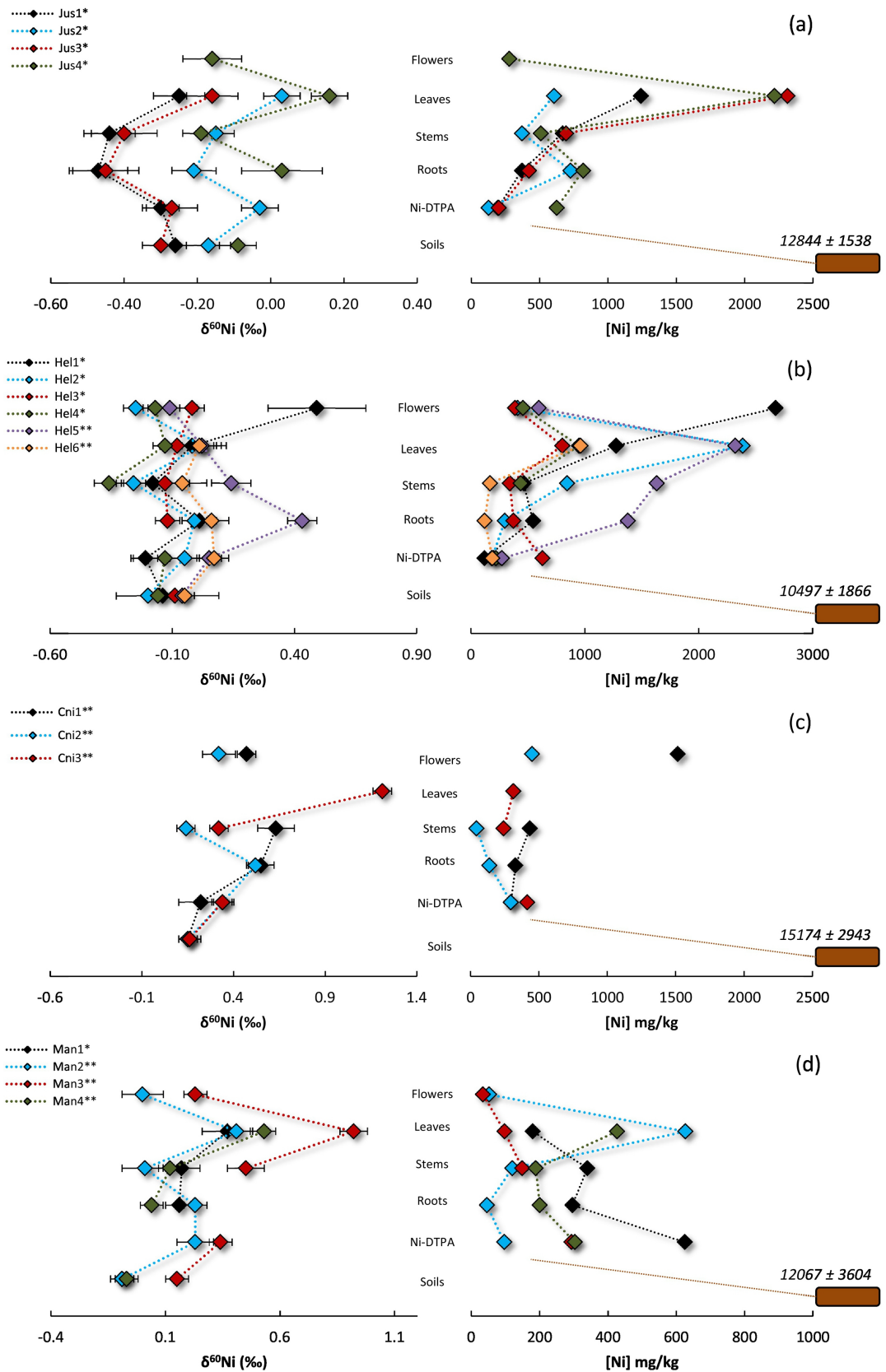


Figure 3