

Predicting duodenal flows and absorption of fatty acids from dietary characteristics in ovine and bovine species: a meta-analysis approach

Lucas Prado, Anne Ferlay, Pierre Noziere, Philippe Schmidely

► To cite this version:

Lucas Prado, Anne Ferlay, Pierre Noziere, Philippe Schmidely. Predicting duodenal flows and absorption of fatty acids from dietary characteristics in ovine and bovine species: a meta-analysis approach. Animal, 2019, 13 (4), pp.727-739. 10.1017/S1751731118001982 . hal-01961394

HAL Id: hal-01961394 https://hal.science/hal-01961394

Submitted on 19 Dec 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Predicting duodenal flows and absorption of fatty acids from dietary characteristics in ovine and bovine species: a meta-analysis approach

L. A. Prado¹, A. Ferlay¹, P. Nozière¹ and P. Schmidely^{2†}

¹Université Clermont Auvergne, INRA, VetAgro Sup, UMR Herbivores, F-63122 Saint-Genès-Champanelle, France; ²AgroParisTech, INRA, UMR0791 Mosar, 16 rue Claude Bernard, F-75231 Paris, France

(Received 24 November 2017; Accepted 15 June 2018)

Dietary and ruminal factors modify the ruminal biohydrogenation (RBH) of polyunsaturated fatty acids (FA), with duodenal FA flows being quantitatively and qualitatively different from FA intake. Using a meta-analysis approach from a database on duodenal flows of FA in ruminants, this study aimed to determine predictive equations for duodenal and absorbed flows of saturated FA, C18:1, C18:2 and C18:3 isomers, odd- and branched-chain FA (OBCFA), C20:5n-3, C22:5n-3 and C22:6n-3 and to quantify the effects of dietary and digestive factors on those equations. The database was divided into four subsets: forage, seed, vegetable oils or animal fats (oil/fat), and fish products (fish) subsets. Models of duodenal and absorbed FA flows were obtained through variance-covariance analysis. Effects of potential interfering factors (conservation mode and botanical families of forages, lipid source, technological processing of lipid supplements, diet composition and animal characteristics) were analysed. We obtained 83 models for duodenal FA flows as a function of FA intake for saturated FA (C14:0, C16:0 and C18:0), C18:1, C18:2 and C18:3 isomers and seven other models for OBCFA. For the seed/oil/fat subset, intakes of total C18:3, C18:2 and starch content increased the duodenal t11-C18:1 flow with 0.08, 0.16 and 0.005 q/kg of dry matter intake (DMI), respectively, whereas intake level [(DMI × 100)/BW] decreased it. The c9c12c15-C18:3 RBH was higher for oil/fat than seed (96.7% v. 94.8%) and a protective effect of Leguminosae v. Gramineae against RBH for that FA appeared in the forage subset. The duodenal C17:0 flow increased with starch content and decreased with ruminal pH, respectively, whereas duodenal iso-C16:0 flow decreased with dietary NDF content for the seed/oil/fat subset. The duodenal C20:5n-3, C22:5n-3 and C22:6n-3 flows depended on their respective intake and the inhibitory effect of C22:6n-3 on duodenal C18:0 flow was quantified. Thirteen models of absorbed FA flows were performed depending on their respective duodenal flows. This study determined the effects of different qualitative and quantitative dietary and digestive factors, allowing for improved predictions of duodenal and absorbed FA flows.

Keywords: biohydrogenation, fatty acids, meta-analysis, rumen, lipid supplementation

Implications

Predictive equations of duodenal and absorbed flows of saturated fatty acids (FA), and unsaturated FA were derived from a large database in ruminants, including experimental factors (forage nature, nature and dose of lipid supplements). These equations depended on FA intake and interfering factors (conservation mode and botanical families of forages, diet composition and animal characteristics). They are the first step to predict the long-chain fatty acid (LCFA) secreted in the milk or the FA profile in meat. They could be included in the renewed Institut National de la Recherche Agronomique (INRA) feed evaluation systems in ruminants.

Introduction

The FA in milk reflects a combination of ruminant specificities of lipid digestion, as well as mammary lipid metabolism (Chilliard *et al.*, 2007). The rumen biohydrogenation (RBH) of polyunsaturated fatty acid (PUFA) leads to the production of geometric and positional isomers of FA with *trans* or *cis* double bonds and saturated FA, mainly C18:0. In the mammary gland, *de novo* FA synthesis produces C4:0 to C16:0, whereas medium-chain fatty acid (MCFA) (50% of C16:0) and LCFA (FA having a number of carbons >16) from chylomicrons or very-low-density lipoproteins are taken up by lipoprotein lipase (Chilliard *et al.*, 2007). In early lactation, when animals are in negative energy balance, adipose reserves are mobilised and thus some plasma FA are

⁺ E-mail: philippe.schmidely@agroparistech.fr

originated from adipose tissue. In parallel with these phenomena, lipid supplementation remains the most effective way to modulate milk FA composition in ruminant.

Due to the extent of RBH, composition of duodenal FA and absorbed FA is very different from the intake. Glasser et al. (2008) and Schmidely et al. (2008) have quantified average regression models for duodenal and absorbed flows of some FA to dietary factors, especially lipid supplementation. They have highlighted the role of additional dietary factors, such as the source of lipids, the physical form (seed v. oil/fat) and the technological processing of lipid supplements on duodenal FA flows. However, the duodenal flows and absorption rates of C18:1, C18:2 and C18:3 isomers have not yet been predicted with accuracy, because of a lack of data and/or an insufficient separation of isomers during chemical analysis. In addition, odd- and branched-chain fatty acid (OBCFA) were not predicted. In the context of the new French INRA feed systems, there is an opportunity to re-evaluate the previous predictive equations of duodenal and absorbed FA flows, including recent studies and characterising the effects of dietary factors on duodenal FA flows with improved precision. Using a metaanalysis approach, the objectives are (1) to quantify duodenal flows and the absorption of different FA including C18:1, C18:2 and C18:3 isomers, OBCFA, MCFA and LCFA; and (2) to quantify more precisely the effects of dietary factors on the relationship between FA intake and duodenal flow.

Material and methods

Selection of publications

The AGRum database (IDDN.FR.001.510032.000.R.C.2011. 000.10300) has been previously described (Glasser *et al.*, 2008; Schmidely *et al.*, 2008). Its update was performed with trials published between 2008 and 2014. The PRISMA diagram (Moher *et al.*, 2009) described the selection process of the peer-reviewed papers considered in this study (Supplementary Figure S1). Publications were taken from the Web of Knowledge and Science Direct research platforms, using the keywords: ruminant, cow, ewe, goat, sheep, biohydrogenation, duodenal FA and digestion. Data with ruminal or postruminal infusion of nutrients were excluded.

The database was then composed of 104 publications (Supplementary Material S1) reporting 167 experiments (N_{exp}) with a total of 437 treatments (N_{trt}). The animal species are divided between bovine ($N_{trt} = 358$) and ovine ($N_{trt} = 79$). Dairy cows are the most studied animals among bovines ($N_{trt} = 202$) followed by beef cattle ($N_{trt} = 156$). The feedstuffs from the database were characterised according to the INRA (2010) feed tables based on their reported chemical composition on a dry matter (DM) basis for CP, NDF, starch, total FA and FA profile (Baumont *et al.*, 2007; Maxin *et al.*, 2013).

Coding

Codes were assigned to each study according to the experimental factors studied and then four database subsets were created: forage, seed, vegetable oils or animal fats (oil/fat) and fish products (fish) subsets. In the forage subset, botanical families and their conservation mode (fresh or pasture, silage and hay) were coded as experimental factors, with all diets based only on one or several forages called forage diets (Supplementary Table S1). In the seed and oil/fat subsets, the lipid source and their technological processing were coded as experimental factors, with two codes designated to each treatment according to lipid source and technological process. The fish subset was created for studies including fish products (oil and meal), with the lipid percentage in the diet (no lipid supplement v. fish products) and the lipid source (fish oil or fish meal v. vegetable oil or animal fat) as experimental factors. In these three subsets, control diets referred to treatments without lipid supplement. The code for lipid source referred to the nature of the main lipid supplement divided as follows: vegetable source, animal fats (including fish products), hydrogenated fat and animal-vegetable blends. The code for technological processes referred to either dry, hydrothermal and mechanical processes, and formaldehyde treatment for seeds, and amide, Ca salts, encapsulated, and formaldehyde treatments for vegetable oils and animal fats, and amide and Ca salts for fish products, respectively (Supplementary Table S1).

Variable selection criteria

A preliminary selection of individual duodenal FA was made to have more than 50 treatments before relating each selected duodenal FA flow (dependent variable) with its respective FA intake (independent variable). Duodenal OBCFA (Fievez et al., 2012) flows that arise mainly from microbial synthesis in the rumen and isomers of C18:1, C18:2 and C18:3 flows were regressed against dietary data (e.g. concentrate, dietary starch and CP contents) or digestive parameters which may reflect or affect ruminal microbial activities, for example, rumen protein balance (RPB), CP microbial synthesis, efficiency of microbial protein synthesis (EMPS), organic matter truly digested in the rumen (OMDr), using the INRA model of feed and diet nutritive values (Sauvant and Nozière, 2013) implemented within the Sytool.fr. application (Chapoutot et al., 2015) and ruminal pH and acetate/propionate (A/P) ratio measured in publications. Absorbed flow was calculated as the difference between duodenal flow and faecal flow or as the difference between ileal flow and faecal flow. Absorbed flows of saturated FA (C14:0, C16:0 and C18:0), total C18:1, C18:2 and C18:3, the main isomers (c9-C18:1, t11-C18:1, c9c12-C18:2 and c9c12c15-C18:3) and LCFA as C20:5n-3 (EPA), C22:5n-3 (docosapentanoic acid (DPA)) and C22:6n-3 (DHA) were predicted from their respective duodenal flows. Different qualitative animal criteria (species and animal physiological stage), quantitative dietary factors (intake level [(dry matter intake $(DMI) \times 100)/BWI$, concentrate percentage, diet chemical composition) and digestive parameters (predicted from Systool. fr) were selected because of their putative effect on the predictive equations.

Statistical analyses

Description of the meta-design. A description of each subset is presented in Supplementary Tables S2 to S5, respectively.

The treatments from the forage subset were presented according to conservation mode (Supplementary Table S2) or botanical families (Supplementary Table S3), whereas those from the seed and oil/fat subsets were described according to the control (Supplementary Table S4) or lipid supplemented diets (Supplementary Table S5). For the fish subset, we separated treatments into three groups: diets supplemented with fish products, other lipid supplements such as vegetable oils or animal fat and control diets without lipid supplement.

Regression models of duodenal fatty acids and absorbed fatty acids. Predictive models for duodenal FA were obtained for each subset using a variance-covariance analysis with a GLM. In the models developed for intra-experiment variations, we considered 'study effect' as a fixed effect (Sauvant *et al.*, 2008). The true intra-experiment response of a dependent variable (Y) to variation in an independent variable (X) was calculated on experiments with a sufficient variation in X: a minimum threshold of variation at intake level $(X_{max} - X_{min})$ was selected with the hypothesis that RBH of PUFA was 90% in order that the minimal variation range ($Y_{max} - Y_{min}$) at duodenal level could be detectable by laboratory analysis. This threshold eliminated 30% to 50% of treatments according to the FA considered, that were further used for external validation (see the 'Model assessments' section). However, this criterion was not applied to regressions using several independent variables to estimate duodenal FA, nor to predict absorbed FA flows from the duodenal flows. The models in equations (1) and (2) were used to describe the relationships between duodenal FA (FA_{duo}, g/kg DMI) and FA intake (FA_{int}, g/kg DMI) and those between absorbed FA (FA_{abs}, g/kg DMI) and FA_{duo}. For absorbed FA models, the analysis included all experiments from the database without taking into account the separation into different subsets:

$$FA_{duo,ij} = \beta_0 + S_i + (\beta_1 + \beta_i) \cdot FA_{int,ij} + \varepsilon_{ij}$$
(1)

$$\mathsf{FA}_{\mathsf{abs},ij} = \beta_0 + \mathsf{S}_i + (\beta_1 + \beta_j) \cdot \mathsf{FA}_{\mathsf{duo},ij} + \varepsilon_{ij} \tag{2}$$

where *i* = experiment index and *j* = treatment index. β_0 is the overall intercept, *S_i* the fixed effect of experiment *i*, β_1 the overall regression slope, β_i the fixed effect of experiment *i* on the slope and ε_{ij} the unexplained residual error.

Identification of potential additional factors. We studied the influence of different factors on three regression parameters: the least square means (LS_{means}) of Y response, slope and residuals of the above equations. First, a *t*-test was used to compare LS_{means} and slope coefficients of the predictive equations between the seed and oil/fat subsets. If the coefficients were not significantly different between the two sets, data were pooled to retain a single regression for both subsets. Second, effects of quantitative (experiment means) and qualitative variables were analysed differently. A Pearson correlation test was applied to determine if the quantitative variables could have an effect on both the slope coefficients and LS_{means} . The effect of qualitative variables on slope coefficients and LS_{means} was assessed with a one-way

ANOVA. To test the influence of qualitative and quantitative factors on previously calculated residuals $(y - \hat{y})$, a one-way ANOVA was conducted for qualitative factors, and a linear global regression model (equation (3)) was used for quantitative factors. For the forage subset, in addition of the previous approach, a global linear regression was performed including two variables, mode of conservation and forage families, and their interactions with the FA intake for duodenal FA flows prediction (equation (4)). To study the pure effect of forage families, treatments consisting of a blend of legumes and grasses were not taken into account. The following model was used:

$$(\mathbf{y} - \hat{\mathbf{y}})_{\text{duo},j} = \beta_0 + \beta_1 \cdot \text{factor}_j + \varepsilon_j$$
 (3)

$$\mathsf{FA}_{\mathsf{duo},j} = \beta_0 + \left(\beta_1 + \mathsf{FAM}_j + \mathsf{CM}_j\right) \cdot \mathsf{FA}_{\mathsf{int},j} + \mathsf{FAM}_j + \mathsf{CM}_j + \varepsilon_j$$
(4)

where FAM and CM are forage families (Gramineae and Leguminosae) and conservation mode (fresh or pasture, silage and hay), respectively. β_0 is the overall intercept, β_1 the regression coefficient and ε_{ij} the random residual error. All statistical analyses were performed with the software Minitab[®] (version 17; State College, PA, USA). We considered P < 0.05 to be statistically significant and P < 0.10 to represent a trend.

Model assessments. To evaluate the models, the coefficients of linear regressions between observed and predicted values were tested using a *F*-test in which the intercept was compared with 0 and the slope was compared with unity. Furthermore, we selected experiments not included in the model (e.g. experiments with a range less than the defined threshold). Then, we calculated a linear regression between observed and predicted values for these experiments, which allowed for the determination of the mean square prediction error (MSPE) from equation (5) (Bibby and Toutenburg, 1977). Three parameters of MSPE (equations (6) to (8)) were studied: (1) the error in central tendency (ECT) that represents the amount the predictive model does not fit; (2) error due to regression (ER) explaining the proportion of slope deviation and (3) error due to disturbance (ED) representing the optimal linear correction of the prediction model:

$$MSPE = \left(\frac{\sum_{n=1}^{i} \left(\overline{[\hat{y}_{i}]} - \overline{[y_{i}]}\right)^{2}}{n}\right)$$
(5)

$$ECT = \left(\overline{[\hat{y}_i]} - \overline{[y_i]}\right)^2$$
(6)

$$\mathsf{ER} = \left(\mathbf{S}_{\hat{y}} - \mathbf{r} - \mathbf{S}_{y}\right)^{2} \tag{7}$$

$$\mathsf{ED} = (1 - r^2) \times S_y^2 \tag{8}$$

where \hat{y} was the predicted duodenal FA flow, y the observed duodenal FA flow and n the number of observations. $S_{\hat{y}}$ and S_y were the standard deviation of the predicted and observed values, respectively. The r refers to the correlation coefficient.

Results

Models from the forage subset

This section deals with duodenal flows of total FA, C18:0, total C18:1, C18:2, C18:3 and related isomers. The global model of duodenal total FA flow (Table 1) depended on the

total FA intake. Duodenal C18:0 flow was dependent on C18:3 intake in the global and intra-experimental tests, without effects of total C18:2, total C18:1 and C18:0 intakes. The duodenal total C18:1 flow was positively dependent on total C18:1 and C18:3 intakes and negatively dependent on total C18:2 intake. Furthermore, the duodenal total C18:1

 Table 1
 Predictions of duodenal flows of total fatty acids (FA), selected saturated FA, total C16:1, C18:1, C18:2 and C18:3, cis and trans isomers of C18:1, C18:2 and C18:3, and odd- and branched-chain fatty acid (OBCFA) from forage diets in ovine and bovine species

Dependent variables (Y, g/kg DMI)	Independent variable(s) (<i>X</i> , g/kg DMI)	Regression	Intercept (β_0)	Coefficient (β_1)	RMSE	R ²	N _{exp}	N _{trt}
Total FA _{duo}	Total FA _{int}	Global	8.11 (1.68)	0.63 (0.09)	2.95	0.62		31
C14:0 _{duo}	C14:0 _{int}	Global	0.09 (0.03)	1.01 (0.15)	0.08	0.69		30
C16:0 _{duo}	C16:0 _{int}	Global	1.19 (0.33)	0.53 (0.08)	0.60	0.57		31
C18:0 _{duo}	C18:3 _{int}	Global	3.87 (0.77)	0.62 (0.08)	2.09	0.61		41
C18:0 _{duo}	C18:3 _{int}	Intra-exp	5.05 (1.24)	0.51 (0.15)	1.41	0.67	10	37
Total C16:1 _{duo}	Total C16:1 _{int}	Global		0.06 (0.02)	0.03	0.86		31
Total C18:1 _{duo}		Global	2.09 (0.43)		1.01	0.84		46
	Total C18:3 _{int}			0.17 (0.07)				
	Total C18:2 _{int}			-0.61 (0.21)				
	Total C18:1 _{int}			1.13 (0.20)				
Total C18:1 _{duo}		Intra-exp			0.45	0.97	13	43
	Total C18:3 _{int}			0.16 (0.05)				
	Total C18:1 _{int}			1.04 (0.28)				
<i>c</i> 9-C18:1 _{duo}	<i>c</i> 9-C18:1 _{int}	Global	-0.82 (0.40)	0.44 (0.03)	5.58	0.60		162
c9-C18:1 _{duo}	c9-C18:1 _{int}	Intra-exp		1.65 (0.71)	0.65	0.78	10	34
c11-C18:1 _{duo}	Total C18:1 _{int}	Intra-exp		0.06 (0.02)	0.02	0.73	9	31
c12-C18:1 _{duo}	Total C18:2 _{int}	Global	0.015 (0.004)	0.006 (0.001)	0.01	0.49		21
c15-C18:1 _{duo}	Total C18:3 _{int}	Global		0.019 (0.003)	0.04	0.79		26
c15-C18:1 _{duo}	Total C18:3 _{int}	Intra-exp	0.05 (0.02)	0.005 (0.003)	0.02	0.94	7	24
t9-C18:1 _{duo}	Total C18:1 _{int}	Global	0.036 (0.004)	0.005 (0.002)	0.01	0.14		26
<i>t</i> 10-C18:1 _{duo}	Total C18:2 _{int}	Intra-exp		0.06 (0.02)	0.04	0.37	6	21
<i>t</i> 11-C18:1 _{duo}		Global			0.15	0.87		26
440	Total C18:3 _{int}			0.17 (0.02)				
	Total C18:2 _{int}			-0.31 (0.06)				
	Total C18:1 _{int}			0.50 (0.13)				
<i>t</i> 11-C18:1 _{duo}		Intra-exp			0.13	0.90	7	26
440	Total C18:3 _{int}			0.19 (0.02)				
	Total C18:2 _{int}			-0.33 (0.06)				
	Total C18:1 _{int}			0.54 (0.13)				
<i>t</i> 12-C18:1 _{duo}	Total C18:3 _{int}	Intra-exp		0.012 (0.002)	0.01	0.69	5	19
t13 + t14-C18:1 _{duo}	Total C18:3 _{int}	Intra-exp		0.03 (0.01)	0.05	0.67	4	14
<i>t</i> 15-C18:1 _{duo}	Total C18:3 _{int}	Global	0.08 (0.02)	0.008 (0.003)	0.05	0.25		21
<i>t</i> 15-C18:1 _{duo}	Total C18:3 _{int}	Intra-exp		0.016 (0.003)	0.02	0.82	6	19
Total C18:2 _{duo}	Total C18:2 _{int}	Global		0.15 (0.01)	0.24	0.85		30
Total C18:2 _{duo}	Total C18:2 _{int}	Intra-exp		0.16 (0.04)	0.12	0.95	8	30
c9c12-C18:2 _{duo}	c9c12-C18:2 _{int}	Global		0.09 (0.02)	0.15	0.37		31
c9c12-C18:2 _{duo}	c9c12-C18:2 _{int}	Intra-exp		0.11 (0.03)	0.08	0.79	7	26
t11t13-CLA _{duo}	Total C18:3 _{int}	Intra-exp		0.0024 (0.0004)	0.003	0.92	5	14
<i>t</i> 11 <i>c</i> 15-C18:2 _{duo}	Total C18:3 _{int}	Global		0.016 (0.005)	0.05	0.39		14
<i>t</i> 11 <i>c</i> 15-C18:2 _{duo}	Total C18:3 _{int}	Intra-exp		0.018 (0.005)	0.04	0.56	5	14
Total C18:3duo	Total C18:3 _{int}	Global		0.09 (0.02)	0.36	0.46		44
c9c12c15-C18:3 _{duo}	c9c12c15-C18:3 _{int}	Global	0.57 (0.16)	0.06 (0.02)	0.19	0.88		26
OBCFA								
C15:0 _{duo}	A/P	Global		0.293 (0.093)	0.10	0.60		7
lso-C15:0 _{duo}	Total FA _{int} $ imes$ total FA _{int}	Global	0.142 (0.017)	0.0002 (0.00004)	0.04	0.51		23
Anteiso-C15:0 _{duo}	Total $FA_{int} \times total FA_{int}$	Global	0.240 (0.035)	0.0003 (0.00008)	0.08	0.51		20
C17:0 _{duo}	Total $FA_{int} \times total FA_{int}$	Global	0.160 (0.013)	0.00014 (0.00003)	0.03	0.51		23
Iso-C17:0 _{duo}	NDF	Global	-0.049 (0.009)	0.00022 (0.00002)	0.004	0.94		10
Anteiso-C17:0 _{duo}	NDF	Global	-0.102 (0.014)	0.00039 (0.00003)	0.006	0.96		10

DMI = dry matter intake; RMSE = root mean square error; N_{exp} = number of experiments; N_{trt} = number of treatments; duo = duodenal flows; int = intake; Intra-exp = intra-experiment; A/P = acetate/propionate ratio.

Data were represented by the mean (standard error).

flow was dependent on intra-variation in total C18:1 and total C18:3 intakes. The global and intra-experiment models of duodenal total C18:2 and *c*9*c*12-C18:2 flows depended on the intake of their respective FA. The conservation mode and forage families did not affect the models of duodenal total C18:1, total C18:2 and *c*9*c*12-C18:2 flows (data not shown). The duodenal total C18:3 flow was positively related to total C18:3 intake. The global model of duodenal *c*9*c*12*c*15-C18:3 flow included an effect of *c*9*c*12*c*15-C18:3 intake (Table 1). Forage families affected the model of duodenal *c*9*c*12*c*15-C18:3 flow, with a significantly higher slope for Leguminosae (0.20 ± 0.03) than for Gramineae (0.03 ± 0.01) (Figure 1).

The global and intra-experiment models of c15-, t12-, t13 + 14- and t15-C18:1 presented a significant effect of total C18:3 intake. For c15- and t15-C18:1, the intra-experiment model was a better fit than the global model (Table 1). The intra-experiment model for duodenal c11-C18:1 flow and the global model of duodenal t9-C18:1 flow was related to total C18:1 intake. The duodenal c12- and t10-C18:1 flows were positively correlated with total C18:2 intake in global and intra-experiments, respectively. The global and intra-experiment models of duodenal t11-C18:1 flow presented a positive effect of total C18:2 intake. Among the C18:2 isomers, the global and intra-experiment models of duodenal t11-C18:1 flow presented a notice effect of total C18:2 intake. Among the C18:2 isomers, the global and intra-experiment models of duodenal t11-C18:1 flow presented and intra-experiment models of duodenal t11-C18:2 isomers, the global and intra-experiment models of duodenal t11-C18:2 isomers, the global and intra-experiment models of duodenal t11-C18:2 isomers, the global and intra-experiment models of duodenal t11-C18:2 isomers, the global and intra-experiment models of duodenal t11-C18:2 isomers, the global and intra-experiment models of duodenal t11-C18:3 intake.

Models from the seed and oil/fat subsets

Duodenal flows of total fatty acids, C18:0, total C18:1, C18:2 and C18:3 and relative isomers. Duodenal total FA flow was significantly related to total FA intake (Table 2), with no difference between the subsets, suggesting that the physical form of lipid supplement had no influence (seed v. oil). For the model of duodenal C18:0 flow, the total C18:3, C18:2,



and C18:1, and C18:0 intakes had significant effects. For duodenal total C18:1 flow, there was a positive intraexperiment effect of total C18:3, C18:2 and C18:1 intakes (Table 2). For total C18:1 model, we obtained a single equation for the seed and oil/fat subset. In contrast, the model of duodenal c9-C18:1 flow depended on c9-C18:1 intake and the physical form affected duodenal flows with a lower slope for the seed than for the oil/fat subsets $(0.25 \pm 0.04 v. 0.32 \pm 0.03$, respectively, Table 2). The intraexperiment model of duodenal c11-C18:1 and t9-C18:1 flows was related to total C18:1 intake. The global model of duodenal *c*11-C18:1 flow presented a significant effect of total C18:1 intake and concentrate percentage for oil/fat subset. The duodenal *c*12-, *c*15-, *t*12-, *t*13 + *t*14-, *t*15-C18:1 flows were positively correlated with total C18:3 intake in intraexperiments. The intra-experiment model of duodenal t10-C18:1 flow presented a significant effect of the interaction between FA intake and concentrate percentage, but interaction between c9c12-C182 intake and either concentrate percentage or starch content of the diet produced lower root mean square error (RMSE). In these two models including c9c12-C18:2 intake, residual analysis indicated on average -0.20 and -0.25 g duodenal t10-C18:1/kg DMI for the lipids fed as seeds, respectively. The duodenal t11-C18:1 flow indicated a significant effect of both total C18:2 and C18:3 intakes. The duodenal total C18:2 and c9c12-C18:2 flows were positively related to their respective FA intakes.

The duodenal total C18:3 and c9c12c15-C18:3 flows depended on their respective FA intake, with slopes of 0.049 ± 0.004 (seed) and 0.035 ± 0.05 (oil/fat) for total C18:3 and 0.052 ± 0.003 (seed) and 0.035 ± 0.004 (oil/fat) for c9c12c15-C18:3, respectively. Concerning the models of duodenal total C18:3 and c9c12c15-C18:3 flows from oil/fat subset, formaldehyde treatment presented an effect on regression residuals. For the c9c12c15-C18:3 intake, the formaldehyde treatment yielded high residuals (0.59 g/kg DMI), but this was only due to two treatments (Figure 2), the Ca salts having no effect. The technological process had no effect on regression residuals for the model of c9c12-C18:2. Duodenal c9t11-CLA flow from the oil/fat subset depended on the total C18:2 intake (slope = 0.008 ± 0.002 , Table 2). We did not obtain a model from the seed subset, perhaps due to insufficient data. There was a significant effect of C18:3 intake on duodenal t11t13-CLA flow, with no effect of the physical form (seed v. oil/fat) on the slope. For duodenal t10c12-CLA flow (Table 2 and Figure 3), data of Kucuk *et al.* (2004) obtained with a combination of 3.2% to 9.4% soyabean oil in high-concentrate diet (80% DM) are clearly higher than all others values in the literature. Without these data, the model for duodenal t10c12-CLA flow included the interaction between FA intake and concentrate percentage. A better prediction may be obtained with the interaction of c9c12-C18:2 and concentrate percentage, but with a smaller number of treatments. In both cases, intercept was not significant, but the inclusion of the interaction in the models reduced the RMSE. Duodenal t11c15-C18:2 flow depended on total C18:3 intake, with no effect of the physical form. In addition, none of the

 Table 2
 Predictions of duodenal flows of total fatty acids (FA), selected saturated FA, total C16:1, C18:1, C18:2 and C18:3, cis and trans isomers of C18:1, C18:2 and C18:3, and odd- and branched-chain FA (OBCFA) from diets supplemented with seeds, vegetable oils or animal fats in ovine and bovine species

Dependent variables (Y, g/kg DMI)	Independent variable(s) (X, g/kg DMI)	Subset	Regression	Intercept (β_0)	Coefficient (β_1)	RMSE	R ²	N _{exp}	N _{trt}
Total FA _{duo}	Total FA _{int}	Seed/oil/fat	Intra-exp	6.87 (1.39)	0.87 (0.03)	6.27	0.93	57	152
C14:0 _{duo}	C14:0 _{int}	Oil/fat	Intra-exp	0.33 (0.04)	0.54 (0.05)	0.12	0.87	20	52
C16:0 _{duo}		Seed/oil/fat	Intra-exp	1.43 (0.19)		0.95	0.97	59	144
	Total C16:1 _{int}				1.07 (0.24)				
	C16:0 _{int}				0.79 (0.02)				
Total C16:1 _{duo}	Total C16:1 _{int}	Oil/fat	Intra-exp	0.14 (0.03)	0.14 (0.03)	0.09	0.72	13	34
C18:0 _{duo}		Seed/oil/fat	Intra-exp	6.48 (0.70)		3.06	0.94	104	267
	Total C18:3 _{int}				0.25 (0.05)				
	Total C18:2 _{int}				0.52 (0.05)				
	Total C18:1 _{int}				0.46 (0.04)				
T	C18:0 _{int}				0.62 (0.08)				
Total C18:1 _{duo}	T-+- C10-2	Seed/oil/fat	Intra-exp		0.20 (0.02)	1.98	0.87	95	232
	Total C18:3 _{int}				0.29 (0.03)				
	Total C18:2 _{int}				0.24 (0.03)				
-0 C10.1		Cood	Intro ovo		0.29 (0.03)	1 6 4	0.01	7	21
C9-C18:1 _{duo}	C9-C18:1 _{int}	Seed	Intra-exp		0.25 (0.04)	1.64	0.91	/	21
C9-C18:1 _{duo}	C9-C18:1 _{int}	Seed	Global		-0.09 (0.04)	0.13	0.95		14
					0.008 (0.001)				
	O C19:1 v starsh				0.047 (0.000)				
a C19-1	a (19.1	Oil/fat	Intra ovn			1 50	0 00	17	12
c11 C19.1	Total C19.2	UII/Iat Sood	Intra-exp	0.12 (0.04)	0.32 (0.03)	0.05	0.09	0	45
c11_C18.1	Total C10.2 _{int}	Sood	Global	0.12 (0.04)	0.013 (0.002)	0.05	0.90	0	25
cir-cio.i _{duo}		Seeu	Giongi		0.018 (0.002)	0.09	0.01		25
c11_C18·1	Total C18:1.	Oil/fat	Intra-ovn	0.21 (0.04)	-0.003 (0.001)	0 13	0 72	1/	36
c11_C18.1	Total C18:1	Oil/fat	Global	0.21 (0.04)	0.017 (0.004)	0.15	0.72	14	/12
	PCO	Olinat	Global	-0.20 (0.00)	0.012 (0.004)	0.10	0.00		42
c12-C18:1duo	Total C18:3int	Seed	Intra-exp	0.09 (0.03)	0.007 (0.002)	0.05	0.77	3	10
c12-C18:1 _{duo}		Oil/fat	Intra-exp		(0.002)	0.07	0.85	12	29
	Total C18:3int				0.009 (0.002)				
	Total C18:2 _{int}				0.016 (0.002)				
<i>с</i> 15-С18:1 _{duo}	Total C18:3 _{int}	Seed/oil/fat	Intra-exp		0.025 (0.004)	0.19	0.7	9	21
<i>t</i> 9-C18:1 _{duo}	Total C18:1 _{int}	Seed/oil/fat	Intra-exp		0.018 (0.005)	0.1	0.3	11	32
<i>t</i> 10-C18:1 _{duo}	Total C18:2 _{int}	Seed	Intra-exp		0.037 (0.006)	0.15	0.92	5	14
<i>t</i> 10-C18:1 _{duo}	Total C18:2 _{int}	Oil/fat	Intra-exp		0.15 (0.03)	0.61	0.92	6	13
<i>t</i> 10-C18:1 _{duo}	$PCO \times total FA_{int}$	Seed/oil/fat	Intra-exp		7.6 <i>E</i> ⁻⁴ (1.1 <i>E</i> ⁻⁴)	1.30	0.75	16	43
<i>t</i> 10-C18:1 _{duo}	PCO × c9c12-C18:2 _{int}	Seed/oil/fat	Intra-exp		1.8 <i>E</i> ⁻³ (0.2 <i>E</i> ⁻³)	1.10	0.80	16	43
<i>t</i> 10-C18:1 _{duo}	Starch × c9c12-C18:2 _{int}	Seed/oil/fat	Intra-exp		5.8 <i>E</i> ⁻⁴ (1.0 <i>E</i> ⁻⁴)	1.03	0.84	8	19
<i>t</i> 11-C18:1 _{duo}		Seed/oil/fat	Intra-exp			0.98	0.82	25	69
	Total C18:3 _{int}				0.12 (0.02)				
	Total C18:2 _{int}				0.07 (0.02)				
<i>t</i> 11-C18:1 _{duo}	Total C18:3 _{int}	Seed/oil/fat	Global		0.08 (0.02)	1.30	0.76		43
	Total C18:2 _{int}				0.16 (0.03)				
	Intake level				-0.93 (0.26)				
	Starch				0.005 (0.002)				
<i>t</i> 12-C18:1 _{duo}	Total C18:3 _{int}	Seed	Intra-exp	0.07 (0.02)	0.023 (0.001)	0.03	0.98	3	10
<i>t</i> 12-C18:1 _{duo}		Oil/fat	Intra-exp		/	0.1	0.9	12	27
	Total C18:3 _{int}				0.018 (0.003)				
	Total C18:2 _{int}				0.020 (0.003)				_
<i>t</i> 13 + <i>t</i> 14-C18:1 _{duo}	Total C18:3 _{int}	Seed	Intra-exp		0.093 (0.005)	0.14	0.98	2	7
$t_{13} + t_{14} - C_{18:1_{duo}}$	Total C18:3 _{int}	Oil/tat	Intra-exp		0.07 (0.01)	0.38	0.89	4	8
715-C18:1 _{duo}	IOTAL CI8:3 _{int}	Uil/fat	intra-exp	1 10 (0 10)	0.035 (0.004)	0.13	0.92	6	12
	10tal C18:2 _{int}	Seed/oil/fat	intra-exp	1.19 (0.16)	0.09 (0.01)	0.69	0.85	59	158
C9CIZ-C18:2duo	C9C12-C18:2 _{int}	Seed/oil/tat	Intra-exp	0.87 (0.09)	0.07 (0.01)	0.31	0.93	23	66
(9612-018:2 _{duo}	८७८। २-८। ४:2 _{int} Starch	Seed/oil/fat	GIODAI		0.06 (0.01) 0.0045 (0.0008)	0.55	0.87		36

Dependent variables (<i>Y</i> , g/kg DMI)	Independent variable(s) (<i>X</i> , g/kg DMI)	Subset	Regression	Intercept (β_0)	Coefficient (β_1)	RMSE	R ²	N _{exp}	N trt
c9t11-CLA _{duo}	Total C18:2 _{int}	Oil/fat	Intra-exp		0.008 (0.002)	0.11	0.6	12	33
<i>t</i> 11 <i>t</i> 13-CLA _{duo}	Total C18:3 _{int}	Seed/oil/fat	Intra-exp		0.0033 (0.0002)	0.01	0.94	7	17
<i>t</i> 11 <i>t</i> 13-CLA _{duo}	Total C18:3 _{int}	Seed/oil/fat	Global	0.14 (0.05)	0.0045 (0.0003)	0.02	0.79		52
	EMPS				-0.006 (0.002)				
<i>t</i> 10 <i>c</i> 12-CLA _{duo}	Total C18:2 _{int} × total C18 :2 _{int}	Seed/oil/fat	Intra-exp		0.00007 (0.00001)	0.02	0.88	17	47
<i>t</i> 10 <i>c</i> 12-CLA _{duo}	$PCO \times total FA_{int}$	Seed/oil/fat	Intra-exp	-0.005 (0.004)	1.2 <i>E</i> ⁻⁵ (1.8 <i>E</i> ⁻⁶)	0.017	0.68	22	66
<i>t</i> 10 <i>c</i> 12-CLA _{duo}	PCO × c9c12-C18:2 _{int}	Seed/oil/fat	Intra-exp		2.1 <i>E</i> ⁻⁵ (2.2 <i>E</i> ⁻⁶)	0.016	0.71	20	59
<i>t</i> 11 <i>c</i> 15-C18:2 _{duo}	Total C18:3 _{int}	Seed/oil/fat	Intra-exp		0.018 (0.005)	0.56	0.66	7	17
Total C18:3 _{duo}	Total C18:3 _{int}	Seed	Intra-exp	0.26 (0.06)	0.049 (0.004)	0.15	0.94	9	28
Total C18:3 _{duo}	Total C18:3 _{int}	Oil/fat	Intra-exp	0.28 (0.05)	0.035 (0.004)	0.17	0.89	18	44
<i>c</i> 9 <i>c</i> 12 <i>c</i> 15-C18:3 _{duo}	<i>c</i> 9 <i>c</i> 12 <i>c</i> 15-C18:3 _{int}	Seed	Intra-exp	0.28 (0.04)	0.052 (0.003)	0.09	0.98	5	17
<i>с</i> 9 <i>с</i> 12 <i>с</i> 15-С18:3 _{duo}	<i>c</i> 9 <i>c</i> 12 <i>c</i> 15-C18:3 _{int}	Oil/fat	Intra-exp	0.27 (0.05)	0.035 (0.004)	0.16	0.90	15	36
OBCFA									
C15:0 _{duo}	A/P	Seed/oil/fat	Global	0.59 (0.06)	-0.07 (0.02)	0.09	0.22		48
lso-C15:0 _{duo}		Seed/oil/fat	Global	0.27 (0.09)		0.03	0.62		27
	DMI				0.0003 (0.0001)				
	A/P				-0.080 (0.015)				
Anteiso-C15:0duo	A/P	Seed/oil/fat	Global	1.30 (0.14)	-0.29 (0.04)	0.10	0.57		31
lso-C16:0 _{duo}	A/P	Seed/oil/fat	Global	-0.09 (0.04)	0.001 (0.0003)	0.03	0.46		21
C17:0 _{duo}		Seed/oil/fat				0.11	0.37		62
ado	СР				0.0017 (0.0005)				
	Total FA _{int}				0.0035 (0.0006)				
lso-C17:0 _{duo}		Seed/oil/fat	Global	0.12 (0.02)		0.02	0.46		21
	Intake level				-0.020 (0.007)				
	Total FA _{int}				0.0007 (0.0002)				
Anteiso-C17:0 _{duo}	Intake level	Seed/oil/fat	Global	0.32 (0.34)	-0.07 (0.01)	0.04	0.50		28

Table 2 (Continued)

DMI = dry matter intake; RMSE = root mean square error; $N_{exp} =$ number of experiments; $N_{trt} =$ number of treatments; duo = duodenal flows; int = intake; Intra-exp = intraexperiment; RPB = rumen protein balance; PCO = percentage of concentrate; EMPS = efficiency of microbial protein synthesis; A/P = acetate/propionate ratio. Data were represented by the mean (standard error).



Figure 2 Intra-experimental relationships between the ruminant duodenal flow of c9c12c15-C18:3 and c9c12c15-C18:3 intake from the oil/fat subset, where (\blacklozenge) are the control treatments without vegetable oils or animal fats, (\blacktriangle) free lipids, (\bullet) calcium salts and (\blacksquare) formaldehydeprotected lipids. DMI = dry matter intake.

selected potential factors had a significant effect on the regression residuals (data not shown).

Odd- and branched-chain fatty acids. The duodenal C15:0 flow was correlated with DMI, A/P ratio, OMDr, and EMPS and ruminal pH (Supplementary Table S6). Among these variables, A/P ratio had significant effect on the duodenal

C15:0 flow (Table 2). The duodenal iso-C15:0 and anteiso-C15:0 flows were negatively related on A/P (Supplementary Table S6). The duodenal iso-C16:0 flow was dependent negatively on NDF content and positively on CP content (Supplementary Table S6). The duodenal C17:0 flow was positively affected by starch content and concentrate percentage, and negatively with DMI, NDF content, A/P ratio and ruminal pH (Supplementary Table S6). For duodenal iso-C17:0 and anteiso-C17:0 flows, intake level had a negative effect on both FA, whereas FA intake had a positive effect on duodenal iso-C17:0 flow (Table 2).

Potential factors affecting duodenal flows of isomers of C18:1, C18:2 and C18:3 from the seed and oil/fat subsets. The duodenal c9-C18:1 flow from seed subset was positively correlated with starch content, concentrate percentage and total FA intake, and negatively correlated with intake level, A/P ratio and ruminal pH (Supplementary Table S7), c9-C18:1 intake and starch content \times c9-C18:1 intake interaction (Table 2). For duodenal c9c12-C18:2 flow from the seed/oil/ fat subset, we observed in the global relationship an effect of starch when associated with c9c12-C18:2 intake. We obtained global and intra-experiment regressions with close intercepts and slopes between the two models. The duodenal



Figure 3 (a) Global relationship between ruminant duodenal flows of *t*10*c*12-CLA from the seed/oil/fat subset and the product between total dietary fatty acid intake (total FA_{int}, *g*/kg dry matter intake (DMI)) and percentage of concentrate (PCO, g/100 g DMI). Data of Kucuk *et al.* (2004) are indicated as black squares. (b) Within-experiment relationship between ruminant duodenal flows of *t*10*c*12-CLA from the seed/oil/fat subset and the product between total FA_{int} (g/kg DMI) and PCO (g/100 g DMI), with the exclusion of data from Kucuk *et al.* (2004) ($Y = -0.005 + 1.2E^{-5}X$, number of treatments = 66, root mean square error = 0.017, $R^2 = 0.68$).

c9c12c15-C18:3 flow from the seed subset was not affected by any factor other than c9c12c15-C18:3 intake. For the oil/ fat subset, none of the correlated factors presented an effect on the duodenal c9c12c15-C18:3 flow when associated with c9c12c15-C18:3 intake (Table 2). The concentrate percentage was correlated with the duodenal flows of c11-, t9-, and t15-C18:1, t10c12-CLA, t11c15-C18:2 and c9c12-C18:2 (Supplementary Table S7). The duodenal c11-C18:1 flow was a function of total C18:1 intake and concentrate percentage from the oil/fat subset (Table 2). The intake level was correlated with the duodenal flows of c9-, c11- (seed subset), t11-, and t13 + t14-C18:1 and t10c12-CLA (seed/oil/fat subset) and positively correlated with t13/t14-18:1 (oil/fat subset) (Supplementary Table S7). The duodenal t11-C18:1 flow from the seed/oil/fat subset was positively dependent on total C18:3 and C18:2 intakes and dietary starch but negatively on intake level (Table 2 and Supplementary Table S7). Furthermore, duodenal t10c12-CLA flow from the seed/ oil/fat subset was affected negatively by concentrate percentage and total FA intake interaction (Table 2). The RPB had a negative effect on duodenal c11-C18:1 flow from the seed subset, whereas the total C18:2 intake had a positive effect (Table 2 and Supplementary Table S7). Only the duodenal t11t13-CLA flow was a function of EMPS when associated with total C18:3 intake (Table 2).

Models from the fish subset

This section deals with duodenal flows of total FA, C18:0, *cis* and *trans* C18:1 isomers, EPA, DPA and DHA. The duodenal total FA flow was predicted, taking into account total FA intake (Table 3). Duodenal C18:0 flow was positively related to total C18:3 intake and negatively to DHA intake. Duodenal *t*11-C18:1 flow was positively dependent on total C18:3 and C18:2, and DHA intakes. Duodenal *t*10-C18:1 flow was negatively correlated to total C18:3 intake and positively to C18:2 and DHA intakes. The duodenal DPA flow had a quadratic relationship with DPA intake and the model for duodenal DHA flow (Figure 4) presented a linear effect of DHA intake × DHA intake, with significant slopes (0.10 ± 0.02 and -0.020 ± 0.006 , respectively).

Absorbed fatty acid prediction

Absorbed total FA and C18:0 flows were linearly related to their respective duodenal flows (Table 4). Similarly, the absorbed flows of total C18:1, *c*9-C18:1, total C18:2, *c*9*c*12-C18:2 and LCFA (EPA + DPA + DHA) depended on their corresponding duodenal FA flow. The model of absorbed *c*9*c*12-C18:2 flow had a slope value higher than that of absorbed total C18:2 flow (1.02 v. 0.70, respectively), as well as a lower RMSE (0.10 v. 0.27 g/kg DMI, respectively). For the models of absorbed total C18:3 and *c*9*c*12*c*15-C18:3 flows, duodenal total C18:3 and *c*9*c*12*c*15-C18:3 flows had a significant effect, respectively.

Evaluated models

The evaluation procedures were only applied to models from the seed and oil/fat subsets because of the amount of available data. We obtained 15 evaluated models from the 32 performed models from the seed and oil/fat subsets (Supplementary Table S8). In all, 10 of the 15 models (c9and *t*13 + *t*14-C18:1, total C18:3, *c*9*c*12*c*15-C18:3 from the seed subset; c9- and t13 + t14-C18:1 from the oil/fat subset; and total FA, total C18:2, c9c12-C18:2 and t9-C18:1 from the seed/oil/fat subset) presented regression coefficients close to the expected value of 1. For the c15-C18:1 and t10c12-CLA models (from the seed/oil/fat subset), the regression coefficient was <1, and for total C18:3 and c9c12c15-C18:3 (from the oil/fat subset) and t11t13-CLA (from the seed/oil/fat subset) the regression coefficients were higher than 1. The regression between observed duodenal flows of total FA and c9c12c15-C18:3 and predicted duodenal flows of these FA

Dependent variables (<i>Y</i> , g/kg DMI)	Independent variable(s) (X, g/kg DMI)	Intercept (β_0)	Coefficient (β_1)	RMSE	R ²	N _{exp}	N _{trt}
Total FA _{duo}	Total FA _{int}	11.46 (3.55)	0.78 (0.07)	5.80	0.85	7	33
C14:0 _{duo}	C14:0 _{int}	0.33 (0.05)	0.46 (0.04)	0.18	0.86	12	38
C16:0 _{duo}				0.69	0.98	13	40
	Total C16:1 _{int}		0.59 (0.16)				
	C16:0 _{int}		0.91 (0.03)				
C18:0 _{duo}		20.1 (1.22)		2.85	0.92	13	40
	Total C18:3 _{int}		0.29 (0.09)				
	DHA _{int} ¹		-6.38 (0.63)				
C18:0 _{duo}		16.2 (1.95)		2.74	0.93	11	35
	Total C18:0 _{int}		5.25 (1.29)				
	LCFA _{int} ²		-2.26 (0.23)				
<i>t</i> 10-C18:1 _{duo}				0.51	0.88	6	21
	Total C18:3 _{int}		-0.08 (0.02)				
	Total C18:2 _{int}		0.06 (0.02)				
	DHA _{int}		0.57 (0.15)				
<i>t</i> 10-C18:1 _{duo}				0.56	0.86	6	21
	Total C18:3 _{int}		-0.07 (0.02)				
	Total C18:2 _{int}		0.07 (0.03)				
	LCFA _{int}		0.21 (0.06)				
<i>t</i> 11-C18:1 _{duo}				1.25	0.86	7	24
	Total C18:3 _{int}		0.20 (0.05)				
	Total C18:2 _{int}		0.12 (0.05)				
	DHA _{int}		2.16 (0.35)				
<i>t</i> 11-C18:1 _{duo}				1.21	0.87	7	24
	Total C18:3 _{int}		0.25 (0.05)				
	Total C18:2 _{int}		0.17 (0.05)				
	LCFA _{int}		0.88 (0.14)				
<i>c</i> 9-C18:1 _{duo}	<i>c</i> 9-C18:1 _{int}	1.24 (0.26)	0.11 (0.03)	0.32	0.74	7	24
<i>c</i> 9 <i>c</i> 12-C18:2 _{duo}	c9c12-C18:2 _{int}	1.64 (0.22)	0.04 (0.01)	0.31	0.93	4	14
Total C18:3 _{duo}	Total C18:3 _{int}	0.51 (0.07)	0.039 (0.004)	0.15	0.93	6	19
<i>c</i> 9 <i>c</i> 12 <i>c</i> 15-C18:3 _{duo}	c9c12c15-C18:3 _{int}	0.49 (0.07)	0.039 (0.004)	0.15	0.93	6	19
EPA _{duo} ³	EPA _{int}	0.05 (0.01)	0.05 (0.01)	0.04	0.81	11	32
DPA _{duo} ⁴	$DPA_{int} \times DPA_{int}$	0.034 (0.006)	0.78 (0.08)	0.03	0.84	11	34
DHA _{duo}		0.031 (0.006)		0.02	0.84	13	39
	DHA _{int}		0.10 (0.02)				
	$DHA_{int} \times DHA_{int}$		-0.020 (0.006)				

 Table 3 Predictions of duodenal flows of total fatty acids (FA) and selected saturated, monounsaturated and polyunsaturated FA from diets

 supplemented with fish products in ovine and bovine species

DMI = dry matter intake; RMSE = root mean square error; $N_{exp} =$ number of experiments; $N_{trt} =$ number of treatments; duo = duodenal flows; int = intake; LCFA = long-chain fatty acid; DPA = docosapentanoic acid.

Data were represented by the mean (standard error).

¹DHA (C22:6n-3). ²LCFA = sum of EPA + DPA + DHA.

³EPA (C20:5n-3).

⁴DPA (C22:5n-3).

from the performed models are described in the Figures 5 and 6, respectively.

Concerning the MSPE parameters, errors due to disturbance accounted for the majority of the errors in 12 models (*c*9- and *t*13 + *t*14-C18:1, and total C18:3 from the seed subset; *c*9- and *t*13 + *t*14-C18:1, total C18:3, *c*9*c*12*c*15-C18:3 and from the oil/fat subset; and total FA, *t*9-C18:1, total C18:2, *c*9*c*12-C18:2 and *t*10*c*12-CLA from the seed/oil/ fat subset). In the models of duodenal flows of *c*15-C18:1 and *t*11*t*13-CLA from seed/oil/fat subset, the majority of the MSPE was allocated to errors due to regression. When the errors due to disturbance accounted for the majority of MSPE, we observed that regression between predicted and observed data presented a good fit with the Y = X model.

Discussion

The separation of the database into different subsets was essential to highlight the FA digestion according to the nature of lipid supplements or forages. For the seed, oil/fat and fish subsets, the intra-experiment relationships were close to the

global ones, indicating that the prediction of these flows was probably not biased. For the forage subset, these relationships were sometimes different, probably because the conservation mode and forage families, were confounded with *c9c12c15*-C18:3 and *c9c12*-C18:2 intakes, respectively. Previous metaanalysis (Glasser *et al.*, 2008; Schmidely *et al.*, 2008) were compared with the present models for total FA, total C18:2, total C18:3 and *c9c12*-C18:2. Overall, the independent variables are the same between the previous models and those proposed in this study, with the exception of *c9c12c15*-C18:3 which was not calculated by Glasser *et al.* (2008). In addition, despite currently having more data, the regression coefficients of the previous and current models are very similar.



Figure 4 Intra-experimental relationships between ruminant duodenal DHA flows and DHA intakes. The bold dashed line shows the quadratic fitted model ($Y = 0.031 \pm 0.006 + 0.10 \pm 0.02 \cdot X - 0.020 \pm 0.006X^2$, number of treatments = 39, root mean square error = 0.02, $R^2 = 0.84$).

Forage subset

Overall slope of the relationship between duodenal flow of total FA and total FA intake was apparently low (0.63) but this is mainly due to the significant intercept of that relationship (8.11 g FA/kg DMI), suggesting that microbial synthesis may contribute largely to the duodenal flow of total FA. It can, however, not be ruled out that methodological considerations during lipid extraction may affect that relationship (Christie and Han, 2010). The RBH of c9c12c15-C18:3 was on average 94%, similar to the mean of 92.7% observed by Doreau et al. (2005), with forage families and conservation mode confounded. Moreover, the different slopes of the duodenal c9c12c15-C18:3 flow models between forage families may be explained by the presence of red clover silage in the diets offered to cattle, a Leguminosae rich in polyphenol oxidase with a likely inhibitory effect on RBH (Van Ranst et al., 2011). However, this difference in RBH



Figure 5 Relationship between observed ruminant duodenal total fatty acid (FA) flows and predicted duodenal total FA flows from the seed/oil/ fat subset ($Y = 0.99 \pm 0.04 \cdot X$, number of treatments = 147, root mean square error = 7.96, $R^2 = 0.77$). DMI = dry matter intake.

Table 4 Predictions of absorbed fatty acid (FA) flows from duodenal flows of total FA, selected saturated FA, total and isomers of C18:1, C18:2 and C18:3, and long-chain FA in ovine and bovine species

Dependent variables (Y, g/kg DMI)	Independent variable(s) (<i>X</i> , g/kg DMI)	Intercept (μ)	Coefficient (β_1)	RMSE	R ²	Nexp	N _{trt}
		1 4 2	4.0			chp	
Total FA _{abs}	Total FA _{duo}	4.29 (0.97)	0.65 (0.02)	3.90	0.96	80	208
C14:0 _{abs}	C14:0 _{duo}	-0.10 (0.01)	0.92 (0.03)	0.03	0.97	32	84
C16:0 _{abs}	C16:0 _{duo}		0.75 (0.02)	0.66	0.97	71	180
C18:0 _{abs}	C18:0 _{duo}	5.25 (0.54)	0.49 (0.02)	2.14	0.96	68	172
Total C18:1 _{abs}	Total C18:1 _{duo}		0.79 (0.03)	0.22	0.87	43	121
c9-C18:1 _{abs}	<i>c</i> 9-C18:1 _{duo}		0.88 (0.01)	0.15	0.99	22	63
Total C18:2 _{abs}	Total C18:2 _{duo}	0.42 (0.11)	0.70 (0.34)	0.27	0.97	71	182
c9c12-C18:2abs	c9c12-C18:2 _{duo}	-0.28 (0.06)	1.02 (0.03)	0.10	0.99	10	32
Total C18:3 _{abs}	Total C18:3 _{duo}		0.76 (0.02)	0.07	0.98	58	149
c9c12c15-C18:3abs	c9c12c15-C18:3 _{duo}		0.92 (0.02)	0.04	0.96	12	35
EPA _{abs} ¹	EPA _{duo}		0.95 (0.01)	0.01	0.99		16
DPA _{abs} ²	DPA _{duo}		0.95 (0.02)	0.003	0.99		5
DHA _{abs} ³	DHA _{duo}		0.96 (0.04)	0.01	0.98		10

DMI = dry matter intake; RMSE = root mean square error; $N_{exp} =$ number of experiments; $N_{trt} =$ number of treatments; abs = absorbed flows; duo = duodenal flows; DPA = docosapentanoic acid.

Data were represented by the mean (standard error).

¹EPA (C20:5n-3).

²DPA (C22:5n-3).

³DHA (C22:6n-3).



Figure 6 (a) Relationship between observed and predicted ruminant duodenal flows of *c*9*c*12*c*15-C18:3 from the seed subset ($Y = 0.82 \cdot X$, number of treatments (N_{trt}) = 114, root mean square error (RMSE) = 0.27, $R^2 = 0.57$). (b) Relationship between observed and predicted duodenal flows of *c*9*c*12*c*15-C18:3 from the oil/fat subset (g/kg of dry matter intake (DMI)) ($Y = 1.29 \cdot X$, $N_{trt} = 94$, RMSE = 0.26, $R^2 = 0.60$).

between Leguminosae and Gramineae could be rather attributed to a modification of the bacteria activity with a decrease in microbial protein synthesis when grass silage was replaced by red clover silage (Halmemies-Beauchet-Filleau *et al.*, 2013). No effect of the conservation mode on the c9c12c15-C18:3 RBH was found in our study, in disagreement with Doreau *et al.* (2005), where the RBH of c9c12c15-C18:3 was more extensive (+6.4%) for fresh grass than hay. The lack of conservation mode effect in our subset could be explained by the fact that low and high intakes of c9c12c15-C18:3 were associated to hay and fresh grass, respectively (Supplementary Table S2).

Seed and oil/fat subsets

For seed and oil/fat subsets, the regression coefficients of total duodenal C18:3 and duodenal c9c12c15-C18:3 on their respective intake were not different. Glasser et al. (2008) found a slope of 0.05 ± 0.01 for duodenal total C18:3, representing 95% of RBH. The c9c12c15-C18:3 model from the seed subset presented a RBH lower than the oil/fat model with 94.8% and 96.5%, respectively, but this difference is relatively low. This slightly limited RBH is probably due to seed hulls delaying the c9c12c15-C18:3 release for the microbes. The models of C18:2 isomers (c9t11-CLA, t11t13-CLA, t10c12-CLA and t11c15-C18:2) were independent of the physical form of lipid supplements (seed v. oil/fat), and therefore, this factor does not seem to affect the first RBH step. The t11c15-C18:2 and t11t13-CLA were produced during the c9c12c15-C18:3 RBH (Shingfield et al., 2010a; Ferlay et al., 2017). Moreover, the duodenal t11c15-C18:2 flow is greater than that of t11t13-CLA, which could be explained by the fact that t11c15-C18:2 comes from a major pathway of linolenic acid RBH, whereas t11t13-CLA comes from a minor pathway (Shingfield et al., 2010a).

The models for the different C18:1 isomers as a function of total C18:2 and total C18:3 intakes, had slopes <0.1, with the exception of c9-, t11- and t10-C18:1, indicating that these three FA are major isomers of C18:1 at the duodenum (Table 2). Indeed, t11-C18:1 is the main C18:1 isomer, originating both from c9c12-C18:2 and c9c12c15-C18:3 RBH

(Harfoot and Hazlewood, 1997). Furthermore, *c*9-C18:1 could be isomerised to *t*11-C18:1. However, we did not find any effect of *c*9-C18:1 intake on duodenal *t*11-C18:1 flow, suggesting that this isomerisation rate is low and/or rapid (Laverroux *et al.*, 2011). For the *t*10-C18:1 model, the duodenal flow was higher with oil/fat than with the seed form. This could be linked to the rate of oil release from raw seed, which is possibly more gradual than directly from the oil (Chilliard *et al.*, 2009). The low slope of the models is valuable for the two subsets forage and seed/oil/fat, suggesting similarities of formation of isomers of C18:1 during the RBH pathway whatever the dietary ingredient (forage or oleaginous).

Our models indicated that the main variable driving the duodenal flows of t10-C18:1 and t10c12-CLA is the combination of total FA intake (or c9c12-C18:2 intake) and percentage of concentrate. More specifically, we quantified the specific effect of starch intake which seems to be a major determinant of the duodenal flows of these FA when associated with c9c12-C18:2 intake. This interaction reflects the changes in microbial population within the rumen associated when high starch with high unsaturated lipid diets are fed, leading to the preferential development of bacterial species such as Propionibacterium acnes (Devillard et al., 2006) and/ or Megasphaera elsdenii (Kim et al., 2002) that are known to synthesise t10c12-CLA from c9c12-C18:2. Although different *in vitro* studies have shown an appearance of *t*10-C18:1 from c9c12c15-C18:3 (Ferlay et al., 2017), we found no effect of c9c12c15-C18:3 intake on duodenal t10-C18:1 flow in our meta-analysis.

We proposed predictive equations for the duodenal OBCFA flows, with a positive effect of starch intake on duodenal C17:0 flow, a positive effect of NDF content on duodenal C15:0 flow, a negative effect of concentrate percentage on duodenal iso-C16:0 flow and a negative effect of A/P on duodenal iso-C15:0 and anteiso-C15:0 flows (Table 3). These effects could be related to bacterial metabolism as increasing starch content in the diet favours amy-lolytic bacterial activity, whereas the NDF content increases that of cellulolytic bacteria (Vlaeminck *et al.*, 2006).

Vlaeminck *et al.* (2015) demonstrated positive relationships between amylolytic bacteria and anteiso-, linear and oddchain FA and between cellulolytic bacteria and iso-FA. In addition, Belanche *et al.* (2012) demonstrated a positive effect of high dietary NDF on duodenal OBCFA flows. We found a positive effect of RPB on duodenal flows of C15:0 and iso-C16:0, in agreement with Vlaeminck *et al.* (2015). An increasing intake level is associated with a decrease in OMDr and thus a decrease in availability of nutrients to bacteria, thereby explaining the negative effects of intake level on duodenal iso- and anteiso-C17:0 flows.

Fish subset

The DPA had a lower RBH than EPA and DHA (67%, 95% and 94%, respectively), in agreement with Kairenius *et al.* (2011). These latter authors suggest that RBH of EPA or DHA involves the initial removal of the double bond between carbon atoms 4 and 5, whereas DPA does not have a double bond in this position, explaining a lower RBH of DPA compared with EPA and DHA. Nevertheless, in a recent *in vitro* study, Aldai *et al.* (2018) indicate that firstly the *cis*-13 double bond of DHA migrates to the adjacent position 14 reversing its geometry from *cis* to *trans*, forming the *trans*-14, *cis*-16 system.

Other models showed inhibitory or positive effects of DHA on the different steps of C18 RBH. The DHA intake as well as the LCFA intake had a negative effect on the duodenal C18:0 flow and a positive effect on duodenal *t*10- and *t*11-C18:1 flows, in agreement with Lourenco *et al.* (2010) and Shing-field *et al.* (2010b), that reported that EPA and DHA inhibited both the last step of RBH and the bacteria growth, increasing the *trans* FA and CLA content in the rumen. This effect was not found in our study with EPA or DPA intake, although they were correlated with DHA intake.

In conclusion, this meta-analysis proposes predictive equations for different FA that have not been predicted previously: isomers derived from RBH that affect animal lipid metabolism, notably the milk fat depression; OBCFA, which are specific bacterial FA used for diagnostic of rumen function, and EPA, DPA and DHA from fish products. These predictive equations add to the body of quantitative knowledge on FA digestion. However, the effects of different dietary factors (e.g. lipid source, conservation mode of forage and forage families) have not been demonstrated in our models due to low amounts of available data. In the current context, understanding and controlling the ruminal digestion of FA, and the effects of different qualitative and quantitative diet variations, is essential to improve the quality of ruminant products (meat and milk).

Acknowledgements

The authors are grateful to C. Gérard and B. Delord that funded this meta-analysis study through Neovia and Limagrain brands, respectively, which are greatly acknowledged for their financial support to Lucas de Ofeu Aguiar Prado CIFRE scholarship. In addition, the authors would like to thank Patrick Chapoutot from INRA-UMR0791 Mosar team for granting us permission to work on the Systool model and its help on feedstuff characterisation.

Declaration of interest

No conflict of interest to declare.

Ethics statement

Not relevant.

Software and data repository resources

The AGRum database has been deposited at Inter Deposit Digital Number (https://www.iddn.org/) under the reference (IDDN.FR.001.510032.000.R.C.2011.000.10300).

Supplementary material

To view supplementary material for this article, please visit https://doi.org/10.1017/S1751731118001982

References

Aldai N, Delmonte P, Alves SP, Bessa RJB and Kramer JKG 2018. Evidence for the initial steps of DHA biohydrogenation by mixed ruminal microorganisms from sheep involves formation of conjugated fatty acids. Journal of Agricultural Food and Chemistry 66, 842–855.

Baumont R, Dulphy JP, Sauvant D, Tran G, Meschy F, Aufrère J, Peyraud JL and Champciaux P 2007. Les tables de la valeur des aliments. In Alimentation des bovins, ovins et caprins - Besoins des animaux - Valeurs des aliments – Tables INRA 2007, pp. 181–275. Editions Quae, Versailles, France.

Belanche A, Doreau M, Edwards JE, Moorby JM, Pinloche E and Newbold CJ 2012. Shifts in the rumen microbiota due to the type of carbohydrate and level of protein ingested by dairy cattle are associated with changes in rumen fermentation. Journal of Nutrition 142, 1684–1692.

Bibby J and Toutenburg H 1977. Prediction and improved estimation in linear models. John Wiley & Sons, Berlin, Germany. p. 201.

Chapoutot P, Martin O, Nozière P and Sauvant D 2015. Systool Web, a new one-line application for the French INRA « systali » project. In Proceedings of the 66th Annual Meeting of the European Society of Animal Science, 31 August to 4 September, Wageningen Academic Publishers, Warsaw, Poland, p. 265.

Chilliard Y, Glasser F, Ferlay A, Bernard L, Rouel J and Doreau M 2007. Diet, rumen biohydrogenation and nutritional quality of cow and goat milk fat. European Journal of Lipid Science and Technology 109, 828–855.

Chilliard Y, Martin C, Rouel J and Doreau M 2009. Milk fatty acids in dairy cows fed whole crude linseed, extruded linseed, or linseed oil, and their relationship with methane output. Journal of Dairy Science 92, 5199–5211.

Christie WW and Han X 2010. Lipid extraction, storage and sample handling. In Lipid analysis, isolation, separation, identification and lipidomic analysis (Chapter 3, ed. PJ Barnes and Associates), 4th edition pp. 55–66. The Oily Press, Bridgwater, UK.

Devillard E, McIntosh FM, Newbold CJ and Wallace RJ 2006. Rumen ciliate protozoa contain high concentrations of conjugated linoleic acids and vaccenic acid, yet do not hydrogenate linoleic acid or desaturate stearic acid. British Journal of Nutrition 96, 697–704.

Doreau M, Lee MRF, Ueda K and Scollan ND 2005. Métabolisme ruminal et digestibilité des acides gras des fourrages. In Proceedings of the 12th Rencontres Recherches Ruminants, 7 and 8 December, Paris, France, pp. 101–104.

Ferlay A, Bernard L, Meynadier A and Malpuech-Brugère C 2017. Production of trans and conjugated fatty acids in dairy ruminants and their putative effects on human health: a review. Biochimie 141, 107–120.

Fievez V, Colman E, Castro-Montoya JM, Stefanov I and Vlaeminck B 2012. Milk odd- and branched-chain fatty acids as biomarkers of rumen function - an update. Animal Feed Science and Technology 172, 51–65.

Glasser F, Schmidely P, Sauvant D and Doreau M 2008. Digestion of fatty acids in ruminants: a meta-analysis of flows and variation factors: 2. C18 fatty acids. Animal 2, 691–704.

Halmemies-Beauchet-Filleau A, Vanhatalo A, Toivonen V, Heikkilä T, Lee MRF and Shingfield KJ 2013. Effect of replacing grass silage with red clover silage on ruminal lipid metabolism in lactating cows fed diets containing a 60:40 forage-to-concentrate ratio. Journal of Dairy Science 96, 5882–5900.

Harfoot CG and Hazlewood GP 1997. Lipid metabolism in the rumen. In The rumen microbial ecosystem (ed. PN Hobson and CS Stewart), pp. 382–426. Chapman and Hall, London, UK.

INRA 2010. Alimentation des bovins, ovins et caprins - Besoins des animaux - Valeurs des aliments -Tables INRA 2007 - mise à jour 2010. Editions Quae, Paris, France.

Kairenius P, Toivonen V and Shingfield KJ 2011. Identification and ruminal outflow of long-chain fatty acid biohydrogenation intermediates in cows fed diets containing fish oil. Lipids 46, 587–606.

Kim YJ, Liu RH, Rychlik JL and Russell JB 2002. The enrichment of a ruminal bacterium (*Megasphaera elsdenii* YJ-4) that produces the *trans*-10, *cis*-12 isomer of conjugated linoleic acid. Journal of Applied Microbiology 92, 976–982.

Kucuk O, Hess BW and Rule DC 2004. Soybean oil supplementation of a highconcentrate diet does not affect site and extent of organic matter, starch, neutral detergent fiber, or nitrogen digestion, but influences both ruminal metabolism and intestinal flow of fatty acids in limit-fed lambs. Journal of Animal Science 82, 2985–2994.

Laverroux S, Glasser F, Gillet M, Joly C and Doreau M 2011. Isomerization of vaccenic acid to *cis* and *trans* C18:1 isomers during biohydrogenation by rumen microbes. Lipids 46, 843–850.

Lourenço M, Ramos-Morales E and Wallace RJ 2010. The role of microbes in rumen lipolysis and biohydrogenation and their manipulation. Animal 4, 1008–1023.

Maxin G, Glasser F, Doreau M and Baumont R 2013. Prévision de la teneur en matières grasses et de la composition en acides gras des fourrages. In Proceedings

of the 20th Rencontres Recherches Ruminants, 4 and 5 December, Paris, France, pp. 49–52.

Moher D, Liberati A, Tetzlaff J and Altman DG 2009. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. Public Library of Science Medicine 6, 264–269.

Sauvant D and Nozière P 2013. Integrative model of the digestive tract including the interactions involved in energy and protein digestion in ruminants. In Energy and protein metabolism and nutrition in sustainable animal production (ed. JW Oltjen, E Kebreab and H Lapierre), pp. 317–318. Wageningen Academic Publishers, Wageningen, The Netherlands.

Sauvant D, Schmidely P, Daudin JJ and St-Pierre NR 2008. Meta-analyses of experimental data in animal nutrition. Animal 2, 1203–1214.

Schmidely P, Glasser F, Doreau M and Sauvant D 2008. Digestion of fatty acids in ruminants: a meta-analysis of flows and variation factors. 1. Total fatty acids. Animal 2, 677–690.

Shingfield KJ, Bernard L, Leroux C and Chilliard Y 2010a. Role of trans fatty acids in the nutritional regulation of mammary lipogenesis in ruminants. Animal 4, 1140–1166.

Shingfield KJ, Lee MRF, Humphries DJ, Scollan ND, Toivonen V, Reynolds CK and Beever DE 2010b. Effect of incremental amounts of fish oil in the diet on ruminal lipid metabolism in growing steers. British Journal of Nutrition 104, 56–66.

Van Ranst G, MRF L and Fievez V 2011. Red clover polyphenol oxidase and lipid metabolism. Animal 5, 512–521.

Vlaeminck B, Fievez V, Cabrita ARJ, Fonseca AJM and Dewhurst RJ 2006. Factors affecting odd- and branched-chain fatty acids in milk: a review. Animal Feed Science and Technology 13, 389–417.

Vlaeminck B, Gervais R, Rahman MM, Gadeyne F, Gorniak M, Doreau M and Fievez V 2015. Postruminal synthesis modifies the odd- and branched-chain fatty acid profile from the duodenum to milk. Journal of Dairy Science 98, 4829–4840.