

Biogeographical Patterns of Species Richness and Abundance Distribution in Stream Diatoms Are Driven by Climate and Water Chemistry

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Manuscript

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Biogeographical patterns of species richness and abundance

Distribution in stream diatoms are driven by climate and water chemistry

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Abstract

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This is the first inter-continental comparative assessment of diatom richness and species abundance distribution (SAD) along spatial, climatic, and water chemistry gradients using stream data from the US and Finland. We tested a number of hypotheses about the causes of variability in richness and by extension, the SAD, including the metabolic theory, the species-energy, the more individuals, the climatic variability, and the climatic tolerance hypotheses. We found no support for any of these concepts, given that the relationships of richness with explanatory variables were non-existent, weak or unexpected. Diatom richness and the SAD responded more strongly to climatic variability, climatic extremes, and water chemistry, especially nutrient levels, than to mean temperature. A consequence of the weak energy dependence was a non-linear latitudinal gradient of richness in both countries, and a non-linear latitudinal gradient of the SAD in the US. The SAD was not entirely driven by richness but displayed distinct responses to spatial, climatic, and water chemistry factors, indicating that the environment had complementary effects on richness and the SAD. This emphasizes the utility of the SAD for ecological research—a notion that has been previously debated. Thus, we found sufficient support for the operation of multiple climate and water chemistry mechanisms in structuring diatom communities, which underscores their complex response to the environment and the necessity for novel predictive frameworks.

Introduction

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The spatial variability in species richness along latitudinal, longitudinal, and elevational gradients has drawn a continued interest among ecologists for centuries, beginning with the works of Alexander von Humboldt and Carolus Linnaeus (Pianka 1966; Gaston 2000; Willig et al. 2003; Rahbek 2005; Mittelbach et al. 2007). There are numerous hypotheses about the nearly universal poleward decline in species richness with ecological and evolutionary rates, energy, and climatic favorability and stability emerging as important underlying factors. The elevation-richness relationship is more variable, most frequently conforming to a monotonically declining or a hump-shaped form, but there are some commonalities in the driving forces of the latitudinal and elevational gradients (Rahbek 2005). Thus, warmer temperatures in tropical and low elevation regions shorten the generation times and accelerate the metabolic and mutation rates, leading to greater speciation (Rohde 1992; Currie et al. 2004; Allen et al. 2006). These ideas were conceptualized within the framework of the Metabolic Theory of Ecology (MTE), which predicted species richness (S) as a function of temperature (T in Kelvin), according to the equation $ln(S) = -E_A/kT$ + I, where E_A = activation energy with an expected value of about -0.65, k = Boltzmann constant (8.62 × 10^{-5} eV K⁻¹), and I = intercept (Allen et al. 2002; Brown et al. 2004). The species-energy theory proposed that areas with greater total available energy, such as the tropics, have more diverse communities because they can support larger species populations with lower likelihood of extinction (Wright 1983). This theory is also known as the more individuals hypothesis (MIH), because larger species populations result in a greater total number of individuals (Srivastava and Lawton 1998). The MIH predicts that richness scales positively with energy because richness is a function of the number of individuals, which is proportional to the available energy, i.e. communities are energylimited. Further elaboration of the species-energy theory recognized that temporal variability in energy may directly impact the richness of an area, given that periods of low energy are marked by higher extinction rates, and consequently, by a lower number of individuals. Thus, models that included total energy and energy variability explained the variability in bird and mammal species richness better than either energy variable alone (Carrara and Vazquez 2010).

Finally, according to the climatic tolerance hypothesis, the tropics harbor greater richness because their more benign warm and humid conditions fall within the physiological tolerance of many more species compared to the cold and dry extra-tropical regions (Currie et al. 2004). However, the latitudinal species patterns can be complicated by longitudinal effects, such as an east-west heterogeneity in rainfall, which is particularly distinct at mid-latitudes (Terborgh 1973).

While the spatial variability of species richness has been extensively studied, we know substantially less about the spatial and environmental dependence of the species abundance distribution (SAD), defined here as the number of individuals across species in a community. In fact, due to a more theoretical and statistical emphasis in the study of the SAD over the past seventy years (May 1975; McGill et al. 2007; Ulrich et al. 2010), the empirical latitudinal, longitudinal, and elevational patterns of the SAD and their underlying mechanisms, including environmental variability, have remained largely unknown.

It is possible that the latitudinal gradient of richness is paralleled by a corresponding gradient of the SAD. Brown (2014) suggested that as richness increases toward lower latitudes, the SAD may transition from a less even log linear (a few very abundant and some very rare species) to a more even curvilinear pattern (greater numbers of both intermediate and low abundance species) as a result of intensified enemy effects. There is evidence that the SADs of real forest communities do become less even at high latitudes when compared to neutrally assembled communities due to strong environmental filtering that favors only a few well-adapted species (Qiao et al. 2015). However, there is no broad empirical support for a tendency in the SAD toward greater evenness and lognormality at low latitudes. On the contrary, a recent study of forest trees showed the opposite trend—preponderance of less even logseries SADs at low latitudes but more even lognormal SADs at high latitudes, possibly as a result of overall greater dispersal in the tropics, and subsequent accumulation of rare species (Ulrich et al. 2016a). A global study of dryland plant communities reported an overall prevalence of lognormal compared to log-series SADs, which was linked to environmental variability and stress but not to latitude (Ulrich et al. 2016b). There are even fewer and generally terrestrial studies of the SAD along elevational gradients,

which either have not reported any distinct patterns (Ulrich et al. 2016a; 2016b) or shown a transition from log-series to lognormal SADs with altitude (Arellano et al. 2017). Therefore, there is clearly a need for further research on the large-scale spatial patterns of the SAD, especially in aquatic ecosystems, which have been largely neglected in this context.

Our overall goal was to explore the large scale spatial patterns in diatom species richness and abundance distribution and determine if they are driven by the same environmental factors and whether these factors are climatic, including average temperature, seasonal and diurnal temperature fluctuation, and extreme temperatures, or water chemistry, i.e. nutrient levels, pH, and specific conductance.

Anthropogenic eutrophication is changing aquatic ecosystems globally by altering physico-chemistry, increasing primary production, and affecting species richness, composition and abundance with potentially strong ecological and evolutionary consequences (Smith and Schindler 2009; Alexander et al. 2017). A comprehensive review of the freshwater literature concluded that water chemistry variables were stronger predictors of diatom community structure than temperature (Soininen 2007). However, climatic factors outperformed water chemistry in predicting the distributions of individual diatom species across northern latitudes, e.g. Finland (Pajunen et al. 2016). Therefore, the roles of climate vs. water chemistry and their interaction with spatial factors in defining diatom community properties are still unresolved and merit further research.

Here, we examined the shape of the SAD in freshwater diatom communities by calculating the standard deviation (parameter σ) of a Poisson lognormal distribution model and skewness. Parameter σ indicates how equitably abundances are distributed across abundance classes with lower values suggesting higher equitability. Skewness measures the symmetry of the SAD compared to a lognormal distribution. Negative values (left skew) indicate prevalence of rare species, while positive values (right skew), greater frequency of abundant species.

We tested the following theories, hypotheses, and predictions. First, we tested whether $ln(S) \sim -E_A/kT$ with $-E_A$ between -0.6 and -0.7, as postulated by the MTE (Brown et al. 2004). We predicted that if this relationship holds, then parameter σ of the SAD would scale positively with E_A/kT ($\sigma \sim$

 E_A/kT), given that $\sigma \sim -\ln(S)$, as empirically shown for freshwater diatoms (Passy 2016). Second, we tested the predictions of the MIH that richness (S) increases with energy (here temperature, T) because S is a positive function of the number of individuals (N), which in turn, is proportional to the amount of energy (Srivastava and Lawton 1998). However, since the effect of T on S is indirect through N, the expectation is that the correlations between T and N and S, would be equal to or greater than the correlation between T and S (Currie et al. 2004; Carrara and Vazquez 2010). Given the above consideration about the parameter σ -ln(S) relationship, we also predicted that if the MIH operates, parameter σ would scale negatively with T and N, i.e. communities would become more even (lower σ) at higher levels of available energy and higher numbers of individuals. Third, we extended our reasoning to the energy variability and climatic tolerance hypotheses and tested if seasonal (and diurnal) energy fluctuation and climatic harshness, respectively, decrease S, but increase parameter σ . Fourth, considering the strong impact of water chemistry on freshwater ecosystems, we predicted that climatic variables alone would be insufficient to describe the species richness and SAD patterns, but a combination of climatic and water chemistry factors would be required. For our objectives 1-4, we assumed that if the shape of the SAD is purely defined by richness, then parameter σ would respond to the same environmental predictors as richness and in a complementary way, e.g. parameter σ would show positive responses to predictors eliciting a negative response in richness and vice versa. However, if the SAD has unique environmental underpinning, parameter σ may display distinct environmental responses. Finally, we examined the spatial effects on richness and the SAD and how they constrained the community responses to climate and water chemistry to gain a deeper understanding of the causes of large scale community variability.

Materials and Methods

Datasets

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US. Data on stream water chemistry and diatom composition were collected from 669 distinct steam localities in the US (fig. A1) by the National Water-Quality Assessment (NAWQA) Program (http://water.usgs.gov/nawqa). Diatoms were sampled from a defined area of hard substrate or

macrophytes. Water chemistry, including total phosphorus, nitrate + nitrite, ammonia, pH, and specific conductance, was measured for the month of algal collection (Table A1). Samples were taken in July and August from 1993 to 2009 along a latitudinal range of 36 degrees, longitudinal range of 83 degrees, and elevational range of 2676 m. Climatic variables, including mean annual temperature (T_{mean}), temperature seasonality (standard deviation, T_{SD}), mean diurnal temperature range (mean difference between monthly maximum and minimum temperatures, T_{diur}), minimum temperature of the coldest month (T_{min}), and maximum temperature of the warmest month (T_{max}), were obtained from the WorldClim database (Hijmans et al. 2005). Diatoms were identified in 590-700 cell counts and the total count was converted to total density (cells·cm $^{-2}$).

Finland. Data on climate (the same variables as in the US) and water chemistry, including total phosphorus, pH, and specific conductance, were available for 100 streams in Finland (fig. A1). Diatoms were sampled in July and August from 2001 to 2004 along a latitudinal range of 10 degrees, longitudinal range of 7 degrees, and elevational range of 302 m. Diatoms were identified in counts of about 500 cells.

Analysis of the species abundance distribution

Using count data, community SADs were fit with a Poisson lognormal distribution model, which is comparatively insensitive to sampling effort and performs equally well to other commonly used models (Sæther et al. 2013; Baldridge et al. 2016). We estimated the standard deviation, parameter σ , with the poilog R-package (Grøtan and Engen 2008). A comparison of the rank of the observed log likelihood with the log likelihood derived from 1000 bootstraps provided a goodness of fit (gof) metric. Values of gof between 0.05 and 0.95 indicate good fit. We also calculated the proportion of the species pool revealed by the sample, which represents the proportion of the total area of the fitted Poisson lognormal distribution.

We calculated the skewness (γ) of the log₂-transformed counts of individuals and the standard error of skewness (SES) as $(6/n)^{0.5}$. Skewness is considered significant if the absolute value of the ratio γ /SES is greater than 2 (SYSTAT Software, Inc. 2009).

Theory testing

We performed a series of regression analyses to test the outlined theories and hypotheses. The metabolic theory was tested with model 1 (Tables 1, 2) using ln(S) as a dependent variable. However, since the richness response to 1/kT can be curvilinear (Algar et al. 2007), in a second model (model 2, Tables 1, 2), we added a quadratic term. We further explored whether the predictions of the MTE can be extended to the SAD parameter σ , given its dependence on ln(S), and calculated models 1 and 2 for parameter σ as well (Tables 1, 2).

Testing the more individuals hypothesis required several equations, including models 3 and 4 from Table 1 (the dependent variable was richness), and equation (1) below, which contained a quadratic term to account for potential nonlinearity in the density (N) response to mean temperature (T_{mean}).

$$ln(N) = ln(T_{mean}) + ln(T_{mean})^2$$
 (1)

Our hypothesis that if the MIH operates, parameter σ would scale negatively with T and N, was tested with models 3 and 4 from Table 1. These ideas were tested only with the US dataset because it contained density data.

The climatic variability hypothesis, suggesting that variability in temperature would constrain richness, and consequently the SAD, was tested with model 5 from Tables 1 and 2. The climatic tolerance hypothesis, implicating extreme temperatures as driving factors of community properties, was tested with model 6 from Tables 1 and 2 for both richness and the SAD.

Additional models from Tables 1 and 2 were calculated in order to test the responses of richness and the SAD to climatic factors (model 7), nutrient (nitrogen and/or phosphorus) enrichment (model 8), overall water chemistry (model 9), climate and nutrient enrichment (model 10), and climate and overall water chemistry (model 11). The remaining models from Tables 1 and 2 determined the latitudinal (model 12), longitudinal (model 13), and elevational trends (model 14). Model 15 tested the overall spatial effect. For the SAD, model 16 estimated the effect of richness on parameter σ . For each multiple regression model in Tables 1 and 2, we performed backward selection of significant terms only to reduce redundancy and collinearity (Table A2). All models were compared using the Akaike Information

Criterion (AIC). Models with lower AIC provided better fits. Variance partitioning assessed to what extent the community responses to environmental factors were due to covariance of the environment with latitudinal, longitudinal, and elevational gradients. Specifically, we estimated how much of the variance explained by models 1-11 from Tables 1 and 2 was contributed by covariance of the respective predictors in these models with the predictors in model 15. For the SAD, variance partitioning using models 1-15 also determined whether the response of parameter σ to the predictors (environmental and spatial) in these models was in fact driven by their covariance with richness (model 16). The response of skewness to climatic and water chemistry predictors was examined with multiple regressions using the backward stepping procedure. Variance partitioning and multiple regressions were performed with SYSTAT 13 (SYSTAT Software, Inc. 2009).

Results

In both the US and Finnish datasets, the Poisson lognormal model fit the abundance data well $(0.36 \le \text{gof} \le 0.53)$. Parameter σ was a comparatively strong and negative function of ln-transformed richness (slope of -0.67 in the US, n = 669 and -0.83 in Finland, n = 100, P < 0.00001, Tables 1, 2), i.e. rich communities had lower parameter σ and were thus more equitable. The relationship of ln-species richness with the Boltzmann temperature factor (1/kT) was not significant in the US and convex in Finland, inconsistent with the prediction of the MTE (Tables 1, 2, models 1 and 2). Parameter σ exhibited a quadratic relationship with the Boltzmann temperature factor (more or less concave in both countries). Thus, our expectation for opposing responses of ln-richness and parameter σ to 1/kT was not well supported.

In testing the species-energy and the MIH, we examined the relationships among richness, mean temperature, and density in the US dataset (Table 1, models 3 and 4). In contrast with the expectations of these hypotheses, richness displayed a U-shaped response to mean temperature (Table 1, fig. 1) and a marginally non-significant decline with density (slope = -0.64, $R^2 = 0.01$, p = 0.06, n = 669), while the density-mean temperature relationship was hump-shaped ($\ln N = 12.98 + 0.04T_{mean} - 0.01T_{mean}^2$, $R^2 = 0.01$).

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0.04, P < 0.02, n = 667). Thus, richness was related weakly to mean temperature but not to density, indicating that the effect of temperature on richness was not due to a temperature dependence of density. Parameter σ was weakly related to mean temperature (Table 1, fig. 2) and density (parameter $\sigma = 1.86 +$ $0.03\ln(N)^2$, $R^2 = 0.03$, P < 0.00005, n = 669), exhibiting a U-shaped response to both. In Finland, the greatest richness was observed at intermediate temperature and the lowest parameter σ , at the highest temperature (Table 2, figs 3, 4). To test the role of climatic variability in defining community patterns, we examined the response of richness and parameter σ to seasonal and diurnal temperature fluctuation (Tables 1, 2, model 5, figs 1-4). In the US and Finland, temperature variability provided a better fit for both variables than mean temperature, as indicated by their lower AIC. With the exception of parameter σ in Finland, diatom richness and abundance equitability exhibited non-linear responses to diurnal and/or seasonal temperature variability (Table A2). The climatic tolerance hypothesis was tested using minimum (T_{min}) and maximum temperatures (T_{max}) as predictors of richness and parameter σ (Tables 1, 2, model 6, figs 1-4). Contrary to the expectation of diminished richness at the low temperature extreme, richness was the highest at both ends of the T_{min} gradient in the US and at lower T_{min} in Finland. Maximum temperature had a notable effect (quadratic) on richness only in Finland. In the US, parameter σ exhibited low values (high equitability) at low T_{min} and high T_{max} levels. In Finland, parameter σ declined with both T_{min} and T_{max} . From all sets of climatic variables, extreme temperatures provided the best fit in Finland, while temperature variability outperformed the other variables in the US, as revealed by a comparison of their respective AIC (Tables 1, 2, models 4-6). However, the model including all significant climatic variables (Table 1, model 7) explained substantially greater amount of the variance in both richness and parameter σ , indicating that these community properties were products of multiple climatic influences. In the US, all of the variance in richness and most of the variance in parameter σ explained by water chemistry was contributed by nutrient levels, which generally had a positive effect on richness and a negative effect on parameter σ (Table 1, models 8 and 9, figs 1, 2). Adding specific conductance to the

parameter σ model increased R² by only 3% (Tables 1, A2, model 9). In Finland, richness showed a unimodal response to total phosphorus, while parameter σ exhibited a marginally non-significant U-shaped response ($R^2 = 0.06$, p = 0.053). Predictability of richness improved by 5% after including pH in the model (Table 2, model 9). Notably, the effect of total phosphorus on parameter σ became significant and contributed 8% to the explained variance when the influence of specific conductance was accounted for in the full water chemistry model (Tables 2, A2, model 9). The overall best environmental model included climatic and nutrient variables only in the US but climatic, nutrient, and water chemistry variables in Finland and explained 21-40% and 25-30% of the variance in richness and parameter σ , respectively (Tables 1, 2, A2, model 11). However, a model with climatic and nutrient variables only provided comparable fits in Finland (Tables 2, A2, model 10).

The individual responses of richness and parameter σ to latitude, longitude, and elevation (Table 1, models 12-14) were weaker in the US (2-19% explained variance) but stronger in Finland (5-22% explained variance). Although the spatial relationships were nonlinear, the richest communities in the US were found at lower latitudes and elevations, and at westernmost and intermediate longitudes. The most equitable communities (lowest parameter σ) in the US were observed at intermediate latitudes, longitudes, and elevations. In Finland, richness was the greatest at intermediate latitudes, extreme longitudes, and higher elevations, while parameter increased with latitude and responded unimodally to longitude and elevation. Nevertheless, the model including all spatial factors provided the best fit for both richness and parameter σ , capturing respectively 16-26% of the variance in the US and 18-24% in Finland (Tables 1, 2, model 15).

We asked next if the behavior of richness and the SAD along climatic and water chemistry gradients (Tables 1, 2, models 1-11) was driven by spatial variability of these gradients. To answer this question, we calculated the covariance of environmental variables (Tables 1, 2, models 1-11) with spatial predictors (Tables 1, 2, model 15) using variance partitioning. The amount of variance in richness and parameter σ explained by individual predictor sets, e.g. mean temperature, temperature fluctuation, extreme temperature, nutrients, and water chemistry, was generally due to covariance of the environment

with space (i.e., spatially structured environment). In more complex models, including a combination of predictor sets, e.g. models 7, 10, and 11 (Tables 1, 2), spatially structured environment explained only a portion of the variance. For parameter σ , which had a comparatively strong negative correlation with richness, we also assessed whether environment and space contributed to the explained variance independently of richness. Variance partitioning showed that in general, a substantial portion of the explained variance in parameter σ was captured by the environment and space after the richness effect was controlled for.

Skewness was positive in all communities in both the US and in Finland indicating a prevalence of common species. Of the skewness values, 50% were significant (γ /SES > 2) in the US and 71% in Finland. Skewness was either negatively (Pearson r = -0.21, P < 0.0001, the US) or non-significantly (Finland) related to richness. In both countries, skewness was positively correlated with parameter σ (Pearson r = 0.42-0.46, P < 0.0001), i.e. equitable communities were more symmetric with lower skewness and these communities tended to be rich in the US. Skewness increased at intermediate T_{SD} and low pH in the US and at intermediate specific conductance and T_{diur} but low total phosphorus in Finland (Table 3).

Discussion

We examined the behavior of both diatom richness and abundance distribution along environmental and spatial gradients in streams from two continents, testing a number of specific hypotheses. The response of richness to the Boltzmann temperature factor and mean temperature contradicted the MTE and the species-energy hypotheses, respectively, predicting a positive relationship (both hypotheses) with a specific rate of increase (MTE). Deviations from the predictions of the MTE were previously reported for both aquatic and terrestrial macroorganisms, where the richness-temperature relationship was found to be curvilinear, linear with a slope significantly different from predictions, or not significantly different from zero (Algar et al. 2007; Hawkins et al. 2007; Pinel-Alloul et al. 2013). In microbes, including lake phytoplankton and soil bacteria, the richness response to temperature was also variable, following

respectively, a segmented (Segura et al. 2015) or linear pattern, flatter than observed in macrobes (Zhou et al. 2016). A systematic analysis of multiple datasets across a broad range of terrestrial macroorganisms, testing the predictions of the MTE, found that the richness-temperature relationship was positive in datasets that included areas with colder winters but non-existent or negative in datasets from tropical, subtropical, and warm temperate regions (Hawkins et al. 2007). A review of the species-energy relationship noted that it is scale-dependent and transitions from unimodal at small scales to monotonically increasing at large scales (Evans et al. 2005). Here, the relationship of richness with the Boltzmann temperature factor and mean temperature was non-significant or weak in the US, but comparatively stronger and unimodal in Finland, even though both datasets covered regional to continental scales and included cold climate streams. The SAD response to the Boltzmann temperature factor and mean temperature was weak and U-shaped in the US, but stronger and monotonic in Finland, where inequality (parameter σ) decreased with temperature.

The present results were also inconsistent with the more individuals hypothesis, which so far, has received mixed support (Srivastava and Lawton 1998; Kaspari et al. 2000; Mönkkönen et al. 2006; Šímová et al. 2011). The hump-shaped behavior of density along the temperature gradient in the US dataset indicated that higher temperatures actually decrease the number of individuals, which is in stark contrast with the MIH prediction for a positive relationship. This pattern could be a consequence of intensified grazing at higher temperatures due to accelerated herbivore metabolic and consumption rates (O'Connor and Bruno 2009; West and Post 2016). Finally, the richness-density relationship was non-significant, indicating that the temperature effect on richness was not indirect through density. These findings suggest that i) the effect of temperature on both richness and the SAD is weak and may depend on latitude; ii) richness and the SAD may exhibit different and not necessarily complementary behavior along the temperature gradient; and iii) an overriding influence of factors other than temperature, given the non-linear density response to temperature.

Temperature variability and temperature extremes were better descriptors of both richness and parameter σ than mean temperature, but did not cause the responses predicted by the energy variability

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and the climatic tolerance hypotheses, respectively. High richness was observed at both low and high seasonality in the US and high seasonality Finland in contrast with the energy variability hypothesis, which proposes that energy reduction during the cold seasons will lead to lower richness. This result can be partially explained from the perspective of coexistence theory, which postulates that temporal heterogeneity in resource supply increases biodiversity because it allows coexistence of species that alternate between dominance and persistence at different times (Sommer 1985; Tilman and Pacala 1993). In both the US and Finland, richness was high at the lowest minimum temperature, contrary to the expectation of the climatic tolerance hypothesis. It is possible that diatoms deviate from the expectation of this hypothesis because they inhabit the comparatively milder aquatic environment, less prone to extreme fluctuations. Conversely, in terrestrial habitats, where temperatures reach much greater extremes, the climatic tolerance hypothesis was supported (Šímová et al. 2011). Furthermore, low temperatures may favor diatoms over cyanobacteria (Peterson and Grimm 1992; Van der Grinten et al. 2005) or green algae (Roberts et al. 2003); therefore, lower temperature minima and stronger temperature seasonality may prevent diatom exclusion and promote higher biodiversity of this algal group. High seasonality, low temperature minima, and low temperature maxima were also associated with the highest abundance equality (lowest parameter σ) in the US, which suggests more equitable resource partitioning. These results indicate that a decrease in the temporal variability of temperature and an increase in the temperature minima due to global warming (Xu et al. 2013), can lead to lower biodiversity and abundance equitability in some stream diatoms.

Although climate had a stronger influence on both richness and parameter σ in the US and Finland, water chemistry captured a notable amount of variance. Nutrients explained 8-14% of the variance of richness and parameter σ , while specific conductance or pH added \leq 5%, indicating that nutrient availability was a more important source of community variability, especially in the US dataset, which comprised primarily non-acidic steams. Richness was generally higher at intermediate or high nutrient values, while equality in the SAD increased with nutrients. An increase of diatom richness with nutrient supply has been previously documented and explained with the ability of more functional groups

(e.g., tolerant and sensitive to nutrient limitation) to coexist at high nutrients (Passy 2008; Soininen et al. 2016). Here, we further report that the SAD was also constrained by nutrient supply and in the US, it was even more responsive than richness. A recent study on the SAD of stream diatom communities, sampled across a land use gradient, revealed that their equitability increased (parameter σ decreased) with the transition from forest to agriculture and suggested that nutrient enrichment was responsible for this pattern (Passy 2016). In the present investigation we found support for this hypothesis and show that communities indeed became more even at higher nutrient supply across a broad range of latitudes. Nevertheless, the best model for richness and parameter σ included both climatic and water chemistry variables. These variables explained substantial proportion of the variance in richness and parameter σ independently of space, indicating that the environment was an important determinant of community organization. While a combined effect of nutrients and temperature on microbial biodiversity has been reported before (Soininen et al. 2016; Wang et al. 2016), this study is the first to demonstrate that nutrients and climate co-controlled the variability in the diatom SAD at a continental scale.

Skewness was positive in both datasets in contrast with a large body of literature, documenting negative skew as the dominant pattern (Gregory and Gaston 2000; Hubbell 2001; Magurran and Henderson 2003). This discrepancy may be due to both sampling intensity and environmental influences. Thus, when too few individuals are sampled, the left side of the SAD remains veiled and the right side exhibits a positive skew (McGill 2003). Although in both datasets a comparatively large number of individuals was sampled following standard protocols, the proportion of species revealed by the sample had a median of 44-76%, indicating that some species remained veiled. However, skewness had detectable correlations with environmental factors (R² = 0.09-0.20), pointing to ecological constraints on the SAD symmetry as well. Skewness responded most strongly to temperature variability and pH in the US, while water chemistry and to a lesser extent temperature variability were the underlying factors in Finland. Thus, temperature variability proved to be a key driver of richness and the SAD, including its evenness and symmetry. Determining the mechanisms controlling the distribution of common species, i.e.

the SAD symmetry, has important practical implications, given that these species are primarily responsible for delivering ecosystem services (Winfree et al. 2015).

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In both the US and Finland, diatom richness did not conform to the classical pattern of monotonic latitudinal decrease, consistent with prior diatom studies (Passy 2010; Soininen et al. 2016). These studies attributed the deviation from the classical pattern to the overriding effect of wetlands and their impact on micronutrient availability. Here the non-linear response of richness to latitude was associated with a nonlinear response of seasonality, which was one of the major predictors of richness. This suggests that temperature variability, in addition to micronutrient availability, may underlie the latitudinal richness distributions. The expectation for increased evenness in the SAD at low latitudes (Brown 2014) was confirmed only in Finland, while in the US, the most equitable communities were observed at mid latitudes. The elevational response of richness—bimodal in the US but peaking at high elevations in Finland—did not follow the common monotonic decline or hump-shaped patterns. Admittedly, the elevational gradient in Finland was short and not sufficient to reveal the full variability in richness. The SAD was most even at intermediate elevations in the US and at lower elevations in Finland, but this response was weak. Thus, the latitudinal effects on diatom richness and the SAD became more prominent at high latitudes (Finland) compared to lower latitudes (the US). At high latitudes, elevation constrained richness as well, while the SAD remained weakly controlled by elevation across latitudes. These results can be explained with the generally weak correlation of richness and the SAD with mean temperature, which is a major covariate of both latitude and elevation. Notably, longitude had the strongest effect on the SAD in the US, explaining 19% of its variance. This finding may be due in part to the longitudinal dependence of total phosphorus with a peak at mid longitudes, where the SAD was most even (lowest parameter σ). Total phosphorus had virtually no relationships with either latitude or elevation. These finding reveal that richness and the SAD had unique responses to spatial variability, e.g. locations with low richness did not necessarily have low equality (high values of parameter σ). This means that richness and the SAD may have distinct environmental underpinning.

Diatom richness and parameter σ were positively correlated, but this correlation did not underlie the responses of parameter σ to environmental and spatial variables, as indicated by variance partitioning. In fact, the covariance of richness with these variables explained generally less than half of the SAD variance, implying that much of the SAD response was independent of richness. Furthermore, in the US, climate, water chemistry, and spatial models explained better the variance of parameter σ , while in Finland, the pattern was reversed and richness was explained better than parameter σ by the respective models. There were also differences in the environmental and spatial predictors that entered the regression models of richness and parameter σ (Table A2) as well as in the shape of the richness and parameter σ responses to common predictors (figs 1-4). For example, parameter σ displayed a monotonic decline with nutrients, while the richness response was non-linear, indicating that eutrophication has a differential effect on richness vs. abundance equality. The form of the SAD is strongly dependent on richness and total abundance (Locey and White 2013) and the usefulness of the SAD for ecological research has been questioned (Yen et al. 2013), in part because of its dependence on richness. The results presented here demonstrate that the abiotic environment has distinct impacts on richness and the SAD, and that the shape of the SAD can be predicted by climatic and water chemistry variables, emphasizing the utility of the SAD in unravelling ecological mechanisms. Although we did not detect scaling relationships between temperature and the SAD, we encourage future research on organisms that are more temperature dependent than diatoms, to explore whether such relationships exist and thus add to the body of knowledge on the temperature effects on community organization.

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Table 1. US diatoms. Regression models (Eqn) testing the responses of ln-transformed species richness, ln(S) (models 1, 2, and 15), species richness (models 3-15) and parameter σ (models 1-15) to environmental and spatial variables. Covariance of the selected predictors (X) in the respective model with i) the selected spatial variables in model 15 is given by $X \cap Spa$ and with ii) ln(S), by $X \cap ln(S)$. Covariance was calculated only for significant X. n = 669. AIC = Akaike Information Criterion. The AIC of the best environmental model is boldfaced. T_{mean} = mean annual temperature, T_{SD} = temperature seasonality (standard deviation), T_{diur} = mean diurnal temperature range, T_{min} = minimum temperature of the coldest month, T_{max} = maximum temperature of the warmest month, T_{max} = total phosphorus, T_{max} = specific conductance, T_{max} = latitude, T_{max} = longitude, T_{max} = elevation, and T_{max} = non-significant.

Eqn	Predictor set	Richness			SAD parameter σ			
		AIC	\mathbb{R}^2	X∩Spa	AIC	\mathbb{R}^2	X∩Spa	X∩ln(S)
1	1/kT		NS			NS		
2	$1/kT + (1/kT)^2$		NS		1245	0.02	0.02	-0.01
3	$ln(N) + ln(N)^2$		NS		1237	0.03	0.06	0.00
4	$T_{mean} + T_{mean}^2$	5387	0.01	0.01	1245	0.02	0.02	-0.01
5	$T_{SD} + T_{SD}^2 + T_{diur} + T_{diur}^2$	5331	0.09	0.08	1165	0.14	0.11	0.07
6	$T_{min} + T_{min}^2 + T_{max} + T_{max}^2$	5351	0.07	0.05	1194	0.09	0.09	0.04
7	$\begin{split} T_{mean} + T_{mean}^2 + T_{SD} + T_{SD}^2 + T_{diur} + \\ T_{diur}^2 + T_{min} + T_{min}^2 + T_{max} + T_{max}^2 \end{split}$	5299	0.15	0.07	1104	0.22	0.15	0.10
8	$TP + TP^2 + NO_x + NO_x^2 + NH_4 +$ NH_4^2	5323	0.11	0.07	1172	0.12	0.12	0.07
9	$TP + TP^2 + NO_x + NO_x^2 + NH_4 +$ $NH_4^2 + pH + pH^2 + Cond + Cond^2$	5323	0.11	0.07	1150	0.15	0.12	0.08
10	$T_{mean} + T_{mean}^2 + T_{SD} + T_{SD}^2 + T_{diur} +$ $T_{diur}^2 + T_{min} + T_{min}^2 + T_{max} + T_{max}^2 +$ $TP + TP^2 + NO_x + NO_x^2 + NH_4 +$	5259	0.21	0.10	1080	0.25	0.18	0.12
11	NH_{4}^{2} $T_{mean} + T_{mean}^{2} + T_{SD} + T_{SD}^{2} + T_{diur} + T_{diur}^{2} + T_{min} + T_{min}^{2} + T_{max} + T_{max}^{2} + T_{ma$	5259	0.21	0.10	1080	0.25	0.18	0.12

12	$Lat + Lat^2 + Lat^3$	5384	0.02	_	1241	0.02	_	-0.01
13	$Long + Long^2 + Long^3$	5352	0.06	_	1114	0.19	_	0.10
14	$Elev + Elev^2 + Elev^3$	5375	0.03	_	1241	0.02	_	0.00
15	$Lat + Lat^{2} + Lat^{3} + Long + Long^{2} +$ $Long^{3} + Elev + Elev^{2} + Elev^{3}$	5287	0.16	_	1061	0.26	_	0.13
16	ln(S)	_	_	_	1024	0.29	0.13	_

Table 2. Finnish diatoms. Regression models (Eqn) testing the responses of ln-transformed species richness, ln(S) (models 1, 2, and 15), species richness (models 3-15) and parameter σ (models 1-15) to environmental and spatial variables. Covariance of the selected predictors (X) in the respective model with i) the selected spatial variables in model 15 is given by X \cap Spa and with ii) ln(S), by X \cap ln(S). n = 100. The AIC of the best environmental model is boldfaced. Abbreviations as in Table 1.

Eqn	Predictor set	R	cichness			SAD parameter σ		
		AIC	\mathbb{R}^2	X∩Spa	AIC	\mathbb{R}^2	X∩Spa	$X \cap ln(S)$
1	1/kT		NS		144	0.07	0.07	-0.03
2	$1/kT + (1/kT)^2$	68	0.07	0.05	144	0.07	0.07	-0.03
3	$ln(N) + ln(N)^2$	-	_	_	_	-	_	_
4	$T_{mean} + T_{mean}^2$	827	0.07	0.06	144	0.07	0.07	-0.03
5	$T_{SD} + {T_{SD}}^2 + {T_{diur}} + {T_{diur}}^2$	814	0.21	0.18	136	0.16	0.13	0.06
6	$T_{min} + T_{min}^2 + T_{max} + T_{max}^2$	812	0.23	0.12	133	0.18	0.17	0.07
7	$\begin{split} T_{mean} + T_{mean}^2 + T_{SD} + T_{SD}^2 + T_{diur} + \\ T_{diur}^2 + T_{min} + T_{min}^2 + T_{max} + T_{max}^2 \end{split}$	801	0.34	0.23	128	0.25	0.17	0.11
8	$TP + TP^2$	819	0.14	0.13		NS		
9	$TP + TP^2 + pH + pH^2 + Cond +$ $Cond^2$	815	0.19	0.13	140	0.13	0.09	0.03
10	$\begin{split} T_{mean} + T_{mean}^2 + T_{SD} + T_{SD}^2 + T_{diur} + \\ T_{diur}^2 + T_{min} + T_{min}^2 + T_{max} + T_{max}^2 + \\ TP + TP^2 \end{split}$	797	0.36	0.22	126	0.30	0.18	0.14
11	$\begin{split} T_{mean} + T_{mean}^2 + T_{SD} + T_{SD}^2 + T_{diur} + \\ T_{diur}^2 + T_{min} + T_{min}^2 + T_{max} + T_{max}^2 + \\ TP + TP^2 + pH + pH^2 + Cond + \\ Cond^2 \end{split}$	792	0.40	0.22	125	0.29	0.16	0.09
12	$Lat + Lat^2 + Lat^3$	813	0.20	-	134	0.18	_	0.06

$Long + Long^2 + Long^3$	812	0.22	_	145	0.06	_	0.05
$Elev + Elev^2 + Elev^3$	813	0.20	_	146	0.05	_	0.00
$Lat + Lat^2 + Lat^3 + Long + Long^2 +$	809	0.24	_	134	0.18	_	0.06
$Long^3 + Elev + Elev^2 + Elev^3$							
ln(S)	_	_	_	128	0.21	0.06	_
	$Elev + Elev^{2} + Elev^{3}$ $Lat + Lat^{2} + Lat^{3} + Long + Long^{2} +$ $Long^{3} + Elev + Elev^{2} + Elev^{3}$	Elev + Elev ² + Elev ³ $Elev + Elev2 + Elev3$ $Lat + Lat2 + Lat3 + Long + Long2 + Elev3$ $Long3 + Elev + Elev2 + Elev3$	Elev + Elev ² + Elev ³ $Elev + Elev2 + Elev3$ $Elev + Elev2 + Lat3 + Long + Long2 + Elev3$ $Elev + Elev2 + Elev3$ $Elev + Elev2 + Elev3$	Elev + Elev ² + Elev ³ $813 0.20 -$ $Lat + Lat^{2} + Lat^{3} + Long + Long^{2} +$ $Long^{3} + Elev + Elev^{2} + Elev^{3}$ $809 0.24 -$	Elev + Elev ² + Elev ³ 813 0.20 - 146 Lat + Lat ² + Lat ³ + Long + Long ² + 809 0.24 - 134 Long ³ + Elev + Elev ² + Elev ³	Elev + Elev ² + Elev ³ 813 0.20 - 146 0.05 Lat + Lat ² + Lat ³ + Long + Long ² + 809 0.24 - 134 0.18 Long ³ + Elev + Elev ² + Elev ³	Elev + Elev ² + Elev ³ 813 0.20 - 146 0.05 - Lat + Lat ² + Lat ³ + Long + Long ² + 809 0.24 - 134 0.18 - Long ³ + Elev + Elev ² + Elev ³

Table 3. Regression models of skewness in the US and Finland. Std. = standardized. Abbreviations as in Table 1.

Effect	Std. coefficient	Standard error	<i>t</i> -value	<i>p</i> -value
US, $R^2 = 0.09$, $n =$	669			
Constant	0.00	0.02	43.57	0.000000
T_{SD}	-0.11	0.01	-2.34	0.02
T_{SD}^{2}	-0.21	0.00	-4.92	0.000001
$T_{ m diur}$	-0.13	0.01	-2.93	0.003
${T_{\text{diur}}}^2$	0.13	0.00	3.07	0.002
NO_x	-0.10	0.01	-2.65	0.01
рН	-0.15	0.03	-3.71	0.0002
Finland, $R^2 = 0.20$	0, n = 100			
Constant	0.00	0.05	20.88	0.000000
T_{diur}^{2}	-0.31	0.08	-2.54	0.013
TP^2	0.31	0.04	2.71	0.008
Cond ²	-0.33	0.07	-2.85	0.005

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Figure legend Figure 1. Relationships between richness and climatic (A-E), water chemistry (F-J), and spatial variables (K-M) in the US dataset. The fits were generated by LOWESS with tension of 0.5. All predictors were centered on the mean. n = 669. $T_{mean} = mean$ annual temperature, $T_{SD} = temperature$ seasonality (standard deviation), $T_{\text{diur}} = \text{mean diurnal temperature range}$, $T_{\text{min}} = \text{minimum temperature of the coldest month}$, T_{max} = maximum temperature of the warmest month, TP = total phosphorus. **Figure 2.** Relationships between parameter σ and climatic (A-E), water chemistry (F-J), and spatial variables (K-M) in the US dataset. The fits are generated by LOWESS with tension of 0.5. All predictors were centered on the mean. n = 669. Abbreviations as in fig. 1. Figure 3. Relationships between richness and climatic (A-E), water chemistry (F-H), and spatial variables (I-K) in the Finnish dataset. The fits are generated by LOWESS with tension of 0.5. All predictors were centered on the mean. n = 100. Abbreviations as in fig. 1. **Figure 4.** Relationships between parameter σ and climatic (A-E), water chemistry (F-H), and spatial variables (I-K) in the Finnish dataset. The fits are generated by LOWESS with tension of 0.5. All predictors were centered on the mean. n = 100. Abbreviations as in fig. 1.

Appendix

Table A1. Basic statistics of climatic, water chemistry and spatial variables included in the regression analyses (Tables 1, 2). T_{mean} = mean annual temperature, T_{SD} = temperature seasonality (standard deviation), T_{diur} = mean diurnal temperature range, T_{min} = minimum temperature of the coldest month, and T_{max} = maximum temperature of the warmest month.

Variable	Minimum	Maximum	Median	Mean	SD
US					
Latitude (degrees)	25.40	61.77	41.64	41.58	4.58
Longitude (degrees)	-154.17	-70.74	-92.08	-95.63	16.48
Elevation (m)	57.00	2733.00	303.00	483.85	507.35
Total phosphorus ($mg \cdot L^{-1}$)	0.002	3.40	0.07	0.16	0.31
Nitrite + nitrate ($mg \cdot L^{-1}$)	0.01	13.95	0.46	1.36	2.22
Ammonia ($mg \cdot L^{-1}$)	0.002	5.80	0.03	0.06	0.25
рН	2.95	9.60	7.90	7.83	0.56
Specific conductance ($\mu S \cdot cm^{-1}$)	11.00	23595.65	476.00	539.10	966.64
T _{mean} (°C)	-0.80	23.70	9.60	9.86	3.59
T_{sd}	3.19	12.58	9.05	8.93	1.70
T_{diur} (°C)	7.80	19.60	12.10	12.54	1.86
T_{min} (°C)	-22.20	13.40	-9.40	-8.70	5.70
T _{max} (°C)	15.20	36.30	29.20	29.35	2.87
Finland					
Latitude (degrees)	60.27	70.06	66.26	65.91	2.81
Longitude (degrees)	23.28	30.44	27.02	27.04	2.09
Elevation (m)	18.00	320.00	180.00	177.49	81.91

Total phosphorus (mg·L ⁻¹)	0.00	0.19	0.02	0.03	0.04
рН	4.50	8.20	7.00	6.80	0.81
Specific conductance ($\mu S \cdot cm^{-1}$)	9.40	277.00	44.75	65.37	59.24
T _{mean} (°C)	-2.70	5.00	-0.50	0.28	2.03
T_{sd}	6.93	10.19	9.52	9.29	0.80
T _{diur} (°C)	6.50	9.20	8.30	8.21	0.67
T_{min} (°C)	-21.30	-9.10	-17.15	-16.60	3.29
T _{max} (°C)	14.80	21.00	18.75	18.78	1.66

Table A2. Significant variables (P < 0.05) included in models 1-15 from Tables 1 (US) and 2 (Finland).

Abbreviations as in Table 1.

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Eqn	1	US	Finland			
	Richness	Parameter σ	Richness	Parameter σ		
1	NS	1/kT	NS	1/kT		
2	NS	$1/kT + (1/kT)^2$	$1/kT + (1/kT)^2$	$(1/kT)^2$		
3	NS	$ln(N)^2$	_	_		
4	T_{mean}^{2}	$T_{mean} + T_{mean}^{2}$	T_{mean}^{-2}	T_{mean}		
5	$T_{SD} + T_{SD}^2$	$T_{SD} + T_{SD}^2 + T_{diur} +$ T_{diur}^2	$T_{SD} + T_{diur} + T_{diur}^{2}$	$T_{SD} + T_{diur}$		
6	$T_{min} + T_{min}^2 + T_{max} $ $+ T_{max}^2$	$T_{min} + T_{min}^{2} + T_{max} +$ T_{max}^{2}	$T_{min} + T_{max} + T_{max}^{2}$	$T_{min} + T_{max}^{2}$		
7	$\begin{split} T_{mean} + T_{mean}^2 + T_{SD} \\ + T_{SD}^2 + T_{diur} + \\ T_{diur}^2 + T_{min} + T_{max} \end{split}$	$\begin{split} T_{SD} + T_{SD}^2 + T_{diur} + \\ T_{min} + T_{min}^2 + T_{max} \end{split}$	$T_{SD} + {T_{diur}}^2 + T_{min} +$ $T_{max} + {T_{max}}^2$	$T_{mean}^2 + T_{diur} + T_{min}^2 + $ T_{max}^2		
8	$TP + TP^2 + NO_x$	$TP + TP^2 + NO_x \\$	TP^2	NS		
9	$TP + TP^2 + NO_x \\$	TP + Cond	$TP^2 + pH^2$	$TP^2 + Cond^2$		
10	$\begin{split} T_{mean} + T_{SD} + T_{SD}^2 \\ + T_{diur} + T_{min} + T_{max} \\ + T_{max}^2 + TP + NO_x \\ + NH_4 \end{split}$	$\begin{split} T_{SD} + T_{SD}^2 + T_{diur} + \\ T_{min} + T_{min}^2 + T_{max} + \\ TP \end{split}$	$T_{mean} + T_{diur} + T_{min} + \\$ $T_{max}^2 + TP^2$	$\begin{split} T_{mean} + T_{SD} + T_{diur} + T_{min} \\ + T_{max}^2 + TP^2 \end{split}$		
11	$\begin{split} T_{mean} + T_{SD} + {T_{SD}}^2 \\ + T_{diur} + T_{min} + T_{max} \end{split}$	$\begin{split} T_{SD} + T_{SD}^2 + T_{diur} + \\ T_{min} + T_{min}^2 + T_{max} + \\ TP \end{split}$	$T_{mean} + T_{diur} + T_{min} +$ $T_{max}^2 + TP^2 + pH^2$	$\begin{split} &T_{SD} + {T_{diur}}^2 + {T_{max}}^2 + TP \\ &+ Cond \end{split}$		

$$+ T_{max}^2 + TP + NO_x$$

$$+ NH_4$$

$$12 \quad Lat + Lat^2 + Lat^3 \quad Lat^2 \quad Lat^2 + Lat^3 \quad Lat^2 + Lat^3$$

$$13 \quad Long^2 + Long^3 \quad Long^2 + Long^3 \quad Long + Long^2 \quad Long^2$$

$$14 \quad Elev + Elev^3 \quad Elev^2 \quad Elev + Elev^2 \quad Elev^2$$

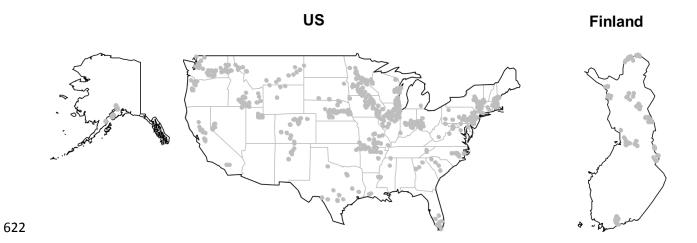
$$Lat + Long^2 + \quad Lat^3 + Long + Long^2$$

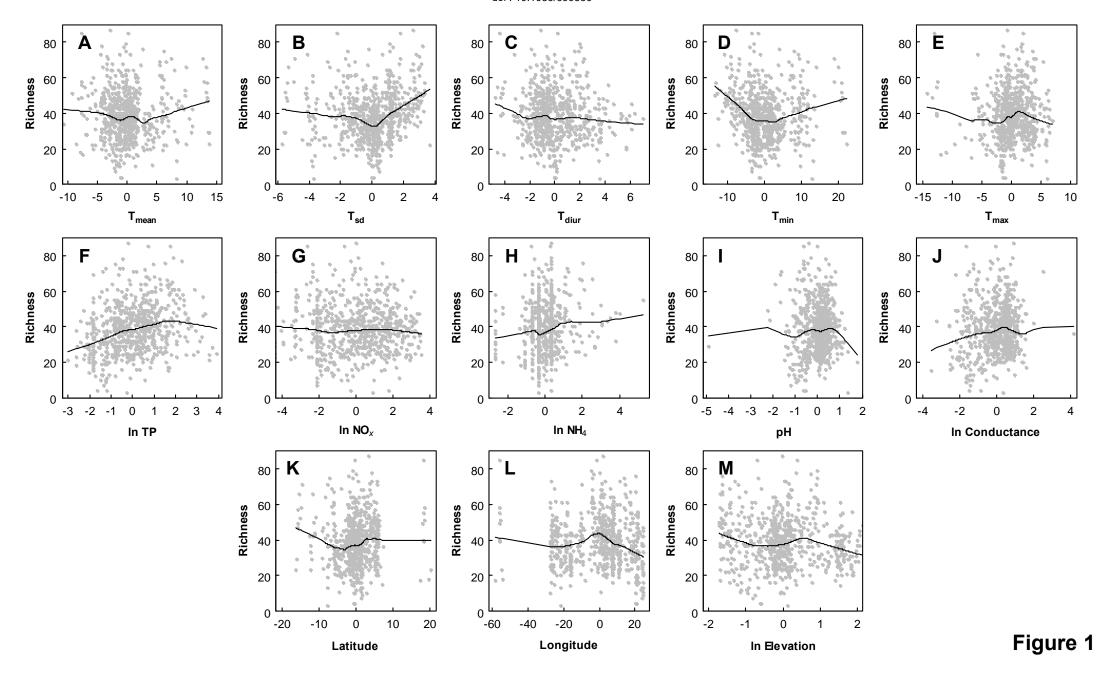
$$15 \quad Long^3 + Elev^2 + \quad + Long^3 + Elev + \quad Lat^2 + Long^2 \quad Lat^2 + Lat^3$$

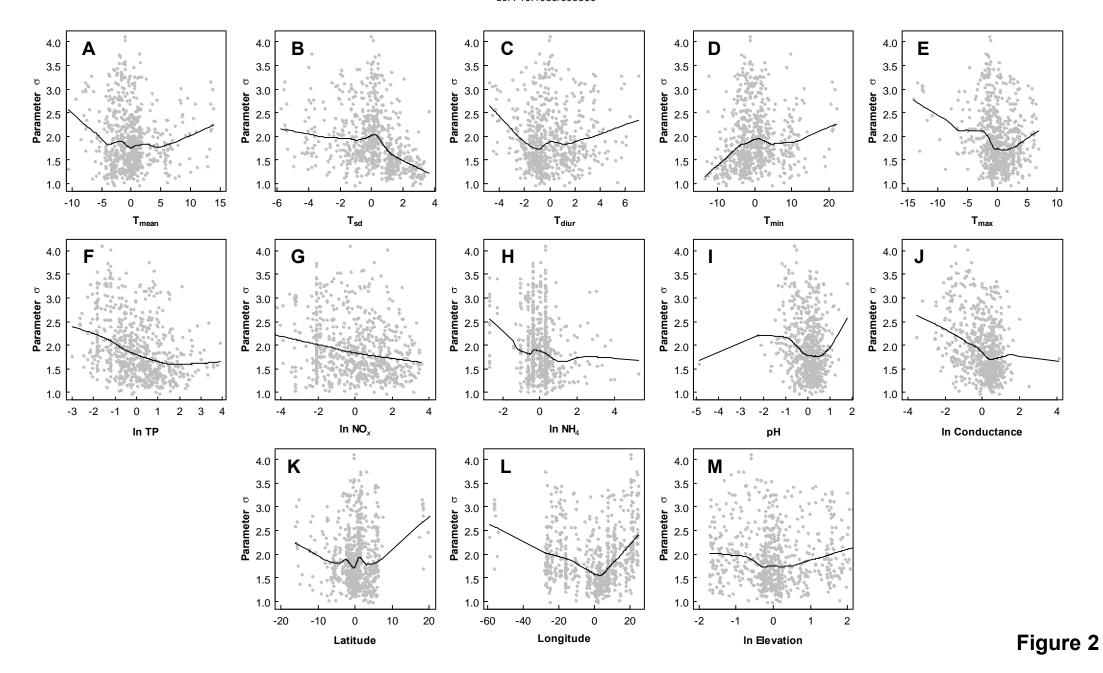
$$Elev^3 \quad Elev^2$$

Figure A1. Map of the study streams in the US and Finland.

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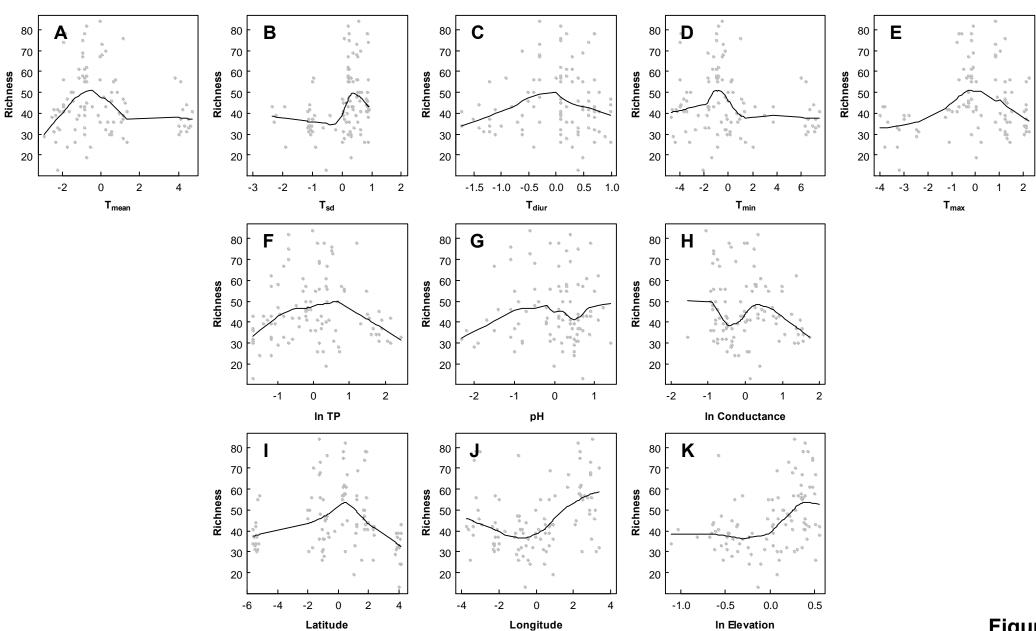


Figure 3

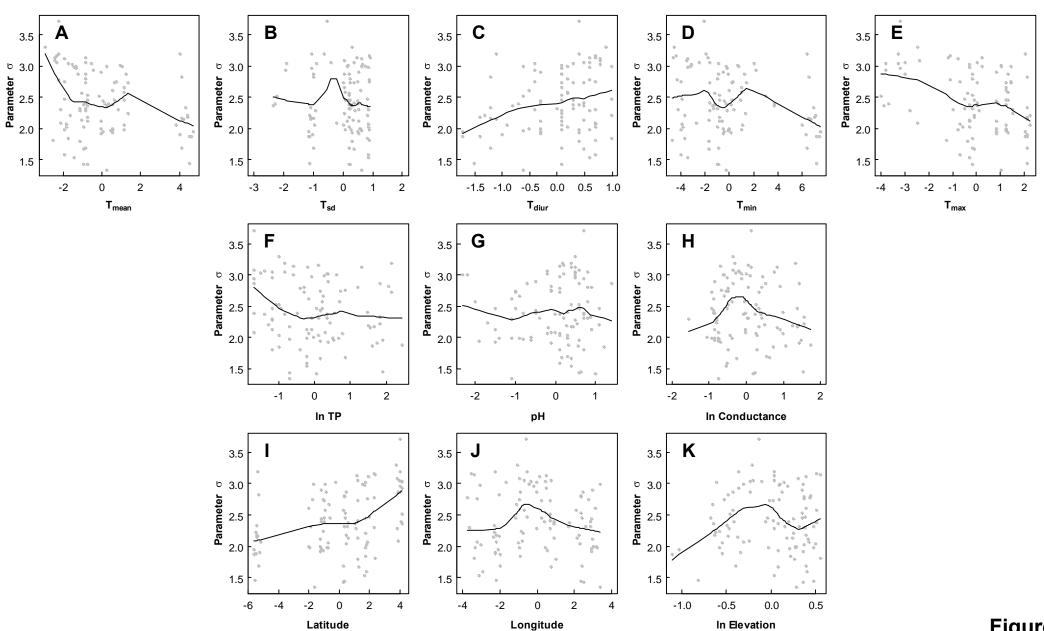


Figure 4