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Aluminum oxide nanowires as safe and effective adjuvants for **next-generation** vaccines

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Improving efficiency of an adjuvant, material that enhances the body's immune response to an antigen, has become vital for the development of safer, cheaper, and more effective **next-generation** vaccines. Commercial vaccines typically use aluminum salt-based adjuvant particles, most commonly aluminum oxyhydroxide (AlOOH) and aluminum hydroxide (Al(OH)₃) based often referred to as "alum". Despite their broad use, their adjuvant properties are rather moderate. This is even worse in the case of aluminum oxide (Al₂O₃)-based adjuvant. While being more robust and less cytotoxic, Al₂O₃ is a significantly less effective adjuvant than above-mentioned Al compounds and is consequently not commonly used. Here, we report on the remarkably enhanced adjuvant properties of Al₂O₃ when produced in the form of nanowires (NWs). Based on recent advances in understanding neutrophil activation by inert nanoscaffolds, we have created ultra-long Al₂O₃ NWs with a high aspect ratio of ~1000. These NWs showed strong humoral immune response with no damaging effect on the microvasculature. Since only the change of shape of Al adjuvants is responsible for the excellent adjuvant properties, our finding holds great promise for rapid implementation as safer and more effective adjuvant alternative for human vaccines. The mechanism behind human blood-derived neutrophil activation with Al₂O₃ NWs was found to be sequestering of Al₂O₃ NWs by neutrophils *via* formation of neutrophil extracellular traps (NETs).

Introduction

Vaccination has made an enormous contribution to global health and is considered one of the greatest medical accomplishments for achieving global safety against dangerous diseases. Since the efficiency of a particular vaccine correlates directly with the percentage of a given population that has been vaccinated, the vaccine market requires cost-effective and efficient solutions to attain broad vaccination coverage [1]. The technologies

involved in manufacturing different types of vaccines influence strongly vaccine costs. The high price of antigen expression makes many vaccines less affordable to (and thus less used by) low-income population [2]. Adding effective adjuvants to vaccine formulation enhances vaccines in two possible ways: (a) it allows the use of smaller antigen doses or physically small adjuvant molecules (meaning synthetic pathogenic components instead of attenuated risky pathogens) and (b) it leads to enhanced antibody titers, decreasing the number or re-vaccinations and maintaining the reliable antibody levels during whole life [3]. Furthermore, improving the efficiency of adju-

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vants to enhance the body's immune response to an antigen would allow the development of next generation of safer, more effective, and more affordable vaccines.

Aluminum (Al) salt-based adjuvants (most commonly in the form of aluminum oxyhydroxide (AlOOH), aluminum hydroxide (Al(OH)₃) are used in human vaccines since 1930s and are commonly referred to as "alum" [4–6]. The presence of oxygen-containing groups on their surface [7–9] provides fast antigen adsorption and yield a vigorous response from the immune system compared to alternatives [10]. While aluminum oxide (Al₂O₃) is more robust, it is significantly less effective than other Al-based adjuvants and is commonly much less used. Furthermore, Al salt-based adjuvants exhibit relatively low cytotoxicity as compared to pathogen stimulants or virusomes and have proven to cause a fewer number of local severities [11,12].

Recent studies showed that the shape and dimensions of the adjuvant particles may affect the immune response against protein and peptide antigens [13,14]. For example, enhanced adjuvant activity of small, elongated particles (nanorods or nanosticks) of AlOOH and Al(OH)₃ with diameters of 4–20 nm and aspect ratio of 8–10 was recently reported [14–16], although the origin of the improved performance remained largely unclear.

Earlier, some of us demonstrated that different size nanoparticles thrust the activity of neutrophils, which boosts the immune system response toward co-injected antigens [17,18]. This mechanism of neutrophils activation is highly dependent on the size of the particles, e.g. bacteria, fungi, nano- and microparticles [18,19]. It results in the formation of neutrophil extracellular traps (NETs), which effectively isolate foreign bodies and suppress local self-limiting inflammation [20]. The mechanism of NETs' action is realized by the entrapping of foreign particles by unbound DNA and histones, both decorated by toxic proteinous content of neutrophils' granules [21]. Formation of NETs was recently demonstrated as an important player responsible for adjuvant action of injected alum, reported to recruit neutrophils, and their activation leading to stimulation of DNA sensors and provoking local inflammatory response [22].

Since the NETs' formation is particles' size-dependent, we hypothesized that the utilization of high-aspect ratio Al-based vaccine adjuvants can greatly contribute to establishing more potent immune responses through neutrophils' activation. Unfortunately, relatively expensive synthesis of nanorods (due to the use of harsh chemicals, elevated reaction temperatures, and low reaction yields), which was demonstrated earlier [14], as well as their potential damages to the smallest blood vessels may become barriers for their use in low-cost vaccination.

In this article, we have chosen to investigate if Al₂O₃ nanowires (NWs) can activate neutrophil cells upon injection to the body and possibly enhance adjuvant properties over other Al-based adjuvants. We choose to compare our findings to aluminum-based adjuvants, commonly used in commercially available vaccine formulations. Our interest was partially triggered by our recent discovery of an innovative approach for ultra-low-cost synthesis of γ -alumina NWs (γ -Al₂O₃ NWs) of variable dimensions and controllable aspect ratio [23,24]. These nanomaterials are produced by exposing bulk Al alloys to alcohols at near room temperature and atmospheric pressure, fol-

lowed by heat treatment in air. No catalysts, toxic metals, external stimuli, or costly synthesis systems are required. We expect the future industrialization will allow formation of these γ -Al₂O₃ NWs at a cost of a few dollars per pound, which is almost negligible for vaccination applications. The tests conducted for the first time on 20- to 60-micron-long, 20–40 nm diameter, ~1000 aspect ratio NWs substantially exceeded our expectations and demonstrated γ -Al₂O₃ NWs to be extremely effective vaccine adjuvant, inducing excellent humoral immune response. Quite remarkably, γ -Al₂O₃ NWs did not damage the microvasculature of the tissues and were found to be easily entrapped by NETs and passivated. When compared with state of the art commercial Al-based adjuvant, the proposed low-cost γ -Al₂O₃ NWs were found to be both more efficient and safer. Our findings highlight great promises of NWs for substantial breakthroughs in the development of safer, more effective, and more affordable vaccines.

Results and discussion

γ -Al₂O₃ NWs have been synthesized by the protocol described by some of us previously [23]. It relies on the de-alloying of bulk Al-Li alloy in ethanol at 60 °C for 24 h, which yields Al ethoxide NWs. Subsequent heat-treatment at 800 °C for 1 h results in the formation of γ -Al₂O₃ NWs, as shown in the scanning electron microscopy (SEM) micrograph (Fig. 1a). The microstructure of the NWs is uniform. The average NWs' thickness is ~20 nm, average length 20–60 μ m, and aspect ratio exceeding ~1000. We believe that the dimensions of the NWs are sufficient to activate neutrophils (typically 12–17 μ m in diameter). In particular, of a typical length of 35 μ m [25], the crystals of monosodium urate, which are formed in the condition of the gout, can be effectively ingested by the neutrophils [26,27]. The crystallinity of the γ -Al₂O₃ NWs was revealed by the transmission electron microscopy micrograph (TEM, Fig. 1b). The γ -Al₂O₃ NWs are polycrystalline with an average crystal diameter of 15–20 nm. The rough surface and sharp ends of the γ -Al₂O₃ NWs can promote the membrane damage in neutrophils, resulting in the promotion of the NET formation [18]. The characteristic selected area electron diffraction (SAED) pattern in Fig. 1b area exhibits strong and prominent diffraction rings indexed as 311, 222, 400, and 440 reflections of γ -Al₂O₃ (Fig. 1c). Thermogravimetric analysis (TGA) shows clearly the conversion of Al(OEt)₃ NWs to Al₂O₃ NWs (Fig. 1c). Organic carbon- and hydrogen-containing groups, which constitute almost ~40 wt.% of the mass, burn in air until 400 °C. In the range of 500–1000 °C, the mass of the Al₂O₃ NWs is stabilized. The high-angle XRD pattern (Fig. 1d) of the Al₂O₃ NWs displays diffraction peaks at 2θ of 14.66°, 37.18°, 39.03°, 45.36°, 60.15°, and 66.70°, indexed as (2 2 0), (3 1 1), (2 2 2), (4 0 0), (5 1 1), and (4 4 0) crystal planes of γ -Al₂O₃ NWs, respectively. The chemical composition of γ -Al₂O₃ NWs was further validated by FTIR (Fig. 1e) and XPS analysis (Fig. 1f). The FTIR spectrum exhibits a broad band centered at 3495 cm⁻¹ due to OH (stretching) band and/or adsorbed water. There is a general lack of peaks related to oxygen-containing groups besides the presence of bands related to AlOOH, such as the bands at 3298, 3091, 1070, and 1165, assigned to $\nu_{as}(\text{Al})\text{-OH}$, $\nu_s(\text{Al})\text{-OH}$, stretching vibrations, and δ_{as} Al-O-H modes for

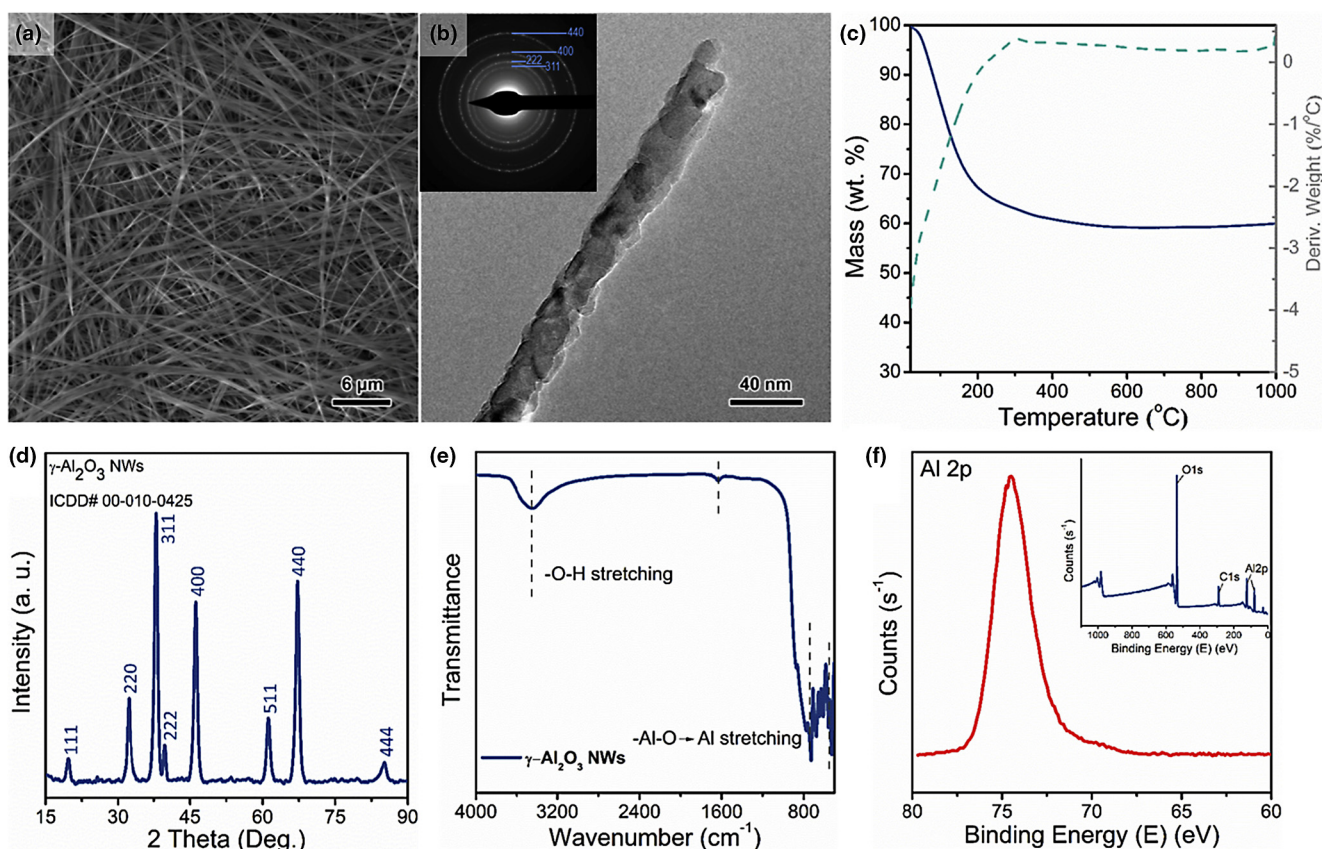


FIG. 1

Characterization of γ - Al_2O_3 nanowires: (a) SEM micrograph; (b) TEM micrograph and SAED; (c) TGA; (d) XRD pattern; (e) FTIR; (f) XPS survey (inset) and characteristic of the Al_{2p} core level.

AlOOH. The bands below 1000 cm^{-1} at 864 , 749 , and 583 cm^{-1} are caused by the formation of the γ - Al_2O_3 phase. Furthermore, the XPS survey spectrum, together with the high-resolution core-level spectrum of Al_{2p} (74.6 eV), is shown in Fig. 1f,g (inset). The atomic ratio between $\text{Al}_{2p}/\text{O}_{1s}$ is 0.65 close to that of the theoretical value of 0.66 for γ - Al_2O_3 .

A commercial Al-based adjuvant (alum) (Fig. S1a) and commercial aluminum oxide microparticles (Al_2O_3 MPs) (Fig. S1b) were used as control materials throughout this study. Fig. S1c shows X-ray diffraction (XRD) patterns of alum. The XRD patterns indicate that alum has an aluminum oxide hydroxide (γ - $\text{AlO}(\text{OH})$) (boehmite) structure with the average crystallite size 2.2 nm (Fig. S1). Zeta-potential measurements in water at $25\text{ }^\circ\text{C}$ (Fig. S1d) indicate that alum adjuvant formulation has a positive surface charge of $+6.6 \pm 0.3\text{ mV}$ (Table S1). This is only slightly higher than that of the γ - Al_2O_3 NWs with a surface charge of $+2.2 \pm 1.5\text{ mV}$. Surprisingly, using Al_2O_3 MPs, a negative surface charge was recorded. The positive surface charge of γ - Al_2O_3 NWs may be advantageous to facilitate the electrostatic binding of negatively charged proteins. Indeed, incubation of γ - Al_2O_3 NWs in water supplemented with 0.2% ovalbumin (OVA) results in a material with negative (about -25.6 mV) surface charge due to the adsorption of OVA. The same was true for the alum adjuvant.

The cell toxicity of γ - Al_2O_3 NWs was tested on HeLa and U87MG cell lines after 24-h incubation (Fig. S2a–b). In addition,

and in the belief that γ - Al_2O_3 NWs will be mainly processed by macrophages, cytotoxicity studies of this cell type were included (Fig. S2c).

The cytotoxicity of the different materials was evaluated based on cell viability assessment by MTS assay relative to controls as proposed by Kong et al. [28]: non-toxic $>90\%$ cell viability, slightly toxic 65 – 90% , toxic 35 – 65% , severe toxic $<35\%$. All the nanostructures appeared nontoxic for HeLa cells after 24 h of exposure even at high concentrations. This was also true for Al_2O_3 MPs alone. Using U87MG human brain cells, in the case of γ - Al_2O_3 NWs at concentrations $>100\text{ }\mu\text{g mL}^{-1}$, the nanostructures proved to be slightly toxic after 24 h of exposure. The same was observed for alum. Interestingly, Al_2O_3 MPs had a different cytotoxic behavior on these cell lines: at concentrations between 200 and $400\text{ }\mu\text{g mL}^{-1}$, the structures were slightly toxic and reached toxic levels for concentrations $\geq 500\text{ }\mu\text{g mL}^{-1}$.

In order to test adjuvant properties of γ - Al_2O_3 NWs, we have determined both parts of immune responses, namely humoral and cellular immune responses. Immunization experiments were performed on Balb/c laboratory mice using chicken egg OVA as model antigen. The presence of anti-ovalbumin antibodies, both IgG and IgM types, was measured before immunization (day 0), as well as after 14 and 35 days of immunization (Fig. 2a,b, Fig. S3, Fig S4a–b). The humoral immune response after a second vaccination caused by the “ γ - Al_2O_3 NWs + OVA” was significantly stronger compared to that effect of “alum + OVA”

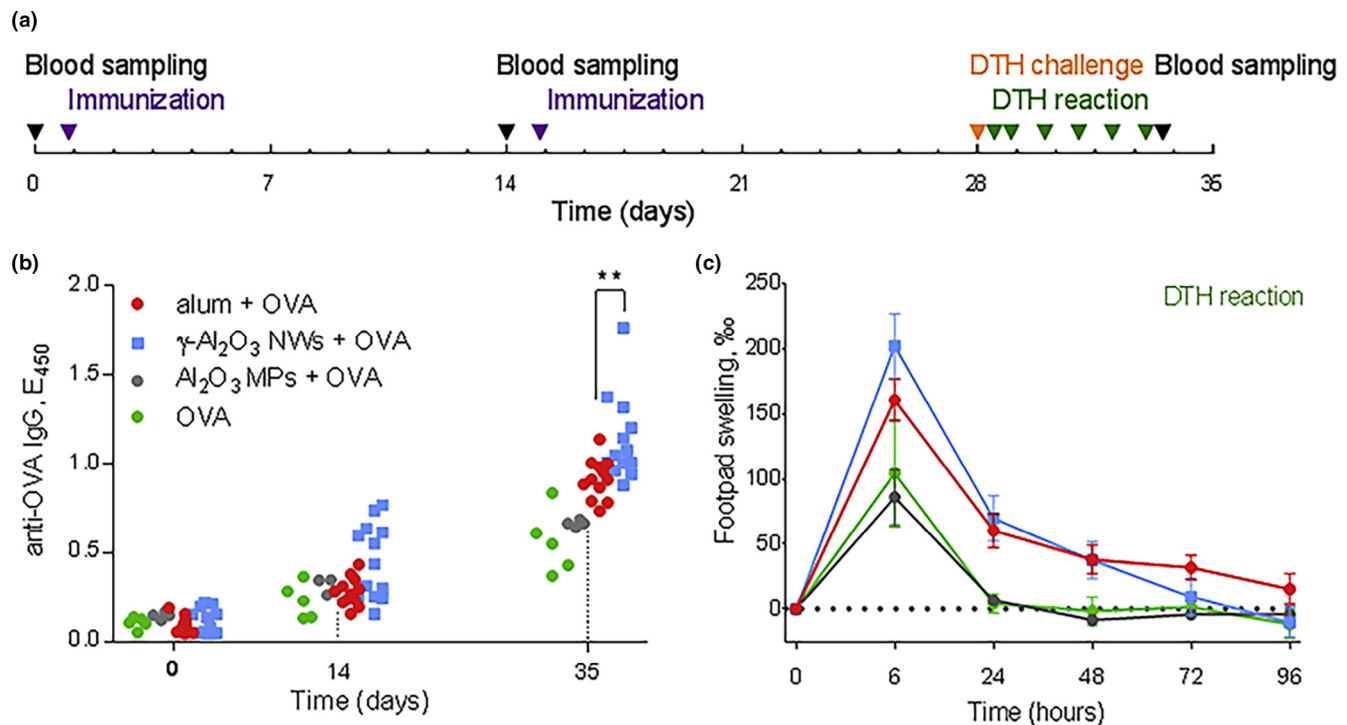


FIG. 2

Humoral and cellular immune responses in Balb/c mice provoked by OVA using alum, γ -Al₂O₃ NWs, Al₂O₃ MPs, or OVA alone acting as adjuvants: (a) experimental design and (b) humoral immune response. Anti-OVA IgG-specific levels in serum (humoral immune response), each dot represent the response of specific animal; sera dilution was 1:2000; (c) cellular immune response; DTH reaction measured as a foot swelling. Data are M ± sem for all studied animals in the group; n = 12, 13, 4, 5, correspondingly.

(Fig. 2b), as indicated by the significantly higher amount of anti-OVA IgG antibodies formed 35 days after immunization at the dilution factor of 1/2000. Further analysis of antibody titers (dilution factor at which antibody is still effective) revealed (Fig. S3) that titers of antibodies obtained after γ -Al₂O₃ NWs + OVA immunization were 1/64000; after the use of alum – 1/16000, for Al₂O₃ MPs and OVA alone, they were as low as 1/2000 and 1/1000, correspondingly; the cut-off was set to 0.5 OD value (Fig. S3). Both adjuvants were not inducing acute-phase antibodies of IgM type (Fig S4b), which signifies that there is no inflammation left.

The cellular immune response was assessed using an established model of delayed type hypersensitivity (DTH) reaction [29], modified by us to be less invasive animal treatment protocol [18]. The DTH is a part of the cell-mediated immune response associated with Th1 reactivity in response to insoluble foreign antigens. It is a localized inflammatory reaction prototypical *in vivo* assay to study cell-mediated host immune response mediated by CD4+ T lymphocytes (small leukocytes, white blood cells). These cells promote T-helper cell type 1 (Th1) production of interferon- γ (a soluble cytokine). This immune response is characterized by an influx of innate immune cells, with macrophages and the dendritic cells predominating. Therefore, after the formation of antibodies (day 28), mice were challenged with OVA injected between the fingers of the footpad (right foot), while PBS served as control and injected to the left foot (Fig. 2a). Footpad swelling, being proportional to the strengths of cellular immune response, was measured in dynamics for

few days, until swelling disappeared. The DTH reaction (usually evaluated at 6–24 h after antigen exposure) was comparable for both groups of animals, immunized with OVA in the presence of either alum or γ -Al₂O₃ NWs (Fig. 2c), while it was significantly lower for Al₂O₃ MPs or OVA alone. The swelling of the footpad faded after 72 h of the DTH challenge, which correlates with the normal responsiveness to the antigen. Absence of exaggerated cellular response with much stronger humoral response for γ -Al₂O₃ NWs makes them immunologically favorable agents for the use in vaccines.

Some of us have recently shown that nanoscaffolds may be able to induce activation of polymorphonuclear neutrophils to form neutrophil extracellular traps [18], networks of extracellular fibers produced as an immune response to neutralize infection [30]. Localization of neutrophils leads to localized self-limiting inflammation, which results in isolation of inflammatory stimuli with NETs [31] and, as discovered recently, is an important trigger of alum action [22]. This phenomenon is mainly responsible for the strong self-limiting (fast resolved) inflammation caused by some nanoparticles [18] (best illustrated by gout attack, caused by monosodium urate needle-like crystals, to be as spontaneously resolved as it appears).

Since γ -Al₂O₃ NWs provoked increased humoral immune response, we were interested to follow up the fate of the γ -Al₂O₃ NWs upon injection in the body. We used a lately described mouse air pouch model [32], where sterile air is injected under the skin at the back of the mouse to form a cavity, which is being infiltrated with immune cells followed by γ -Al₂O₃

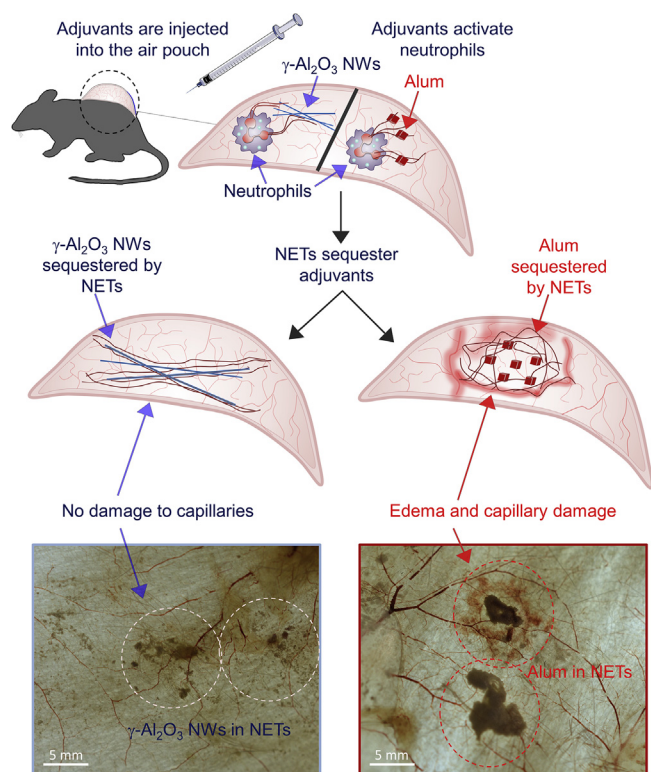


FIG. 3

Formation of neutrophil extracellular traps (NETs): sequestering of $\gamma\text{-Al}_2\text{O}_3$ NWs and alum by aggregated NETs after 24 h of incubation in mouse air pouch. Dotted red circles show capillary damage and edema. No significant damage to capillaries is observed after $\gamma\text{-Al}_2\text{O}_3$ NWs' treatment.

NWs' injection (1 mg $\gamma\text{-Al}_2\text{O}_3$ NWs per mouse). Within 24 h after NWs' injection, mice were sacrificed, and their skin was observed for aggregated NETs and visual damage by microphotography. For both $\gamma\text{-Al}_2\text{O}_3$ NWs and alum, we have observed abundant formation of aggregated NETs entrapping injected Al compounds (Fig. 3., S5). However, micro-capillary damage and edema were significantly less visible for $\gamma\text{-Al}_2\text{O}_3$ NWs than for commercial alum. It is known that swelling and edema often arise at the place of vaccine injection, which is accompanied by the release of histamines [33]. However, the exact molecular mechanism of how injection of alum results in edema remains unknown [22,34]. Therefore, the remarkable properties of $\gamma\text{-Al}_2\text{O}_3$ NWs of not causing edema are highly beneficial for potential applications as adjuvants. Their utilization will result in less swelling, edema, and irritation at the place of injection.

To elucidate the mechanism of the $\gamma\text{-Al}_2\text{O}_3$ NWs' adjuvant properties and related neutrophil activation, we tested their ability to stimulate NETs' formation in human blood-derived neutrophils. For this reason, we obtained a venous blood from 3 healthy volunteers and isolated neutrophil population in sterile conditions. Then neutrophils were placed in multi-well plate containing Sytox Green, a fluorescent dye, which senses extracellular DNA (those externalized from neutrophils in the case of NETs' formation). Positive strong (phorbol myristyl acetate, PMA) and weak (48 mM bicarbonate) inducers of NETs' formation were applied to cell well, simultaneously with $\gamma\text{-Al}_2\text{O}_3$ NWs, alum, or Al_2O_3 MPs. Fig. 4 shows the data of neutrophil

activation, isolated from 3 different healthy donors. From the mean fluorescence intensity (MFI) level, it is evident that $\gamma\text{-Al}_2\text{O}_3$ NWs exhibit immediate to strong NET formation upon the contact with the isolated neutrophils. The same was true for alum, the structure of which can also be regarded as 1D material (please see elongated shapes of the fibers in Fig. S1A). In contrast, the Al_2O_3 MPs did not induce such an effect. This is an important finding, which confirms that the improved humoral response of $\gamma\text{-Al}_2\text{O}_3$ NWs is most likely to be linked to their outstanding structural characteristics. Based on our previous findings, we suspect that a combination of high aspect ratio and small diameter of $\gamma\text{-Al}_2\text{O}_3$ NWs effectively activates neutrophils by causing local disturbances in lipid bilayer composition followed by membrane re-cycling. The same level of neutrophils' activation, although seen for alum, does result in the membrane damaging and lower levels of humoral immune response as compared to the $\gamma\text{-Al}_2\text{O}_3$ NWs. It also corroborates our hypothesis that not only the chemical composition, but also the shape and size of the adjuvant may have a profound effect on activation and interaction with NETs.

Based on obtained data, we propose the mechanism of enhanced action of $\gamma\text{-Al}_2\text{O}_3$ NWs as an adjuvant. $\gamma\text{-Al}_2\text{O}_3$ NWs adsorb antigens and upon injection release them; this results in local macrophage and neutrophil activation (Fig. 5a). Macrophages process the antigens and stimulate antibody production through plasma cells (Fig. 5b). Additionally, at the early time upon neutrophil contact with $\gamma\text{-Al}_2\text{O}_3$ NWs, the former release NETs and cytokines, and significantly enhance antibody production (Fig. 5b). Within 24 h, $\gamma\text{-Al}_2\text{O}_3$ NWs are sequestered by NETs, which limits the inflammation caused by the presence of the foreign $\gamma\text{-Al}_2\text{O}_3$ NWs' particles (Fig. 5c). The $\gamma\text{-Al}_2\text{O}_3$ NWs reside locally within the traps without damaging the microvasculature or causing local inflammation. Sufficient concentration of antibodies is achieved in the body as the result of the " $\gamma\text{-Al}_2\text{O}_3$ NWs' adjuvant + antigen" vaccination.

Conclusions

This study demonstrates for the first time the extremely promising performance of long (20–60 microns) small diameter (20–40 nm), high aspect ratio (>1000) $\gamma\text{-Al}_2\text{O}_3$ nanowires (NWs) as a material that enhances the body's immune response to an antigen. The results are particularly encouraging because Al_2O_3 is commonly considered to be rather ineffective as an adjuvant. Compared to conventional Al_2O_3 microparticles, $\gamma\text{-Al}_2\text{O}_3$ NWs exhibited noticeably reduced cell toxicity and dramatically stronger neutrophil activation. Furthermore, $\gamma\text{-Al}_2\text{O}_3$ NWs showed improved humoral immune responses, resulting in 4-fold (1/64000 vs 1/16000) higher antibody titers and significantly less damages to microvasculature when compared to alum, a commonly chosen vaccine adjuvant. Our study further revealed that $\gamma\text{-Al}_2\text{O}_3$ NWs can effectively activate polymorphonuclear neutrophils to form neutrophil extracellular traps (NETs), thereby explaining their outstanding properties. The following neutralization of the NWs in the aggregated NETs explains the self-limiting nature of the NWs' inflammatory effect, which makes NWs safer candidates for common vaccines. We expect the ultra-low cost of the NWs' synthesis in combination with their

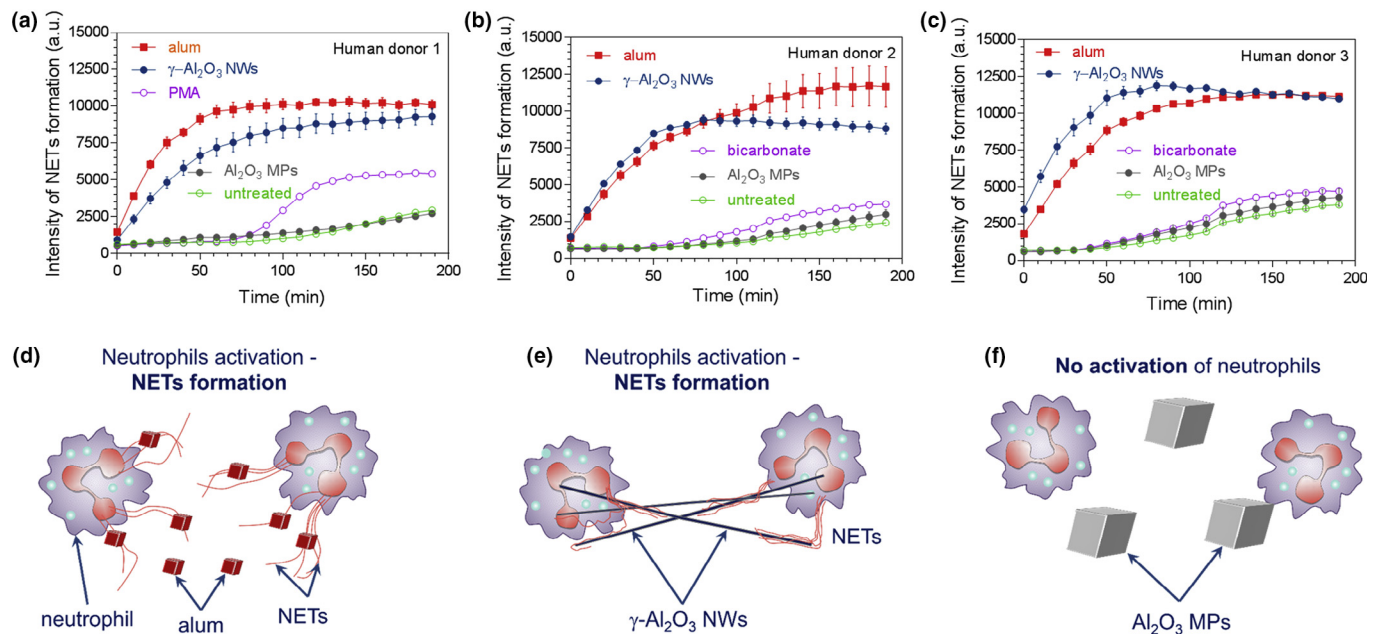


FIG. 4

Activation of peripheral blood-derived neutrophils upon the contact of Al-based adjuvants. Data of three different healthy donors (a–c) are presented. Intensity of NETs' formation is presented as a mean fluorescent intensity of Sytox green dye upon the contact with extracellular DNA: γ - Al_2O_3 NWs and alum but not Al_2O_3 MPs cause rapid activation of human peripheral blood neutrophils (d–f), which is noticeable during the first minutes of incubation. Strong (PMA) and weak (48 mM bicarbonate) inducers of NETs' formation show their response after 120 min.

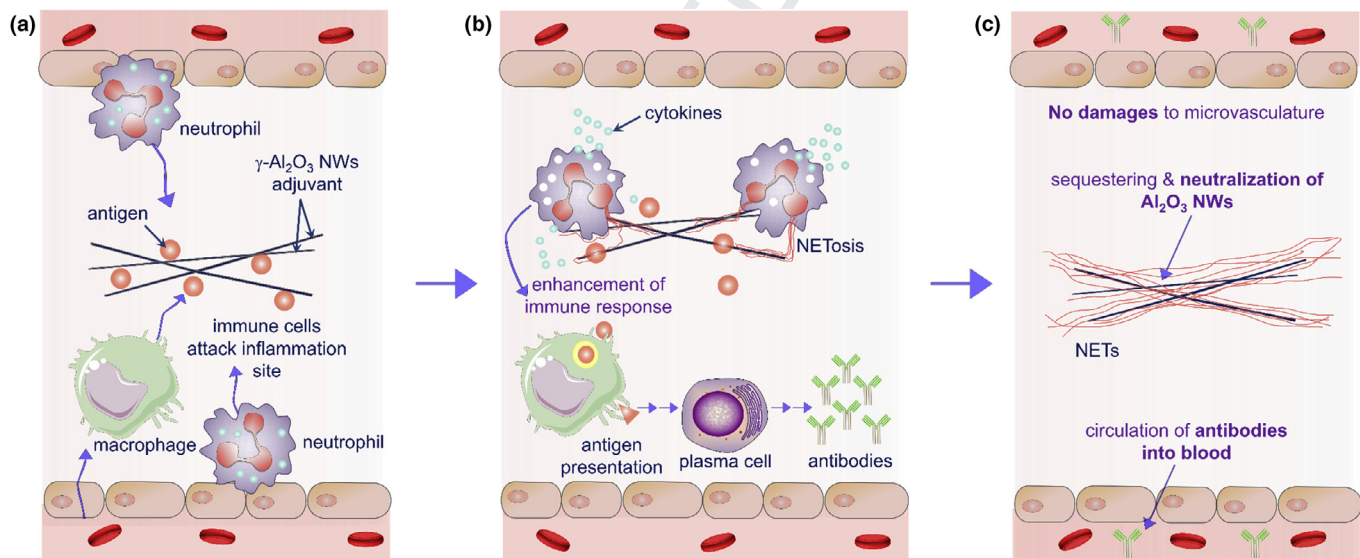


FIG. 5

Brief schematics of the process of the immunization by the γ - Al_2O_3 NWs-OVA (see text for description).

extremely promising performance may open the door to new families of next-generation adjuvants for better, safer, and cheaper human vaccines. We also expect that modifying NWs' surface chemistry and optimizing their dimensions and morphology may boost their performance further.

Materials and methods

Materials

Aluminum oxide (Al_2O_3), Aluminum hydroxide ($\text{Al}(\text{OH})_3$), aluminum (99.999% pure), lithium (99.99% pure), anhydrous etha-

nol (EtOH), and ovalbumin (OVA) were purchased from Sigma-Aldrich and used as received.

Fabrication of γ - Al_2O_3 NWs

According to the previous study of transformation from bulk alloys to oxide nanowires [23], a bimetallic Al-Li alloy (50 at.% lithium) was exposed to anhydrous ethanol ($\geq 99.5\%$) to yield soluble Li alkoxide and white precipitate Al alkoxide nanowires ($\text{Al}(\text{EtO})_3$ NWs). After initial delithiation, Li alkoxides were decanted and then $\text{Al}(\text{EtO})_3$ NWs were washed by extra dry alcohol. The produced $\text{Al}(\text{EtO})_3$ NWs were cast on polyethylene, and

376 ethanol was evaporated at room temperature under argon protec-
377 tion. The final step of the Al(EtO)₃ NWs' transformation into
378 Al₂O₃ NWs involved calcination at 1000 °C in air at atmospheric
379 pressure.

380 *Materials characterization*

381 Scanning electron microscopy (SEM) images were obtained using
382 an electron microscope ULTRA 55 (Zeiss, France) equipped with a
383 thermal field emission emitter and three different detectors (EsB
384 detector with filter grid, high efficiency In-lens SE detector, and
385 Everhart-Thornley Secondary Electron Detector). Transmission
386 electron microscopy (TEM) images were recorded on a JEOL
387 TEM 4000EX (JEOL, Japan) operating at 300 kV. Thermogravi-
388 metric analysis (TGA) was conducted on TGA Q50 analyzer (TA
389 instruments, USA) under air atmosphere at a heating rate of 5 °-
390 C·min⁻¹. The powder X-ray diffraction (X'Pert PRO Alpha-1,
391 USA) was performed using Cu-K α radiation to identify the crys-
392 talline phase of the composite. X-ray photoelectron spectroscopy
393 (XPS) was performed using a PHI 5000 VersaProbe-Scanning
394 ESCA Microprobe (ULVAC-PHI, Japan/ USA) instrument at a base
395 pressure below 5 × 10⁻⁹ mbar. Core-level spectra were acquired
396 at pass energy of 23.5 eV with a 0.1 eV energy step. All spectra
397 were recorded with 90° between X-ray source and analyzer and
398 with the use of low-energy electrons and low-energy argon ions
399 for charge neutralization. After subtraction of the linear back-
400 ground, the core-level spectra were decomposed into their com-
401 ponents with the mixed Gaussian-Lorentzian (30:70) shape
402 lines using the CasaXPS software. Quantification calculations
403 were conducted using sensitivity factors supplied by PHI. Fourier
404 transform infrared (FTIR) spectra were recorded using a ThermoS-
405 cientific FTIR instrument (Nicolet 8700) at a resolution of
406 4 cm⁻¹. Dried sample (1 mg) was mixed with KBr powder
407 (100 mg) in an agate mortar. The mixture was pressed into a pel-
408 let under 10-ton load for 2–4 min, and the spectrum was recorded
409 immediately. Sixteen accumulative scans were collected. The sig-
410 nal from a pure KBr pellet was subtracted as the background.
411 Zeta-potential measurements were performed using a Zeta-sizer
412 Nano-ZS (Malvern Instruments Inc. Worcestershire, UK) in clear
413 disposable zeta cell. Samples were diluted to 10 µg mL⁻¹ for
414 Al₂O₃ MPs, γ -Al₂O₃ NWs, and 0.2% for alum in Milli-Q water
415 and were run at 25 °C in quintuplicate with automatic parameter
416 selection (>12 runs).

417 *Cell viability and internalization*

418 The HeLa cell line derived from cervical carcinoma [ATCC[®] CCL-
419 2[™], ECACC, Sigma-Aldrich, Saint-Quentin Fallavier, France] and
420 the U-87 MG cell line derived from human brain (glioblastoma
421 astrocytoma) [ATCC[®] HTB-14[™], Sigma-Aldrich, Saint-Quentin
422 Fallavier, France] were cultured and maintained in Dulbecco's
423 Modified Eagle's medium (DMEM, Gibco[®]) supplemented with
424 10% fetal bovine serum (FBS, Gibco[®]) and 1% penicillin-strepto-
425 mycin (Gibco[®]) in a humidified incubator at 37 °C and 5% CO₂.

426 Cells were seeded at a density of 10⁴ cells/well in a 96-well
427 plate and grown for 24 h before assay. The culture medium was
428 replaced with a fresh medium that contains γ -Al₂O₃ NWs and
429 alum hydroxide 2% at different concentrations. After 24 h, the
430 old medium was aspirated, washed with PBS and replaced with

a fresh medium. The cell viability was evaluated using Uptiblu
(Interchim) method. Briefly, 10 µL of the Uptiblu solution was
added to each well containing 100 µL of DMEM with 10% FBS
and the plate was incubated for 4 h in the humidified incubator.
The fluorescence of each well (excitation 540 nm/emission
590 nm) was measured using a microplate reader (PHERAstar
FS, BMG LABTECH GmbH, Germany). Each condition was repli-
cated five times and the mean absorbance value of non-exposed
cells was taken as 100% cellular viability.

Cytotoxicity studies on macrophages

The human monocytic THP-1 cells were cultured and main-
tained in culture in Roswell Park Memorial Institute medium
(RPMI 1640, Invitrogen) supplemented with 10% fetal bovine
serum (FBS, Gibco[®]), 1% penicillin-streptomycin (Gibco[®]) and
50 µM β -mercaptoethanol in a humidified incubator at 37 °C
and 5% CO₂. THP-1 monocytes were differentiated into macro-
phages by a 5-day incubation with 20 nM phorbol 12-myristate
13-acetate (PMA, Sigma) and seeded at a density of 1.5 × 10⁴-
cells/well in a 96-well plate. The culture medium was replaced
with a fresh medium without β -mercaptoethanol that contains
the compounds from 0 to 400 µg mL⁻¹. After 24 h, the cell viabil-
ity was evaluated using Calcein-AM and PI co-staining. The cells
were stained without any washing by addition of 10 µL of
Calcein-AM/PI mix solution (45 µM/30 µM) in each well and
incubated at 37 °C for 30 min. Subsequently, the stained cells
were observed with 4× objective (GFP and Texas Red filter cubes)
using a Cytation[™] 5 Cell Imaging Multi-Mode Reader (BioTek
Instruments SAS, France).

Animals

Six-week-old male Balb/c mice were used. The mice were housed
in cages covered with air filters in a temperature-controlled room
with a 12-h light and 12-h dark schedule and kept on a standard
diet with drinking water available ad libitum. Studies involving
animals, including housing and care, method of euthanasia,
and experimental protocols, were approved by the Ethics com-
mittee of Danylo Halytsky Lviv National Medical University, pro-
tocol 20180226/2 & 20130624/6; all experiments were designed
to comply with principles of the 3Rs (Replacement, Reduction
and Refinement). Totally, 34 mice were analyzed: 12 mice immu-
nized with alum + OVA, 13 mice immunized with γ -Al₂O₃ NWs
+ OVA, 4 mice immunized in Al₂O₃ MPs + OVA, 5 mice immu-
nized with OVA alone; immunization was done in 3 indepen-
dent experiments.

Mice immunization model

For correct comparison, Al₂O₃, Al₂O₃ NW, and Al(OH)₃ were
used as pure compounds and prepared as dispersions just before
immunization after pre-heating to 150 °C for 2 h to eliminate
LPS. The dispersions of 'alum + OVA', 'Al₂O₃ MPs + OVA', and
' γ -Al₂O₃ NWs + OVA' were prepared by mixing OVA and an adju-
vant (Al₂O₃, γ -Al₂O₃ NWs or alum) in phosphate-buffered saline
(PBS) by means of the sonication at 44 kHz for a duration of
10 min at 25 °C until a homogenous clear dispersion is achieved.
Suspensions were prepared immediately before the injection.
The 50 µL of 'adjuvant (20 mg mL⁻¹) + 50 µL OVA (1 mg mL⁻¹)'

in PBS were injected intraperitoneally (PI) in mice at day 1 and day 15 of the experiment. The DTH test was performed 28 days after the 1st immunization by injection of 50 μL of OVA in PBS (100 $\mu\text{g mL}^{-1}$) into the right hind paw. Injection of 50 μL of PBS into the left hind paw was used as a control. The thickness of the paw was measured with a caliper (B110T; Kroeplin Laengenmesstechnik) before the injection and 6, 24, 48, 72, and 96 h after it, as described previously [18]. Blood was collected from mouse tail before experiment started (day 0), before second immunization (day 14), and at the end of experiment (day 35). No more than 50 μL of blood was collected to prevent damage to the animal. Serum was isolated (usually 20 μL per mice) and immediately frozen.

Air pouch model

Air pouch model [32] was performed by injection of 5 mL of sterile air subcutaneously into the back of Balb/c mice to form an air pouch. In two days, an additional 2 mL of sterile air were blown into the pouch. Two days later, 4 mg of $\gamma\text{-Al}_2\text{O}_3$ NWs, 4 mg of alum in PBS, 4 mg of Al_2O_3 MPs, or PBS only was injected into the air pouches of each mice. Next day, mice were sacrificed, and the air pouch membranes were analyzed.

ELISA procedure

For determination of serum anti-OVA IgG and IgM, immunosorbent NUNC MaxiSorp plates were coated with 50 μL of a 5 $\mu\text{g/mL}$ solution of chicken egg albumin in 100 mM carbonate-bicarbonate buffer (pH 9.6). Plates were incubated overnight at 4 $^\circ\text{C}$ and washed three times with phosphate-buffered saline supplemented with 0.05% Tween (PBS-T). The plates were blocked with 100 μL of 4 % BSA in PBS-T for 2 h at 37 $^\circ\text{C}$. To follow the dynamics of Ab production all serum samples were diluted 1:2000 in washing buffer. Detection was done with goat anti-mouse IgG(H+L)-HRP (Jackson ImmunoResearch, 115-035-003) for IgG or goat anti-mouse IgM-HRP (Jackson ImmunoResearch, 115-035-020) for IgM, diluted in washing buffer (1:10000). For determination of IgG antibody titers, samples collected at day 35, after second immunization, were diluted 1:1000, 1:2000, 1:4000, 1:8000, 1:16000, 1:32000, 1:64000, and 1:128000. 50 μL of prepared solution was added to the well and incubated at 37 $^\circ\text{C}$ for 1 h, after that the plates were washed again. Detection was done using goat anti-mouse IgG(H+L)-HRP (Jackson ImmunoResearch, 115-035-003), which were diluted in washing buffer (1:10000), added to the plates (50 μL per well) and incubated at room temperature for 1 h. After the corresponding washings, the assay was developed with TMB (3,3',5,5'-tetramethylbenzidine) containing H_2O_2 as a substrate (50 μL per well). The reaction was stopped with 50 μL of 1 N sulfuric acid and absorbance was read at 450 and 620 nm reference wavelength at Perkin–Elmer HST 7000 BioAssay reader. To account for differences between plates, values were corrected using a standard serum applied on each plate.

Analysis of DNA externalization

Human granulocytes were isolated from heparinized venous blood of normal healthy donors by Lymphoflot (Bio-Rad) density gradient centrifugation, as described [35]. The granulocyte-rich layer on top of RBCs was taken and subjected to hypotonic lysis

of RBCs. Isolated granulocytes were cultured in 96-well culture plates at 5×10^6 cells per milliliter with special inducers of NETosis and with the fluorescent nucleic acid dye Sytox green (Life Technologies). The fluorescence was measured in kinetics for 3 h using BioAssay Reader HTS7000 (Perkin Elmer), excitation 480/15 nm, emission 535/20 nm.

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Author contributions

RoBi and SS conceived the idea; RoBi, SP, and TD performed immunization, animal experiments, neutrophil isolation, and ex vivo experiments; RaBo, AB, FW, and SZ performed physical characterization; AB performed cell culture testing; KT and FW synthesized NWs; RB, SS, GU, and KT wrote the manuscript.

Data availability

The raw/processed data required to reproduce these findings cannot be shared at this time due to technical or time limitations.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.mattod.2018.10.034>.

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