



Wolbachia Divergence and the Evolution of Cytoplasmic Incompatibility in *Culex pipiens*

Célestine Atyame Nten, Pierrick Labbe, Emilie E. Dumas, Pascal Milesi,
Sylvain Charlat, Philippe Fort, Mylène Weill

► To cite this version:

Célestine Atyame Nten, Pierrick Labbe, Emilie E. Dumas, Pascal Milesi, Sylvain Charlat, et al..
Wolbachia Divergence and the Evolution of Cytoplasmic Incompatibility in *Culex pipiens*. PLoS
ONE, 2014, 9 (1), pp.e87336. 10.1371/journal.pone.0087336 . hal-01938100

HAL Id: hal-01938100

<https://hal.science/hal-01938100>

Submitted on 28 Nov 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

Wolbachia Divergence and the Evolution of Cytoplasmic Incompatibility in *Culex pipiens*

Célestine M. Atyame^{1*}, Pierrick Labbé¹, Emilie Dumas¹, Pascal Milesi¹, Sylvain Charlat², Philippe Fort³, Mylène Weill¹

1 CNRS, University Montpellier 2, ISEM - UMR 5554, Montpellier, France, **2** CNRS, University Lyon 1, LBBE - UMR 5558, Villeurbanne, France, **3** CNRS, University Montpellier 2, CRBM - UMR 5237, Montpellier, France

Abstract

Many insect species harbor *Wolbachia* bacteria that induce cytoplasmic incompatibility (CI), i.e. embryonic lethality in crosses between infected males and uninfected females, or between males and females carrying incompatible *Wolbachia* strains. The molecular mechanism of CI remains unknown, but the available data are best interpreted under a *modification-rescue* model, where a *mod* function disables the reproductive success of infected males' sperm, unless the eggs are infected and express a compatible *resc* function. Here we examine the evolution of CI in the mosquito *Culex pipiens*, harbouring a large number of closely related *Wolbachia* strains structured in five distinct phylogenetic groups. Specifically, we used a worldwide sample of mosquito lines to assess the hypothesis that genetic divergence should correlate with the divergence of CI properties on a low evolutionary scale. We observed a significant association of *Wolbachia* genetic divergence with CI patterns. Most *Wolbachia* strains from the same group were compatible whereas those from different groups were often incompatible. Consistently, we found a strong association between *Wolbachia* groups and their *mod-resc* properties. Finally, lines from the same geographical area were rarely incompatible, confirming the conjecture that the spatial distribution of *Wolbachia* compatibility types should be constrained by selection. This study indicates a clear correlation between *Wolbachia* genotypes and CI properties, paving the way toward the identification of the molecular basis of CI through comparative genomics.

Citation: Atyame CM, Labbé P, Dumas E, Milesi P, Charlat S, et al. (2014) *Wolbachia* Divergence and the Evolution of Cytoplasmic Incompatibility in *Culex pipiens*. PLoS ONE 9(1): e87336. doi:10.1371/journal.pone.0087336

Editor: Kostas Bourtzis, International Atomic Energy Agency, Austria

Received: October 10, 2013; **Accepted:** December 20, 2013; **Published:** January 31, 2014

Copyright: © 2014 Atyame et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study is the contribution of the Institut des Sciences de l'Evolution de Montpellier (Unité Mixte de Recherche, CNRS-IRD-UM2 5554). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors declare that they have no competing interests.

* E-mail: celestine.atyame-nten@univ-reunion.fr

‡ Current address: University La Réunion/CRVOI, Ste Clotilde, Réunion Island, France

Background

Wolbachia bacteria are among the most common endosymbionts of arthropods and filarial nematodes [1–4]. Maternally inherited through the egg cytoplasm, they manipulate their host reproduction by various means, all increasing the proportion of infected females over generations, thus favoring their own dispersal [5,6]. The most commonly described *Wolbachia*-induced phenotype in arthropods is cytoplasmic incompatibility (CI) [2]. CI is a form of conditional sterility resulting in embryonic lethality in diploid organisms [7] or in the production of male offspring in some haplo-diploid species [8]. CI occurs either in crosses between *Wolbachia* infected males and uninfected females or in crosses between males and females infected with incompatible strains of *Wolbachia*. CI is termed bidirectional if the death of embryos occurs in the two reciprocal crosses, or unidirectional, if only one cross is incompatible.

The molecular mechanisms underlying CI are currently unknown. However, cytological studies commonly show paternal chromosome condensation failure and abnormal segregation in the first mitotic division, leading to embryonic death [9]. These observations are currently best interpreted under a toxin/antitoxin model [10,11]. According to this model, *Wolbachia* in males modify

the sperm (the so-called modification, or *mod* factor) by depositing a kind of “toxin” during its maturation. *Wolbachia* in females, on the other hand, deposit an “antitoxin” (the *rescue*, or *resc* factor) in the eggs, so that the offspring of infected females can develop normally. The simple compatibility patterns seen in several insect host species [12–14] have initially led to the view that CI relied on a single pair of *mod/resc* genes. However, more complex patterns, such as those described in the mosquito *Culex pipiens* [15–17] suggest that CI is controlled by multiple *mod/resc* factors that interact in complex ways [17–20]. Here we are interested in the processes that shape the evolution of compatibility types within *Cx. pipiens*.

Mosquitos of the *Cx. pipiens* complex are infected by a variety of strains from the *wPip* *Wolbachia* clade. This diversity represents an ideal model to study the relationship between *Wolbachia* genetics and CI properties for the following reasons: (1) all *wPip* strains share a monophyletic origin within the *Wolbachia* B group as evidenced by *Wolbachia* multilocus strain typing methodology [21]; (2) the recent sequencing of fast evolving genes indicates that five phylogenetic groups can be distinguished within the *wPip* clade, referred to as *wPip*-I to *wPip*-V, [21,22]; (3) multiple infections have never been evidenced despite the use of sensitive polymorphic markers [17,21–26]; (4) finally, this system is characterized by

an unrivalled variability of compatibility types, including compatible as well as uni- and bi-directionally incompatible lines [15–17,27,28]. Such a variability relies on the rapid diversification of crossing types [29] and is independent from the nuclear background [16,17,30] or from other inherited symbionts known to manipulate insect reproduction [29].

In this study, we took opportunity of the recently worked-out *wPip* phylogeny to address the correlation between *wPip* genetic divergence and crossing properties.

Methods

Mosquito Collection and Isofemale Lines Maintenance

Culex pipiens larvae and pupae were collected in three countries (Tunisia in 2007, 2008 and 2009, Algeria in 2006 and 2008 and in New Mexico in 2012). None of the samples in any location were collected in protected areas, and these field studies did not involve endangered or protected species. No specific permission was required to collect mosquito larvae in public areas, and when collected on private land or in private residences, the owners or residents gave permission for the study to be conducted on their land or in their residences. Samples were reared to adulthood in laboratory and females were blood-fed to establish isofemale lines. Each egg raft (containing 100–300 eggs) was individually isolated for hatching and *Wolbachia* was genotyped by analysing two first-instar larvae (L1) (see below). For each locality, two isofemale lines carrying the same *wPip* group were maintained, whenever possible, to constitute replicates. Using the same procedure, two isofemale lines were established from samples collected in China in 2003. A total of 29 isofemale lines were thus established for the present study. In addition, 22 isofemale lines from laboratory stocks of various geographical origins were also used. They include one line from Tunisia [24]; two lines from La Reunion island [17]; four lines from Lebanon; four lines from Mauritius [22]; four lines from Mayotte [22]; three lines from France [22,24]; two lines from California [24,31]; one line from Italy [32] and one line from Turkey [24] (Table S1 in File S1). Isofemale lines were reared in 65 dm³ screened cages kept in a single room at 22 to 25°C, under a 12-h light/12-h dark cycle. Larvae were fed with a mixture of shrimp powder and rabbit pellets, while adults were fed with a honey solution.

wPip Strain Identification

Mosquito DNA was extracted using a CetylTrimethylAmmonium Bromide protocol (CTAB) [33]. The genotyping of *wPip* strains infecting isofemale lines was performed through PCR/RFLP tests on two ankyrin-domain genes, *ank2* and *pk1* [26,34]. Both genes clearly differentiate the five previously identified *wPip* groups (*wPip*-I to *wPip*-V) [21]. The *HinfI* restriction enzyme was used for the *ank2* gene, whereas the discrimination of the five *wPip* groups with the *pk1* gene was performed using a combination of *TaqI* and *PstI* restriction enzymes [22]. Digested DNA fragments were separated on 2% agarose gel electrophoresis.

Crossing Properties

Isofemale lines were reared for at least four generations before crossing to allow acclimation to laboratory conditions and to optimize mating and blood feeding. Reciprocal crosses were performed using 25–50 virgin females and an equivalent number of males. All individuals were 2–5 days old. Females were allowed to blood-feed five days after caging and their egg rafts were collected five days later and stored individually until hatching. Crossing relationships between isofemale lines were determined by examining eggs' hatching rate (HR) under a binocular microscope.

All unhatched egg rafts were checked for fertilization through observation of embryonic development as described by Duron & Weill [35].

The crossing relationships between two given isofemale lines were categorized as follows:

- Compatible (C) when HR was >90% in the two reciprocal crosses;
- Incompatible (IC), with two CI patterns: uni-directionally incompatible crosses (UIC), when HR was 0% in one of the reciprocal crosses and >90% in the other, and bi-directionally incompatible crosses (BIC), when HR was 0% in both reciprocal crosses. Note that crosses with intermediate HR (90% > HR > 0%) represented less than 5% of all crosses and were discarded from the analysis.

We examined the variability in the crossing properties of isofemale lines through reciprocal crossing of each line with 4 reference isofemale lines: Lv (*wPip*-II), Mc and SI (*wPip*-III) and Is (*wPip*-IV), already used as references in a previous investigation [17]. For each studied line, the outcome of crossing males with females of the 4 reference lines defines the male crossing type or CT (*mod* ability) while the outcome of crossing of females with males of the 4 reference lines defines the female CT (*resc* ability). The resulting cytotypes, referred to herein as 4-ref-cytotypes (4RCTs), correspond to the combination of male and female CTs (8 crosses for each one).

Statistical Analyses

IC frequency in intra-group and in inter-group crosses. Using the data shown in Table 1, we performed two analyses to understand how phylogenetic groups affect compatibility, first between C vs IC crosses and second by distinguishing among IC crosses between UIC and BIC.

• C vs IC crosses

We first tested if the probability for two strains to be compatible was different if they belonged to the same *wPip* group (Intra*wPip*) or if they were from two different groups (Inter*wPip*). We computed the generalized linear model (GLM) $\text{PROPIC} = \text{CROSS} + \epsilon$, where PROPIC is a two-level variable corresponding to the proportions of IC and C crosses (with IC = UIC+BIC), and CROSS a two-level factor indicating whether the crosses are intra or inter *wPip* groups. ϵ is the error parameter, following a binomial distribution to take over-dispersion into account, if present. We tested the significance of the CROSS factor using likelihood ratio tests (LRT), as described in Crawley [36].

We then tested just for an Intra*wPip* group effect on PROPIC: we used the same model and procedures as above, with CROSS being a five-level factor (corresponding to the five *wPip* groups).

Finally, in Inter*wPip* crosses, we tested whether PROPIC of a given *wPip* group depends on the *wPip* group it was crossed with. We again used the same model and procedures as above, with CROSS being a ten-level factor (corresponding to the ten possible *wPip*_i × *wPip*_j crosses between two of the five *wPip* groups).

• C vs UIC vs BIC crosses

We then tested whether the probability for two strains to be bi-directionally rather than uni-directionally incompatible was different in Inter*wPip* than in Intra*wPip* crosses (i.e. dividing IC crosses between UIC and BIC). We computed the multinomial log-linear model $\text{PROPIC} = \text{CROSS} + \epsilon$. PROPIC is a three-level variable corresponding to the proportions of C, UIC and BIC

Table 1. Crossing relationships of *Culex pipiens* isofemale lines according to wPip groups.

Categories of crosses		Total	C		UIC		BIC	
			N	Mean (SD)	N	Mean (SD)	N	Mean (SD)
Within wPip groups	wPip-I/wPip-I	168	160	0.94 (0.016)	8	0.06 (0.015)	0	0
	wPip-II/wPip-II	6	6		0		0	
	wPip-III/wPip-III	9	5		4		0	
	wPip-IV/wPip-IV	19	19		0		0	
	wPip-V/wPip-V	1	1		0		0	
	Total	203	191		12		0	
Between wPip groups	wPip-I/wPip-II	45	29	0.45 (0.03)	11	0.32 (0.03)	5	0.23 (0.026)
	wPip-I/wPip-III	67	41		26		0	
	wPip-I/wPip-IV	36	0		2		34	
	wPip-I/wPip-V	10	10		0		0	
	wPip-II/wPip-III	14	8		6		0	
	wPip-II/wPip-IV	22	0		16		6	
	wPip-II/wPip-V	7	4		3		0	
	wPip-III/wPip-IV	42	15		17		10	
	wPip-III/wPip-V	6	6		0		0	
	wPip-IV/wPip-V	3	0		0		3	
	Total	252	113		81		58	

Total indicates the total number of reciprocal crosses performed to established CI patterns, and N the number of crosses that were compatible (C), uni-directionally incompatible (UIC) and bi-directionally incompatible (BIC). SD = standard deviation. In incompatible crosses, HR = 0%; in compatible crosses, HR >90%. For more details about crosses within wPip groups see Tables S2, S3, S4, S5 and S6 in File S1 whilst for crosses between wPip groups see Tables S7, S8, S9 and S10 in File S1.

doi:10.1371/journal.pone.0087336.t001

crosses and CROSS a two-level factor (Intra w Pip vs Inter w Pip). ε is the error parameter, following a multinomial distribution. We tested the significance of the CROSS factor using LRT as above.

As above, we then tested for a wPip group effect (considering only the Intra w Pip crosses) and for an Inter w Pip-cross effect on PROPIC, using multinomial log-linear models instead of GLM.

Distribution of the 4-ref-cytotypes among the wPip groups. We performed a Fisher's exact test [37] to test for independence between 4RCTs and wPip groups. We next used pairwise comparisons using Fisher's exact test to compare the 4RCT distributions between wPip groups. P-values were corrected using Hommel's sequential Bonferroni correction to take multiple testing into account [38].

Results and Discussion

The purpose of this study was to examine the correlation between genetic divergence and compatibility among wPip strains, motivated by our recent work showing a monophyletic origin of wPip strains and their organization into five genetic groups [21]. We examined a large dataset of crosses between *Cx. pipiens* isofemale lines from different geographic origins, infected either with strains from the same wPip group (Intra w Pip crosses) or with strains from different wPip groups (Inter w Pip crosses). All *Wolbachia* strains were unambiguously assigned to one wPip group using the PCR/RFLP assay on the two ankyrin-domain genes, *ank2* and *pk1* described in Dumas et al. [22] (see Methods).

We analyzed crosses of 72 isofemale lines infected with wPip strains from various groups and collected in 18 countries: 35 wPip-I from ten countries (Benin, Tunisia, Philippines, Greece, France [metropolitan, Reunion and Mayotte islands], Spain, Lebanon and Mauritius), four wPip-II from three countries (Australia, France and Cyprus), six wPip-III from three countries (California,

New Mexico and France), 24 wPip-IV from five countries (Algeria, Tunisia, Turkey, Italy and China) and three wPip-V from two countries (China and Philippines) (Table S1 in File S1). A total of 455 reciprocal crosses (i.e. 910 crosses) including 260 new reciprocal crosses (i.e. 520 crosses) and 195 reciprocal crosses (i.e. 390 crosses) from previous surveys [16,17,26] were examined.

Compatibility among wPip Strains Correlates with their Genetic Relatedness

To the noticeable exception of the wPip-III group, most crosses involving lines infected with strains from the same wPip group (Intra w Pip crosses) were compatible (Figure 1 and Table 1). CI in the wPip-I group occurred in less than 5% crosses (8 out of 168). However, intragroup CI was not found in crosses between strains of the same area and only affected strains from different geographic origins (Table 2). This supports the theoretical prediction that because of selection, only compatible *Wolbachia* strains can stably coexist in panmictic host populations [39]. A striking case of CI pattern is the Is line, infected with a wPip-IV strain and long known to induce CI when crossed with most other lab lines [16,17,40]; however, crosses between Is and other wPip-IV-infected lines were all compatible (Table 1 and Table S5 in File S1).

The wPip-III group significantly differed from other groups ($P = 0.006$), showing higher CI levels (Table 1 and Table S4 in File S1). However, this was estimated from a limited number of crosses between six strains of the wPip-III group and the high CI level mainly pertained to two lines (Sl and Mc), which induce opposed CI patterns when crossed with other wPip groups (Tables S7–S9 in File S1). They both originate from California, where CI was reported in 1980 [27], suggesting that the wPip-III group might be more heterogeneous than measured with polymorphic markers

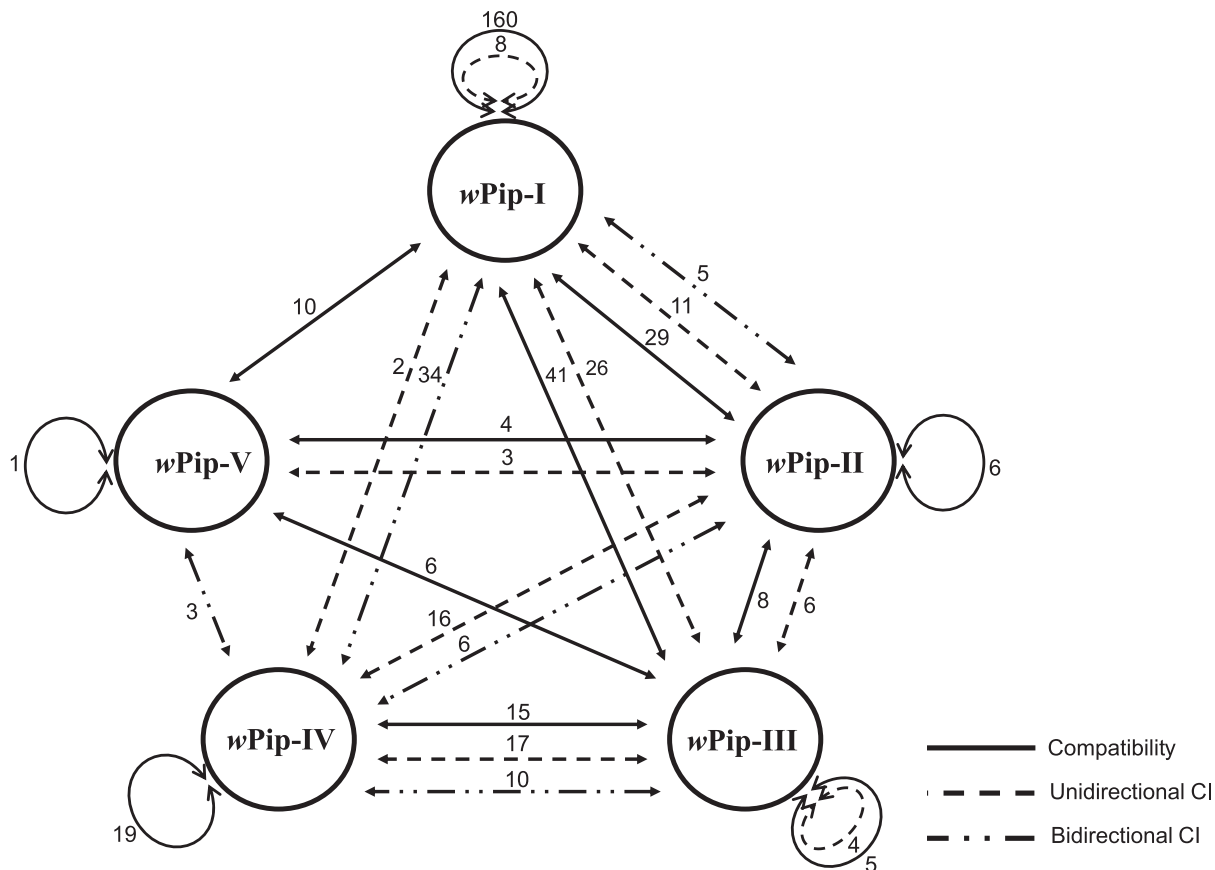


Figure 1. Schematic representation of the crossing relationships between *Culex pipiens* lines infected with different *wPip* groups. Numbers indicates the number of reciprocal crosses analyzed. In all compatible crosses, hatching rate (HR) >90% and in incompatible crosses, HR = 0%.
doi:10.1371/journal.pone.0087336.g001

used in this study. On the other hand, the difference between SI and Mc lines may result from genetic drift since their sampling in 1950 and 1984, respectively [24,31]. Indeed, *Cx. pipiens* lines can modify their crossing types in only 50 generations in laboratory conditions [29]. Would it be the case, evolution in the laboratory would more likely concern the SI line since Mc displays CI patterns identical to the *wPip-III* group line Albu-3 sampled in 2012 from New Mexico (Table S4 and Tables S8–S9 in File S1).

In contrast to intra-group crosses, CI occurred more frequently in crosses between lines infected with different *wPip* groups (Inter*wPip* crosses) (mean frequency of compatible crosses of 0.94 ± 0.016 in Intra*wPip* crosses *vs.* 0.45 ± 0.03 in Inter*wPip* crosses, $P < 0.001$). A significant effect of *wPip* group combinations on the extent of CI was detected ($P < 0.001$). An illustration is the *wPip-I* group, half-compatible (29/45) with the *wPip-II* group, fully incompatible (36/36) with the *wPip-IV* group and fully compatible (10/10) with the *wPip-V* group (Table 1 and Tables S7 in File S1). Similar results were also obtained in other group combinations. This variability mainly relies on the polymorphism of *wPip* genomes because the stability of CI properties over *Cx. pipiens* life span observed in previous investigations excluded the role of other factors such as density levels, nuclear background or sperm competitive ability [41,42]. In addition, males from five *wPip* infected lines [Tn (*wPip-I*), Lv (*wPip-II*), Mc and SI (*wPip-III*) and Is (*wPip-IV*)] displaying incompatibility with infected females always show full compatibility when uninfected [16,35].

Taken together, these results establish that the genetic proximity of *wPip* strains correlates with their compatibility. This issue could not be addressed without the knowledge of the *wPip* phylogeny, only recently worked-out [21]. Although such a correlation was hypothesized, previous surveys in *Drosophila* produced contrasting results: Charlat et al. [43] found compatibility between genetically close *Wolbachia* strains, whilst in other investigations, closely related bacteria appeared totally or partially incompatible [20,44]. However, comparing our large survey to these previous studies cannot be straightforward since we examined a much higher number of crosses and *wPip* strains displayed a much lower level of genetic divergence than the strains used in the other studies. Indeed, Charlat et al. [43] compared two *Wolbachia* sister strains considered as genetically identical from analysis of the *wsp* gene only, while in the two other studies, the *Wolbachia* strains were genetically closely related yet showed differences in their *wsp* sequences [20,44]. By contrast, all *wPip* strains studied here have strictly identical *wsp* genes and could be only discriminated on the basis of other fast evolving markers such as ankyrin genes and mobile genetic elements including prophages and transposable elements [21].

In conclusion, this analysis shows that except for group III, mosquito lines infected with the same *wPip* groups have a very high probability to be compatible. By contrast, one cannot predict the CI outcome of crosses between mosquito lines infected with different *wPip* groups, despite the frequent occurrence of CI.

Table 2. Crossing relationships between isofemale lines infected with strains from the wPip-I group and from different geographic origins.

wPip-I females	wPip-I males													
	La Réunion				Mauritius				Mayotte				Tunisia	
	Leu-118				Mau-2				Mau-4				May-6	
	Pie-11	C (8)*	C (8)*	C (8)*	C (16)	C (21)	C (17)	C (18)	C (24)	C (24)	C (23)	C (23)	C (24)	C (24)
	Leu-118	C (8)*	C (18)	C (15)	C (12)	C (11)	C (12)	C (15)	C (16)	C (16)	C (19)	C (19)	C (18)	C (24)
	Mau-2	C (18)	C (15)	C (11)	C (12)	C (8)	C (11)	C (12)	C (15)	C (16)	C (11)	C (11)	C (10)	C (16)
	Mau-4	C (17)	C (11)	C (12)	C (15)	C (16)	C (16)	C (12)	C (12)	C (19)	C (19)	C (19)	C (18)	C (21)
	Mau-5	C (9)	C (6)	C (10)	C (14)	C (11)	C (16)	C (12)	C (17)	C (17)	C (17)	C (17)	C (13)	C (12)
	Mau-7	C (15)	C (10)	C (10)	C (14)	C (11)	C (16)	C (12)	C (17)	C (17)	C (17)	C (17)	C (13)	C (12)
	May-4	C (9)	C (10)	C (10)	C (14)	C (11)	C (16)	C (12)	C (17)	C (17)	C (17)	C (17)	C (13)	C (12)
	May-5												C (15)	C (17)
	May-6												C (14)	C (16)
	May-17	C (14)	C (6)											
	Tn	C (24)	C (18)	C (24)	C (15)	C (15)	C (23)	C (22)					C (24)	C (25)
	Sok	C (7)	C (16)	C (22)	C (21)	C (18)	C (17)	C (17)					C (30)	C (18)
	Zer11-1	C (12)	C (16)	C (24)	C (18)	C (18)	C (16)	C (16)					C (17)	C (22)
	Zer11-2	C (27)	C (18)	C (17)	C (17)	C (21)	C (20)	C (20)					C (18)	C (17)
	AinT11-1	C (11)	C (18)										C (12)	
	AinT11-2	C (12)	C (12)										C (25)	
	Lib-1	C (13)	C (10)	C (12)	C (12)	C (15)	C (16)	C (16)					C (20)	C (13)
	Lib-2	C (13)	C (14)	C (12)	C (19)	C (17)	C (15)	C (15)					C (15)	C (12)
	Lib-3	C (21)	C (12)	C (16)	C (16)	C (14)	C (14)	C (14)					C (16)	C (15)
	Lib-4	C (10)	C (16)	C (15)	C (14)	C (10)	C (16)	C (16)					C (12)	C (17)

Crosses were classified either compatible (C, hatching rate (HR) >90%) or incompatible (IC, HR = 0%, in bold). The number of egg rafts collected in each cross is bracketed. Boxed crosses were performed between mosquito lines from the same population. * Crosses corresponding to data from Atayme et al. [17]. Note that crosses between mosquitoes from the same isofemale line are always compatible.

doi:10.1371/journal.pone.0087336.t002

Table 3. Summary of 4-ref-cytotypes (4RCTs) and male and female crossing types (*mod* and *resc* abilities) identified among the 51 *Culex pipiens* isofemale lines infected with the five *wPip* groups.

4-ref-cytotypes	males crossing types					females crossing types					Distribution of 4RCTs in <i>wPip</i> groups				
	Lv	Mc	SI	Is	<i>mod</i>	Lv	Mc	SI	Is	<i>resc</i>	<i>wPip</i> -I	<i>wPip</i> -II	<i>wPip</i> -III	<i>wPip</i> -IV	<i>wPip</i> -V
I	C	C	C	C	i	IC	IC	C	IC	1	1				
II	IC	C	IC	IC	ii	IC	IC	C	IC	1	5				
III	C	C	C	IC	iii	IC	IC	C	IC	1	2				
IV	IC	C	IC	IC	ii	C	C	C	IC	2	2		1		
V	C	C	C	IC	iii	C	C	C	IC	2	12	1			2
VI	C	IC	C	IC	iv	C	C	C	IC	2	1				
VII	IC	IC	IC	IC	v	C	C	C	IC	2	1				
VIII	C	C	IC	IC	vi	C	C	C	IC	2	1	1			
IX	C	C	IC	C	vii	C	C	C	IC	2		1	2		
X	C	C	C	C	i	C	C	C	IC	2		1			
XI	C	C	C	C	i	IC	C	IC	C	3				7	
XII	C	C	IC	C	vii	IC	C	IC	C	3				7	
XIII	IC	IC	C	C	viii	IC	C	IC	C	3				2	
XIV	C	C	C	IC	iii	IC	IC	C	C	4			1		
Total	14				8					4	25	4	4	16	2

The cytotypes were determined based on reciprocal crosses between the 51 isofemale lines and four reference laboratory lines (see text): Lv (*wPip*-II), Mc and SI (*wPip*-III) and Is (*wPip*-IV). C = compatible cross (all hatching rate, HR >90%); IC = incompatible cross (bolded cells, HR = 0%).

doi:10.1371/journal.pone.0087336.t003

Bi-directional CI only Occurs between Mosquito Lines Infected with Divergent *wPip* Groups

To test if *wPip* groups also predict CI patterns, incompatible crosses were subdivided in UIC and BIC. BIC was never observed among Intra*wPip* crosses, all incompatible crosses (6%, $n = 12/203$) being UIC. Among the 139 incompatible Inter*wPip* crosses, 58% ($n = 81$) and 42% ($n = 58$) were respectively UIC and BIC, and most BIC ($n = 53$) involved the *wPip*-IV group (Figure 1 and Table 1). We found a significant effect of the nature of the *wPip* group combination on CI patterns ($P < 0.001$). For instance, the *wPip*-I group showed more UIC than BIC with the *wPip*-II group ($n = 11$ vs. $n = 5$), only UIC with the *wPip*-III group ($n = 26$) and more BIC than UIC with the *wPip*-IV group ($n = 34$ vs. $n = 2$) (Figure 1, Table 1 and Table S7 in File S1). Although generating the highest BIC rates, the *wPip*-IV group nevertheless showed variable rates, from 27.3% (*wPip*-II crosses, $n = 22$) to 94.4–100% (*wPip*-I and *wPip*-III crosses, $n = 36$ and $n = 3$, respectively). This extends further the heterogeneity in the CI patterns of each *wPip* group when confronted to other groups.

Table 4. *P*-values for the pairwise comparisons of 4-ref-cytotypes (4RCTs) distributions between *wPip* groups.

	<i>wPip</i> -I	<i>wPip</i> -II	<i>wPip</i> -III	<i>wPip</i> -IV
<i>wPip</i> -II	0.12			
<i>wPip</i> -III	0.008	1		
<i>wPip</i> -IV	<i>8.05 × 10⁻¹⁰</i>	0.0014	0.0006	
<i>wPip</i> -V	1	1	0.2	0.013

Fisher's exact tests were computed from the Table 3 data. Significant *P*-values (<0.05) are in italics. *P*-values still significant after Hommel's sequential Bonferroni correction for multiple testing are bolded.

doi:10.1371/journal.pone.0087336.t004

Our finding that bi-directional CI only affects crosses between genetically different *wPip* groups corroborates results of previous studies showing bi-directional CI between divergent *Wolbachia* strains [14,45]. These data fit the model according to which multiple *mod/resc* functions control CI patterns in *Wolbachia* infecting *Cx. pipiens* [17–19]. Although the *mod/resc* functions responsible for mutual compatibility are expected to show little variability within a same *wPip* group, other *mod/resc* functions not involved in mutual compatibility should be neutral thus more prone to diverge between *wPip* groups and might occasionally produce BIC.

Wolbachia Genetic Divergence and the Evolution of *mod* and *resc* Properties

Wolbachia strains can be characterized by their crossing types (CT) or cytotypes (i.e. compatible, uni-directionally or bi-directionally incompatible) with different strains [39]. Cytotypes can be divided into male CT (*mod* ability) and female CT (*resc* ability). We reported previously that *Wolbachia* strains from the *wPip*-I group with identical genotypes could nevertheless display distinct male and female CTs when crossed with genetically distant *wPip* strains [17]. To examine how *mod* and *resc* abilities evolved within and among the five *wPip* groups, we specifically tested whether cytotypes were distributed at random (i.e. different *wPip* groups share same cytotypes) or showed preferential distribution into specific *wPip* groups.

Since we could not reasonably examine by reciprocal crossing the 51 *Cx. pipiens* isofemale lines (25 infected with *wPip*-I strains, 4 with *wPip*-II, 4 with *wPip*-III, 16 with *wPip*-IV and 2 with *wPip*-V), we used the restricted 4RCT (4-ref-cytotype), corresponding to the combination of four male and four female CTs identified by reciprocal crossing with 4 isofemale lines arbitrarily chosen as references: Lv (*wPip*-II), Mc and SI (*wPip*-III) and Is (*wPip*-IV). Overall, we identified eight distinct male CTs (i to viii, *mod*

abilities, Table 3) and four distinct female CTs (1 to 4, *resc* abilities) combined into fourteen 4RCTs (I to XIV). *wPip* groups globally displayed fewer *resc* than *mod* abilities, *wPip*-II being an extreme case with a single *resc* and all different *mod* abilities. Theory predicts that the evolution of *mod* functions should be more constrained by selection than the evolution of *resc* functions [46]. Indeed, changing a *resc* function is counter-selected because it renders the mutant unable to ensure its transmission. On the contrary, changing a *mod* function only makes infected males incompatible with resident strains, which is neutral in a panmictic population because males do not transmit the infection. Consistent with this view, we observed a larger polymorphism of the *mod* than the *resc* function in the data set.

The 4RCTs were not randomly distributed between the *wPip* groups (Fisher's exact test, $P < 0.001$): ten 4RCTs were specific to a single group (for example 4RCTs I and XI are specific to *wPip*-I and *wPip*-IV, respectively), while four 4RCTs were shared by several groups, such as the 4RCT V shared by *wPip*-I, *wPip*-II and *wPip*-V. The *wPip*-IV group harbors three specific 4RCTs (Table 3), which makes it significantly different from the others in pairwise comparisons (Fisher's exact test, P -value < 0.05 for all four comparisons, only three remaining significant after Hommel's sequential Bonferroni correction, Table 4).

Analysis of the *mod* and *resc* abilities (Table 3) showed a clear partitioning between *wPip* groups: *wPip*-IV displayed exclusively the *resc* 3 ability (16/16) and almost exclusively the *mod* i and *mod* vii abilities (14/16), whereas *wPip*-I, -II, -III and -V mainly displayed the *resc* 2 ability (26/35) and the *mod* ii and *mod* iii abilities (26/35). The special situation of *wPip*-IV is consistent with the fact that it is involved in 91.4% of crosses that produced BIC (Table 1).

We further examined the independence between the *mod* and *resc* abilities using their respective frequencies deduced from Table 3. As expected from their linked transmission, the two variables were not independent (Fisher's exact test, $P < 0.001$).

Worldwide Distribution of *wPip* Strains does not Correlate with CI Patterns

We recently highlighted a clear spatial structure of *wPip* groups over *Cx. pipiens* distribution range: *wPip*-I and *wPip*-III are largely spread over different continents, whereas *wPip*-II is restricted to Western Europe, *wPip*-V to Asia, and *wPip*-IV sporadically present in Europe, Asia and North Africa [22]. We then asked whether the large geographic distribution of the *wPip*-I and *wPip*-III groups could be due to more invasive CI properties. The theory on *Wolbachia* dynamics in a panmictic host population predicts that a strain X can invade a population infected by a strain Y if males X induce CI (mod^{X+}) and if females X rescue CI induced by most of the males Y ($resc^{X+,Y+}$) [46]. As shown in Table S7 in File S1, when crossed with the *wPip*-II and *wPip*-III strains infected females, *wPip*-I males induced modest CI (28.9% (13/45) and 25.4% (17/67), respectively) while *wPip*-I females efficiently rescued CI (82.2% (37/45) and 86.6% (58/67), respectively). Almost all of crosses with *wPip*-IV were bi-directionally incompatible, while crosses with *wPip*-V were fully compatible, a situation which does not favor invasion in either case. Taken together, this suggests that the large geographic distribution of *wPip*-I is independent from invasive CI properties. The same conclusion stands for *wPip*-III (Tables S8–S9 in File S1), which was fully compatible with *wPip*-V, induced low to moderate CI with *wPip*-II and *wPip*-IV females (14.3%, 2/14; 50%, 21/42, respectively) and rescued quite efficiently CI with *wPip*-II and *wPip*-IV males (71.4%, 10/14; 61.9%, 26/42, respectively). Therefore, considering that all known *Cx. pipiens* populations are infected by *Wolbachia*, reasons other

than CI properties should be invoked to explain the present large distribution of *wPip*-I and *wPip*-III. This may be a consequence of passive migration due to human activities, a process shown to be responsible for long-distance gene flow [47]. Alternatively, *wPip*-I and *wPip*-III infections might confer selective advantages, e.g. higher female fecundity as is the case with the mosquito *Aedes albopictus* [48], or protection against natural enemies as described in *Drosophila melanogaster* [49,50].

Conclusion

In this study, we show a clear correlation between genetic divergence of *Wolbachia* strains infecting *Cx. pipiens* mosquitos and crossing relationships: crosses within same genetic groups were mostly compatible and showed no bi-directional CI. Future investigations using theoretical models like parsimony inference models [18,19] should help addressing how *mod* and *resc* determinants in each *wPip* group may interplay to explain the observed phenotypes. This is a critical issue for the development of new control strategies of arthropod disease-vector and pest populations, for which *Wolbachia* are now considered as promising tools [51]. The large database of CI relationships in the *Cx. pipiens* complex described here should help identifying candidate genes responsible for CI properties by testing their correlation with distinct *mod* and *resc* abilities groups.

Supporting Information

File S1 Supporting file contains Tables S1–S10. Table S1. *Culex pipiens* isofemale lines. Table S2. Reciprocal crosses between isofemale lines infected with *wPip* strains from the *wPip*-I group. (A) Reciprocal crosses between isofemale lines from La Réunion Island according to Atyame et al [17]. (B), Crosses between isofemale lines from Tunisia (Tn), Philippines (Ma-B), France (Bf-A), Grece (Ko), Spain (Ep-A and Ep-B) were performed in previous studies [16,26] and Cotonou (Cot-A and Cot-B) were performed for this study. Crosses were classified either compatible (C, hatching rate (HR) $> 90\%$) or incompatible (IC, HR = 0%, shaded). The number of egg-rafts collected in each cross is bracketed. Note that crosses between mosquitoes from the same isofemale line are always compatible. Table S3. Reciprocal crosses between isofemale lines infected with *wPip* strains from the *wPip*-II group. Isofemale lines were isolated from samples collected in France (Lv), Brisbane (Au) and Cyprus (Ke-A and Ke-B). All crosses were performed by Duron et al. [16]. C = compatible crosses (HR $> 90\%$). The number of egg-rafts collected in each cross is bracketed. Table S4. Reciprocal crosses between isofemale lines infected with *wPip* strains from the *wPip*-III group. Isofemale lines were isolated from samples collected in California (Sl and Mc), New Mexico (Albu-3) and France (Bf-B, Trio-2 and Trio-7). *, Crosses corresponding to data from Duron et al. [16]. Crosses were classified either compatible (C, for HR $> 90\%$) or incompatible (IC, HR = 0%, shaded). The number of egg-rafts collected in each cross is bracketed. Table S5. Reciprocal crosses between isofemale lines infected with *wPip* strains from the *wPip*-IV group. (A and B), reciprocal crosses between the isofemale line Is (from Turkey) and the isofemale lines from Tunisia (Bou-1, Bou-2, Kef-1, Kef-2, Tab-1, Tab-2), from Algeria (Dou-1, Dou-2, Guel-1, Guel-2, Kal-1, Kal-2, Lac-1, Lac-2, Souk-2, Ha) and from Italy (CAA). (C), reciprocal crosses between the isofemale lines from Tunisia. C = compatible crosses (HR $> 90\%$). The number of egg-rafts collected in each cross is bracketed. Table S6. Reciprocal crosses between isofemale lines infected with *wPip*

strains from the *wPip-V* group. These crosses correspond to data from Duron et al. [16] and isofemale lines were from China (Kara-C) and Philippines (Ma-A). C = compatible crosses (HR >90%). The number of egg-rafts collected in each cross is bracketed. **Table S7. Crossing relationships between mosquito lines infected with *wPip* strains from the *wPip-I* group and lines infected with strains from *wPip-II*, *wPip-III*, *wPip-IV* and *wPip-V* groups.** (A) Between *wPip-I* infected males and females infected with *wPip-II*, *wPip-III*, *wPip-IV* and *wPip-V* groups. (B) Between *wPip-I* infected females and males infected with *wPip-II*, *wPip-III*, *wPip-IV* and *wPip-V* groups. *, Crosses corresponding to data from Duron et al. [16,26]. Crosses were classified either compatible (C, HR >90%) or incompatible (IC, HR = 0%, shaded). Bi-directionally incompatible crosses are underlined. The number of egg-rafts collected in each cross is bracketed. **Table S8. Crossing relationships between mosquito lines infected with *wPip* strains from the *wPip-II* group and lines infected with strains from *wPip-III*, *wPip-IV* and *wPip-V* groups.** (A) Between *wPip-II* infected males and females infected with *wPip-III*, *wPip-IV* and *wPip-V* groups. (B) Between *wPip-II* infected females and males infected with *wPip-III*, *wPip-IV* and *wPip-V* groups. *, Crosses corresponding to data from Duron et al. [16,26]. Crosses were classified either compatible (C, HR >90%) or incompatible (IC, HR = 0%, shaded). Bi-directionally incompatible crosses are underlined. The number of egg-rafts collected in each cross is bracketed. **Table S9. Crossing relationships between mosquito lines infected with *wPip* strains from the**

***wPip-III* group and lines infected with strains from *wPip-IV* and *wPip-V* groups.** (A) Between *wPip-III* infected males and females infected with *wPip-IV* and *wPip-V* groups. (B) Between *wPip-III* infected females and males infected with *wPip-IV* and *wPip-V* groups. *, Crosses corresponding to data from Duron et al. [16]. Crosses were classified either compatible (C, HR >90%) or incompatible (IC, HR = 0%, shaded). Bi-directionally incompatible crosses are underlined. The number of egg-rafts collected in each cross is bracketed. **Table S10. Crossing relationships between mosquito lines infected with *wPip* strains from the groups *wPip-IV* and *wPip-V*.** *, Crosses corresponding to data from Duron et al. [16]. IC = incompatible crosses (HR = 0%, shaded). Bi-directionally incompatible crosses are underlined. The number of egg-rafts collected in each cross is bracketed. (PDF)

Acknowledgments

We are very grateful to Nicole Pasteur, Olivier Duron, Mathieu Sicard, Anna Sophie Fiston-Lavier for helpful comments on the manuscript, and to Patrick Makoundou and Sandra Unal for technical assistance.

Author Contributions

Conceived and designed the experiments: CMA MW. Performed the experiments: CMA MW ED. Analyzed the data: CMA PL PM PF MW. Wrote the paper: CMA MW PF PL PM SC.

References

- Taylor MJ, Bandi C, Hoerauf A (2005) *Wolbachia* bacterial endosymbionts of filarial nematodes. *Adv Parasitol* 60: 245–84.
- Werren JH, Baldo L, Clark ME (2008) *Wolbachia*: master manipulators of invertebrate biology. *Nat Rev Microbiol* 6(10): 741–51.
- Hilgenboecker K, Hammerstein P, Schlattmann P, Telschow A, Werren JH (2008) How many species are infected with *Wolbachia*? A statistical analysis of current data. *FEMS Microbiol Lett* 281(2): 215–20.
- Zug R, Hammerstein P (2012) Still a host of hosts for *Wolbachia*: analysis of recent data suggests that 40% of terrestrial arthropod species are infected. *PLoS One* 7(6): e38544.
- Stouthamer R, Breeuwer JA, Hurst GD (1999) *Wolbachia pipiensis*: microbial manipulator of arthropod reproduction. *Annu Rev Microbiol* 53: 71–102.
- Engelstadter J, Hurst GDD (2009) The ecology and evolution of microbes that manipulate host reproduction. *Annu Rev Ecol, Evol and Systematics* 40: 127–49.
- Callaini G, Dallai R, Riparbelli MG (1997) *Wolbachia*-induced delay of paternal chromatin condensation does not prevent maternal chromosomes from entering anaphase in incompatible crosses of *Drosophila simulans*. *J Cell Sci* 110(2): 271–80.
- Tram U, Fredrick K, Werren JH, Sullivan W (2006) Paternal chromosome segregation during the first mitotic division determines *Wolbachia*-induced cytoplasmic incompatibility phenotype. *J Cell Sci* 119(17): 3655–63.
- Serbus LR, Casper-Lindley C, Landmann F, Sullivan W (2008) The genetics and cell biology of *Wolbachia*-host interactions. *Annu Rev Genet* 42: 683–707.
- Werren JH (1997) Biology of *Wolbachia*. *Annu Rev Entomol* 42: 587–609.
- Poinot D, Charlat S, Mercot H (2003) On the mechanism of *Wolbachia*-induced cytoplasmic incompatibility: confronting the models with the facts. *Bioessays* 25(3): 259–65.
- Riegler M, Stauffer C (2002) *Wolbachia* infections and superinfections in cytoplasmically incompatible populations of the European cherry fruit fly *Rhagoletis cerasi* (Diptera, Tephritidae). *Mol Ecol* 11(11): 2425–34.
- Mercot H, Charlat S (2004) *Wolbachia* infections in *Drosophila melanogaster* and *Drosophila simulans*: polymorphism and levels of cytoplasmic incompatibility. *Genetica* 120(1–3): 51–9.
- Dobson SL, Marsland EJ, Rattanadechakul W (2001) *Wolbachia*-induced cytoplasmic incompatibility in single- and superinfected *Aedes albopictus* (Diptera: Culicidae). *J Med Entomol* 38(3): 382–7.
- Laven H (1967) Speciation and Evolution in *Culex pipiens*. In: Wright J, Pal R, eds. *Genetics of Insect Vectors of Disease*. Amsterdam: Elsevier.
- Duron O, Bernard C, Unal S, Berthomieu A, Berticat C, et al. (2006) Tracking factors modulating cytoplasmic incompatibilities in the mosquito *Culex pipiens*. *Mol Ecol* 15(10): 3061–71.
- Atyame CM, Duron O, Tortosa P, Pasteur N, Fort P, et al. (2011) Multiple *Wolbachia* determinants control the evolution of cytoplasmic incompatibilities in *Culex pipiens* mosquito populations. *Mol Ecol* 20 (2): 286–298.
- Nor I, Hermelin D, Charlat S, Engelstadter J, Reuter M, et al. (2012) Mod/Resc Parsimony Inference: theory and application. *Information and Computation* 213: 23–32.
- Nor I, Engelstadter J, Duron O, Reuter M, Sagot MF, et al. (2013) On the genetic architecture of cytoplasmic incompatibility: inference from phenotypic data. *American Naturalist* 182(1): E15–E24.
- Zabalou S, Apostolaki A, Pattas S, Veneti Z, Paraskevopoulos C, et al. (2008) Multiple rescue factors within a *Wolbachia* strain. *Genetics* 178(4): 2145–60.
- Atyame CM, Delsuc F, Pasteur N, Weill M, Duron O (2011) Diversification of *Wolbachia* endosymbiont in the *Culex pipiens* mosquito. *Mol Biol Evol* 28(10): 2761–72.
- Dumas E, Atyame CM, Milesi P, Fonseca DM, Shaikevich EV, et al. (2013) Population structure of *Wolbachia* and cytoplasmic introgression in a complex of mosquito species. *BMC Evol Biol* 13(1): 181.
- Sanogo YO, Dobson SL (2004) Molecular discrimination of *Wolbachia* in the *Culex pipiens* complex: evidence for variable bacteriophage hyperparasitism. *Insect Mol Biol* 13(4): 365–9.
- Duron O, Lagnel J, Raymond M, Bourtsis K, Fort P, et al. (2005) Transposable element polymorphism of *Wolbachia* in the mosquito *Culex pipiens*: evidence of genetic diversity, superinfection and recombination. *Mol Ecol* 14(5): 1561–73.
- Duron O, Fort P, Weill M (2006) Hypervariable prophage WO sequences describe an unexpected high number of *Wolbachia* variants in the mosquito *Culex pipiens*. *Proc Biol Sci* 273(1585): 495–502.
- Duron O, Boureux A, Echaubard P, Berthomieu A, Berticat C, et al. (2007) Variability and expression of ankyrin domain genes in *Wolbachia* variants infecting the mosquito *Culex pipiens*. *J Bacteriol* 189(12): 4442–8.
- Barr AR (1980) Cytoplasmic incompatibility in natural populations of a mosquito, *Culex pipiens* L. *Nature* 283: 71–2.
- O'Neill SL, Paterson HE (1992) Crossing type variability associated with cytoplasmic incompatibility in Australian populations of the mosquito *Culex quinquefasciatus* Say. *Med Vet Entomol* 6(3): 209–16.
- Duron O, Bernard J, Atyame CM, Dumas E, Weill M (2012) Rapid evolution of *Wolbachia* incompatibility types. *Proc Biol Sci* 279(1746): 4473–80.
- Walker T, Song S, Sinkins SP (2009) *Wolbachia* in the *Culex pipiens* group mosquitoes: introgression and superinfection. *Journal of Heredity* 100: 192–6.
- Georgiopoulos GP, Metcalf RL, Gidden FE (1966) Carbamate-resistance in mosquitoes: selection of *Culex pipiens fatigans* Wied (= *Culex quinquefasciatus*) for resistance to Baygon. *Bulletin of the World Health Organization* 35: 691–708.
- Calvitti M, Moretti R, Lampazzi E, Bellini R, Dobson SL (2010) Characterization of a new *Aedes albopictus* (Diptera: Culicidae)-*Wolbachia pipiensis* (Rickettsiales: Rickettsiaceae) symbiotic association generated by artificial transfer of the *wPip* strain from *Culex pipiens* (Diptera: Culicidae). *J Med Entomol* 47(2): 179–87.

33. Rogers SO, Bendich AJ (1988) Extraction of DNA from plant tissues. In: Gelvin SB, Schilperoort RA, eds. Plant Molecular Biology Manual. Boston: Kluwer Academic Publishers: 1–10.
34. Sinkins SP, Walker T, Lynd AR, Steven AR, Makepeace BL, et al (2005) *Wolbachia* variability and host effects on crossing type in *Culex* mosquitoes. *Nature* 436(7048): 257–60.
35. Duron O, Weill M (2006) *Wolbachia* infection influences the development of *Culex pipiens* embryo in incompatible crosses. *Heredity* 96(6): 493–500.
36. Crawley M (2007) The R Book. John Wiley & Sons Ltd, Chichester, UK.
37. Fisher RA (1935) The logic of inductive inference. *Journal of the Royal Statistical Society* 98: 39–54.
38. Hommel G (1988) A stagewise rejective multiple test procedure based on a modified Bonferroni test. *Biometrika* 75: 383–6.
39. Rousset F, Raymond M, Kjellberg F (1991) Cytoplasmic incompatibilities in the mosquito *Culex pipiens*: how to explain a cytotype polymorphism? *J Evol Biol* 4: 69–81.
40. Atyame CM, Pasteur N, Dumas E, Tortosa P, Tantely ML, et al. (2011) Cytoplasmic incompatibility as a means of controlling *Culex pipiens quinquefasciatus* mosquito in the islands of the South-Western Indian Ocean. *PLoS Negl Trop Dis* 5(12): e1440.
41. Rasgon JL, Scott TW (2003) *Wolbachia* and cytoplasmic incompatibility in the California *Culex pipiens* mosquito species complex: parameter estimates and infection dynamics in natural populations. *Genetics* 165(4): 2029–38.
42. Duron O, Fort P, Weill M (2007) Influence of aging on cytoplasmic incompatibility, sperm modification and *Wolbachia* density in *Culex pipiens* mosquitoes. *Heredity* 98(6): 368–74.
43. Charlat S, Nirgianaki A, Bourtzis K, Mercot H (2002) Evolution of *Wolbachia*-induced cytoplasmic incompatibility in *Drosophila simulans* and *Drosophila sechellia*. *Evolution* 56(9): 1735–42.
44. Charlat S, Riegler M, Baures I, Poinot D, Stauffer C, et al. (2005) Incipient evolution of *Wolbachia* compatibility types. *Evolution* 58(9): 1901–8.
45. Bordenstein SR, Werren JH (2007) Bidirectional incompatibility among divergent *Wolbachia* and incompatibility level differences among closely related *Wolbachia* in *Nasonia*. *Heredity* 99(3): 278–87.
46. Charlat S, Calmet C, Mercot H (2001) On the *mod resc* model and the evolution of *Wolbachia* compatibility types. *Genetics* 159(4): 1415–22.
47. Raymond M, Callaghan A, Fort P, Pasteur N (1991) Worldwide migration of amplified insecticide resistance genes in mosquitoes. *Nature* 350: 151–3.
48. Dobson SL, Marsland EJ, Rattanadechakul W (2002) Mutualistic *Wolbachia* infection in *Aedes albopictus*: accelerating cytoplasmic drive. *Genetics* 160(3): 1087–94.
49. Hedges LM, Brownlie JC, O'Neill SL, Johnson KN (2008) *Wolbachia* and virus protection in insects. *Science* 322(5902): 702.
50. Teixeira L, Ferreira A, Ashburner M (2008) The bacterial symbiont *Wolbachia* induces resistance to RNA viral infections in *Drosophila melanogaster*. *PLoS Biol* 6(12): e2.
51. Vavre F, Charlat S (2012) Making (good) use of *Wolbachia*: what the models say. *Curr Opin Microbiol* 15(3): 263–8.