



Ion channels and properties of large neuronal networks: a computational study of re.nal waves during development

D Karvouniari, Lionel Gil, Olivier Marre, Serge Picaud, Bruno Cessac

► To cite this version:

D Karvouniari, Lionel Gil, Olivier Marre, Serge Picaud, Bruno Cessac. Ion channels and properties of large neuronal networks: a computational study of re.nal waves during development. Symposium on Ion channels and Channelopathies - IPMC, Nov 2018, Sophia Antipolis, France. hal-01925829

HAL Id: hal-01925829

<https://hal.science/hal-01925829v1>

Submitted on 17 Nov 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Ion channels and properties of large neuronal networks: a computational study of retinal waves during development.

D. Karvouniari, Biovision team, INRIA and LJAD, UCA

L. Gil, INLN, Sophia Antipolis

O. Marre, Institut de la Vision, Paris

S. Picaud, Institut de la Vision, Paris

B. Cessac, Biovision team, INRIA, Sophia Antipolis



Ion channels and properties of large neuronal networks: a computational study of retinal waves during development.

D. Karvouniari, Biovision team, INRIA and LJAD, UCA

L. Gil, INLN, Sophia Antipolis

O. Marre, Institut de la Vision, Paris

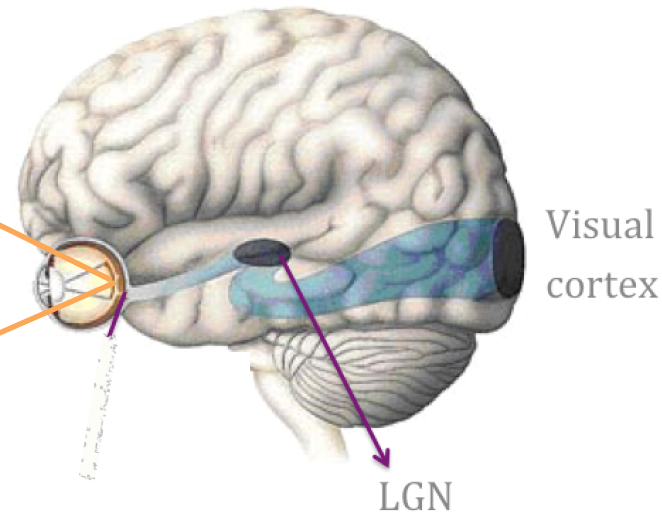
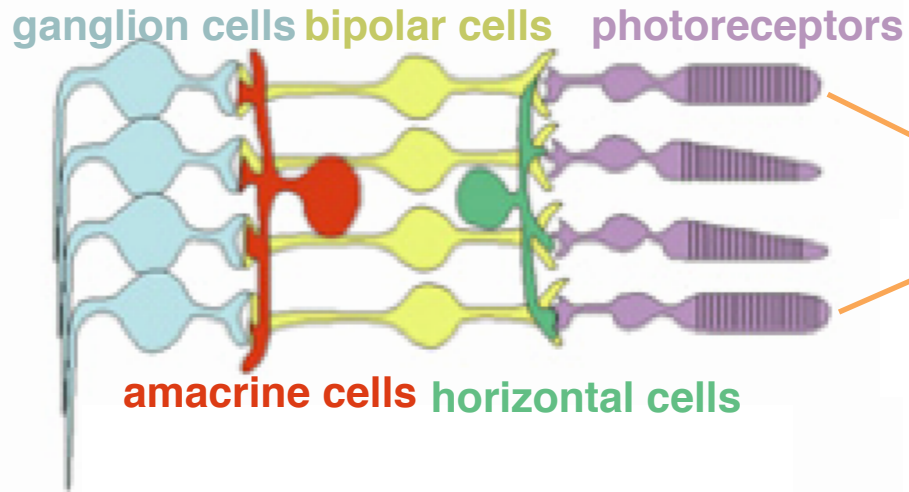
S. Picaud, Institut de la Vision, Paris

B. Cessac, Biovision team, INRIA, Sophia Antipolis



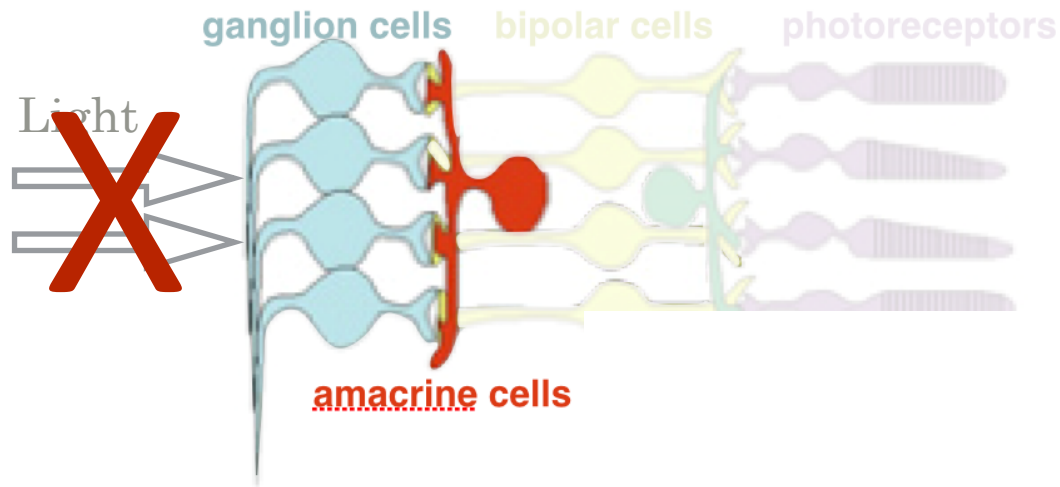
The structure of the adult retina

Retina's layered structure



The structure of the retina during development

Retina's layered structure
is shaped during development



But How?

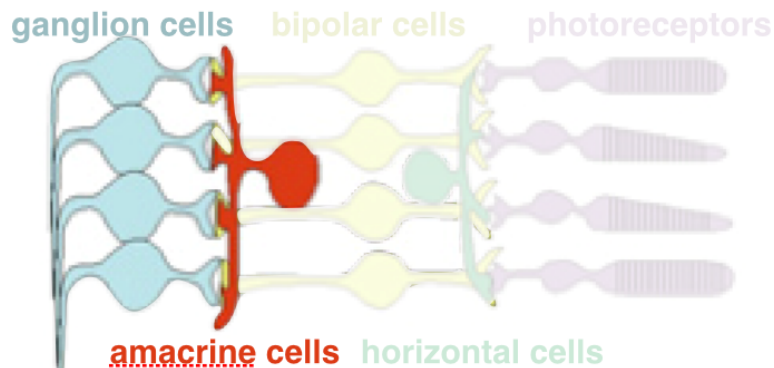


Retinal waves!

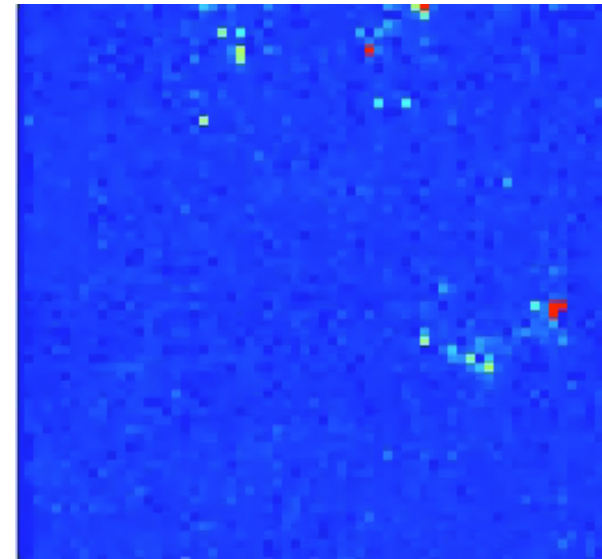
Retinal waves

Spontaneous spatio-temporal waves during development
Disappear short after birth
when vision is functional

Recordings from the retina



**Multi-electrode
array
(MEA)**



(Maccione et al. 2014)

*MEA recording of the voltage from
a P11 mouse retina in the presence
of 10 μ M bicuculline.*

Stages of Retinal Waves During Development

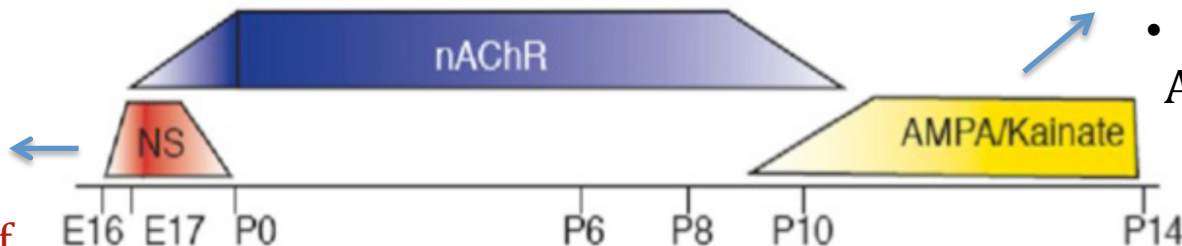
Stage II

- Retinotopic mapping
- Nicotinic Acetylcholine Receptors (nAChR)

Stage III

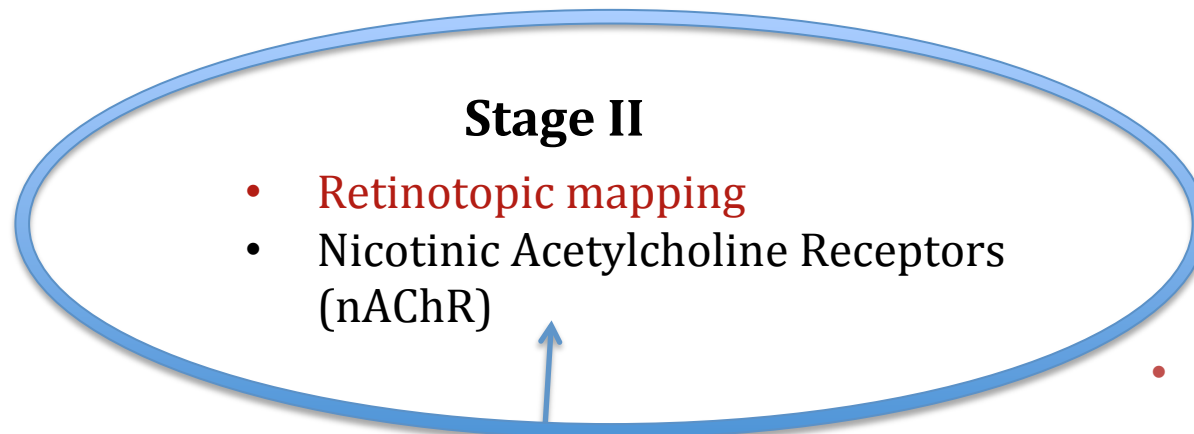
- Disappear when vision is functional
- Glutamate – AMPA receptors

Stage I



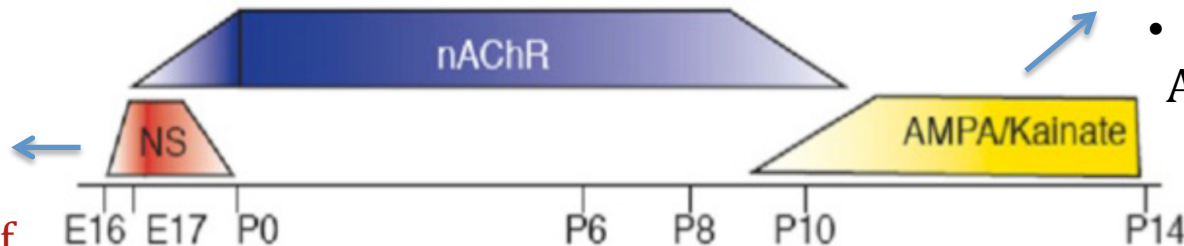
- Formation of retina circuitry
- Chemical synapses not formed yet
- Gap junction-mediated

Stages of Retinal Waves During Development



- Stage III**
- Disappear when vision is functional
 - Glutamate – AMPA receptors

Stage I

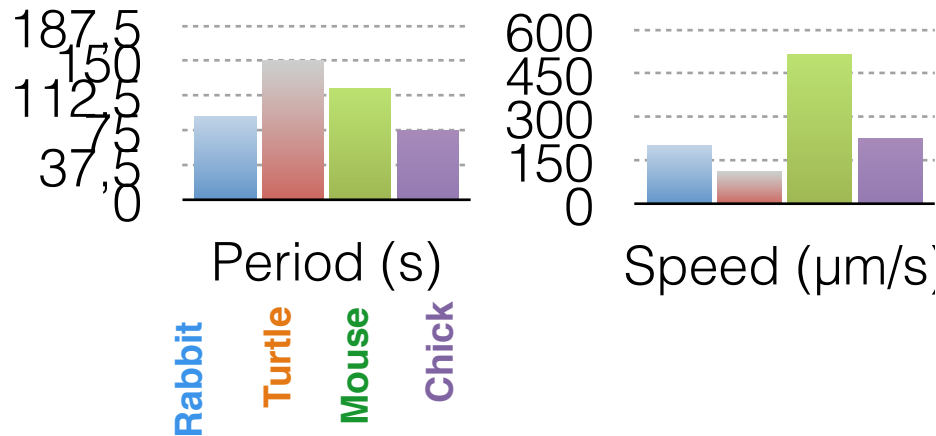


- Formation of retina circuitry
- Chemical synapses not formed yet
- Gap junction-mediated

Variability within retinal waves

i) Across species

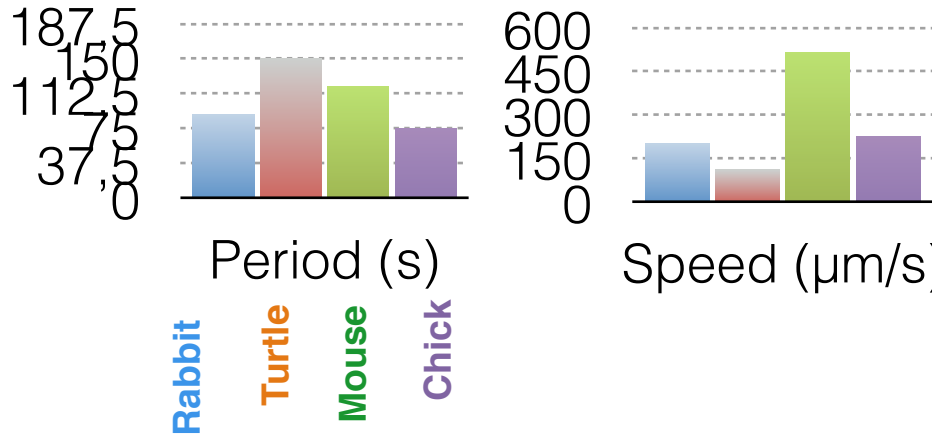
Godfrey et al. 2007



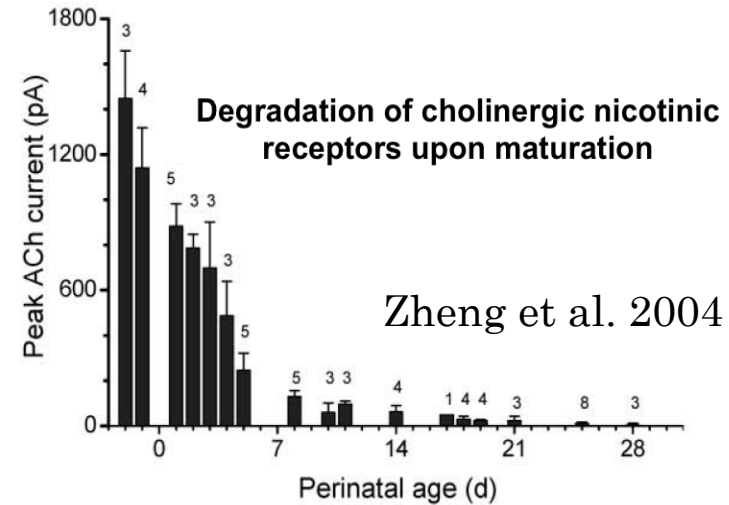
Variability within retinal waves

i) Across species

Godfrey et al. 2007



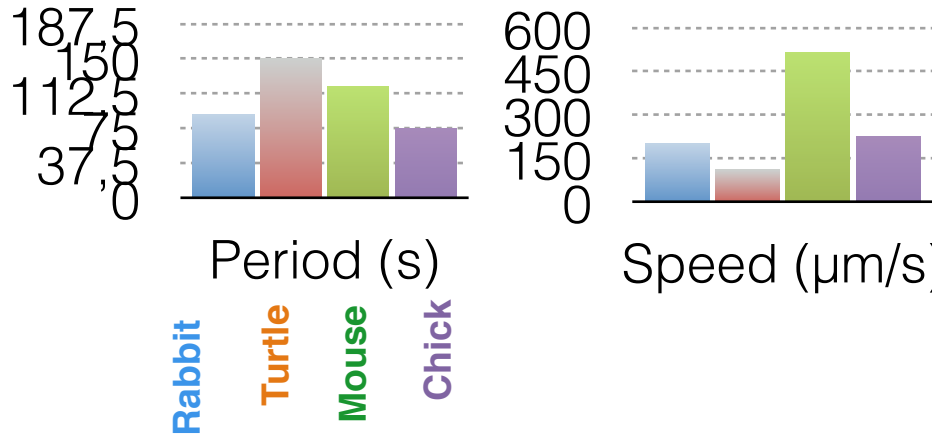
ii) Development



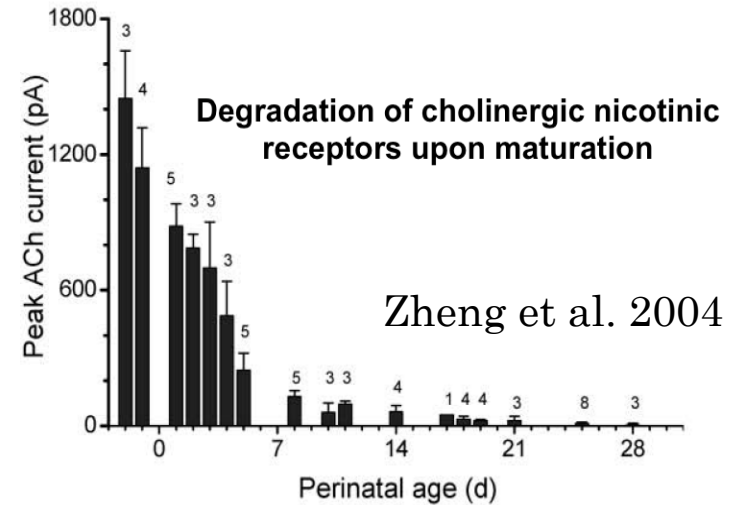
Variability within retinal waves

i) Across species

Godfrey et al. 2007

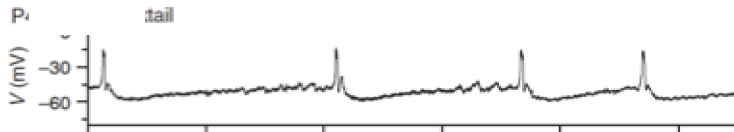


ii) Development

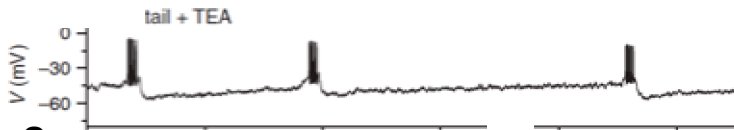
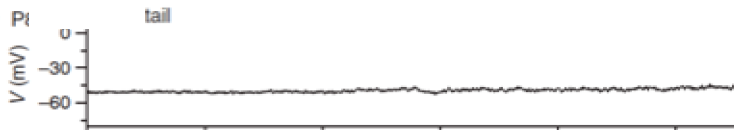


iii) Pharmacology

P4



P8

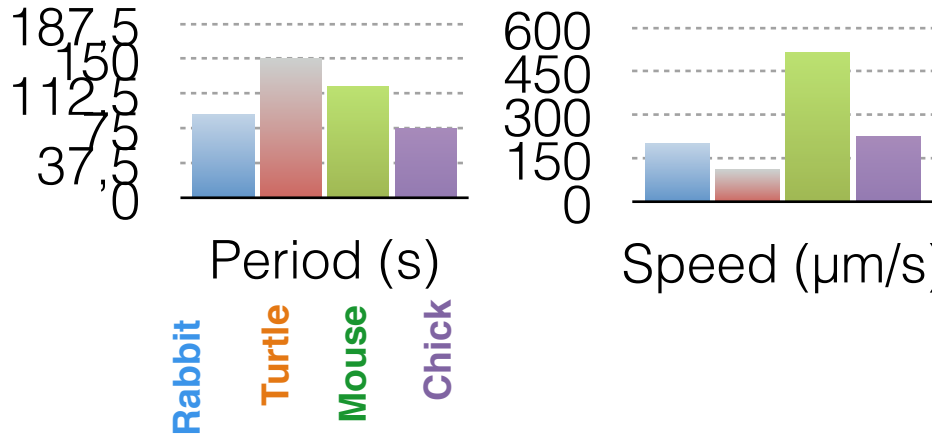


0 Time (sec) 50

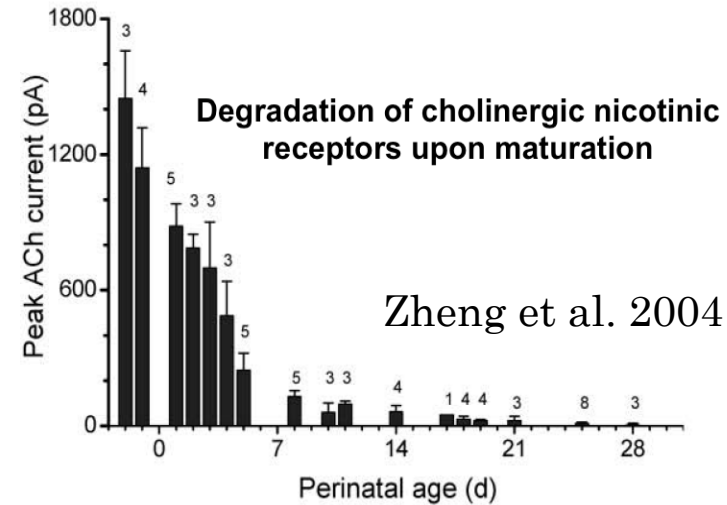
Variability within retinal waves

i) Across species

Godfrey et al. 2007

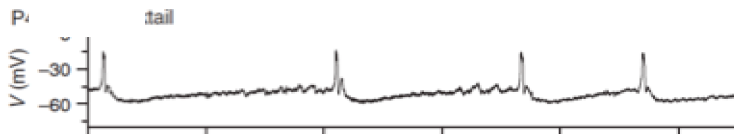


ii) Development

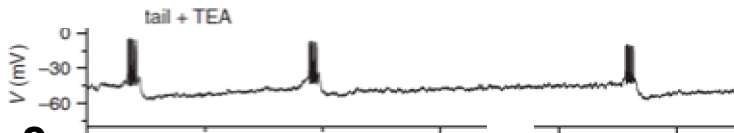
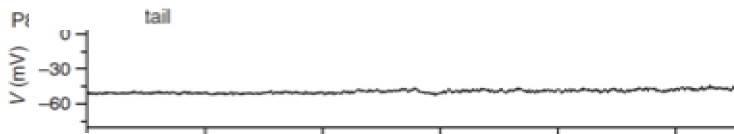


iii) Pharmacology

P4

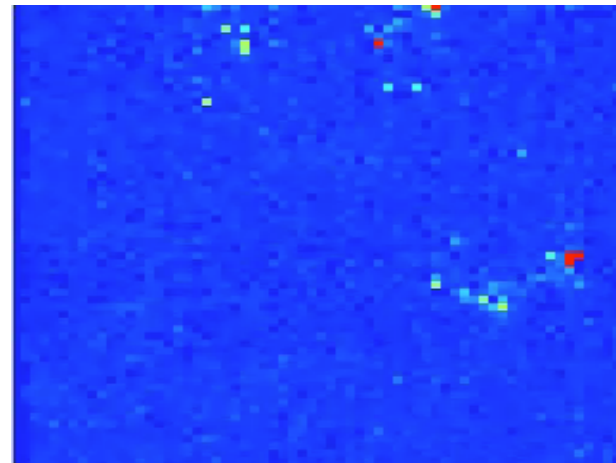


P8



0 Time (sec) 50

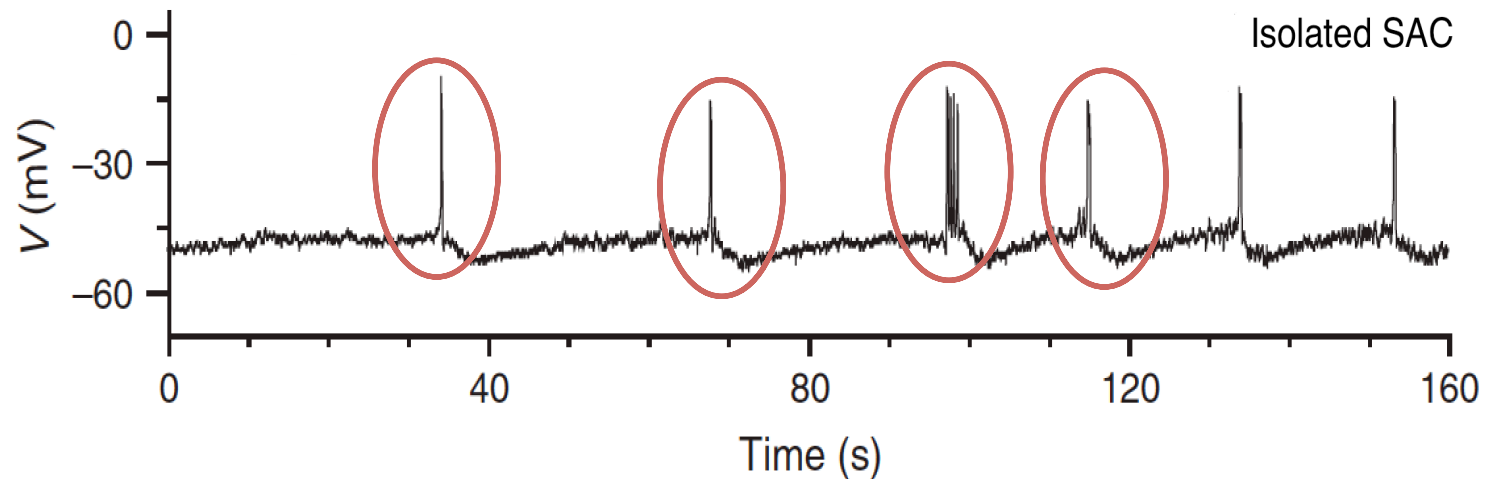
iv) Spatial Variability



Experiments for the emergence of retinal waves

Retinal waves require three components:

i) Spontaneous bursting activity

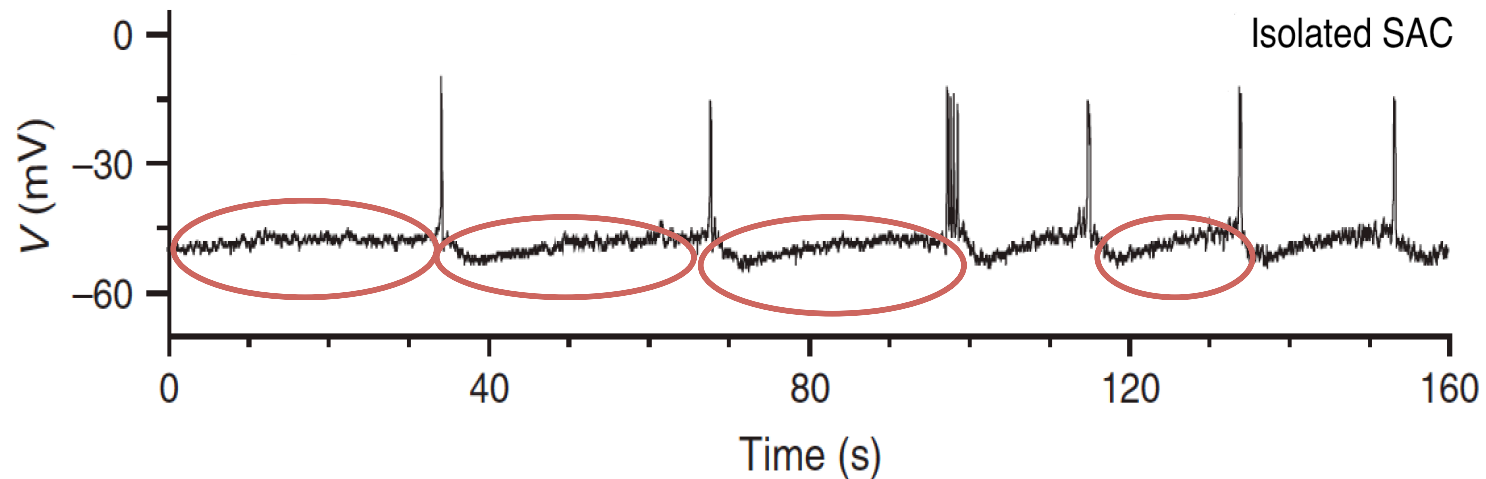


Experiment for isolated neurons,
Zheng et al., 2006, Nature

Experiments for the emergence of retinal waves

Retinal waves require three components:

- i) Spontaneous bursting activity
- ii) Refractory mechanism (slow After HyperPolarisation- sAHP)

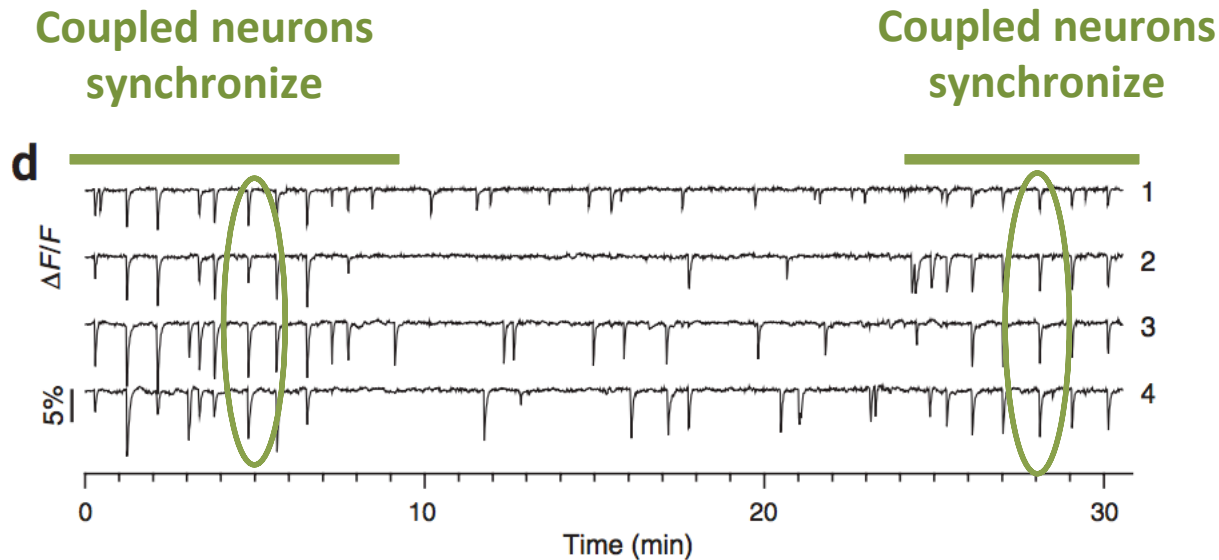


Experiment for isolated neurons,
Zheng et al., 2006, Nature

Experiments for the emergence of retinal waves

Retinal waves require three components:

- i) Spontaneous bursting activity
- ii) Refractory mechanism (slow After HyperPolarisation- sAHP)
- iii) Coupling (through Acetylcholine neurotransmitter)

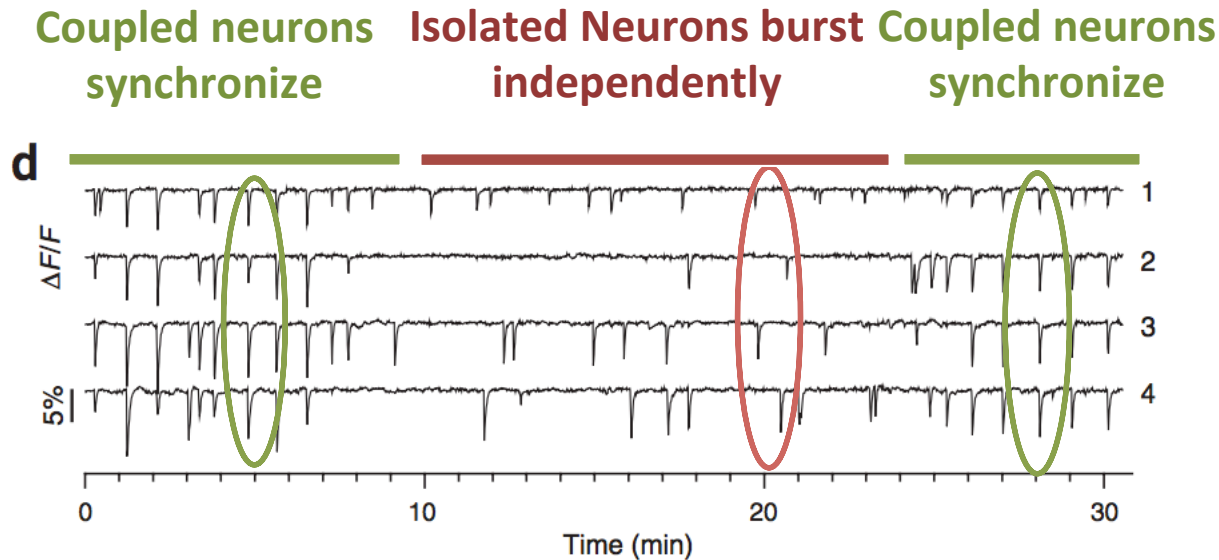


Experiment for coupled and isolated neurons, Zheng et al., 2006, Nature

Experiments for the emergence of retinal waves

Retinal waves require three components:

- i) Spontaneous bursting activity
- ii) Refractory mechanism (slow After HyperPolarisation- sAHP)
- iii) Coupling (through Acetylcholine neurotransmitter)



Experiment for coupled and isolated neurons, Zheng et al., 2006, Nature

Why study retinal waves?

Strategy:

To propose a model (i) sufficiently close from biophysics to explain and propose experiments and (ii) sufficiently well posed mathematically to analyse its dynamics upon varying biophysical parameters (development - pharmacology).

- Modelling one cell bursting
- Modeling cells coupling
- Modelling waves generation, propagation and termination.

Why study retinal waves?

Strategy:

*To propose
its*

Multiscale modelling

*and
analyse
t -*

From ionic channel to neuron to retina scale

Non linear dynamics, dynamical systems theory,
statistical physics.

- Modelling cells coupling
- Modelling waves generation, propagation and termination.

Main assumption

There are a few physiological parameters controlling retinal waves dynamics, evolution and variability.

Find these parameters from a mathematical analysis

Bifurcation theory

A network model for stage II retinal waves

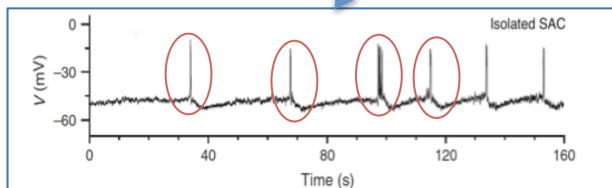
A network model for stage II retinal waves

Membrane potential dynamics

$$C_m \frac{\partial V}{\partial t} = -g_L^M (V - V_L) - g_{Ca}(V)(V - V_{Ca}) - g_K N (V - V_K)$$

$$\tau_N \frac{\partial N}{\partial t} = \Lambda(V)(N_\infty(V) - N)$$

**Morris-Lecar &
Fast K⁺ channels**



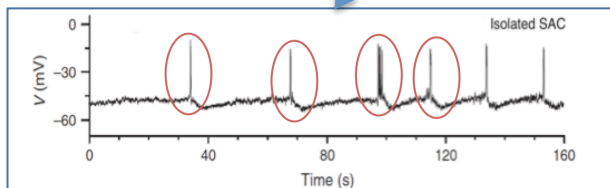
A network model for stage II retinal waves

Membrane potential dynamics

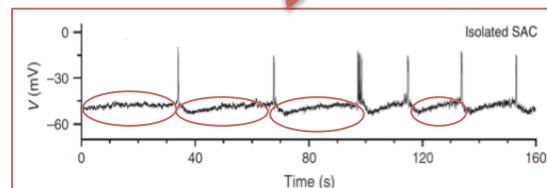
$$C_m \frac{\partial V_i}{\partial t} = -g_L^M (V_i - V_L) - g_{Ca} (V_i) (V_i - V_{Ca}) - g_K N_i (V_i - V_K) - g_{sAHP} R_i^4 (V_i - V_K)$$

$$\tau_N \frac{\partial N}{\partial t} = \Lambda(V) (N_\infty(V) - N)$$

**Morris-Lecar &
Fast K⁺ channels**

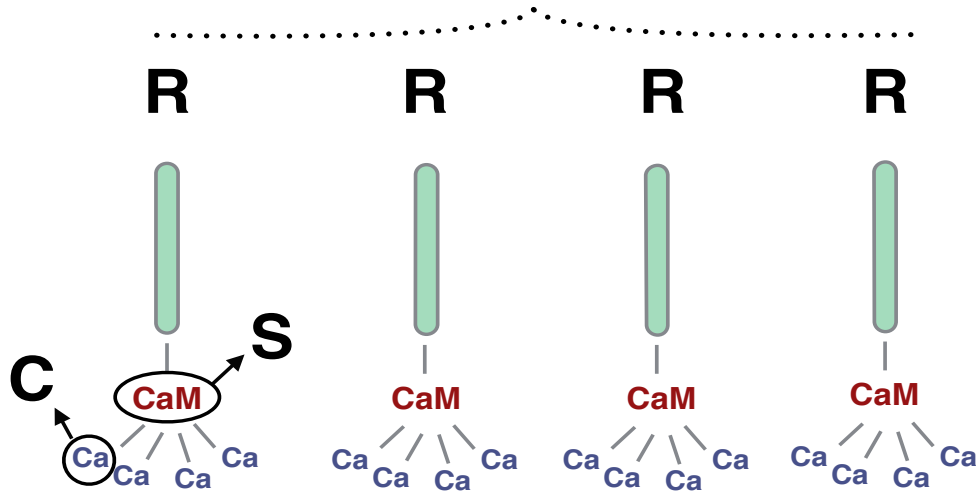


**sAHP current
Refractory
mechanism**



Set of equations for sAHP current

Activated Ca-gated K⁺ channel



Gating mechanism

All 4 subunits R of the channel are bound to activate the channel



Saturated Calmodulin molecule (CaM) bind to each channel subunit



4 Calcium ions bound to each saturated Calmodulin complex S

SACs Network

Model synaptic interactions

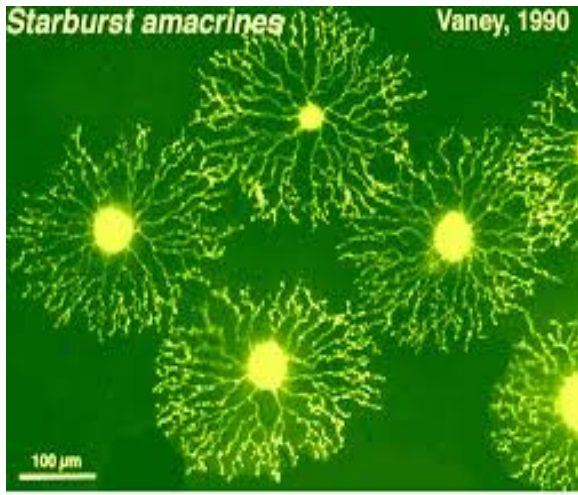
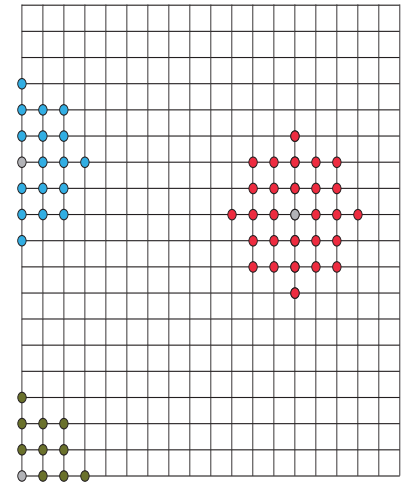
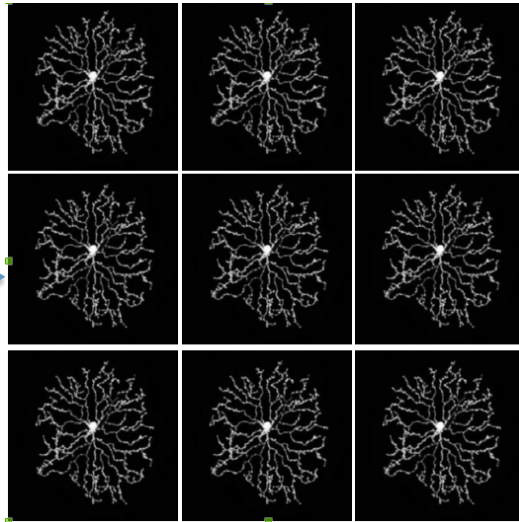


Fig 7. Starburst amacrine cells are stained with lucifer yellow in whole-mount rabbit retina.



SACs realistic
connections

SACs on a lattice

SACs become points on
a lattice

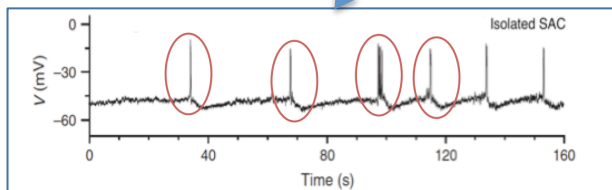
A network model for stage II retinal waves

Membrane potential dynamics

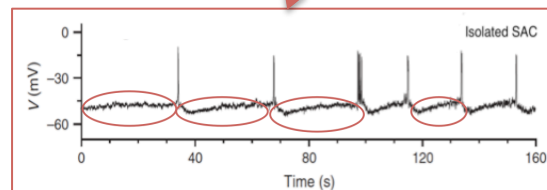
$$C_m \frac{\partial V_i}{\partial t} = -g_L^M (V_i - V_L) - g_{Ca} (V_i) (V_i - V_{Ca}) - g_K N_i (V_i - V_K) - g_{sAHP} R_i^4 (V_i - V_K) - \sum_j g_{j,Ach} (A_j) (V_i - V_{Ach})$$

$$\tau_N \frac{\partial N}{\partial t} = \Lambda(V) (N_\infty(V) - N)$$

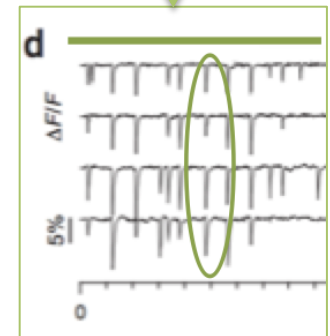
**Morris-Lecar &
Fast K⁺ channels**



**sAHP current
Refractory
mechanism**

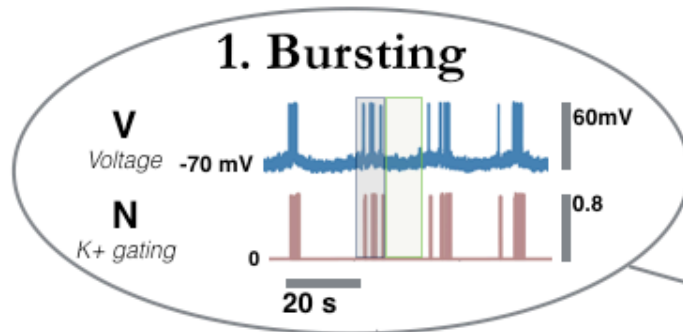


**Ach current
Network
effect**

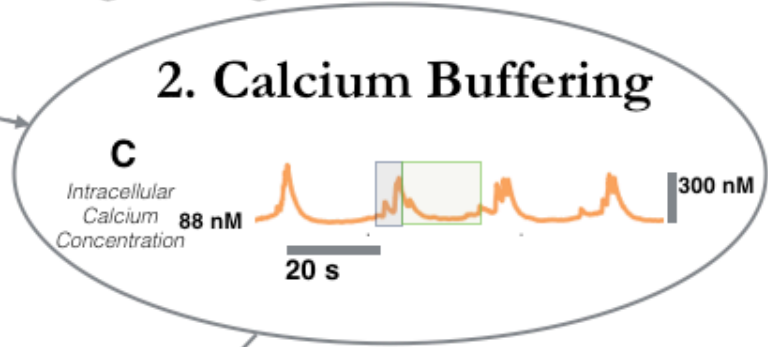


Spontaneous Bursting Mechanism of SACs

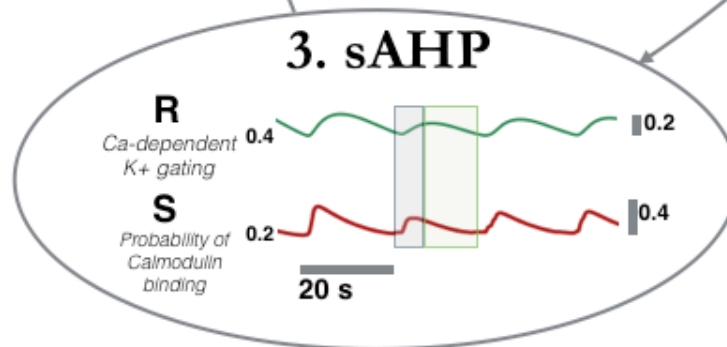
Bursting
 sAHP



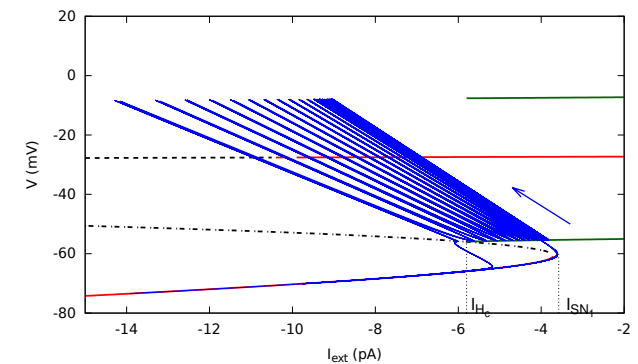
Increase of Calcium load during bursting



Bursting starts again when the Calcium load is low enough



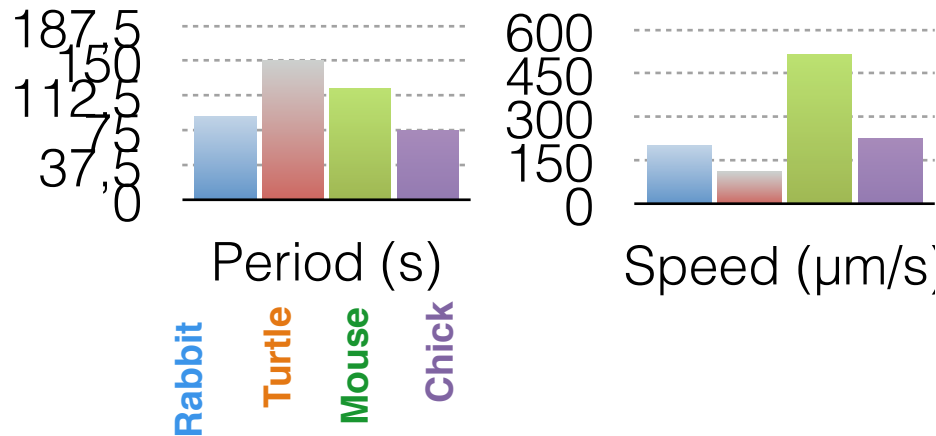
Calcium controls the sAHP phase.
Voltage is low and Calcium starts offloading



Variability within retinal waves

i) Across species

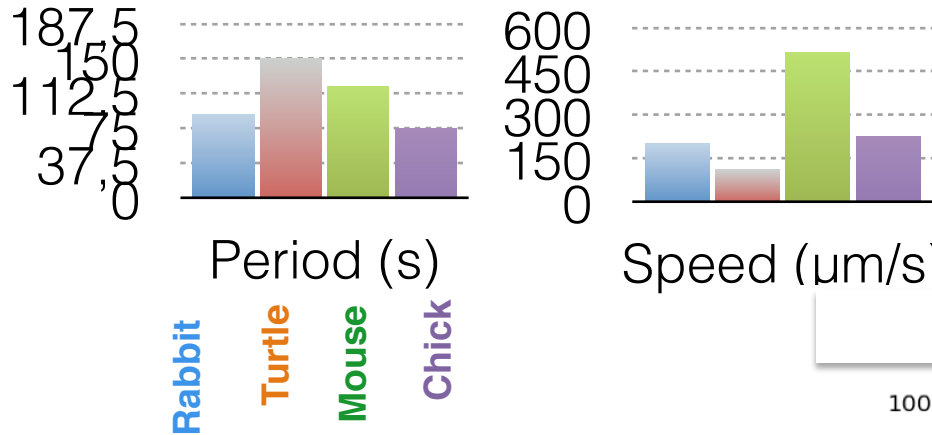
Godfrey et al. 2007



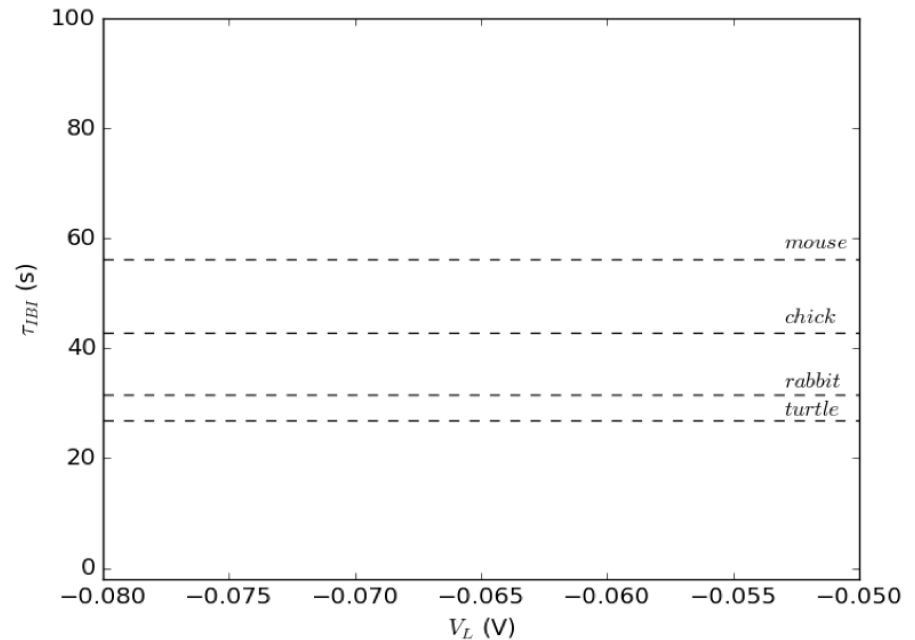
Variability within retinal waves

i) Across species

Godfrey et al. 2007



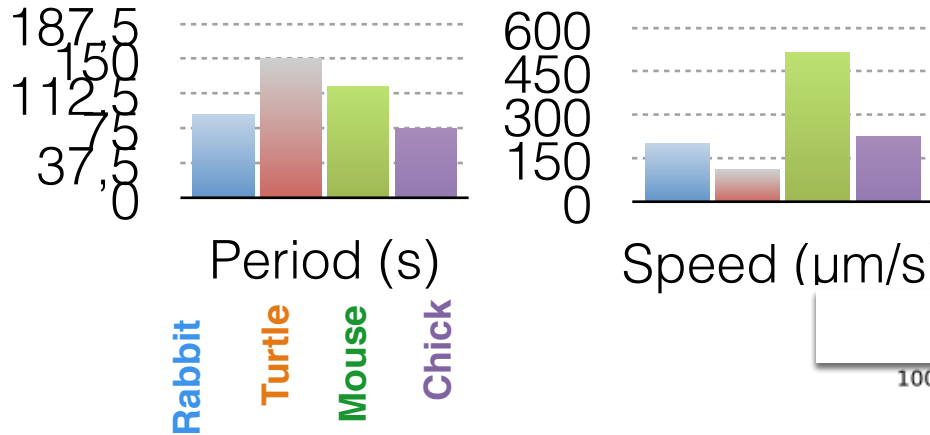
Interburst intervals



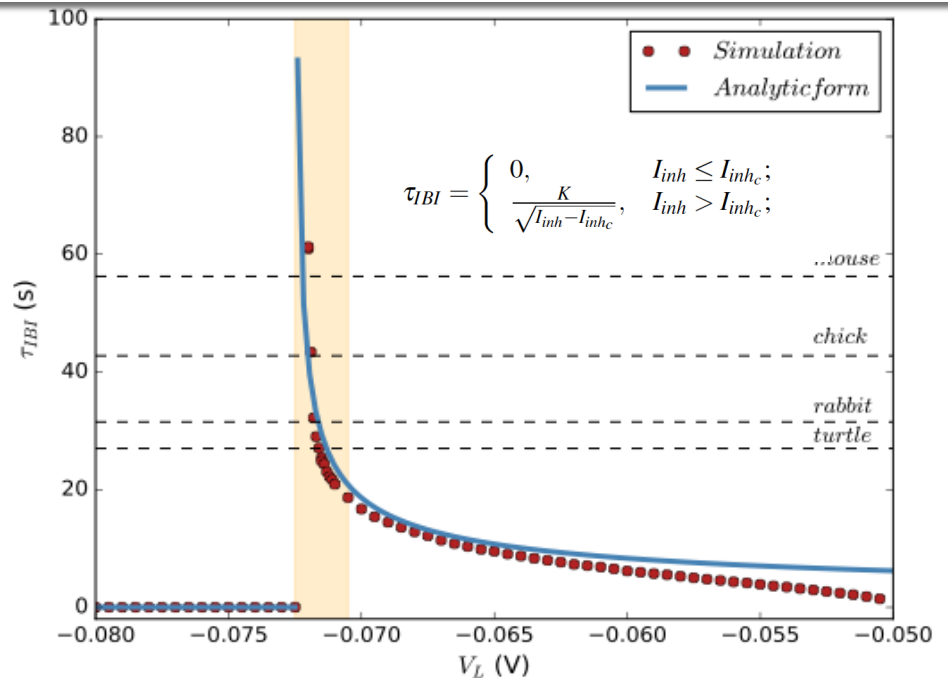
Variability within retinal waves

i) Across species

Godfrey et al. 2007



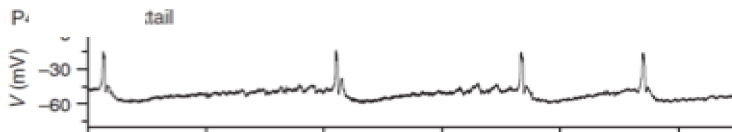
Interburst intervals



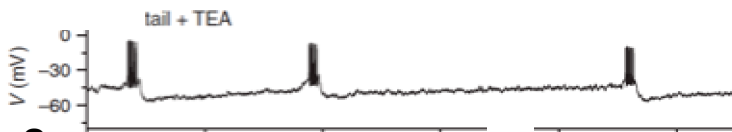
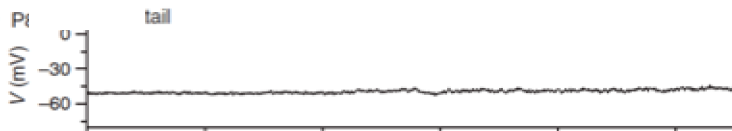
Variability within retinal waves

iii) Pharmacology

P4



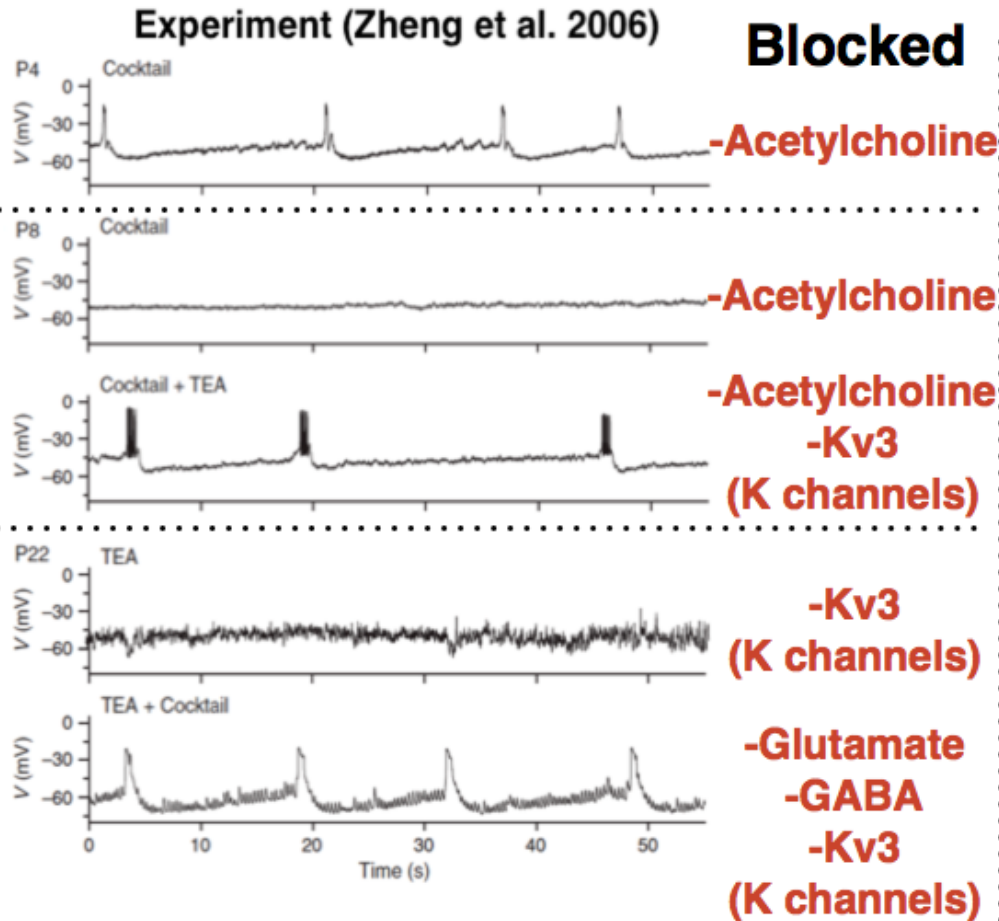
P8



0 Time (sec) 50

How do SACs lose their excitability?

A



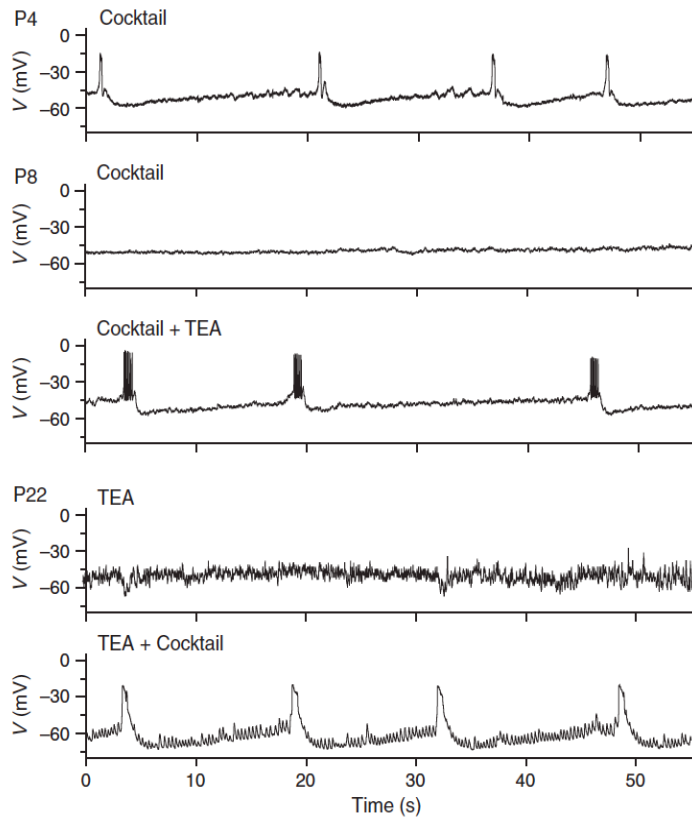
**Potassium channels
(potentially Kv3)
are modelled by 3
parameters
gK, V3, V4**

***TEA blocks Kv3 channels**

Predict the role of Kv3 channels in the loss of SACs excitability

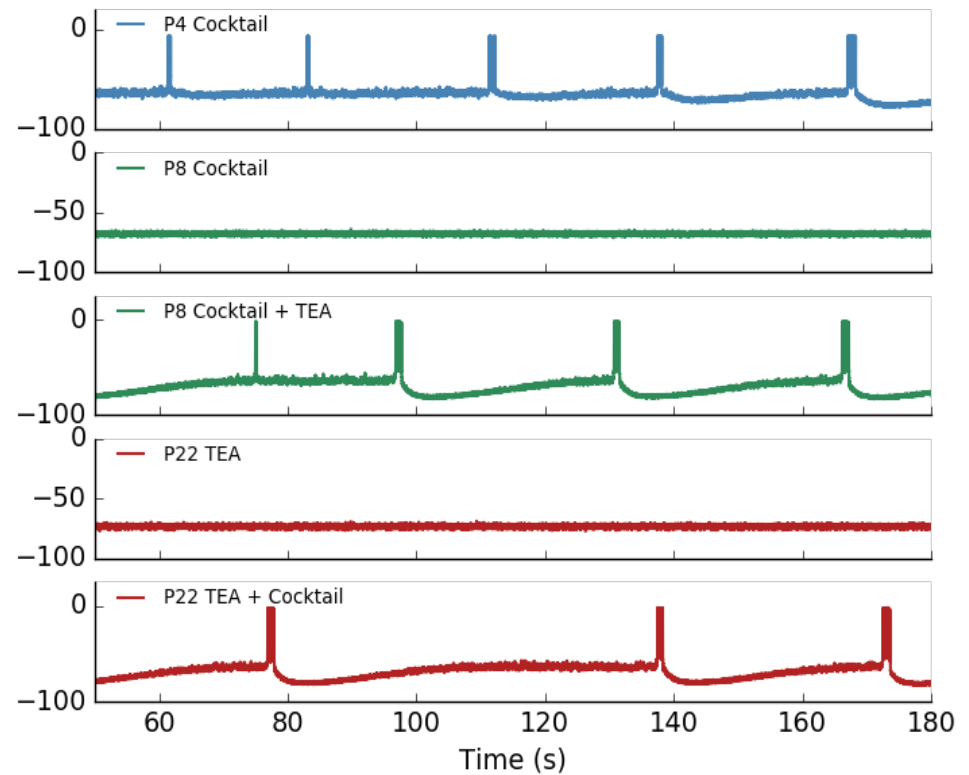
A

Experiment (Zheng et al. 2006)



B

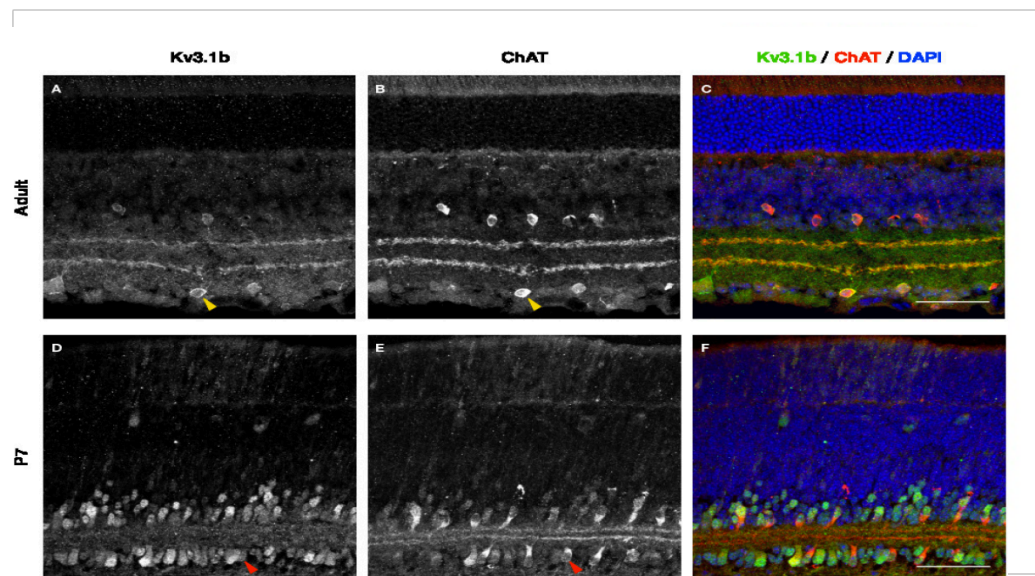
Model



How do SACs lose their excitability?

Hypothesis: The expression of the potassium Kv3 channels increases upon maturation possibly leading to the loss of SACs excitability

Preliminary experimental exploration



(O.Marre-E. Orendorff)

Localization of Kv3.1b in adult and P7 retinas.

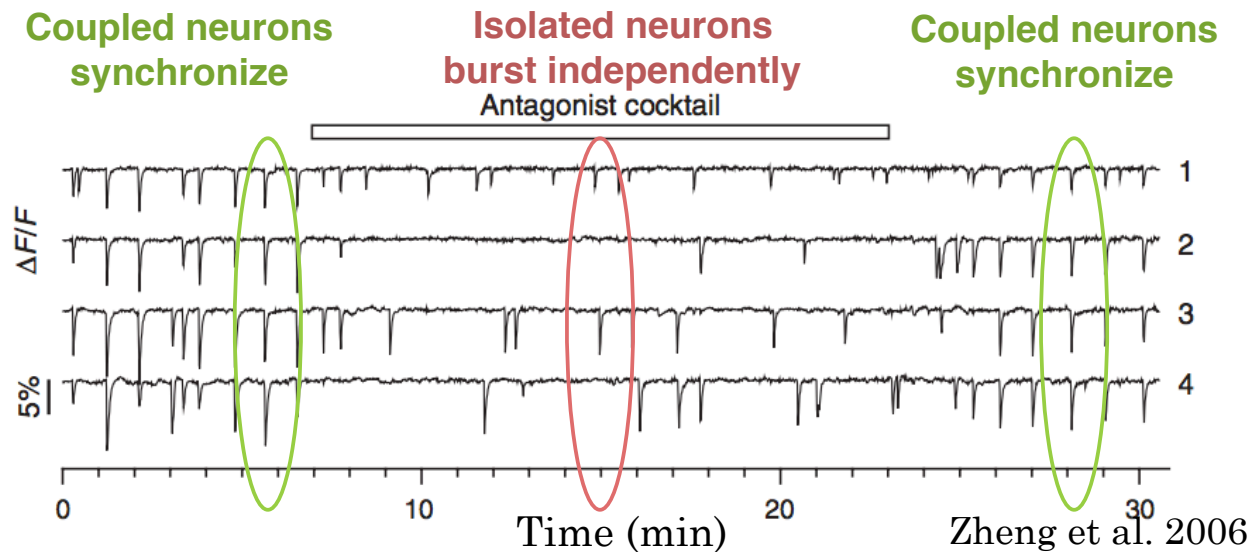
(A-C) Adult retina section showing Kv3.1b (green) and choline acetyltransferase (ChAT, red) reactivity in starburst amacrine cells. Cell nuclei stain (DAPI, blue).

(D-F) P7 retina section showing little colocalization of Kv3.1b with ChAT in starburst amacrine cells.

However, results are preliminary as experiment is not yet conclusive

Cellular mechanisms of stage II retinal waves

C. Synchrony through Acetylcholine

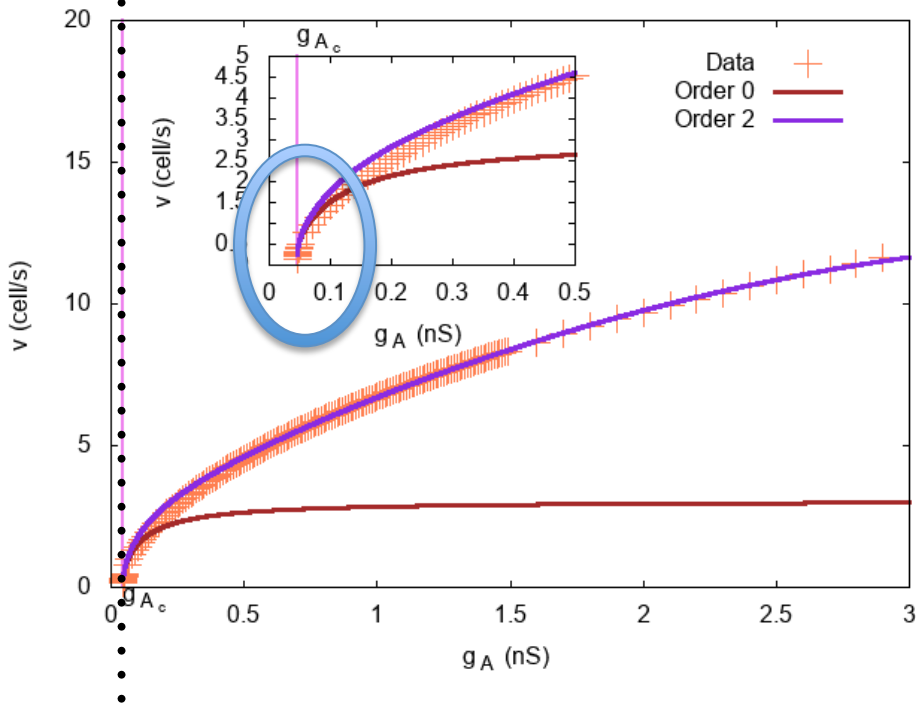


**Mutual excitatory
connections between SACs through
Acetylcholine**

Waves speed

$$v_0 = \frac{1}{t_C - \frac{1}{\mu} \log \left(1 + \frac{1}{g_A} \frac{2\mu\sqrt{\gamma_A}}{d\beta\Omega} \left(\frac{I_{SN} + g_{sAHP} R^4 (V_- - V_K)}{(V_- - V_A)} \right) \right)}$$

Waves speed

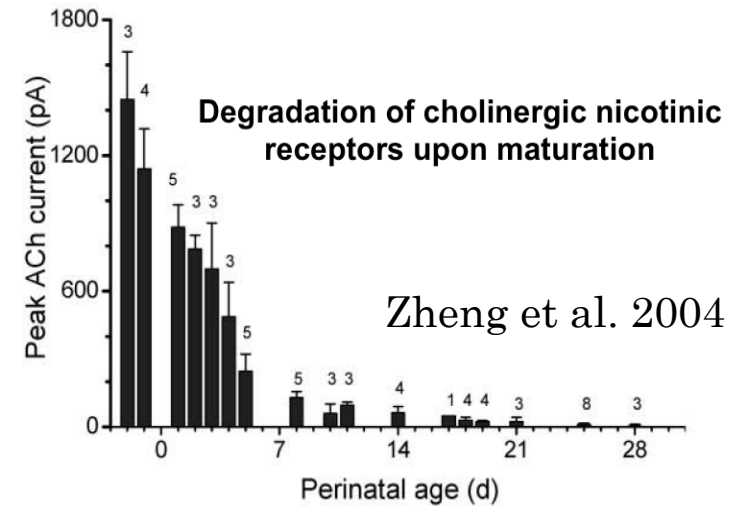


Propagation threshold

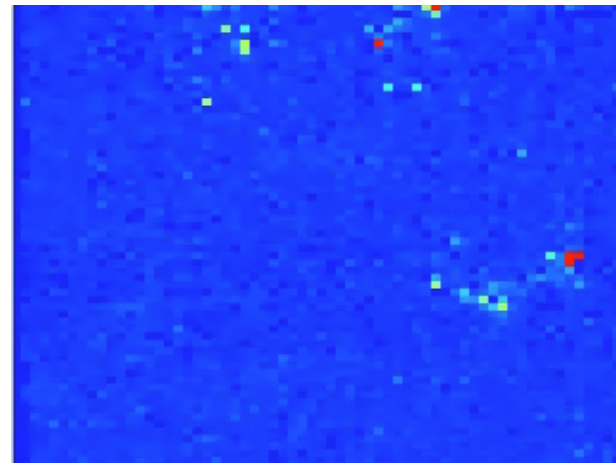
$$g_{A_c} = - \frac{2\mu\sqrt{\gamma_A}}{n_i\beta\Omega} \frac{I_{SN} + g_S R^4 (V_- - V_K)}{V_- - V_A}$$

Variability within retinal waves

ii) Development



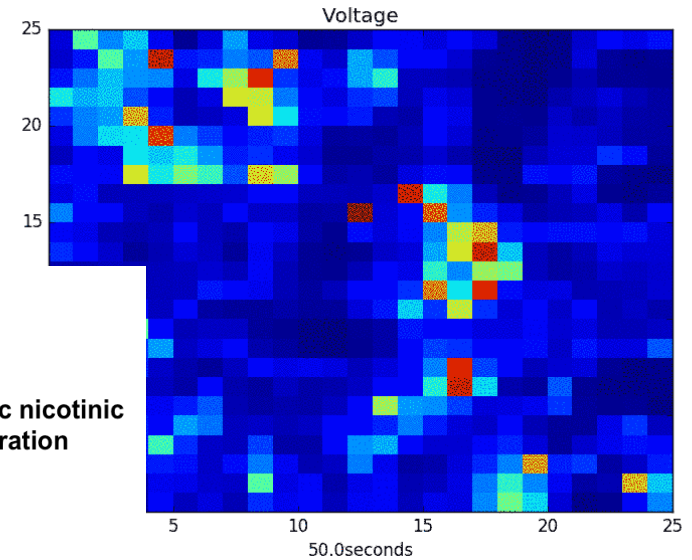
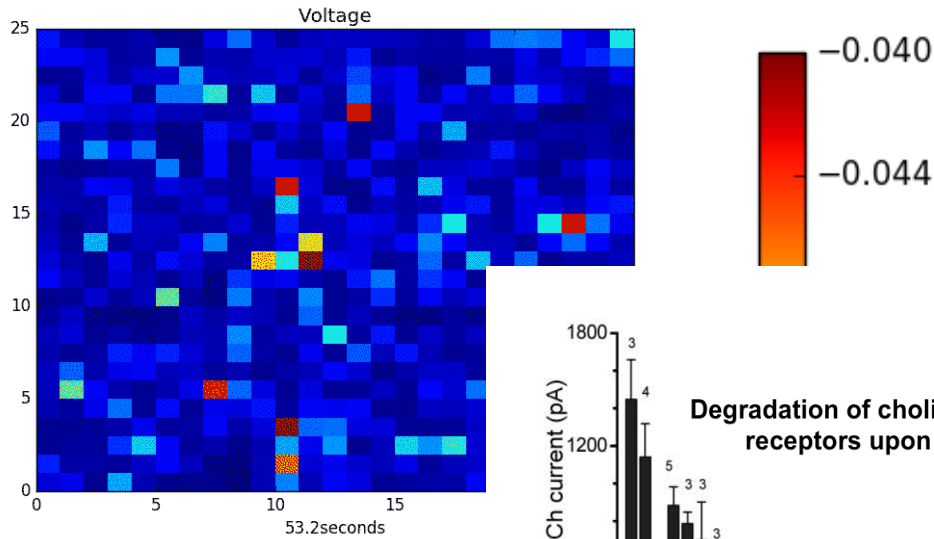
iv) Spatial Variability



Network of SACs : Simulated Voltage

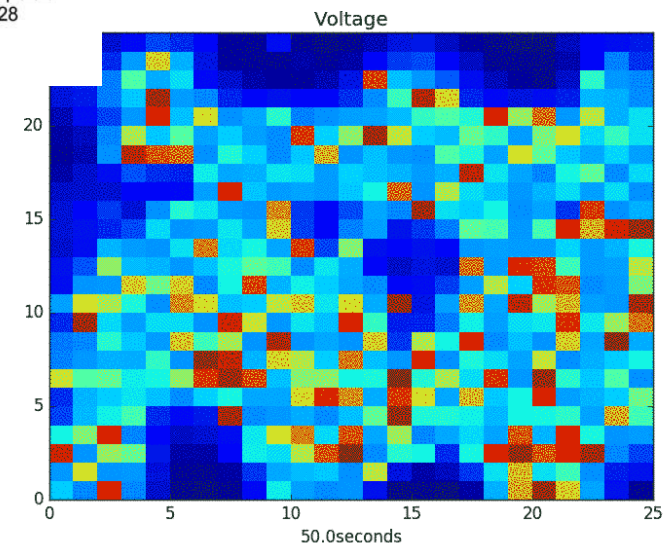
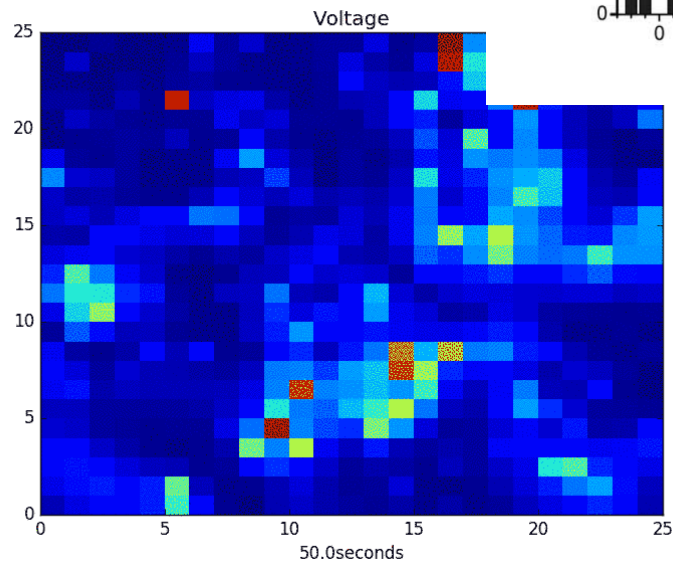
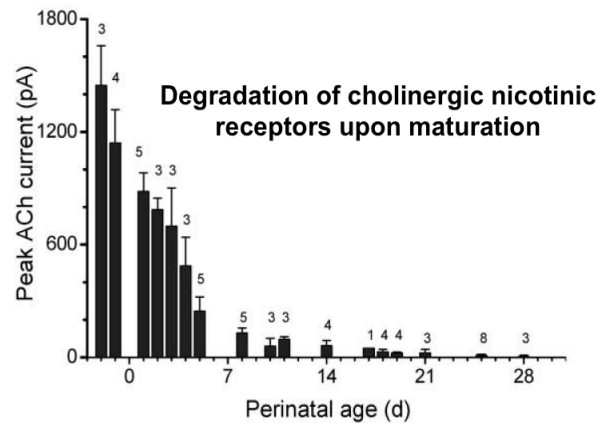
Isolated Neurons

$g_{ACh} = 0.126 \text{ nS}$



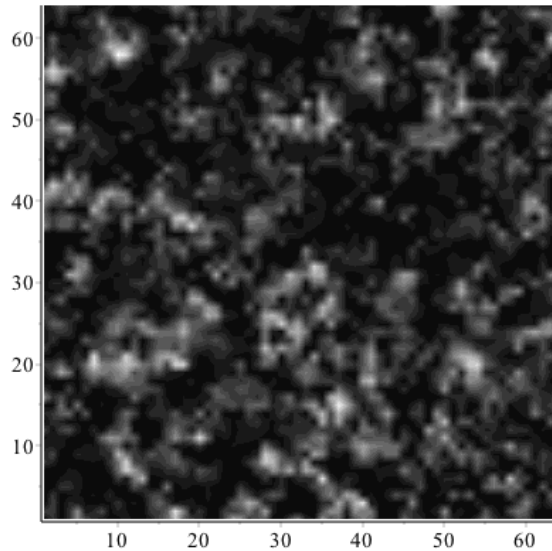
$g_{ACh} = 0.168 \text{ nS}$

$g_{ACh} = 0.21 \text{ nS}$

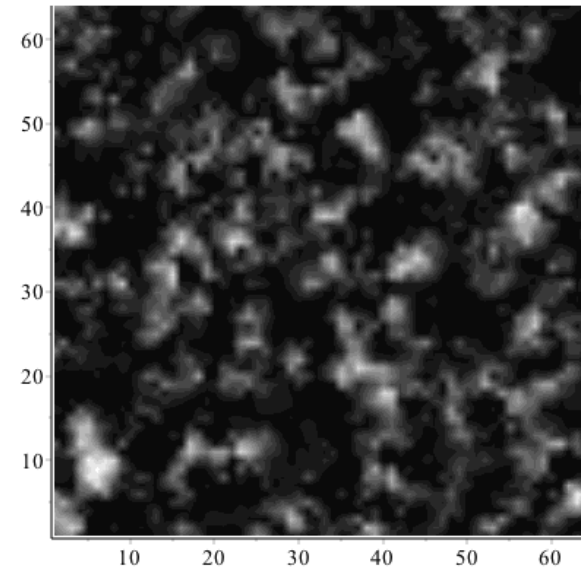


Network of SACs : Simulated Calcium Concentration

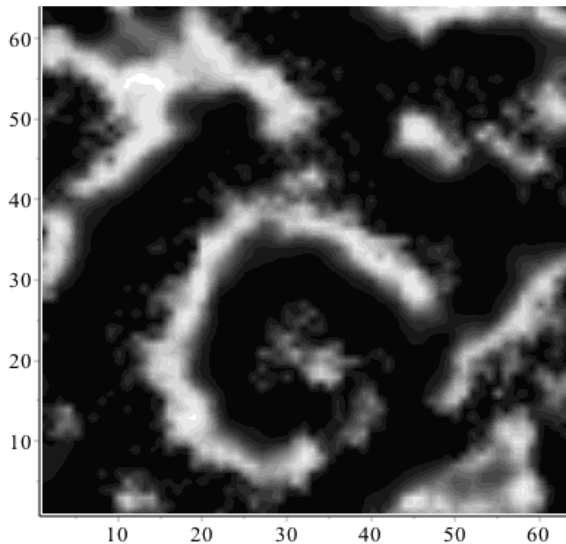
$g_{ach}=0.102$ nS



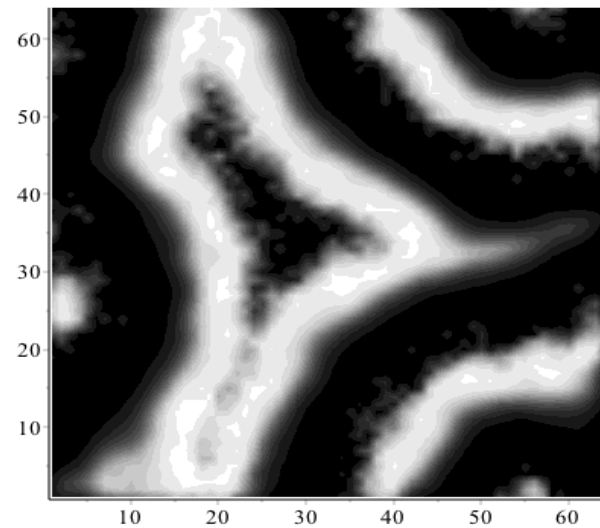
$g_{ach}=0.126$ nS



$g_{ach}=0.168$ nS



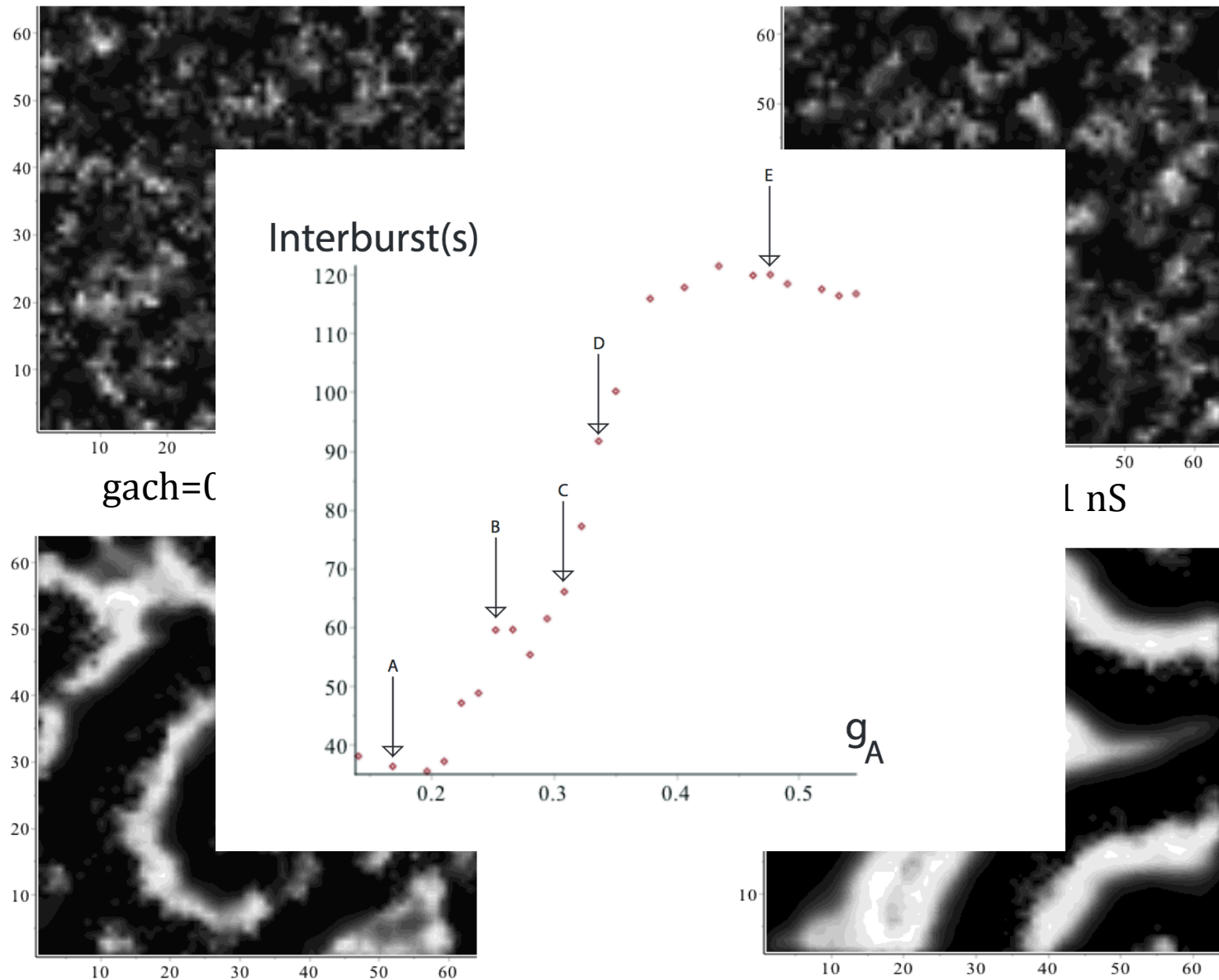
$g_{ach}=0.21$ nS



Network of SACs : Simulated Calcium Concentration

$g_{ACH}=0.102 \text{ nS}$

$g_{ACH}=0.126 \text{ nS}$

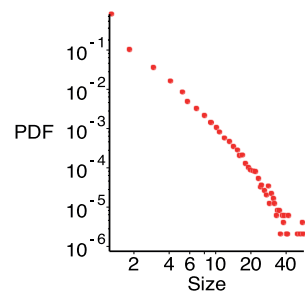
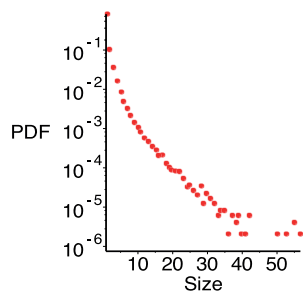


Waves size distribution

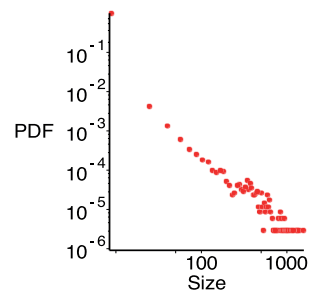
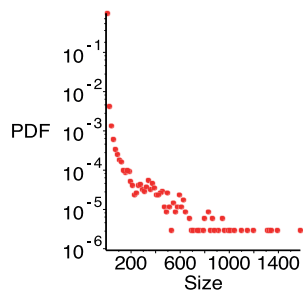
lin-log

log-log

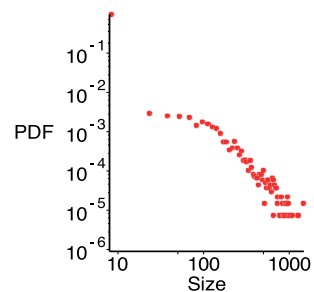
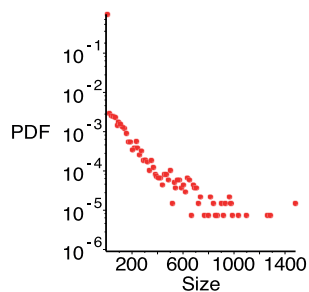
A



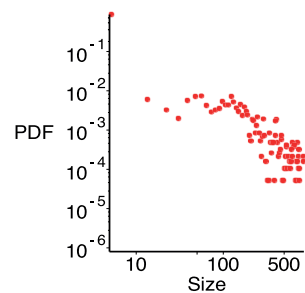
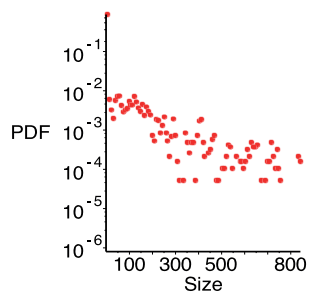
B



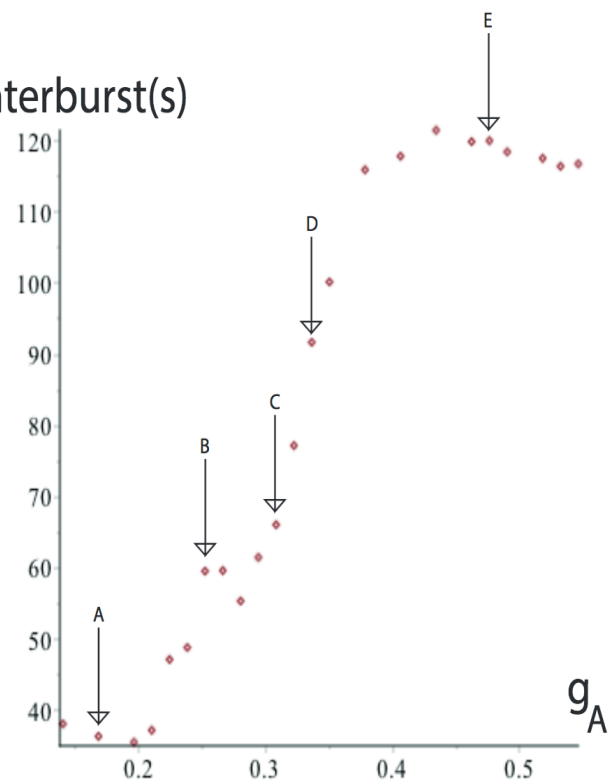
C



E



Interburst(s)

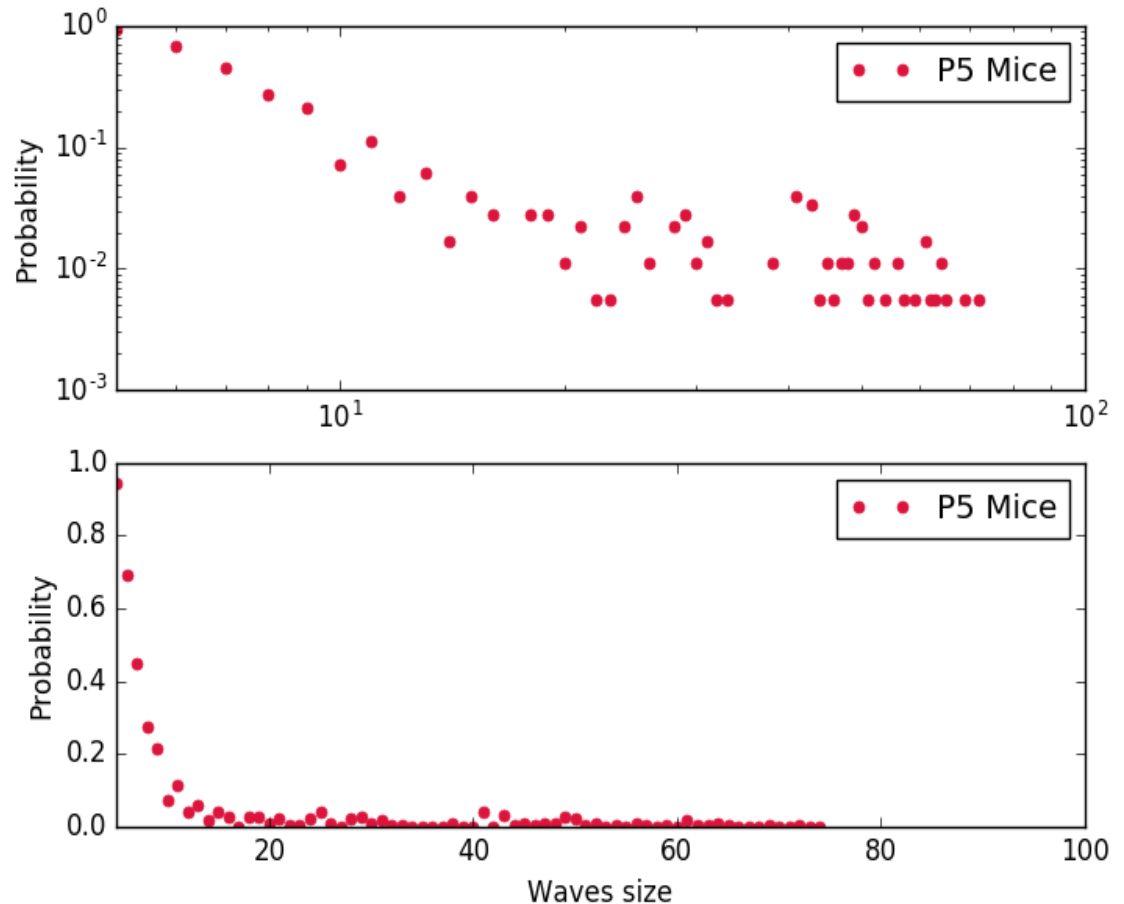


N-neuron model

- There is a **competition** between 2 mechanisms:
 - Period variability which tends to desynchronise
 - Acetylcholine which tends to synchronise
- There is an intermediate regime of coupling, where **variability** is maximum
- Therefore there is a wide repertoire of patterns
 - **Weak coupling** leads to small **localised activity**
 - **Moderate coupling** leads to **propagating patterns**
 - **Strong coupling** leads to **complete synchrony** of neurons

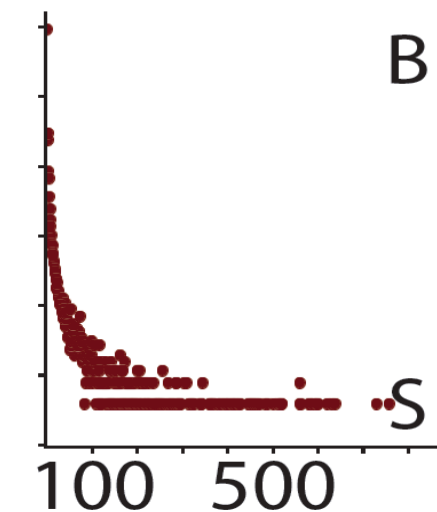
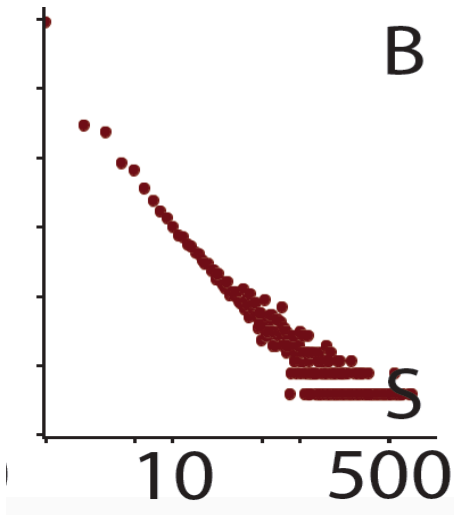
Experimentally varying Ach conductance (Data D. Karvouniari + Institut de la Vision)

Experiment

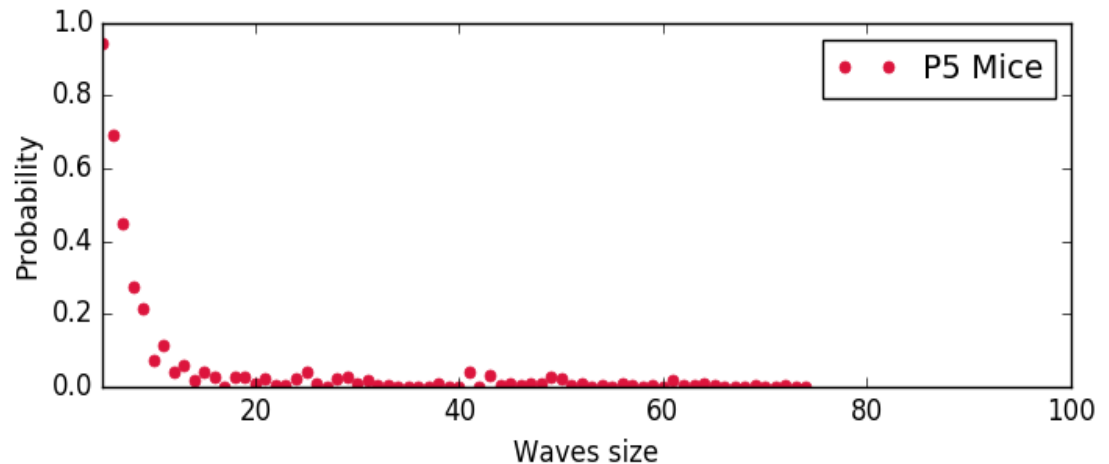
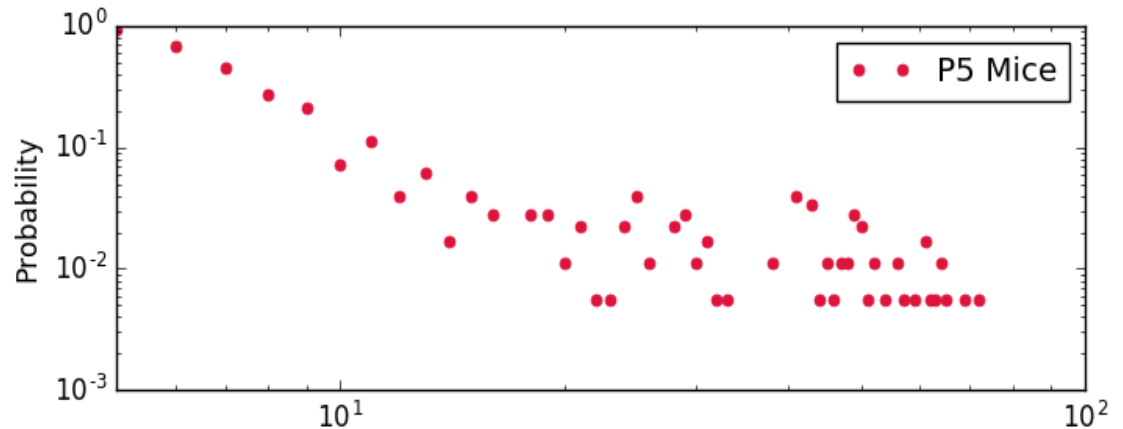


Experimentally varying Ach conductance (Data D. Karvouniari + Institut de la Vision)

Model



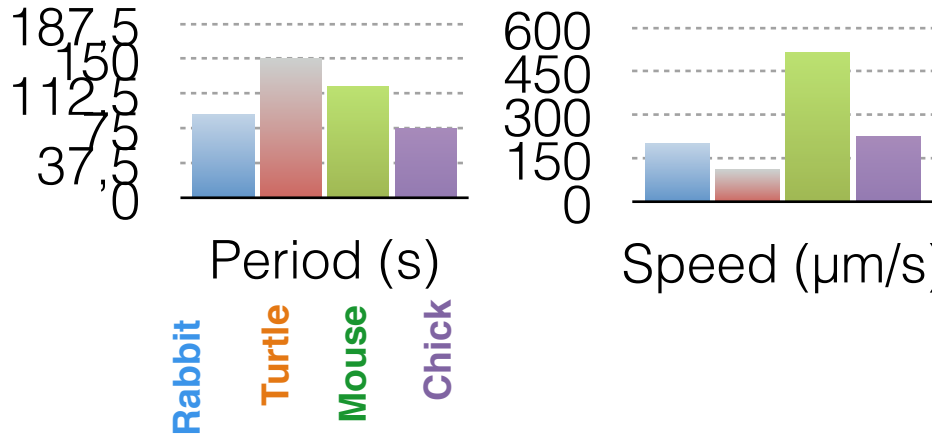
Experiment



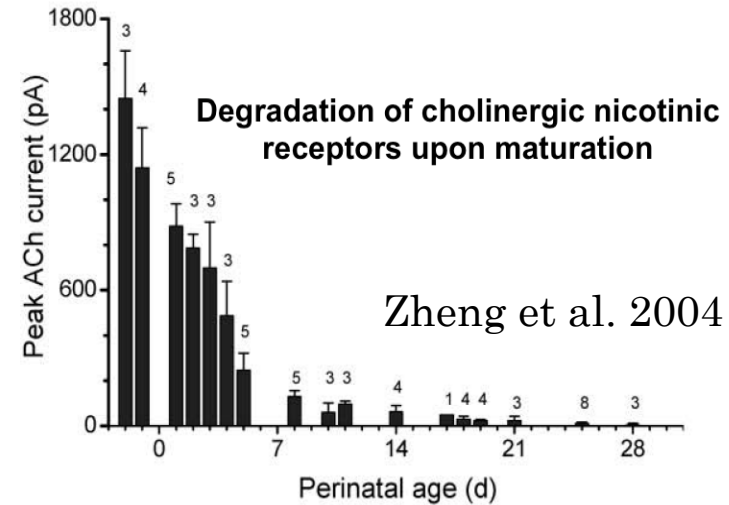
Variability within retinal waves

i) Across species

Godfrey et al. 2007

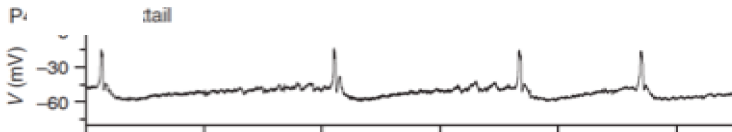


ii) Development

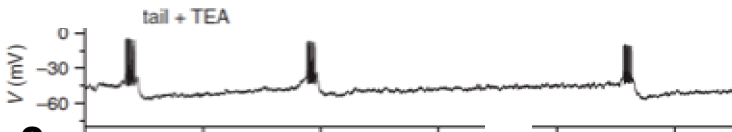
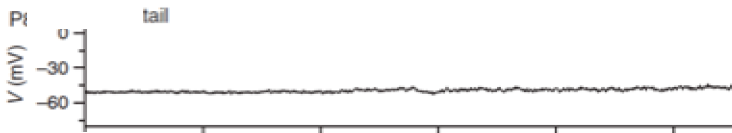


iii) Pharmacology

P4

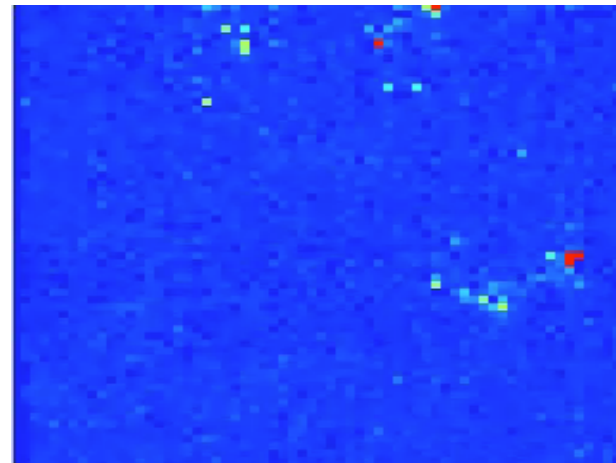


P8



0 Time (sec) 50

iv) Spatial Variability



Conclusion

- **Biophysical model of stage II retinal waves relevant at:**
 - The cells scale (bursting, experimental match and predictions)
 - The network scale (waves propagation)
 - The developmental level (evolution of ionic channels and synapses).
- **Theoretical description via bifurcation theory**
 - Bursting
 - Interburst variability
 - Waves patterns
- **Next step: reactivating waves in adults.**