



HAL
open science

During a short window of *Drosophila* oogenesis, piRNA biogenesis may be boosted and mobilization of transposable elements allowed.

Jeremy Dufourt, Chantal Vaury

► **To cite this version:**

Jeremy Dufourt, Chantal Vaury. During a short window of *Drosophila* oogenesis, piRNA biogenesis may be boosted and mobilization of transposable elements allowed.. *Frontiers in Genetics*, 2014, 5, pp.385. <10.3389/fgene.2014.00385>. <hal-01923996>

HAL Id: hal-01923996

<https://hal.science/hal-01923996v1>

Submitted on 10 Oct 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



HAL Authorization



During a short window of *Drosophila* oogenesis, piRNA biogenesis may be boosted and mobilization of transposable elements allowed

Jérémy Dufourt^{1,2†} and Chantal Vaury^{1,2*}

¹ Centre National de la Recherche Scientifique, Unité Mixte de Recherche 6293, Institut National en Santé et Recherche Médicale, Unité Mixte de Recherche 1103, Clermont Ferrand, France

² Laboratoire GRéD, Clermont Université, Université d'Auvergne, Clermont Ferrand, France

*Correspondence: chantal.vaury@udamail.fr

[†] Present address: Jérémy Dufourt, mRNA Regulation and Development, Institute of Human Genetics, CNRS UPR1142, Montpellier, France

Edited by:

Nelson Cabej, University of Tirana, Albania

Reviewed by:

Molly Hammell, Cold Spring Harbor Lab, USA

Keywords: transposable elements, piRNA, Piwi, evolution, spatio-temporal regulation

INTRODUCTION

In *Drosophila*, PIWI-interacting RNAs (piRNAs) form a distinct class of small 24–30 nt single-stranded non-coding RNAs that are associated with the PIWI proteins Piwi, Aubergine (Aub), and Argonaute 3 (AGO3), which are expressed in the germline (Brennecke et al., 2007). Genetic studies have shown that the PIWI/piRNA ribonucleoprotein (RNP) complex has an evolutionary conserved role in the silencing of transposable elements (TEs) in the germline of animals ranging from sponges to humans (Cerutti and Casas-Mollano, 2006; Aravin et al., 2007; Grimson et al., 2008; Luteijn and Ketting, 2013).

piRNAs arise from specific genomic clusters called piRNA clusters that are mainly localized in pericentromeric regions. These clusters are transcribed uni/bi-directionally as long single-stranded RNA precursors by the RNA polII (Sienski et al., 2012; Goriaux et al., 2014; Mohn et al., 2014). In the germline, these precursors are processed into piRNAs within specific cytoplasmic regions called the nuage prior to being loaded onto Piwi and Aub proteins. The piRNAs generated from this initial transcription of piRNA clusters are called primary piRNAs. These RNP complexes can repress expression of TEs at the transcriptional (TGS) (Sienski et al., 2012; Rozhkov et al., 2013) and/or post-transcriptional (PTGS) level (Chambeyron et al., 2008; Haase et al., 2010; Dufourt et al., 2011; Rozhkov et al., 2013).

TGS control results from a compact repressive chromatin structure characterized by an enrichment of histone H3, which is tri-methylated on Lysine 9, the so-called H3K9me3 repressive mark. It has been suggested that Piwi loaded onto piRNAs identifies TE targets by a homology-dependent base-recognition mechanism. It recruits SU(Var)3-9, the major heterochromatin-specific HMTase in *Drosophila* (Huang et al., 2013), which tri-methylates histone H3 on Lysine 9 (H3K9me3) of the target. Several studies have reported that this heterochromatic structure is labile (Dufourt et al., 2011; Sienski et al., 2012), indicating that TEs can occasionally bypass repression (Haase et al., 2010; Rozhkov et al., 2013; Klenov et al., 2014).

piRNAs also serve as guides for post-transcriptional repression (PTGS) of TEs. PTGS involves two other PIWI proteins, Aub, and Ago3. The Aub/piRNA RNP complex targets cytoplasmic mRNAs encoded from TEs and cleaves the mRNA by virtue of its slicer activity. This gives rise to new piRNAs called secondary piRNAs, which are sense with respect to the canonical transposon mRNAs. These secondary piRNAs are loaded onto Ago3. The Ago3/piRNA RNP complex targets precursor transcripts arising from piRNA clusters. Subsequent cleavage prompts the biogenesis of new piRNAs bound to Aub whose sequence is identical to that of the initiator piRNA. This loop of amplification, called the ping-pong cycle, amplifies

silencing-competent piRNAs (Brennecke et al., 2007).

Acting together, these two mechanisms, TGS and PTGS, tightly repress TE transposition in the germ line and thus act as guardians of genome integrity. Interestingly, since the pool of piRNAs produced in the oocyte is deposited in the embryo, TE repression is transmitted from the mother to her progeny, who are immediately protected against TE mobilization. (Ronsseray et al., 1993; Malone et al., 2009; Handler et al., 2011; de Vanssay et al., 2012; Grentzinger et al., 2012; Le Thomas et al., 2014).

Since TE transcription is blocked by TGS in the germ line, the current model fails to define when the ping-pong cycle is active and if a rebootstrap has to occur to increase the stock at each generation. It also fails to explain why, despite this tight repression, high genetic variability, mainly due to TE insertions, is observed in each genome sequenced to date, evidence suggesting that TEs have a means of overcoming repression in the germ line and transpose.

In the germline, at the anterior side of *Drosophila* ovaries, the germline stem cell (GSC) divides to give a daughter cell called the cystoblast. This latter undergoes four cycles of mitotic division to form cysts of successively 2, 4, 8, and 16 germ cells. The mature egg chamber consisting of 16 germ cells including the oocyte and 15 nurse cells then leaves the germline. Recently, we observed that transient

downregulation of Piwi occurs during early oogenesis when cysts divide (Dufourt et al., 2014). In this region called the Piwiless pocket (Pilp), the absence of Piwi is correlated with a decrease in germline repression exerted on sensor transgenes used as read-out of repression of two TEs, the LTR retrotransposon Idefix and the P-transposon. We propose that this short window of oogenesis could correspond to the moment at which mRNAs are synthesized from TEs, as a consequence of which the ping-pong cycle is boosted, resulting in an increasing pool of piRNAs, and TE replication cycles are allowed.

ENHANCING THE POOL OF GERMLINE piRNAs

It has been shown that ping-pong processing ensures that a large pool of piRNAs will be produced during oogenesis and transmitted to protect the embryo as soon as it starts developing. Because

of the TGS exerted on TEs in the germ line, the moment at which this increase occurs remains unknown. It could be speculated that the pool of piRNAs is increased within the primordial germ cells (PGC), where maternally-deposited piRNAs are present and transcription of piRNA clusters is active (Le Thomas et al., 2014). However, the repressive heterochromatin structure embedding TEs would be expected to prevent the production of TE mRNAs and thereafter any new round of secondary piRNA synthesis from TE mRNAs.

We propose that, in the Pilp, the decrease in Piwi diminishes the TGS exerted on TEs and leads to their transcription. The resulting mRNAs serve as targets for the ping-pong cycle, which is thus kicked-up and the piRNA pool that will be ultimately transmitted to the progeny is amplified. This phase is transient and restricted to the dividing cysts

because Piwi expression is restored to normal at the end of the mitotic divisions (Figure 1).

Interestingly, we recently observed that Aub, a major component of PTGS, is required for TE silencing during the germinal stages of oogenesis whereas its depletion after this stage has no impact on TE silencing (Dufourt et al., 2014). In contrast, Piwi involved in TGS is required throughout oogenesis. Together with the existence of the Pilp, where TGS is weakened, these data suggest that the PTGS exerted on TEs mostly takes place in the small group of cells where TE mRNAs are produced.

Altogether, these results designate dividing cysts and more specifically the Pilp as a window of the germ line development during which mRNAs encoded by TEs may be produced, the ping-pong cycle boosted and the pool of *de novo* piRNAs that will be inherited amplified.

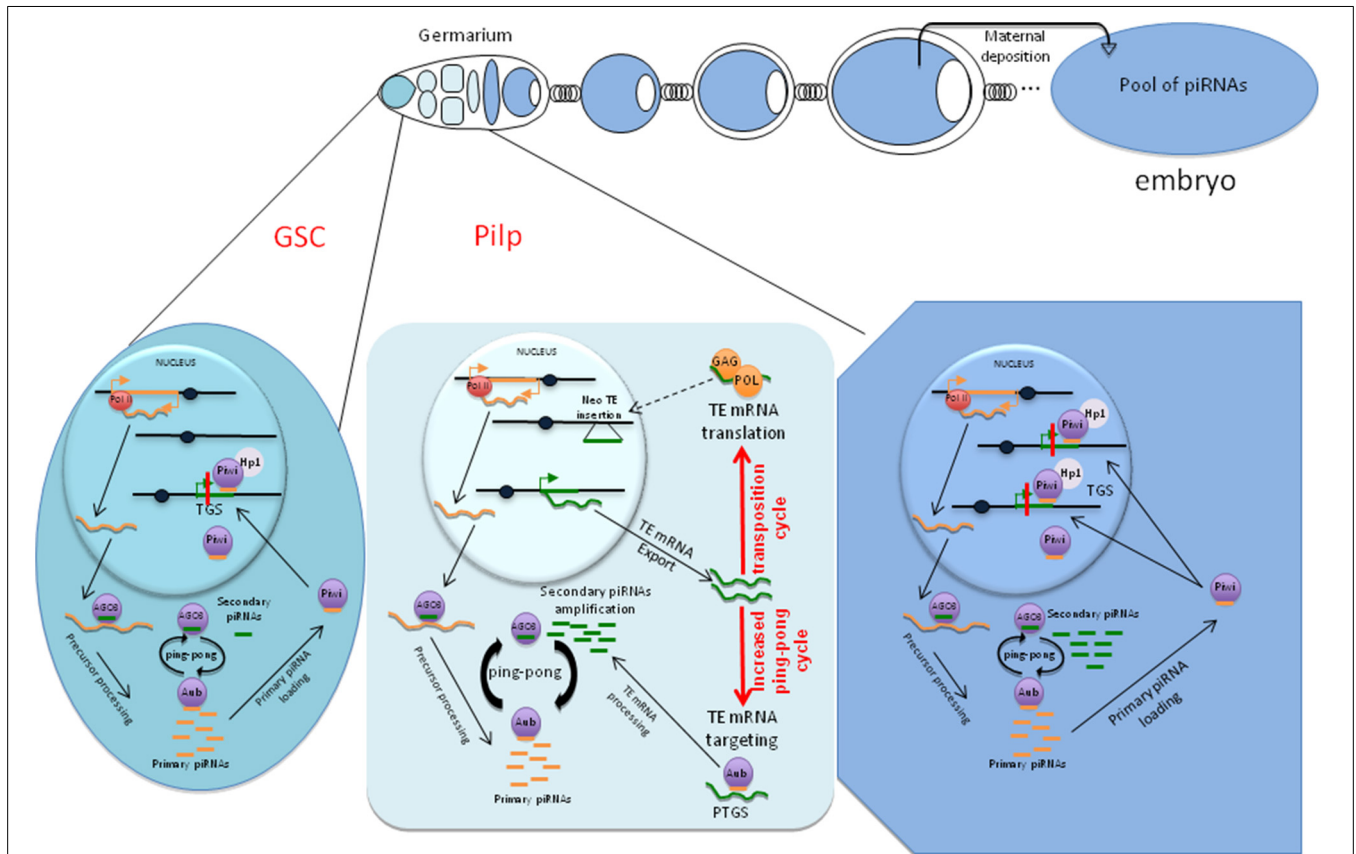


FIGURE 1 | A schematic representation of *Drosophila melanogaster* egg chambers. In the germarium (left part of the upper panel), the Pilp is shown as light blue cells. Transcriptional gene silencing (TGS) and post-transcriptional gene silencing (PTGS) targeting TEs are presented below. In

the Pilp (mid cell, lower panel), the decrease in Piwi allows TE transcription, which has two consequences on the TE/host relationship (red arrows): (1) an increased pool of piRNAs produced by the ping-pong mechanism and (2) increased transposition cycles leading to neo-TE insertions.

ALLOWING TEs TO TRANPOSE IN THE GERM LINE

Although they represent a constant threat for genome stability, TEs have efficiently colonized all the eukaryotic genomes and are considered as major tools for genome evolution and plasticity. This implies that they find a way to bypass host defense mechanisms and mobilize in the germ cells thereby ensuring their propagation to the next generation.

We believe that the best moment when TEs can escape piRNA silencing and insert the genome is when cysts divide, thus within the Pilp. Three lines of evidence support this assumption. First, loss of control of TEs in the Pilp will not affect the potential of the stem cell to continuously produce new viable germline cysts. Mobilization in the GSC would guarantee new insertions to the whole progeny but could also create severe genome damage that might lead to loss of GSC and sterility or lethal effects on descendants. Thus, protecting the stem cell from which all the future germ cells will derive appears to be essential for the species. Second, in the Pilp, the oocyte is not yet in a condensed state that could prevent TE activity. The oocyte nucleus will be condensed and blocked in meiosis after the cystoblast has completed the four rounds of mitotic division to create a cyst. Third, TE silencing is weakened owing to a decrease in Piwi. Transcription is then allowed and replication cycles may start from the pool of synthesized mRNAs.

Because of its property to display a weakened RNA silencing, the Pilp may thus ensure the constant and controlled permissiveness for *de novo* TE genomic integrations (Figure 1) and also sporadic bursts of TE transpositions, as occasionally mentioned in the literature (Biemont and Vieira, 2006). It is noteworthy that TE activation has been reported in the germline of several species (Zamudio and Bourc'his, 2010). In *Arabidopsis*, the maternal endosperm genome is hypomethylated, resulting in transient transposon activation (Hsieh et al., 2009). In pollen from *Arabidopsis*, TEs are also reactivated and transpose but only in the pollen vegetative nucleus thereby avoiding dramatic events in the sperm cells, which give rise to the progeny (Slotkin et al., 2009). In mice,

genome-wide loss of DNA methylation accompanies the acquisition of pluripotent states in PGC, which opens a window of opportunity for TEs to escape from host restraint (Rougier et al., 1998; Hajkova et al., 2002). L1 transcripts and proteins are found in male germ cells entering meiosis but are repressed in differentiated somatic tissues (Branciforte and Martin, 1994). As a last example, the MT family of LTR retrotransposons, although it represents only <5% of the genome, accounts for 13% of the transcriptome of the mature mouse oocyte (Peaston et al., 2004). In all these examples, TE activation is not only linked to the availability of key transcription factors but also to a relaxation of epigenetic control in the cells.

PERSPECTIVE

The two potential functions of the Pilp, i.e., increasing the pool of piRNAs and allowing TE transposition, are shown in Figure 1.

To have a better understanding of the close relationship between TE, evolution, germline protection and transmission to the progeny, it will be important to show whether the piRNA pool in the Pilp is different from that in the GSC and/or the rest of the ovaries. It would be expected for piRNAs arising from piRNA clusters to be highly abundant in GSC but unable to complete the ping-pong cycle because of TGS repressing TE transcription. In contrast, piRNAs transmitted to the Pilp by the GSC should be able to complete the ping-pong cycle with piRNA partners arising from TE mRNAs. To have a fuller understanding of this process, studies should be made to investigate whether these mechanisms are conserved across species to maintain a harmonious balance between TEs and their host genome.

Several studies have recently shown the involvement of the piRNA pathway in additional functions such as germ line development and sex determination (Rouget et al., 2010) (Kiuchi et al., 2014). We also reported that the downregulation of Aub after the germarium stage leads to sterility whereas, at the same stage, TEs are repressed (Dufourt et al., 2014). A recent study demonstrated that Piwi/Su(Var)3-7 genetic interaction leads to an increase in sterility and embryo defects but, importantly, does not correlate

with TE derepression (Basquin et al., 2014). Hence, it will be important to explore the role of this short window of germ line development not only in TE control but also in total mRNA regulation and germ line development.

ACKNOWLEDGMENTS

We thank E. Brassat and E. Théron for helpful discussions, and S. Chambeyron, S. Ronsseray, N. Sambrani, and G. Sanchez for critical comments on the manuscript. Work in C Vaury's lab is supported by grants from CNRS, INSERM, and the Association Nationale de la Recherche (ANR) (project plasTiSiPi) and Ligue contre le Cancer. The present work was supported by the Fondation ARC pour la recherche sur le cancer (to Jérémy Dufourt).

REFERENCES

- Aravin, A. A., Hannon, G. J., and Brennecke, J. (2007). The Piwi-piRNA pathway provides an adaptive defense in the transposon arms race. *Science* 318, 761–764. doi: 10.1126/science.1146484
- Basquin, D., Spierer, A., Begeot, F., Koryakov, D. E., Todeschini, A. L., Ronsseray, S., et al. (2014). The *Drosophila* Su(var)3-7 gene is required for oogenesis and female fertility, genetically interacts with piwi and aubergine, but impacts only weakly transposon silencing. *PLoS ONE* 9:e96802. doi: 10.1371/journal.pone.0096802
- Biemont, C., and Vieira, C. (2006). Genetics: junk DNA as an evolutionary force. *Nature* 443, 521–524. doi: 10.1038/443521a
- Branciforte, D., and Martin, S. L. (1994). Developmental and cell type specificity of LINE-1 expression in mouse testis: implications for transposition. *Mol. Cell. Biol.* 14, 2584–2592. doi: 10.1128/MCB.14.4.2584
- Brennecke, J., Aravin, A. A., Stark, A., Dus, M., Kellis, M., Sachidanandam, R., et al. (2007). Discrete small RNA-generating loci as master regulators of transposon activity in *Drosophila*. *Cell* 128, 1089–1103. doi: 10.1016/j.cell.2007.01.043
- Cerutti, H., and Casas-Mollano, J. A. (2006). On the origin and functions of RNA-mediated silencing: from protists to man. *Curr. Genet.* 50, 81–99. doi: 10.1007/s00294-006-0078-x
- Chambeyron, S., Popkova, A., Payen-Groschene, G., Brun, C., Laouini, D., Pelisson, A., et al. (2008). piRNA-mediated nuclear accumulation of retrotransposon transcripts in the *Drosophila* female germline. *Proc. Natl. Acad. Sci. U.S.A.* 105, 14964–14969. doi: 10.1073/pnas.0805943105
- de Vanssay, A., Bouge, A. L., Boivin, A., Hermant, C., Teyssat, L., Delmarre, V., et al. (2012). Paramutation in *Drosophila* linked to emergence of a piRNA-producing locus. *Nature* 490, 112–115. doi: 10.1038/nature11416
- Dufourt, J., Brassat, E., Desset, S., Pouchin, P., and Vaury, C. (2011). Polycomb group-dependent, heterochromatin protein 1-independent, chromatin

- structures silence retrotransposons in somatic tissues outside ovaries. *DNA Res.* 18, 451–461. doi: 10.1093/dnares/dsr031
- Dufourt, J., Dennis, C., Boivin, A., Gueguen, N., Theron, E., Goriaux, C., et al. (2014). Spatio-temporal requirements for transposable element piRNA-mediated silencing during *Drosophila* oogenesis. *Nucleic Acids Res.* 42, 2512–2524. doi: 10.1093/nar/gkt1184
- Goriaux, C., Desset, S., Renaud, Y., Vaury, C., and Brasset, E. (2014). Transcriptional properties and splicing of the flamenco piRNA cluster. *EMBO Rep.* 15, 411–418. doi: 10.1002/embr.201337898
- Grentzinger, T., Armenise, C., Brun, C., Mugat, B., Serrano, V., Pelisson, A., et al. (2012). piRNA-mediated transgenerational inheritance of an acquired trait. *Genome Res.* 22, 1877–1888. doi: 10.1101/gr.136614.111
- Grimson, A., Srivastava, M., Fahey, B., Woodcroft, B. J., Chiang, H. R., King, N., et al. (2008). Early origins and evolution of microRNAs and Piwi-interacting RNAs in animals. *Nature* 455, 1193–1197. doi: 10.1038/nature07415
- Haase, A. D., Fenoglio, S., Muerdter, F., Guzzardo, P. M., Czech, B., Pappin, D. J., et al. (2010). Probing the initiation and effector phases of the somatic piRNA pathway in *Drosophila*. *Genes Dev.* 24, 2499–2504. doi: 10.1101/gad.1968110
- Hajkova, P., Erhardt, S., Lane, N., Haaf, T., El-Maarri, O., Reik, W., et al. (2002). Epigenetic reprogramming in mouse primordial germ cells. *Mech. Dev.* 117, 15–23. doi: 10.1016/S0925-4773(02)00181-8
- Handler, D., Olivieri, D., Novatchkova, M., Gruber, F. S., Meixner, K., Mechtler, K., et al. (2011). A systematic analysis of *Drosophila* TUDOR domain-containing proteins identifies Vreteno and the Tdrd12 family as essential primary piRNA pathway factors. *EMBO J.* 30, 3977–3993. doi: 10.1038/emboj.2011.308
- Hsieh, T. F., Ibarra, C. A., Silva, P., Zemach, A., Eshed-Williams, L., Fischer, R. L., et al. (2009). Genome-wide demethylation of Arabidopsis endosperm. *Science* 324, 1451–1454. doi: 10.1126/science.1172417
- Huang, X. A., Yin, H., Sweeney, S., Raha, D., Snyder, M., and Lin, H. (2013). A major epigenetic programming mechanism guided by piRNAs. *Dev. cell* 24, 502–516. doi: 10.1016/j.devcel.2013.01.023
- Kiuchi, T., Koga, H., Kawamoto, M., Shoji, K., Sakai, H., Arai, Y., et al. (2014). A single female-specific piRNA is the primary determinant of sex in the silkworm. *Nature* 509, 633–636. doi: 10.1038/nature13315
- Klenov, M. S., Lavrov, S. A., Korbut, A. P., Stolyarenko, A. D., Yakushev, E. Y., Reuter, M., et al. (2014). Impact of nuclear Piwi elimination on chromatin state in *Drosophila melanogaster* ovaries. *Nucleic Acids Res.* 42, 6208–6218. doi: 10.1093/nar/gku268
- Le Thomas, A., Stuwe, E., Li, S., Du, J., Marinov, G., Rozhkov, N., et al. (2014). Transgenerationally inherited piRNAs trigger piRNA biogenesis by changing the chromatin of piRNA clusters and inducing precursor processing. *Genes Dev.* 28, 1667–1680. doi: 10.1101/gad.245514.114
- Luteijn, M. J., and Ketting, R. F. (2013). PIWI-interacting RNAs: from generation to transgenerational epigenetics. *Nat. Rev. Genet.* 14, 523–534. doi: 10.1038/nrg3495
- Malone, C. D., Brennecke, J., Dus, M., Stark, A., McCombie, W. R., Sachidanandam, R., et al. (2009). Specialized piRNA pathways act in germline and somatic tissues of the *Drosophila* ovary. *Cell* 137, 522–535. doi: 10.1016/j.cell.2009.03.040
- Mohn, F., Sienski, G., Handler, D., and Brennecke, J. (2014). The rhino-deadlock-cutoff complex licenses noncanonical transcription of dual-strand piRNA clusters in *Drosophila*. *Cell* 157, 1364–1379. doi: 10.1016/j.cell.2014.04.031
- Peaston, A. E., Evsikov, A. V., Graber, J. H., De Vries, W. N., Holbrook, A. E., Solter, D., et al. (2004). Retrotransposons regulate host genes in mouse oocytes and preimplantation embryos. *Dev. Cell* 7, 597–606. doi: 10.1016/j.devcel.2004.09.004
- Ronsseray, S., Lemaitre, B., and Coen, D. (1993). Maternal inheritance of P cytotype in *Drosophila melanogaster*: a “pre-P cytotype” is strictly extrachromosomally transmitted. *Mol. Gen. Genet.* 241, 115–123. doi: 10.1007/BF00280208
- Rouget, C., Papin, C., Boureux, A., Meunier, A. C., Franco, B., Robine, N., et al. (2010). Maternal mRNA deadenylation and decay by the piRNA pathway in the early *Drosophila* embryo. *Nature* 467, 1128–1132. doi: 10.1038/nature09465
- Rougier, N., Bourc’his, D., Gomes, D. M., Niveleau, A., Plachot, M., Paldi, A., et al. (1998). Chromosome methylation patterns during mammalian preimplantation development. *Genes Dev.* 12, 2108–2113. doi: 10.1101/gad.12.14.2108
- Rozhkov, N. V., Hammell, M., and Hannon, G. J. (2013). Multiple roles for Piwi in silencing *Drosophila* transposons. *Genes Dev.* 27, 400–412. doi: 10.1101/gad.209767.112
- Sienski, G., Donertas, D., and Brennecke, J. (2012). Transcriptional silencing of transposons by Piwi and maelstrom and its impact on chromatin state and gene expression. *Cell* 151, 964–980. doi: 10.1016/j.cell.2012.10.040
- Slotkin, R. K., Vaughn, M., Borges, F., Tanurdzic, M., Becker, J. D., Feijo, J. A., et al. (2009). Epigenetic reprogramming and small RNA silencing of transposable elements in pollen. *Cell* 136, 461–472. doi: 10.1016/j.cell.2008.12.038
- Zamudio, N., and Bourc’his, D. (2010). Transposable elements in the mammalian germline: a comfortable niche or a deadly trap? *Heredity (Edinb)* 105, 92–104. doi: 10.1038/hdy.2010.53

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 03 September 2014; accepted: 22 October 2014; published online: 06 November 2014.

Citation: Dufourt J and Vaury C (2014) During a short window of *Drosophila* oogenesis, piRNA biogenesis may be boosted and mobilization of transposable elements allowed. *Front. Genet.* 5:385. doi: 10.3389/fgene.2014.00385

This article was submitted to *Epigenomics and Epigenetics*, a section of the journal *Frontiers in Genetics*.

Copyright © 2014 Dufourt and Vaury. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.