



Influence of raw milk microflora and starter cultures in cheese on protein hydrolysis and bioactive peptide generation during digestion

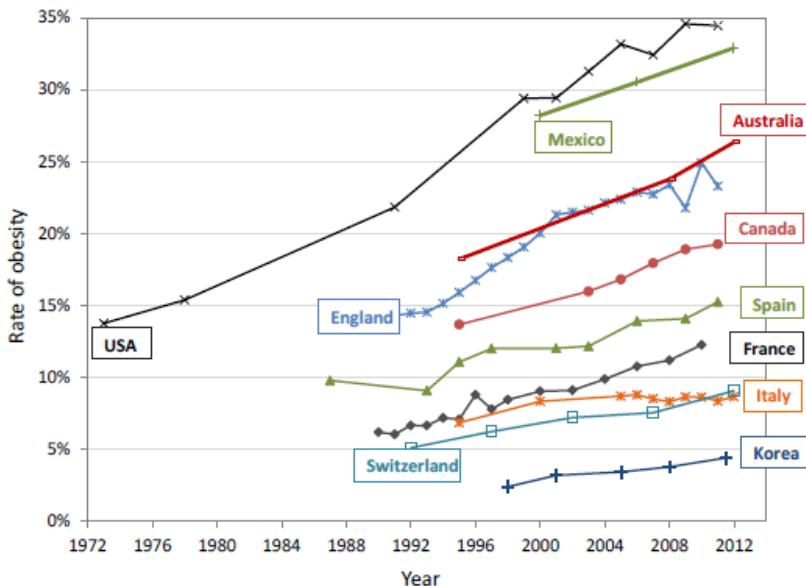


L Egger¹, O Ménard², L Abbuehl¹, D Duerr¹, R Badertscher¹, H Stoffers¹, H Berthoud¹, M Meola¹, G Vergères¹, D Dupont², R Portmann¹

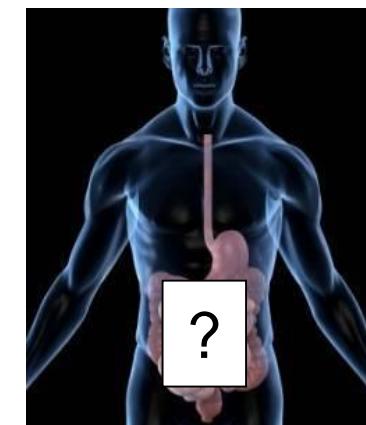
¹Agroscope, CH-3003 Berne, www.agroscope.ch; ²STLO, UMR 1253, INRA, Agrocampus Ouest, 35000 Rennes, France



Food and human health: the key role of digestion



Diet-related diseases ↑
Prevent these pathologies rather than
cure them



Gut = interface between food and human body

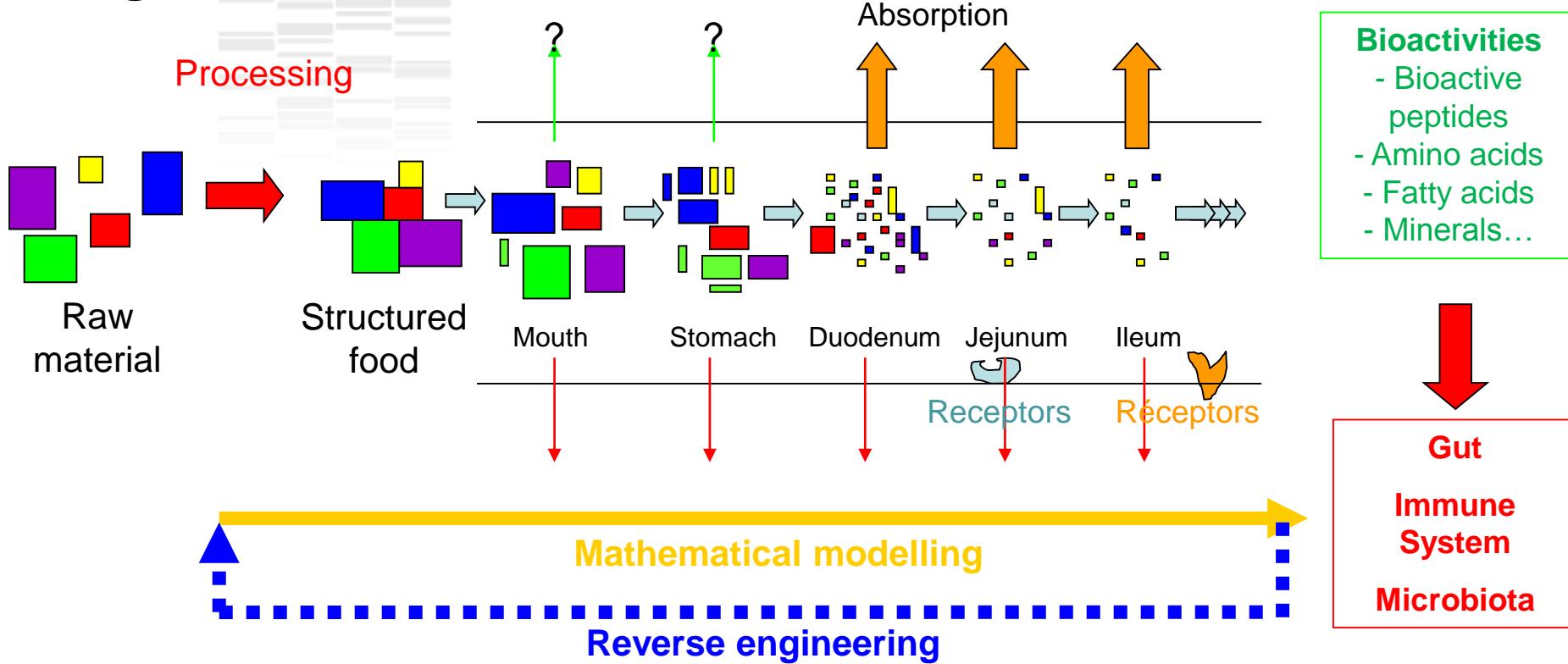
Digestion releases food components that can have a beneficial or a deleterious effect on human health

... but the mechanisms of food disintegration in the gastrointestinal tract remain unclear and the digestive process has been considered as a black box so far

By increasing our knowledge on food digestion, we will increase our knowledge on the effect of food on human health

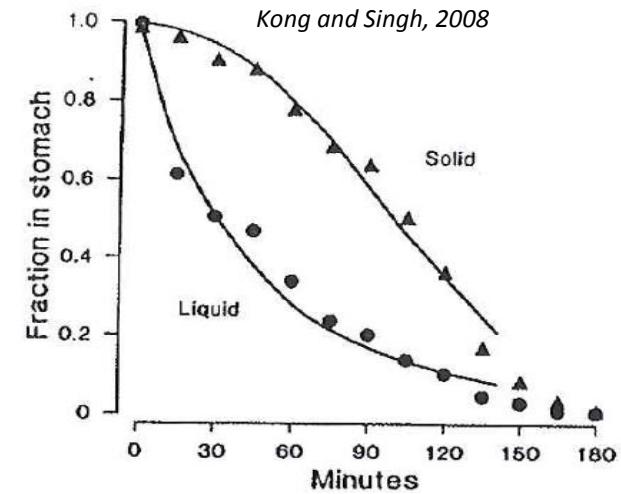
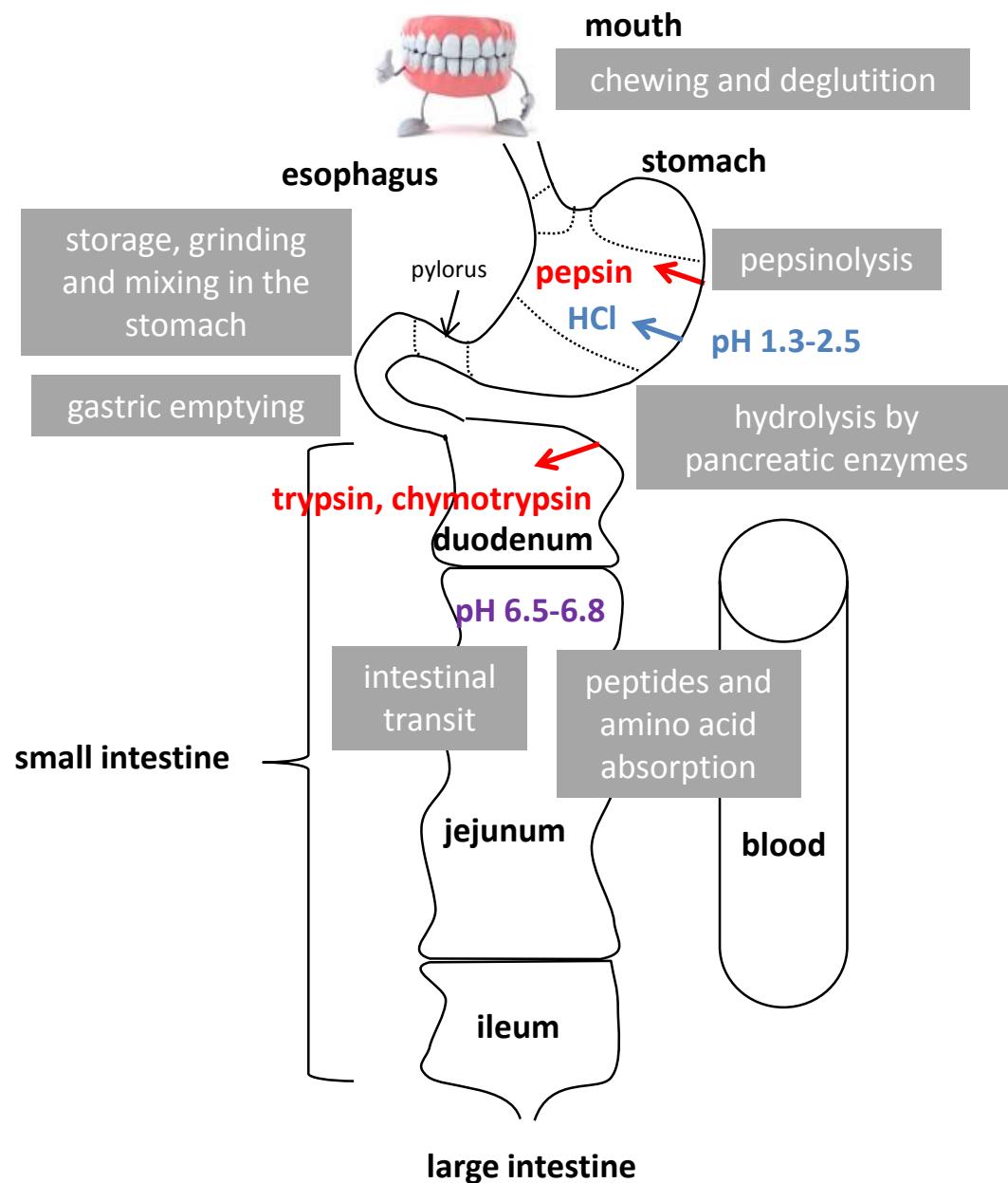
Our goals

Neonate/ Elderly



- To understand the mechanisms of breakdown of food matrices and their constituents in the gut and identify the beneficial/deleterious food components released during digestion
- To determine the impact of the structure of food matrices on these mechanisms
- To model these phenomena in order to develop a reverse engineering approach

The digestive process



→ Gastric phase = key step for the whole digestion process

Models available at INRA for simulating digestion

Dupont et al.
2010ab,
Mol Nutr Food Res

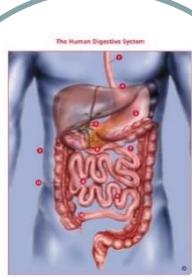


*In vitro static models
(infant, adult)*

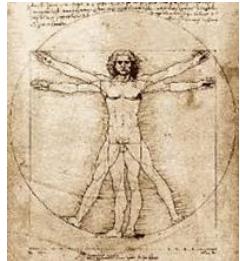
Le Feunteun et al.
2014
Food Bioprocess
Tech

$$\Phi_{12} = k_{12\text{whey}} \times (V_1 - m_{caswpd1} \times \alpha) + k_{12\text{aggr}} \times m_{caswpd1} \times \alpha$$

*In silico
models*



*In vitro dynamic models
(infant, adult, elderly)*



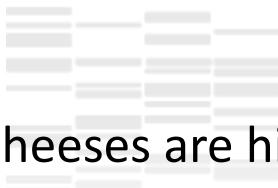
*Human
models*



Animal models

De Oliveira et al. 2016
Am J Clin Nutr
De Oliveira et al. 2017
Clin Nutr

Menard et al. 2014,
Food Chem
Sanchez et al. 2015
Food Res Int



Context of the study

Cow milk cheeses are highly-consumed products all over Europe

They provide highly digestible proteins and lipids, bioavailable calcium as well as a complex microbial ecosystem to the body

However, cheese digestion has been scarcely studied so far and their digestion products have never been exhaustively characterized

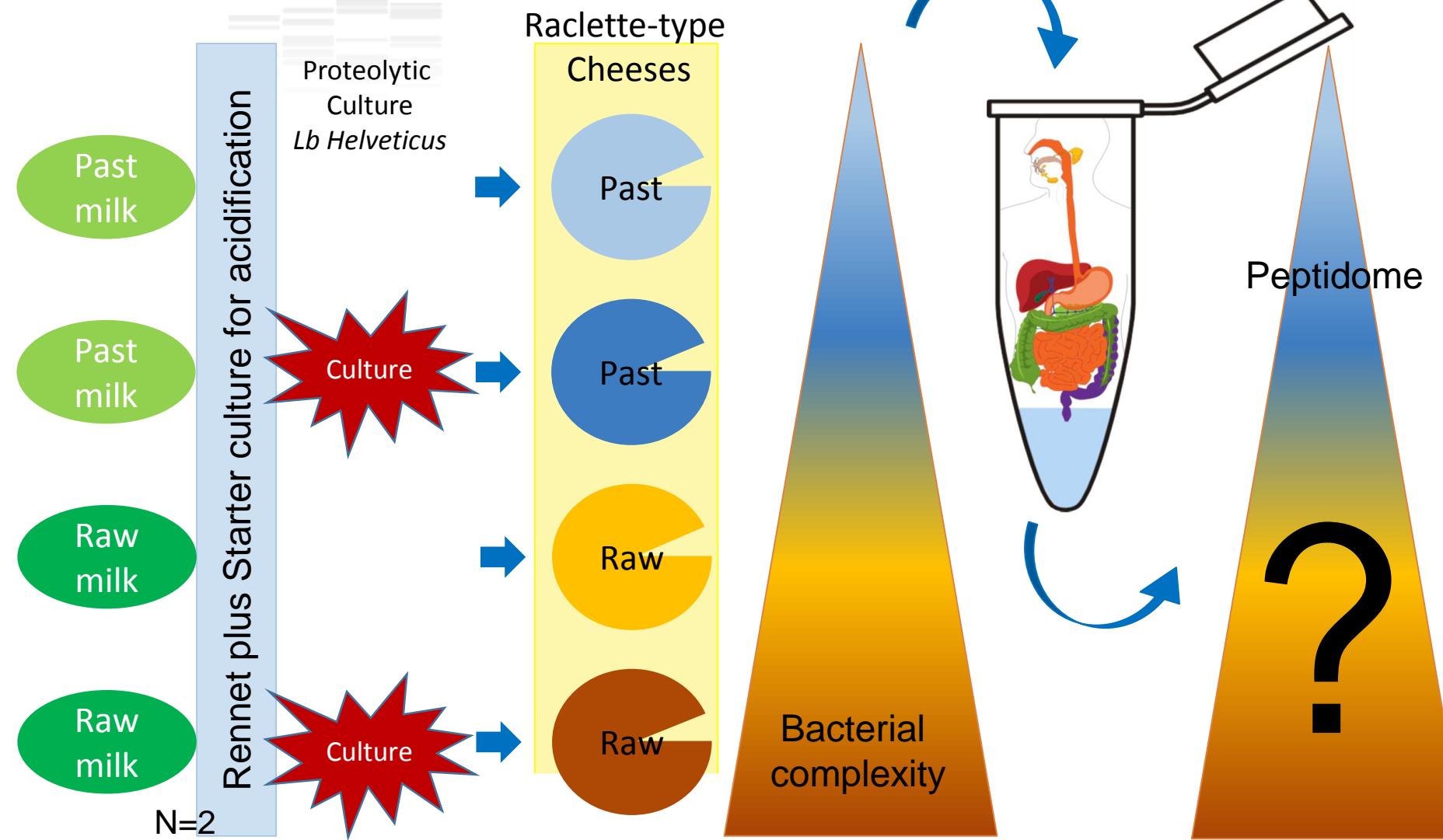
Research Questions

Do bacterial strains in cheese have an impact on protein hydrolysis during human digestion?

Does a higher microbial diversity lead to the generation of a higher number and diversity of peptides after digestion?

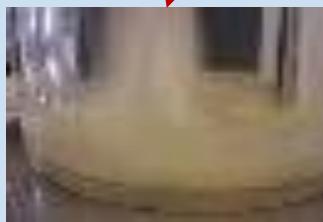
Are there differences at a molecular level after digestion of cheeses with a different bacterial background?

Experimental Setting



In vitro digestion methods

Dynamic (DIDGI®)



- Gastric :
- Pepsin (2000U/ml) 0.5ml/min
 - HCl 1M to achieve pH=1.68+3.52^(-t/42)
 - Emptying t_{1/2}=85min β=1.8
 - duration: 150min

- Intestinal digestion:
- Bile (2%) 0.5ml/min
 - Pancreatin (7%) 0.25ml/min
 - NaHCO₃ 1M to achieve pH=6.8
 - Emptying t_{1/2}=250min β=2.5
 - duration: 240min

Kinetic digestions n=2, cheeses: 4 X 2

Static (INFOGEST)

PAPER



A standardised static *in vitro* digestion method suitable for food – an international consensus†

M. Minekus,^{‡a} M. Alminger,^{‡b} P. Alvito,^{‡c} S. Ballance,^{‡d} T. Bohn,^{‡e} C. Bourlieu,^{‡f} F. Carrière,^{§g} R. Boutrou,^{‡h} M. Corredig,^{‡i} D. Dupont,^{§h} C. Dufour,^{‡j} L. Egger,^{‡k} M. Golding,^{||l} S. Karakaya,^{‡m} B. Kirkhus,^{§n} S. Le Feunteun,^{‡o} U. Lesmes,^{‡p} A. Macierzanka,^{‡q} A. Mackie,^{‡r} S. Marze,^{§s} D. J. McClements,^{||l} O. Ménard,^{‡f} I. Recio,^{‡u} C. N. Santos,^{‡w} R. P. Singh,^{||x} G. E. Vigarud,^{‡y} M. S. J. Wickham,^{‡z} W. Weitschies,^{‡aa} and A. Brodkorb,^{*ab}

- Gastric:
- Pepsin (2000U/ml)
 - pH=3
 - duration: 120min

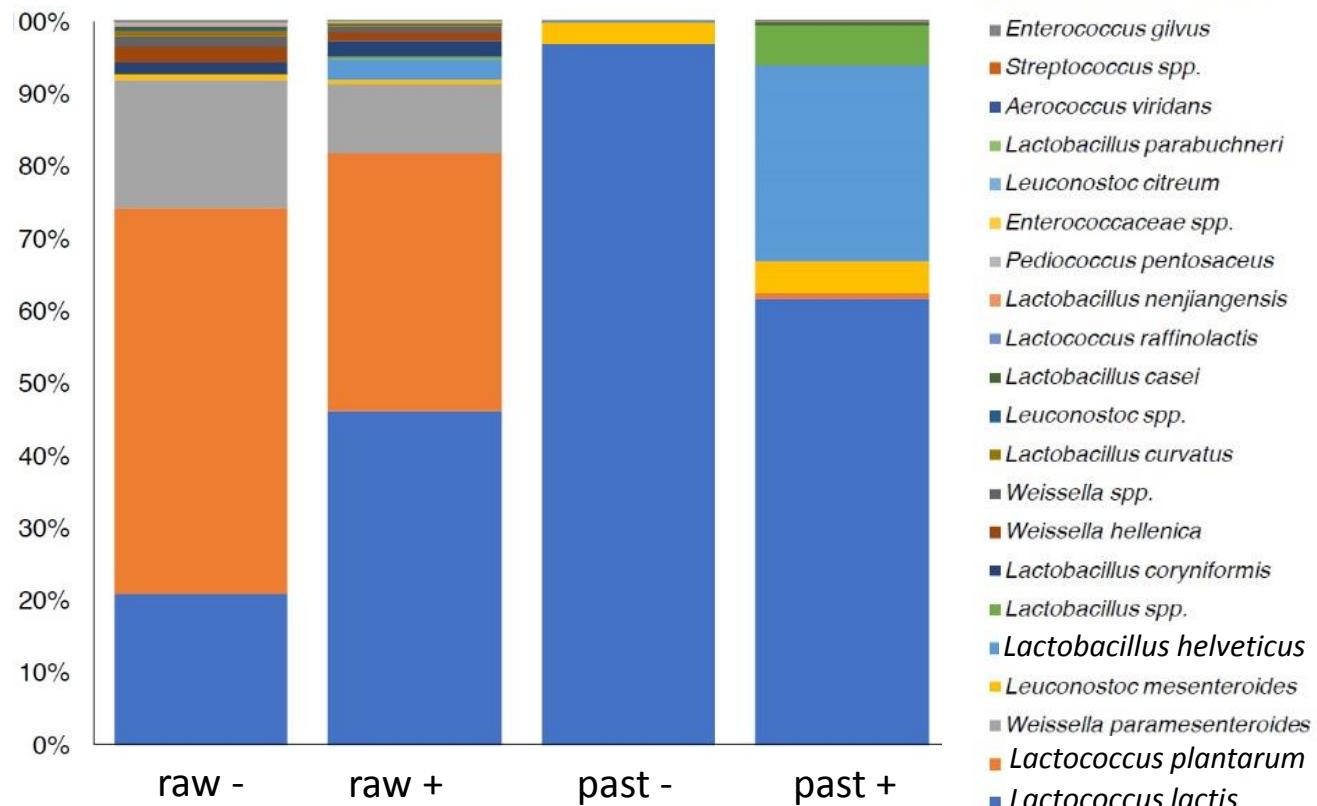
- Intestinal digestion:
- Bile salts 10mM
 - Pancreatin: to react 100 U of trypsin/ml digesta
 - pH=7
 - duration: 120min

Kinetic digestion n=1, Endpoints n=3, cheeses: 4 X 2

Characterization of Cheeses

Microbial Diversity

Bacterial community based on 16S rRNA sequencing (v1v2) after 120 days of ripening:

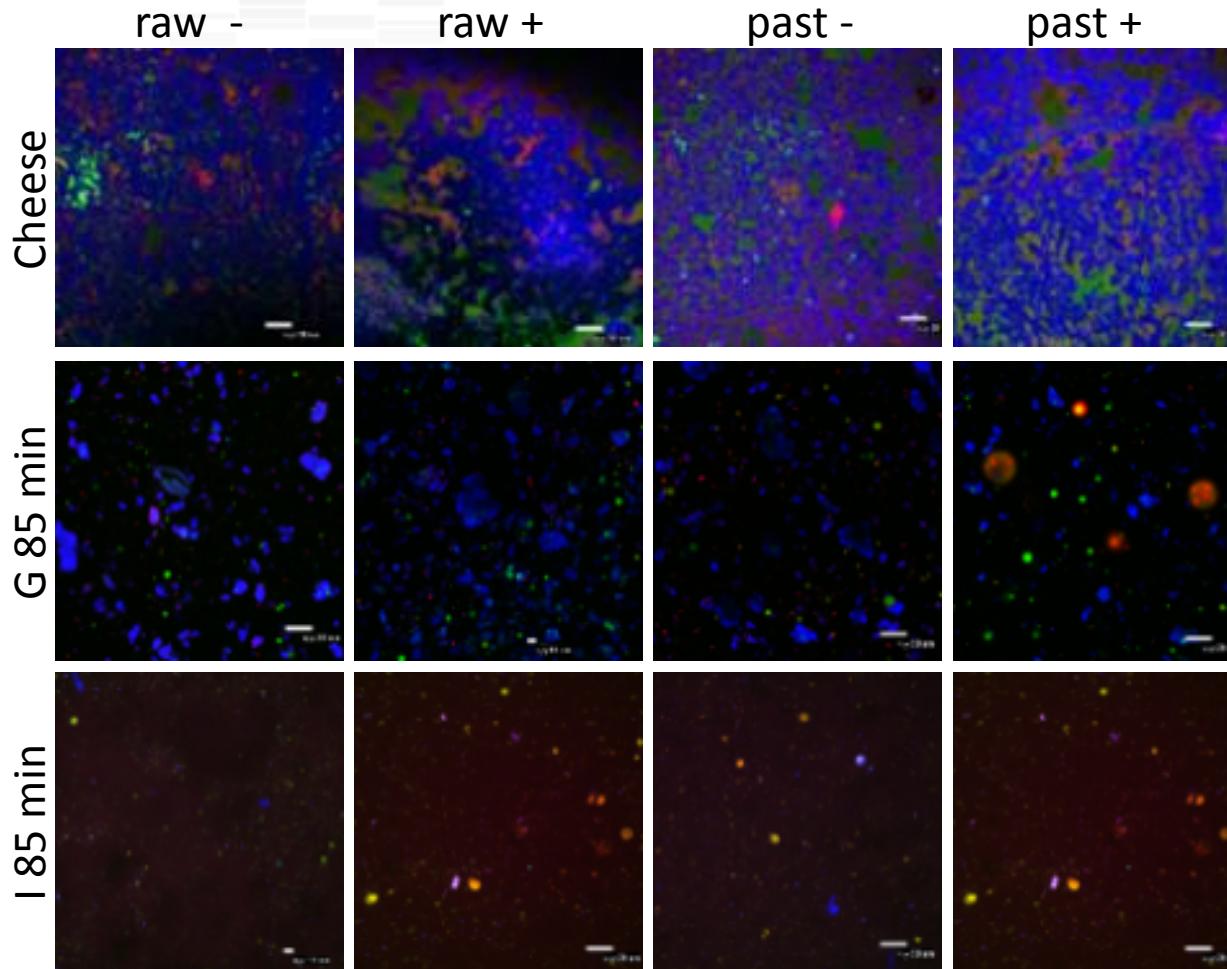


Composition

of four different (n=2) Swiss Raclette cheeses
After 120 days of ripening:

Cheese		sample	fat	NaCl	true protein	NPN	pH	Free NH ₂
milk	LH		g/kg	g/kg	g/kg	g/kg		mmol/kg
	no	raw -	290.3	18.9	244.37	5.7	5.1	151.3
raw	yes	raw +	295.8	17.9	250.26	5.6	5.1	151.5
	no	past -	284.8	21.0	247.41	5.7	5.1	143.2
past	yes	past +	288.3	18.9	238.26	5.8	5.1	145.2

Structural evolution of the cheeses during dynamic IVD



Apolar lipids
Proteins
Amphiphilic compounds

Cheese:

- network of proteins, lipids
- amphiphilic compounds at the interfaces

Soluble gastric phase

- Protein aggregates
- lipid droplets and peptides/FFA
- lipid layer on top of the gastric bowl

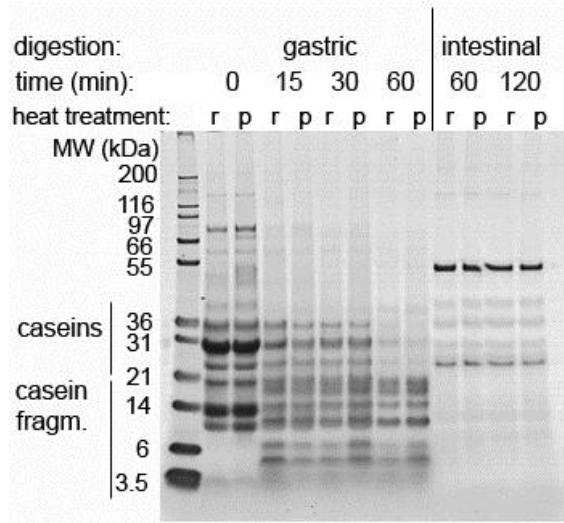
Intestinal phase:

- few protein aggregates
- mixed micelles of apolar lipids + amphiphilic compounds (bile salts and hydrolysis products)

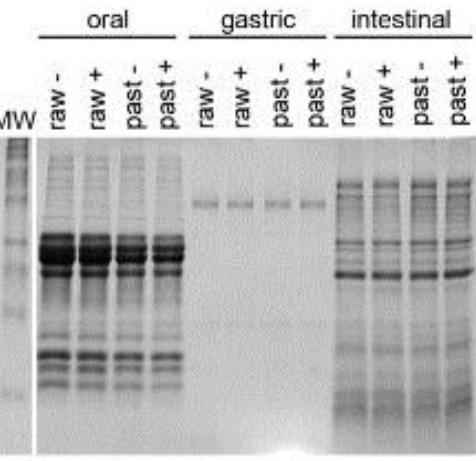
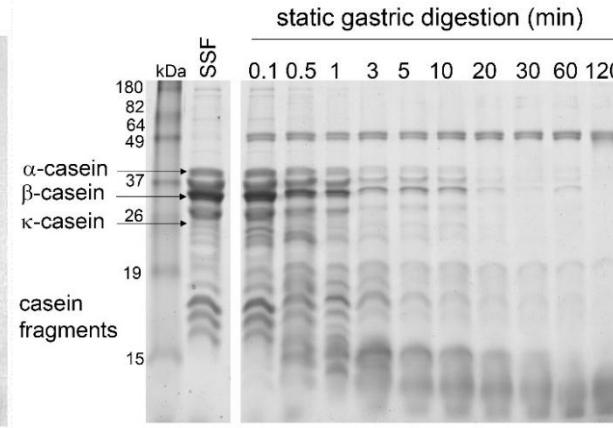
→ No structural differences visible between the four cheeses

Protein Hydrolysis

Dynamic



Static



Similar proteolysis pattern in dynamic and static *in vitro* digestion

Gastric digestion:

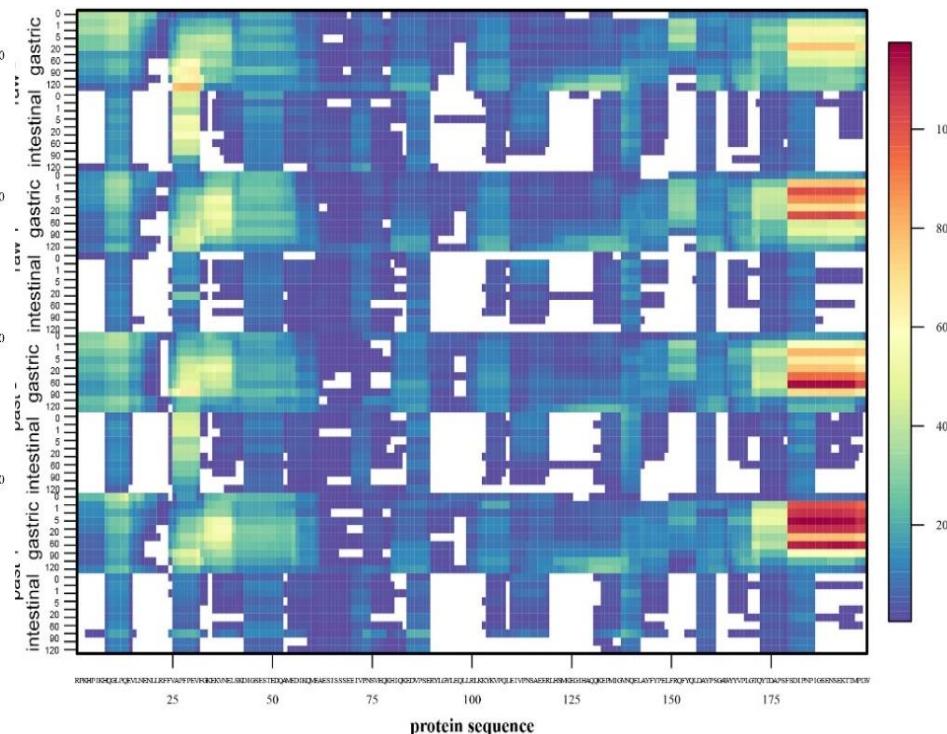
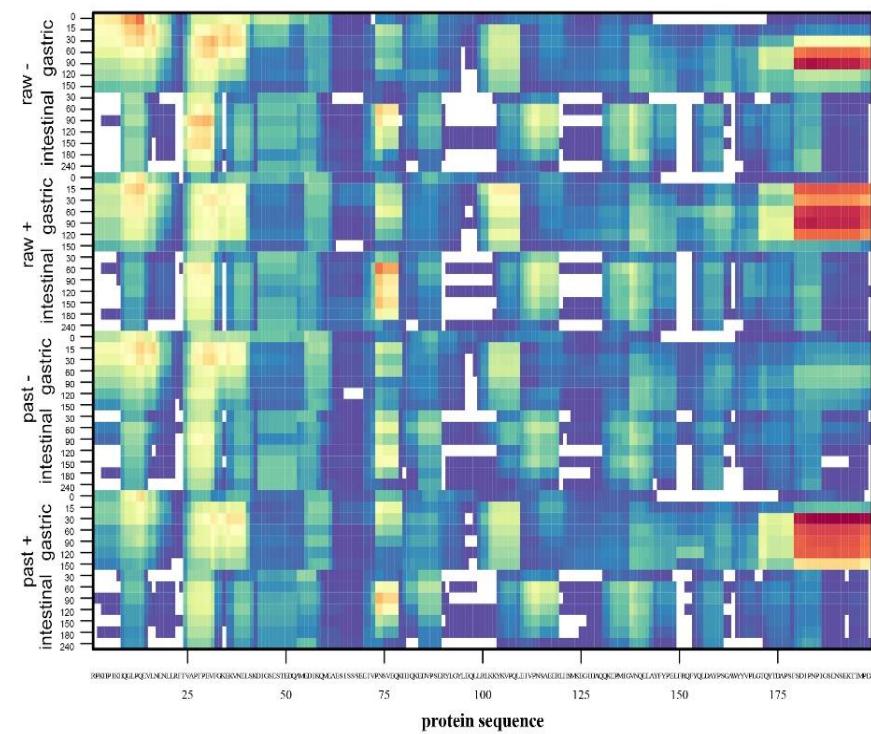
- Fast hydrolysis of intact caseins in all cheeses
- At 60 min of digestion no intact caseins remain but large fragments present

Intestinal digestion:

- Only proteins from pancreatin visible

Peptide generation during IVD

α_{s1} -casein



Gastric digestion:

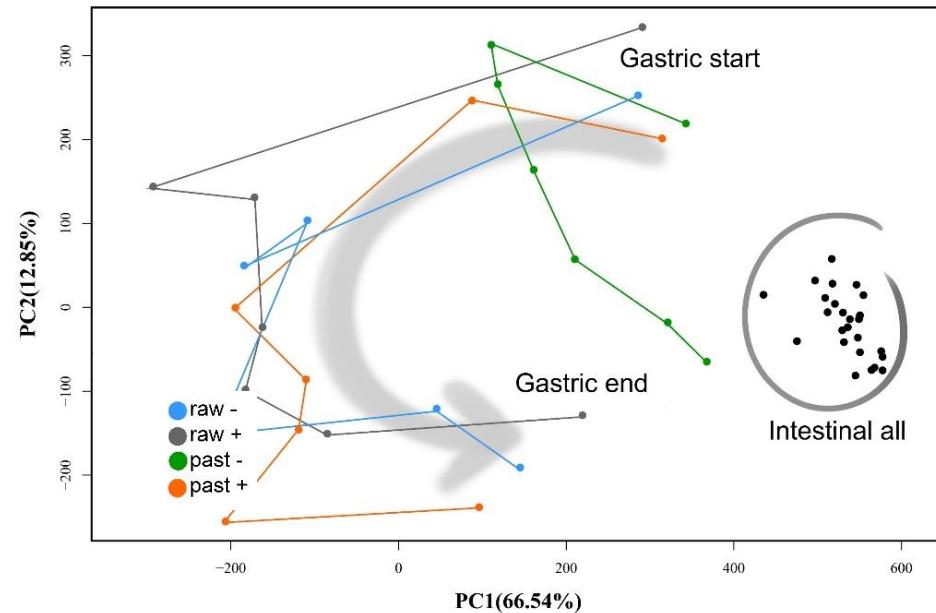
- Higher number of peptides at the C-terminus of the cheese samples with the proteolytic strain (observed in all caseins)

Intestinal digestion:

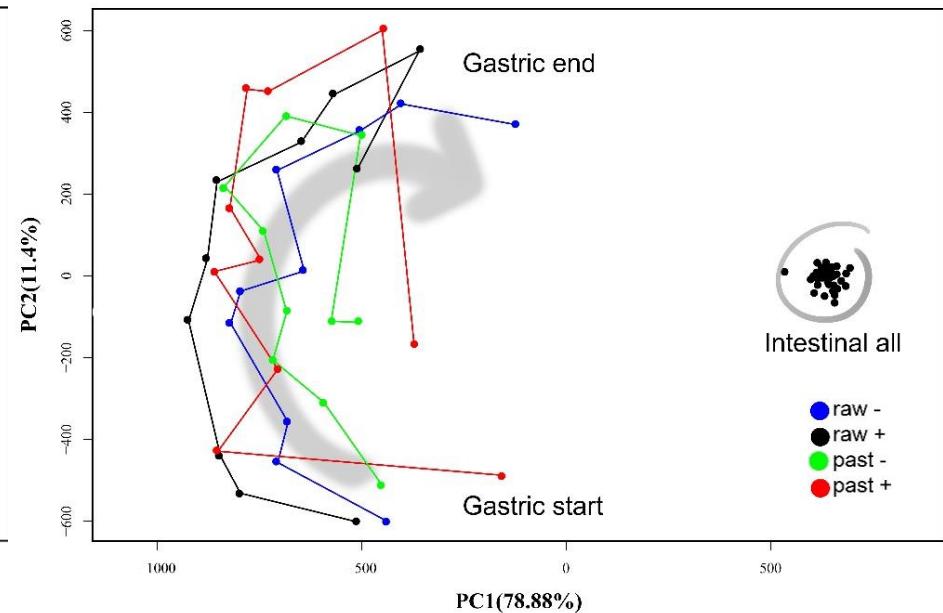
- Differences have been wiped off by the action of intestinal proteases

Peptide generation during IVD

Dynamic



Static



Gastric digestion:

- Peptidomes of cheese samples show an evolution in both digestion models

Intestinal digestion:

- No differences observed in the intestinal phase for the four different cheeses in both digestion models

Bioactive Peptides

β -casein, dynamic digestion

Opioid¹

Hypertension²

DPP-IV
inibitory³



Peptide	Cheese								raw / past
	1	3	2	4	5	7	6	8	
raw -	raw +	past -	past +						
EELNVPGEI	2	2	1	0	0	0	0	0	0.04
LNVPGEI	1	1	0	0	0	0	0	0	0.13
NVPGEIVES	0	0	0	0	2	1	1	0	0.05
NVPGEIVES <u>L</u>	10	10	10	6	5	7	8	0	0.10
FQSEEQQQT	0	0	0	0	1	1	0	0	0.13
QTEDEL	1	1	0	3	4	6	3	1	0.11
TEDEL	18	18	20	13	11	15	11	11	0.03
TEDELQD	2	2	5	7	11	8	7	5	0.08
PFAOTOSLV	1	1	0	0	0	0	0	0	0.13
YFPFPGPIN	0	0	0	2	3	3	2	0	0.13
PFPGPIP	5	5	5	4	1	0	3	1	0.00
PQNIPPLTQTP	1	1	0	0	0	0	0	0	0.13
PQNIPPL	1	1	0	0	0	0	0	0	0.13
NIPPL	11	11	8	5	7	7	6	3	0.13
NIPPLT	8	8	3	0	0	2	1	0	0.10
PPI TQT	0	0	0	0	0	1	0	1	0.13
PVVPVP	7	6	5	3	2	2	4	3	0.04
VVVPPPLQPEV	1	1	1	2	0	0	0	0	0.00
PPFLQPE	7	7	9	13	14	16	14	18	0.01
PFLQPEV	0	0	0	2	2	4	3	4	0.01
SLTLTDVENLHLPLPLL <u>QSWM</u> HQPHQPLPT	1	1	1	0	0	0	0	0	0.02
HLPLP	0	0	0	0	2	0	0	3	0.15
LLQSWMHQPHQPLPT	2	2	1	0	1	0	0	0	0.11
MHQPHQPLPT	30	30	31	18	22	15	9	1	0.03
QPHQLPPT	0	0	0	1	3	2	3	0	0.06
FPPQSV	14	14	13	21	12	10	7	13	0.07
<u>SVLSL</u> SQS K VLPVP <u>Q</u> KAVPY	1	1	0	0	0	0	0	0	0.13
LPVPQ	7	7	8	5	5	4	6	6	0.11
MFPPQ	2	2	3	2	1	2	1	1	0.03
MFPPQS	4	4	2	2	1	0	0	0	0.00
AVPYPP	5	5	5	5	0	0	0	5	0.02

¹Nongonierma, 2015; ²Aluko, 2015, ³Nongonierma, 2016

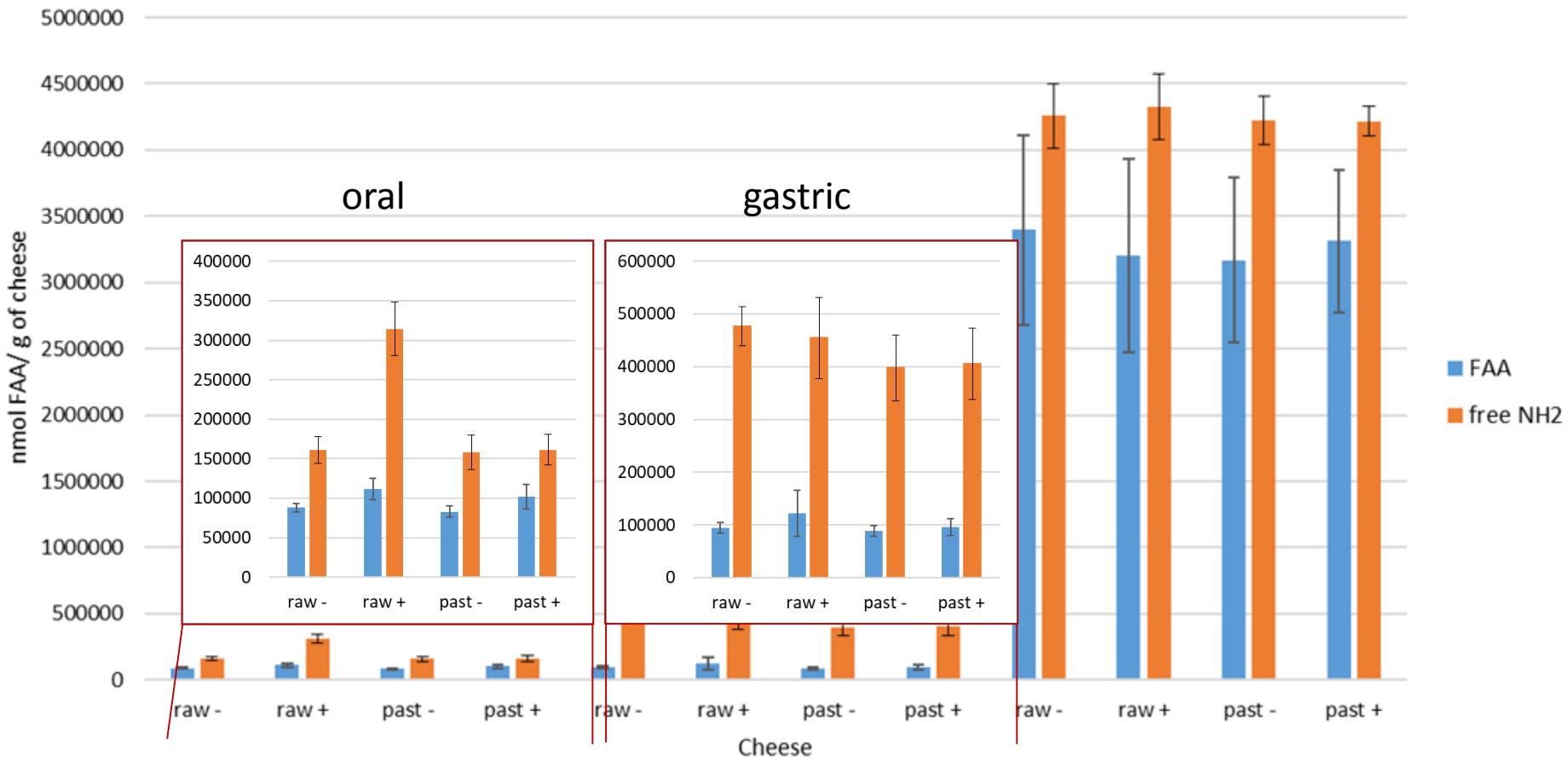
Peptide	Cheese								culture +/-
	1	3	5	7	2	4	6	8	
minus culture	plus culture								
YPVEP	3	3	2	4	0	1	3	2	0.10
TLTDVEN	4	4	4	3	4	1	3	1	0.11
LHLPLP	0	0	1	0	1	2	0	3	0.12
LLQSWMHQPHQPLPT	2	2	1	0	1	0	0	0	0.11
<u>SVLSL</u> SQS K VLPVP <u>Q</u> KAVPY	1	1	0	0	0	0	0	0	0.13
RDMPI	0	0	0	0	1	0	0	1	0.13
QEPVLGPV	0	0	3	2	4	2	3	3	0.09
LNVPGEIVES <u>L</u>	3	3	2	2	2	0	1	0	0.02
LNVPGEI	1	1	0	0	0	0	0	0	0.13
PVLGPV	3	3	5	4	8	5	3	6	0.18
FQSEEQQQT	0	0	1	1	0	0	0	0	0.13
DELQDKI	1	1	0	2	0	0	0	0	0.05
PFAOTOSL V	1	1	0	0	0	0	0	0	0.13
VYPFPGPIN	11	11	10	10	8	9	8	0	0.09
VYPFPGP	9	9	9	7	93	11	16	27	0.19
PPFCPI	16	16	12	14	13	10	10	11	0.03
IPONIPPL	9	9	6	5	2	2	7	2	0.05
PQNIPPLTQTP	1	1	0	0	0	0	0	0	0.13
PQNIPPL	1	1	0	0	0	0	0	0	0.13
NIPPL	11	11	7	7	8	5	6	3	0.07
NIPPLT	8	8	0	2	3	0	1	0	0.16
TQTPVVPFPLQPEV	0	0	1	0	1	3	0	1	0.19
TQTPV	3	3	2	1	1	2	2	0	0.19
TPVVVPP	2	2	2	0	0	0	0	0	0.02

Tendency: ➔ Bioactive Peptides

- raw milk
- without proteolytic culture

Release of free amino acids

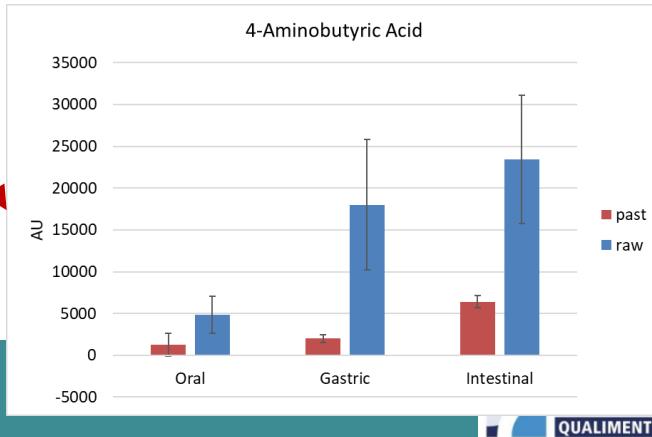
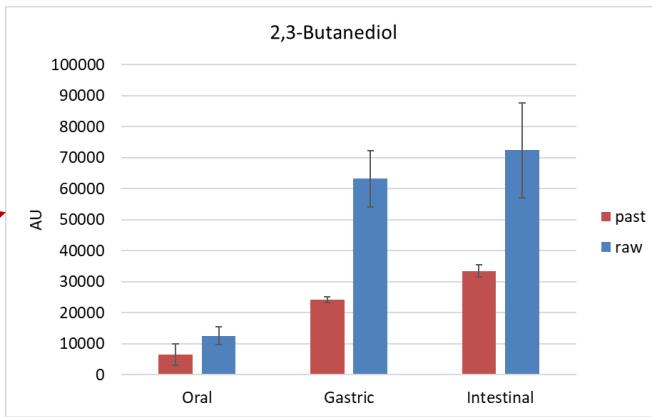
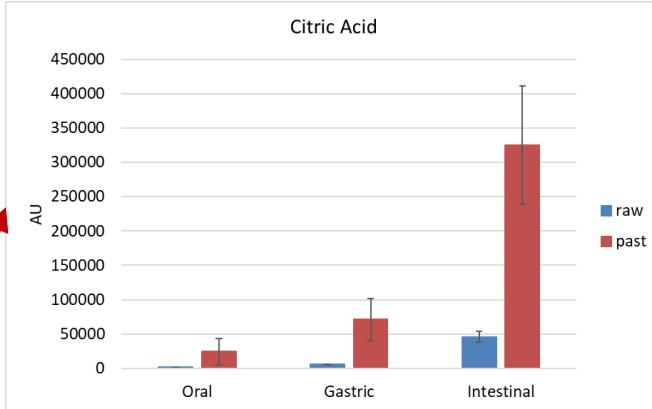
FAA in 1 g cheese gastric digest



→ Some differences between cheeses are visible in the oral or gastric phase but are wiped off after the intestinal phase

Metabolome

		Gastric		Intestinal	
	Name	t-Test	Kruskal-Wallis	t-Test	Kruskal-Wallis
Increased in Past milk Cheese	L-Serine	0.007	0.003	0.748	0.753
	Glyceric acid	0.000	0.001	0.077	0.115
	L-Serine	0.000	0.001	0.021	0.016
	Malic acid	0.000	0.001	0.158	0.172
	L-Serine	0.000	0.001	0.085	0.093
	L-Arabitol	0.000	0.001	0.472	0.016
	L-Glutamine	0.001	0.003	0.749	0.753
	Citric acid	0.001	0.001	0.000	0.001
	D-Tagatose	0.000	0.001	0.000	0.002
	D-Tagatose	0.000	0.001	0.829	0.401





Conclusions

- Raw milk cheeses have a higher microbial diversity after 4 month of ripening, resulting in a higher content of free amino acids and differences in specific metabolites
- After gastric digestion, cheeses with higher microbial complexity have more complex peptide patterns and a slightly higher proteolysis
- All cheese digesta have only minor differences after intestinal IVD under dynamic and static conditions and these differences originate from non digestable metabolites present in the cheeses



Reto Portman
Agroscope, Switzerland

Head

Didier DUPONT - Senior Scientist

Scientists

Rachel BOUTROU – Scientist

Amélie DEGLAIRE – Lecturer

Juliane FLOURY – Lecturer

Catherine GUERIN - Lecturer

Joëlle LEONIL – Senior Scientist

Steve LE FEUNTEUN – Scientist

Françoise NAU – Professor

Frédérique PEDRONO – Lecturer

Ashkan MADADLOU – Post-doc

The Bioactivity & Nutrition team



Lotti Egger
Agroscope, Switzerland

PhD students

Lucie LORIEAU (2016-2019)

Linda LEROUX (2016-2019)

Manon HIOLLE (2016-2019)

Yohan REYNAUD (2016-2019)

Jun WANG (2018-2021)

Lea SALELLES (2018-2021)

Technicians

Gwenaële HENRY

Yann LE GOUAR

Nathalie MONTHEAN

Engineers

Julien JARDIN

Olivia MENARD

Jordane OSSEMOND

Masters students

Improving health properties of food by sharing our knowledge on the digestive process

International Network

Dr. Didier DUPONT, Senior Scientist, INRA, France





Industry involvement

→ ~ 50 private companies are following INFOGEST





INFOGEST



Chair
Didier Dupont - France



Vice-chair
Alan Mackie - UK

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lipases and
lipid digestion
WG4

Digestive
amylases and
starch
digestion
WG5

In silico
models of
digestion
WG6



Didier Dupont



Alan Mackie



Uri Lesmes



Myriam Grundy



Nadja Siegert



Choi-Hong Lai

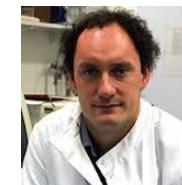


Guy Vergeres

The “Mind-
the-Gap”
group



Frederic Carriere



Fred Warren



Steven Le Feunteun



INFOGEST
International Conference
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April 2019

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