



**HAL**  
open science

## Equilibrium and Out-of-Equilibrium Adherence of Hydrogels against Polymer Brushes

Jennifer Macron, Bruno Bresson, Yvette Tran, Dominique Hourdet,  
Costantino Creton

► **To cite this version:**

Jennifer Macron, Bruno Bresson, Yvette Tran, Dominique Hourdet, Costantino Creton. Equilibrium and Out-of-Equilibrium Adherence of Hydrogels against Polymer Brushes. *Macromolecules*, 2018, 51 (19), pp.7556 - 7566. 10.1021/acs.macromol.8b01063 . hal-01904369

**HAL Id: hal-01904369**

**<https://hal.science/hal-01904369v1>**

Submitted on 23 Dec 2018

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

This document is confidential and is proprietary to the American Chemical Society and its authors. Do not copy or disclose without written permission. If you have received this item in error, notify the sender and delete all copies.

## Equilibrium and out-of-equilibrium Adherence of Hydrogels against Polymer Brushes

Journal:	<i>Macromolecules</i>
Manuscript ID	Draft
Manuscript Type:	Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Macron, Jennifer; ESPCI Paris, SIMM Bresson, Bruno; ESPCI, CNRS, LPQ/ESPCI Tran, Yvette; ESPCI Paris, SIMM Hourdet, Dominique; SIMM, Université Pierre et Marie Curie, Sorbonne-Universités, Creton, Costantino; E.S.P.C.I., Laboratoire SIMM-UMR 7615

SCHOLARONE™  
Manuscripts

1  
2  
3  
4  
5  
6  
7  
8 **Equilibrium and out-of-equilibrium Adherence of Hydrogels against**  
9 **Polymer Brushes**  
10  
11  
12  
13  
14  
15  
16  
17

18 Jennifer Macron<sup>a,c</sup>, Bruno Bresson<sup>a</sup>, Yvette Tran<sup>a</sup>, Dominique Hourdet<sup>a,b</sup> and Costantino  
19 Creton<sup>a,b,\*</sup>  
20  
21  
22  
23  
24  
25  
26  
27

28 <sup>a</sup>Laboratoire Sciences et Ingénierie de la Matière Molle, ESPCI Paris, PSL University,  
29 Sorbonne Université, CNRS, F-75005 Paris, France  
30

31 <sup>b</sup> Global Station for Soft Matter, Global Institution for Collaborative Research and Education,  
32 Hokkaido University, Sapporo, Japan  
33  
34

35 <sup>c</sup> current address: Bertarelli Foundation Chair in Neuroprosthetic Technology, Laboratory for Soft  
36 Bioelectronic Interfaces, Institute of Microengineering, Institute of Bioengineering, Center for  
37 Neuroprosthetics, Ecole Polytechnique Fédérale de Lausanne (EPFL), 1202 Geneva, Switzerland  
38

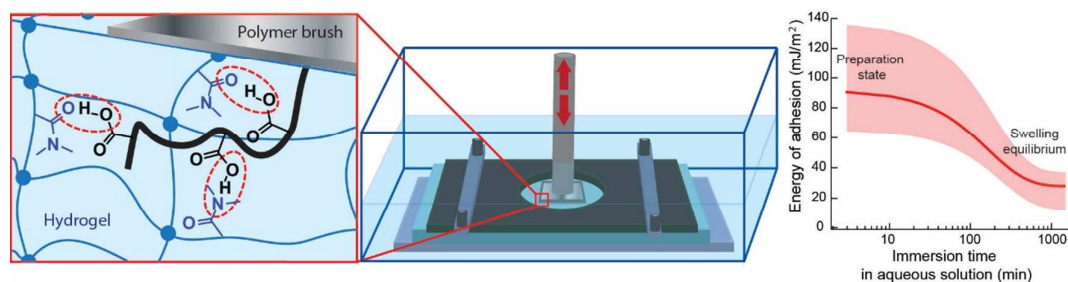
39 emails: [Jennifer.Macron@epfl.ch](mailto:Jennifer.Macron@epfl.ch), [Bruno.Bresson@espci.fr](mailto:Bruno.Bresson@espci.fr), [Yvette.Tran@espci.fr](mailto:Yvette.Tran@espci.fr),  
40 [Dominique.Hourdet@espci.fr](mailto:Dominique.Hourdet@espci.fr), [Costantino.Creton@espci.fr](mailto:Costantino.Creton@espci.fr)  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

## ABSTRACT

At low pH, underwater adherence between poly(*N,N*-dimethylacrylamide) hydrogels and poly(acrylic acid) brushes is due to the formation of multiple hydrogen bonds. The effect of the key parameters controlling the formation of these interactions (contact time and composition of the hydrogel) was investigated with a contact mechanics test using a flat probe. We specifically quantified the loss of adherence during the progressive swelling to equilibrium of the gels and, at fixed contact time, found a significant decrease in adherence between the preparation state and the swelling equilibrium even in the case of relatively low dilution factors. This adherence loss was attributed to the slowdown of the kinetics of formation of multiple H-bond interactions as the gel approached its equilibrated state. In both limiting conditions the energy of adherence scaled with the polymer concentration, independent of the crosslinks density of the gel, suggesting that the Lake and Thomas amplification factor is not relevant for these weak bonds.

**Keywords:** underwater adherence, hydrogels, polymer brush, weak interactions, out-of-equilibrium, swelling equilibrium.

## TOC



## 1. INTRODUCTION

The use of hydrogels as surgical adhesives<sup>1</sup>, drug-loaded skin or mucoadhesives<sup>2</sup>, injectable adhesive hydrogels<sup>3</sup> or tailor-made artificial organs through 3D bioprinting<sup>4</sup> has raised some fundamental questions about the hydrogel adherence with mucus in wet environments and its evolution over time. More recently some work has been done to control and improve the adherence of synthetic hydrogels to hydrophilic and hydrophobic surfaces for the engineering purpose of assembling multimaterials<sup>5-8</sup>. However, in all these cases, the main objective was to design and demonstrate a recipe or method that works, rather than to obtain a deep and general understanding of the underlying principles behind gel adherence.

In parallel some more systematic studies have also been carried out with model systems<sup>9-11</sup>, investigating among other things the effect of the detailed composition of the gel, of the environment (pH in particular), ionic strength or temperature on the measured adherence with the objective to develop robust models. Our study fits well in this latter category and focuses on the process of equilibration of the gel with its environment and its consequences on adhesive interactions.

Synthetic hydrogels are model hydrophilic soft materials that are typically swollen with water (80-95% of water) and have moduli in the kPa range. They are normally composed of flexible water-soluble polymer chains synthesized or crosslinked in the presence of a fixed concentration of water (constituting the preparation conditions) and, if left then in excess water, will swell to equilibrium. This equilibration with the environment until the chemical potential of the water and mobile species inside and outside the gel is the same, has been well studied and leads to changes in mechanical properties such as the elastic modulus and also to changes in the large strain properties<sup>12-13</sup>. However changes in surface properties and adherence due to this equilibration process in aqueous conditions have not been addressed systematically to the best of our knowledge.

The process of swelling to equilibrium of the polymer gel results in a dilution of the flexible chains and therefore in a reduction in their concentration. In the absence of heterogeneities, this decrease in polymer concentration also occurs on the surface of the hydrogel.

Taking a naïve picture of adherence, one can hypothesize that if adherence between two hydrogels or between a hydrogel and a polymer brush occurs at preparation conditions and at swelling equilibrium, the areal density of interactions should scale with the polymer concentration. When the hydrogel is separated from the surface in controlled conditions, the energy needed to perform this debonding test will in turn depend on the strength and areal density of these interactions (the stress that can be sustained by the interface) and also on the bulk dissipative mechanisms in the hydrogel. If the interactions are weak and the gels are elastic, bulk dissipation is negligible. In this case, the energy dissipated during the debonding (which we call work of adherence) can be attributed to the irreversible energy dissipation occurring during the breakup of weak bonds attached to flexible chains. This scenario has been discussed in detail by Chaudhury for hydrophobic systems<sup>14-15</sup> and assumes that this

1 bond scission is rate dependent as proposed originally by Evan Evans<sup>16</sup>, and depends also on the  
2 length of the chain to which the interacting group is attached, as proposed by Lake and Thomas<sup>17</sup> for  
3 the scission of main chain bonds in crosslinked flexible networks.  
4  
5

6  
7 It is the purpose of this article to explore different conditions under which underwater macroscopic  
8 adherence can be measured between a neutral hydrogel and a polymer brush in conditions where  
9 gel/brush molecular interactions can occur by hydrogen bonding. The model gel is made of poly(*N,N*-  
10 dimethylacrylamide) (PDMA) and the brush is made of poly(acrylic acid) (PAA). As reported  
11 previously this type of combination gives a weak and reversible adherence which depends strongly on  
12 the pH and temperature of the medium<sup>11, 18</sup>. Above the pK<sub>a</sub> of the acid/base functions of the brush, the  
13 carboxylic groups of PAA are negatively charged and no adherence is observed with neutral PDMA.  
14 However in acidic conditions (well below the pK<sub>a</sub> of the acid/base functions), the PAA brush is fully  
15 protonated and multiple hydrogen bonds can be formed between the brush and the gel, even in water,  
16 leading to a measurable macroscopic adherence.  
17  
18  
19  
20  
21

22 In a previous study<sup>11</sup>, we investigated the dependence of pH on the underwater adherence of neutral  
23 hydrogels swollen at equilibrium, i.e. in a situation where brush and gel were immersed for several  
24 hours in the medium before being put in contact. In those conditions, the adherence was measured  
25 below 100 mJ/m<sup>2</sup> and required several minutes of contact to be observed.  
26  
27  
28

29 We address here more specifically two important aspects: the variation in adherence energy as the gel  
30 preparation conditions are varied (concentration of monomer and degree of crosslinking) and the  
31 variations in the observed adherence energy as the gel, initially in preparation conditions,  
32 progressively swells to equilibrium in an acid solution.  
33  
34  
35  
36  
37

## 38 **2. EXPERIMENTAL SECTION**

### 39 2.1 Mold design for hydrogel layer preparation

40  
41 Performing a flat–flat adherence test between a hydrogel and a nanometrically smooth surface such as  
42 a polymer brush requires a macroscopic gel with a low amplitude surface roughness. To obtain this  
43 low roughness, we used a mold formed by two hydrophobically modified glass substrates (dimensions:  
44 75 x 25 mm), separated by a silicone rubber gasket (1.8 mm thick) and held together using binder clips  
45 (see suppl. info Figure S1). The hydrophobic substrates were obtained by chemically grafting  
46 octadecyltrichlorosilane (OTS, Sigma-Aldrich, ≥ 90%) on glass. Before any chemical modification,  
47 the glass substrates were cleaned in a “Piranha” solution (mixture of 70 vol % of sulfuric acid (Sigma-  
48 Aldrich, 97%) and 30 vol% of hydrogen peroxide (Sigma-Aldrich, 35%)), heated at 150 °C for 30  
49 minutes, then rinsed, sonicated in Milli-Q water bath and dried under nitrogen flow. Silanization  
50 occurred using a liquid phase method: clean substrates were immersed in a sealed reactor for 2 hours  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

at room temperature in a 0.5 vol% OTS solution prepared in extra-dry toluene. The glass substrates were then removed from the bath, rinsed with Milli-Q water and finally dried under nitrogen flow.

## 2.2 Synthesis of the PDMA hydrogels

The synthesis of poly(*N,N*-dimethylacrylamide) hydrogels was carried out in Milli-Q water at room temperature by free radical polymerization of  $x$  wt% of *N,N*-dimethylacrylamide (DMA, Sigma-Aldrich, 99%) monomers with  $y$  mol% of *N,N'*-methylenebisacrylamide (MBA, Sigma-Aldrich, 99%) as cross-linkers. As redox initiator system, we used 1 mol% potassium persulfate (KPS, Sigma-Aldrich,  $\geq 99\%$ ) and 1 mol% *N,N,N',N'*-tetramethylethylenediamine (TEMED, Sigma-Aldrich, 99%). After the dissolution of monomer, cross-linker and KPS in Milli-Q water, the solution was deoxygenated for 30 minutes with a bubbling of nitrogen. TEMED was then rapidly added to the formulation under stirring, and the solution was directly poured into the mold described in the previous section that was placed in a reactor under a nitrogen atmosphere, to avoid oxygen inhibition of radicals. Polymerization and cross-linking occurred simultaneously, and molds were left in the reactor for 10 hours at room temperature to ensure complete reaction. The hydrogels were kept into the mold closed with paraffin film for a maximum of 3 days, to avoid water evaporation, before being used for swelling and adhesive tests. The resulting synthesized gels will be referred to as: **P $x$ -CL $y$** , with  $x$  being the polymer concentration in wt% and  $y$  the cross-linking ratio in mol% relative to monomer. Table 1 summarizes the composition of the **P $x$ -CL $y$**  hydrogels that were synthesized for this study.

<b>P<math>x</math>-CL<math>y</math></b> PDMA hydrogels	DMA/g	H <sub>2</sub> O/g	KPS/mg	MBA/mg	TEMED/ $\mu$ L	Initial mass swelling ratio $\Lambda_0$
<b>P10-CL1</b>	1	9	27.3	15.6	15.2	10.3
<b>P10-CL2</b>	1	9	27.3	31.1	15.2	10.2
<b>P10-CL3</b>	1	9	27.3	46.7	15.2	10.0
<b>P15-CL3</b>	1.5	8,5	40.9	70.0	22.8	6.6
<b>P20-CL3</b>	2	8	54.5	93.3	30.4	4.9
<b>P30-CL3</b>	3	7	81.8	140	45.7	3.3

**Table 1: Chemical composition of P $x$ -CL $y$  PDMA hydrogels and swelling ratio in their preparation state.**

## 2.3 Kinetics and Swelling Measurements

The swelling behavior of the PDMA hydrogels was studied from the preparation state to swelling equilibrium as a function of immersion time. To quantify the kinetics of swelling, layers of PDMA hydrogels (dimensions: 73 x 23 x 1.8 mm<sup>3</sup>) were weighed in their preparation state before immersion

1 into a large volume of acid solution at pH 2 (adjusted by adding hydrochloric acid). The sample was  
2 regularly weighed until reaching its swelling equilibrium. After the study of the swelling kinetics, the  
3 sample was dried for 48 hours in a desiccator under vacuum at 80°C and the dry weight of polymer  
4 was measured. The mass swelling ratio  $\Lambda(t)$  as a function of immersion time was calculated from the  
5 weight data obtained with Equation 1. The results of these bulk swelling experiments are displayed in  
6 Figure S2 of the supplementary information.  
7  
8  
9

$$\Lambda(t) = \frac{m_{gel}(t)}{m_{polymer}^{dry}} = \frac{m_{polymer}^0 + m_{water}(t)}{m_{polymer}^{dry}} \quad \text{Eq 1}$$

#### 19 2.4 Synthesis of PAA brushes

20  
21 The synthesis of PAA brushes on silicon wafers using the “grafting onto” approach has been described  
22 in more details in a previous paper<sup>19</sup>. Briefly, a self-assembled monolayer of 3-  
23 glycidoxypropyltrimethoxysilane (GPS) was first formed. In a second step, the grafting of poly(*tert*-  
24 butyl acrylate) (PtBA) chains was obtained after the esterification of PtBA-COOH with surface-  
25 attached GPS. In the final step, the PtBA brushes were converted into PAA brushes by a pyrolysis  
26 reaction. The PAA brushes we prepared for our study of adherence as a function of immersion time,  
27 were 6 nm-thick in the dry state with a grafting density of 0.15 chains/ nm<sup>2</sup> and a degree of  
28 polymerization DP ~ 300.  
29  
30  
31  
32  
33

#### 34 2.5 Mechanical Characterization

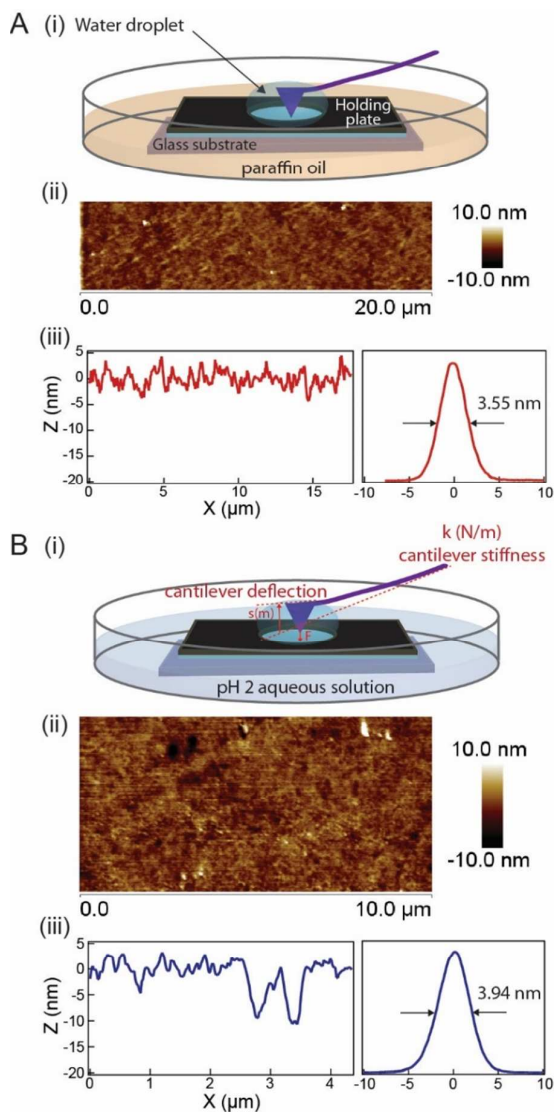
35  
36  
37  
38 Uniaxial compression tests were performed on PDMA hydrogel cylinders with an Instron model 3340  
39 fitted with a 10 N load cell in both preparation and equilibrated states. The preparation state gels were  
40 10 mm-high cylinders with an 8 mm-diameter. The samples were coated with paraffin oil both to  
41 avoid water evaporation and to lubricate the interface between the gel and the testing plates during the  
42 uniaxial compression. The compression tests were performed at a constant displacement rate of 50  
43  $\mu\text{m}\cdot\text{s}^{-1}$  corresponding to  $\dot{\lambda} = 0.05 \text{ s}^{-1}$ . Swollen samples were tested after a week of swelling in a pH 2  
44 aqueous solutions (the pH was adjusted using hydrochloric acid) that were changed every day.  
45 Dimensions of the swollen gel samples during testing depend on their swelling ratio; for each swollen  
46 cylinder, the height and diameter were measured with a caliper before testing and used for the data  
47 treatment.  
48  
49  
50  
51  
52  
53

#### 54 2.6 Characterization of hydrogel free surfaces by AFM

55  
56 Atomic Force Microscopy measurements in water (at 25°C) were performed on a ICON Microscope  
57 equipped with a Nanoscope V controller (Bruker). Both out-of-equilibrium and swollen P20-CL3 gels  
58  
59



were investigated. The probe used for the measurement was a pyramidal silicon nitride nanometric tip of 20 nm radius and 6  $\mu\text{m}$  height. The QNM (Quantitative NanoMechanics) was used for both conditions (out-of-equilibrium and swollen states) with a cantilever stiffness of 0.1 N/m (Figure 1). The setpoint force was 2 nN, the scanning frequency was 1 Hz and the oscillation frequency of the cantilever was 1 kHz. The gel was held by a Teflon coated plate with a centered circular hole, preventing sliding of the gel during the acquisition. For equilibrium conditions, the swollen hydrogel was supported on a glass substrate and surrounded by a pH 2 acid solution in a Petri dish (Figure 1b). In the preparation state, the gel was surrounded by paraffin oil to avoid water evaporation (Figure 1a). In both cases, the contact between the cantilever and the gel surface was established thanks to a Milli-Q water droplet deposited at the interface.



**Figure 1:**

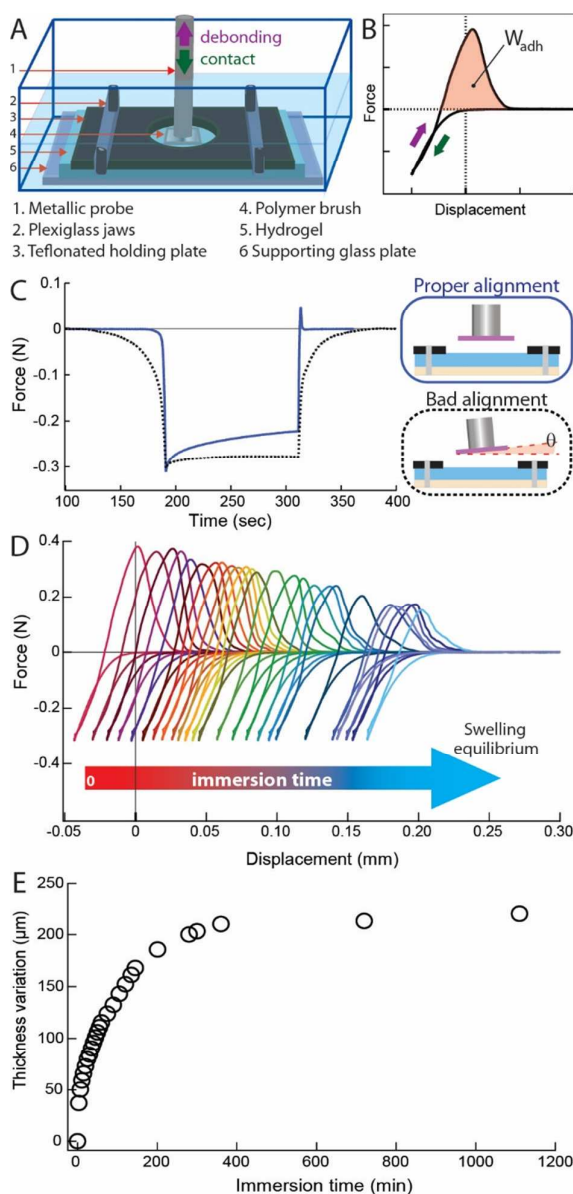
**Characterization of the free surface of the gel by Atomic Force Microscopy (AFM).** The AFM experiments were performed on a P20-CL3 hydrogel in water using a cantilever stiffness of 0.1 N/m. The schemes of the setup are illustrated for the preparation state (A) and in pH 2 aqueous solution (B). Also are displayed the AFM images in QNM modes (ii) and the corresponding height profile and histogram with the mean roughness (iii).

## 2.7 Adherence testing

To perform the adherence tests we have used an experimental setup based on the flat–flat contact test and specifically designed by Sudre et al.<sup>11</sup> to characterize the underwater adherence of soft materials. The full description of this device has been detailed in a previous paper, and we only recall here the main elements necessary for a proper understanding of our present work. The general principle is the same as that of a regular probe test, with some modifications to address the needs of working underwater with swollen hydrogels. Hence, the setup was designed to be able to work both in air (for alignment purposes) and in immersed conditions. The temperature and pH of the liquid medium are easily adjustable *in situ* and measurable without disturbing the test. A commercial Instron machine model 3340 was used for all the adherence tests. The load cell of 10 N is sensitive enough to measure very small forces reproducibly. The noise level of the load cell is on the order of 0.1 mN.

To conduct the adherence test, we used a planar silicon wafer of 1 x 1 cm<sup>2</sup> glued on the flat part of a stainless steel cylindrical punch with a polystyrene solution (45 wt% in toluene) and functionalized on its smooth front side with a PAA brush. The punch was fixed to the load cell with a mandrel and could be pulled off at a constant velocity. During the experiment, the displacement of the punch and the force applied on it are measured as a function of time.

To perform measurements on non-equilibrated hydrogels during swelling, we developed a new sample-holder system, efficiently maintaining the sample from the preparation state to its swelling equilibrium (Figure 2A). A layer of soft hydrogel is laid on a glass substrate and mechanically held by a Teflon-coated aluminum plate, presenting a circular opening of 18 mm of diameter and clamped with Plexiglas jaws in the trough of the initial setup (Figure 2). As the diameter of the hole is large enough, the mechanical holding has no significant consequences on the swelling of the maintained gel compared to the swelling of a free sample. The modified hard substrate attached on the mobile punch is moved down to come into contact with the soft material and subsequently removed at a constant velocity.



**Figure 2: Underwater adherence test between PDMA hydrogel and PAA brush.** (A) Illustration of the setup used to mechanically hold the hydrogel layer with a Teflon-coated plate during the adherence test. (B) Typical force-displacement curves obtained during an adherence test and used to calculate the energy of adherence  $W_{adh}$  (red area). (C) Consequences of the alignment between the hydrogel and the polymer brush on the force response in aqueous solution during the contact and the detachment steps for a correct alignment (—) and for a misalignment (---). (D) Force-displacement data obtained from the kinetic study of adherence of **P10-CL2** in pH 2 aqueous conditions at  $T = 21^\circ\text{C}$  from the preparation state to the swelling equilibrium of the hydrogel. Each force-displacement curve corresponds to a different immersion time point of the gel. (E) Increase of thickness of **P10-CL2** hydrogel during the adherence test in pH 2 aqueous conditions at  $T = 21^\circ\text{C}$  as a function of the immersion time.

The reproducibility and therefore the quantitative aspect of the adhesive measurements are dependent on the quality of the alignment between the hydrogel and the wafer surface. This alignment has to be done in air (to see the contact) and can be finely tuned with the help of three micrometric screws placed under the device. The punch was approached to the gel surface and the contact was observed for a contact force lower than the values used for the adherence tests. If properly aligned, the two

1 materials come into contact, with a contact area equal to the area of the silicon wafer. In that case, the  
2 molecular adhesion at the interface causes a peak of force that appears during the detachment step. In  
3 the case of misalignment, where the angle between the two surfaces  $\theta$  is significant, we observed a  
4 lack of adhesive response of the system during the detachment of the punch; the parts of the punch  
5 taking off that could contribute to the global adhesive force are offset by those still under compression.  
6 An alternative way to check the alignment is the time required to reach a set compressive force. When  
7 punch and gel are properly aligned, the time is at a minimum (less than 4 seconds for an approach  
8 speed of  $10 \mu\text{m/s}$  in our case). We considered that the alignment was adequate when the contact was  
9 complete for a stress lower than 20% of the contact stress applied during the actual adherence test (see  
10 Figure 2C).

11 After adjusting the alignment, the setup was immersed in an aqueous solution at pH 2 (at  $21^\circ\text{C}$ ) and  
12 the immersion time was monitored. After 3 minutes of immersion ( $t_{imm}^1$ ), a first measurement at the  
13 preset experimental conditions (e.g. with a compressive contact stress of 0.3 N and a contact time of 2  
14 minutes fixed for all the adhesive tests unless explicitly mentioned) was carried out. This measurement  
15 corresponds to the minimum dilution of the PDMA hydrogel immersed underwater (i.e. initial out-of-  
16 equilibrium state). The speed at which the probe moves during the compression and detachment stage  
17 has been set respectively at  $10 \mu\text{m}\cdot\text{s}^{-1}$  and  $100 \mu\text{m}\cdot\text{s}^{-1}$  for all the cycles of contact-detachment that have  
18 been repeated regularly during 16 hours (see Figure 2D) as the gel sample swells and increases in  
19 thickness (Figure 2E). The last measurement of adherence has been performed when the PDMA  
20 hydrogel has reached its swelling equilibrium.

21 Following the convention used for soft adhesives, the adherence energy is obtained from the raw data  
22 according to Equation 2 and the scheme in Figure 2B.

$$W_{adh} = \frac{1}{A} \int_{x>x_0} F(x) \cdot dx \quad \text{Eq 2}$$

23 where A is the contact area between the brush and the hydrogel and F is the force needed to apply to  
24 completely detach the brush from the surface of the PDMA gel.

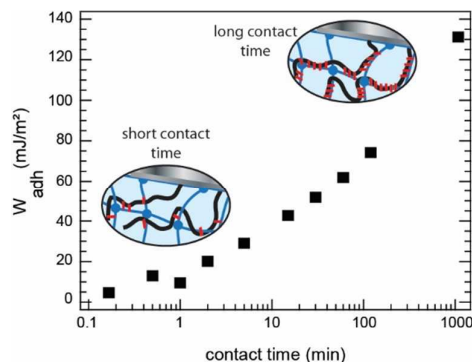
### 25 **3. RESULTS**

#### 26 *3.1 Effect of contact time on adherence*

27 Before we report the changes in adherence as the gel swells, we need to address the question of the  
28 effect of contact time on adherence in equilibrium conditions. For this purpose, adherence tests were  
29 carried out between a fully equilibrated **P10-CL2** gel and a PAA brush for different contact times  
30 (from 10 seconds to 17 hours). The results of these tests are presented on Figure 3 and show a  
31 progressive increase of  $W_{adh}$  with contact time. Given the thickness of the swollen polymer brush ( $\sim 12$

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18

nm), the time needed for the complete diffusion of the grafted chains inside the hydrogel do not exceed a few milliseconds and cannot be responsible for such a slow dependence of  $W_{adh}$  with contact time.



19  
20  
21  
22  
23

**Figure 3** : Energy of adherence as a function of the contact time between the PAA polymer brush and the surface of a **P10-CL2** PDMA hydrogel swollen at equilibrium in an aqueous solution at pH 2.

24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43

In this system, the energy of adherence between the two surfaces is due to the formation of hydrogen bonds between the PAA brush and the PDMA hydrogel. At short contact times, the measured energy of adherence is quite small ( $W_{adh} \sim 0$  for  $t_c = 10$  sec). This suggests that while individual hydrogen bonds may form quite rapidly, they also have a very short lifetime (few picoseconds). Increasing the contact time enhances the probability to create multiple hydrogen bonds that now have an increasingly longer lifetime and are increasingly stable. In turn, the creation of multiple hydrogen bonds limits the mobility of the polymer in the brush and in the hydrogel so that further increase in multiple bonds slows down logarithmically. The energy of adherence of the equilibrated system is only sensitive to the more stable bonds and increases continuously with contact time to reach, after 17 hours of contact, a level of adherence  $W_{adh} \sim 130 \text{ mJ.m}^{-2}$ . For the swelling equilibrium we needed to use a contact time long enough to measure some adherence but significantly shorter than the equilibrium swelling time and we picked 120 seconds.

### 44 3.2 Effect of immersion time on the adherence between hydrogel and brush

45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Now that the effect of contact time has been addressed, the evolution of the energy of adherence between a polymer brush and a hydrogel was determined during its progressive swelling from its preparation conditions to equilibrium into water. To check the methodology, we first performed the test with a 1.8 mm thick **P10-CL2** hydrogel layer in its preparation state in contact with a 12 nm thick swollen PAA brush. The physical adhesion at the interface of the brush and the hydrogel is provided at pH 2 by the formation of multiple hydrogen bonds between the chains of PDMA and PAA, through their hydrophilic sites<sup>11</sup>.

To measure the evolution of the energy of adherence of the out-of-equilibrium system, cycles of compression/traction between **P10-CL2** gel and the PAA brush have been repeated every 3 minutes during the first hour of immersion and then every 30 min/1hr during the next sixteen hours, applying the test parameters defined in the experimental section, i.e. a compression force of 0.3 N, contact time of 120 seconds and a debonding velocity of 100  $\mu\text{m/s}$ . The origin of the displacement for the set of experiments has been defined as the contact point between the two surfaces at zero-force before immersion. The raw data obtained from this study, representing the force applied on the gel as a function of the displacement of the probe (Figure 2D), have been used to calculate the energy of adherence of the system ( $W_{adh}$ ) for each cycle as a function of immersion time ( $t_{imm}$ ) (Figure 4). The evolution of the position of the first contact point at zero-force between the two surfaces during the test series is directly related to the increase in thickness of the **P10-CL2** gel as it progressively swells ( $T = 21^\circ\text{C}$ ,  $\text{pH } 2$ ) (Figure 2E). The change in thickness of the gel has been used to estimate the *in situ* degree of swelling of the **P10-CL2** hydrogel during the adhesive test. In this experiment performed for 5 different **P10-CL2**/PAA systems, the average energy of adherence continuously decreased from 90  $\text{mJ}\cdot\text{m}^{-2}$ , estimated after 3 min of immersion in the aqueous medium (time referred as  $t_{imm}^1$ ), to an average minimal value of 28  $\text{mJ}\cdot\text{m}^{-2}$  after 16 hours of immersion. At this point, the gel is swollen to equilibrium and the energy of adherence measured between **P10-CL2** and the PAA brush, for those contact and debonding conditions remained constant.

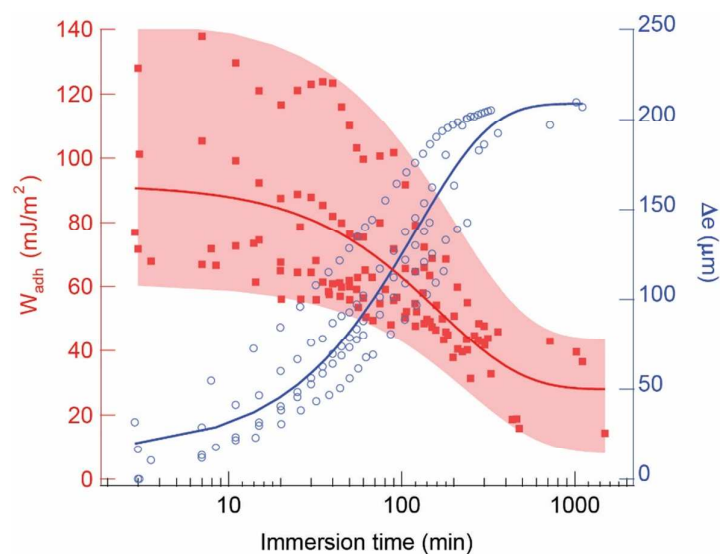


Figure 4: Kinetics of adherence and swelling of **P10-CL2** hydrogels ( $n = 5$ ) from preparation state to swelling equilibrium. On the left axis, the energy of adherence of the system measured as a function of immersion time is represented with the experimental data points ( $\blacksquare$ ), the dispersion ( $\blacksquare$ ) and the best fit of the data points with an arbitrary function to guide the eye ( $\text{---}$ ) ( $W_{adh} = A_0 + A_1 \times \exp(-t_{imm}/\tau_1) + A_2 \times \exp(-t_{imm}/\tau_2)$  with  $A_0 = 27.9 \text{ mJ/m}^2$ ;  $A_1 = 48.3 \text{ mJ/m}^2$ ;  $A_2 = 15.5 \text{ mJ/m}^2$ ;  $\tau_1 = 20 \text{ min}$ ,  $\tau_2 = 90 \text{ min}$ ). On the right axis, the *in situ* variation of thickness of the sample is represented with the different experimental data points ( $\circ$ ) and the fit of the data points ( $\text{---}$ ) ( $\Delta e = B_0 + B_1 \times \exp(-t_{imm}/\tau_3) + B_2 \times \exp(-t_{imm}/\tau_4)$  with  $B_0 = 209 \text{ }\mu\text{m}$ ;  $B_1 = -192 \text{ }\mu\text{m}$ ;  $B_2 = -2 \text{ }\mu\text{m}$ ;  $\tau_3 = 125 \text{ min}$ ,  $\tau_4 = 15 \text{ min}$ ).

1 Simultaneously to the decrease in adherence, the thickness of the hydrogel, mechanically maintained  
2 with the Teflon coated plate, increased by approximately 215  $\mu\text{m}$  corresponding to a swelling of  
3 12% of the unconstrained part of the gel layer, where the contact is made with the polymer brush  
4 (Figure 4). This swelling ratio is very close to what is measured in one direction when the sample is  
5 free to swell in water (in unconstrained conditions Figure S2).  
6  
7  
8

9 The progressive swelling of the hydrogel implies a dilution of the bulk concentration of polymer near  
10 the interface between the PDMA hydrogel and the PAA brush: in the case of the **P10-CL2** hydrogel,  
11 this bulk dilution of the polymer is estimated at 29% from the preparation state to the swelling  
12 equilibrium, and should lead to a decrease of the probability of forming hydrogen bonds between the  
13 two surfaces.  
14  
15  
16

### 17 Evolution of the topography of the PDMA hydrogel

18 During the *in situ* swelling, the mechanical holding of the hydrogel causes a slight macroscopic  
19 curvature of the top surface. The height difference caused by this curvature was estimated to  
20 approximately 10  $\mu\text{m}$  by comparing the indentation measurements of **P10-CL2** in its preparation state  
21 and at swelling equilibrium (Fig. S3).  
22  
23  
24  
25

26 To complete our investigation of the evolution of the topography of the hydrogel once it is immersed  
27 in the aqueous solution and see if it can have an impact on the evolution of the adhesive properties of  
28 the gel, we also performed comparative AFM measurements in the liquid state on the **P20-CL3**  
29 PDMA hydrogels (Figure 1). The difference in surface roughness between the gel in its preparation  
30 state (Figure 1A) and the gel at swelling equilibrium (Figure 1B) is of the order of a few nanometers.  
31 We also noticed the presence of holes, randomly distributed on the surface of swollen **P20-CL3**. The  
32 height of these holes is comprised between 12 and 25 nm and their average diameter is 350 nm. The  
33 same structures have already been observed in the case of polymer thin films. The existence of these  
34 holes at swelling equilibrium is attributed to the partial dissolution of un-crosslinked polymer chains  
35 on the surface of the gel. The reorganization of the network by aggregation of hydrophilic units,  
36 leading to channels formation, helping the water transfer during the swelling, is another explanation  
37 discussed in the literature<sup>20-23</sup>.  
38  
39  
40  
41  
42  
43  
44

45 The probe penetrates about 50  $\mu\text{m}$  deep in the gel during the contact phase. The roughness of a few  
46 nanometers and the curvature of 10  $\mu\text{m}$  of the swollen PDMA are completely flattened during the  
47 contact phase between the brush and the gel, regardless of its swelling state. The changes in  
48 topography observed on the PDMA hydrogel, due to its swelling and its mechanical holding during its  
49 immersion into aqueous medium, are unlikely to be responsible for the observed decrease of the  
50 energy of adherence. This assumption was confirmed by the comparison with the results of the same  
51 kinetic study performed on a **P10-CL2** in its preparation state, maintained by chemical grafting on a  
52 glass substrate which does not involve any deformation of the gel (Figure S4-A).  
53  
54  
55  
56  
57  
58  
59  
60

### Reversibility of the swelling

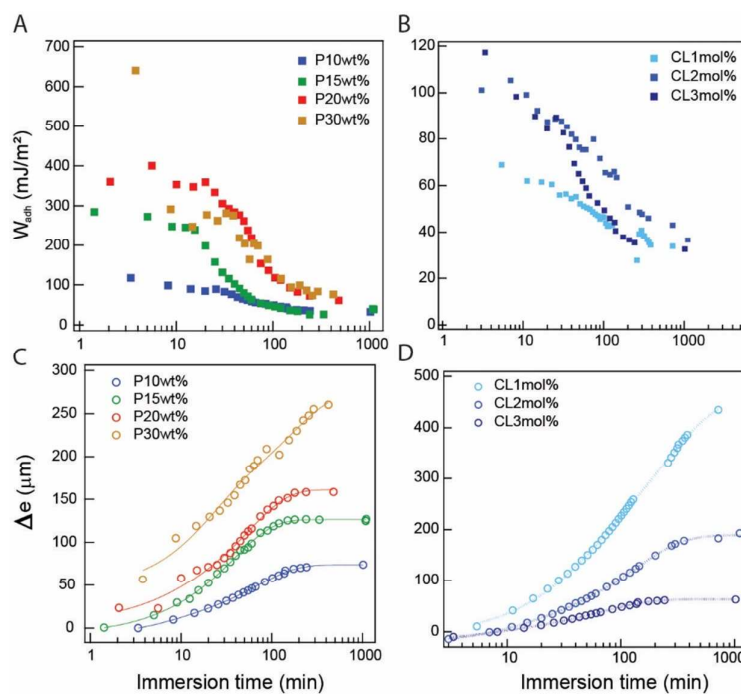
Because the gels in preparation conditions may contain unreacted molecules, we considered the possibility of a decrease in adherence due to a surface contamination or to the polymer degradation by hydrolysis. In order to check this hypothesis, we carefully and slowly dried a gel sample after equilibration to recover the initial degree of swelling in the preparation state. A new kinetic study of adherence during equilibration was then performed on this partially dried sample. The results of the adhesive test showed that after the drying step, the **P10-CL2** hydrogel recovered the same level of adherence than it previously had in its preparation state (Figure S4-B). The evolution of the energy of adherence as a function of immersion time was identical during the second swelling test, and we observed the same decrease of adherence until a plateau value around  $40 \text{ mJ.m}^{-2}$ .

### 3.3 Effect of crosslinks density and polymer concentration on adherence

To investigate the effect of the polymer concentration in the PDMA hydrogel on its adhesive properties with the PAA brush, kinetic studies of adherence and *in situ* swelling of PDMA hydrogels between preparation state and swelling equilibrium were carried out for a series of gels with a fixed monomer concentration (10 wt%) and different crosslinks density (1, 2 and 3 mol% relative to monomer) and for another series with a fixed crosslinking ratio (3 mol% relative to monomer) and different initial concentrations of monomer (10, 15, 20 and 30 wt%). Figure 5 shows the *in situ* swelling kinetics and the change in work of adherence as a function of immersion time for both cases as obtained from the same experimental protocol.

The kinetic studies of adherence performed on **P10-CL<sub>y</sub>** (5B) and **P<sub>x</sub>-CL3** hydrogels in their preparation state (Figure 5A) confirmed the results already observed for **P10-CL2**. During the swelling process, the energy of adherence between the gel and the brush continuously decreases from an initial value corresponding to an out-of-equilibrium state close to the preparation state of the sample, to a minimal plateau value once the swelling equilibrium of the gel has been achieved.





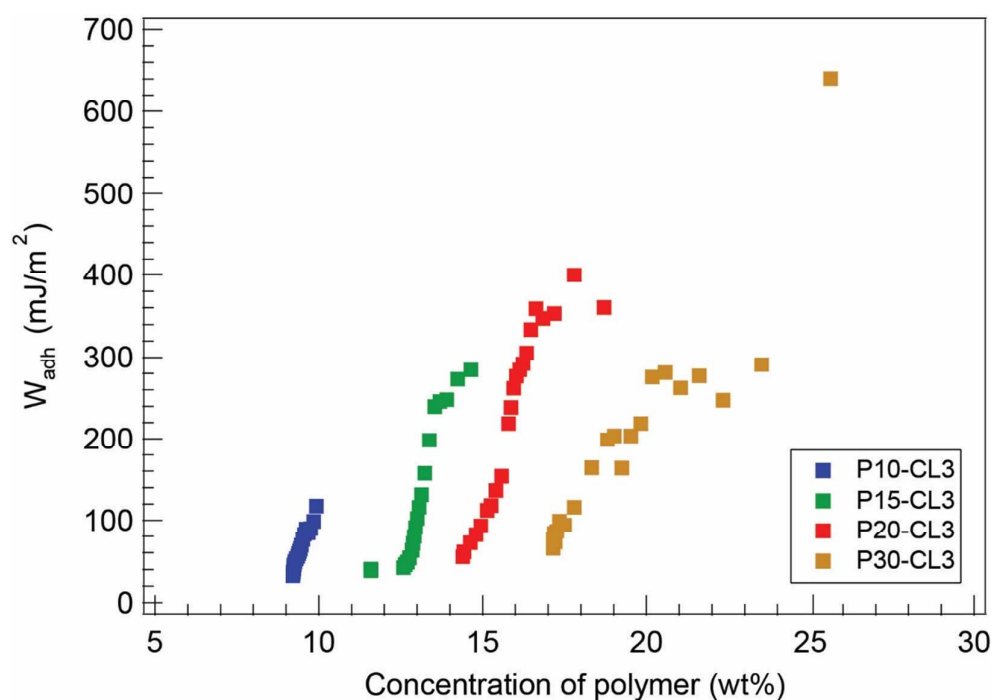
**Figure 5 : Kinetics of adherence and swelling of Px-CLy hydrogels from preparation state to swelling equilibrium.** (A) and (C) Energy of adherence  $W_{adh}$  and variation of the hydrogel thickness as a function of immersion time for different concentrations of polymer in the preparation state (10, 15, 20 and 30 wt%) and a constant crosslinking ratio fixed at 3 mol%. (B) and (D) Energy of adherence and variation of the hydrogel thickness as a function of immersion time for a constant initial concentration of polymer at preparation state of 10 wt% and different crosslinking ratios of 1, 2 and 3 mol%.

At short (2-3 minutes) immersion times (Figure 5A), the energy of adherence of the PDMA hydrogel with the PAA brush increased more than 5 fold (from 117  $\text{mJ}/\text{m}^2$  to 640  $\text{mJ}/\text{m}^2$ ) with increasing initial concentration of monomer in the macroscopic gel (from 10 to 30 wt%). In the same short immersion time conditions, a smaller increase of the energy of adherence was also observed (from 69 to 117  $\text{mJ}/\text{m}^2$ ) with increasing crosslinks density at fixed monomer concentration (Figure 5B). For long immersion times (equilibrium swelling) the same qualitative observations can be made concerning the influence of the concentration of polymer and of the crosslinks density but all values are much lower.  $W_{adh}$  at equilibrium swelling vary from 27  $\text{mJ}/\text{m}^2$  for **P10-CL1** to 55  $\text{mJ}/\text{m}^2$  for **P30-CL3** as summarized on Table 2. It is interesting to note that an equivalent level of adherence than **P10-CL2** after 17 hours of contact (Figure 3), has been reach after 2 minutes of contact by the same gel but in its preparation state (Figure 5A).

PDMA Px-CLy	Out-of-equilibrium initial state ( $t_{imm}^1$ )			Swelling equilibrium state			
	$W_{adh}^{t_{imm}^1}$ ( $\text{mJ}\cdot\text{m}^{-2}$ )	$G_0$ (kPa)	$C(t_{imm}^1)$ (wt%)	Swelling equilibrium ratio $\Lambda_{eq}$	$W_{adh}^{eq}$ ( $\text{mJ}\cdot\text{m}^{-2}$ )	$G_{eq}$ (kPa)	$C_{eq}$ (wt%)
<b>P10-CL1</b>	69	13.5	8.8	16.5	27	11	6.0
<b>P10-CL2</b>	105	26	9.2	13.8	38	23	7.3
<b>P10-CL3</b>	117	36	9.8	10.9	33	37	9.1
<b>P15-CL3</b>	285	79	14.6	8.6	39	84	11.6
<b>P20-CL3</b>	400	127	18.7	6.8	48	122	14.4
<b>P30-CL3</b>	640	232	25.6	5.8	55	192	17.1

**Table 2: Adherence energy and gel composition at the bulk and near the interface in preparation and equilibrium conditions.**  $W_{adh}^{t_{imm}^1}$  refers to the energy of adherence measured at  $t_{imm}^1 \approx 3$  min.  $G_0$  is the shear modulus obtained by compression test at state preparation of the hydrogel and  $G_{eq}$  at swelling equilibrium.  $C(t_{imm}^1)$  is the bulk concentration of polymer in the PDMA hydrogel at  $t_{imm}^1 \approx 3$ min and  $C_{eq}$  is the one at swelling equilibrium.

Given these results one may be tempted to attribute the decrease in adherence energy to the dilution of the polymer in the gel taking place during swelling and therefore to the decrease of the probability to form H-bond interactions between the gel and the PAA brush. It is therefore instructive to plot the adherence energy data, no longer as a function of swelling time but as a function of polymer concentration in the gel. Figure 6 shows the adherence energy as a function of polymer concentration for four different gels. Each color symbol corresponds to a different gel and experiments at all immersion times are compiled. It is immediately obvious from the data that the polymer concentration alone cannot explain the data.



**Figure 6 :** Energy of adherence of **Px-CL3** hydrogels as a function of the concentration of polymer in the hydrogel for 10, 15, 20 and 30 wt%.

A meaningful example is the comparison between **P15-CL3** in its preparation state which has a polymer concentration around 14.4 wt% for  $t_{imm} = 1$  minute and **P20-CL3** in its equilibrium state which has a polymer concentration of 14.6 wt% (Table 2 and Figure 6). The polymer concentration in the network is nearly identical but there is a large difference in adherence energy. The sample in its preparation state has an adherence energy  $W_{adh} = 285$  mJ/m<sup>2</sup>, while the sample at its swelling equilibrium has only  $W_{adh} = 48$  mJ/m<sup>2</sup>.

## 4. DISCUSSION

Let us now try to determine what are the possible parameters influencing the measured adherence energy. An adherence test such as the one performed here can be divided in two stages. In the first stage, the interfacial interactions (hydrogen bonds in our case) between the two surfaces progressively form and we have seen that this process is rather slow at equilibrium (see Figure 3). In a second stage a stress is applied to the interface and the two surfaces are separated. Given the low value of adherence energy measured (well below 1 J/m<sup>2</sup>) and, since our gels are quite elastic (Figure S5), one expects that the dissipated energy is more related to a molecular mechanism such as breakage of the multiple hydrogen bonds formed during the contact than to bulk viscoelastic dissipation. At the interface between the gel and the brush, hydrogen bonds keep forming and breaking based on kinetic mechanisms controlled by the association and dissociation constants of physical bonds. Given the strong dependence of the adherence energy on the contact time one can suppose that the concentration and strength (multiple bonds) of hydrogen bonds formed at the interfaces both increase with time.

For a given bond, the connection between the thermally activated bond constants and the bond scission when a force is applied has been made by Evans in his seminal series of paper on single bond scission as a function of loading rate<sup>24-25</sup>. In his picture, the activation energy for bond scission is modified by a factor  $F\lambda$  when a force is applied. Chaudhury showed in a simplified model targeted at elastomer adhesion, that by applying this time-dependent bond scission to an extensible polymer chain, it is possible to account for the rate-dependent energy storage in the chain before bond breakage<sup>14-15</sup>. He considered a chain tethered permanently at one end and with a labile bond at the other end. When a force is applied, the chain stretches (it is an entropic spring), decreasing the energy barrier for breakage and consequently the probability of breakage. If the polymer chain is stretched at a constant speed  $V$ , the labile bond is also stretched at a constant speed  $V_{bond}$  proportional to  $V$ . As the chain stretching rate increases the loading rate increases as well and the chain can be stretched further before breaking since the probability to break the bond is time and force dependent. In other words as  $V$  increases, the time to break decreases but the force to break (and hence the stored elastic energy at the point of break) increases.

His model predicts for the adherence energy:

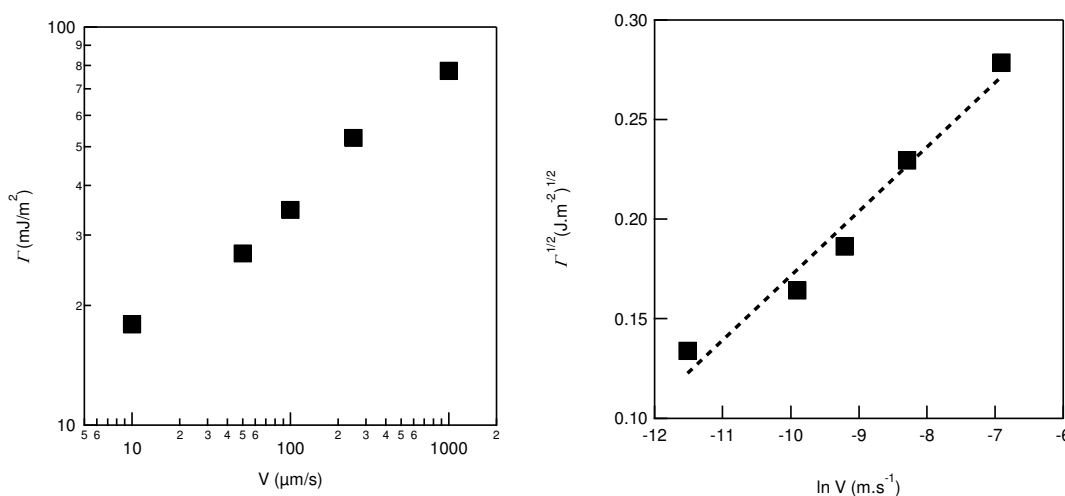
$$W_{adh} = \left( \frac{\Sigma_{HB}}{2k_s} \right) \left[ \left( \frac{kT}{\lambda} \right) \ln \left( \frac{k_s V_{bond} \lambda \tau_{diss}}{kT} \right) \right]^2 \quad \text{Eq.3}$$

where  $\Sigma_{HB}$  is the areal density of hydrogen bonds formed at the interface,  $\lambda$  is the activation length of the labile bond itself (a few nm),  $\tau_{diss}$  is the characteristic time of bond dissociation (depending on the bond strength),  $k_s$  is the spring constant of the polymer chain and  $V_{bond}$  is the pulling velocity on the bond. The term between square parentheses is the velocity dependent force at which the H-bonds break.

If one assumes that the pulling velocity on the bond is proportional to the macroscopic pulling velocity, this equation predicts that for a given interface and time of contact:

$$W_{adh} \sim (\ln V)^2 \quad \text{Eq.4}$$

which is in reasonable agreement with our experimental data for the adherence of **P10-CL2** from a PAA brush in equilibrium conditions (**Figure 7b**). The slope of the linear fit of  $\sqrt{W_{adh}}$  as a function of  $\ln V$  is equal to  $0.032 \pm 0.003 \text{ J}^{1/2}\text{m}^{-1}$  and the intercept point is equal to  $0.46 \pm 0.15 \text{ J}^{1/2}\text{m}^{-1}$  with a correlation coefficient  $R^2 = 0.984$  and the probe velocity in m/s. Hence experimentally, the faster the polymer chains are loaded, the higher is the force where the H-bond scission occurs and therefore the elastic energy dissipated during this bond scission process is higher.



**Figure 7** : Left: energy of adherence as a function of detachment speed of the polymer brush from the surface of **P10-CL2** hydrogel at its swelling equilibrium. Right: Representation of the square root of the same data as a function of logarithm of detachment speed.

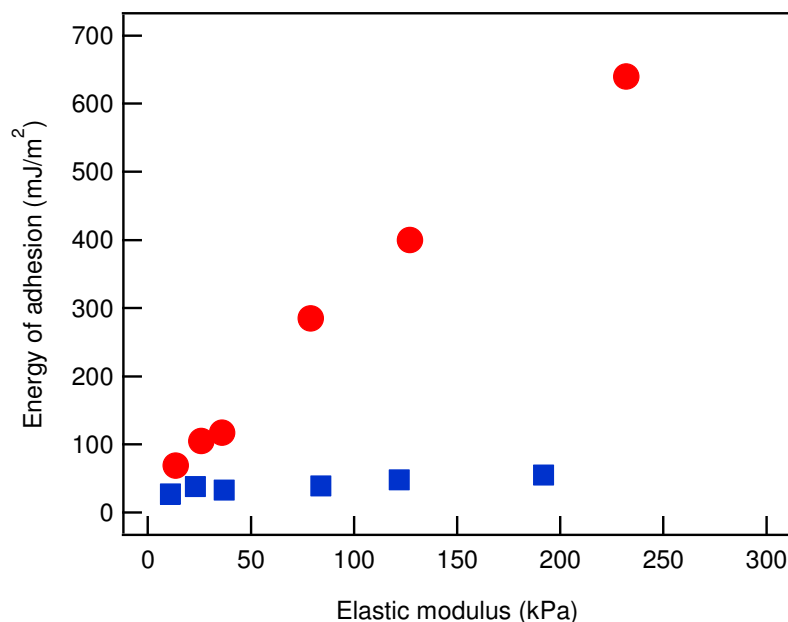
Using equation 3 and our fit of the square root of the energy of adherence as a function of logarithm of detachment speed, we can also estimate the order of magnitude of the dissociating constant of hydrogen bonds in our system. However we have three unknowns :  $\Sigma_{HB}$ ,  $k_s$  and  $\tau_{diss}$  and only two equations. Following Chaudhury<sup>14</sup> we assume that  $\Sigma_b$  cannot be too different from the areal density of strands  $\sim 10^{18} \text{ strands/m}^2$ . This gives then a value of  $k_s = 0.8 \text{ N/m}$  close to the spring constant one would expect for a fully stretched chain<sup>26</sup> (see supplementary information for a more detailed discussion). This reflects the fact that to account for a measurable value of the adhesion energy the chain needs to be stretched before breaking and the bond strength cannot be too low. With this value of  $k_s$  the dissociation time becomes  $\tau_{diss} = 8.7 \cdot 10^{-5} \text{ s}$ . We can then estimate with equation 5 the activation energy of dissociation of hydrogen bonds between the PDMA hydrogel and the PAA brush:

$$\tau_{diss} = \left(\frac{h}{kT}\right) \exp\left(\frac{E_a}{kT}\right) \quad \text{Eq. 5}$$

which for P10-CL2 at its swelling equilibrium after a contact time of 120 seconds gives  $E_a = 20 kT$  which is in a good agreement with what one would expect for multiple hydrogen bonds. It should be noted that for a different contact time the nature and areal density of formed bonds may be different and its activation energy might change.

In principle, equation 3 predicts also a Lake-Thomas like amplification factor of the adherence energy. In the prefactor  $\left(\frac{\Sigma_{HB}}{2k_s}\right)$ ,  $\Sigma_{HB}$  should scale with the concentration of polymer and increase with contact time while the spring constant of the polymer chain  $k_s$  should scale with  $N_0^{-1}$ . This gives a prediction of  $W_{adh} \propto \Sigma_{HB}(t_c)N_0$ , i.e. the adherence energy should scale with the inverse of the modulus of the gel<sup>17, 27</sup>. However, for these weakly adhering systems the energy of the C-C covalent bond which is in the Lake and Thomas model, is replaced by a rate dependent strain energy which results from a balance between spring constant of the polymer strand and labile bond energy<sup>14</sup>.

Figure 8 shows that experimentally the adherence energy increases almost linearly with the modulus. Since the value of the spring constant  $k_s$  in equation 3 has to scale with  $1/N_0$ , this suggests that the concentration and strength of multiple hydrogen bonds formed at the interface represented by  $\Sigma_{HB}$  and  $E_a$  must increase more than linearly with polymer concentration to compensate the spring constant effect.



**Figure 8** : Energy of adherence as a function of elastic modulus  $G$  of the PDMA hydrogel: red symbols are for hydrogels in their preparation state, and blue symbols for hydrogels swollen at equilibrium.

1 In other words, for this system where adhesion is due to the formation of hydrogen bonds, the  
2 adherence energy increases with elastic modulus because more concentrated gels are able to form a  
3 higher concentration of stronger bonds faster.  
4  
5

6  
7  
8 This brings us now back to the question of why does  $\Sigma_{HB}$  for a given contact time changes between  
9 the preparation conditions and the equilibrium conditions.  
10

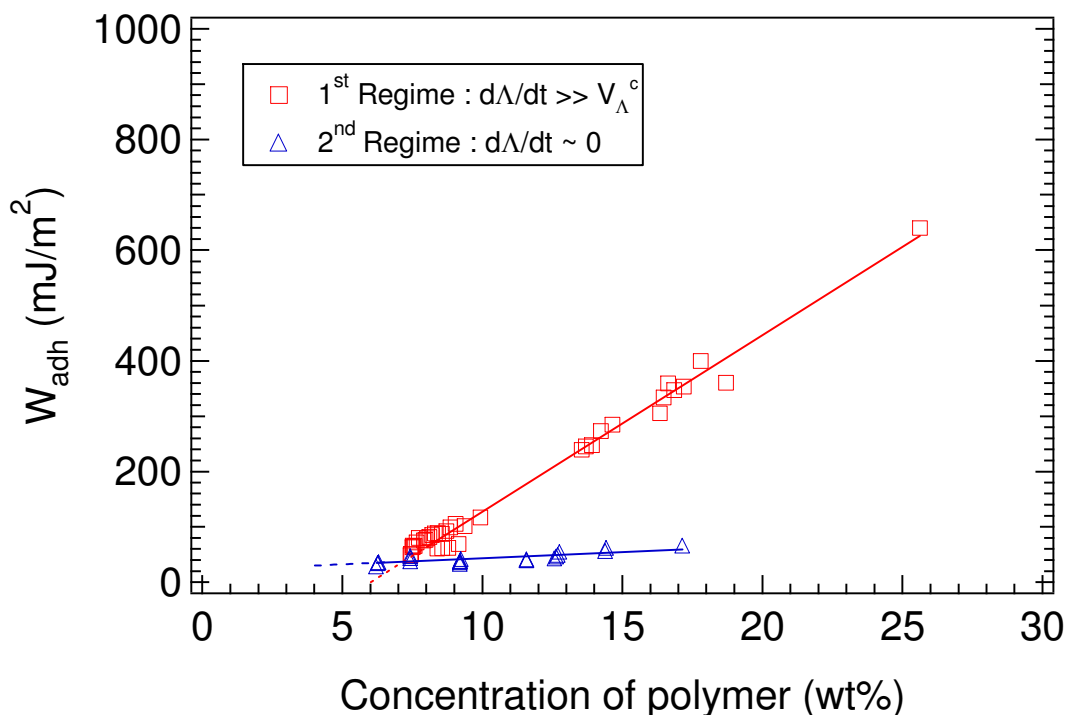
11  
12 It has been argued<sup>28</sup> that because the swelling stretches the polymer chains of the crosslinked network  
13 of the PDMA hydrogel, there is a decrease of degree of freedom for the polymer chains of the gel.  
14 This in turn would increase the entropic cost of forming multiple interactions. This thermodynamic  
15 argument is appealing but cannot explain how a very different adherence can be obtained with only a  
16 very small change in swelling ratio as for the **P10-CL3** hydrogel.  
17  
18

19  
20 An alternative argument relies on kinetics. The swelling rate of the PDMA gel, defined as the  
21 derivative of the swelling ratio  $\Lambda(t)$  as a function of immersion time is:  
22

$$23 \quad V_{\Lambda}(t) = \frac{d\Lambda}{dt} \quad \text{Eq.6}$$

24  
25  
26  
27 If we plot the adherence energy as a function of  $V_{\Lambda}$  (Figure S6), we are able to define for each  
28 composition a critical swelling flux, above which the gel is considered in an out-of-equilibrium state.  
29 We can also define a second regime when the swelling speed is close to zero, where the gel can be  
30 considered in its swelling equilibrium state.  
31  
32

33  
34 Considering those two different regimes, we plotted the adherence energy (for the same contact time)  
35 as a function of the polymer concentration (Figure 9): for both conditions the adherence energy actually  
36 increases linearly with polymer concentration. However a significant difference exists in the  
37 magnitude of the effect. The slope is more than ten times larger in the case of the out-of-equilibrium  
38 state. If the expected dependence of the energy of adherence on the concentration of polymer is indeed  
39 observed, it cannot be the only factor controlling the level of adherence and the swelling state seems to  
40 play an important and significant part to the results of our study.  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

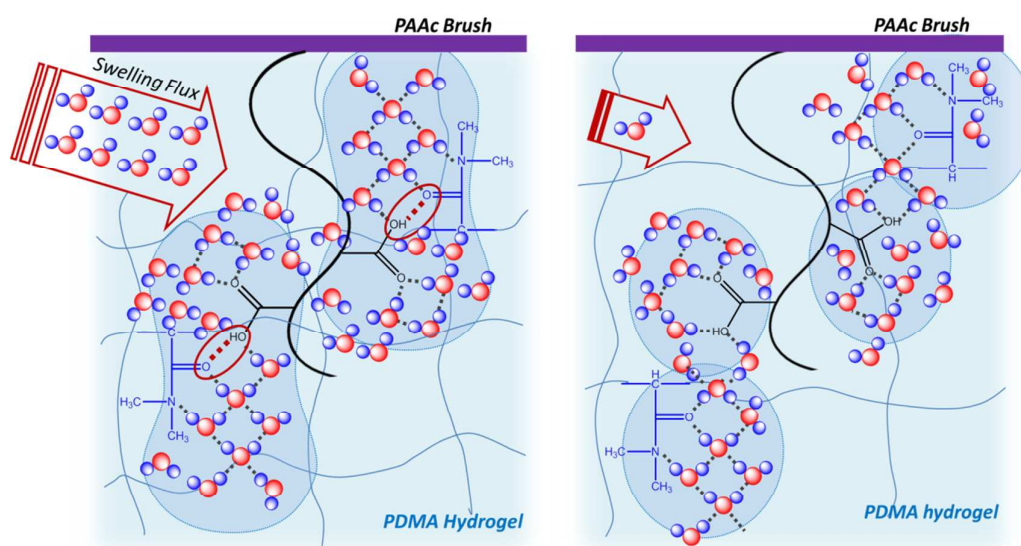


**Figure 9 : Energy of adherence as a function of concentration of polymer for the PDMA hydrogel system.** Two linear regimes are being fitted. The first regime corresponds to the systems that are out-of-equilibrium with a swelling rate above the critical value  $V_{\Lambda}^c$ . The second regime corresponds to the systems that have reached the swelling equilibrium with a swelling rate close to zero.

One hypothesis that could be explored is linked to the non-fouling properties of hydrogels,

It is well-known that because of the hydration of the hydrophilic sites, the water shells formed along the polymer chains make the adherence of proteins through weak interactions impossible, acting like a repulsive barrier<sup>29</sup>. The expulsion of water molecules from both surface and protein is necessary to facilitate protein adsorption by reducing the free energy barrier arising from dehydration entropic effects.

In our case, all the chains are hydrophilic and shells of bound water are organized all around in both states. However, at  $t_{imm}^1$ , the swelling is responsible of a water flux entering the gel. This flux could favor the destabilization of the water shells and increase the replacement and exchange of water molecules. We hypothesize that this flux could accelerate the reorganization of the H-bond interactions, in particular with the merging of hydration shells of hydrophilic sites and the formation of multiple intramolecular interactions between polymer chains, which are responsible of the adherence measured. Once the swelling equilibrium has been achieved, we hypothesize again that the exchange of bound water with free water slows down the probability to form multiple hydrogen between the polymer chains of the brush and the gel decreases. This scenario is drawn schematically in Figure 10.



**Figure 10: Schematic of the gel and brush with their water hydration shell.** We speculate that the presence of a water flux may speed up the formation of hydrogen bonds.

If this hypothesis is correct one expects that the adherence that we measure has a different dependence on contact time if the contact is made in as-prepared or in equilibrated conditions. Therefore comparisons made at the same contact time may give indeed lower values of  $W_{adh}$  equilibrated gels than for dilute gels.

## CONCLUSION

We have carried out systematic adherence experiments (at pH 2) between a neutral PDMA gel and a PAA brush by using a probe tack tester in immersed conditions. Our key finding is that regardless of the composition of the gels (monomer and cross-linker concentration), the measured value of adherence energy systematically decreases with increasing immersion time, i.e; as the gel swells to equilibrium. Interestingly for each specific condition (short or long immersion time) the adherence energy increases linearly with the elastic modulus of the gel i.e. with the bulk density of elastic chains which controls the adhesive interactions at the interface. When we analyze the debonding process however we find two interesting but contradictory results: on one hand for a given gel the adherence energy increases linearly with  $(\ln V)^2$  suggesting that chain stretching occurs before failure, and on the other hand at a given debonding rate the adherence energy increases with the modulus suggesting that the dissipated energy does not depend on the length of the chain being stretched but rather on the density  $\Sigma_{HB}$  of hydrogen bonds formed at the interface.

To reconcile these two results, we propose that the rate of formation of multiple hydrogen bonds at the interface and their activation energy (strength of the bond) may be a highly kinetic effect so that



1  $\Sigma_{HB}(t)$  and  $E_a(t)$  increase faster in the presence of a flux of water entering the gel during the  
2 swelling/deswelling process and with polymer concentration. If this is the case the lower adherence  
3 energy measured at equilibrium conditions (when the flux is low) may simply be due to the slower  
4 kinetics of multiple H-bond formation.  
5  
6  
7  
8  
9  
10

## 11 REFERENCES

- 12 1. Annabi, N.; Tamayol, A.; Shin, S. R.; Ghaemmaghami, A. M.; Peppas, N. A.; Khademhosseini,  
13 A., Surgical materials: Current challenges and nano-enabled solutions. *Nano Today* **2014**, *9* (5), 574-  
14 589.
- 15 2. Andrews, G. P.; Laverty, T. P.; Jones, D. S., Mucoadhesive polymeric platforms for controlled  
16 drug delivery. *Eur J Pharm Biopharm* **2009**, *71* (3), 505-518.
- 17 3. Zhu, F.; Wang, C.; Yang, S.; Wang, Q.; Liang, F.; Liu, C.; Qiu, D.; Qu, X.; Hu, Z.; Yang, Z.,  
18 Injectable tissue adhesive composite hydrogel with fibroblasts for treating skin defects. *Journal of*  
19 *Materials Chemistry B* **2017**, *5* (13), 2416-2424.
- 20 4. Kang, H.-W.; Lee, S. J.; Ko, I. K.; Kengla, C.; Yoo, J. J.; Atala, A., A 3D bioprinting system to  
21 produce human-scale tissue constructs with structural integrity. *Nat Biotechnol* **2016**, *34*, 312.
- 22 5. Yuk, H.; Zhang, T.; Lin, S.; Parada, G. A.; Zhao, X., Tough bonding of hydrogels to diverse non-  
23 porous surfaces. *Nat Mater* **2016**, *15* (2), 190-196.
- 24 6. Tang, J.; Li, J.; Vlassak, J. J.; Suo, Z., Adhesion between highly stretchable materials. *Soft*  
25 *Matter* **2016**, *12* (4), 1093-1099.
- 26 7. Li, J.; Celiz, A. D.; Yang, J.; Yang, Q.; Wamala, I.; Whyte, W.; Seo, B. R.; Vasilyev, N. V.; Vlassak,  
27 J. J.; Suo, Z.; Mooney, D. J., Tough adhesives for diverse wet surfaces. *Science* **2017**, *357* (6349), 378-  
28 381.
- 29 8. Jiawei, Y.; Ruobing, B.; Zhigang, S., Topological Adhesion of Wet Materials. *Adv Mater* **2018**, *0*  
30 (0), 1800671.
- 31 9. La Spina, R.; Tomlinson, M. R.; Ruiz-Perez, L.; Chiche, A.; Langridge, S.; Geoghegan, M.,  
32 Controlling network-brush interactions to achieve switchable adhesion. *Angew. Chem.-Int. Edit.*  
33 **2007**, *46* (34), 6460-6463.
- 34 10. Alfahid, L.; Seddon, W. D.; Williams, N. H.; Geoghegan, M., Double-network hydrogels  
35 improve pH-switchable adhesion. *Soft Matter* **2016**, *12* (22), 5022-5028.
- 36 11. Sudre, G.; Olanier, L.; Tran, Y.; Hourdet, D.; Creton, C., Reversible adhesion between a  
37 hydrogel and a polymer brush. *Soft Matter* **2012**, *8* (31), 8184 - 8193.
- 38 12. Lin, W. C.; Fan, W.; Marcellan, A.; Hourdet, D.; Creton, C., Large Strain and Fracture  
39 Properties of Poly (dimethyl acrylamide)/silica Hybrid Hydrogels. *Macromolecules* **2010**, *43*, 2554-  
40 2563.
- 41 13. Rose, S.; Dizeux, A.; Narita, T.; Hourdet, D.; Marcellan, A., Time Dependence of Dissipative  
42 and Recovery Processes in Nanohybrid Hydrogels. *Macromolecules* **2013**, *46* (10), 4095-4104.
- 43 14. Chaudhury, M. K., Rate-Dependent Fracture at Adhesive Interface. *The Journal of Physical*  
44 *Chemistry B* **1999**, *103* (31), 6562-6566.
- 45 15. Ghatak, A.; Vorvolakos, K.; She, H.; Malotky, D. L.; Chaudhury, M. K., Interfacial rate  
46 processes in adhesion and friction. *Journal of physical chemistry B* **2000**, *104*, 4018-4030.
- 47 16. Merkel, R.; Nassoy, P.; Leung, A.; Ritchie, K.; Evans, E., Energy landscapes of receptor-ligand  
48 bonds explored with dynamic force spectroscopy. *Nature* **1999**, *397*, 50-53.
- 49 17. Lake, G. J.; Thomas, A. G., The strength of highly elastic materials. *Proceedings of the Royal*  
50 *Society of London, series A: Mathematical and Physical Sciences* **1967**, *A300*, 108-119.
- 51 18. Sudre, G.; Tran, Y.; Creton, C.; Hourdet, D., pH/Temperature control of interpolymer  
52 complexation between poly(acrylic acid)  
53  
54  
55  
56  
57  
58  
59  
60

- 1 and weak polybases in aqueous solutions. *Polymer* **2012**, *53*, 379-385.
- 2 19. Sudre, G.; Siband, E.; Hourdet, D.; Creton, C.; Cousin, F.; Tran, Y., Synthesis and
- 3 Characterization of Poly(acrylic acid) Brushes: "Grafting-Onto" Route. *Macromolecular Chemistry and*
- 4 *Physics* **2012**, *213*, 293-300.
- 5 20. Tani, K.; Fujiyoshi, Y., Water channel structures analysed by electron crystallography.
- 6 *Biochimica et Biophysica Acta (BBA) - General Subjects* **2014**, *1840* (5), 1605-1613.
- 7 21. Kim, S. H.; Marmo, C.; Somorjai, G. A., Friction studies of hydrogel contact lenses using AFM:
- 8 non-crosslinked polymers of low friction at the surface. *Biomaterials* **2001**, *22* (24), 3285-3294.
- 9 22. Kim, S. H.; Opdahl, A.; Marmo, C.; Somorjai, G. A., AFM and SFG studies of pHEMA-based
- 10 hydrogel contact lens surfaces in saline solution: adhesion, friction, and the presence of non-
- 11 crosslinked polymer chains at the surface. *Biomaterials* **2002**, *23* (7), 1657-1666.
- 12 23. Beines, P. W.; Klosterkamp, I.; Menges, B.; Jonas, U.; Knoll, W., Responsive Thin Hydrogel
- 13 Layers from Photo-Cross-Linkable Poly(N-isopropylacrylamide) Terpolymers. *Langmuir* **2007**, *23* (4),
- 14 2231-2238.
- 15 24. Evans, E.; Ritchie, K., Dynamic Strength of Molecular Adhesion Bonds. *Biophys J* **1997**, *72*,
- 16 1541-1555.
- 17 25. Evans, E. A.; Calderwood, D. A., Forces and bond dynamics in cell adhesion. *Science* **2007**, *316*
- 18 (5828), 1148-1153.
- 19 26. Hui, C. Y.; Tang, T.; Lin, Y. Y.; Chaudhury, M. K., Failure of elastomeric polymers due to rate
- 20 dependent bond rupture. *Langmuir* **2004**, *20* (14), 6052-6064.
- 21 27. Akagi, Y.; Sakurai, H.; Gong, J. P.; Chung, U.-i.; Sakai, T., Fracture energy of polymer gels with
- 22 controlled network structures. *The Journal of Chemical Physics* **2013**, *139* (14), 144905.
- 23 28. Rose, S.; Prevoteau, A.; Elziere, P.; Hourdet, D.; Marcellan, A.; Leibler, L., Nanoparticle
- 24 solutions as adhesives for gels and biological tissues. *Nature* **2014**, *505* (7483), 382-385.
- 25 29. Chen, S.; Li, L.; Zhao, C.; Zheng, J., Surface hydration: Principles and applications toward low-
- 26 fouling/nonfouling biomaterials. *Polymer* **2010**, *51* (23), 5283-5293.
- 27
- 28
- 29
- 30
- 31
- 32
- 33
- 34
- 35
- 36
- 37
- 38
- 39
- 40
- 41
- 42
- 43
- 44
- 45
- 46
- 47
- 48
- 49
- 50
- 51
- 52
- 53
- 54
- 55
- 56
- 57
- 58
- 59
- 60