



HAL
open science

Evidence of human leptospirosis cases in a cohort of febrile patients in Bangui, Central African Republic: a retrospective study, 2012–2015

Pierre-Alain Rubbo, Marie-Estelle Soupé-Gilbert, Davy Martial Golongba, Florent Mbombo, Dominique Girault, Emmanuel Nakouné, Jean-Pierre Lombart, Sébastien Breurec, Cyrille Goarant

► To cite this version:

Pierre-Alain Rubbo, Marie-Estelle Soupé-Gilbert, Davy Martial Golongba, Florent Mbombo, Dominique Girault, et al.. Evidence of human leptospirosis cases in a cohort of febrile patients in Bangui, Central African Republic: a retrospective study, 2012–2015. *BMC Infectious Diseases*, 2018, 18 (1), pp.376. 10.1186/s12879-018-3298-z . hal-01856179

HAL Id: hal-01856179

<https://hal.science/hal-01856179v1>

Submitted on 10 Aug 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

RESEARCH ARTICLE

Open Access



Evidence of human leptospirosis cases in a cohort of febrile patients in Bangui, Central African Republic: a retrospective study, 2012–2015

Pierre-Alain Rubbo^{1*}, Marie-Estelle Soupé-Gilbert², Davy Martial Golongba¹, Florent Mbombo¹, Dominique Girault², Emmanuel Nakouné¹, Jean-Pierre Lombart¹, Sébastien Breurec³ and Cyrille Goarant²

Abstract

Background: In spite of a local favorable environment, leptospirosis has never been described in Central African Republic so far mainly because of the weakness of diagnostic tests and differential diagnostic strategy for febrile jaundice cases negative for yellow fever virus. Here we bring a complementary insight to conclusions of Gadia CLB et al. regarding the presence of leptospirosis in Central African Republic in YFV-negative febrile icteric patients.

Methods: Our study included 497 individuals presenting with fever and jaundice but negative for yellow fever infection, retrospectively selected from the national surveillance biobank for yellow fever in Institut Pasteur de Bangui, Central African Republic.

A combination of serological (ELISA, agglutination) and molecular biology techniques (quantitative real-time polymerase chain reaction) was used to identify *Leptospira* or the patient's immune response to the bacteria. Statistical analyses were done using the non parametric Mann-Whitney U test with a 5% statistical threshold.

Results: ELISA test results showed 46 positive serum samples while 445 were negative and 6 remains equivocal. In addition, the reference microscopic agglutination test for leptospirosis diagnostic confirmed that 7 out of 32 samples tested were positive. Unfortunately, all 497 serum samples tested for leptospirosis were negative using the molecular techniques.

Conclusions: Unlike Gadia et al., we confirmed that leptospirosis is circulating in Central African Republic and therefore may be responsible for some of the unexplained cases of febrile jaundice in the country. Thus, leptospirosis needs to be investigated to improve identification of aetiological pathogens. Our study also suggests a need to improve sample transportation and storage conditions.

Keywords: Leptospirosis, Bacteria, Infection, Central African Republic, Sub-Saharan Africa, Surveillance, Emergence

Background

Cases of human leptospirosis have never been described in Central African Republic despite local suitable environmental conditions for disease dissemination and human infection. In addition, among cases suspected of yellow fever, a large majority are negative for Yellow Fever Virus (YFV) highlighting the need to improve

differential diagnostic and identify other pathogens responsible for febrile jaundice, including *Leptospira*. Gadia CLB et al. have recently explored pathogens that could be responsible for febrile jaundice in individuals negative for YFV infection in Central African Republic (CAR).

Their retrospective study highlighted that almost half of the 198 samples tested were positive for at least one pathogen other than YFV [1]. This informative article showed that hepatitis viruses are predominant in YFV-negative Yellow Fever suspicions. It confirmed the

* Correspondence: pierrealainrubbo@gmail.com

¹Institut Pasteur, BP 923 Avenue de l'Indépendance, Bangui, Central African Republic

Full list of author information is available at the end of the article



importance of establishing a differential diagnostic algorithm for patients with febrile jaundice, and of improving the management of such cases by healthcare workers, especially in resources-limited countries, such as in sub-saharan countries where national surveillance of malaria and YFV do not include differential testing of related pathogens. Finally, their study also highlighted the scarcity of reliable data from African countries about the role of pathogens other than YFV and *Plasmodium falciparum* in febrile jaundice cases.

CAR is one of the poorest countries in the world facing a chronic healthcare crisis due to successive political conflicts in the past years. Infections frequently associated to febrile jaundice, such as YFV, malaria, hepatitis or leptospirosis for example, are major public health issues in this country. More importantly, although very few YFV-infected individuals are confirmed each year in the country [2] along the World Health Organization (WHO) epidemiological surveillance program, no public health strategies have been implemented for differential diagnostic of febrile jaundice. Therefore it remains crucial to better evaluate the impact and prevalence of each pathogen associated with febrile jaundice in CAR with the aim to improve therapeutic management of such patients. Here we bring a complementary insight regarding the presence of leptospirosis in Central African Republic in individuals uninfected by YFV.

Methods

We designed a retrospective study using 497 YFV-negative serum samples from individuals with febrile jaundice, and stored in the YFV surveillance program biobank between 2012 and 2015 in Institut Pasteur de Bangui.

All 497 individual samples were tested with the Panbio Leptospira Immunoglobulin M (IgM) Enzyme-linked immunosorbent assay (ELISA) kit (Alere, Jouy-en-Josas, France) according to manufacturer’s instructions.

In addition, a panel of 32 serum samples selected from those previously tested in ELISA were also tested using the reference microscopic agglutination test (MAT) for leptospirosis diagnostic [3] using a 24-serovar panel based on the WHO recommendation [4]. This panel included serovars Australis, Autumnalis, Ballum, Bataviae, Canicola, Panama, Pomona, Pyrogenes, Tarassovi, Celledoni, Cynopteri, Djasiman, Gryppotyphosa, Hebdomadis, Javanica, Louisiana, Mini, Sejroe, Shermani (all from eponymous serogroups) as well as Icterohaemorrhagiae and Copenhageni (serogroup Icterohaemorrhagiae), Hardjo (serogroup Sejroe) and Bratislava (serogroup Australis) and the saprophytic strain Patoc (serogroup Semaranga).

All 497 serum samples were also tested for leptospirosis using the LipL32-based quantitative Polymerase Chain Reaction (qPCR) [5]. A subset was also tested with the RNaseP qPCR as an external control PCR [5]

Table 1 Characteristics of individuals with febrile jaundice but negative when tested for YFV according to results of ELISA IgM tests for leptospirosis

Characteristics	Positive Leptospirosis IgM ELISA test		Negative Leptospirosis IgM ELISA test		Equivocal Leptospirosis IgM ELISA test		P value*	
	No	%	No	%	No	%		
Age (years)	0–9	6	14.3	190	43.8	3	14.3	0.07
	10–19	13	31.0	92	21.2	8	38.1	0.86
	20–29	13	31.0	77	17.7	5	23.8	0.17
	30–39	4	9.5	35	8.1	2	9.5	0.35
	40–49	2	4.7	27	6.2	2	9.5	0.76
	> 50	4	9.5	13	3.0	1	4.8	0.36
Time interval between symptoms and serum sampling (days)	< 1	0	0.0	6	1.4	1	4.8	NA
	1–4	14	33.3	180	41.5	7	33.3	0.09
	5–10	28	66.6	247	56.9	13	61.9	0.26
	> 10	0	0.0	1	0.2	0	0.0	NA
Time interval between serum sampling and lab storage (days)	< 1	7	16.7	120	27.6	3	14.3	NA
	1–4	25	59.5	227	52.3	13	61.9	0.53
	5–10	9	21.4	71	16.4	3	14.3	0.34
	> 10	1	2.4	16	3.7	2	9.5	0.35
	Total	42	100	434	100	21	100	

*Comparison was done using the nonparametric Mann-Whitney U test between negative and positive Leptospirosis IgM ELISA tests. Statistical significance was assessed at P value < 0.05

as well as with additional molecular detection assays targeting lipL32 [6] or the 16S rRNA gene [7].

Statistical analyses were done using the non parametric Mann-Whitney U test with a 5% significance threshold.

Results

A majority of individuals selected in this study were below 19 years old (62.8%) while a minority of individuals were above 30 years old (18.1%) (Table 1). Only 7 out of the 497 (1.4%) individual samples were sampled within the day of symptoms onset, and only 130 out of the 497 (26.2%) individual samples were received at the laboratory and stored within the same day of serum sampling (Table 1).

Among them, 42 were positive in ELISA with a mean absorbance of 1.76 (range 1.2–4.5), 434 were negative with a mean absorbance of 0.31 (range 0.1–0.8), and 21 were equivocal (i.e. with a value between 0.9 and 1.1) with a mean of 0.97 (range 0.9–1.1). Globally, median age of individuals included in the study was 14 years old (range 0–76). Median age of individuals with a positive ELISA test was 21.5 years, while it was 12 years old for individuals with a negative ELISA test. All the samples with an equivocal ELISA results were retested with the same ELISA kit: 11 samples became negative, 4 became positive and 6 remained equivocal.

Among the 32 serum samples tested in MAT, 9 were positive with a titer ≥ 50 including 7 with a titer ≥ 100 , suggesting the presence of specific antibodies against *Leptospira*. All 7 samples with a titer ≥ 100 in MAT were also positive in ELISA confirming an earlier infection in these individuals. Two samples had titer of 800 (Patoc) and 1600 (Bratislava) suggesting a recent infection and demonstrating a circulation of the disease in the country. The titer of 800 for the non-pathogenic serovar Patoc suggests that the MAT panel used did not include an appropriate representative of the infecting serovar.

Regarding molecular detection of leptospirosis, all 497 serum samples were negative with the LipL32 qPCR. The RNaseP qPCR gave positive amplification, however with late cycle thresholds (Ct), suggesting that the DNA extracts only contained low amounts of DNA.

Discussion

Suboptimal sampling, transportation (from remote field sites and between laboratories) and storage conditions may have led to the degradation of *Leptospira* DNA, leading to a very low sensitivity. Other molecular tests also failed to evidence *Leptospira* DNA in the specimens. Thus, efforts must be made to optimize pre-analytical management of samples, notably the time between sampling on the field and diagnostic test in the laboratory to improve identification of pathogens by limiting the risk

of sample alteration, which may lead to false negative molecular tests for leptospirosis and other pathogens.

Although we failed to detect *Leptospira* DNA by qPCR possibly because of the degradation of bacterial DNA due to time between sampling and testing, serological tests confirmed that leptospirosis is circulating in Central African Republic. This is the first study revealing proofs of circulation of *Leptospira* in Central African Republic. Globally, very few information is available on leptospirosis incidence, distribution and burden in sub-saharan countries [8].

In addition to hepatitis viruses as demonstrated by Gadia et al. [1], *Leptospira* may also be responsible for some of the unexplained cases of febrile jaundice disease identified in the country and would deserve to be investigated within a differential diagnostic strategy, for example within the national surveillance program of YFV.

Conclusions

This study showed that, despite leptospirosis is rarely mentioned, strains of *Leptospira* circulate in Central African Republic. Leptospirosis diagnostic should be included in the toolkit of laboratories when febrile jaundice disease is suspected. If left untreated, the disease may cause organ failure leading to death. Rigorous procedures for pre-analytical management of samples may also improve the capacity of laboratories to identify pathogens such as *Leptospira*.

Abbreviations

CAR: Central African Republic; DNA: DesoxyriboNucleic Acid; ELISA: Enzyme-linked immunosorbent assay; IgM: Immunoglobulin M; MAT: microscopic agglutination test; PCR: polymerase chain reaction; qPCR: quantitative polymerase chain reaction; WHO: World Health Organization; YFV: Yellow Fever virus

Acknowledgements

We thank local health authorities of Central African Republic as well as the Direction of Institut Pasteur International Network for constant support. We also thank the WHO and non governmental organizations for transporting samples to the laboratory in Institut Pasteur de Bangui. We also grateful to all the participants in this study.

Availability of data materials

The datasets analysed during the current study are available from the corresponding author on reasonable request.

Funding

This work was supported by the Institut Pasteur International Network (AICP grant 2012-A22) and Institut Pasteur of Bangui. National surveillance of yellow fever is funded by the WHO.

Authors' contributions

PAR and CG contributed to design and coordinate the study and the biological analysis, to collect, analysed and interpreted the data, as well as to the writing, review and edition of this manuscript. MESH, DMG, FM and DG run the biological tests, contributed to collect the data and participated to the analysis and interpretation of the data. EN and JPL participated in discussions related to the study, and to the review of this manuscript. SB obtained the funding, contributed to the design of the study, participated to run the experiments, as well as to the review of the manuscript. All authors approved the final version of the manuscript.

Ethics approval and consent to participate

The Central African Republic scientific committee for validation of protocols, results and studies approved the study and allowed utilization and exportation of serum samples stored in Institut Pasteur de Bangui for the ACIP22/2012 relating to the Leptospirosis diagnostic study. The surveillance program for the YFV was approved by the Ministry of Health in Central African Republic and it was subject to mandatory declaration (yellow fever and other haemorrhagic fevers). In this context, patient consent was not necessary. The references of the approval and validation notifications from the Central African Republic scientific committee are 04/UB/FACSS/CSCVPER/17 and 05/UB/FACSS/CSCVPER/17.

Consent for publication

Not applicable.

Competing interests

Pierre-Alain RUBBO is the Founding Chief Executive Officer, and one of the shareholders, of the Omunis start-up company. Omunis did not provide any support in the form of salaries for authors, and did not have any additional role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript. The other authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Institut Pasteur, BP 923 Avenue de l'Indépendance, Bangui, Central African Republic. ²Institut Pasteur, Noumea, New Caledonia. ³University Hospital of Pointe-à-Pitre/Abymes and University of Antilles, Pointe-à-Pitre, Guadeloupe, France.

Received: 28 February 2018 Accepted: 1 August 2018

Published online: 07 August 2018

References

- Gadia CLB, Manirakiza A, Tekpa G, Konamna X, Vickos U, Nakouné E. Identification of pathogens for differential diagnosis of fever with jaundice in the Central African Republic: a retrospective assessment, 2008-2010. *BMC Infect Dis.* 2017;17(1):735.
- Rachas A, Nakouné E, Bouscaillou J, Paireau J, Selekon B, Senekian D, et al. Timeliness of yellow fever surveillance, Central African Republic. *Emerg Infect Dis.* 2014;20(6):1004–8.
- Levett PN. Leptospirosis. *Clin Microbiol Rev.* 2001;14(2):296–326.
- World Health Organization WHO, International Leptospirosis Society ILS. Human Leptospirosis: guidance for diagnosis, surveillance and control. World Health Organization; 2003. <http://apps.who.int/iris/handle/10665/42667>
- Stoddard RA. Detection of pathogenic *Leptospira* spp. through real-time PCR (qPCR) targeting the LipL32 gene. *Methods Mol Biol.* 2013;943:257–66.
- Levett PN, Morey RE, Galloway RL, Turner DE, Steigerwalt AG, Mayer LW. Detection of pathogenic leptospires by real-time quantitative PCR. *J Med Microbiol.* 2005;54(Pt 1):45–9.
- Merien F, Amouriaux P, Perolat P, Baranton G, Saint Gl. Polymerase chain reaction for detection of *Leptospira* spp. in clinical samples. *J Clin Microbiol.* 1992;30:2219–24.
- Allan KJ, Halliday JE, Cleaveland S. Renewing the momentum for leptospirosis research in Africa. *Trans R Soc Trop Med Hyg.* 2015;109:605–6.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

