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1	Sources and spatial distribution of dissolved aliphatic and polycyclic aromatic
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26 Abstract

27 The goal of this work is to investigate the spatial distribution and sources of dissolved aliphatic and polycyclic aromatic hydrocarbons (AHs and PAHs), as well as their 28 relationships with a series of biogeochemical parameters (total chlorophyll a, phaeopigment 29 a, suspended particulate matter, particulate and dissolved organic carbon, and nutrients), in 30 surface coastal waters of the Gulf of Gabès (Tunisia, Southern Mediterranean Sea). Samples 31 32 were collected off the Northern and Southern coasts of Sfax city, as well as in the Gabès-Ghannouch area, from October-November 2014 and in March 2015. Total dissolved AH and 33 PAH concentrations ranged from 0.02 to 6.3 μ g L⁻¹ and from 8.9 to 197.8 ng L⁻¹, respectively. 34 35 These data suggest that this area has been moderate-to-highly impacted by hydrocarbons compared to other Mediterranean coastal environments. The highest AH concentrations were 36 measured off of the Sfax Northern coast (near Ezzit wadi) and in the Gabès-Ghannouch area, 37 38 whereas the highest PAH concentrations were recorded in the Sfax Southern coast (near commercial and fishing harbours and the Sidi Salem channel). Analysis of the AH molecular 39 patterns reveals that low molecular weight (LMW) compounds in these samples are 40 dominated by n-C₁₆, n-C₁₇ and n-C₁₈, which suggests that these samples feature high biogenic 41 42 activity (in the form of bacteria or algae), and that high molecular weight (HMW) compounds display monomodal distributions centred on n-C₂₅ and n-C₂₆, which can be attributed to 43 petrogenic inputs. The PAH molecular patterns are characterized by the dominance of LMW 44 (2-3 rings) compounds and their alkylated homologues, which account for $90 \pm 7\%$ of total 45 PAHs. The observed PAH distribution and isomer ratios thus indicate the presence of major 46 petrogenic contaminations. The concentrations of AHs, PAHs and dissolved organic carbon 47 (DOC) do not correlate with each other, thus demonstrating that these three pools likely 48 originated from different sources and subsequently underwent different processes in the Gulf 49 of Gabès. 50

52 Keywords: Aliphatic hydrocarbons, polycyclic aromatic hydrocarbons, Gulf of Gabès, Mediterranean Sea, anthropogenic signatures, biogenic inputs. 53 54 **1. Introduction** 55 56 Hydrocarbons, including aliphatic hydrocarbons (AHs) and polycyclic aromatic 57 hydrocarbons (PAHs), are among the most ubiquitous organic compounds in marine 58 environments (Lipiatou and Saliot, 1991a; Roose and Brinkman, 2005; Dachs and Méjanelle, 59 60 2010). AHs, which may be defined as straight chained, branched, or cyclic non-aromatic, are herein considered to be comprise a series of resolved compounds (R), including linear *n*-61 alkanes $(n-C_x)$ and two branched *n*-alkanes (isoprenoids), pristane (Pr) and phytane (Phy), as 62

63 well as unresolved complex mixture (UCM). While AHs have several biogenic sources,

64 including higher plant waxes, marine phyto- and zoo-plankton, bacteria and diagenetic

transformations (Cripps, 1989; Rieley et al., 1991; Volkman et al., 1992; Wang et al., 1999),

their anthropogenic sources mainly comprise unburned petroleum/oils (Mazurek and

67 Simoneit, 1984; Bouloubassi and Saliot, 1993; Wang et al., 1997; Readman et al., 2002).

68 PAHs consist of two or more fused benzene rings and are composed of parent compounds and their alkylated homologues. They are synthesized during the formation of oil 69 (petrogenic PAHs), during the incomplete combustion of fossil fuels and biomass (pyrogenic 70 71 PAHs) (Wang et al., 1999; Wurl and Obbard, 2004; Pampanin and Sydnes, 2013), and 72 biologically produced in soils by woody plants and termites or in oceanic environments through microbial degradation of organic matter (Chen et al., 1998; Wilcke et al., 2002). 73 74 Petrogenic PAHs encompass low molecular weight (LMW) 2-3 ring compounds and their alkylated homologues, whereas pyrogenic PAHs encompass high molecular weight (HMW) 75

4-6 ring compounds with a low proportion of alkylated derivatives (Neff, 1979; Wang et al., 76 77 1999). PAHs, which are deleterious to living organisms (Kennish, 1992; Samanta et al., 2002), have long been recognized as high-priority contaminants by both the European Union 78 79 (Official Journal of the EU 24/08/2013, Directive 2013/39/EU) and the United States Environmental Protection Agency (US EPA, 40 CFR Part 423, Appendix A to Part 423). 80 81 The Mediterranean Sea has long been considered to be one of the most polluted seas in the world (Hinrichsen, 1990; UNEP/MAP, 2012). It is believed that anthropogenic AHs and 82 PAHs released from urban, industrial, and agricultural activities may enter Mediterranean 83 waters through rivers (Lipiatou and Saliot, 1991b; Tolosa et al., 1996), surface runoffs, 84 85 effluents (Guigue et al., 2011), atmospheric deposition (Castro-Jiménez et al., 2012) and seabased or shipping-related activities (Tsapakis et al., 2010; Tornero and Hanke, 2016). Because 86 of their low solubility in water, AHs and PAHs are considered to be preferentially associated 87 88 with particles in coastal marine waters (May et al., 1978; Gustafsson et al., 1997; Adhikari et al., 2015). Therefore, most of studies dealing with hydrocarbons in the Mediterranean Sea 89 have focused on sediments and particles, whereas relatively little work has studied the 90 dissolved phase of the water column (Dachs and Méjanelle, 2010; The Mermex group, 2011). 91 However, the dissolved phase is considered as a significant compartment for 92 93 hydrocarbon dynamics in the ocean. First, some studies have shown that the concentrations of hydrocarbons, mostly PAHs, in the dissolved phase are equal to or higher than those in the 94 particulate phase of coastal and open ocean waters (Bouloubassi and Saliot, 1991; Tedetti et 95 96 al., 2010; Berrojalbiz et al., 2011; Guigue et al., 2011; 2015). Second, dissolved PAHs in surface waters represent a relevant compartment to investigate the dynamics of air-water 97 98 fluxes (González-Gaya et al., 2016). Third, dissolved hydrocarbons become bioconcentrated by passive diffusion and respiration and are thus more bioavailable for marine biota than are 99 particulate hydrocarbons, for which bioaccumulation by ingestion occurs within a smaller 100

variety of organisms (i.e. filter feeders) (Tilseth et al., 1984; Akkanen et al., 2012). Finally, in 101 102 the dissolved phase, hydrocarbons are subjected to a variety of processes, such as volatilization (via air-water gas exchanges) (Guitart et al., 2010; González-Gaya et al., 2016), 103 104 biodegradation (Jiménez et al., 2011; Sauret et al., 2016), phototransformation (Al-Lihaibi, 2003; King et al., 2014) and complexation with dissolved organic matter (DOM) (Sabbah et 105 106 al., 2004; Kim and Kwon, 2010), which may in turn modify their bioavailability and thus 107 modify their fate in marine waters (Berrojalbiz et al., 2011; Adhikari et al., 2015). The Gulf of Gabès (Southeastern Tunisia) is one of the most productive ecosystems of 108 the Mediterranean Sea and is the main fishing area of Tunisia (Jabeur et al., 2000; Bel Hassen 109 110 et al., 2009; D'Ortenzio and Ribera d'Alcalà, 2009; The Mermex group, 2011). At the same time, the coastal zone of the Gulf of Gabès has been strongly affected by anthropogenic 111 activity due to the recent significant expansion of the population and the intensification of 112 113 industrial and agricultural activities. These anthropogenic inputs can be attributed to contributions from great urban centres (i.e., Sfax and Gabès cities), fishing and commercial 114 harbours, several phosphoric acid production plants, municipal landfills and wastewater 115 effluents, and industrial and agricultural waste issued from the production and storage of olive 116 oil, chemical products, textiles, soap and paint (Aloulou et al., 2012). The Gulf of Gabès has 117 thus recently become a site of growing interest for investigations of contaminants. Numerous 118 studies have reported the levels of hydrocarbons in sediments (Aloulou et al., 2010; Amorri et 119 al., 2011; Louati et al., 2001; Zaghden et al., 2005; 2007; 2014). Nonetheless, to our 120 knowledge, no data concerning the distribution of dissolved hydrocarbons in surface waters of 121 this ecosystem currently exist. Therefore, the overall objective of this work is to evaluate, for 122 the first time, the origins and spatial distributions of dissolved AHs and PAHs, along with 123 several biogeochemical parameters, in the surface waters of the Sfax and Gabès coastal areas. 124 125

127 2. Materials and Methods

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129 **2.1. Study area**

The Gulf of Gabès, located in the Southeastern region of Tunisia (within the Southern 130 Mediterranean Sea), extends from the city of Chebba (35°14'N, 11°09'E) to the Tunisian-131 132 Libyan border (33°10'N, 11°34'E) and encompasses the Kerkennah and Djerba islands (Fig. 1). It is characterized by several unique geomorphologic, climatic and oceanographic 133 conditions. First, the tidal range is one of the highest within the Mediterranean Sea, due to the 134 135 width (~ 250 km) and very low slope of its continental shelf (Sammari et al., 2006; Hattour et al., 2010). Second its basin is very shallow: in some areas, the water depth does not exceed a 136 few meters over a distance of several kilometres, thus making the Gulf of Gabès very 137 sensitive to atmospheric changes (Hattab et al., 2013) and inducing weak currents, high 138 temperatures (14-29 °C) and high salinities (38-40) in surface waters (Hamdi et al., 2015). 139 140 The climate in the Gulf of Gabès is arid to semiarid (i.e., dry and sunny) with strong easterly winds (INM, 2008), resulting in severe aeolian erosion and the transportation of Saharan dusts 141 into the sea (Jedoui, 1980; Bouaziz et al., 2003). Finally, the Gulf of Gabès is one of the most 142 143 productive areas of the Mediterranean Sea [with a total chlorophyll a concentration (TChl a) that can reach 20 μ g L⁻¹] and accounts for 65% of all Tunisian fish production (Jabeur et al., 144 2000; Bel Hassen et al., 2009; D'Ortenzio and Ribera d'Alcalà, 2009; DGPA, 2010; The 145 Mermex group, 2011). Because the Gulf of Gabès is an essential region for the Tunisian 146 economy, this coastal area has been strongly influenced by anthropogenic activity from 147 various urban, industrial and agricultural outputs for many decades. These activities are 148 particularly pronounced in the two largest cities of the Gulf, Sfax and Gabès (Fig. 1). 149

Sfax (34°43'N, 10°46'E; Fig. 1) is the second-largest city in Tunisia (with 730,000 150 151 inhabitants) and represents its economic centre. The Sfax Northern coast (which extends from the commercial harbour to Ezzit wadi and beyond; Fig. 1) was recently restored through the 152 *Taparura* project. This area features the outlet of the rainwater drain channel ("PK4"), the 153 outlet of the Ezzit wadi, and the "Poudrière" industrial zone (Fig. 1). The Sfax Southern coast 154 extends from the commercial harbour to the southern boundary of the Salina/Thyna 155 agglomeration and containts the "SIAPE" phosphoric acid industry (Fig. 1). Near this industry 156 are also the Thyna municipal landfill and the "ONAS" wastewater treatment plant (WWTP). 157 SIAPE washing water, the WWTP effluent, and the municipal landfill leaching waters are 158 159 released into the sea through a channel that joins the El Hakmouni wadi eastward of the WWTP. Moreover, the Sfax Southern coast encompasses several industrial zones, including 160 "Madagascar", the fishing harbour and "Sidi Salem" (Fig. 1). 161 162 The Gabès-Ghannouch area (33°56'N, 10°03'E; Fig. 1) contains approximately 130,000 inhabitants and is characterized by important industrial and harbour activities. The "GCT-163 Gabès" phosphoric acid industry is situated along the commercial harbour. This area also 164 houses a thermal power plant in the commercial harbour zone, petrochemical and off-shore 165 petroleum infrastructures, and industries related to agribusiness, building materials, energy 166 167 production and textiles. The El-Tine and Gabès wadis receive a variety of urban, industrial and agricultural wastes (Fig. 1) (see Text S1 for complete description of Sfax and Gabès 168 169 coastal areas).

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171 **2.2. Sampling**

Thirty-six stations were sampled in the coastal areas of Sfax (October 2014 and March
2015) and Gabès cities (November 2014) (Fig. 1; Table 1). For each transect, the most coastal
station was located approximately 100 m (S1, S5, S8, S14, S15, S19) or 300 m (S12, S16,

S22, S25, S27, S30, S33, S35) from land. With the exception of S1-S2, S5-S6, and S8-S9, 175 176 which were sampled from a distance of 200 m, all stations were located at 1000 m intervals along each transect (Fig. 1; Table 1). Sampling was performed on board the vessel 177 "Taparura" between 10:30 am and 3:30 pm (18 and 23 October 2014), 8:30 am and 12:30 pm 178 (13 November 2014), and 14 pm and 16.30 pm (8 March 2015). This sampling occurred 179 around high tide and under calm sea and sunny conditions (18 and 23 October, 13 November 180 2014) or in cloudy weather (8 March 2015). Seawater samples were collected at a depth of ~ 181 0.1 m using 4-L Nalgene[®] polycarbonate bottles (ThermoFisher Scientific, Waltham, USA), 182 which were opened below the water surface to avoid sampling of the surface microlayer. 183 184 Before their use, these bottles were extensively washed with 1 M hydrochloric acid (HCl) and ultrapure water (i.e., Milli-Q water from Millipore system, final resistivity: $18.2 \text{ M}\Omega \text{ cm}^{-1}$). 185 They were then rinsed three times with each respective sample before being filled and placed 186 187 in a cold and dark area after collection. On board, in situ measurements of temperature, salinity and pH were obtained using measuring cells with the TetraCon[®] 4-electrode system 188 and a refractometer (WTW, Weilheim, Germany). Temperature and salinity data are not 189 available for stations S1, S5, S8, S14, S15 and S19 (see data in Table 2). 190

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2.3. Filtration, handling and storage of samples

Once in the laboratory, samples were immediately filtered under low vacuum conditions (< 50 mm Hg) through pre-combusted (500 °C, 4 h) GF/F (~ 0.7 μ m) glass fibre filters (25 or 47 mm diameter, Whatman, Fisher Scientific, Illkirch, France) using glassware filtration systems. To analyse dissolved hydrocarbons (AHs and PAHs), 2 L of samples were filtered through 47-mm diameter GF/F filters. Filtered samples were then stored in 2.5 L amber glass bottles with 50 mL of dichloromethane (CH₂Cl₂) at 4 °C, in the dark, before undergoing solvent extraction (see §2.4). It should be noted that these filters were also used for particulate

hydrocarbon analyses, but these results are not presented in this study. For analyses of TChl a 200 and phaeopigment a (Phaeo a), 250-300 mL of samples were filtered through 25-mm 201 diameter GF/F filters. Filters were stored in Nunc[®] cryogenic vials (Dutscher, Brumath, 202 France) at -20 °C until analysis. For analyses of suspended particulate matter (SPM) and 203 particulate organic carbon (POC), between 250 and 1100 mL of sample, depending on their 204 station of origin, were both filtered with pre-weighted 25-mm diameter GF/F filters. After this 205 filtration, filters were dried at 60 °C for 24 h and stored in Petrislides MilliPore[®] (Saint-Louis, 206 USA) until analysis. Filtered seawater issued from this SPM and POC filtration was 207 transferred in 100-mL Schott[®] glass bottles (Fisher Scientific) and in 60-mL HDPE bottles for 208 209 nutrient and dissolved organic carbon (DOC) analyses, respectively. Glass and HDPE bottles were stored at -20 °C until analysis. Nutrient, TChl a, Phaeo a, SPM, and POC analyses were 210 not conducted for samples obtained from S1, S5, S8, S14, S15 and S19, whereas DOC 211 212 analyses were not performed on samples from stations S1, S5 and S14 (see analytical procedures in § 2.7 and data in Table 2). 213

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2.4. Hydrocarbon extraction, purification and instrumental analysis

Dissolved hydrocarbons in the $< 0.7 \mu m$ size fraction were extracted from seawater 216 using liquid-liquid extraction with CH₂Cl₂ (2 x 50 mL per litre), according to the procedure of 217 Guigue et al. (2011; 2014; 2015). These extracts were then concentrated by using a rotary 218 evaporator before changing the solvent to *n*-hexane. Hexane extracts were then purified to 219 separate the hydrocarbon fraction from more polar compounds. Extracts were fractionated 220 within a 500 mg silica column (Silica gel extra pure, Merck, Darmstadt, Germany), which had 221 previously been activated at 500 °C for 4 h and then partially deactivated with ultrapure water 222 (2% w/w). AHs and PAHs were eluted together using 2 mL of *n*-hexane followed by 3 mL of 223

224	<i>n</i> -hexane/CH ₂ Cl ₂ (3:1 v/v) (Guigue et al., 2014; 2015). All solvents were of organic trace
225	analysis quality (Rathburn Chemicals Ltd, Walkerburn, Scotland).
226	Purified hydrocarbon fractions (AHs + PAHs) were analysed by a gas chromatograph-
227	mass spectrometer (GC-MS) (5977A Series GC/MSD System, Agilent Technologies, Santa
228	Clara, USA) running at an ionization energy of 70 eV for a m/z range of 50-400, and using
229	the selected ion monitoring (SIM) mode. The GC-MS used helium at a flow rate of 1.2
230	mL/min as a carrier gas and an HP-5 MS column (30 m x 0.25 mm x 0.25 $\mu m,$ J&W
231	Scientific, Agilent Technologies). The injector (used in splitless mode) and detector
232	temperatures were set at 270 and 320 °C, respectively. The non-linear temperature program
233	used for the column was as follow: 70 °C for 3 min, 15 °C/min (ramp 1) to 150 °C, and then 7
234	°C/min (ramp 2) to a final temperature of 320 °C, held for 10 min. The identification and
235	quantification of hydrocarbons in samples and calibrations were performed in SIM mode
236	(Guigue et al., 2014; 2015).
237	
238	2.5. Quality assurance/Quality control
239	During the experiments and analyses, gloves were worn and care was taken to avoid
240	contamination. All the glassware was washed with 1 M HCl and ultrapure water before being
241	baked at 500 °C for 4 h. All materials that could not be baked were washed with 1 M HCl and
242	ultrapure water and dried at room temperature.
243	AH and PAH external calibration standards (04071, Fluka and 47543-U, Supelco,
244	among others, purchased from Sigma-Aldrich) were run for peak identification, and sample

245 quantifications were processed using internal standards. Deuterated standard mixtures, which

were introduced in samples prior to extraction (n-C₁₆- d_{34} , n-C₂₄- d_{50} , n-C₃₆- d_{74} for AHs, and

247 Naph- d_8 , Phe- d_{10} , Per- d_{12} for PAHs) and prior to injection (Ace- d_{10} , Flu- d_{10} and Chrys- d_{12}),

248 were used to determine procedural recoveries and to quantify hydrocarbons in samples.

Average recoveries of n-C₁₆- d_{34} , n-C₂₄- d_{50} and n-C₃₆- d_{74} were 93% for each standard, while 249 250 those of Naph- d_8 , Phe- d_{10} , Per- d_{12} were 51, 71, and 84%, respectively. These procedural recoveries, which include the processes of extraction, evaporation, and purification, do not 251 take into account the potential losses of HMW dissolved hydrocarbons by sorption onto 252 polycarbonate bottles during the few hours between the sampling and filtration steps, which 253 254 are very difficult to quantify in natural samples (Delle Site, 2001). Blanks were run for the 255 entire procedure, including extraction, solvent concentration, and purification. The total mass of all AH and PAH compounds measured in these procedural blanks represented on average 256 19 and 4% of the total mass of all AH and PAH compounds measured in these samples, 257 respectively. For AHs, the procedural blank was mainly due to $n-C_{23}$, $n-C_{24}$, $n-C_{30}$, and $n-C_{32}$. 258 All concentration values were blank- and recovery-corrected. Detection limits of the method 259 used for individual compounds varied from 1 to 30 pg L^{-1} in water. 260

261

262 **2.6. Determination of hydrocarbon indices and ratios**

For AHs, we determined the concentration of resolved compounds (R) (i.e., *n*-alkanes 263 from $n-C_{15}$ to $n-C_{40}$, including Pr and Phy) and the concentration of the UCM by integrating 264 the hump (if present) using the mean response factors of the resolved compounds. Biogenic 265 266 AHs were distinguished from petrogenic AHs mainly by relying on their molecular pattern distribution and then by determining several ratios and indices. First, we used the R/UCM 267 ratio as an indicator of recent petroleum contamination (when > 0.3) or severe degraded 268 petroleum contamination (when < 0.05) (Simoneit and Mazurek, 1982; Commendatore and 269 Esteves, 2004; Asia et al., 2009; Commendatore et al., 2012). We also used the Pr/Phy ratio 270 as an indicator of biogenic AHs (when > 1) (Cripps, 1989; Commendatore and Esteves, 2004; 271 Cincinelli et al., 2008). However, due to the controversial origin of these isoprenoids, it is not 272 relevant to employ this ratio as an indicator of petrogenic AHs (when it is < 1). Finally, we 273

- used the carbon preference index (CPI₁₅₋₄₀), computed as the odd/even ratio in the n-C₁₅-n-
- 275 C_{40} range, as an indicator of biogenic AHs (CPI₁₅₋₄₀ > 1 or < 1) or AHs issued from petroleum 276 contaminations (CPI₁₅₋₄₀ ~ 1) (Commendatore et al., 2012).
- 277 We quantified 29 PAH compounds (\sum_{29} PAHs_{-P+A}) in these samples. These include 19
- 278 parent PAHs (PAHs_{-P}), which are naphthalene (Naph), acenaphthylene (Acy), acenaphthene
- 279 (Ace), fluorene (Flu), dibenzothiophene (DBT), phenanthrene (Phe), anthracene (Ant),
- 280 fluoranthene (Flt), pyrene (Pyr), benz[a]anthracene [BaA], chrysene (Chr),
- benzo[b]fluoranthene [BbF], benzo[k]fluoranthene [BkF], benzo[a]pyrene [BaP],
- benzo[e]pyrene [BeP], perylene (Per), dibenzo[a,h]anthracene [DA], benzo[g,h,i]perylene
- [BP] and indeno[1,2,3-cd]pyrene [IP]. We also quantified 10 alkylated homologues (PAHs_{-A}),
- including methyl (C_1), dimethyl (C_2) and trimethyl (C_3) of Naph and Phe, and C_1 and C_2 of
- Flu and Pyr. Petrogenic and pyrogenic PAHs were discriminated mainly by relying on their
- 286 molecular pattern distribution and then by determining selected ratios (Lipiatou and Saliot,
- 1991; Budzinski et al., 1997; Mille et al., 2007). First, we used the ratio between alkylated
- homologues and parent compounds (Alk/Par); generally, petrogenic PAHs are characterized
- by the dominance of alkylated homologues over their parent compounds (Alk/Par > 1),
- whereas the inverse pattern is observed for pyrogenic PAHs (Youngblood and Blumer, 1975).
- 291 Second, we used the Ant/(Ant + Phe) and Flt/(Flt + Pyr) ratios, as indicators of pyrogenic (>
- 0.1 and > 0.5, respectively) or petrogenic PAHs (< 0.1 and < 0.4, respectively) (Budzinski et
- al., 1997; Yunker et al., 2002; Asia et al., 2009; Li et al., 2015). It is worth noting that all
- these ratios and indices have to be used with precaution since hydrocarbon presence patterns
- 295 may be substantially modified by many processes.
- 296

297 **2.7. Biogeochemical analyses**

298	Nutrients, such as nitrites (NO ₂ ⁻), nitrates (NO ₃ ⁻), phosphates (PO ₄ ³⁻), and silicates
299	[Si(OH) ₄] were analysed using the automated colorimetric method described in Aminot and
300	Kérouel (2007). To obtain reproducible nutrient measurements, we used standards and
301	compared their results to those of commercial products (OSIL Ltd, Havant, UK). The
302	detection limits of NO ₂ ⁻ , NO ₃ ⁻ , and PO ₄ ³⁻ were 0.05, 0.05, and 0.02 μ M, respectively.
303	TChl <i>a</i> and Phaeo <i>a</i> were analysed using the fluorescence method following a methanol
304	extraction (Raimbault et al., 2004). Five mL of methanol (RP Prolabo, VWR France,
305	Fontenay sous Bois, France) were added to the filter, and, after 30 minutes of extraction in the
306	dark at 4 °C, a fluorescence measurement was performed using a fluorometer from model 10
307	Turner Designs (Sunnyvale, USA) equipped with the Welschmeyer kit to avoid Chl b
308	interference (Welschmeyer, 1994). As the monochromatic fluorescence method cannot
309	separate divinyl chlorophyll from chlorophyll, results are given in terms of total Chl a (TChl
310	a) concentrations, representing the sum of Chl a and divinyl Chl a. The blank "methanol +
311	filter" was close to zero, yielding a very low detection limit of approximately 0.01 μ g L ⁻¹ .
312	Calibrations were made using a pure Sigma Chl a standard. We applied the acidification
313	method to determine Phaeo a concentrations (Raimbault et al., 2004).
314	After spending 24 h in the desiccator, filters were reweighed on the same balance, and
315	the SPM concentration was calculated as the difference between the filter weight before and
316	after sample filtration, normalised to the filtration volume (Neukermans et al., 2012). POC
317	was then determined with an AutoAnalyser II Technicon (New York, USA) using the wet-
318	oxidation procedure established by Raimbault et al. (1999). The POC detection limit was 0.50
319	μΜ.
320	To determine DOC, samples were acidified with orthophosphoric acid and analysed
321	using a TOC-5000 total carbon analyser (Shimadzu, Kyoto, Japan) (Sohrin and Sempéré,

322 2005). Average TOC concentrations in Deep Atlantic Water and low carbon water reference

standards were 43 \pm 2 μ M C, n = 24 and 1 \pm 0.3 μ M C, n = 24, respectively. The analytical precision of the procedure was \leq 2%.

- 325
- 326 **2.8. Statistical treatment**

Colour maps of dissolved AH and PAH concentrations were performed using Ocean 327 Data View (ODV) software version 4.6.5 (Schlitzer, R., http://odv.awi.de, 2014.). The spatial 328 329 interpolation/gridding of data was conducted using Data-Interpolating Variational Analysis (DIVA) (Barth et al., 2010; Troupin et al., 2012). Because all variables (except pH) displayed 330 non-normal distributions and high-amplitude values (Joliffe, 1986), it was preferable to use 331 332 Spearman's rank-order correlation over Pearson's correlation. Hierarchical ascendant classification (HAC) was used to find homogenous groups of individuals (stations) within the 333 dataset, with respect to selected variables. These groups (or clusters) present a low within-334 335 variability and are different than others (displaying high between-variability). The dissimilarity measurement between clusters was based on Ward's method, which uses squared 336 Euclidean distance. Variable values were centred and reduced beforehand. Two different 337 HAC were then applied: one on physical and biogeochemical parameters (i.e., T, salinity, pH, 338 NO_2^{-} , NO_3^{-} , PO_4^{-3-} , Si(OH)₄, TChl *a*, Phaeo *a*, SPM, POC, DOC) and the other one on selected 339 hydrocarbon parameters (i.e., R, Pr/Phy, CPI₁₅₋₄₀, for AHs, \sum_{29} PAHs_{-P+A}, Alk/Par, Flt/(Flt + 340 Pyr) for PAHs). Clusters (groups) issued from these HAC were represented in the form of 341 box-and-whisker plots. Principal component analysis (PCA) was also applied to these 342 hydrocarbon variables, based on Spearman's rank-order correlation matrix. Normality tests 343 (including Shapiro-Wilk, Jarque-Bera and Kolmogorov-Smirnov tests), Spearman's rank-344 order correlation matrices, HAC, and PCA were performed using XLSTAT 2013.5.01 (a 345 Microsoft Excel add-in program), while box-and-whisker plots were completed with 346

347 StatView 5.0. The significance threshold for these different analyses and tests was set at p <
348 0.05.

349

350 **3. Results**

- 351
- 352

3.1. Spatial distribution of physical and biogeochemical parameters

353 Physical and biogeochemical parameters are reported in Table 2. Applying HAC to this dataset reveals the presence of six groups of stations (G1-G6; Fig. 1; Fig. 2a-i). The first 354 group (G1) includes stations S2 and S6, which are located on the Sfax Northern coast, in front 355 356 of Ezzit wadi and between Ezzit wadi and PK4 channel, respectively (Fig. 1). These stations are characterized by the highest observed POC (245 and 208 µM, respectively; Fig. 2h) and 357 SPM concentrations (44 and 59 mg L^{-1} , respectively; Fig. 2i), as well as the highest measured 358 salinity values (~ 42.0), and high TChl a (16 μ g L⁻¹ in S2; Fig. 2e), Phaeo a (3.5 μ g L⁻¹ in S6; 359 Fig. 2f) and DOC concentrations (181 µM in S2; Fig. 2g) (Table 2). G2 includes all other 360 361 stations from the Sfax Northern coast (S3, S4, S7, S9-S13) as well as station S35 from the Gabès-Ghannouch area (Fig. 1). This group is marked by the lowest observed PO_4^{3-} (10.4 ± 362 2.3 μ M; Fig. 2c), TChl a (3.7± 1.4 μ g L⁻¹; Fig. 2e), and Phaeo a concentrations (1.3 ± 0.6 μ g 363 L⁻¹; Fig. 2f) (Table 2). G3 encompasses stations S16 (in front of the fishing harbour), S20 (in 364 365 front of the Sidi Salem channel) and S22 (in front of the El Maou wadi) on the Sfax Southern coast (Fig. 1). These stations yield the highest measured concentrations of NO₂⁻ (0.8 ± 0.1 366 μ M; Fig. 2a), PO₄³⁻ (107.5 ± 22.0 μ M; Fig. 2c), TChl *a* (27.8 ± 13.5 μ g L⁻¹; Fig. 2e), Phaeo *a* 367 $(6.8 \pm 3.4 \ \mu g \ L^{-1};$ Fig. 2f), and DOC (194.8 $\pm 10.7 \ \mu M;$ Fig. 2g), as well as high 368 concentrations of NO₃, POC, and SPM (Fig. 2b, h, i; Table 2). Therefore, it is likely that 369 370 these three stations were subjected to intense eutrophication. G4 includes the remaining stations from the Sfax Southern coast (S17, S18, S21, S23-S26) (Fig. 1), which are marked by 371

372	the lowest concentrations of Si(OH) ₄ (3.1 \pm 1.4 μ M; Fig. 2d), POC (30.7 \pm 12.8 μ M; Fig. 2h),
373	and SPM (10.2 \pm 1.7 mg L ⁻¹ ; Fig. 2i), the highest T values (26.8 \pm 0.3 °C), and the lowest pH
374	values (8.0 \pm 0.05) (Table 2). G5 comprises stations from the Gabès-Ghannouch area (S27-
375	S32, S36) (Fig. 1), which are characterized by the lowest NO ₂ ⁻ concentrations (~ 0. μ M; Fig.
376	2a), the lowest salinity values (34.7 \pm 2.4), and high DOC concentrations, with the highest
377	value for S30 (217.6 μ M; Fig. 2g) (Table 2). Finally, G6 includes stations S33 and S34, which
378	are located between the fishing and commercial harbours of the Gabès-Ghannouch area (Fig.
379	1). They record the highest concentrations of Si(OH) ₄ (27.6-33.4 μ M; Fig. 2d) and NO ₃ ⁻ (3.2-
380	4.0 μ M; Fig. 2b), as well as high concentrations of PO ₄ ³⁻ (35.9-42.0 μ M; Fig. 2c) and TChl <i>a</i>
381	(7.7-8.7 µg L ⁻¹ ; Fig. 2e) (Table 2).

383

3.2. Spatial distribution of dissolved hydrocarbons

Dissolved total AH and PAH concentrations and ratios are reported in Table 3. HAC and PCA applied to the main hydrocarbon variables from these tables reveal the presence of three groups of stations (G'1-G'3). These three groups are represented by PCA (Fig. 3) (HAC dendrogram not shown), in the form of box-and-whisker plots (Fig. 4) and colour maps (Fig. 5). AH and PAH molecular profiles are shown in Fig. 6 and Fig. 7, respectively. Stations displaying similar molecular profiles were grouped together. Below, we describe the hydrocarbon content of each of these three HAC/PCA groups.

391

392 *3.2.1. Group G'1*

Group G'1 includes the northernmost stations of Sfax (S1-S6) and all stations from the Gabès-Ghannouch region (S27-S36) (Fig. 1, 3, 5). These stations are characterized by the highest *R* concentrations, including Pr and Phy, with an average of $4.7 \pm 0.7 \mu g L^{-1}$ (Table 3; Fig. 3, 4a, 5). UCM is present in all stations, except S3 and S6, with concentrations varying

from 5.42 (S2) to 14 μ g L⁻¹ (S34), representing 63.4 ± 5.7% of the total AH pool (Table 3). When present, *R*/UCM values are all > 0.30 (Table 3). This group exhibits the lowest Pr/Phy ratios, which vary from 0.27 (S3) to 0.36 (S30, S33) (Table 3; Fig. 4b). AH molecular profiles display monomodal distributions centred on *n*-C₂₅–*n*-C₂₆ compounds (Fig. 6a). An equal amount of even and odd carbon numbered *n*-alkanes is also observed, resulting in CPI₁₅₋₄₀ values close to 1 (Table 3; Fig. 4c).

Concentration of \sum_{29} PAHs_{-P+A} was 29.7 ± 12.1 ng L⁻¹ (Table 3; Fig. 3, 4d, 5). Alk/Par 403 ratios range from 0.6 (S3) to 5.5 (S33) but are mainly > 2 (Table 3; Fig. 4e). PAHs_{P+A} pattern 404 distributions are marked by the dominance of LMW compounds (2-3 rings) over HMW 405 406 compounds (≥ 4 rings) (Fig. 7a, f), the former representing $88 \pm 4\%$ of total PAHs. Naphthalenes (C_2 -Naph > Naph > C_3 -Naph > C_1 -Naph) represent the major compounds, 407 followed by phenanthrenes (C_1 -Phe > C_2 -Phe > Phe > C_3 -Phe). Nevertheless, the heavy 408 409 compounds C₁-Pyr and Pyr are also present in significant proportions in these stations (6.4 \pm 4.8 and $3.8 \pm 3.9\%$, respectively) (Fig. 7a, f). The Flt/(Flt + Pyr) ratios range from 0.08 to 410 411 0.38 (Table 3; Fig. 4f).

412

413 *3.2.2. Group G'2*

Group G'2 contains the southernmost stations of the Sfax North coast (S7-S13) as well 414 as most stations from the Sfax Southern coast (S16-S18, S20-S26) (Fig. 1, 3, 5). These 415 stations are distinguished by the lowest R concentrations $(0.13 \pm 0.25 \ \mu g \ L^{-1})$ and the absence 416 of UCM (Table 3; Fig. 3, 4a, 5). Pr/Phy ratios vary from 0.51 (S7) to 2.43 (S21) but are 417 mainly > 1 (Table 3; Fig. 4b). AH molecular profiles also vary within this group. S7 and S11 418 reveal distributions similar to those of G'1 stations (i.e., monomodal distribution centred on 419 the n-C₂₅-n-C₂₆ range) (Fig. 6a) and do not display any odd-to-even carbon-numbered 420 dominance (CPI₁₅₋₄₀ ~1) (Fig. 3, 4c). The molecular profiles of S8 and S17 are marked by a 421

regular distribution of *n*-alkanes in the $n-C_{15}$ - $n-C_{38}$ range, with the maximal abundance of *n*-422 C_{29} (Fig. 6c) resulting in a slight odd-to-even carbon-numbered predominance ($CPI_{15-40} = 1.3$ 423 and ~1, respectively) (Fig. 3, 4c). A bimodal distribution of n-alkanes is observed at S26 and 424 S13, in which light compounds are centred on $n-C_{16}$ (S26) or $n-C_{15}$ (S13) and heavy 425 compounds are centred on $n-C_{28}$ (S26) or $n-C_{33}$ (S13) (Fig. 6d,e). In contrast, the molecular 426 427 profiles of stations S9, S10, S12, S16, S18 and S20-S25 also display monomodal distributions but feature a strong even-to-odd predominance within the $n-C_{16}-n-C_{22}$ range (Fig. 6f). This 428 trend results in low CPI₁₅₋₄₀ values (0.11-0.89), with n-C₁₆ being the major compound, 429 followed by *n*-C₁₈ (Table 3; Fig. 3, 4c). Globally, CPI_{15-40} values for this group are < 1 (Table 430 3, Fig. 4c). 431 G'2 stations record PAH concentrations similar to those of G'1 stations, with \sum_{29} PAHs. 432 $_{P+A}$ of 28.9 ± 16.2 ng L⁻¹ (Table 3; Fig. 3, 4d, 5). Alk/Par ratios are lowest in G'2, ranging 433 434 from 0.25 to 2.25 (but mainly falling between 1 and 2) (Table 3; Fig. 3, 4e). PAH molecular profiles reveal the predominance of LMW compounds (Fig. 7a-e), as LMW compounds 435 represent $89.2 \pm 8.2\%$ of all total PAHs. Naphthalenes are the dominant compounds, followed 436 by phenanthrenes (Fig. 7a-e). Heavy compounds are also present in these stations, with C₁-437 Pyr and Pyr in S7, Flt in S8, Flt and Chr in S21-S26 (Fig. 7a, b, e). Flt/(Flt + Pyr) ratios are 438 439 the highest in this group, with values exceeding 0.5, except at S7 and S11 (Table 3; Fig. 3, 4f). 440

441

442 *3.2.3. Group G'3*

Group G'3 comprises only three stations from the Sfax Southern coast: S14, S15 and S19 (Fig. 1, 3, 5). *R* concentration is high in S14 (4.5 μ g L⁻¹) and low in S15 and S19 (0.04 and 0.03 μ g L⁻¹, respectively) (Table 3; Fig. 3, 4a, 5). S14 displays a molecular distribution similar to those of G'1 stations (i.e., monomodal distribution centred on the *n*-C₂₅–*n*-C₂₆

447	range), with UCM present at a concentration of 7.4 μ g L ⁻¹ , representing 62.2% of total AHs
448	(Table 3; Fig. 6a). Stations S15 and S19 exhibit different molecular profiles, with an almost
449	regular distribution from $n-C_{15}$ to $n-C_{36}$ and the slight emergence of $n-C_{17}$ and Phy (Table 3;
450	Fig. 3, 6b). Within these three stations, Pr/Phy are low (0.34-0.48) and equal amounts of
451	even- and odd-carbon-numbered <i>n</i> -alkanes can be found (CPI \sim 1) (Table 3; Fig. 4b, c).
452	These stations contain the highest recorded PAH concentrations, with 175.8 \pm 20.7 ng
453	L ⁻¹ (Table 3; Fig. 3, 4d, 5). Molecular profile of the PAHs reveals the predominance of LMW
454	compounds (Fig. 7d), as inferred by the presence of the highest Alk/Par ratios (6.6 ± 0.6)
455	(Table 3; Fig. 3, 4e). The LMW compounds represent 97.6 \pm 0.3% of all total PAHs.
456	Naphthalenes are the dominant compounds, followed by phenanthrenes and fluorenes,
457	whereas the HMW compounds are almost undetectable (Fig. 7d). The Flt/(Flt + Pyr) ratios are
458	among the lowest measured values (0.08-0.13; Table 3; Fig. 3, 4f).
459	Additionally, we can also use the cross plot of Ant/(Ant + Phe) versus Flt/(Flt + Pyr)
460	ratios to distinguish different origins of PAHs (Yunker et al., 2002, 2015), as shown in Fig. 8.
461	Most stations reveal mixed origins of PAHs. All stations in G'3 and G'1 yield Ant/(Ant +
462	Phe) > 0.1 and Flt/(Flt + Pyr) < 0.4 (mixed origin), except S2, S3, S4, S6, which are strictly of
463	petrogenic origin. In contrast, G'2 stations record Ant/(Ant + Phe) < 0.1 and Flt/(Flt + Pyr) $>$
464	0.5 (mixed origin), except S8, which has a pyrogenic origin, and S7 and S11, which are
465	strictly of petrogenic origin. Therefore, the clustering of the highly contaminated $G'1 + G'3$
466	stations in an area where $Flt/(Flt + Pyr) < 0.4$ along with the clustering of the less
467	contaminated G'2 group (displaying more degraded hydrocarbons) in an area where
468	
	Ant/Ant+Phe < 0.1 , very likely suggests that PAHs have different origins in G'1 + G'3 than
469	Ant/Ant+Phe < 0.1, very likely suggests that PAHs have different origins in $G'1 + G'3$ than they do in $G'2$.

3.3. Correlation between biogeochemical parameters and dissolved hydrocarbons

472	Table 4 displays the Spearman's rank-order correlation matrix for biogeochemical
473	parameters and dissolved hydrocarbons. R concentration is significantly negatively correlated
474	with T (r = -0.68, n = 30, p < 0.05) and significantly positively correlated with $Si(OH)_4$
475	concentration (r = 0.56, n = 30, p < 0.05). The concentration of \sum_{29} PAHs _{-P+A} shows a
476	significant negative correlation with pH (r = -0.47, n = 30, p < 0.05) and significant positive
477	correlation with concentrations of NO ₃ ⁻ , PO ₄ ³⁻ , Si(OH) ₄ and TChl <i>a</i> (r = 0.39-0.66, n = 30, p <
478	0.05). Neither <i>R</i> concentrations nor \sum_{29} PAHs _{-P+A} concentrations are correlated to DOC
479	concentrations (r = 0.01-0.20, n = 30, $p > 0.05$). No significant relationship was found
480	between <i>R</i> and \sum_{29} PAHs _{-P+A} concentrations (r = -0.07, n = 36, p > 0.05), suggesting that AHs
481	and PAHs in this coastal area are decoupled. Evidence for this decoupling can also be seen in
482	results of the PCA, in which AH and PAH variables are located on principal component 1
483	(PC1) and principal component 2 (PC2), respectively, thus underscoring their absence or very
484	low correlation (Fig. 3). TChl a concentrations are significantly positively correlated to those
485	of NO ₃ ⁻ , PO ₄ ³⁻ , Si(OH) ₄ , Phae <i>a</i> , SPM, POC and DOC ($r = 0.40-0.70$, $n = 30$, $p < 0.05$). This
486	underlines the strong coupling between phytoplankton biomass, nutrients and the organic
487	carbon pool (Table 4). The absence of any correlation between POC and DOC concentrations,
488	as well as the positive correlation observed between DOC and SPM concentrations, suggest
489	the presence of slightly different sources of particulate and dissolved organic carbon in the
490	surface waters.
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493	4. Discussion
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495	4.1. Comparisons with previously published data
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4.1.1. Physical and biogeochemical parameters

498	Measured temperature and pH values in this study, which range from 18.0-27.3 °C and
499	7.90-8.30, respectively (Table 2), are within the same order of magnitude as those reported in
500	other studies in the Gulf of Gabès (Drira et al., 2014; 2016; Rekik et al., 2014) and in the
501	Western Mediterranean basin (Pujo-Pay et al., 2011). Salinity, however, displays a wider
502	range (32-42) than has been observed by previous works in this coastal area (i.e., 36-40 by
503	Drira et al., 2016). This is likely because stations in this study are located closer to the coast
504	and are thus more influenced by terrestrial freshwater inputs for the lower values and by a
505	thinner water column subjected to greater evaporation for the higher ones.
506	NO_2^- concentrations measured here (0.00-0.90 μ M; Table 2) are similar to those already
507	reported in these coastal waters (0.01-0.83 μ M), whereas NO ₃ ⁻ and Si(OH) ₄ concentrations
508	(0.01-4.0 and 1.4-33.4 μ M, respectively; Table 2) are lower than previously reported values
509	(1.2-13.4 and 2.6-55.2 μ M, respectively) (Rekik et al., 2013; Drira et al., 2016). PO ₄ ³⁻
510	concentrations measured here (5.5-120.3 μ M; Table 2) are higher than those that have
511	previously been recorded in this area (0.1-39.7 μ M) (Rekik et al., 2012, 2013; Drira et al.,
512	2016). The highest PO_4^{3-} concentrations are found in the Sfax Southern coast (G3 group:
513	stations S16, S20, S22; Fig. 1, 2c; Table 2) and can be attributed, in part, to the release of
514	phosphogypsum from the SIAPE industry, which is known to be enriched in PO_4^{3-} , as well as
515	to the release of domestic and industrial wastes from the Sidi Salem channel. The lowest
516	PO_4^{3-} concentrations are observed mainly in the Sfax Northern coast (G2 group; Fig. 1, 2c;
517	Table 2), where phosphoric acid activities have not occurred for several years.
518	TChl <i>a</i> concentrations measured in the present work (1.7-40.7 μ g L ⁻¹ ; Table 2) are
519	higher than those previously found in the same coastal waters (0.0-25.6 μ g L ⁻¹) (Rekik et al.,
520	2013; 2014), while our SPM concentrations (8.3-59.4 mg L^{-1} ; Table 2) are on the same order

of magnitude (0.7-30 mg L^{-1}) (Katlane et al., 2013) and lower than those measured in the Northern coast of Sfax (14-434 mg L^{-1}) (Rekik et al., 2014).

Therefore, these data suggest that this coastal area is characterized by an enrichment in 523 PO₄³⁻, TChl *a*, POC, DOC and SPM concentrations compared to other Mediterranean systems 524 (The Mermex group; Pujo-Pay et al., 2011; Karydis and Kitsiou, 2012). PO₄³⁻ anthropogenic 525 inputs from various industrial sources have led to the eutrophication (or the enhancement of 526 biological production) of these coastal waters (Ben Mustapha et al., 1999; Drira et al., 2014; 527 2016). The high variability in concentrations from station to station may be related not only to 528 different anthropogenic input contributions, but also to the complex bathymetry and 529 hydrodynamics of this area. 530

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4.1.2. Dissolved n-alkane and PAH concentrations

Compared to PAHs, only a few studies have investigated dissolved *n*-alkanes in marine 533 waters. Dissolved total R concentrations ($n-C_{15}-n-C_{40}$ with Pr and Phy) determined here in the 534 surface waters of the Sfax and Gabès coastal areas (0.02 to 6.3 μ g L⁻¹; Table 3) yield higher 535 values than those recorded in surface waters of the Marseille coastal area (Northwestern 536 Mediterranean Sea, France) (0.04-0.58 µg L⁻¹) (Guigue et al., 2011; 2014), in offshore surface 537 waters of the Northwestern Mediterranean Sea (0.7-2.4 μ g L⁻¹) (Marty and Saliot, 1976) and 538 in surface waters of the Gulf of Tunis (Tunisia) (0.25-1 µg L⁻¹; Mzoughi and Chouba, 2011). 539 Dissolved total PAH concentrations measured here in the surface waters of the Sfax and 540 Gabès coastal areas, i.e., 8.9-197.8 ng L⁻¹ for \sum_{29} PAHs_{-P+A} and 3.6-56.4 ng L⁻¹ for \sum_{19} PAHs₋ 541 _P, are high compared to those reported in the Western Mediterranean (\sum_{16} PAHs_{-P+A}: 0.4-2.2 542 ng L⁻¹ and \sum_{19} PAHs_{-P+A}: 0.16-0.8 ng L⁻¹) and the Eastern Mediterranean (\sum_{19} PAHs_{-P+A}: 543 0.16-8.8 ng L⁻¹) (Dachs et al., 1997; Berrojalbiz et al., 2011), as well as in coastal waters of 544 Barcelona (Spain) and Banyuls-sur-Mer (France) (\sum_{15} PAHs_{-P+A}: 2.5-41.7 ng L⁻¹ in surface 545

546	micro layer and 3.6-30.7 ng L^{-1} in surface waters) (Guitart et al., 2007). However, they are
547	within the same range as those observed in surface waters of the Marseille coastal area (\sum_{32}
548	PAHs _{-P+A} : 5-162 ng L ⁻¹) (Guigue et al., 2011), the Rhone delta (France) (\sum_{80} PAHs _{-P+A} : 15-
549	290 ng L ⁻¹) (Bouloubassi and Saliot, 1991), and the Venice Lagoon (Italy) (\sum_{20} PAHs _{-P+A} :
550	12.4-266.8 ng L ⁻¹) (Manodori et al., 2006). Our measured concentrations are also lower than
551	those found in the Gulf of Tunis (\sum_{24} PAHs _{-P+A} : 139-694 ng L ⁻¹ in summer and 150-1008 ng
552	L^{-1} in winter) (Mzoughi and Chouba, 2011) and the Saronikos Gulf (Greece) ($\sum_{17} PAHs_{-P+A}$:
553	103-459 ng L ⁻¹) (Valavanidis et al., 2008). El Nemr and Abd-Allah (2003) also reported
554	higher concentrations of PAHs. _P in surface waters of the Alexandria coast (Egypt) (\sum_7 PAHs.
555	_P : 13-120 ng L^{-1}). Consequently, these AH and PAH concentration data indicate that the
556	surface coastal waters of the Gulf of Gabès are likely moderately-to-highly contaminated
557	when compared to other Mediterranean regions. Hence concentration levels of hydrocarbons
558	recorded in surface waters of the Gulf of Gabès are not so high compared to other
559	Mediterranean coastal environment despite strong anthropogenic releases. This could be
560	explained by the influence of hydrodynamic conditions, in particular tides. Tides of the Gulf
561	of Gabès, which present the highest amplitude of all the Mediterranean Sea, with the upper
562	Adriatic, may have a role of dispersing hydrocarbons at sea.
563	

4.2. Origin and potential sources of dissolved hydrocarbons

4.2.1. Group G'1

AH and PAH concentrations and ratios (*R*/UCM > 0.3; CPI ~ 1 and Alk/Par > 2)
indicate the presence of recent and chronic petrogenic contaminations (Simoneit and
Mazurek, 1982; Wang et al., 1999; Commendatore and Esteves, 2004; Commendatore et al.,
2012). Additionally, the cross plot and occurrence of Pyr and C₁-Pyr suggest, for many of

these stations, a contribution of pyrogenic (combustion) sources (Fig. 7, 8). AH profiles 571 572 display a molecular distribution very close to that recorded for classic heavy fuels or residuals used as marine diesel and for industrial power generation (Wang and Fingas, 2003). This 573 574 observed high proportion of alkylated homologues was also reported in Sfax surficial sediments by Zaghden et al. (2014). The hydrocarbon contamination found in the 575 576 northernmost stations of Sfax and in the Gabès-Ghannouch stations may have several sources, 577 especially wadis, uncontrolled discharges, and harbour activities. The Ezzit wadi, which may transport various uncontrolled domestic, industrial, and agricultural wastes, would thus 578 represent a significant source of petrogenic hydrocarbons for the Northern coast of Sfax. 579 580 Within the Gabès coastal area, the main sources of hydrocarbons include industrial discharges (from the thermal power plant), commercial and fishing harbours (by means of oil spills, 581 ballast water, shipping, and air emissions) and wadis (El-Tine and Gabès wadis) (Fig. 1). 582 583 Despite the weak correlation between biogeochemical and hydrocarbon individual parameters, this observed G'1 station contamination is supported by the presence of 584 widespread anthropisation occurring in the northernmost Sfax stations (G1) and in the Gabès-585 Ghannouch area (G5 + G6 stations). 586

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588 *4.2.2. Group G'2*

This group is characterized by a range of diverse AH profiles, from the most petrogenic distribution (S7, S11, close to G'1 profiles; Fig. 3, 6a) to increasingly biogenic origin markers (away from G'1 profiles, Fig. 3, 6c-f). The emergence of odd *n*-alkanes n-C₁₅, n-C₁₇, n-C₂₉, n-C₃₃ may be attributed to phytoplankton or higher plant debris lysis (Douglas and Eglington, 1966; Blumer et al., 1971; Goutx and Saliot, 1980), as well as the suspected presence of small-size phytoplankton cells (picophytoplankton). Since micro and macro algae are abundant in the Gulf of Gabès (Bel Hassen et al., 2008; Drira et al., 2014; Rekik et al., 2014),

this assumption seems plausible. Even *n*-alkanes can be attributed to microbial activities such 596 597 as bacteria, fungi and yeast (Elias et al., 1997). Stations S9, S10, S12, S16, S18, S20-25, which yield the lower CPI, exhibit higher abundances of these microbial signatures (Fig. 3, 598 599 6f). The hypothesised biogenic signatures of AHs within this group are supported by the presence of Pr/Phy ratios > 1 (Fig. 4b, c). Also, the relatively low concentration levels of AHs 600 601 and PAHs, associated with low Alk/Par values (compared to the two other groups), suggest at 602 these sites less petrogenic inputs, and, at the same time, more pronounced weathering processes, very likely through biodegradation. Indeed, it has been shown that the bacterial 603 degradation of PAHs could lead to a depletion of alkylated PAHs compared to their non 604 605 alkylated counterparts (Vila et al., 2010; Jiménez et al., 2011).

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4.2.3. Group G'3

608 This group is mainly characterized by the highest PAH concentrations, LMW PAH abundances, and Alk/Par values associated with CPI ~ 1, which indicate the importance of 609 610 local petrogenic inputs. These stations were sampled in March 2015 close to the coast and were thus likely affected by anthropogenic inputs from Sidi Salem channel as well as by 611 fishing and commercial harbours located in the Southern coast of Sfax. The Sidi Salem 612 613 channel contains, among other things, untreated wastewater from slaughter-house, whose flow rate is estimated at 55 m³ day⁻¹, sludge from the WWTP, and pretreated wastewaters of 614 various industries (DGQEV, 2008). Therefore, all these inputs may substantially contribute to 615 the PAH content in this area. 616

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4.2.4. Differences between Sfax and Gabès environments

In the Sfax region, we observe a combination of petrogenic inputs (G'1, G'3) and
biogenic sources (G'2), as well as the occurrence of weathering processes which are

supported by the decrease in both AH and PAH concentrations and Alk/Par ratio off of the 621 622 coast (G'2: S1-S3/S4, S5-S7, S8-S11, S12-S13, S16-S18, S19-S26). Even though these weathering processes seem to be driven by biodegradation (Vila et al., 2010; Jiménez et al., 623 2011), we cannot exclude other processes such as volatilisation (Jiménez et al., 2012), 624 photodegradation (King et al., 2014) and particle sequestration (Dachs et al., 2002). These 625 626 biogenic AHs and weathering processes are not observed in the Gabès-Ghannouch region; we 627 assume that this is due to either the recurrence of larger releases in Gabès, with continuous and direct discharge of phosphogypsum into the sea, or to a particular ecosystem in Sfax that 628 is more suitable for the biodegradation of hydrocarbons. Measurements of bacterial 629 630 abundance, diversity and activities in these two ecosystems should thus be considered as complements to hydrocarbon analysis in future studies. Moreover, in the Sfax waters, we did 631 not observe coupling between the hydrocarbon and biogeochemical contents, whereas 632 633 hydrocarbons in the Gabès-Ghannouch area (G'1 stations) are associated with biogeochemical groups G5 and G6, which exhibit the highest measured values of silica. These data may thus 634 reflect the influence of terrigenous inputs (Gabès and El-Tine wadis), chronic releases or 635 spills/plumes (harbour activities, phosphogypsum releases). 636

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4.2.5. Inputs of PAHs from particles

For each of the three hydrocarbon groups, the cross plot shows the superposition of pyrogenic inputs in addition to the widespread existence of petrogenic sources (Fig. 8). This pyrogenic component probably reflects the influence of either high suspended matter contents or of the proximity of sediment. Given the dynamics of the area, we may assume that we are not in a particle-dissolved equilibrium position and thus that rearrangements may occur in cases of environmental disturbance (Dong et al., 2015). Dissolved PAHs found here in surface waters may partially originate from surficial sediments. Indeed, in these very shallow waters,

exchanges between sediment and the water column are exacerbated. Studies carried out on 646 647 surficial sediments of the Sfax coastal area have demonstrated that phenanthrenes (especially alkylated homologues) are among the most abundant PAHs (Zaghden et al., 2007; 2014). 648 Consequently, some LMW PAHs recorded in surface waters may be released from sediments, 649 either by the resuspension of particulate matter or by passive diffusion. This process of 650 651 partitioning between the two matrixes (water and sediment) is highly related to the 652 octanol/water partition coefficients (Kow) of PAHs, as well as to the organic carbon content in sediment that controls this partitioning through a linear free energy relationship (Zhou and 653 654 Maskaoui, 2003).

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4.3. PAH partitioning in the dissolved phase

DOC is considered as one of the most important factors controlling the partitioning of 657 658 hydrocarbons in the dissolved phase. Although we record high DOC concentrations in these coastal waters, no linear correlation is found between DOC and dissolved \sum_{29} PAHs._{P+A} 659 concentrations (r = 0.01, n = 30, p > 0.05). In addition, no linear correlation is observed 660 between individual PAH and DOC concentrations, except for Ace (data not shown). This lack 661 of correlation between DOC and dissolved PAHs may a reflect different sources for these two 662 663 pools in surface waters of the Gulf of Gabès, but also their low affinity at the molecular level. Therefore, a fraction of dissolved LMW PAHs in this area could be under freely form (i.e., 664 not complexed to dissolved organic matter), which is in agreement with their physico-665 chemical properties (relatively low K_{ow} and high solubility) (Shi et al., 2007). Besides, 666 significant negative linear relationships are found between individual PAH concentrations and 667 their respective K_{ow} values (data not shown), which suggests that the molecular distribution 668 pattern observed, i.e., dominance of LMW PAHs in all samples, is driven by the hydrophobic 669 character of compounds. Interestingly, significant positive correlations are detected between 670

several dissolved PAHs (C₂-Naph, C₃-Naph, Ace, Flu, C₁-Flu, C₂-Flu and DBT) and POC or 671 SPM concentrations (data not shown). This could underline a "particle concentration effect" 672 (Zhou et al., 1999). The latter is observed when particle partition coefficient (K_p or K_{oc}) is 673 inversely proportional to particle concentration. The increase in particle concentration is 674 generally associated to an increase in DOC concentration, which may explain this distribution 675 676 of PAHs towards the dissolved phase (Zhou et al., 1999). However, in our case, the increase 677 in POC/SPM concentrations was not concomitant to an increase in DOC concentrations. Our results thus illustrate the complexity of the relationships between dissolved PAHs, DOC and 678 POC in marine coastal waters. 679

680

681 **5.** Conclusions

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While some works have already reported hydrocarbon levels in surficial sediments of 683 the Sfax coastal area (Zaghden et al., 2007; 2014), this present study represents to our 684 knowledge the first assessment of concentrations of dissolved hydrocarbons (28 AHs and 29 685 PAHs) in surface waters of the Gulf of Gabès (in the Sfax and Gabès coastal areas), and their 686 associations with biogeochemical parameters. Overall, these areas are moderate-to-highly 687 688 impacted by hydrocarbons and show a relatively high eutrophication level compared to other Mediterranean coastal environments. The highest hydrocarbon concentrations can be 689 attributed to recent petrogenic sources from industrial, domestic and harbour activities. 690 691 Interestingly, biogenic AHs and signatures of weathering processes are found in Sfax, but not in Gabès, with lower hydrocarbon contents and decreasing Alk/Par ratios off the coast. 692 Concentrations in AHs, PAHs, and DOC do not correlated with each other, thus highlighting 693 that these three pools have different sources and are then submitted to different processes in 694 the Gulf of Gabès. Despite these relatively high DOC concentrations, dissolved PAHs do not 695

seem to show an affinity towards dissolved organic matter. These results demonstrate that the
dissolved phase can provide very interesting information about the dynamics of PAHs in
coastal waters. Further investigations should take into consideration the particulate organic
matter of water column to obtain a global view of PAH dynamics in coastal waters of the Gulf
of Gabès.

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Figure 1. Location of the study stations in the coastal areas of Sfax and Gabès cities, in the Gulf of Gabès (Southeastern Mediterranean Sea, Tunisia). The detailed characteristics of the stations are given in Table 1. The six groups of stations (G1-G6) identified according to the hierarchical ascendant classification applied to biogeochemical parameters are presented with different colours.

1049 Figure 2. Box-and-whisker plots of biogeochemical parameters with regard to the six clusters

identified from the hierarchical ascendant classification: a) NO_2^- , b) NO_3^- , c) PO_4^{3-} , d)

1051 Si(OH)₄, e) TChl *a*, f) Phaeo *a*, g) SPM, h) POC and i) DOC concentrations. The six clusters

are as follow: G1 (S2, S6), G2 (S3, S4, S7, S9-S13), G3 (S16, S20, S22), G4 (S17, S18, S21,

1053 S23-S26), G5 (S27-S32; S36) and G6 (S33, S34). The black and red lines represent the

1054 median and mean values, respectively.

Figure 3. Principal component analysis (PCA) of hydrocarbons dataset: a) correlation circle
and b) projections of samples on the first factorial plane (PC1 *versus* PC2).

1057 Figure 4. Box-and-whisker plots of main hydrocarbon parameters with regard to the three

1058 clusters identified from the hierarchical ascendant classification: a) total resolved *n*-alkanes

1059 (*R*), b) Pr/Phy, c) CPI₁₅₋₄₀, d) \sum_{29} PAH_{-P+A}, e) Alk/Par and f) Flt/(Flt + Pyr). The three clusters

1060 are as follow: G'1 (S1-S6, S27-S36), G'2 (S7-S13, S16-S18, S20-S26) and G'3 (S14, S15,

1061 S19). The black and red lines represent the median and mean values, respectively.

1062 Figure 5. Spatial distribution of dissolved AH and PAH concentrations in the Sfax and Gabès

1063 coastal areas. The three groups of stations identified according to the hierarchical ascendant

- 1064 classification applied to hydrocarbon parameters are presented in black (G'1), red (G'2) and
- blue (G'3). Ocean Data View software version 4.6.5, Schlitzer, R., http://odv.awi.de, 2014.

- 1066 Figure 6. Relative abundances of dissolved AHs (%) for stations a) S1-S7, S11, S14, S27-
- 1067 S36, b) S15, S19, c) S8, S17, d) S26, e) S13 and f) S9, S10, S12, S16, S18, S20-S25.
- 1068 Figure 7. Relative abundances of dissolved PAHs (%) for stations a) S1-S7, b) S8, c) S9-S13,
- 1069 d) S14-S20, e) S21-S26 and f) S27-S36.
- 1070 **Figure 8.** PAH cross-plot between Flt/(Flt + Pyr) and Ant/(Ant + Phe) ratios. Stations S1,
- 1071 S13, S29-S31, S35 and S36, for which Ant was not detected, are not represented.

Table 1. Characteristics of the study stations in the coastal areas of Sfax and Gabès cities, in the Gulf of Gabès (Southeastern Mediterranean Sea,

1073 Tunisia).

Stations	Area	Position	Date of	Site depth	Sampling depth
			sampling	(m)	(m)
S1	Northern Sfax, in front of Ezzit wadi	34°45'34.20"N, 10°48'30.06"E	08/03/2015	0.5	0.1
S2	Northern Sfax, in front of Ezzit wadi	34°45'29.31"N, 10°48'38.11"E	23/10/2014	0.9	0.1
S3	Northern Sfax, in front of Ezzit wadi	34°45'04.18"N, 10°49'05.26"E	23/10/2014	1.2	0.1
S4	Northern Sfax, in front of Ezzit wadi	34°44'40.06"N, 10°49'30.42"E	23/10/2014	1.8	0.1
S5	Northern Sfax, in front of Taparura fishing harbour	34°45'16.47"N, 10°47'55.98"E	08/03/2015	0.5	0.1
S6	Northern Sfax, in front of Taparura fishing harbour	34°45'12.08"N, 10°48'01.54"E	23/10/2014	2	0.1
S7	Northern Sfax, in front of Taparura fishing harbour	34°44'50.57"N, 10°48'33.99"E	23/10/2014	2	0.1
S8	Northern Sfax, in front of PK4 channel	34°44'57.08"N, 10°47'20.72"E	08/03/2015	0.5	0.1
S9	Northern Sfax, in front of PK4 channel	34°44'48.13"N, 10°47'28.79"E	23/10/2014	1.2	0.1
S10	Northern Sfax, in front of PK4 channel	34°44'25.94"N, 10°48'02.75"E	23/10/2014	2	0.1
S11	Northern Sfax, in front of PK4 channel	34°44'06.98"N, 10°48'30.00"E	23/10/2014	1	0.1
S12	Northern Sfax, in front of buried phosphogypsum	34°44'11.50"N, 10°47'17.78"E	23/10/2014	2	0.1
S13	Northern Sfax, in front of buried phosphogypsum	34°43'54.81"N, 10°47'49.23"E	23/10/2014	2	0.1
S14	Southern Sfax, in front of commercial harbour	34°43'15.18"N, 10°46'55.91"E	08/03/2015	0.5	0.1
S15	Southern Sfax, between commercial and fishing harbours	34°42'53.31"N, 10°46'36.94"E	08/03/2015	0.5	0.1
S16	Southern Sfax, in front of fishing harbour	34°42'33.58"N, 10°46'10.56"E	18/10/2014	3	0.1
S17	Southern Sfax, in front of fishing harbour	34°42'13.32"N, 10°46'43.64"E	18/10/2014	5	0.1
S18	Southern Sfax, in front of fishing harbour	34°41'55.94"N, 10°47'17.71"E	18/10/2014	9	0.1
S19	Southern Sfax, in front of Sidi Salem channel/industrial zone	34°42'32.34"N, 10°45'22.38"E	08/03/2015	0.5	0.1
S20	Southern Sfax, in front of Sidi Salem channel/industrial zone	34°42'15.90"N, 10°45'54.53"E	18/10/2014	1	0.1
S21	Southern Sfax, in front of Sidi Salem channel/industrial zone	34°41'58.75"N, 10°46'30.02"E	18/10/2014	6	0.1
S22	Southern Sfax, in front of El Maou wadi/SIAPE phosphogysum area	34°41'35.05"N, 10°45'25.26"E	18/10/2014	1	0.1
S23	Southern Sfax, in front of El Maou wadi/SIAPE phosphogysum area	34°41'22.70"N, 10°46'01.65"E	18/10/2014	8	0.1
S24	Southern Sfax, in front of El Maou wadi/SIAPE phosphogysum area	34°41'08.60"N, 10°46'37.37"E	18/10/2014	6.2	0.1
S25	Southern Sfax, in front of El Hakmouni wadi/SIAPE phosphogysum area	34°40'40.42"N, 10°45'00.41"E	18/10/2014	4	0.1
\$26	Southern Sfax, in front of El Hakmouni wadi/SIAPE phosphogysum area	34°40'25.81"N, 10°45'37.67"E	18/10/2014	5.5	0.1
\$27	Gabès, in front of El-Tine wadi	33°56'17.26"N, 10°05'09.35"E	13/11/2014	5.5	0.1
S28	Gabès, in front of El-Tine wadi	33°56'25.23"N, 10°05'47.31"E	13/11/2014	8.5	0.1
S29	Gabès, in front of El-Tine wadi	33°56'32.81"N, 10°06'25.37"E	13/11/2014	10	0.1
S30	Gabès, in front of commercial harbour and phosphogysum discharges	33°55'03.65"N, 10°06'25.72"E	13/11/2014	8	0.1
531	Gabès, in front of commercial harbour and phosphogysum discharges	33°55'16.48"N, 10°07'01.99"E	13/11/2014	9.5	0.1

	S32 S33 S34 S35 S36	Gabès, in front of commercial harbour and phosphogysum discharges Gabès, between commercial and fishing harbours Gabès, between commercial and fishing harbours Gabès, in front of fishing harbour and Gabès wadi Gabès, in front of fishing harbour and Gabès wadi	33°55'27.79"N, 10°07'36.81"E 33°54'24.48"N, 10°06'37.27"E 33°54'37.77"N, 10°07'13.43"E 33°53'50.15"N, 10°07'20.52"E 33°54'06.24"N, 10°07'55.26"E	13/11/2014 13/11/2014 13/11/2014 13/11/2014 13/11/2014	10 2.8 8 7.5 10.5	0.1 0.1 0.1 0.1 0.1	
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Table 2. Physical and biogeochemical parameters.

Stations	T (°C)	Salinity	pН	NO_2^-	NO ₃	PO_{4}^{3-}	Si(OH) ₄	TChl a	Phae a	SPM	POC	DOC
				(µM)	(µM)	(µM)	(µM)	$(\mu g L^{-1})$	$(\mu g L^{-1})$	$(mg L^{-1})$	(µM)	(µM)
S1	nd	Nd	8.30	nd	nd	nd	nd	nd	nd	nd	nd	nd
S2	21.0	42	8.11	0.24	0.11	6.2	8.5	16.1	2.8	43.6	244.8	180.8
S3	21.8	42	8.19	0.06	0.06	7.5	6.1	5.7	1.7	16.5	53.2	148.3
S4	21.0	42	8.14	0.10	0.09	7.8	6.0	5.3	0.6	13.4	43.5	144.9
S5	nd	Nd	8.12	nd	nd	nd	nd	nd	nd	nd	nd	nd
S6	21.5	42	8.06	0.07	0.18	9.7	8.1	6.0	3.4	59.4	207.6	139.7
S 7	22.0	40	8.21	0.04	0.12	8.6	5.1	2.3	0.6	9.7	30.3	129.1
S 8	nd	Nd	8.17	nd	nd	nd	nd	nd	nd	nd	nd	106.7
S9	22.5	42	8.11	0.05	0.26	11.6	4.1	2.9	1.4	17.6	66.5	122.2
S10	22.6	40	8.14	0.03	0.13	10.0	4.0	1.7	0.7	23.3	28.2	122.1
S11	22.0	38	8.14	0.07	0.17	9.4	6.9	3.1	2.2	25.8	101.8	135.2
S12	23.2	42	8.01	0.07	0.50	14.7	4.3	4.1	2.0	16.4	45.3	146.8
S13	21.5	40	8.00	0.07	0.29	11.9	3.1	2.9	0.6	15.7	23.6	124.1
S14	nd	Nd	8.08	nd	nd	nd	nd	nd	nd	nd	nd	nd
S15	nd	Nd	8.22	nd	nd	nd	nd	nd	nd	nd	nd	115.0
S16	26.7	40	8.02	0.68	1.50	120.0	13.4	40.7	9.7	27.1	177.5	190.9
S17	26.5	40	8.03	0.00	0.06	17.4	2.5	7.7	3.8	11.8	49.4	127.3
S18	27.3	40	7.94	0.01	0.07	5.5	1.4	2.0	0.4	10.6	14.9	107.3
S19	nd	Nd	8.19	nd	nd	nd	nd	nd	nd	nd	nd	132.1
S20	26.9	40	8.06	0.90	1.76	120.3	7.3	29.0	7.8	32.2	168.2	186.6
S21	26.5	38	8.00	0.23	0.86	24.0	3.2	8.7	2.4	13.0	42.4	126.3
S22	26.9	32	8.11	0.66	1.20	82.1	7.2	13.8	3.0	19.5	77.0	206.9
S23	26.8	38	8.00	0.15	0.45	6.4	2.8	2.8	0.8	9.0	24.7	126.2
S24	26.9	40	7.90	0.01	0.09	10.2	2.3	3.2	1.0	8.3	17.0	121.1
S25	26.7	35	7.90	0.35	1.31	14.5	5.3	3.9	2.5	10.1	30.5	126.2
S26	26.8	38	7.94	0.16	0.47	12.0	4.6	4.9	1.9	8.8	35.7	117.4
S27	22.0	36	7.97	0.02	0.89	18.7	13.1	6.1	1.7	17.0	34.1	137.4
S28	22.0	35	7.99	0.01	0.31	14.9	11.2	5.7	1.7	19.5	34.6	110.6
S29	21.0	32	8.14	0.00	0.01	6.8	2.6	3.0	0.7	18.7	24.5	112.9
S30	20.0	32	8.08	0.00	0.97	19.9	15.2	7.1	1.1	21.8	36.8	217.6

S31	20.0	34	8.00	0.00	0.14	14.1	7.9	6.7	1.2	22.7	39.1	124.3
S32	21.0	35	8.02	0.00	0.01	15.0	8.1	8.1	2.1	22.2	45.1	122.8
S33	18.0	36	7.97	0.00	4.00	42.1	33.4	7.7	1.4	19.5	46.2	110.6
S34	18.0	38	8.02	0.03	3.22	35.9	27.6	8.7	1.6	19.3	54.8	118.9
S35	18.0	40	8.13	0.00	0.39	12.1	6.6	5.0	1.5	21.9	36.8	127.8
S36	18.0	39	8.13	0.00	1.47	21.8	14.9	6.5	1.6	18.8	54.4	187.1

nd: not determined. NO₂⁻: nitrites; NO₃⁻: nitrates; PO₄³⁻: phosphates; Si(OH)₄: silicates; TChl *a*: total chlorophyll *a*; Phae *a*: phaeopigment *a*; 1097 POC: particulate organic carbon; SPM: suspended particulate matter; DOC: dissolved organic carbon.

							-					
	$R (\mu g L^{-1})$	UCM ($\mu g L^{-1}$)	<i>R</i> /UCM	Pr/Phy	CPI-15-40	MH	$\sum_{16} PAH$		$\sum_{29} PAH_{-P+A}$	Alk/Par		Flt/
							$(ng L^{-1})$	$(ng L^{-1})$	$(ng L^{-1})$		(Ant + Phe)	(Flt + Pyr)
S1	4.46	7.95	0.56	0.32	0.99	$n-C_{25}-C_{26}$	6.25	6.25	34.84	4.58	nd	0.08
S2	5.34	5.42	0.99	0.34	0.96	$n-C_{25}-C_{26}$	12.73	12.98	27.88	1.15	0.05	0.21
S 3	5.88	Np	nd	0.27	1.05	$n-C_{25}-C_{26}$	10.39	10.51	17.16	0.63	0.04	0.21
S4	6.31	7.96	0.79	0.34	1	$n-C_{25}-C_{26}$	10.66	10.80	19.52	0.81	0.02	0.21
S5	4.93	8.30	0.59	0.33	0.97	$n-C_{25}-C_{26}$	7.58	7.58	37.67	3.97	0.78	0.10
S6	3.84	Np	nd	0.36	0.98	$n-C_{25}-C_{26}$	7.90	8.08	18.14	1.24	0.03	0.20
S7	1.09	Np	nd	0.51	1.09	$n-C_{25}-C_{26}$	10.89	11.08	20.23	0.83	0.04	0.19
S 8	0.033	Np	nd	1.02	1.30	$n-C_{29}$	14.50	14.50	45.21	2.12	0.75	0.91
S9	0.030	Np	nd	1.13	0.31	$n-C_{16}-C_{18}$	8.38	8.64	24.58	1.84	0.05	0.57
S10	0.021	Np	nd	0.97	0.49	$n-C_{16}-C_{18}$	3.69	3.79	9.86	1.6	0.07	0.58
S11	0.144	Np	nd	0.79	0.93	$n-C_{25}-C_{26}$	4.44	4.50	9.73	1.16	0.08	0.40
S12	0.045	Np	nd	1.06	0.41	$n-C_{16}$	16.67	17.45	53.70	2.08	0.04	0.55
S13	0.053	Np	nd	1.51	0.97	$n-C_{33}$	3.53	3.63	8.94	1.46	nd	0.51
S14	4.51	7.41	0.61	0.34	0.97	$n-C_{25}-C_{26}$	19.52	19.52	156.82	7.03	0.80	0.13
S15	0.044	Np	nd	0.43	0.94	n-C ₁₇ -Phy	25.65	25.65	197.83	6.71	0.79	0.10
S16	0.041	Np	nd	1.57	0.89	$n-C_{15}-C_{16}$	7.66	8.08	26.25	2.25	0.08	0.56
S17	0.094	Np	nd	1.73	0.86	$n-C_{29}$	5.68	6.00	19.41	2.23	0.05	0.53
S18	0.16	Np	nd	1.51	0.11	$n-C_{16}$	7.42	7.71	19.84	1.57	0.03	0.61
S19	0.035	Np	nd	0.48	0.99	n-C ₁₇ -Phy	24.90	24.90	172.76	5.94	0.78	0.08
S20	0.25	Np	nd	0.90	0.14	$n-C_{16}-C_{18}$	7.79	8.29	25.97	2.13	0.05	0.49
S21	0.034	Np	nd	2.43	0.59	$n-C_{16}$	19.65	20.33	37.29	0.83	0.03	0.88
S22	0.077	Np	nd	1.25	0.33	$n-C_{16}$	13.59	14.30	37.85	1.65	0.04	0.78
S23	0.038	Np	nd	1.64	0.53	$n-C_{16}$	18.84	19.21	30.13	0.57	0.02	0.85
S24	0.026	Np	nd	1.76	0.52	$n-C_{16}$	9.98	10.40	22.79	1.19	0.03	0.74
S25	0.032	Np	nd	1.66	0.42	$n-C_{16}$	53.73	56.37	70.60	0.25	0.04	0.86
S26	0.045	Np	nd	1.85	0.62	$n-C_{16}$	18.24	19.08	29.26	0.53	0.02	0.84
S27	4.66	12.03	0.39	0.29	0.99	$n-C_{26}-C_{25}$	6.63	6.63	37.49	4.66	0.31	0.14
S28	4.53	7.37	0.61	0.35	0.91	$n-C_{26}-C_{25}$	6.36	6.36	39.40	5.2	0.62	0.12
S29	4.41	7.32	0.60	0.30	0.96	$n-C_{26}-C_{25}$	4.03	4.03	15.48	2.84	nd	0.06
S30	4.92	8.34	0.59	0.36	0.98	$n-C_{26}-C_{25}$	6.07	6.07	24.09	2.97	nd	0.19
S31	4.28	8.13	0.53	0.35	0.99	$n-C_{26}-C_{25}$	4.23	4.23	16.56	2.91	nd	0.09

Table 3. Dissolved aliphatic hydrocarbons (AHs) and polycyclic aromatic hydrocarbons (PAHs): concentrations and ratios.

S32	4.44	7.89	0.56	0.34	0.98	$n-C_{26}-C_{25}$	6.37	6.37	29.24	3.6	0.38	0.15
S33	3.61	6.53	0.55	0.36	0.98	$n-C_{26}-C_{25}$	7.33	7.33	47.42	5.47	0.40	0.17
S34	5.08	14.03	0.36	0.31	0.95	$n-C_{26}-C_{25}$	9.38	9.38	56.79	5.05	0.22	0.38
S35	4.38	7.47	0.59	0.32	0.99	$n-C_{26}-C_{25}$	5.09	5.09	20.45	3.02	nd	0.10
S36	3.91	7.05	0.55	0.31	0.98	$n-C_{26}-C_{25}$	7.25	7.25	33.10	3.56	nd	0.18

np: not present; *nd*: not determined. *R*: total resolved *n*-alkanes from $n-C_{15}$ to $n-C_{40}$ including two isoprenoids, pristane (Pr) and phytane (Phy); UCM:

1117 unresolved complex mixture; CPI₁₅₋₄₀: carbon preference index: \sum odd *n*-alkanes *n*-C₁₅-*n*-C₄₀/ \sum even *n*-alkanes *n*-C₁₅-*n*-C₄₀; MH: major hydrocarbons. \sum_{16}

1118 PAH: sum of the 16 priority PAHs listed by EPA; \sum_{19} PAH_{-P}: sum of 19 parent PAHs; \sum_{29} PAH_{-P+A}: sum of 29 parent and alkylated PAHs; Alk/Par: sum of

- 1119 alkylated PAHs/sum of parent PAHs; Ant: anthracene; Phe: phenanthrene; Flt: fluoranthene; Pyr: pyrene.

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Table 4. Spearman's rank-order correlation matrix for biogeochemical parameters and dissolved hydrocarbons (n = 30).

	T (°C)	Salinity	pН	NO_2^-	NO_3^-	PO_4^{3-}	Si(OH) ₄	TChl a	Phae a	SPM	POC	DOC	R
Salinity	0.09												
pH	-0.35	0.32											
NO ₂	0.54	0.30	-0.04										
NO ₃	0.00	-0.26	-0.30	0.31									
PO ₄ ³⁻	-0.03	-0.37	-0.25	0.03	0.74								
Si(OH) ₄	-0.56	-0.24	0.07	-0.06	0.54	0.53							
TChl a	-0.18	-0.16	-0.05	0.17	0.41	0.70	0.65						
Phae a	0.23	0.04	-0.07	0.47	0.32	0.50	0.33	0.69					
SPM	-0.18	0.29	0.36	0.34	0.27	0.36	0.56	0.69	0.72				
POC	-0.39	0.00	0.37	0.01	0.15	0.25	0.63	0.49	0.41	0.64			
DOC	-0.03	0.23	0.45	0.38	0.24	0.25	0.36	0.46	0.45	0.56	0.35		
R	-0.68	-0.06	0.33	-0.32	-0.16	-0.06	0.56	0.35	-0.08	0.22	0.32	0.20	
$\sum_{29} PAH_{-P+A}$	0.09	-0.30	-0.47	0.22	0.66	0.55	0.39	0.40	0.34	0.15	-0.15	0.01	-0.07^{*}

1138 Significant correlations (p < 0.05) are in bold. Stations S1, S5, S8, S14, S15 and S19 are not included in these correlations (absence of biogeochemical parameter measurements). * For the correlation between *R* and \sum_{29} PAH_{P+A} all stations are taken into account (i.e., n = 36).