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GENE EXPRESSION ANALYSIS OF DUCK LIVER STEATOSIS FROM HYBRID AND PARENTAL SPECIES

2. Samples



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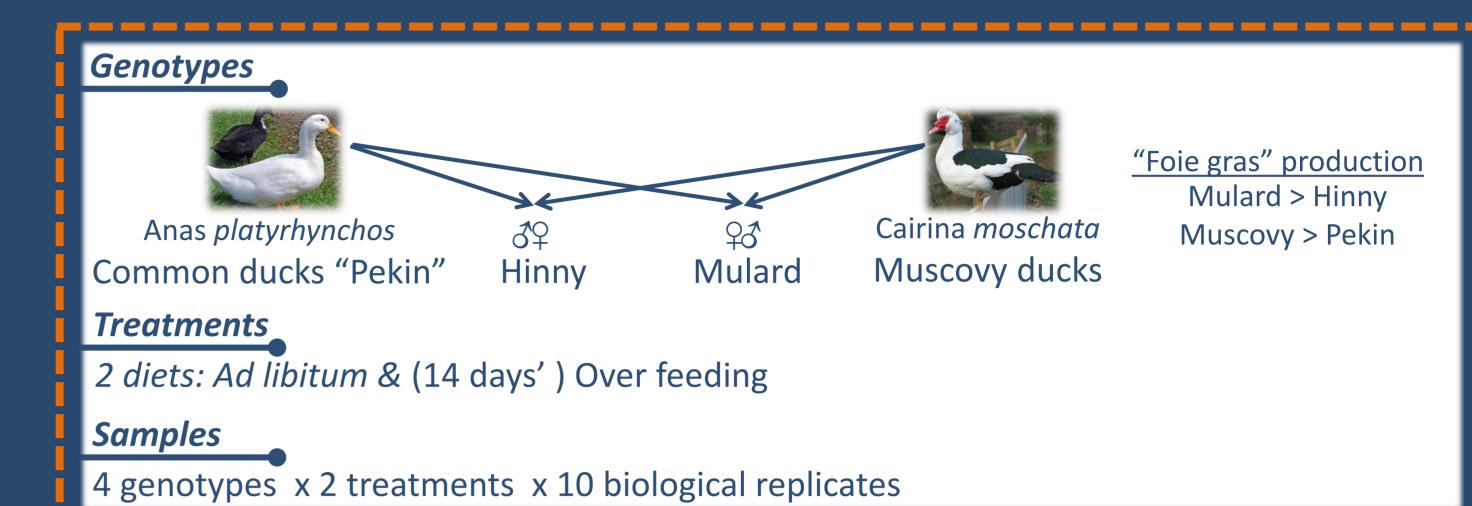
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1. Background

3. Assemblies

iver steatosis can occur spontaneously in wild waterfowls as a result of energy storage before migration. This ability is exploited since thousand years in domesticated birds to produce "foie gras" by overfeeding. However, different abilities for fatty liver production are known according to species. To better describe the mechanisms involved in hepatic steatosis development and differences between species and hybrids, next-generation sequencing and analyses was performed on RNAs extracted from the livers of ducks fed ad libitum or overfed.

Usually, such RNA-seq analyses involve the mapping of reads on a reference genome. However, when two different species are involved some mapping biases could be expected. Thus, alternative methods must be developed to be more appropriate for transcriptome analyses and comparisons between the two species and their hybrids. Analyses using both reference genome and *de novo* methods point out a good performance of the *de novo* method for the different species treatment revealing a new set of genes differentially expressed.



Keywords: RNA-seq, Inter-species Assembly, Gene expression, Liver steatosis

Sequencing

Illumina HiSeq 2000: 100bp paired-end x 80 libraries

Reference genome assembly

Available reference genome: Anas platyrhynchos BGI_duck_1.0, INSDC Assembly GCA_000355885.1

Assembly Adapter trimming ↔ **TopHat2 + Cufflinks + Cuffmerge**^[1] Nb sequences **41K** Nb residues **65M** N50 **2545** Figure 1 | Average of mapping rate on reference Reference genome is only available for Pekin and other genome in function of the species don't have a satisfying mapping rate on this species by TopHat2. Similar result is shown by mapping reference, reference genome assembly is limited by the reads back to reference mapping biases. genome assembly. De novo assembly Raw reads Mulard **Muscovy** Pekin Hinny Input read pairs 555M 514M 545M 639M

> Preprocessing: adapter removing, quality trimming and filtering Trimmomatic^[2]

Both surviving 98,34% 98,24% 98,23% 98,22%

	DEG	Pekin	Muscovy	Mule	Hinny	common
DRAP	up-regulated	2281	3450	4907	3901	539
transcriptome	down-regulated	1468	2717	4013	3795	364
	all	3749	6167	8920	7696	906
Cufflinks	up-regulated	1553	1371	1592	1314	520
transcriptome	down-regulated	680	773	953	924	235
	all	2233	2144	2545	2238	758
Increase (%)	up-regulated	147	252	308	297	104
DRAP vs Cufflinks	down-regulated	216	351	421	411	155
	all	168	288	350	344	120

Table 2 | comparison of differentially expressed genes (DEG) of reference genome and De novo assemblies. For both transcriptomes, statistical tests are realized at gene level by edgeR^[8] following a normalization step, then the results are filtered at p-value<0.05 and FC≥2. For DRAP assembly, methods DESeq2 and edgeR show similar results. As shown for Cufflinks assembly, 2281 and 1468 genes are found respectively up and down-regulated by overfeeding for Pekin, 758 DEG are in common for the 4 genotypes (also shown in Fig. 4C).

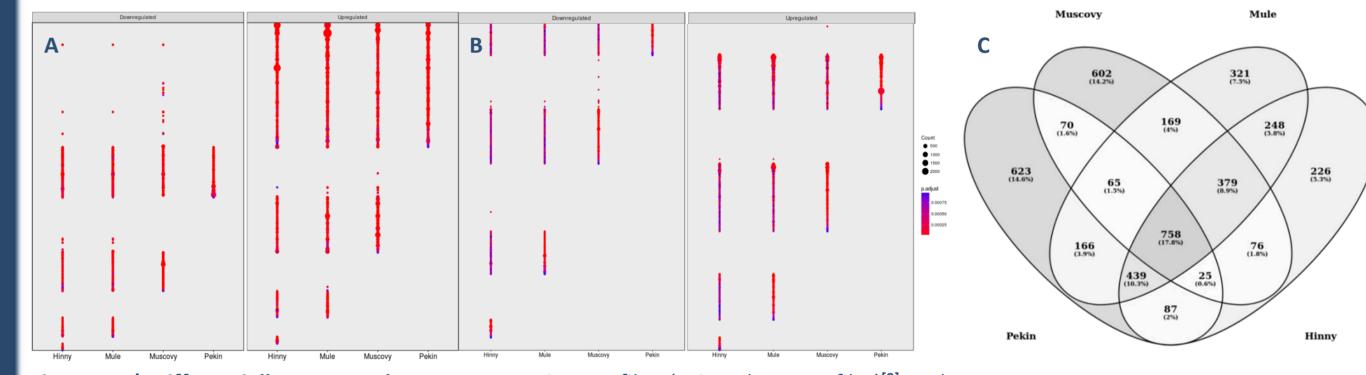
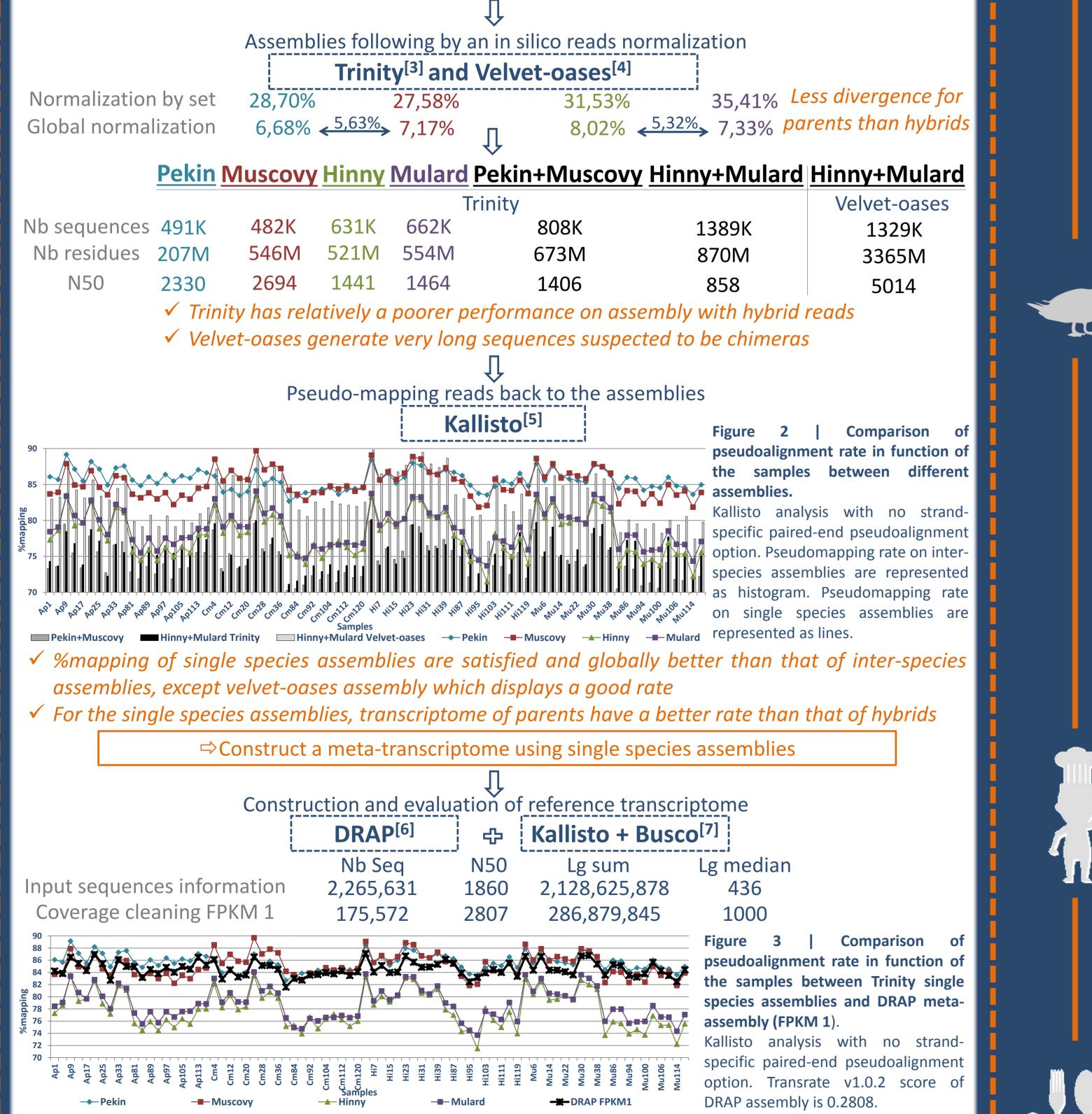
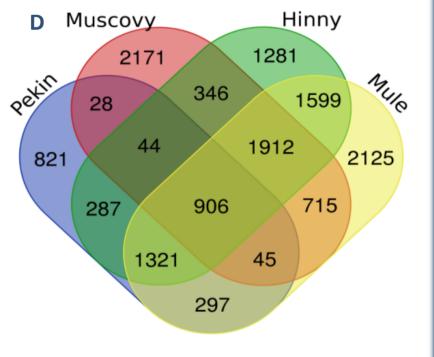


Figure 4 | Differentially expressed genes. Expression profiles (using clusterProfiler)^[9] and



Venn diagram^[10] for reference genome approach (A and C) and for *de novo* assembly (B and D). A and B: left side down-regulated DEG by overfeeding, right side up-regulated DEG by overfeeding. Dot size corresponds to the count of FC. The p-values are shown in color bar, values increase from hot to cold color.

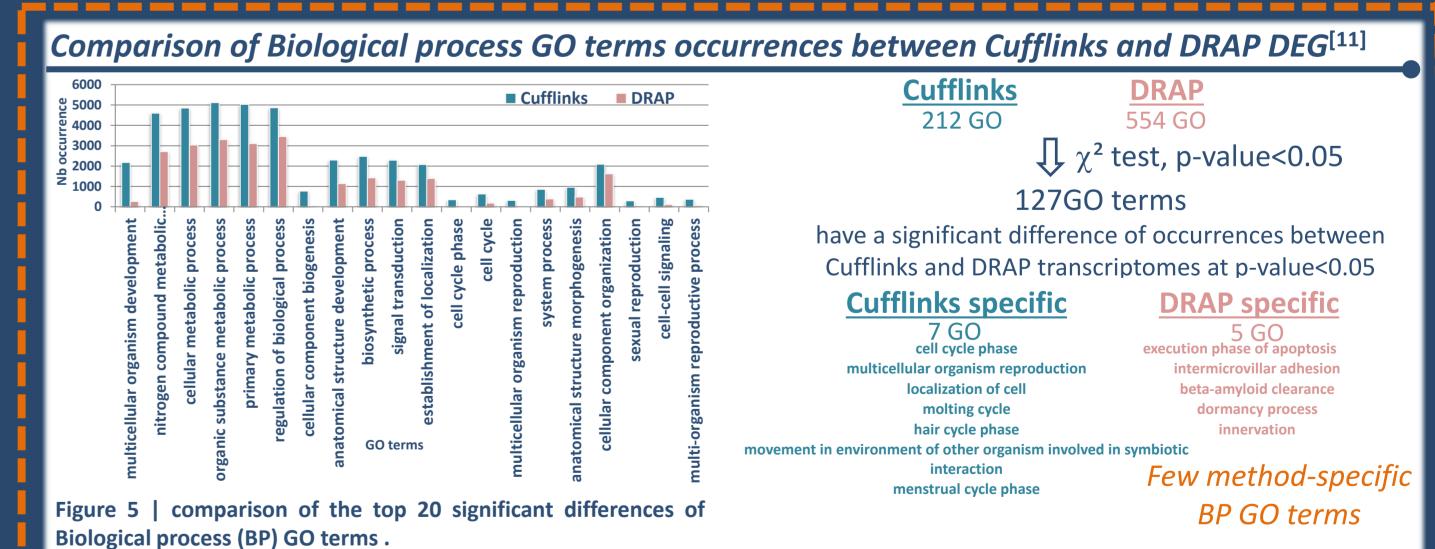
✓ Globally more DEG of de novo transcriptome (see Tab. 2) but a similar expression profiles (Fig. 4a and Fig. 4b): A slight increase of DEG for Pekin (168%) and especially great increase for the other samples. \Rightarrow Reference genome approach seems pertinent with homologues species for DEG identification, but not with heterologs.



✓ More DEG in common of the 4 genotypes for de novo assembly (34% vs 24%)

4. Differential analysis

Perspective



Significant BP GO terms occurrences using DRAP (annotated by Trinotate) are overestimated using Cufflinks. As there is more annotation with DRAP, our results suggest that there are more GO terms found overrepresented with DRAP, when excepted top 20 GO terms.

DRAP assembly possesses a satisfied pseudoalignment rate for all the samples

BUSCO categories	Reference	Cufflinks	DRAP	De novo approach discovers more		
buscu categories	genome	assembly	FPKM1	new genes (97% completeness),		
Complete	82.9%	81.5%	97.1%	provides a transcriptome more		
Complete and single-copy	81.2%	42.9%	59.1%	complete (0 missing) and less		
Complete and duplicated	1.7%	38.6%	38.0%	fragmented (only 3%).		
Fragmented	9.9%	11.2%	3.0%	DRAP assembly was chosen		
Missing	7.2%	7.3%	0.0%	as the reference transcriptome		
Table 1 BUSCO notation assessme	for further analyses.					

5. Functional annotation

✓ Focus on the new species-specific DEG discovered by de novo approach and Go-enrichment analyses which may bring new functions.

 \checkmark De novo approach bring us the opportunity to analyze genotype effect in addition to feeding effect. An ANOVA test will be realized to study the genotype and diet effect, also their interaction.

[1] Trapnell, Cole et al. Differential Gene and Transcript Expression Analysis of RNA-Seq Experiments with TopHat and Cufflinks: Nature Protocols 7.3 (2012): 562–578. PMC. Web. 26 June 2018. [2] Bolger, A. M. et al. Trimmomatic: A flexible trimmer for Illumina Sequence Data. Bioinformatics, btu170 (2014). [3] Grabherr, M. G. et al. Full-length transcriptome assembly from RNA-Seq data without a reference genome. Nat Biotech advance online publication, (2011). [4] Schulz MH, Zerbino DR, Vingron M, Birney E. Oases: robust de novo RNA-seq assembly across the dynamic range of expression levels. Bioinformatics 2012 Apr 15;28(8):1086-92. [5] Nicolas L Bray, Harold Pimentel, Páll Melsted and Lior Pachter, Near-optimal probabilistic RNA-seq quantification, Nature Biotechnology 34, 525–527 (2016), doi:10.1038/nbt.3519 [6] Cabau C, Escudié F, Djari A, Guiguen Y, Bobe J, Klopp C. (2017) Compacting and correcting Trinity and Oases RNA-Seq de novo assemblies. PeerJ 5:e2988. [7] Felipe A. Simão, Robert M. Waterhouse, Panagiotis Ioannidis, Evgenia V. Kriventseva, Evgeny M. Zdobnov; BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs, Bioinformatics, Volume 31, Issue 19, 1 October 2015, Pages 3210–3212, https://doi.org/10.1093/bioinformatics/btv351 [8] Robinson MD, McCarthy DJ, Smyth GK (2010). "edgeR: a Bioconductor package for differential expression analysis of digital gene expression data." Bioinformatics, 26(1), 139-140. [9] Yu G, Wang L, Han Y, He Q (2012). "clusterProfiler: an R package for comparing biological themes among gene clusters." OMICS: A Journal of Integrative Biology, 16(5), 284-287. doi: 10.1089/omi.2011.0118. [10] http://bioinformatics.psb.ugent.be/webtools/Venn/ [11] https://github.com/anlebreton/GO_terms_FisherTest

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