



HAL
open science

What is the relationship between the bioaccumulation of chemical contaminants in the variegated scallop *Mimachlamys varia* and its health status? A study carried out on the French Atlantic coast using the Path ComDim model

Marine Breitwieser, Evelyne Vigneau, Amélia Viricel, Vanessa Becquet, Camille Lacroix, Marina Erb, Valérie Huet, Carine Churlaud, Stéphane Le Floch, Benoit Guillot, et al.

► **To cite this version:**

Marine Breitwieser, Evelyne Vigneau, Amélia Viricel, Vanessa Becquet, Camille Lacroix, et al.. What is the relationship between the bioaccumulation of chemical contaminants in the variegated scallop *Mimachlamys varia* and its health status? A study carried out on the French Atlantic coast using the Path ComDim model. *Science of the Total Environment*, 2018, 640-641, pp.662 - 670. 10.1016/j.scitotenv.2018.05.317 . hal-01842065

HAL Id: hal-01842065

<https://hal.science/hal-01842065v1>

Submitted on 11 Dec 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Title**

2
3
4 What is the relationship between the bioaccumulation of chemical contaminants in the
5
6 variegated scallop *Mimachlamys varia* and its health status? A study carried out on the French
7
8 Atlantic coast using the Path ComDim model.
9

10
11 **Authors**

12
13
14 ^{1*} Marine Breitwieser, ² Evelyne Vigneau, ¹ Amélia Viricel, ¹ Vanessa Becquet, ³ Camille
15
16
17 Lacroix, ³ Marina Erb, ¹ Valérie Huet, ¹ Carine Churlaud, ³ Stéphane Le Floch, ⁴ Benoit Guillot,
18
19
20 ¹⁺ Marianne Graber, ¹⁺ Hélène Thomas
21
22
23
24
25
26

27 **Affiliations**

28
29 ¹ Littoral Environnement et Sociétés (LIENSs), UMR 7266, CNRS - Université de La
30
31 Rochelle, 2 rue Olympe de Gouges, F-17042 La Rochelle Cedex 01, France
32

33 ² StatSC, ONIRIS, INRA, 44322, Nantes, France
34
35

36 ³ Cedre, Centre de Documentation, de Recherche et d'Expérimentations sur les Pollutions
37
38 Accidentelles des Eaux, 715 rue Alain Colas, CS 41836, Brest, Cedex 2, France
39

40
41 ⁴ UMR CNRS 5805 EPOC-OASU-Université de Bordeaux, Allée Geoffroy Saint-Hilaire, CS
42
43 50023 33615 Pessac Cedex, France
44

45 ⁺ Equal contribution
46

47
48 * Corresponding author
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

56
57

58
59
60
61
62
63
64
65

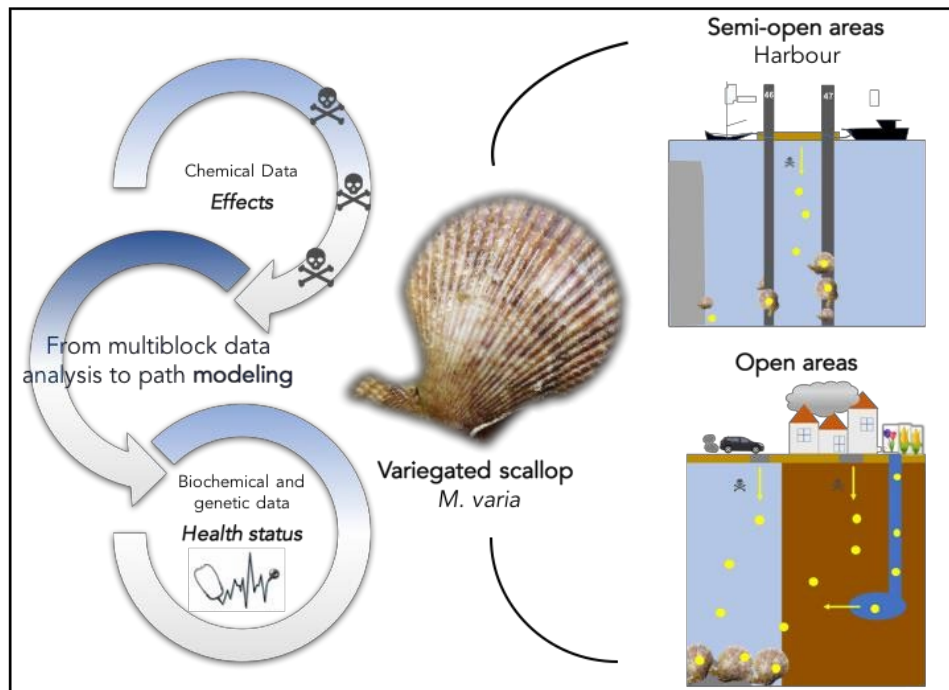
Abstract

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52

Increasing activity along the French Atlantic coast has led to chronic pollution with, in particular, mixtures of contaminants such as hydrocarbons, phytosanitary products, PCBs and heavy metals. Based on previous research, pollution biomarkers were used in this study as they can indicate health status when monitoring the impact of pollutants on coastal species such as the marine bivalve *Mimachlamys varia*. Mollusc bivalves were sampled in March 2016, in open and semi-open areas (a harbour zone), from thirteen sites which differed in terms of their level of pollution, and were located along the Atlantic coast from Brittany down to the Nouvelle-Aquitaine region. First, analyses of heavy metals and organic contaminants (e.g. pesticides, polycyclic aromatic hydrocarbons, polychlorobiphenyl) in the digestive gland of bivalves were performed. Second, biochemical assays were used to study defence biomarkers: oxidative stress with Superoxide Dismutase (SOD), detoxification of organic compounds with Glutathione-S Transferase (GST), lipid peroxidation with Malondialdehyde (MDA), and immune processes with Laccase. In addition to the biochemical assays, a genetic approach was used to measure genetic diversity (haplotype and nucleotide diversity) at each site. Biomarker assays and genetic diversity were correlated with the chemical contaminants in bivalves using the Path-ComDim statistical model. Our results showed specific correlations between biochemical assays in the digestive glands with heavy metal contaminants, and between genetic diversity and organic pollution. Blocks of responses were analysed for correlations in order to develop standardized tools and guidelines that could improve our understanding of the short-term and long-term impact of contaminants on physiological parameters.

53
54
55
56
57

Keywords: *Mimachlamys varia*, chronic chemical contamination, Atlantic coast, genetic diversity, biochemical assays, Path-ComDim model



Integrative approach undertaken in this study to assess effects of a chronic chemical contamination on natural populations of the marine bivalve *Mimachlamys varia*.
Photo credit: Thierry Guyot.

Highlights

- *Mimachlamys varia* was shown to be a useful bioindicator for environment quality assessment
- Scallops were collected from 13 areas along the French Atlantic coast, contaminated to different degrees with organic compounds and heavy metals
- Relationships between blocks of variables (contaminants, biomarkers of pollution, genetic diversity) were demonstrated with the Path-ComDim model
- The health of marine bivalves is modulated by polymetallic pollution
- Genetic diversity was negatively correlated to organic contamination

1 1. Introduction

2 Run-off water containing various chemical contaminants accumulates on the coast.

3 The degree of long-term chemical contamination of coastal waters may vary depending on the
4 effects of dilution as a result of tides, currents and climate variation. Since the 1970s
5 (Convention for the Prevention of Marine Pollution from Land-Based Sources, 1974) there
6 has been a growing awareness of the disastrous consequences of chemical contaminants on
7 our health and on the well-being of the environment.

8 Aside from the cross-disciplinary policies on issues related to the coast and the maritime
9 economy, framed by the French National Strategy for the Sea and the Coast and that was
10 introduced in 2016 (by the Minister for the Environment, Energy and the Sea in August
11 2016), the regulations concerning the quality of coastal waters are important both at the
12 national and the European Community level. The regulations pertaining to water quality
13 define the principles that must be adhered to by European Union member states in order to
14 achieve a good ecological water status by 2020. These principles are defined in the Marine
15 Strategy Framework Directive (MSFD), the Water Framework Directive (WFD), in measures
16 taken by the farming sector (better management of fertilisation, the nitrate directive, the *plan*
17 *Ecophyto*), and measures taken at the level of catchment areas. The aim is to reduce the use of
18 phytosanitary, heavy metal, polychlorobiphenyl and hydrocarbon products and to help the
19 local stakeholders, farmers and port managers to improve the quality of the water.

20 In this context, the aim of the present study was to assess the health of marine bivalves, which
21 are excellent sentinel organisms for measuring the quality of coastal environments (Grosell
22 and Walsh, 2006). Additionally, bivalves such as the variegated scallop *Mimachlamys varia*
23 are commercialised along the French coast. Initial *in situ* ecotoxicological studies were
24 carried out by our laboratory to assess the presence of pollutants in these organisms
25 (biomarkers of exposure) and/or to measure the impact of pollutants (biomarkers of effects)

26 on this species, using both biochemical markers and genetic diversity measurements
27 (Bustamante et al., 2005; Milinkovitch et al., 2015; Breitwieser et al., 2016, submitted-a).
28 Here, a biomonitoring study was performed on 13 sites in the Nouvelle-Aquitaine and
29 Brittany regions in March 2016 (Figure 1). Antioxidant enzyme activity (SuperOxyde
30 Dismutase biomarker), detoxication enzyme activity (Glutathion S-transferase biomarker),
31 lipid peroxidation (MalonDiAldehyde biomarker) and immunomodulation (Laccase
32 biomarker) were used as pollution markers. Malondialdehyde and SOD form part of the
33 antioxidant defences and have been used to show the impact of exposure to metals and a wide
34 range of organic compounds in the environment (Amiard-Triquet et al., 2013). Glutathione S-
35 transferases are involved in the phase II detoxification of organic contaminants, and also play
36 a protective role in oxidative stress by catalysing a selenium-dependent glutathione
37 peroxidase (Tappel et al., 1982; Prohaska, 1980). Lipid peroxidation in the MDA assay
38 indicates damage to cellular membrane lipids caused by ROS and is useful for assessing
39 exposure to and the effects of pollutants in marine species (Sureda et al., 2011; Capo et al.,
40 2015). Moreover, in bivalves such as oysters, mussels and variegated scallops, there is a
41 depletion of key enzymes such as laccase, which is involved in the modulation of the immune
42 system in the bivalve in the presence of different kinds of contaminants (Luna-Acosta et al.,
43 2010, 2011; Milinkovitch et al., 2015; Breitwieser et al., 2016). In addition, previous studies
44 on other aquatic organisms have shown that chemical contamination can cause genetic
45 damage at the molecular level and can affect population genetic diversity (e.g. Guttman,
46 1994; Gardeström et al., 2008). A decrease in genetic variability can result from a reduction in
47 population size due to an increased mortality rate and/or reduced fertility, and from natural
48 selection of individuals with more resistant genotypes (van Straalen et al., 2002; Nowak et al.,
49 2009; Bickham, 2011; Brady et al., 2017). As a result, investigating the genetic effects of

50 chemical contamination at the population level has been recognized as an important aspect in
51 impact assessment (Bickham et al., 2000).

52 In contrast to previous work investigating the link between chemical contaminants and
53 genetic diversity in variegated scallops (Breitwieser et al., 2016; Breitwieser et al., submitted-
54 a), the present study is able, for the first time, to relate both inorganic and organic pollutants
55 with genetic diversity measured in the same specimens across a large number of sites along
56 the Atlantic coast. In addition, our study uses an innovative statistical analysis (Path-
57 ComDim) to take into consideration the effect of chronic chemical pollution (pesticides,
58 polycyclic aromatic hydrocarbons, polychlorobiphenyl and heavy metals) on (1) genetic
59 diversity, and (2) the metabolic and physiological response through the combined use of
60 several biomarkers of pollution (SOD, GST, MDA, Laccase) of *M. varia* specimens collected
61 from 13 sites.

62 This study is an essential step toward monitoring the health of marine coastal environments
63 along the French Atlantic coast. Our work follows the recent implementation of different
64 strategies and operating procedures developed as part of cross-disciplinary policies on issues
65 relating to the coast and the maritime economy, as it provides a measure of environmental
66 quality through the use of a bioindicator marine bivalve.

67

68 **2. Materials and Methods**

69

70 *2.1- Field collection and tissue sampling*

71 The study sites are shown in Fig. 1. Scallops were collected from 13 sampling sites from
72 Brest down to the Nouvelle-Aquitaine Region. “Loix” was chosen as the less-contaminated
73 site. The open areas, “Troveoc” and “Plougastel”, which are situated in Brest Bay, are
74 subjected to anthropogenic pressure from urban activities and the Elorn estuary. “Erdeven”

75 and “Auray”, situated in southern Brittany, are subject to heavy contamination from
76 agriculture and pig farming. “Port-Neuf” is situated near a water-treatment plant and the
77 industrial area of La Rochelle. “Angoulins” may be exposed to currents and pollution from
78 adjacent urban tourist areas. “Aix Isle”, “Les Palles” and “Oleron” are situated across the
79 Charente estuary, which is impacted by sources of pesticide pollution, in particular from
80 treatment of the vines.

81 Finally, for the semi-open areas, “outlet dredging” is an area of outlet dredging near a
82 touristic beach and close to a harbour, “outlet rainwater” corresponds to the outlet of
83 rainwater in a harbour (without a water-filtering system), “outlet fairing” is located below the
84 careening area, which has a water-filtering system (carbon, sand, UV and paper to remove
85 organic, inorganic and bacterial contamination).

86 Tissue samples from variegated scallops (*M. varia*) were collected from the 13 sites described
87 above. For bioaccumulation and chemical biomarker assays, the digestive glands and gills
88 were sampled from 130 individuals (10 individuals per site) collected in March 2016 (adult
89 size mean of 4.04 ± 0.32 cm). For the genetic approach, muscle tissue was sampled from the
90 same 130 individuals and from additional variegated scallops collected at these sites in 2014
91 and 2016, to estimate population diversity indices. Thus, genetic data were obtained from 459
92 individuals (25 to 59 individuals per site) and these data are described in Breitwieser et al.,
93 submitted-a).

94 Sampling and protein assays were carried out in March 2016. For each sampling site, the
95 organs of the bivalves were removed *in situ* and stored at -80 °C for further analysis. Each
96 gill was cut into two samples and examined for trace elements and a biochemical assessment
97 of biomarkers was performed; each digestive gland was cut into two samples in order to
98 assess organic contaminants and perform a biochemical assessment of biomarkers
99 (Breitwieser et al., 2016). Samples for trace element analysis (gills) were freeze-dried for 48

100 h. For each scallop, muscle tissue was collected and stored in absolute ethanol for genetic
101 analysis.

102

103 2.2- *Assays of heavy metals*

104 Analyses of trace elements were based on previous work by Unrine et al. (2007). Assays
105 of Ag, As, Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, Se, Sn, V and Zn were performed on gills with a
106 Varian Vista-Pro ICP-OES and a Thermofisher Scientific XSeries 2 ICP-MS.

107

108 2.3- *Assays for organic contaminants*

109 The digestive glands of 7 scallops per site were analysed for 21 PAHs, 14 PCBs and 8
110 pesticides by stir bar sorptive extraction-thermal desorption-gas chromatography-tandem
111 mass spectrometry (SBSE-GC-MS/MS) using a method adapted from Lacroix et al. (2014).
112 The results are expressed as µg analytes/kg dry weight (d.w.). Limits of quantification (LOQ)
113 were calculated using the calibration curve method (Shrivastava et al., 2011) and the limit of
114 detection (LOD) was estimated by dividing LOQ by 3. Analytical quality control was made
115 using the Standard Reference Materials (1974c) “Organics in Mussel Tissue (*Mytilus edulis*)”
116 provided by the National Institute of Standards and Technology (NIST, Gaithersburg, USA).

117

118 2.4- *Biochemical assays*

119 2.4.1. *Sample processing*

120 For biomarker assessment in gills and digestive glands, samples were homogenized in
121 ice-cold phosphate buffer (100 mM, pH 7.2, 1100 mOsm). The homogenates were centrifuged
122 at 12,500g / 4 °C for 15 min and the Final Fractions (FF) were used for biochemical assays.

123

124 *2.4.2 Total protein concentration*

125 Total protein concentration was determined using an adaptation of the BCA Kit
126 method (Bicinchononique Acid Kit, Sigma Aldrich). The kit contained bovine serum albumin
127 (BSA) as a standard and involved the reduction of alkaline Cu²⁺ by proteins (Smith et al.,
128 1985) at an absorbance of 562 nm using a spectrofluorometer (SAFAS Flx-Xenius).

129

130 *2.4.3 Antioxidant enzyme activity*

131 SOD activity was determined in the FF using the method developed by Paoletti et al.
132 (1986). The results are presented in U of SOD/mg of protein.

133 GST activity in the organs was determined by adapting a Sigma assay kit method and
134 the results are expressed in U of GST/mg of protein (Mannervik et al., 1988; Habig et al.,
135 1974).

136

137 *2.4.4 Laccase assay*

138 Using a method developed by Luna-Acosta et al. (2010), this activity was measured in
139 the FF. The reaction was assessed at 420 nm using a spectrofluorometer (SAFAS Flx-
140 Xenius). In parallel, the non-enzymatic auto-oxidation (oxidation in the absence of FF) was
141 subtracted. Laccase-type PO activity is expressed as U of laccase/mg of protein (one unit is
142 defined as the amount of enzyme that catalyses the appearance of 1 µmol of product per min).

143

144 *2.4.5 Lipid peroxidation*

145 Damage, and in particular lipid peroxidation, was estimated by assessing the MDA
146 concentration in the final fraction. The product was quantified by measuring absorbance at
147 586 nm (Gerard-Monnier et al., 1998) using the SAFAS Flx-Xenius spectrofluorimeter. The
148 results are expressed in µM.mg protein⁻¹.

149

150 2.5- *Genetic approach*

151 Genetic diversity of scallops was assessed using sequences of a 444 base-pair portion of
152 the mitochondrial gene coding for cytochrome c oxidase I (cox1). DNA extraction, PCR and
153 sequencing protocols were described in Breitwieser et al. (2016) and Breitwieser al.
154 (submitted-a). The haplotype diversity (Hd), and nucleotide diversity (π) were calculated
155 using Arlequin version 3.5.2.2 (Excoffier and Lischer, 2010), for each sampling site
156 separately.

157

158 2.6- *Data processing*

159 First, preliminary statistical analyses were carried out using R v. 3.1.2 (R Core Team,
160 2016). In all cases, normality was tested on residuals using the Kolmogorov-Smirnov test and
161 homogeneity of variance was assessed using the Bartlett test. Moreover, a multi-correlation
162 analysis was performed to highlight correlations between chemical contaminant
163 concentrations and biomarker values.

164 Second, a multiblock statistical analysis with a path modelling approach was used to take
165 into account specific patterns of directed relations among the blocks of variables. In our case
166 study, five blocks of variables were considered: assays for heavy metals, assays for organic
167 contaminants, biochemical assays on the gills, biochemical assays on the digestive glands and
168 genetic data. The advantage of the multiblock data method of analysis used herein, named
169 ComDim (Mazerolles et al., 2002; Cariou et al., 2017), is to simultaneously exhibit common
170 dimensions shared by the blocks and introduce a system of weighting that allows each block
171 to have a specific weight for each common component. Moreover, as reported in Cariou
172 (2017), the combination of this multiblock analysis with a path modelling approach, known as
173 the Path-ComDim approach, takes into account the existence of an *a priori* specific pattern of

174 directed relations among the datasets at hand. Thus, Path-ComDim aims to reduce the
175 dimensionality of data organized in several blocks and to estimate the importance of the
176 assumed effects of one block on another block of information. For this analysis, we used
177 contaminant and biomarker data obtained from 91 of the collected specimens, and the genetic
178 diversity indices obtained for each site (estimated from 25 to 59 individuals per site) (Figure
179 2).

180 We considered a path diagram with arrows directed from heavy metals to biochemical
181 assays in the gills, digestive glands and the genetic data on the one hand, and organic
182 contaminants to biochemical assays in both organs and genetic diversity, on the other (Figure
183 2). These arrows reflect the fact that the chemical contamination measurements (heavy metals
184 and organic contaminants) are assumed to have an influence on the health status measured by
185 the biochemical assays and the genetic approach.

186 From a practical point of view, each block of data was subjected to a pre-treatment: at a
187 given site, organic contaminants for which assay values were regularly under the LOQ
188 (except for two or three observations) were summed. Thus, several PAH variables were
189 aggregated into a variable named “PAHothersum”, and several PCBs were aggregated into
190 the variable “PCBothersum”. All eight pesticides were also summed together. Afterwards all
191 the variables were individually standardized and each block was set at the same total variance
192 to account for the fact that the number of variables is different from one block to another.

193 **3. Results**

194 Data used in the Path-ComDim analysis are shown in the supplementary data (Tables
195 S1 and S2, Figure S1). For this analysis, we used heavy metals contaminants (Figure S1),
196 organic contaminants concentrations (Table S1), the genetic diversity indices obtained for
197 each site (Table S2) and biomarker data (Table S3).

198 Table 1 illustrates the results of the first three common components, obtained using
199 Path-ComDim. It turns out that the first two components (dim. 1 and 2) have an
200 overwhelming importance (47.9 % for dim. 1 and 34.5 % for dim. 2) in comparison to the
201 third component (7.0 % relative importance). Table 1 also shows the contribution of each pair
202 of linked blocks of variables to the definition of each common component. It appears that the
203 first common component (dim.1) mainly highlights the link between organic contaminants
204 and the genetic data (99.3%). The second common component mainly highlights the link
205 between heavy metals and biochemical assays in the digestive gland (83.8%), whereas a weak
206 relationship was observed between organic contaminants and biochemical assays in this organ
207 (8.6%). The links between heavy metals and biochemical assays in the gills, between heavy
208 metals and genetic diversity and between biochemical assays and organic contaminants in the
209 gills do not really contribute to the two first common components.
210 The third component mainly accounts for the link between heavy metals and biochemical
211 assays in the gills, but as the relative importance of this dimension is very small (0.4 %), this
212 result will not be taken into account in the following analysis.

213 From now on, we shall focus on the first two components, denoted t1 and t2. Figure 3
214 shows the configuration of the samples, in relation to their respective site, on the basis of t1
215 and t2. Figure 3 also shows correlations between the common components t1 and t2 and the
216 variables according to the block they belong to (heavy metals, organic contaminants,
217 biochemical assays in the gills and digestive gland, genetic approach). As each common
218 component is built to take account as much as possible of the relationship between the blocks
219 of variables, the subplots 3b to 3f in Figure 3 make it possible to illustrate the links between
220 the blocks of variables depicted in Table 1.

221 Concerning the block of heavy metal variables (Figure 3b), three clusters of heavy
222 metals can be observed along the second component, t2, namely Cu/Se/Zn/As, Cd/Ag, and

223 Pb/Cr/Co/Fe/Mn/Ni. This partition was confirmed using a clustering approach for variables
224 (Vigneau et al., 2015). The Sn and V values were not taken into account because of the lower
225 quantification with respect to the LOD value. The three clusters of heavy metals seem to be
226 associated with location. Indeed, the sites “Outlet rainwater” and “Outlet Fairing”, which are
227 situated in a harbour area, could modulate the Cu/Se/Zn/As cluster, whereas “Outlet
228 dredging” and the majority of the sites situated near the Charente Estuary (“Oleron”,
229 “Angoulins”, “Port-Neuf”, “les Palles”) could influence the Pb/Cr/Co/Fe/Mn/Ni cluster.

230 Moreover, it was interesting to show (Table 1) the strong contribution link between
231 heavy metals and biochemical assays in the digestive glands. Indeed, the Path-ComDim
232 model showed that the biochemical values for the digestive gland had a common dimension
233 (were of a similar order of magnitude) on axe 2 (Figure 3e). In the digestive glands, there was
234 a positive correlation between the biomarkers GST and MDA, but a negative correlation
235 between the group GST/MDA and the Laccase biomarker. The SOD biomarker involved in
236 oxidative stress did not correlate with the other pollution biomarkers (laccase, MDA, GST).

237 The organic contaminants (pesticides, hydrocarbons, polychlorobiphenyls) are well
238 depicted along the common dimension t1. The correlation plot on Figure 3c shows that the
239 Polycyclic Aromatic Hydrocarbon (PAH) variables (except biphenyl), several PCB variables
240 (PCB28, PCB77 and the sum of the rare PCBs) and the variable summing all the pesticide
241 assays correlated well (negatively) with t1. All organic contaminants are situated to the left-
242 hand side of the plot, and in greater quantity in the “Plougastel” site (for pesticides and a rare
243 PCB group named “PCB other sum” in Figure 3c), “Outlet fairing”, “Outlet rainwaters” and
244 “Outlet dredging” sites concerning PCB 28, PCB 77 and all hydrocarbons (Figures 3.a and
245 3.c).

246

247 The two genetic variables on Figure 3f (haplotype diversity and nucleotide diversity)
248 are positively correlated with t1 and are expressed more in sites such as “Loix”, “Angoulins”,
249 “Les Palles” and “Oleron” on the right-hand diagram (Figure 3a). The genetic variables are
250 not correlated with t2 and thus seem not to be linked with heavy metal contaminants (Figure
251 3b). In contrast, t1 illustrated the negative relationship between organic contaminants (Figure
252 3c) and the genetic block (Figure 3f). The greater the quantity of organic contaminants, the
253 lower the indices of genetic diversity (haplotype diversity, nucleotide diversity). This link
254 contributed greatly to t1, as already shown in Table 1.

255

256 It is interesting to note that the results of the biochemical assays on the digestive
257 glands (Figure 3e) were different to those in the gills (Figure 3d). Indeed, it seems that there
258 was a significant background noise in the results of the biochemical assays on the gills. As a
259 consequence, the block of biochemical assays on the gills appeared to have no strong links
260 with the other blocks.

261 These analyses are particularly interesting in comparison with heavy metals, which had a link
262 contribution of 83.8% on common dimension 2 (Table 1). Indeed, a greater contamination of
263 Co, Cr, Pb and Ni led to a positive immunomodulation in the “Outlet dredging” site, whereas
264 a decrease in the specific activity of laccase was influenced by As and Se at “Plougastel”.
265 Moreover, the higher contamination with As and Se induced a higher MDA and GST
266 response, both of which are involved in lipid peroxidation and the detoxification of xenobiotic
267 compounds, unlike laccase.

268

269 **4. Discussion**

270 In this work, an innovative analysis of the effects of chemical contamination (inorganic
271 and organic pollutants) on the biology of the variegated scallop *M. varia* was carried out using

272 a statistical approach involving the Path-ComDim model to: i) clearly study responses to both
273 contaminants (heavy metals and organic pollutants); ii) identify the biochemical and genetic
274 effects for each type of contaminant; and iii) understand the combined effect of the
275 contaminants on health status.

276

277 4.1- Chemical contaminants

278 Our study was carried out using scallops collected *in situ* in March 2016, and for each
279 specimen, the level of contamination with heavy metals and organic pollutants was estimated
280 in the gills and digestive glands. A small number of bio-monitoring investigations of coastal
281 waters have been carried out by the local stakeholders on the Atlantic coast to evaluate the
282 environment quality (e.g. the water department of the local authorities, the harbour
283 authorities). Using the variegated scallop *M. varia*, our results showed higher concentrations
284 of Pb, Cr, Fe, Ni, Co in the gills of this bivalve in the Pertuis Charentais (“Oleron”,
285 “Angoulins”, “Port-Neuf”, “Les Palles” sites) and in particular in “Outlet dredging” zones in
286 La Rochelle harbour in comparison with the least-contaminated area, “Loix”, described as a
287 reference site in the literature (Bustamante and Miramand, 2005). Moreover, Cu and Zn
288 concentrations were higher in “Troveoc”, “Erdeven”, “Auray” and “Outlet rainwater” than in
289 “Loix”. These results confirm the hypothesis that the Atlantic coast is contaminated by trace
290 elements from anthropogenic activities in wastewaters and inputs from drainage areas directly
291 adjacent to the sea, and watershed-derived inputs (e.g erosion of contaminated land, effluents
292 from highly-urbanized rivers, domestic and/or industrial effluents from wastewater treatment
293 plants, urban runoff and sewers). Furthermore, the capacity of heavy metal contaminants to
294 bio-accumulate in bivalve molluscs (Bryan, 1973) depends on the main function of each
295 organ i.e. trapping nutrients (gills), or digesting and excreting nutrients (digestive gland).
296 Indeed, previous studies have shown a greater bioaccumulation of chemical contaminants in

297 the digestive gland in comparison with the gills (Breitwieser et al., submitted-b). However, it
298 is important to underline the difference between non-essential and essential metals: non-
299 essential heavy metals such as As, Pb, Co, Cd could be more toxic for the scallop (Rainbow et
300 al., 2002) and on a larger scale, for Human. In fact, these trace elements are used particularly
301 in industrial sectors, wood preservatives, electrical battery manufacturing, alloy production,
302 and are classified in the first group of carcinogenic compounds (International Agency for
303 Research on Cancer). According to a report by the National Institute for Public Health (Tome
304 2: heavy metals and metalloids, December 2017), their major impregnation by human could
305 be due to seafood consumption.

306 Organic contaminant levels in the digestive gland of *M. varia* scallop provided
307 information on anthropogenic sewage in 13 separate sites on the Atlantic coast. In this work,
308 the data obtained showed high levels of pesticides in Plougastel (sum of dieldrin, aldrin,
309 isodrin, endrin, 2-4 dde, 4-4 dde and rare PCBs (52, 101, 135, 118, 153, 105, 138, 156, 180,
310 169)) collected together in a single variable (Figure 3c). This might be explained by the slow
311 and insidious degradation of these components, and in particular PCBs, which are now
312 prohibited because they are responsible for acute toxicity. Concerning PCBs 28 and 77,
313 measurements in the digestive glands of *M. varia* indicated that the mean of these two PCBs
314 was significantly higher in Plougastel, outlet rainwater and fairing areas. Moreover,
315 measurements of pyrene and other hydrocarbon congeners in the digestive glands showed
316 anthropogenic pressure (S1 Table of supplementary data). These results might be due to the
317 proximity of urban zones along the Atlantic coast, despite the fact that all of these
318 contaminants are still considered as a priority by the WFD in France.

319 Indeed, on the correlation plot on Figure 3c, all the contaminants are situated on the
320 left of the plot in the common dimension 1. All the organic components could potentially
321 originate from various activities in these sectors (fishing harbours, tourist harbours, petroleum

322 refinery, waste treatment plant) with tidal currents, particularly from the Gironde Estuary and
323 from Brittany (Luna-Acosta et al., 2017). Moreover, previous research indicated that the
324 concentration of pesticides, PCBs and PAHs in sediments was higher than in the surrounding
325 water, and this might account for the observation in the digestive glands of *M. varia*, which is
326 a sedentary benthic species (Luna-Acosta et al., 2017).

327 These data are in agreement with previous results showing that other marine bivalves
328 (mussels, oysters and scallops) collected on the Atlantic coast have a strong ability to ingest
329 and bio-accumulate more hydrophobic high molecular weight PAHs and heavy metals in their
330 digestive organs than in other organs as gills and mantle (Luna-Acosta et al., 2017;
331 Breitwieser et al., 2017, Lacroix et al., 2017).

332

333 4.2- *Health status response*

334 Because benthic marine molluscs are exposed to multiple stressors such as contaminant
335 cocktails and it is not always possible to test for their presence within tissues, there has been a
336 search for indicators that demonstrate the toxicity of chemical pollutants. The advantage and
337 originality of our study is that indicators of health status and contaminants were assessed
338 using the Path-ComDim statistical model in distinct tissues. As previously suggested in *M.*
339 *varia*, indicators of health status are pollutant biomarkers for the short-term (GST, MDA,
340 SOD, Laccase). It is for this reason too that biomarkers are currently being used to assess the
341 condition of European coastal waters (Hagger et al., 2009).

342 From the results obtained with the Path-ComDim model, the biomarkers in the gills of the
343 variegated scallop were not modulated, in contrast to the biomarkers in the digestive gland
344 and the genetic diversity indices. This may be linked to the fact that the ability to deal with
345 oxidative stress, detoxification and immunomodulation revealed by pollution biomarkers may
346 depend on the accumulation and the kind of contamination (Hannam et al., 2010).

347 In this work, the biomarkers of oxidative stress used were SOD and MDA. It was
348 demonstrated that these indicators could provide some of the first signals of stress in molluscs
349 (Regoli et al., 2000; Valavanidis et al., 2006). Furthermore, GST is a phase II conjugation
350 enzyme that intervenes in the metabolism of xenobiotics and in reactions which are catalysed
351 by membranes, or cytosolic enzymes. According to previous studies, GST-specific activity
352 increases in exposed organisms in relation to the concentration of xenobiotics in the medium.
353 Indeed, GSTs, which correlated positively with MDA in this study, are enzymes whose
354 activity is used as a biomarker of exposure to organic substances, especially in molluscs,
355 where EROD activity (a marker of cytochrome P450 involved in the metabolism of
356 potentially toxic compounds) is not routinely assessed (Cajaraville et al., 2000). Laccase
357 proved to be a sensitive biomarker and phenoloxidases seem to play an important role in
358 bivalve immunity when they are activated by ROS (Luna-Acosta et al., 2010). Moreover,
359 Laccase contains metal coenzymes which can be affected by inorganic contamination (heavy
360 metals) in a similar fashion to the activation pathway of SOD in the short term, which
361 explains the contrasting response of this enzyme activity to that of the other biomarkers, GST
362 and MDA.

363 We proposed to use genetic diversity as an indicator of population health because
364 populations with high levels of genetic variation tend to have a better viability (Frankham et
365 al., 2002), a better capacity to resist environmental stress (e.g. Nowak et al., 2012) and
366 possess a greater adaptive potential when facing environmental changes (Eizaguirre and
367 Baltazar-Soares, 2014). Since historical processes and dispersal (i.e. gene flow) influence the
368 genetic diversity of populations, information on the species phylogeography and population
369 structure is required to interpret differences in genetic diversity across sampling sites
370 (Theodorakis, 2003). With that goal, we recently assessed the genetic structure of *M. varia*
371 using *cox1* sequences and did not detect genetic differentiation among sites along the French

372 coasts of the Bay of Biscay and English Channel (Breitwieser et al., submitted-a). Results
373 based on this mitochondrial DNA marker suggest that all individuals sampled in the present
374 study belong to the same population and that differences in genetic diversity observed among
375 our sampling sites (Supplementary Table S2) are not due to genetic drift or historical events.

376 Different regions of France and many countries of the European Union have established
377 an integrated ecosystem-based approach to the management of marine resources. Any
378 assessment of the cumulative short-term or long-term effect of all the activities that can
379 potentially impact an environment that is prone to large natural variations is a relevant but
380 difficult task. In recent years, pragmatic approaches have been initiated by urban authorities,
381 and local and regional stakeholders of Charente-Maritime (France). The aim of these
382 voluntary actions is to monitor quantitative chemical concentrations but not to study the
383 qualitative effects on environmental quality. Today, several pollutant biomarkers are being
384 employed in biomonitoring studies but the results are difficult to compare. This is why our
385 research was focused on refining and standardising current biomarker techniques for specific
386 contaminant types using a statistical model as this will allow comparisons to be made.

387

388 *4.3- Relationship between health status and chemical contaminants*

389 Thanks to the Path-ComDim model, a comprehensive analysis of our results could be
390 performed. Based on previous research, the discussion will be divided into two parts to
391 explain the relationship between health status and contaminants in this study. In contrast to
392 others studies (Breitwieser et al., 2016), all heavy metals and organic pollutants were
393 considered in the model because all of the data come from the same 91 specimens, which is a
394 relevant approach.

395 Concerning biomarkers of pollutants, only the results in the digestive gland showed a
396 significant response with heavy metal contaminants. Our model accounts for 83.8 % of link

397 contribution, suggesting that MDA, which reflects lipid peroxidation, and GST, which shows
398 detoxification of xenobiotics, were positively correlated and were modulated by Cu, Se, As
399 and Zn. Furthermore, based on the model, these four heavy metals, and a group of specific
400 organic contaminants (pesticides, and PCBs 28 and 77), could significantly diminish Laccase
401 activity, in agreement with previous work (Luna-Acosta et al., 2017; Luna-Acosta et al.,
402 2015; Milinkovitch et al., 2015). These results could explain the high functionality of the
403 xenobiotic bio-transformation process by GST on the one hand, and also the decrease in anti-
404 oxidant protection and immune modulation by an increase in MDA and a decrease in Laccase
405 on the other hand (Pan et al., 2005; Pan et al., 2009; Breitwieser et al., 2016). These enzyme-
406 specific activities and concentration levels could be related to the high contaminant
407 concentrations in the harbours, outlet rainwater and touristic areas (Breitwieser et al., 2017).
408 However, it should not be forgotten that abiotic factors could modulate biomarker activities
409 and bioaccumulation. Indeed, data on physico-chemical parameters collected at the same date
410 are shown on a map of work presented in Breitwieser et al., (submitted-a). Only a significant
411 difference in salinity was discernible between Erdeven, Auray and all other sites, but these
412 two sites are mixed with others on the plot of sites (Figure 3a).

413 Concerning genetic effects of pollution, it has already been stressed that chemical
414 contaminants can reduce genetic diversity (e.g. Nowak et al., 2009), or induce deleterious
415 mutations that can be heritable (e.g. Cronin and Bickham, 1998), and per consequence, can
416 impact the population health (Bickham et al., 2000; Bickham, 2011). However, some studies
417 have observed greater genetic diversity in polluted environments (Pedrosa et al., 2017)
418 illustrating that the relationship between chemical contamination and genetic diversity is not
419 straightforward and will depend on the species demography, dispersal, and level of
420 contamination.

421 In our study, low levels of genetic diversity were clearly correlated with high levels of
422 organic contaminants in variegated scallops from outlets fairing, dredging and Plougastel.
423 Since gene flow does not seem to be restricted among our sampling sites (Breitwieser et al.,
424 submitted-a), a reduction in genetic diversity at more polluted sites cannot be explained by a
425 population bottleneck. Conversely, selective mortality of individuals possessing
426 mitochondrial haplotypes susceptible to organic pollutants is plausible. Indeed, mitochondria
427 seem to be particularly vulnerable to environmental contaminants (Meyer et al., 2013 and
428 references therein), therefore natural selection may be acting on the mitochondrial genome in
429 organisms exposed to chemical contaminants.

430 To further test our hypothesis, extensions of this study could include i) obtaining genetic
431 diversity estimates from neutral markers (nuclear loci not under natural selection) and
432 conducting a more fine-scale investigation of gene flow among sites using multiple
433 independent genetic markers such as microsatellites, and ii) assessing experimentally
434 differential mortality of distinct mitochondrial haplotypes following organic contaminant
435 exposure. Additionally, genome-wide data could be used to investigate local adaptation to
436 pollution (Oziolor et al., 2017), which may occur despite gene flow (Tigano and Friesen,
437 2016). Finally, our study focused on the adult stage *in situ*, but chemical contamination also
438 affects early life stages (larvae and juveniles) in bivalves (e.g. Calabrese et al., 1977). Thus, a
439 multigenerational controlled experiment submitting variegated scallops to chemical
440 contaminants, and measuring mortality and genetic diversity of the different life-stages would
441 provide further insights on the resistance and potential adaptation of this bioindicator species
442 to contamination.

443 Overall, the statistical model used for monitoring health status is relevant. Indeed,
444 contrary to other models such as PLS-Path Modelling (Tenenhaus et al., 2005; Bertrand et al.,
445 2017) which extract one dimension by block and fit a unique common model, the Path-

446 ComDim model produces several common dimensions, each of them defining specific
447 weights for the blocks with regard to the links investigated. Furthermore, the graphical output
448 of Path-ComDim can be read like a principal component analysis (PCA). This leads to rather
449 simple and interpretable media for the scientific community.

450

451 **5. Conclusion**

452 In conclusion, the maritime economy serves a wide range of diversified activities, both
453 traditional (fishing, shell fishing, harbour activities, tourism etc.) and emergent (renewable
454 marine energy, biotechnology etc.). In addition to these activities, which are the mainstay of
455 the maritime economy, indirect activities such as logistics, industrial port zones and transport
456 have to be considered. In France, and in particular in the majority of our study sites, the south
457 Atlantic seaboard, with its 720 kilometres of coastland and the Charente-Maritime islands, is
458 an attractive space. Out of interest, the population is constantly growing in the French South-
459 Atlantic, at a rate of 9.5% between 1999 and 2011, in comparison with the population growth
460 in France, which was 7.9% during the same interval.

461 In recent years, shellfish farmers have experienced repeated crises, and this has raised
462 serious issues. France is the biggest oyster producer and the third biggest mussel producer in
463 Europe according to the “Comité National de la Conchyliculture, [http://www.cnc-](http://www.cnc-france.com)
464 [france.com](http://www.cnc-france.com)”. Apart from the health risk, a decrease in the quality of the water, the living
465 environment of marine species, would damage marine environments, biodiversity and the
466 development of economic stakeholders (the aquaculture sector, fishing and also tourism). So,
467 it is now crucial to include an assessment of coastal water quality no less frequently than
468 annually. Indeed, using new tools such as Path-ComDim applicated in this study, it is now
469 necessary to address coastal water contamination; this has been undertaken by some harbour

470 authorities, in particular in La Rochelle, which created a Department of the Environment 10
471 years ago.

472

473 **Acknowledgements**

474 The authors would like to thank the people involved in sampling: V. Huet, A. Combaud, E.
475 Dubillot, T. Guyot, P. Pineau, N. Lachaussée, J. Pigeot, C. and P. Follenfant. We would also
476 like to acknowledge Marine Barbarin, Melvin Le Bihan and Elodie Boussinet for help
477 performing the biochemical assays. Finally, the authors would like to thank Frank Healy for
478 proof-reading the manuscript.

479

480 **Funding:** This work was supported by the PECTIMPACT Project of the University of La
481 Rochelle; the INCREASE CNRS Federation, and by a research grant from the “Communauté
482 d’Agglomération of La Rochelle”.

483

484 **Bibliography**

485 Amiard-Triquet C., Amiard J.C., Rainbow, P., 2013. Ecological Biomarkers: Indicators of
486 Ecotoxicological Effects. CRC Press, Taylor and Francis Group, International Standard
487 Book Number 978-1-4398-8017-3 (Hardback).

488 Bertrand, C., Devin, S., Mouneyrac, C., Giambérini, L., 2017. Eco-physiological responses to
489 salinity changes across the freshwater-marine continuum on two euryhaline bivalves:
490 *Corbicula fluminea* and *Scrobicularia plana*. Ecol. indicators 74, 334-342.

491 Bickham, J.W., Sandhu, S., Hebert, P.D.N., Chikhi, L., Athwal, R., 2000. Effects of chemical
492 contaminants on genetic diversity in natural populations: implications for biomonitoring
493 and ecotoxicology. Mutat Res. 463(1), 33–51.

494 Bickham, J.W., 2011. The four cornerstones of Evolutionary Toxicology. *Ecotox.* 20,497-
495 502.

496 Brady, S.P., Monosson, E., Matson, C.W., Bickham, J.W., 2017. Evolutionary toxicology:
497 Toward a unified understanding of life's response to toxic chemicals. *Evol. Applications*
498 10, 745-751.

499 Breitwieser, M., Viricel, A., Graber, M., Murillo, L., Becquet, V., Churlaud, C., Fruitier-
500 Arnaudin, I., Huet, V., Lacroix, C., Pante, E., Le Floch, S., Thomas-Guyon, H., 2016.
501 Short-term and long-term biological effects of chronic chemical contamination on natural
502 populations of a marine bivalve. *PLoS ONE* 11 (3), e0150184.

503 Breitwieser, M., Viricel, A., Churlaud, C., Guillot, B., Martin, E., Stenger, P-L., Huet, V.,
504 Fontanaud, A., Thomas-Guyon, H. 2017. First data on three bivalve species exposed to
505 an intra-harbour polymetallic contamination (La Rochelle, France). *Comp. Biochem. and*
506 *Physiology, Part C, 199C*, 28-37.

507 Breitwieser, M., Becquet, V., Thomas-Guyon, H., Pillet, V., Sauriau, P-G., Graber, M.,
508 Viricel, A. (submitted-a). Genetic evidence for a single population of variegated scallop
509 *Mimachlamys varia* across a biogeographic break on the French coastline. *Submitted to J.*
510 *Mollus. Stud.*

511 Breitwieser, M., Huet, V., Barbarin, M., Dubillot, E., Graber, M., Thomas, H. (submitted-b).
512 Comparative biomarkers study in two scallops' organs to establish guidelines for
513 evaluating French Atlantic coastlines water quality. *Short research note submitted to*
514 *Mar. Poll. Bulletin.*

515 Bryan, G.W., 1973. The occurrence and seasonal variation of trace metals in the scallops
516 *Pecten maximus* (L.) and *Chlamys opercularis* (L.). *J. Mar. Biol. Assoc. U. K.* 53 (01),
517 145–166.

518 Bustamante, P., Miramand, P., 2005. Evaluation of the variegated scallop *Chlamys varia* as a
519 biomonitor of temporal trends of Cd, Cu, and Zn in the field. *Environ. Pollut.* 138 (1),
520 109–120.

521 Cajaraville, M.P., Bebianno, M.J., Blasco, J., Porte, C., Sarasquete, C., Viarengo, A., 2000.
522 The use of biomarkers to assess the impact of pollution in coastal environments of the
523 Iberian Peninsula: A practical approach. *Sci. Total Environ.* 247, 295-311.

524 Calabrese, A., Mac Innes, J.R., Nelson, D.A., Miller, J.E., 1977. Survival and growth of
525 bivalve larvae under heavy-metal stress. *Mar Biol.* 41(2), 179–84.

526 Capo, X., Tejada, S., Box, A., Deudero, S., Sureda, A., 2015. Oxidative status assessment of
527 the endemic bivalve *Pinna nobilis* affected by the oil spill from the sinking of the Don
528 Pedro. *Marine Environm. Res.* 110, 19-24.

529 Cariou, V., Qannari, E.M., Rutledge, D.N., Vigneau, E., 2017. ComDim: from multiblock
530 data analysis to path modeling, *Food Qual. Prefer.*, doi:
531 <http://dx.doi.org/10.1016/j.foodqual.2017.02.012>

532 Cronin, M.A., Bickham, J.W., 1998. A population genetic analysis of the potential for a crude
533 oil spill to induce heritable mutations and impact natural populations, *Ecotoxicol.* 7, 259
534 – 278.

535 Eizaguirre, C., Baltazar-Soares, M., 2014. Evolutionary conservation—evaluating the
536 adaptive potential of species. *Evol Appl*, 7: 963–967. doi:10.1111/eva.12227.

537 Excoffier, L., Lischer, H.E.L., 2010. Arlequin suite ver 3.5: a new series of programs to
538 perform population genetics analyses under Linux and Windows. *Molecular Ecology*
539 *Resources*, 10: 564–567. doi: 10.1111/j.1755- 0998.2010.02847.x PMID: 21565059.

540 Frankham, R., Briscoe, D.A., Ballou, J.D., 2002. *Introduction to Conservation Genetics*.
541 Cambridge University Press.

542 Gardeström, J., Dahl, U., Kotsalainein, O., Maxson, A., Elfwing, T., Grahn, M., Bengtsson,
543 B.E., Breitholtz, M., 2008. Evidence of population genetic effects of long-term exposure
544 to contaminated sediments—A multi-endpoint study with copepods. *Aquatic Toxicology*
545 86, 426-436.

546 Gerard-Monnier, D., Erdelmeier, I., Chaudiere, J., Yadan, J.C., 1998. Method of colorimetric
547 analysis of malonic dialdehyde and 4-hydroxy-2-enaldehydes as indexes of lipid per-
548 oxidation, kits for carrying out said method, substituted indoles for use in said method
549 and their preparation. US Patent No. US5726063. United States Patent and Trademark
550 Office, Washington, DC.

551 Grosell, M., Walsh, P.J. 2006., Sentinel species and animal models of human health.
552 *Oceanography* 19, 126-133.

553 Guttman, S.I., 1994. Population genetic structure and ecotoxicology. *Environ Health Perspect.*
554 102(Suppl 12): 97–100.

555 Habig, W.H., Pabst, M.J., Jakoby, W.B., 1974. Glutathione S-transferases. The first
556 enzymatic step in mercapturic acid formation. *J. Biol. Chem.* (section 24).

557 Hagger, J.A., Galloway, T.S., Langston, W.J., Jones, M.B., 2009. Application of biomarkers
558 to assess the condition of European marine sites. *Environ. Pollut.* 157, 2003-2010.

559 Hannam, M.L., Bamber, S.D., Moody, A.J., Galloway, T.S., Jones, M.B., 2010.
560 Immunotoxicity and oxidative stress in the Arctic scallop *Chlamys islandica*: effect of
561 acute oil exposure. *Ecotoxicol. Environ. Saf.* 74, 1440-1448.

562 Lacroix, C., Le Cuff, N., Receveur, J., Moraga, D., Auffret, M., Guyomarch, J., 2014.
563 Development of an innovative and green stir bar sorptive extraction–thermal desorption–
564 gas chromatography–tandem mass spectrometry method for quantification of polycyclic
565 aromatic hydrocarbons in marine biota. *J. Chromatogr. A* 1349, 1–10.

566 Lacroix, C., Duvieilbourg, E., Guillou, N., Guyomarch, J., Bassoulet, C., Moraga, D.,
567 Chapalain, G., Auffret, M., 2017. Seasonal monitoring of blue mussel (*Mytilus spp.*)
568 populations in a harbor area: A focus on responses to environmental factors and chronic
569 contamination. *Mar. Environ. Res.* 129, 24-35.

570 Luna-Acosta, A., Rosenfeld, E., Amari, M., Fruitier-Arnaudin, I., Bustamante, P., Thomas-
571 Guyon, H., 2010. First evidence of laccase activity in the Pacific oyster *Crassostrea*
572 *gigas*. *Fish Shellfish Immunol.* 28 (4), 719–726.

573 Luna-Acosta, A., Saulnier, D., Pommier, M., Haffner, P., De Decker, S., Renault, T.,
574 Thomas- Guyon, H., 2011. First evidence of a potential antibacterial activity involving a
575 laccase- type enzyme of the phenoloxidase system in Pacific oyster *Crassostrea gigas*
576 haemocytes. *Fish and Shellfish Immunol.* 31 (6), 795–800.

577 Luna-Acosta, A., Bustamante, P., Budzinski, H., Huet, V., Thomas-Guyon, H., 2015.
578 Persistent organic pollutants in a marine bivalve on the Marennes–Oléron Bay and the
579 Gironde Estuary (French Atlantic Coast)—Part 2: Potential biological effects. *Sci Total*
580 *Environ.* 514, 511–522. doi: 10.1016/j.scitotenv. 2014.10.050.

581 Luna-Acosta, A., Breitwieser, M., Renault, T., Thomas-Guyon, H., 2017. Recent findings on
582 Phenoloxidases in Bivalves. *Mar. Poll. Bull.* 122, 5-16.

583 Mannervik, B., Helena-Danielson, U., Ketterer, B., 1988. Glutathione transferases-structure
584 and catalytic activity. *Crit. Rev. Biochem. Mol. Biol.* 23 (3), 283–337.

585 Mazerolles, G., Devaux, M. F., Dufour, E., Qannari, E. M., Courcoux, P., 2002. Chemometric
586 methods for the coupling of spectroscopic techniques and for the extraction of the
587 relevant information contained in the spectral data tables. *Chemom. and Intell. Lab.*
588 *Systems*, 63, 57–68.

589 Meyer, J.N., Leung, M.C.K., Rooney, A.S., Hengartner, M.O., Kisby, G.E., Bess, A.S., 2013.
590 Mitochondria as a target of Environmental Toxicants. *Toxicol. Scien.* 134, 1-17.

591 Milinkovitch, T., Bustamante, P., Huet, V., Reigner, A., Churlaud, C., Thomas-Guyon, H.,
592 2015. In situ evaluation of oxidative stress and immunological parameters as
593 ecotoxicological biomarkers in a novel sentinel species (*Mimachlamys varia*). *Aquat.*
594 *Toxicol.* 161, 170–175.

595 Nowak, C., Vogt, C., Pfenninger, M., Scwenk, K., Oehlmann, J., Streit, B., Oetken, M., 2009.
596 Rapid genetic erosion of pollutants-exposed experimental chironomid populations.
597 *Environ. Poll.* 157, 881-886.

598 Nowak, C., Vogt, C., Oehlmann, J., Pfenninger, M., Schwenk, K., Streit, B., et al., 2012.
599 Impact of genetic diversity and inbreeding on the life-history of *Chironomus* midges over
600 consecutive generations. *Chemosphere* 88, 988–993.

601 Oziolor, E.M., Bickham, J.W., Matson, C.W., 2017. Evolutionary toxicology in an omics
602 world. *Evol Appl.* 10:752–761.

603 Pan, L., Ren, J., Liu, J., 2005. Effects of benzo(k)fluoranthene exposure on the biomarkers of
604 scallop *Chlamys farreri*. *Comp. Biochem. Physiol. Part–C*, 141, 248–256.

605 Pan, L.Q., Ren, J., Zheng, D., 2009. Effects of benzo(a)pyrene exposure on the antioxidant
606 enzyme activity of scallop *Chlamys farreri*. *Chinese J Oceanol Limn.* 43–53.

607 Paoletti, F., Aldinucci, D., Mocali, A., Caparrini, A., 1986. A sensitive spectrophotometric
608 method for the determination of superoxide dismutase activity in tissue extracts. *Anal.*
609 *Biochem.* 154 (2), 536–541.

610 Pedrosa, J.A.M., Machado, A.N., Cocchiararo, B., Soares, A.M.V.M., Nowak, C., Pestana,
611 J.L.T., 2017. Assessing the suitability of genetic diversity of *Chironomus riparius*
612 (Meigen) as an indicator of environmental pollution. *Ecol. Indic.* 77, 115-124.

613 Prohaska, J.R., 1980. The glutathione peroxidase activity of glutathione S-transferases.
614 *Biochimica and Biophysica. Acta, Enzymology* 611 (1), 87–98.

615 R Core Team. R: A language and environment for statistical computing. R Foundation for
616 Statistical Computing, Vienna, Austria. 2014. Available from: URL [http://www.R-](http://www.R-project.org/)
617 [project.org/](http://www.R-project.org/)

618 Rainbow, P.S., 2002. Trace metal concentrations in aquatic invertebrates: why and so what?
619 Environ. Pollut. 120, 497–507.

620 Regoli, F., Nigro, M., Bompadre, S., Winston, G.W., 2000. Total oxidant scavenging capacity
621 (TOSC) of microsomal and cytosolic fractions from Antarctic, Arctic, and Mediterranean
622 scallops: differentiation between three potent oxidants. Aquat. Toxicol. 49, 13-25.

623 Shrivastava, A. and Gupta, V. B., 2011. Methods for the determination of limit of detection
624 and limit of quantification of the analytical methods. Chron. of Young Sci. 2 (1), 21–25.

625 Smith, P.K., Krohn, R.I., Hermanson, G.T., Mallia, A.K., Gartner, F.H., Provenzano, M.D.,
626 Fujimoto, E.K., Goeke, N.M., Olson, B.J., Klenk, D.C., 1985. Measurement of protein
627 using bicinchoninic acid. Anal. Biochem. 150 (1), 76–85.

628 Sureda, A., Box, A., Tejada, S., Blanco, A., Caixach, J., Deudero, S., 2011. Biochemical
629 responses of *Mytilus galloprovincialis* as biomarkers of acute environmental pollution
630 caused by the Don Pedro oil spill (Eivissa Island, Spain). Aqua. toxicol. 101(3-4), 540–
631 549.

632 Tappel, M.E., Chaudiere, J., Tappel, A.L.L., 1982. Glutathione peroxidase activities of animal
633 tissues. Comp. Biochem. and Physiology. 73 (4), 945–949.

634 Tenenhaus, M., Esposito Vinzi, V., Chatelin, Y. M., Lauro, C., 2005. PLS path modeling.
635 Comput. Stat. and Data Analysis 48, 159–205.

636 Theodorakis, C.W., 2003. Establishing Causality between Population Genetic Alterations and
637 Environmental Contamination in Aquatic Organisms. [Human and Ecological Risk](#)
638 [Assessment: An International Journal](#) 9, 37-58.

639 Tigano, A., Friesen, V.L., 2016. Genomics of local adaptation with gene flow. *Mol Ecol.* 25,
640 2144–2164. doi:10.1111/mec.13606.

641 Unrine, J.M., Hopkins, W.A., Romanek, C.S., Jackson, Brian P., 2007. Bioaccumulation of
642 trace elements in omnivorous amphibian larvae: implications for amphibian health and
643 contaminant transport. *Environ. Pollut.* 149 (2), 182–192.

644 Valavanidis, A., Vlahogianni, T., Dassenaki, M., Scoullou, M., 2006. Molecular biomarkers
645 of oxidative stress in aquatic organisms in relation to toxic environmental pollutants.
646 *Ecotoxicol. Environ. Saf.* 64, 178-189.

647 van Straalen, N.M., Timmermans, M.J.T.N., 2002. Genetic Variation in Toxicant-Stressed
648 Populations: An evaluation of the “Genetic Erosion” Hypothesis. *Hum. Ecol. Risk.*
649 *Assess. Int. J.* 1; 8(5):983–1002.

650 Vigneau, E., Chen, M., Qannari, E.M., 2015. ClustVarLV: an R package for the clustering of
651 variables around latent variables. *The R Journal* 7, 134-148.

652

Table 1: Importance of the three first common components of Path-ComDim and the contribution of each pair of kinked blocks to the common components.

| Importance of the common components | | | Contribution of each link (%) | | | | | | |
|-------------------------------------|---------------------|-----------------------------------|-------------------------------|---------------------------|------------------------|----------------------------------|------------------|---------------|-------------------------|
| dimension | Absolute importance | Relative importance for each dim. | → | Heavy metals to BCH gills | Heavy metals to BCH dg | Heavy metals to genetic approach | ORG to BCH gills | ORG to BCH dg | ORG to genetic approach |
| 1 | 0.0080 | 47.9% | → | <0.1% | <0.1% | 0.5% | <0.1% | 0.1% | 99.3% |
| 2 | 0.0057 | 34.5% | → | 0.3% | 83.8% | 5.2% | <0.1% | 8.6% | 2.10% |
| 3 | 0.0012 | 7.0% | → | 96.8% | 2.6% | <0.1% | 0.5% | <0.1% | <0.1% |

Caption for table 1:

BCHdg: biochemical assays in the digestive glands

BCHgills: biochemical assays in the gills

ORG corresponds to the organic contaminants

Values in the first part of table (importance of the common components) were evaluated as the sum of squares of the saliences of the blocks to the common component extracted at each step (i.e. dimension)

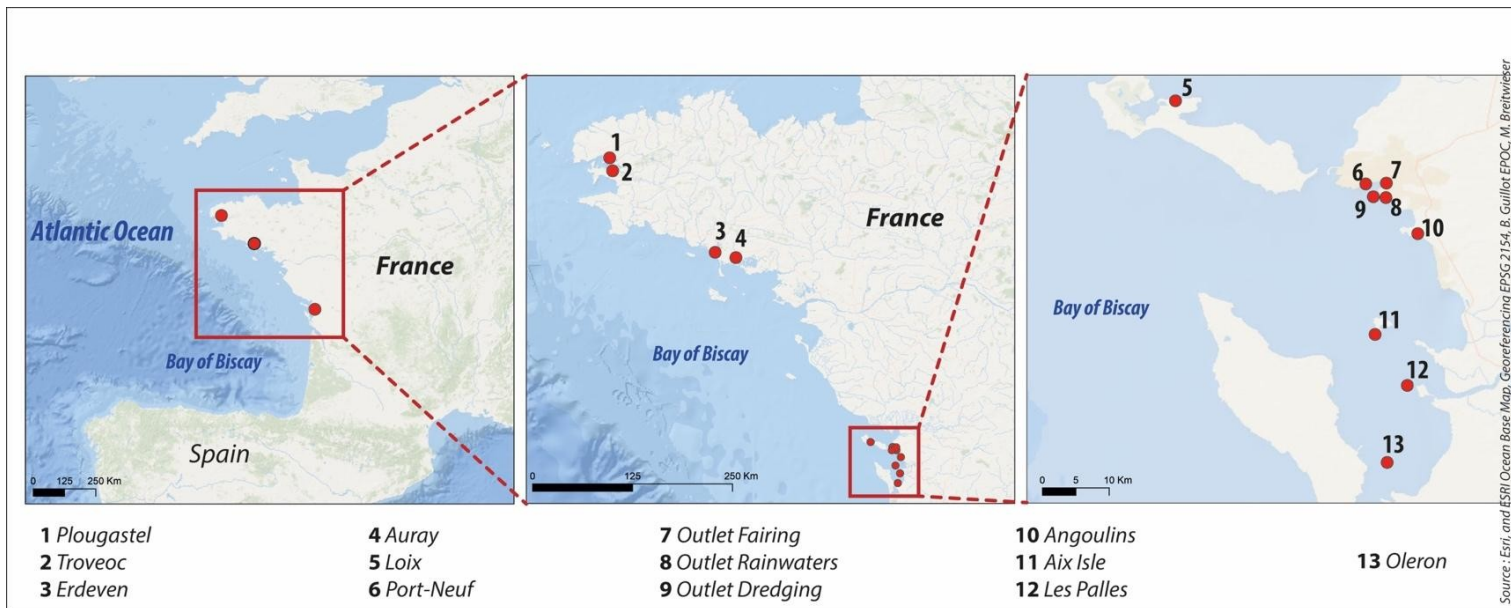


Figure 1: Map of France with the location of the study sites in the Brittany and Nouvelle-Aquitaine regions.

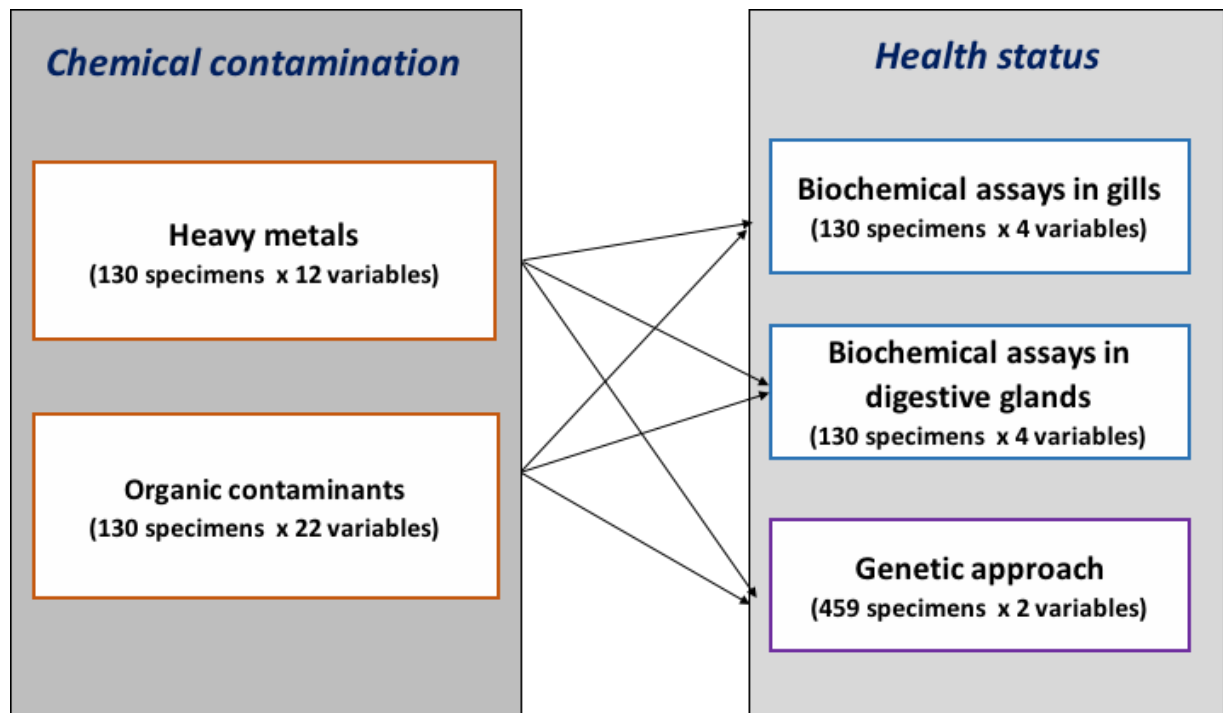


Figure 2: Path diagram for the five data blocks report summarizing chemical contamination with two blocks (heavy metals and organic contamination) and health status with three blocks (biochemical assay in two organs and the genetic approach). For each block, we analysed the same 130 specimens for bioaccumulation and biochemical aspects and 459 individuals (25 to 59 per site) for genetic study of the variegated scallop *Mimachlamys varia*.

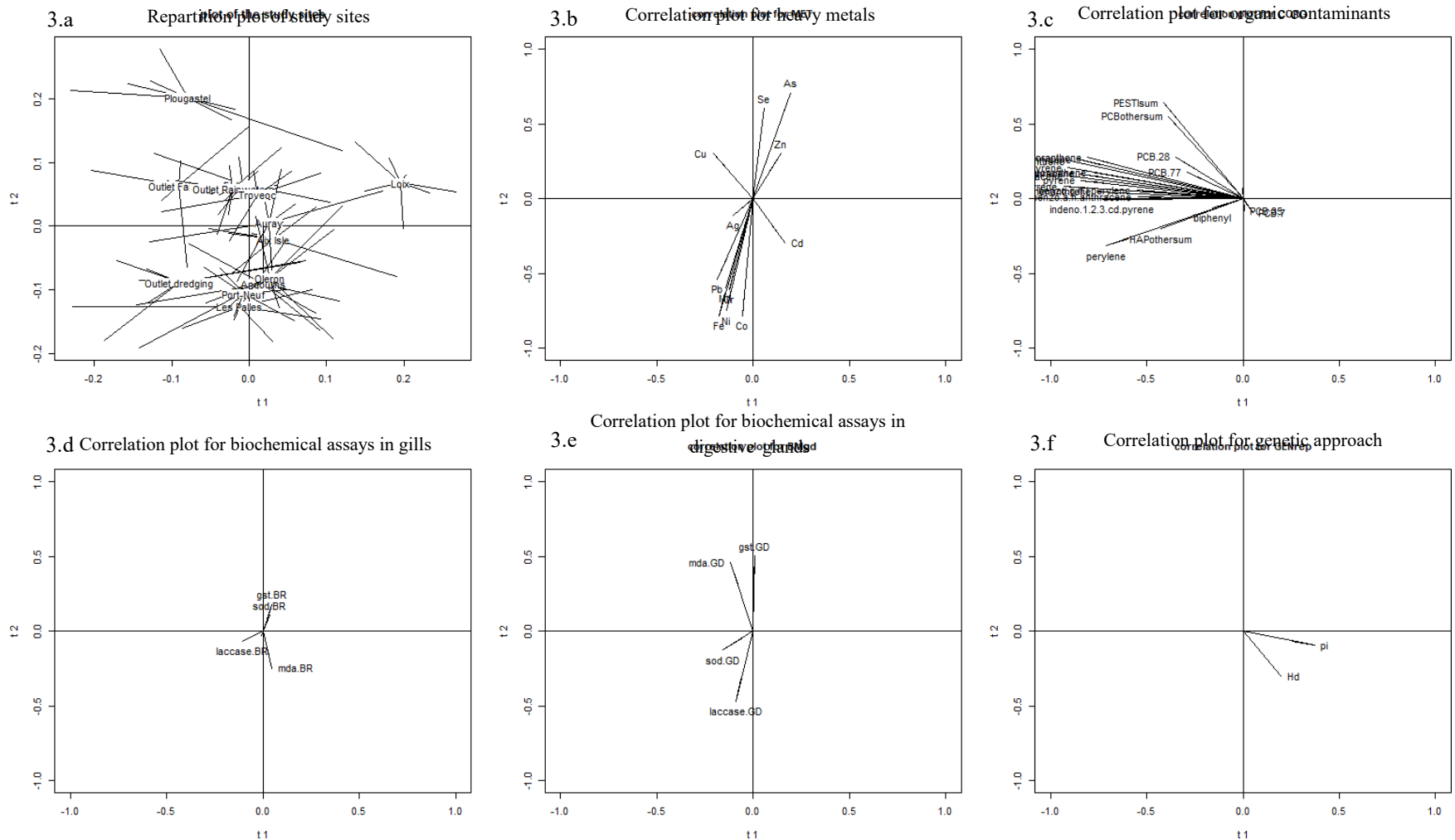


Figure 3: Multiblock dataset (Path-ComDim) analysis of biomarker responses (MDA: MDA contents, SOD: SOD activity, GST: GST activity, LAC: Laccase activity) and genetics, heavy metals, and organic contaminants accounting for the variation in the individual biomarker matrix for individuals of *M. varia* collected in March 2016 at 13 sampling sites.

