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Coudray

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Effects of high fat diet intake and a mitochondrial antioxidant (MitoQ)

on liver mitochondrial activity and cardiolipin content in rats



<u>C Feillet-Coudray¹</u>, G Fouret¹, E Tolika², J Lecomte³, M Aoun⁴, MP Murphy⁵, C Ferreri², E Dubreucq⁶, C Coudray¹

¹INRA UMR 866 DMEM, Montpellier, Fr, ² ISOF Bio Free Radicals, Bologna, Italy, ³CIRAD, UMR IATE, Montpellier, Fr, ⁴INSERM U1046, CNRS UMR9214, Université de Montpellier, Fr, ⁵MRC Mitochondrial Biology Unit/MRC Building, Cambridge UK, ⁶Montpellier SupAgro, UMR IATE, Montpellier, Fr.







A-Introduction

cfeillet@supagro.inra.fr

Excessive intake of dietary fat is one of the major cause of appearance and progression of metabolic syndrome features, such as liver steatosis. These features are associated with structural and functional alterations of mitochondria. Recent studies indicate that some of these alterations are related to the content and quality of cardiolipin (CL), a phospholipid that occurs mainly in the inner membrane of mitochondria and is involved in the organization and functioning of the mitochondrial respiratory chain complexes [1]

B-Protocol and methods



1-control	(40g s	soybean	oil/kg	diet)	

2-HF (4% Soybean oil + 31% lard)

24 rats **Sprague Dawley**

3-HF + Mito-Q (375 µmol/kgdiet)

8 weeks

1-Characterisation of rats

- Oral glucose tolerance test (OGTT) was completed 4 days before rat sacrifice.
- Plasma insulin was quantified with ELISA kits (Merck Millipore, Allemagne).
- Liver TG levels were quantified by enzymatic methods (TG PAP kit, Biomerieux, Fr)

2-Liver mitochondrial lipid content

•Mitochondrial PL, including CL, were separated by TLC (Merck plates) revelated with Mb blue spray and quantified using Scanner 3 CAMAG [3]

•Fatty acid content of mitochondrial PL was determined by GC after separation of the PL fraction by TLC

•Reverse transcription q-PCR analyses were to evaluate gene expression of performed CL synthesis and involved enzymes in remodeling

3-Liver mitochondrial characteristics

•Membrane fluidity was assessed by ESR using 16-NS [2]

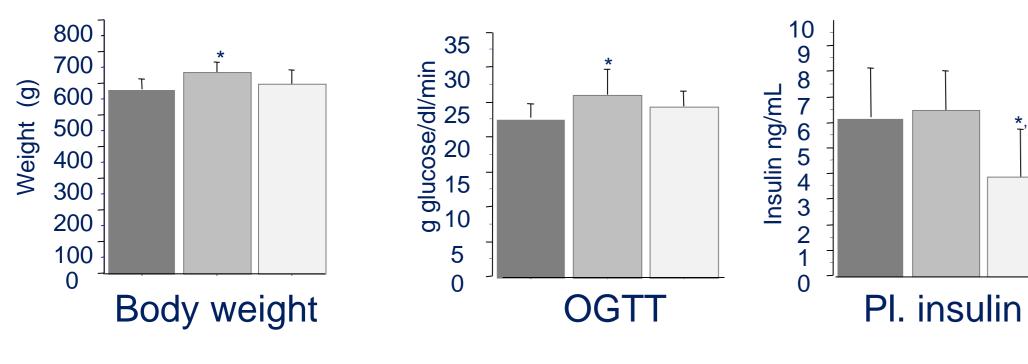
•Membrane potential and ROS production were monitored using respectively Rhodamine (RH-123) and 2',7'-Dichlorofluorescein diacetate

•Respiration was monitored in thermo-stated chambers of high resolution Oxygraph (Oroboros Oxygraph2k, Autriche)

•ATPase determined activity was spectrophotometrically

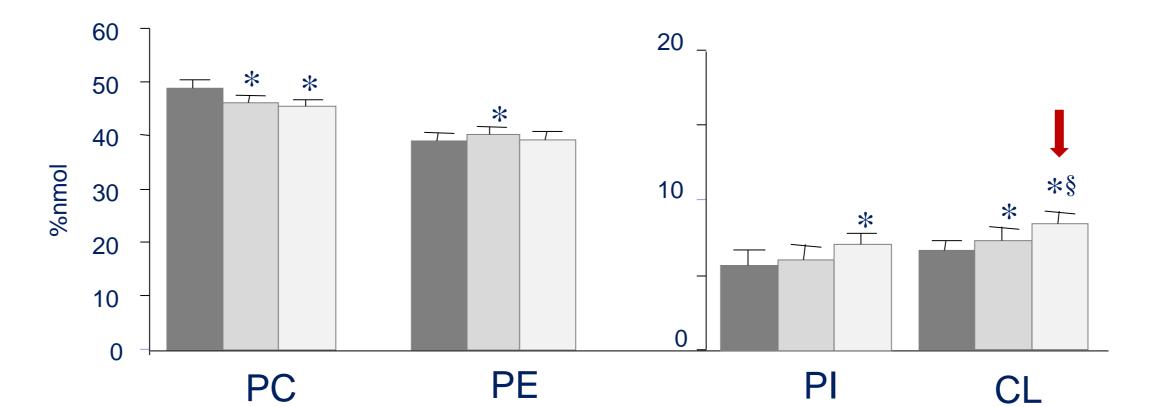
C-Results

1-Characteristics of the rats

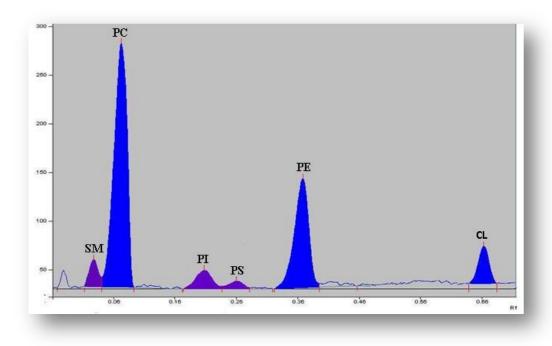


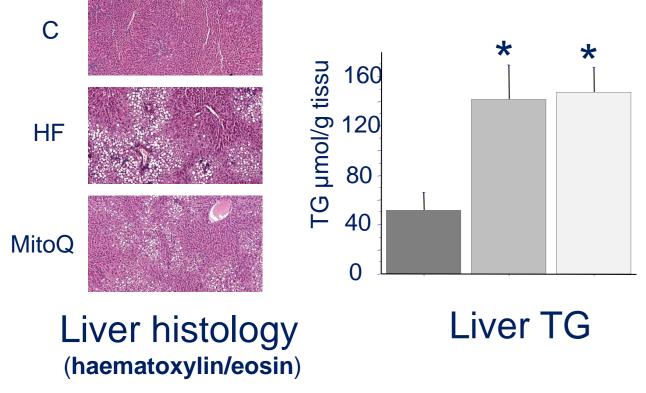
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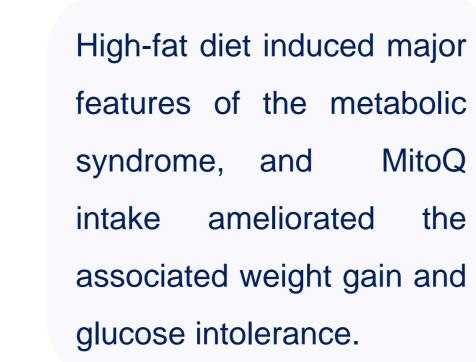
2-Liver mitochondrial cardiolipin content

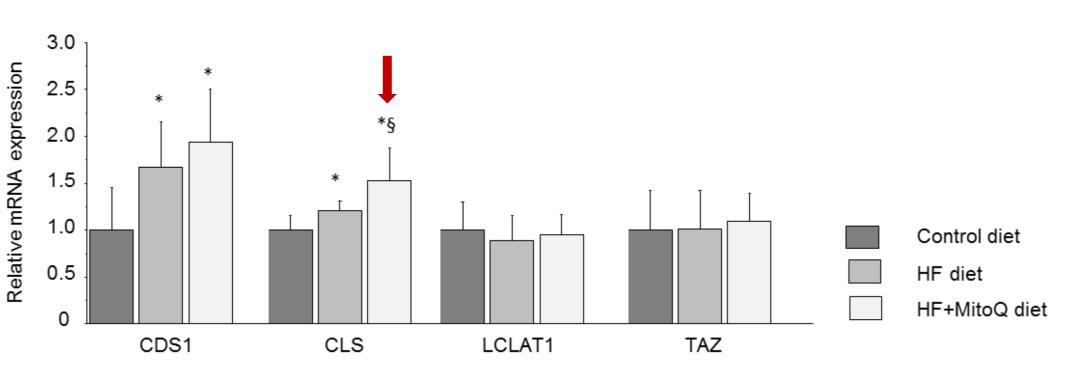


Scan of TLC plate





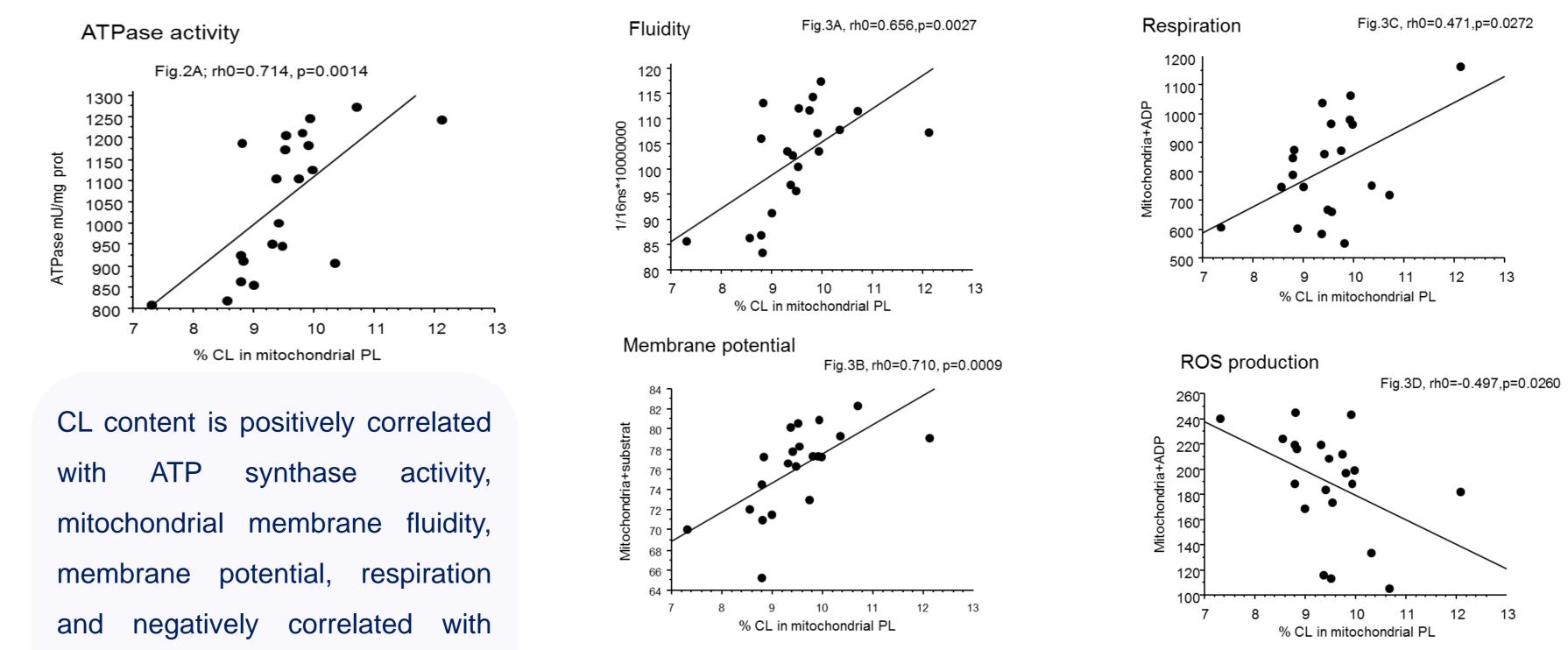




Results are mean + SD; n=8. Unpaired t-test. Significance was set at p<0.05.* vs control; §vs HF

MitoQ both increased CL content of liver mitochondria in the mitochondrial phospholipids and CL synthase gene expression by comparison to the HFD.

4-Liver mitochondrial characteristics



3-Fatty acid composition of mitochondrial PL

MitoQ increased 18:2n-6 (linoleic acid) percent in the mitochondrial phospholipids.

	Control diet	HF diet	HF-MitoQ diet
Total SFA	41.0 <u>+</u> 1.64	42.7 <u>+</u> 0.9	41.7 <u>+</u> 1.8
16:0	18.3 <u>+</u> 0.6	17.0 <u>+</u> 0.9*	16.2 <u>+</u> 0.6*
18:0	22.7 <u>+</u> 2.1	25.7 <u>+</u> 1.0*	25.5 <u>+</u> 1.8*
Total MUFA	8.39 <u>+</u> 0.82	6.37 <u>+</u> 0.76*	5.96 <u>+</u> 0.44*
16:1 n-7	1.28 ± 0.17	0.33 <u>+</u> 0.05*	$0.22 \pm 0.04 * $
18:1 n-9	2.99 <u>+</u> 0.28	$4.00 \pm 0.60^{*}$	3.87 <u>+</u> 0.30*
18:1 n-7	3.98 <u>+</u> 0.79	1.92 <u>+</u> 0.18*	1.75 <u>+</u> 0.16*
Total PUFA	50.6 <u>+</u> 1.05	50.9 <u>+</u> 1.16	52.3 <u>+</u> 1.68
18:2 n-6	9.94 + 1.53	11.5+1.16	12.8 + 0.81*
18:3 n-6	0.106 ± 0.044	0.047 <u>+</u> 0.023*	$0.054 \pm 0.023*$
18:3 n-3	1.021 ± 1.541	0.426 ± 0.198	0.314 ± 0.102
20:3 n-6	0.881 <u>+</u> 0.304	0.892 <u>+</u> 0.119	0.865 <u>+</u> 0.184
20:4 n-6	27.4 <u>+</u> 1.3	28.0 <u>+</u> 0.5	28.6 <u>+</u> 1.11
20:5 n-3	0.114 <u>+</u> 0.037	0.099 <u>+</u> 0.058	0.082 <u>+</u> 0.015
22:5 n-3	0.761 <u>+</u> 0.044	0.954 <u>+</u> 0.047*	0.975 <u>+</u> 0.199*
22:6 n-3	9.75+1.21	8.56+0.57*	8.36+0.41*

mitochondrial ROS production.

Correlations were performed with the Spearman method.

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D-Conclusion

Our studies show that the high fat diets impair liver mitochondrial functions and modify membrane structure and characteristics. MitoQ ameliorates some

metabolic syndrome features, and in particular increases mitochondrial cardiolipin content. Antioxidant-independent mechanisms of MitoQ may be involved

in addition to its own antioxidant action.

Acknowledgments

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[1] Schlame M et al. (2009) BBA. 1788(10):2080-3., [2] Aoun M et al (2012). J Bioenerg Biomembr.44(4):439-52., [3] Fouret G et al (2015). Biochim Biophys Acta. 2015 May 28.