



HAL
open science

Size-characterization of natural and synthetic polyisoprenes by Taylor dispersion analysis

Jean-Philippe Biron, Frédéric Bonfils, Luca Cipelletti, Hervé Cottet

► **To cite this version:**

Jean-Philippe Biron, Frédéric Bonfils, Luca Cipelletti, Hervé Cottet. Size-characterization of natural and synthetic polyisoprenes by Taylor dispersion analysis. *Polymer Testing*, 2018, 66, pp.244-250. 10.1016/j.polymertesting.2018.01.017 . hal-01837505

HAL Id: hal-01837505

<https://hal.science/hal-01837505>

Submitted on 26 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

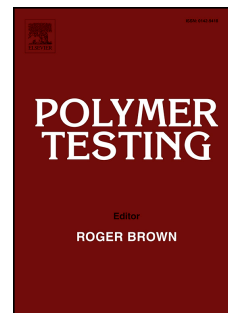


Distributed under a Creative Commons Attribution 4.0 International License

Accepted Manuscript

Size-characterization of natural and synthetic polyisoprenes by Taylor dispersion analysis

Jean-Philippe Biron, Frédéric Bonfils, Luca Cipelletti, Hervé Cottet



PII: S0142-9418(17)31611-2

DOI: [10.1016/j.polymertesting.2018.01.017](https://doi.org/10.1016/j.polymertesting.2018.01.017)

Reference: POTE 5303

To appear in: *Polymer Testing*

Received Date: 12 November 2017

Accepted Date: 22 January 2018

Please cite this article as: J.-P. Biron, Fr  . Bonfils, L. Cipelletti, Herv  . Cottet, Size-characterization of natural and synthetic polyisoprenes by Taylor dispersion analysis, *Polymer Testing* (2018), doi: 10.1016/j.polymertesting.2018.01.017.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Comment citer ce document :

Biron, J.-P., Bonfils, F., Cipelletti, L., Cottet, H. (2018). Size-characterization of natural and synthetic polyisoprenes by Taylor dispersion analysis. *Polymer Testing* , 66, 244-250. , DOI : 10.1016/j.polymertesting.2018.01.017

Size-characterization of natural and synthetic polyisoprenes by Taylor dispersion analysis

Jean-Philippe Biron¹, Frédéric Bonfils², Luca Cipelletti³ and Hervé Cottet^{1*}

¹ IBMM, University of Montpellier, CNRS, ENSCM, Montpellier, France

² IATE, University of Montpellier, CIRAD, INRA, Montpellier Sup Agro, Montpellier, France

³ L2C, University of Montpellier, CNRS, Montpellier, France

* CORRESPONDING AUTHOR: Prof. Hervé Cottet

Tel: +33 4 6714 3427, Fax: +33 4 6763 1046. E-mail: herve.cottet@umontpellier.fr

ABSTRACT: Non-aqueous Taylor dispersion analysis (TDA) was used for the size-characterization of natural and synthetic polyisoprenes (4×10^3 - 2×10^6 g/mol molar mass). Not only the weight-average hydrodynamic radius (R_h), but also the probability distribution of the hydrodynamic radius, were both derived from the Taylorgrams by a simple integration of the elution profile and by a more sophisticated constrained regularized linear inversion of the Taylorgram, respectively. Results in terms of size characterization (hydrodynamic radii between 2 and 100 nm) were compared to size exclusion chromatography coupled to a refractive index-based mass detector. Multimodal size distributions were resolved by TDA for industrial and natural polyisoprenes, with the advantage over the chromatographic technique that, in TDA, there is no abnormal elution of microaggregates (hydrodynamic radii ~ 40-50 nm). Considering the importance and the difficulty of characterizing polyisoprene microaggregates, TDA appears as a promising and simple technique for the characterization of synthetic and natural rubber.

Key Words: polyisoprene; natural rubber; Taylor dispersion analysis; hydrodynamic radius; diffusion coefficient; size distribution.

Title Running Head. Taylor dispersion analysis of synthetic and natural rubbers

Natural rubber (NR) is a biopolymer produced from latex of *Hevea brasiliensis* trees. NR is used to prepare many goods, the main one being tires that absorb nearly 70 % of the 12 million tons annually worldwide. As an agro-material produced by a tree, NR exhibits rather variable properties for many reasons (genotype of the tree [1-2], season [3], maturation of coagula [4-6], etc.). Thus, as for other polymeric materials used in industrial applications, it is important to characterize the NR macromolecular structure as a prelude to predicting NR end-use properties [7-8].

Natural rubber has a very specific associative structure. When dispersed in good solvents for polyisoprene, NR presents a soluble fraction and an insoluble fraction, often referred to as "macrogel" [9-10] or "gel phase" [11-12]. The composition of the soluble part is also specific as it contains two main entities: (i) polyisoprene linear macromolecules of different lengths, with a random coil structure, and (ii) very compact microaggregates or microgels (sphere-like structure) [13-15]. Links between non-isoprene compounds (lipids, proteins, and/or mineral elements) and the polyisoprene chains are thought to be the main reason for the macrogel formation in NR [12, 16-17]. Although an international standard (ISO 16564) exists to characterize the NR macromolecular structure by size exclusion chromatography (SEC) or gel permeation chromatography (GPC), the development of new analytical techniques is still required to improve the characterization of NR using orthogonal or complementary methods.

This paper describes a novel method to characterize the NR macromolecular structure: Taylor dispersion analysis in organic solvent. Taylor Dispersion Analysis (TDA) is an absolute sizing method requiring no calibration and allowing the determination of the diffusion coefficient (or, equivalently, the hydrodynamic radius) of solutes of any size between one angstrom and a few hundred nanometers. TDA is based on the dispersion of a sample band in an open tubular column under Poiseuille flow. The combination of the dispersive parabolic velocity profile with the molecular diffusion of the solute leads to a specific dispersion, called Taylor dispersion. Started with the pioneering work of Taylor on macroscopic tubing [18], TDA is now implemented in narrow capillaries (typically 50 μm internal diameter), leading to much faster analysis (a few min) and requiring only tiny sample amounts (a few nL) [19-22]. In its modern miniaturized version, TDA can be very easily implemented on commercial capillary electrophoresis apparatus. Alternatively, fast TDA can be performed on HPLC equipment at a high linear velocity ($\sim 30 \text{ cm s}^{-1}$) in a coiled open capillary tube of 250 μm i.d. \times 30 m typical dimensions, as recently introduced by the Holyst group [23]. In the latter

configuration, the genuine conditions for the applicability of TDA are not fulfilled [24], however, an empirical correction can be used to get the correct diffusion coefficient [23].

TDA was applied to the characterization of proteins [25-27], macromolecules [28-30], drug delivery systems [31, 32], nanoparticles [21, 33, 34], polyelectrolyte complexes and polyplexes [35], pharmaceutical compounds [36, 37], and micelles or microemulsions [38, 39]. It can also be used for the study of molecular interactions [40, 41]. The application of TDA is not limited to water-soluble solutes, as demonstrated by the size characterization of polystyrenes in THF [28], asphaltenes/bitumen in hexane or THF/acetonitrile mixtures [42, 43], and drugs in pharmaceutical solvents [36]. Basically, TDA yields the weight average hydrodynamic radius when using a mass sensitive detector, such as a UV detector, for a polymer absorbing via the repeating unit [44]. This information is obtained by simple integration of the elution temporal profile (also called Taylorgram). However, we recently demonstrated that more sophisticated data analyses allow one to determine the full hydrodynamic radius distribution, thus providing valuable information on the sample polydispersity [45, 46].

In this work, we demonstrate that TDA can be used to get, within a few minutes, the hydrodynamic radius distributions of commercial natural and synthetic polyisoprenes. The data obtained by TDA are compared to those derived from size exclusion chromatography (SEC) coupled to multi-angle static light scattering (MALS).

2 Experimental

2.1 Materials

The two NR samples used for this study were all TSR (Technically Specified Rubber). Sample NR1 was industrial TSR10 grade prepared by natural coagulation of latex followed by several days of coagulum maturation before processing. This sample was treated with neutral hydroxylamine sulfate (NHS) to stabilize its properties during storage, yielding a TSR10CV grade (CV stands for constant viscosity), a special grade not prone to storage hardening. Sample NR2 was prepared by acid coagulation (formic acid) of fresh field latex. Neutral hydroxylamine sulfate (NHS) was added to the latex prior to coagulation to inhibit storage hardening of rubber [47]. Sample NR2 was TSR5CV grade. Three synthetic industrial cis-1,4-polyisoprenes (PI) were used directly as received: IR2200 (NatSyn 2200, Goodyear chemical), IR307 and IR309 (Kraton polymers). Standard linear polyisoprene (PI) of 4.5, 22, 111, 307, 361, 608 and 766×10^3 g/mol weight-average molar masses were obtained from Polymer Standard Service (PSS, Germany) and were used as received.

2.2 Determination of the number- and weight-average molar mass by size exclusion chromatography coupled to multi-angle light scattering (SEC-MALS)

For all samples, the macromolecular structure was characterized by SEC-MALS, except for standard polyisoprene, for which the number average, M_n , and mass-average, M_w , molar mass were given by the supplier. The samples (30 ± 5 mg), from NR pellets, were dissolved in tetrahydrofuran (THF, 30 mL, HPLC grade, VWR France) stabilized with 2,6-di-tert-butyl-4-methylphenol (BHT). The solutions were stored for 7 days in the dark at 30°C (during storage, the samples were stirred for 1 h/day), they were then filtered (Acrodisc 1 μ m, glass fiber, Pall France) and finally injected into the SEC-MALS apparatus. For each sample, three solutions were independently prepared and measured. The SEC equipment comprises an online degasser (EliteTM, Alltech), a Waters 515 pump, a refractive index detector (Waters 2410) and a multi-angle light scattering detector (Dawn DSP, Wyatt Technology). Two Waters HMW6E columns (Styragel HMW columns packed 20 μ m particles, 300 mm x 7.8 mm I.D.), maintained at 45°C, and a guard column (same stationary phase as the two separation columns) were used. The mobile phase was stabilized THF injected at a flow rate of 0.65 mL.min⁻¹; the injected volume was 150 μ L. The MALS detectors at all 18 angles were calibrated using a THF solution of a polystyrene standard with low polydispersity ($M_w = 30.3$ kg.mol⁻¹, Wyatt technology). The same solution was used to determine the interconnection volume between the two detectors (0.235 mL). The basic theory of determining the weight-average molar mass and radius of gyration for a dilute solution of a macromolecule is well known and described in numerous papers in the literature [48, 49]. The weight average molar masses and radius of gyration at each slice of the chromatogram were calculated using the Berry method for extrapolation, as implemented in ASTRA software (version 5.3.1, Wyatt technology). The order of polynomial fit for the Berry analysis was two. Twelve angles, from 38.8° to 138.8°, were used for the analysis. The differential refractive index increment at $\lambda = 633$ nm was $dn/dc = 0.130$ mL.g⁻¹ [50].

2.3 Determination of the weight-average hydrodynamic radius R_h and of the R_h distribution by Taylor dispersion analysis

TDA experiments were performed on a PACE MDQ Beckman Coulter (Fullerton, CA) apparatus. Capillaries were prepared from bare silica tubing purchased from Composite Metal Services (Worcester, United Kingdom). The capillary dimensions were 40 cm (30 cm to the detector) \times 50 μ m I.D. New capillaries were conditioned with the following flushes: 1 M

NaOH for 30 min and water for 10 min. Before sample injection, the capillary was filled with cyclohexane (8.62×10^{-4} Pa.s viscosity). Between two TDA analyses, the capillary was flushed with cyclohexane (30 psi for 2 min). A mobilization pressure of 1 psi (~ 71 mbar) was applied with cyclohexane vials at both ends of the capillary, resulting in a flow rate of $184 \text{ nL}\cdot\text{min}^{-1}$. Samples were dissolved in cyclohexane at a concentration of 1 g/L. Sample injection (2.8 nL) was performed hydrodynamically on the inlet side of the capillary (0.3 psi for 3 s; $\sim 1\%$ of the capillary volume). Solutes were monitored by UV absorbance at a wavelength of 200 nm. The elution time was systematically corrected for the delay in the application of the pressure (pressure ramp time, 15 s) by subtracting 7.5 s (half-time of the pressure ramp) to the recorded elution time. The temperature of the capillary cartridge was set at 25 °C.

Two different methods were used to determine the weight-average hydrodynamic radius by TDA. The first method (*method 1*) is based on the integration of the Taylorgram [22], and the second method (*method 2*) is based on a deconvolution of the Taylorgram using a recently published Constrained Regularized Linear Inversion (CRLI) method [45].

In method 1, the average diffusion coefficient D is obtained by integration of the Taylorgram (or temporal elution profile) in order to calculate the temporal variance of the Taylorgram, σ^2 . The integration of the elution profile was performed using the discrete form of the following equation [22]:

$$\sigma^2 = \frac{\int S(t)(t-t_0)^2 dt}{\int S(t)dt} = \frac{\sum_{i=n}^m S_i (t_i - t_0)^2 (t_{i+1} - t_i)}{\sum_{i=n}^m S_i (t_{i+1} - t_i)} \quad (1)$$

where $S(t)$ is the detector response, t_i is the elution time for a given point i of the Taylorgram and t_0 is the average elution time. n and m are the starting and ending points that are considered for the integration of the Taylorgram.

The integration of the Taylorgram was only performed on the left part of the elution profile (the end point was t_0), before and after subtracting the Gaussian contribution due to the small molecules (see results and discussion section). Corrections of the temporal variance and average detection time due to the injected volume were also taken into account, as described elsewhere [29], but these corrections are negligible when the injected volume is $\leq 1\%$ of the capillary volume to the detection point [22], as is the case here. Finally, the so called Taylor average diffusion coefficient $\langle D \rangle_{TDA}$ is obtained from:

$$\langle D \rangle_{TDA} = \frac{R_c^2 t_0}{24\sigma^2} \quad (2)$$

where R_c is the capillary radius.

Equation (2) is valid as far as $\frac{\langle D \rangle_{TDA} t_0}{R_c^2} \geq 1.25$ [18, 24] to ensure that the analysis time is

larger than the characteristic diffusion time of the solutes over the capillary cross section.

Equation (2) is also only valid if the axial diffusion can be neglected compared to the Taylor

dispersion contribution. This latter condition is fulfilled if $\frac{R_c u}{\langle D \rangle_{TDA}} \geq 40$, where u is the linear

velocity of the mobile phase [18, 24]. It is worth noting that the Taylor average diffusion coefficient obtained by integration of the Taylorgram (method 1) is a weight harmonic average diffusion coefficient defined by:

$$\langle D \rangle_{TDA} = \left[\overline{D^{-1}} \right]^{-1} = \left[\int_0^\infty P_D(D) D^{-1} dD \right]^{-1} \quad (3)$$

where $P_D(D)$ is the (mass-weighted) probability distribution function (PDF) of the diffusion coefficient. The TDA weight-average hydrodynamic radius $\langle R_h \rangle_{TDA}$ is finally determined using the Stokes-Einstein relationship:

$$\langle R_h \rangle_{TDA} = \frac{k_B T}{6\pi\eta \langle D \rangle_{TDA}} \quad (4)$$

where k_B is the Boltzmann constant, T the temperature (K) and η the viscosity of the carrier liquid (Pa.s).

Method 2 is based on a recently developed Constrained Regularized Linear Inversion (CRLI) approach [45]. This method allows the PDF of the hydrodynamic radius to be extracted from the experimental Taylorgrams. In brief, this method consists of introducing additional constraints in the usual least-squares fitting of the Taylorgram ("regularization"), in order to reject non-physical solutions, finally converging to the desired PDF of the hydrodynamic radius, $P_R(R_h)$. Typically, the constraints penalize those solutions for which $\partial^2 P_R / \partial R^2$ is, on average, too large, i.e. those PDFs that exhibit too detailed a structure, see Ref. [45] for more details.

The CRLI algorithm inverts the following relationship relating the Taylorgram to $P_R(R_h)$ [45]:

$$S(t) = \frac{k_R m_{tot}}{Q} \sqrt{\frac{\lambda_R}{\pi}} \int_0^\infty \frac{P_R(R_h)}{\sqrt{R_h}} \exp\left[-\frac{\lambda_R (t-t_0)^2}{R_h}\right] dR_h \quad (5)$$

where k_R is the detector response factor, m_{tot} the total sample mass injected, Q the carrier liquid flow-rate and with

$$\lambda_R = \frac{2k_B T}{\pi R_c^2 t_0 \eta} \quad (6)$$

After normalization, eq. (5) reduces to:

$$s(t) = \frac{S(t)}{S(t_0)} = c_R \int_0^{\infty} \frac{P_R(R_h)}{\sqrt{R_h}} \exp\left[-\frac{\lambda_R(t-t_0)^2}{R_h}\right] dR_h \quad (7)$$

with

$$c_R = \left[\int_0^{\infty} \frac{P_R(R_h)}{\sqrt{R_h}} dR_h \right]^{-1} = \left[R_h^{-1/2} \right]^{-1} \quad (8)$$

The PDFs of R_h reported in the following are obtained by averaging individual distributions measured in 2 to 5 independent repetitions. For each sample, a mass-average R_h value was calculated from $P_R(R_h)$, via:

$$\langle R_h \rangle_{CRLI} = \frac{\int_0^{\infty} R_h \times P_R(R_h) dR_h}{\int_0^{\infty} P_R(R_h) dR_h} \quad (9)$$

3 Results and discussion

Our main objective is to compare the average radii and the R_h PDFs obtained by SEC and by TDA on different PI standards, industrial PI and natural rubbers (NR).

3.1 Determination of the average hydrodynamic radius by TDA

Examples of raw Taylorgrams obtained for the polyisoprene (PI) standards are displayed in Figure 1A. Each Taylorgram has been normalized by the maximum absorbance at $t=t_0$. A quite good baseline return and peak symmetry is seen in Figure 1A. It is clearly observed that the higher the molar mass, the broader the Taylorgram, due to the higher Taylor dispersion for the species with lower diffusion coefficient. Nevertheless, some PI standards contain a tiny amount of small molecules (possibly monomers, residual solvent) absorbing at the detection wavelength (200 nm). This small molecule contribution appears as a thin peak, most notable for the 361 and 766×10^3 g/mol PI standards (blue and green lines in Figure 1A). The contribution due to the small molecules should not be taken into account in the data analysis. Accordingly, we systematically corrected for it by subtracting the corresponding Gaussian peak from the raw data, leading to the Taylorgrams presented in

Figure 1B [51]. The corresponding weight-average hydrodynamic radii obtained by integration of the Taylorgrams presented in Figure 1B (*method 1*, $\langle R_h \rangle_{TDA}$) are gathered in Table 1. For the sake of comparison, the R_h values obtained without subtraction of the small molecule contribution are also given in Table 1. The impact of the small molecule contribution on the average R_h value is less than 3%.

<Figure 1>

The same data processing was applied to the industrial PI samples (IR2200, IR307 and IR309) and the natural rubber samples (NR1 and NR2). The raw Taylorgrams and the corrected ones are displayed in Figures 2A and 2B for industrial PI, and Figures 3A and 3B for NR, respectively. It should be noticed that the symmetry of the Taylorgrams was not perfect, especially in the case of NR. This may be due to some polymer adsorption onto the capillary surface. This is why the Taylorgrams were analyzed on the left part, where adsorption has a much lower impact. The contribution of the small molecules to the average R_h value is less than 6% (see Table 1). The correlation between the hydrodynamic radius (*method 1*, $\langle R_h \rangle_{TDA}$) and the PI weight average molar mass (M_w , SEC) is given in Figure 4. For a random coil in a good solvent, the theoretical value of the exponent b of the power law $R_h = aM_w^b$ is predicted to be 0.588 [52]. This value was confirmed experimentally using light scattering, for PI in cyclohexane (Fetters *et al.*, $b=0.592$ [53]) and in tetrahydrofuran (Kim *et al.*, $b = 0.606$ [14]). In our experiments, we find an exponent $b=0.54\pm 0.06$ (confidence interval at 95%) for the seven PI standards, slightly lower than the expected theoretical value. Industrial PI grades and NR samples are close to the scaling law obtained for the PI standards (see Figure 4).

<Table 1>

<Figure 2>

<Figure 3>

<Figure 4>

3.2 Hydrodynamic radius distributions obtained by TDA and comparison with SEC

The elution profile in SEC gives access to the size distribution of the polymers, since the separation parameter in SEC is the hydrodynamic volume of the solute. For the SEC

measurements, THF was used as the eluent. THF gives a higher signal for the LS and refractive index analysis, thanks to a higher dn/dc compared to cyclohexane. For the TDA analysis, by contrast, cyclohexane is the best choice, since THF is incompatible with UV detection and with the TDA setup (THF dissolves the MDQ Beckman Coulter plastic interface in contact with the sample and eluent vials). In spite of the need to use different solvents, the comparison between the TDA and SEC-MALS results for the same PI and rubber samples sheds light on the strengths and limitations of each method.

In SEC, the logarithm of the hydrodynamic radius is a decreasing linear function of the elution volume. In the elution profile, the refractive index (RI) detector yields a signal proportional to the polymer mass concentration. The MALS detector combined with the RI detector allows the polymer molar mass to be measured. All the 3 industrial PI analyzed by SEC-MALS exhibited a main peak eluting at about 16-17 min, followed by one or two other modes eluting between 18-22 min, which were more or less separated depending on the sample. IR309 appeared as a tri-modal elution profile (see Figure 5A), while the two other samples (IR307 and IR2200) presented unresolved shoulders. Figure 5B shows hydrodynamic radius (R_h) mass-distributions obtained by SEC assuming that $R_g/R_h = 1.78$, as expected for monodisperse random coils in good solvent [52]. Indeed, SEC coupled to a MALS detector allows determining, for each slice of the chromatogram, both the molar mass (M) and the radius of gyration (R_g). The R_{hi} were calculated from the R_{gi} for each slice of the chromatogram using $R_g/R_h = 1.78$ relationship. In the SEC analysis, all samples display rather broad distributions with an unresolved shoulder for IR2200 and IR307, and three partially resolved modes for IR309. The R_h corresponding to the peak maximum for the mass-distribution of R_h determined from the SEC data (Figure 5B) were 32 nm for IR2200, 45 nm for IR309 and 47 nm for IR307.

Figure 5C displays the hydrodynamic radius distributions obtained by TDA after data processing of the Taylorgrams from Figure 2A according to *method 2* (CRLI, see section 2.3). Size distributions obtained by TDA display two well-defined main modes, and a less important (minor) third mode (see inset of Figure 5C), corresponding to low molar mass polymers (oligomers) or monomers with R_h below 3 nm. These oligomers/monomers appear as a sharp peak in the Taylorgrams presented in Figure 2A. All IR samples display a main mode at R_h ranking between 30 and 50 nm (peak apex at 34.8 nm for IR 2200, 41.4 nm for IR 309 and 42.3 for IR 307). These R_h values are in reasonably good agreement with SEC analysis, given that the solvents are different and taking into account the uncertainty in the actual value of R_g/R_h required to calculate the R_h PDF from the SEC data. A second mode in the R_h distribution was observed by TDA, corresponding to sizes between 5 and 11 nm (peak apex at 10.6 nm for IR 2200, 9.6 for IR 309 and at 5.6 nm for IR 307). This peak is not

resolved in the PDF calculated from the SEC data, except perhaps for the IR 309 sample, which exhibits a pronounced shoulder in the 10-20 nm range. Overall, TDA showed was better resolution for the size characterization of the IR samples compared to SEC.

<Figure 5>

For natural rubber (NR) samples, the TDA analysis again gave R_h profiles with better size resolution than those retrieved from the SEC elution profiles (Figure 6). The SEC data exhibited elution profiles rather unimodal, with a more or less pronounced shoulder. Unlike the PI industrial samples, for the NR samples, it is not possible to assume that each slice of the chromatogram is only composed of monodisperse random coils, because of the co-elution of microaggregates and random coil macromolecules [13]. For this reason, it was not possible to calculate the R_h PDF. By contrast, in TDA, R_h is directly determined, with no need of making any assumptions on the nature of the sample (e.g. on the presence of aggregates). As displayed in Figure 6B, the NR samples had trimodal R_h profiles with small oligomers (R_h below 2-3 nm) and two larger polymer modes at 7–10 nm and 20–60 nm, respectively. Remarkably, the TDA analysis reveals unambiguous differences in the position and the width of the peaks for the two samples, not captured by SEC. In particular, the peak for the largest sizes shifts from 37 nm for NR1 to 40 nm for NR2. As explained above, it was not possible to determine the R_h PDF from the SEC data. However, we note that the ranking in the SEC elution curves is consistent with the TDA data, since NR2 is eluted before NR1, indicative of a larger size.

From the R_h distribution obtained by *method 2*, it is also possible to determine a weight-average R_h by integration on the whole distribution (see Eq. 9). We exclude the contribution of the small molecules, $R_h < 3$ nm, consistentl with the subtraction performed in *method 1*. The results are presented in Table 1: we find a difference of less than 3% between the two methods. This demonstrates the excellent agreement between the two different ways to process the Taylorgrams, thereby confirming the robustness of the methods of analysis presented here.

<Figure 6>

4 Conclusions

We have demonstrated that TDA can be used in non-aqueous media (cyclohexane in the present study) for the analysis of PI in industrial and natural rubbers. Because TDA is implemented in open capillary tubes, it is much less influenced by undesirable interactions as compared to SEC, where microaggregates from NR samples (hydrodynamic radii ~40-50 nm)

may result in abnormal elution curves at large elution times. By leveraging on the constrained regularized linear inversion (CRLI) method, not only does TDA give access to the weight average hydrodynamic radius, it also provides the full hydrodynamic radius distribution. While under some conditions SEC data can yield the PDF of R_h , the present work shows that TDA has a broader applicability and a superior ability to resolve the various modes contributing to the sample size distribution in a large range of hydrodynamic radii from ~ 1 to 100 nm. Given the strengths of the method and the advantages of modern TDA in its miniaturized version (low sample volume $\sim nL$, low solvent consumption, fast analysis (a few min), absolute determination, simple instrumentation and straightforward methodology), there is no doubt that TDA holds great potential for quality control and production monitoring of PI in the rubber industry. Possible future work could be to investigate new capillary coatings and/or capillary i.d. to improve the peak symmetry, especially in the case of NR analysis.

References

- [1] C. Kim, M. Morel, J. Sainte Beuve, S. Guilbert, F. Bonfils, Better Characterization of Raw Natural Rubber by Decreasing the Rotor Speed of Mooney Viscometer: Role of Macromolecular Structure, *Polym Eng Sci*, 50 (2010) 240-248.
- [2] C.M. Lotti, R.M.B. Moreno, P.S. Gonçalves, S. Bhattacharya, L.H.C. Mattoso, Extensional Rheology of Raw Natural Rubber From New Clones of *Hevea brasiliensis*, *Polym Eng Sci*, 52 (2012) 139-148.
- [3] M. Ferreira, R.M.B. Moreno, P.S. Gonçalves, L.H.C. Mattoso, Evaluation of natural rubber clones of *Hevea brasiliensis*, *Rubb Chem Technol*, 75 (2002) 171-178.
- [4] E.E. Ehabe, Y. Leroux, F. Ngolemasango, F. Bonfils, G. Nkeng, B. Nkouonkam, J. Sainte Beuve, Effect of the maturation on the bulk viscosity and molecular chain length of cuplump NR, *J Appl Polym Sci*, 86 (2002) 703-708.
- [5] Jie-Ping Zhong, Si-Dong Li, Ling-Xue Kong, Lei Yang, Shuang-Quan Liao, Xiao-Dong She, Study on the Properties of Natural Rubber during Maturation, *J Polym Mater*, 26 (2009) 351-360.
- [6] J. Intapun, J. Sainte Beuve, F. Bonfils, V. Tanrattanakul, E. Dubreucq, L. Vaysse, Effect of micro-organisms during initial coagula maturation of *Hevea* natural rubber, *J Appl Polym Sci*, 118 (2010) 1341-1348.
- [7] K.N.G. Fuller, Rheology of raw rubber, in: A. Roberts (Ed.) *Natural Rubber Science and Technology*, Oxford University Press, Oxford, 1988, pp. 141-176.
- [8] L. Vaysse, F. Bonfils, J. Sainte-Beuve, M. Cartault, Natural Rubber, in: K. Matyjaszewski, M. Möller (Eds.) *Polymer Science, A Comprehensive Reference*, Elsevier BV, Amsterdam, 2012, pp. 281-293.
- [9] P.W. Allen, G.M. Bristow, The gel phase in natural rubber, *Rubb Chem Technol*, 36 (1963) 1024-1034.
- [10] E.E. Ehabe, F. Bonfils, Novel insight into the gel phase of *hevea* natural rubber, *J Rubb Res*, 14 (2011) 1-10.
- [11] S. Amnuaypornsrri, J. Sakdapipanich, S. Toki, B. Hsiao, N. Ichikawa, Y. Tanaka, Strain-induced crystallization of natural rubber: Effect of proteins and phospholipids, *Rubb Chem Technol*, 81 (2008) 753-766.
- [12] Y. Tanaka, L. Tarachiwin, Recent advances in structural characterization of natural rubber, *Rubb Chem Technol*, 82 (2009) 283-314.

- [13] C. Kim, M. Morel, J. Sainte Beuve, A. Collet, S. Guilbert, F. Bonfils, Characterization of natural rubber using size-exclusion chromatography with online multi-angle light scattering Study of the phenomenon behind the abnormal elution profile, *J Chromato A*, 1213 (2008) 181-188.
- [14] C. Kim, J. Sainte Beuve, S. Guilbert, F. Bonfils, Study of chain branching in natural rubber using size-exclusion chromatography coupled with a multi-angle light scattering detector (SEC-MALS), *Eur. Polym. J.*, 45 (2009) 2249–2259.
- [15] S. Dubascoux, C. Thepchalerm, E. Dubreucq, S. Wisunthorn, L. Vaysse, S. Kiatkamjornwong, C. Nakason, F. Bonfils, Comparative study of mesostructure of natural and synthetic polyisoprene by SEC-MALS and AF4-MALS, *J Chromato A*, 1224 (2012) 27-34.
- [16] V.A. Grechanovskii, I.P. Dmitrieva, N.B. Zaitsev, Separation and preliminary characterisation of the protein component from commercial varieties of hevea rubber, *International polymer science and technology*, 14 (1987) 1-4.
- [17] S.N. Gan, Storage hardening of Natural Rubber, *J Membrane Sci, Pure Appl Chem*, A33 (1996) 1939-1948.
- [18] G. Taylor, Dispersion of Soluble Matter in Solvent Flowing Slowly through a Tube, *Proceeding of the Royal Society of London Series A*, 219 (1953) 186-203.
- [19] M. Bello, R. Rezzonico, P. Righetti, Use of Taylor-Aris Dispersion for Measurement of a Solute Diffusion Coefficient in Thin Capillaries, *Science*, 226 (1994) 773-778.
- [20] U.G. Sharma, N; Carbeck, J, Diffusivity of Solutes Measured in Glass Capillaries Using Taylor's Analysis of Dispersion and a Commercial CE Instrument, *Anal Chem*, 77 (2005) 806-813.
- [21] F.V. d'Orlye, A. Varenne; P. Gareil, Determination of nanoparticle diffusion coefficients by Taylor dispersion analysis using a capillary electrophoresis instrument, *J Chromatogr A*, 1204 (2008) 226-232.
- [22] H. Cottet, J. Chamieh, Comparison of Single and Double Detection Points of Taylor Dispersion Analysis for Monodisperse and Polydisperse Samples, *J Chromatogr A*, 1241 (2012) 123-127.
- [23] A.M. Lewandrowska, A. Majchere, A. Ochab-Marcinek, *et al.* Taylor Dispersion Analysis in Coiled Capillaries at High Flow Rates, *Anal Chem*, 85 (2013) 4051-4056.
- [24] H. Cottet, J-P Biron, M. Martin, On the optimization of operating conditions for Taylor dispersion analysis of mixtures, *Analyst*, 139 (2014) 3552-3562.
- [25] W.L. Hulse, J. Gray, R.T. Forbes, Evaluating the inter and intra batch variability of protein aggregation behaviour using Taylor dispersion analysis and dynamic light scattering, *International Journal of Pharmaceutics*, 453 (2013) 351-357.
- [26] W.L. Hulse, R.T. Forbes, A Taylor dispersion analysis method for the sizing of therapeutic proteins and their aggregates using nanolitre sample quantities, *International Journal of Pharmaceutics*, 416 (2011) 394-397.
- [27] A. Lavoisier, J-M. Schlaeppli, Early developability screen of therapeutic antibody candidates using Taylor dispersion analysis and UV area imaging detection, *mAbs*, 7 (2015) 77-83.
- [28] E.P.C. Mes, W.T. Kok, H. Poppe, *et al.*, Comparison of methods for the determination of diffusion coefficients of polymers in dilute solutions: The influence of polydispersity, *J Polym Sci, Part B Polym Physics*, 37 (1999) 593-603.
- [29] H. Cottet, M. Martin, A. Papillaud, *et al.*, Determination of dendrigraft poly-L-lysine diffusion coefficients by Taylor dispersion analysis, *Biomacromolecules*, 8 (2007) 3235-3243.
- [30] J. Deschamps, S.G. Dutremez, B. Boury, *et al.*, Size-Based Characterization of an Ionic Polydiacetylene by Taylor Dispersion Analysis and Capillary Electrophoresis, *Macromolecules*, 42 (2009) 2679-2685.
- [31] F. Oukacine, S. Bernard, I. Bobe, *et al.*, Physico-chemical characterization of polymeric micelles loaded with platinum derivatives by capillary electrophoresis and related methods, *J Control Release*, 196 (2014) 139-145.

- [32] A. Ibrahim, R. Meyrueis, G. Pouliquen, et al., Size and charge characterization of polymeric drug delivery systems by Taylor dispersion analysis and capillary electrophoresis, *Anal Bioanal Chem*, 405 (2013) 5369-5379.
- [33] B. Belongia, J. Baygents, Measurements on the Diffusion Coefficient of Colloidal Particles by Taylor-Aris Dispersion, *J Colloid Interf Sci*, 195 (1997) 19-31.
- [34] F. Oukacine, A. Morel, I. Desvignes, et al. , Size-based characterization of nanoparticle mixtures by the inline coupling of capillary electrophoresis to Taylor dispersion analysis, *J Chromatogr A*, 1426 (2015) 220-225.
- [35] L. Leclercq, S. Reinhard, J. Chamieh, et al., Fast Characterization of Polyplexes by Taylor Dispersion Analysis, *Macromolecules*, 48 (2015) 7216-7221.
- [36] F. Ye, H. Jensen, S.W. Larsen, et al., Measurement of drug diffusivities in pharmaceutical solvents using Taylor dispersion analysis, *J Pharmaceut Biomed Analysis*, 61 (2012) 176-183.
- [37] H. Jensen, S.W. Larsen, C. Larsen, et al., Physicochemical profiling of drug candidates using capillary-based techniques *J Drug Deliv Sci Tec*, 23 (2013) 333-345.
- [38] J. Chamieh, F. Davanier, V. Jannin, et al., Size characterization of commercial micelles and microemulsions by Taylor dispersion analysis, *Int J Pharmaceu*, 492 (2015) 46-54.
- [39] K. MacEwan, D.G. Leaist, Quaternary mutual diffusion coefficients for aqueous solutions of a cationic-anionic mixed surfactant from moments analysis of Taylor dispersion profiles, *Physical Chemistry Chemical Physics*, 5 (2003) 3951-3958.
- [40] N.N. Poulsen, N.Z. Andersen, J. Ostergaard, et al. , Flow induced dispersion analysis rapidly quantifies proteins in human plasma samples, *Analyst*, 140 (2015) 4365-4369.
- [41] S. Latunde-Dada, R. Bott, D. Barker, et al., Methodologies for the rapid determination of the diffusion interaction parameter using Taylor dispersion analysis, *Anal Methods*, 8 (2016) 386-392.
- [42] W.T. Kok, A.J. Tudos, M. Grutters, et al. , Characterization of Asphaltenes by Nonaqueous Capillary Electrophoresis, *Energy & Fuels*, 25 (2011) 208-214.
- [43] M. Ghanavati, H. Hassanzadeh, J. Abedi, Application of taylor dispersion technique to measure mutual diffusion coefficient in hexane plus bitumen system, *AIChE Journal*, 60 (2014) 2670-2682.
- [44] H. Cottet, J-P Biron, M. Martin, Taylor dispersion Analysis of mixtures, *Anal Chem*, 79 (2007) 9066-9073.
- [45] L. Cipelletti, J-P Biron, M. Martin, et al., Measuring Arbitrary Diffusion Coefficient Distributions of Nano-Objects by Taylor Dispersion Analysis, *Anal Chem*, 87 (2015) 8489-8496.
- [46] L. Cipelletti, J-P Biron, M. Martin, et al., Polydispersity Analysis of Taylor Dispersion Data: The Cumulant Method, *Anal Chem*, 86 (2014) 6471-6478.
- [47] B.C. Sekhar, Inhibition of hardening in natural rubber, *Rubb Chem Technol*, 35 (1962) 889-895.
- [48] P. Wyatt, Light scattering and the absolute characterization of macromolecules, *Anal Chim Acta*, 272 (1993) 1-40.
- [49] Y. Yu, P. DesLauriers, D.C. Rohlffing, SEC-MALS method for the determination of long-chain branching and long-chain branching distribution in polyethylene, *Polymer*, 46 (2005) 5165-5182.
- [50] C. Kim, A. Deratani, F. Bonfils, Determination of the refractive index increment of natural and synthetic poly(cis-1,4-isoprene) solutions and its effect on structural parameters, *J Liq Chromatogr Related Technol*, 33 (2010) 37-45.
- [51] H. Cottet, J-P Biron, L. Cipelletti, et al. , Determination of Individual Diffusion Coefficients in Evolving Binary Mixtures by Taylor Dispersion Analysis: Application to the Monitoring of Polymer Reaction, *Anal Chem*, 82 (2010) 1793-1802.
- [52] W. Burchard, Solution Properties of Branched Macromolecules, *Adv Polym Sci*, 143 (1999) 113.

[53] L.J. Fetters, N. Hadjichristidis, J.S. Lindner, J.W. Mays, Molecular Weight Dependence of Hydrodynamic and Thermodynamic Properties for Well-Defined Linear Polymers in Solution, J Phys Chem Ref Data, 23 (1994) 619-640.

ACCEPTED MANUSCRIPT

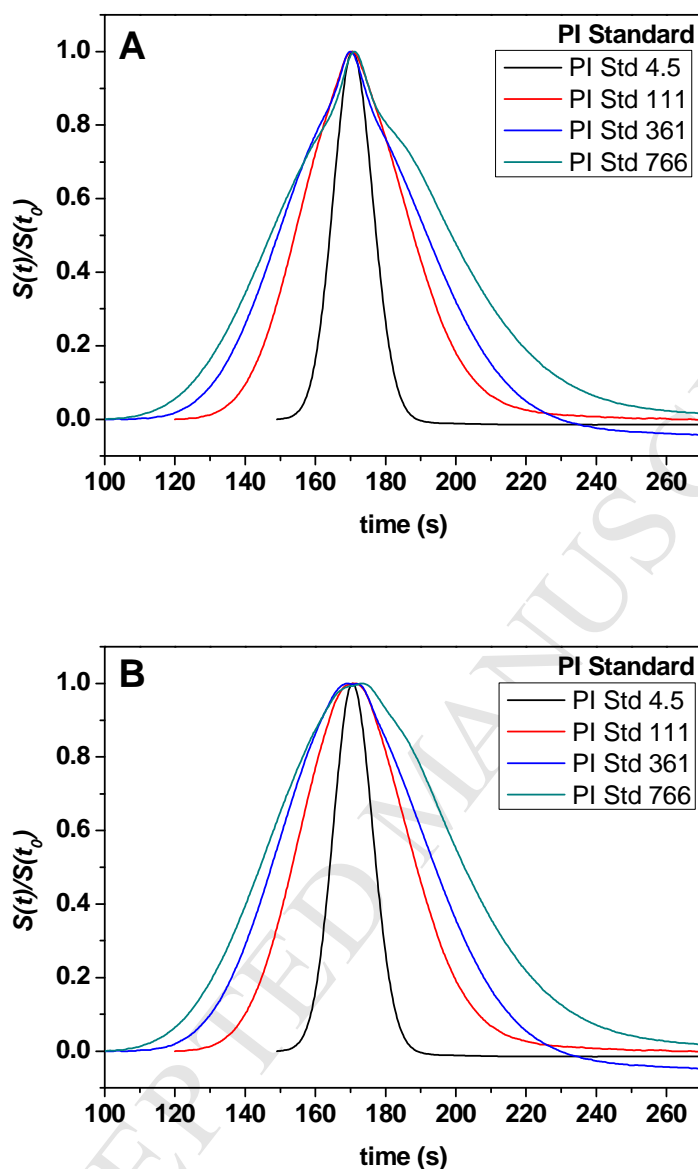


Figure 1. Taylorgrams obtained for PI standards in cyclohexane before (A) and after subtraction of the contribution due to small molecules (B). Experimental conditions: fused silica capillary 40 cm (30 cm to the detector) x 50 μm i.d. Mobile phase: cyclohexane. Hydrodynamic injection: 0.3 psi, 3 s. Mobilization pressure: 1.0 psi. Temperature: 25 $^{\circ}\text{C}$. Samples: PI standards 4.5, 111, 361, and 766 $\times 10^3$ g/mol at 1 g/L in cyclohexane. Detection at 200 nm. The normalized Taylorgram is presented ($S(t)/S(t_0)$).

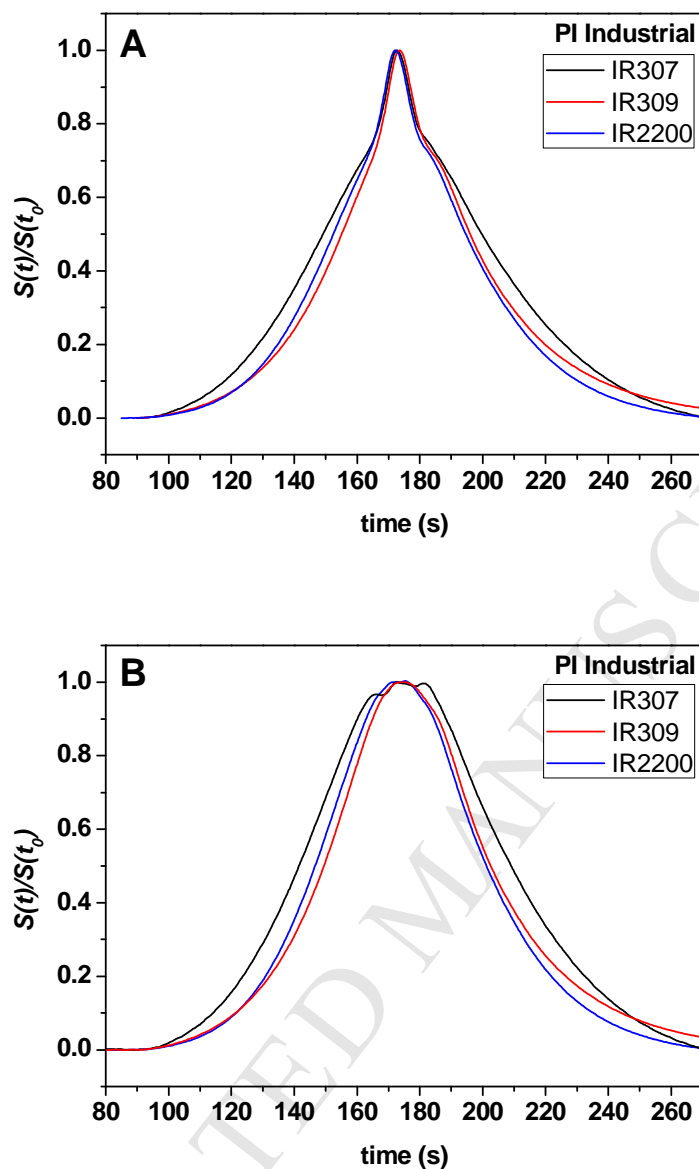


Figure 2: Taylorgrams obtained for industrial PI in cyclohexane before (A) and after subtraction of the contribution due to small molecules (B). Same conditions as in Figure 1.

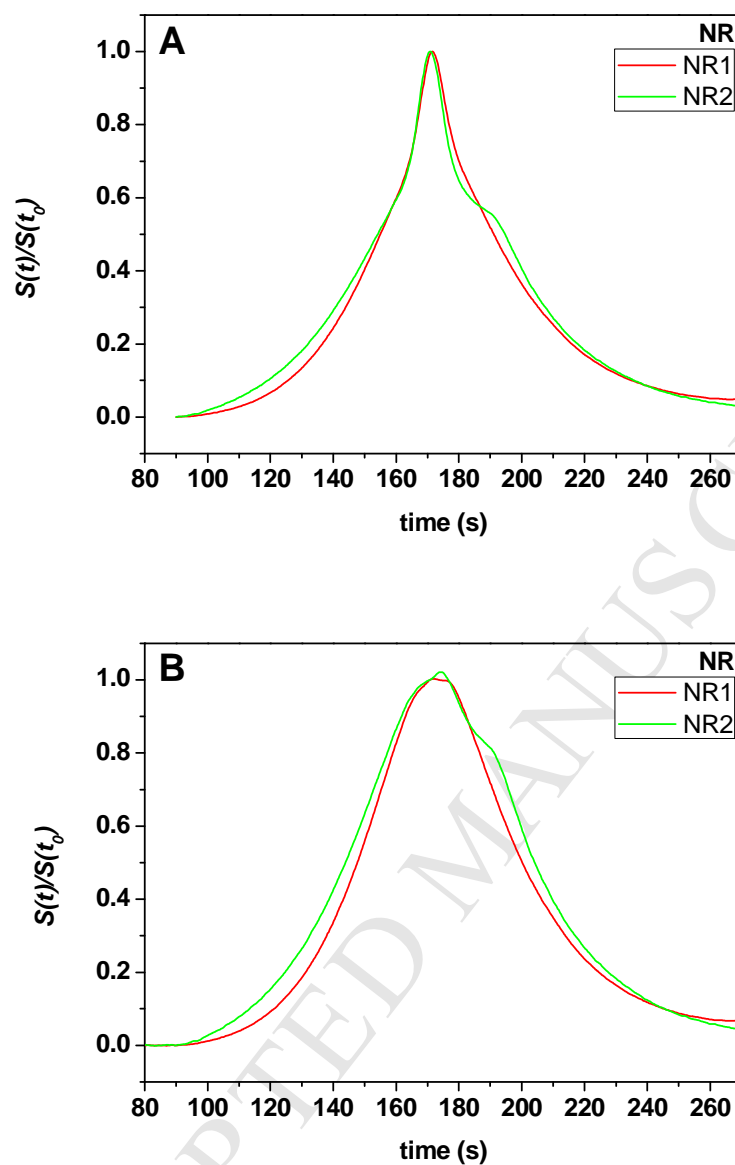


Figure 3. Taylorgrams obtained for natural rubbers (NR) in cyclohexane before (A) and after subtraction of the contribution due to small molecules (B). Same conditions as in Figure 1.

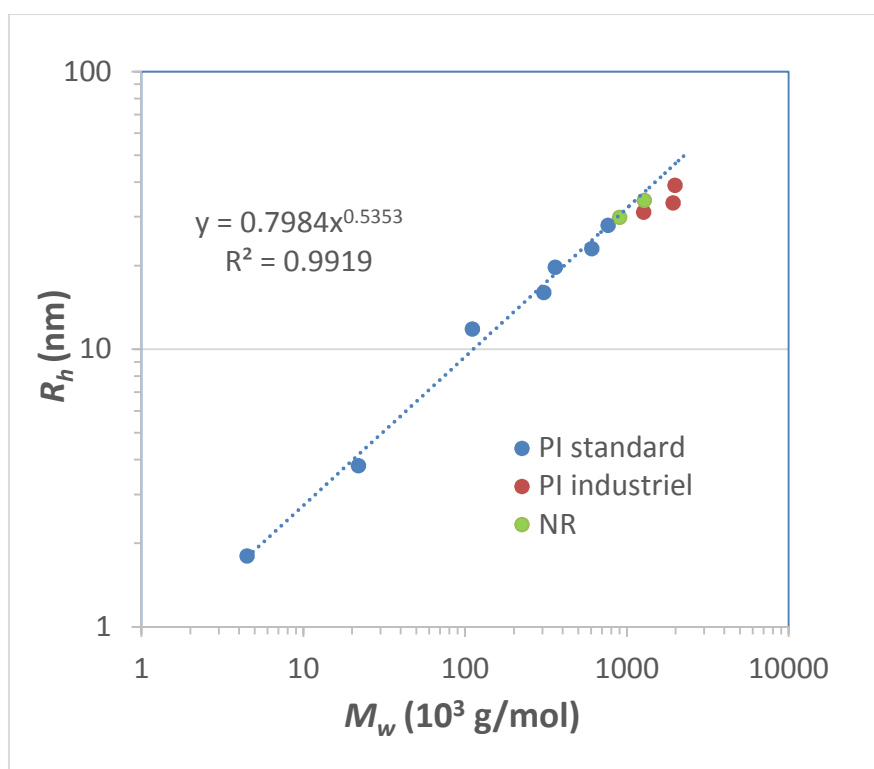


Figure 4: Log-log plot of the TDA hydrodynamic radius (method 1, after subtraction of the small molecule contribution) versus the molar mass for the PI standards, PI industrial and for NR. The dotted line is a power law fit for the PI standards. See Table 1 for the numerical values.

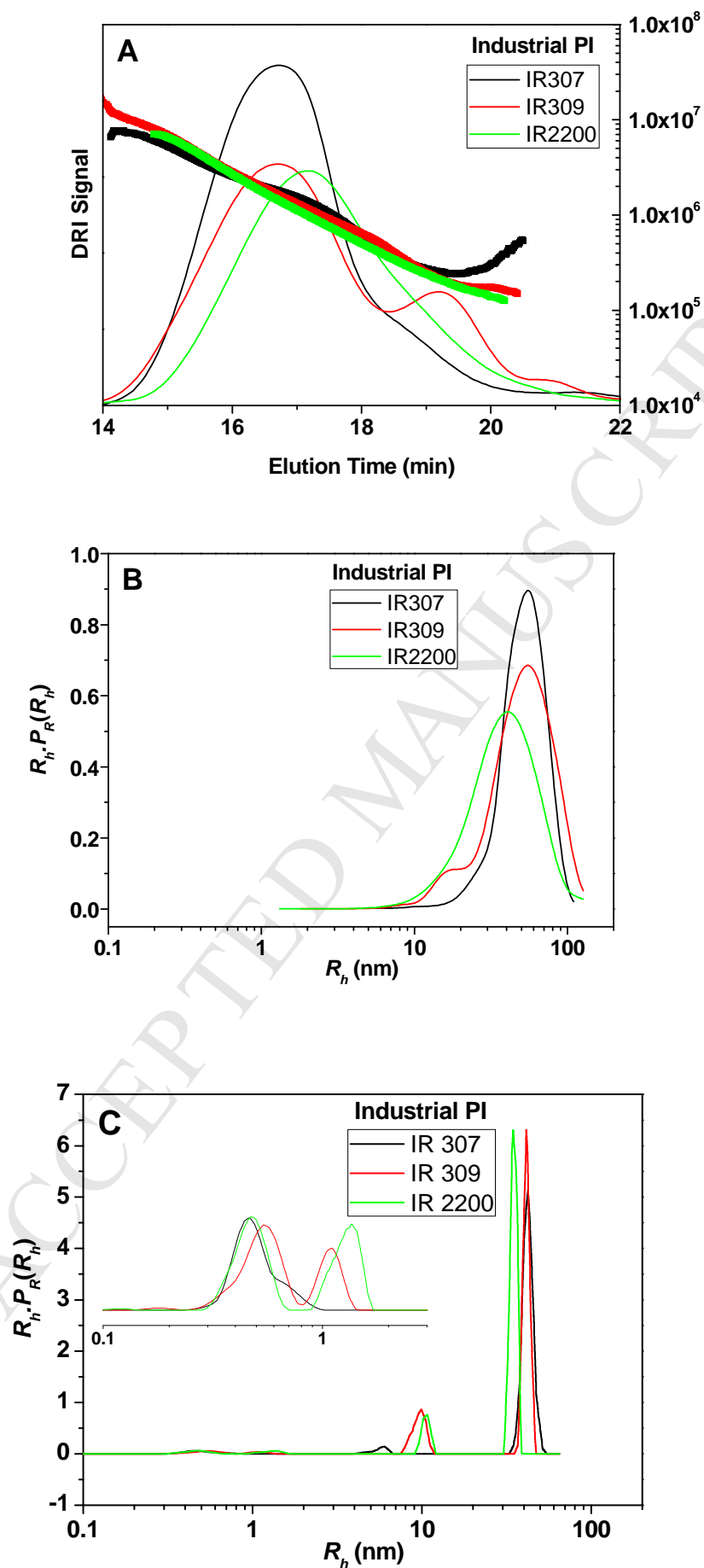


Figure 5: Elution profiles obtained by SEC (RI signal, A) and R_h distributions obtained by SEC-MALS (B) or TDA using *method 2* (C), for the 3 industrial PI samples. Experimental conditions: SEC in tetrahydrofuran, two Waters HMW6E columns (20 μm , 300 mm x 7.8 mm I.D.) maintained at 45°C, flow rate of 0.65 mL.min⁻¹; TDA conditions as in Figure 2. The insert in Figure 2C is an expanded view. R_h distributions obtained by SEC-MALS are based on the assumption that $R_g/R_h = 1.78$.

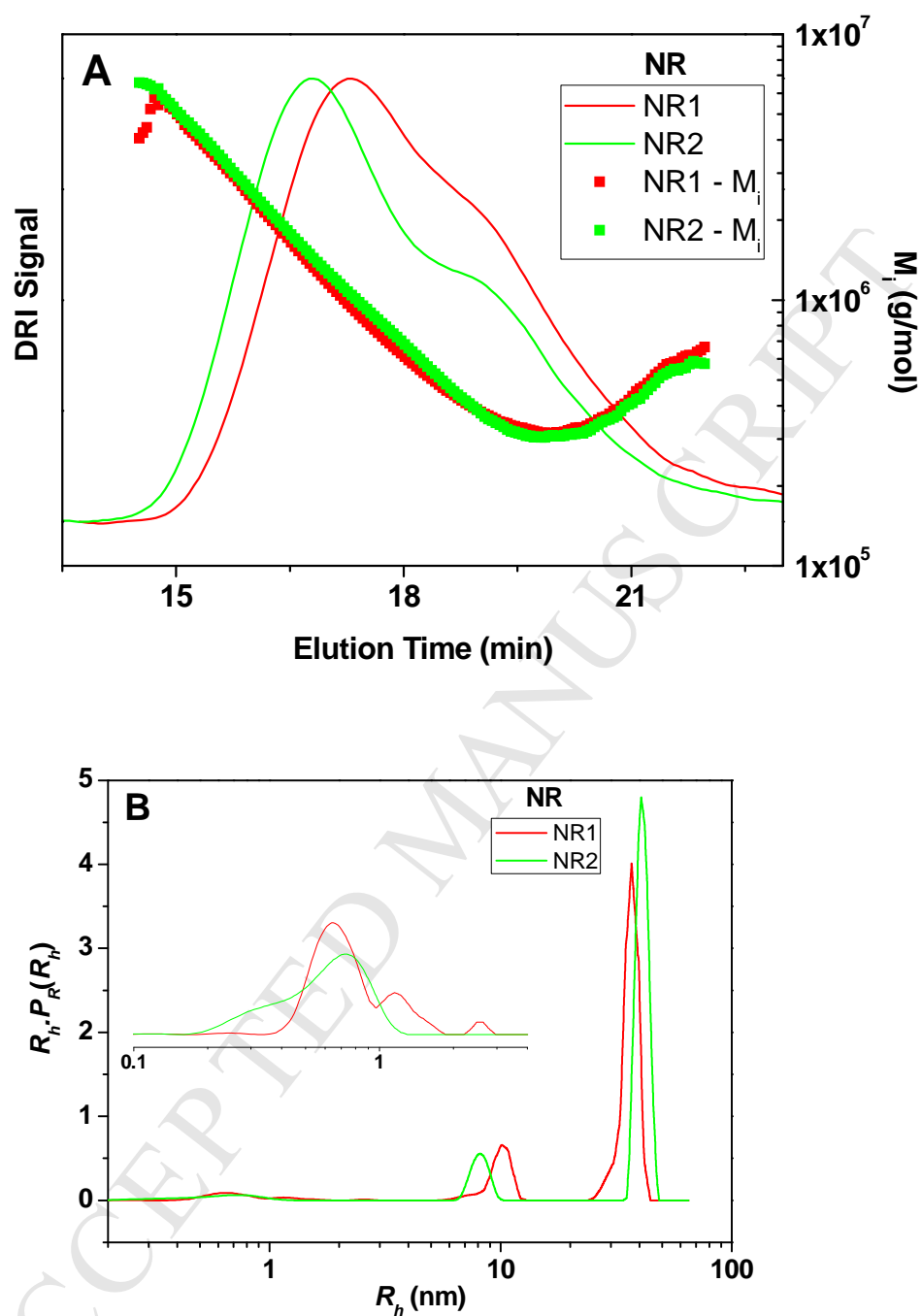


Figure 6: Elution profiles obtained by SEC (RI signal, A) and R_h distributions obtained by TDA using *method 2* (B) for two NR samples. Experimental conditions: SEC, see Figure 5; TDA, as in Figure 3. Insert in Figure 6B is an expanded view.

Table 1: Number- and weight-average molar masses (M_n , M_w), hydrodynamic radius (R_h +/- one standard deviation) for natural and synthetic PI. M_n , M_w are determined by SEC-MALS, except for the PI standards^a. R_h is determined by TDA, using both method.

Sample	Type	M_n (kg.mol ⁻¹) (1)	M_w (kg.mol ⁻¹) (1)	$\langle R_h \rangle_{TDA}$ (nm) ^b Method 1		$\langle R_h \rangle_{CRLI}$ (nm) ^b Method 2	
				with subtraction ^c	without subtraction ^d	Integration for $R_h > 3$ nm ^e	Integration for $R_h > 0$ ^f
PI4.5	PI Standard	4.5	4.5	1.8 ± 0.01	1.8 ± 0.01	1.8 ± 0.01	1.8 ± 0.01
PI22	PI Standard	22	22	3.8 ± 0.1	3.8 ± 0.1	3.8 ± 0.04	3.8 ± 0.04
PI111	PI Standard	111	111	11.8 ± 0.2	11.6 ± 0.2	11.5 ± 0.1	11.4 ± 0.1
PI307	PI Standard	307	307	16.0 ± 0.2	15.7 ± 0.2	16.6 ± 0.1	16.2 ± 0.1
PI361	PI Standard	361	361	19.7 ± 0.8	19.4 ± 0.8	20.2 ± 0.4	19.9 ± 0.4
PI608	PI Standard	608	608	23.0 ± 1.6	16.7 ± 1.3	22.0 ± 0.2	16.2 ± 0.5
PI766	PI Standard	766	766	27.9 ± 0.7	27.3 ± 0.6	28.5 ± 0.7	28.0 ± 0.7
PI1040	PI Standard	1040	1040	28.9 ± 0.05	27.4 ± 0.2	29.9 ± 0.2	28.5 ± 0.3
IR307	PI Industrial	1079	1986	40.1 ± 0.5	38.9 ± 0.5	41.0 ± 0.5	39.8 ± 0.4
IR309	PI Industrial	652	1935	35.0 ± 0.1	33.6 ± 0.2	34.9 ± 0.06	33.5 ± 0.03
IR2200	PI Industrial	584	1271	32.4 ± 0.3	31.1 ± 0.3	31.7 ± 0.2	30.3 ± 0.3
NR1	NR Industrial	595	902	31.7 ± 1.5	29.8 ± 1.4	31.4 ± 1.5	29.5 ± 1.4
NR2	NR Industrial	711	1282	36.2 ± 0.4	34.3 ± 0.5	36.4 ± 0.6	34.4 ± 0.6

^a For standard polyisoprene, all M_n and M_w were given by the supplier.

^b Three repetitions in average

^c Average R_h obtained by integration of the taylorgram after subtraction of the small molecule contribution (*method 1* applied to Figures 1B, 2B, 3B).

^d Average R_h obtained by integration from the whole taylorgrams including the small molecule contribution (*method 1* applied to Figures 1A, 2A, 3A).

^e Average R_h obtained by *method 2* (CRLI, eq. (9)) by integration of the $P_R(R_h)$ distribution for R_h higher than 3 nm.

^f Average R_h obtained by *method 2* (CRLI, eq. (9)) by integration of the whole $P_R(R_h)$ distribution.

1
2
3
4
5
6
7
8
9
10
11

Highlights

- Sizing natural or synthetic polyisoprenes by miniaturized Taylor dispersion analysis
- Full hydrodynamic radius distribution of polyisoprenes from Taylor dispersion analysis
- Taylor dispersion analysis has a better size-resolution compared to SEC
- No abnormal elution of the microaggregates in Taylor dispersion analysis
- Short analysis time (a few min) and low sample and solvent consumption

ACCEPTED MANUSCRIPT

Comment citer ce document :

Biron, J.-P., Bonfils, F., Cipelletti, L., Cottet, H. (2018). Size-characterization of natural and synthetic polyisoprenes by Taylor dispersion analysis. *Polymer Testing* , 66, 244-250. , DOI : 10.1016/j.polymertesting.2018.01.017