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Mitral valve disease—morphology and mechanisms

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Abstract

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The authors declare no competing interests.

Author contributions

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Mitral valve disease is a frequent cause of heart failure and death. Emerging evidence indicates that the mitral valve is not a passive structure, but—even in adult life—remains dynamic and accessible for treatment. This concept motivates efforts to reduce the clinical progression of mitral valve disease through early detection and modification of underlying mechanisms. Discoveries of genetic mutations causing mitral valve elongation and prolapse have revealed that growth factor signalling and cell migration pathways are regulated by structural molecules in ways that can be modified to limit progression from developmental defects to valve degeneration with clinical complications. Mitral valve enlargement can determine left ventricular outflow tract obstruction in hypertrophic cardiomyopathy, and might be stimulated by potentially modifiable biological valvular–ventricular interactions. Mitral valve plasticity also allows adaptive growth in response to ventricular remodelling. However, adverse cellular and mechanobiological processes create relative leaflet deficiency in the ischaemic setting, leading to mitral regurgitation with increased heart failure and mortality. Our approach, which bridges clinicians and basic scientists, enables the correlation of observed disease with cellular and molecular mechanisms, leading to the discovery of new opportunities for improving the natural history of mitral valve disease.

Introduction

Mitral valve insufficiency is a major source of morbidity and death worldwide and a frequent cause of heart failure, with complications that include arrhythmia, endocarditis, and sudden cardiac death.^{1,2} Structural deficiencies in the mitral valve and secondary changes induced by abnormal ventricular size and deformation are implicated in the development of these valvular lesions.³ However, the effect of mitral insufficiency on cardiac function is more than purely mechanical, whereby pump function is maintained at the expense of elevated filling pressures, but extends to impaired contractility and electrical instability.

Mitral valve diseases (MVDs) that lead to valve insufficiency have long been conceived as inexorable processes (Figure 1a) for which therapeutic options are limited to surgical valve repair or replacement, with the underlying mechanisms inaccessible. The underlying genetic mutations and mechanisms of mitral valve dysfunction associated with myxomatous degeneration have remained elusive.⁴ In mitral valve lesions associated with ventricular disease, the leaflets are considered biologically passive and fixed in size relative to the enlarged or narrowed ventricle and not an available target for therapy.⁵ Valve plasticity—defined as the potential for change in cellular phenotype and behaviour—and altered leaflet matrix and micromechanics, have been considered in studies of valve development. However, MVD had not been associated with modified adult valve biology until the 2004 discovery in a mouse model of Marfan syndrome, in which a mutation in an extracellular matrix (ECM) protein alters the regulation of aorta and valve cell biology and thereby creates opportunities for modifying the course of disease.⁶ We believe that the mitral valve is not a passive structure, but—even in adult life—a ‘living valve’ that undergoes active changes.³ The interface between valve imaging, genetics, biology, and biomechanics can help us to understand the mechanisms of mitral valve function and those that might be applied to novel therapies for mitral regurgitation (Figure 1b). Our mission is to reduce the clinical progression of MVD by early detection and modification of underlying mechanisms.

This approach can also be applied to conditions that lead to mitral regurgitation, in which the mitral leaflets are either too long or small in relation to the remodelling left ventricle, or where they are too stiff or compliant. For example, in patients with mitral valve prolapse (MVP), the leaflets are excessively long and typically show myxomatous degeneration with increased leaflet compliance.^{7–9} In ischaemic and functional mitral regurgitation, which often occurs in patients surviving a myocardial infarction (MI) or with dilated cardiomyopathy,¹⁰ the leaflet area is deficient relative to the dilated left ventricle,^{11,12} and the leaflets are stiffer and undergo fibrosis,¹³ so that any compensatory leaflet remodelling is insufficient to prevent regurgitation. In individuals with hypertrophic cardiomyopathy (HCM), although the underlying genetic mutation is associated with a sarcomeric protein, elongated leaflets contribute to both mitral regurgitation and left ventricular outflow tract (LVOT) obstruction, and are disproportionate to the reduced cavity size and abnormal LVOT.^{14–16}

Imaging, most commonly cardiac ultrasonography, has a major role in the understanding of these structural mechanisms,¹⁷ and reveals mitral valve biology by allowing correlations of valve structure, biomechanics, and haemodynamics with basic cellular, molecular, and genetic findings and, consequently, the development of new physiological therapies. In this Review, we provide an understanding of the mitral valve as a functional and interactive organ, beginning with consideration of how normal valve structure and biomechanical function are ideally matched as a result of choreographed development. We then review how inherited and acquired perturbations alter valve function, and consider how we can access controlling pathways to maintain physiological heart function and prevent clinically apparent disease.

Mitral valve structure

The mitral valve has two leaflets: the anterior leaflet, contiguous with the aorto-mitral curtain; and the posterior leaflet composed of three scallops, P1–P3 (Figure 2a). Both leaflets attach to the dynamic mitral annulus at their basal ends, whereas multiple chordae tendineae emerge from the ventricular surfaces and attach distally to the papillary muscles. This complex shape enables a delicate balance of force to sustain unidirectional blood flow through the large and dynamic orifice of the mitral valve. Both leaflets are divided into three radially located zones from annulus to free edge, with locally unique ultrastructural composition and biomechanics. The annulus-to-leaflet transition zone, which spans ~1 cm from the mitral annulus radially toward the leaflet edge, contains atrial myocytes^{18,19} and cells that express smooth muscle α actin.²⁰ Nerve fibres and terminals extend into this region from the mitral annulus, maintaining electro-physiological continuity with the rest of the heart,^{21,22} and small blood vessels span this leaflet region.^{19,23–25} The translucent smooth zone that forms the central belly of the leaflet consists of thin, but dense, collagenous networks and vimentin-positive fibroblasts that actively synthesize the ECM in response to mechanical loading.²⁶ The distal rough zone at the leaflet edge is thick and rich in glycosaminoglycans and retains water to sustain the compressive stresses from coaptation.²⁷ The cross-sectional structure of the normal mitral leaflets is analogous to the aortic valve, with three well-defined tissue layers—the atrialis, spongiosa, fibrosa/ventricularis (Figure 2b and Figure 3)—each with a different thickness, characteristic cells, matrix composition, and

biomechanical features that contribute to its proper function. The atrialis layer is covered with endothelium and contains a subendothelial connective tissue layer of lamellar collagen and elastin sheets, which extends from the left atrial endocardium into the leaflet. This layer sustains high-frequency, large tensile deformations in systole, with the interspersed elastin enabling elastic recoil in diastole. The presence of left atrial myocytes and smooth muscle cells adjacent to nerve endings can transiently modulate leaflet stiffness and deformation during closure.²⁸ Just beneath the atrialis is the spongiosa, which contains loosely arranged collagen and is rich in glycosaminoglycans; this layer gets thinner towards the mitral annulus, and becomes prominent towards the leaflet free edge. Retention of water in the spongiosa aids in resisting compressive loading imposed by coaptation of the leaflets.²⁷ The fibrosa is connected to the mitral annulus and faces the high-pressure left ventricle; this layer is composed predominantly of collagen fibres that are arranged parallel to the leaflet free edge, are densely packed, and flexibly crimped or wavy.²⁹ At the sites of chordal insertion, the fibrosa transitions from a planar collagenous structure to a cylindrical collagenous chord that enables the gradual transition of forces between the chordae and leaflets without regional kinking.^{24,30} Conversely, the chordae tendineae are composed of a cylindrical collagen core within an elastin sheath, and have demonstrable viscoelastic properties—their extensibility decreases with increasing strain rates and varies with chordal size and location.^{31,32}

The leaflets are populated with cells that have unique features and sustain valve homeostasis throughout life.³³ The leaflets are covered by a continuous endothelial cell layer on both atrial and ventricular aspects. The deep sub-endothelial layers contain mainly valvular interstitial cells (VICs), a population of cells unique to cardiac valves³⁴ that originate from endocardial endothelial cells.^{35–38} Quiescent VICs are noncontractile fibroblastlike cells, do not express smooth muscle α actin, but are positive for vimentin and chondromodulin1, and contribute to homeostatic remodelling of matrix constituents with relatively low turnover.³⁹ During tissue remodelling in embryonic development and conditions such as myxomatous degeneration, epicardial-derived cells migrate into the posterior leaflet,^{40,41} and VICs transition to an activated myofibroblast-like phenotype, expressing smooth muscle-associated contractile proteins, matrix metallo-proteinases, and inflammatory cytokines that rapidly remodel the extracellular milieu.^{42–44} Many of the same factors that modulate postnatal VIC phenotype in MVD also have important functions during embryonic development, suggesting that the VIC phenotype is a common link between primitive embryonic mesenchyme and postnatal plasticity to adapt to the dynamic valve environment. Mitral valve endothelial and interstitial cells have reciprocal interactions that favour normal homeostasis.⁴⁵

Mitral valve function during the cardiac cycle is remarkably dynamic in response to myocardial contraction and varying transvalvular pressure. The mitral annulus undergoes a reduction in presystolic area contributed by left atrial contraction, which facilitates early leaflet motion and adequate leaflet coaptation.⁴⁶ When such atriogenic annular reduction is ablated, equivalent reduction in the mitral annulus is possible via ventriculogenic contraction, but this contraction is delayed and results in increased mitral regurgitation.⁴⁶ The dynamic interaction between the surrounding myocardial structures and the valve leaflets governs the leaflet coaptation⁴⁷ and, ultimately, the mechanical strain and stress on

the valve extracellular matrix and cells.^{48–50} Under normal conditions in healthy individuals, the anterior leaflet undergoes large deformations with peak circumferential strains (parallel to the mitral annulus) ranging from 2.5% to 5.0%, and peak radial strains (perpendicular to the mitral annulus) ranging from 7.8% to 22.0%.^{29,51–53} In the circumferential direction, peak strains are concentrated in the free edge of the leaflet, as the leaflets are stretched between the marginal chordae. In the radial direction, peak strains are concentrated in the soft belly region of the anterior leaflet, because that leaflet has the largest systolic curvature.^{51,54} Principal leaflet strains rotate from the circumferential direction during diastole to the radial direction in systole; this rotation corresponds with the diastolic tension in the secondary chordae that controls leaflet motion into the LVOT, and the large systolic leaflet curvature and chordal tethering that increase radial leaflet strain.^{51,54} These dynamic shifts in leaflet strains from diastole to systole align with orientation of the collagen and elastin fibril networks in the leaflet. Although tempting to postulate that optimal valve biomechanics are supported by such favourable fibre orientation, this orientation might actually be the result of deformations during fetal and neonatal valve development. Anecdotal evidence of this hypothesis is seen in patients with stunted left ventricular growth, in whom loss of transmitral blood flow or strain results in an immature valve with chordal fusion and immobile leaflets.⁵⁵ An abundance of glycosaminoglycans in the spongiosa indicates that an inherent viscoelastic and strain rate-dependent behaviour exists in the valve leaflets.

The adaptive matching of leaflet biomechanics to the needs for normal leaflet excursion and closure indicates why altered biomechanics in disease might impair coaptation. Transfer of the mechanical strains imposed on the ECM to the resident valve cells might also govern valve homeostasis and behaviour. While the matrix might dampen the magnitude of cellular strains, VICs cultured at cyclic matrix-level strains respond with altered expression of collagens, proteoglycans, and growth factors such as those of the transforming growth factor β (TGF β) family.^{56–60} Furthermore, VIC-mediated tissue stiffness can be modulated by the endothelium-dependent mediators nitric oxide and endothelin acting on contractile VIC phenotypes.⁶¹ In addition to this interplay between mechanics and biology, nonphysiological oscillatory flows induce inflammatory activation of the valvular endothelium that can induce VIC activation, but the precise mechanisms are still unknown.⁶² Different shear stress patterns on the inflow and outflow valve surfaces might induce heterogeneous mechanical responses.³³ The inherent dynamism of valve action, involving both tissue and fluid stresses, strongly suggests that these diverse signals are integrated during normal valvular cell function. Disease states perturb the mechanical strains and ECM composition and can shift these dynamic equilibria towards pathological remodelling. Understanding this interplay of biomechanics and valve biology can substantially inform long-term repair strategies, to promote beneficial as opposed to detrimental changes.

Normal mitral valve dynamics and competence also determine the fluid dynamics of left ventricular filling and ejection. During early-to-mid diastolic filling, vortical flow structures that form beneath the anterior leaflet direct the inflow towards the aortic valve and enable systolic ejection without substantial kinetic energy losses.⁶³ Elongation of the anterior leaflet, which occurs in patients with hypertrophic obstructive cardiomyopathy, or loss of annular dynamics and restricted leaflet motion, as seen in individuals with functional mitral

regurgitation, might alter these vortical structures and impair efficient ventricular pumping.⁶⁴ Loss of normal vortical structures might also increase residual blood in the left ventricle between consecutive cardiac beats, and therefore increase diastolic filling pressures, cause adverse ventricular dilatation, and biological remodelling.⁶⁵ As flow-based imaging to detect cardiac dysfunction emerges, the role of the mitral valve and its lesions in governing efficient ventricular pumping might be further revealed.⁶⁶

Mitral valve development

Early mitral valve development is complex and errors in early morphogenesis can contribute to MVD.^{67–72} The first cell-fate decision in the embryo that contributes to mitral valve formation is the segregation of the endocardial and myocardial lineages.^{73,74} This process occurs before, or at the onset of, gastrulation within the cranial and cardiac lateral plate mesoderm, marked by the transcription factor mesoderm posterior protein 1, or earlier in the mesoderm when haematopoietic and cardiac lineages are determined by brachyury protein-expressing cells.⁷⁴ These prospective valve cells emerge as vascular endothelial growth factor receptor 2 (VEGFR2)-positive cells.⁷⁴ Some VEGFR2-positive cells arising when cells of the second heart field migrate from the pharyngeal mesoderm are also likely to contribute to the endocardium (Figure 4).⁷⁴

Mitral valve development begins soon after looping of the early embryonic heart tube. In response to growth factors including several proteins of the TGF β super family (TGF β 1–3, BMP2–4), a subset of the endocardial cells of the early embryonic heart become activated and transform into a mesenchymal phenotype, known as endothelial-to-mesenchymal transition (EMT; Figure 5).^{71,75} The endothelial cells that do not undergo EMT express NFATc1, which suppresses important transcriptional factors needed for the phenotypic transition of the cells to proliferate and elongate to form the maturing leaflets.⁷⁶ NFATc1 limits EMT by suppression of its positive regulators Snail and Slug.⁶⁹ The molecular and cellular events that occur after this initial EMT are complex and poorly understood. Post-EMT leaflet maturation involves migration and proliferation of mesenchymal cells to distend the forming cushions into the lumen, under the control of growth factors,^{72,75,77} followed by their gradual thinning and elongation. One hypothesis for this leaflet maturation is that cell proliferation results in polarized, aligned cell arrays that collectively migrate and extrude leaflet tissue into the lumen, referred to as a convergent extension process.⁶⁹ Indeed, microscopic analysis of developing valves shows alignment of cells during post-EMT and neonatal development.⁶⁹ Concomitant with the early remodelling phase is differentiation of the cushion mesenchyme into interstitial cells.⁷⁴ Based on this fundamental change in cell phenotype, the matrix surrounding these fibroblasts matures into a highly organized lamellar and fibrous tissue, making the valve more rigid and thereby able to handle the increasing haemodynamic load of the beating heart.⁷⁴ Slight defects in any of these fundamental processes might lead to structural and functional valve disease that presents in postnatal life and can progress into degenerative changes in adult life.

Mitral valve ageing

While mitral valve structure remains largely preserved with age, ultrastructural and cellular changes that indicate chronic mechanical loading have been reported. For example, some investigators have reported high cellularity, closely packed, parallel, and thick collagen fibres, very few elastic fibres, and dense muco-polysaccharides in the valves of children aged >10 years.⁷⁸ Conversely, less cellular, thick, dense, nonparallel collagen bundles, with slightly increased number of elastic fibres, and reduced density of muco-polysaccharides were observed in mitral valves of individuals in their second to fifth decades of life.⁷⁸ Finally, marked reduction in cellularity, disoriented collagen fibres, and increased elastin fibres with severely reduced muco-polysaccharides were seen in valves of those aged ≥60 years.⁷⁸ Other investigators have identified age-related changes that differ between the anterior and posterior leaflets.⁷⁹ A study from patients aged 72–91 years without cardiac disease confirmed the well-known calcification concentrated along the annulus or chordal insertion regions, but remarkably minimal changes in leaflet mechanical properties.⁸⁰ Unlike the semilunar cardiac valves, age-dependent biological changes in the mitral valve are not well understood.

Mitral valve prolapse

Echocardiographic diagnosis

MVP affects nearly one in 40 individuals,⁸¹ and is the leading surgical indication for mitral regurgitation.⁸² Although screening of the general population indicates that many individuals with MVP have few clinical symptoms,^{81,83} in large clinical populations the disease is strongly associated with heart failure, need for valve surgery, endocarditis, and death in the ~20% of those with moderate-to-severe mitral regurgitation and left ventricular ejection fraction <50%.⁸⁴

Echocardiography has been central to the diagnosis of MVP as leaflet displacement beyond the normal range relative to the mitral annulus.¹⁷ Diagnostic specificity is required to study the genetic mutations associated with this MVP. Older studies lacked this specificity and suggested that MVP prevalence was as high as 10–15% or more,⁸⁵ often on the basis of 2D views that seemed to show prolapse because of the normal saddle shape of the valve, without actual pathological leaflet displacement.^{81,85–89} 3D echocardiographic demonstration of this saddle shape led to recognition that prolapse could be defined most specifically by leaflet displacement relative to the anterior and posterior high points of the annular saddle when imaged in long-axis views (Figure 6a).^{87–89} Additional assessment focuses on leaflet displacement relative to each other, leading to malcoaptation.^{90,91} New diagnostic criteria have revised the prevalence of MVP to 2.4% without loss of sensitivity.⁸¹ Improved diagnostic specificity has also provided a firm foundation for genetic studies of this frequently inherited disease.⁹²

For cardiologists or sonographers, mitral prolapse is defined by an abnormal systolic bulging or ‘billowing’ of one or both leaflets towards the left atrium (≥2 mm beyond a line connecting the annular hinge points in a long-axis view; Figures 6b,c).^{88,93} For cardiac surgeons, ‘billowing’ designates bulging of the leaflet bodies into the atrium while the

leaflet tips and coaptation point remain on the ventricular side. ‘Prolapse’ indicates an overriding of the annulus by a leaflet edge with displacement of the coaptation point into the atrium, and is therefore restricted to MVP complicated by moderate-to-severe mitral regurgitation, which does not include the overall spectrum of MVP disease, particularly in familial studies.⁹³ Leaflet ‘flail’ implies coaptation failure with eversion of the free edge of a leaflet into the left atrium, usually consequent to chordal rupture (Figure 6d). Flail most frequently affects the posterior leaflet and is generally associated with severe mitral regurgitation, leading to ventricular remodelling and conveying a poor outcome with medical management.^{94–99}

Other valve characteristics evaluated by echocardiography in MVP are leaflet length and thickness, mitral annulus diameter, and mitral regurgitation severity. Normal leaflet length averages are 22–23 mm for the anterior and 12–13 mm for the posterior leaflet; normal annulus diameter is 28–30 mm.^{100,101} In patients with MVP, leaflets are generally elongated and thickened, and the annulus is enlarged.¹⁰² Mid-leaflet thickness in the parasternal long-axis view in diastole (which is normally 2–3 mm) is increased in 14% of patients with moderate MVP (to ~3–8 mm) and up to 40% of those with severe forms of the disease.⁸⁸ Notably, leaflet thickness measured by echocardiography depends on the acquisition mode (harmonics) and acoustic wavelength,¹⁰³ typically overestimating the dimensions measured during surgery. Thickness also decreases as the leaflet elongates in systole by Poisson’s effect in soft materials.¹⁰⁴ ‘Classic’ MVP refers to superior leaflet overshoot >2 mm and echocardiographic leaflet thickness ≥5 mm, associated with more severe dysfunction.^{81,105} Mitral regurgitation varies from trivial to severe according to the failure of coaptation driven by chordal elongation or rupture and annular enlargement.¹⁰⁶ Regurgitation in patients with MVP is typically greatest in mid-to-late systole, whereas functional mitral regurgitation is often seen during early-to-mid systole.^{107,108} Severe mitral regurgitation seems to occur in ~10% of patients with MVP and most with flail. Mitral valve lesions also seem to worsen with patient age, with leaflet thickening and increasing mitral regurgitation, in about one-quarter to one-third of patients, although an asymptomatic phase limits accurate knowledge of natural history.^{109,110} Interactions among mitral apparatus components augment mitral regurgitation. For example, annular flattening, observed with severe mitral regurgitation, can increase chordal stresses and promote chordal rupture;^{111,112} left ventricular remodelling can exacerbate malcoaptation through apical tethering of nonprolapsed leaflet segments (Figure 6c);⁹⁰ and mitral regurgitation can beget mitral regurgitation through leaflet remodelling induced by nonphysiological strains and shear stress.

Genetic studies of MVP

Familial genetic studies have improved our understanding of MVP and its diagnosis.¹¹³ First, some patients with MVP do not have diagnostic leaflet displacement into the left atrium, but have features that suggest early, potentially prodromal forms.^{114,115} These prodromal forms are based on anterior displacement of the coaptation point as a manifestation of excessive posterior leaflet length; in the familial context, such individuals share genetic variation with relatives manifesting fully developed MVP.^{114,115} The prodromal morphology of MVP has allowed identification of a new locus for autosomal dominant MVP on chromosome 13.¹¹⁵ Although the natural history of prodromal forms of

MVP is unknown, they might indicate genetically susceptible individuals in whom early therapy can limit clinical progression. Recognizing that leaflet thickness, displacement, and mitral regurgitation vary widely within the same family allows us to include all family members who will be informative for genetic linkage.^{114–116}

Histopathology and biomechanics

Degenerative MVD leads to mitral valve incompetence with annular dilatation, excess valvular tissue resulting in hooding, doming, or billowing of a leaflet portion or entire leaflet, especially the posterior, and chordal rupture resulting in a flail leaflet with prominent mitral regurgitation (Figures 6 and 7). Two forms of degenerative MVD have been reported. So-called Barlow disease is associated with excess connective tissue, with redundant, thickened, yellow, and gelatinous leaflets, marked annular dilatation, elongated and thin (sometimes ruptured) or thick (frequently calcified) chordae, disrupted collagen and elastic layers, and excess acidic mucopolysaccharides. Fibroelastic deficiency is characterized by thin, translucent leaflets deficient in collagen, elastin, and proteoglycans with only moderate annular dilatation and with focal chordal elongation or rupture.^{44,117–120} Pathologically thickened lesions in these otherwise thin leaflets appear in leaflet segments exposed to mitral regurgitation, leading to the hypothesis that these are secondary leaflet changes.⁴⁴

In myxomatous leaflet degeneration, leaflet thickness varies with the degree of fibrosis in the atrialis layer and the accumulation of myxoid ECM rich in glycosaminoglycans.^{121–127} The most consistent marker of degenerative MVD is disruption of the fibrosa by myxomatous ECM (Figure 8).^{44,121} Movat's stain, which delineates ECM components, shows expansion of the spongiosa by loose, amorphous ECM strongly positive for proteoglycans, with diminished staining for collagen and fragmented elastin.¹²² Similarly, myxoid replacement of the collagen core of chordae tendineae is frequent, leading to their rupture (Figure 8a).¹²² Loss or severe disruption of the elastic lamina beneath the atrialis and the spongiosa is prominent (Figure 8a).¹²² Expansion of the spongiosa and plaque formation lead myxomatous leaflets to thicken (2.3 mm versus 0.9 mm in the normal valve; $P < 0.0001$), with increased cell density.¹²² VICs in diseased valves are mostly in an activated myofibroblastlike phenotype characterized by expression of vimentin and smooth muscle α actin, but not smooth muscle myosin heavy chain isoforms 1 or 2, markers of differentiated smooth muscle cells.¹²³ Cells that stain positive for the haematopoietic marker CD45⁺ are also detected and might be fibrocytes, which are engrafted blood-borne cells responsible for fibrosis in several diseases (such as fibrotic lung disease and nephropathy) and can differentiate into myofibroblasts that can both secrete matrix and degrade collagen and elastin.^{122,123} In myxomatous degeneration, expression of proteoglycans and catabolic enzymes is increased, including matrix metalloproteinases (MMP-1, MMP-2, MMP-9, and MMP-13) and cathepsin K, which contributes to fragmentation of collagen and elastin fibres (Figure 8b).^{121–124} A substantial increase in hyaluronic acid content with reduced sulphated glycosaminoglycan content has been reported in degenerated posterior mitral leaflets in comparison with normal valve leaflets in humans.^{125,126} These biological changes alter the mechanical properties of the diseased leaflets, with an increase in circumferential extensibility from 15.5% in healthy valves to 37.2% in myxomatous valves, increase in radial extensibility from 24.1% to 39.7%, and an important reduction in leaflet stiffness and

failure strain.⁷ However, echocardiographic classification of flail versus nonflail valves did not demonstrate differences in the leaflet thickness, length or thicknesses, volume or weight, but measurable differences in leaflet extensibility and basal chordal stiffness.¹²⁷ In that study, flail leaflets had highly extensible leaflets and less stiff basal chordae than nonflail valves.¹²⁷

Surgical presentation

Surgical mitral valve repair involves removal, restraint, or reconstruction of a part or whole valve scallop that is most redundant (Figure 7a). Degenerative processes resulting in mitral regurgitation have two main phenotypes: diffuse myxomatous degeneration (so-called Barlow disease), which can present as a genetic disorder, and fibroelastic deficiency (FED),^{44,117–119,128,129} which might be caused by an accelerated ageing process. Patients with Barlow disease usually present in their fourth or fifth decade for surgery after a long history of murmur and regurgitation; those with FED are typically older with more abrupt onset of mitral regurgitation and associated symptoms owing to chordal rupture and flail.^{44,117,119,120} Although most of the leaflet is thin in FED, localized myxomatous thickening occurs within the typically posterior flail scallop; whether this change is primary or secondary to the regurgitant lesion is unknown.⁴⁴ Other differences between these two expressions of mitral valve degeneration exist (Table 1). Of note, however, the echocardiographic definition of these entities has not been standardized or validated, nor are there known genetic differences or recognized variations in basic mechanisms.

Genetic presentation

Prolapse and mitral valve leaflet thickening occur in several inherited connective tissue disorders, including Marfan syndrome, Ehlers–Danlos syndrome, osteogenesis imperfecta, and pseudo-xanthoma elasticum.¹³⁰ Familial occurrence of MVD without syndromic features was first described >40 years ago.¹³⁰ In addition to the X-linked form of the disease, to date, only three autosomal loci have been reported to be associated with familial MVD: chromosome 16p12.1-p11.2, chromosome 11p15.4, and chromosome 13q31.3-q32.1.^{115,131,132} Identifying genes with mutations in these chromosomal locations will eventually lead to improved understanding of the complex pathogenesis of MVD.⁹² A promoter polymorphism in the matrix metalloproteinase 3 gene (*MMP3*) is a modifying factor that affects the severity MVD.¹³³

The frequency of familial MVD inheritance is unknown, but familial clustering has been proved.¹³⁴ While myxomatous and prolapsing mitral valves often seem to be isolated, familial studies reveal that many affected individuals are not clinically recognized, so heritability might be more frequent than is evident without screening.¹³⁵ The pattern of inheritance is typically autosomal dominant with reduced, age-dependent, penetrance.¹³⁶ The genetic susceptibility of heterogeneous MVD might be complex, with several common susceptibility variants explaining its frequency of 2.4% in the general population and rare, penetrant variants present in families.^{81,137} A genome-wide case–control study is underway, but several loci have been identified in families with Mendelian transmission of MVD, which is consistent with heterogeneous genetic pathways that lead to the myxomatous phenotype. Genome-wide association studies might reveal the interrelated pathways

responsible for normal and disordered valve development that can be targeted to improve disease natural history.¹³⁸

Marfan syndrome, fibrillin-1, and TGF- β —Investigations into the pathogenesis of Marfan syndrome, in particular, have led to improved understanding of genetic contributions to MVD. Marfan syndrome results from heterozygous mutation in *FBN1*, the gene that encodes fibrillin-1, which is the principal component of microfibrils in the ECM.¹³⁹ The occurrence of myxomatous mitral valve thickening and MVP in this disorder has been variably reported as >50%¹⁴⁰ versus 28% with more-recent diagnostic criteria.¹⁴¹ MVD has also been reported to be associated with subdiagnostic variants of Marfan syndrome.^{142,143} Affected individuals often have mitral valve elongation and thickening similar to Barlow disease morphology.^{44,143} Some phenotypic manifestations such as ocular lens dislocation are readily explained by a deficiency of fibrillin-1, a structural protein. Although such a structural deficiency might explain leaflet prolapse itself, the myxomatous leaflet thickening suggests additional changes have occurred in nonstructural pathways.¹⁴⁴

Several lines of evidence indicate that excessive signalling by members of the TGF β superfamily leads to many features of Marfan syndrome.^{6,145,146} Fibrillin1 is similar to the latent TGF β binding proteins (LTBP), and binds to several LTBP isoforms (Figure 9).¹⁴⁷ Mice with a cysteine-substitution mutation in *Fbn1* have longer and thicker mitral valves than wild-type control mice.⁶ Antagonism of TGF β signalling prevents the pathological prolongation and thickening of mitral valves.⁶ TGF β family members can also be inhibited by blocking type1 angiotensin II receptor,^{148,149} a pathway that is also implicated in nonsyndromic myxomatous MVP.¹⁵⁰

Many Marfan syndrome manifestations can be treated by antagonism of TGF β signalling in mouse models, which has led to the investigation of this pathway in human patients with this disease. Loeys–Dietz syndrome, which has phenotypic overlap with Marfan syndrome, leads to aortic aneurysms, dissection, and arterial tortuosity in addition to midline facial abnormalities such as widely spaced eyes (hypertelorism), often with long bone overgrowth.¹⁵¹ This disease is associated with heterozygous mutations in genes encoding the components of the TGF β receptors, *TGFBR1* and *TGFBR2*, or the TGF β 2 ligand, *TGFB2*.^{151,152} Four of 14 (29%) affected individuals with Loeys–Dietz syndrome had substantial mitral valve prolapse.¹⁵¹ In another investigation in which the prevalence of MVP among individuals with *FBN1* mutations was compared with that in individuals with *TGFBR2* mutations, more people with *FBN1* mutation had MVP: 105/232 (45%) in those with *FBN1* mutations versus 15/66 (21%) with *TGFBR2* mutations.¹⁵³ Furthermore, in a report that identified mutations in *MADH3*, which encodes mothers against decapentaplegic homologue 3 (commonly known as Smad3), this gene was associated with a very similar condition to Loeys–Dietz syndrome that was termed aneurysms–osteoarthritis syndrome,¹⁵⁴ in which MVP has been reported in 18 of 36 (50%) patients.^{154,155}

Examination of affected tissues in Marfan, Loeys–Dietz, and aneurysms–osteoarthritis syndromes consistently shows increased TGF β activation and signalling.^{153–156} The loss of TGF β receptor function or its Smad3 signalling factor should have the opposite effect, but paradoxically does not. An attempt to address and understand this paradox arose from

investigation of mice with a fibrillin-1 mutation and haploinsufficiency for Smad4.¹⁵⁷ This combination of mutated genes was expected to improve the phenotype, but the mice actually had more aortic dissection and shorter lifespan than mice with the fibrillin-1 mutation alone.¹⁵⁷ Further investigation led to the discovery that this combination of mutant alleles leads to a unique increase in mitogen-activated protein kinase signalling, most notably the extracellular signal-regulated kinase (ERK), and that selective antagonism of this pathway reverses this effect.^{156,157} Several potential mechanisms have been outlined in another review.¹⁵⁸ One possibility is that mutations in TGF β receptors selectively decrease canonical signalling that leads to a loss of feedback inhibition.¹⁵⁸ Consequently, increased production and activation of the TGF β ligand might lead to excessive noncanonical signalling. Another hypothesis posits that impaired TGF β receptor function leads to upregulation of ligand production and activation.¹⁵⁶ This situation might have disparate effects on different cell types depending on their embryological origin and their relative potential to maintain TGF β signalling. The interaction between different cell types seems to correlate with specific regions of the vasculature and specific tissues that are typically affected in these conditions of increased TGF β signalling, namely Marfan, Loeys–Dietz, and aneurysms–osteoarthritis syndromes.¹⁵⁸

Filamin A mutation syndrome—An X-linked form of myxomatous valvular dystrophy which affects all four cardiac valves was first reported in 1969,¹⁵⁹ and has led to exploration of its molecular pathogenesis. Affected men typically develop dysfunction of the mitral or aortic valves or both, which requires replacement with a prosthetic valve.¹⁶⁰ A large extended pedigree was identified in western France, and the trait mapped along with mild haemophilia A using techniques of genetic genealogy.¹⁶⁰ In this family, the identification of two cousins surgically treated for severe valvulopathy, both also affected by mild haemophilia A, led to familial screening that identified 10 out of the 44 tested men as having progressive MVP, and four with moderate-to-severe aortic regurgitation.¹⁶¹ Among 47 women in the same family, 10 had asymptomatic mitral or aortic valve abnormalities. All individuals with valvular disease had mild haemophilia A (factor VIII activity 15–50%).¹⁶¹ To increase the size of this family for genetic studies, databases were screened to identify patients with mild haemophilia A, surgically treated for valvulopathy, with the same geographical origin as the initial family. This genealogical approach linked two kindreds to a common ancestor born in the 18th century. All family members with MVP had mild haemophilia A, except one individual whose factor VIII activity was normal (>50%), demonstrating that the myxomatous valvulopathy was a separate entity.¹⁶⁰ Finally, an 8 cM interval on chromosome Xq28 was investigated and the association refined to the *FLNA* gene encoding filamin A.^{161,162} Three distinct missense mutations and a 1,944 bp deletion were identified in this gene, each of which is suspected to affect the same antiparallel β -strand organization of filamin A.^{161,162} Notably, individuals with these mutations do not have nodular periventricular heterotopia, a neurological phenotype previously associated with mutations in other regions of the filamin A molecule.^{161–163}

Filamin proteins have an Nterminal actinbinding domain, followed by 24 immunoglobulinlike filamin repeats that fold into antiparallel β sheets that function as interfaces for protein interactions.¹⁶³ Filamin A is an actin-binding protein that stabilizes cortical F-actin networks

and links them to cellular membranes, thereby conferring membrane integrity and protecting cells against mechanical stress.¹⁶³ Filamin A binds to >70 additional cellular proteins, including trans-membrane receptors and signalling molecules. FLNA, therefore, has essential scaffolding functions and integrates multiple cellular behaviours during embryonic development, cellular migration, and mechanical stress responses.^{164–166} N-terminal filamin A mutations causing valvular dystrophy also regulate small Rho-GTPases, so that cells with these mutations are less able to spread and migrate on culture surfaces. Mutated filamin A alters the balance between RhoA and Rac1 GTPases in favour of RhoA, potentially altering the role of Rac1-specific Rho GTPaseactivating protein 24 (also known as FilGAP) and downstream trafficking of β 1 integrins to the cellular membrane, and thus mechanobiology.¹⁶⁷

Complete loss of *FLNA* leads to embryonic lethality with a pleomorphic array of cardiac malformations involving the cardiac ventricles, atria, and outflow tracts as well as widespread aberrant vascular patterning.^{168,169} Abnormal epithelial and endothelial organization and aberrant adherens junctions in developing blood vessels in heart and other tissues support the essential roles for FLNA in intercellular junctions and provide a possible mechanism for the valvular defects.^{168,169} The human disease mutations in *FLNA* also influence many signalling pathways that modulate valve growth, causing excessive TGF β and 5-hydroxytryptamine activation that provides a developmental basis and potential therapeutic opportunities for valve disease.¹⁷⁰

Progressive mitral valve degeneration

While genetic mutations are increasingly shown to underlie the pathogenesis of MVP,^{92,171,172} severe MVP that requires surgical correction often manifests after the fifth decade of life, frequently with a decades-long progressive increase in severity.¹⁷³ Mechanical and biological factors can contribute to disease. Mitral annular flattening has been reported in patients with MVP who develop greater mitral regurgitation compared with those with less regurgitation and normal individuals.¹¹¹ Such annular flattening increases stress on the leaflets and chordae,^{112,174} which can accelerate the degenerative processes initiated by genetic mutations.

In studies of surgically excised human myxomatous tissues, members of the TGF β superfamily are overexpressed, and VICs are transformed into myofibroblasts that increase expression of matrix metalloproteases that can propagate degeneration of collagen and elastin structures.^{122,150,175–177} Changes in these severely affected valves might be a secondary effect, and amendable only to surgical therapy. However, genetic associations between particular genes and MVP have the potential to reveal pathways and modifying factors that can be modulated to limit valve degeneration, even before it is triggered by developmental and signalling defects.¹⁷³ Clues might also be gathered from correlative studies with heritable canine MVP, a cause of severe heart failure and mortality.^{178,179}

Mitral valve in HCM

Phenotype and LVOT obstruction

Echocardiography has enabled the understanding that the mitral valve contributes to subaortic obstruction in patients with HCM via systolic anterior motion (SAM) of the mitral leaflets towards the hypertrophied upper septum. SAM has two main consequences: firstly, flow is accelerated within the narrowed LVOT, creating a pressure gradient measurable by Doppler echocardiography; and secondly, disruption of normal leaflet coaptation, causing mitral regurgitation of variable severity.^{91,180} Subaortic obstruction is present in two-thirds of patients at rest or with exercise, and leads to symptoms of, and predicts progression to, heart failure and sudden cardiac death.^{181,182}

SAM, first described as a diagnostic feature of obstructive HCM by echocardiography, was long thought to be caused simply by mitral valve suction towards the hypertrophied septum owing to accelerated LVOT velocities (known as the Venturi effect).¹⁸³ The importance of structural abnormalities in the mitral valve in generating SAM was recognized after observations that SAM can be present without HCM;⁹ and that SAM begins before LVOT flow acceleration.^{64,184} Furthermore, isolated anterior and inward displacement of the papillary muscles combined with leaflet elongation, especially of the posterior leaflet, can lead to SAM, the degree of which is related to leaflet length, even in the absence of septal hypertrophy.^{15,17,185–192}

SAM results from the combination of septal hypertrophy, creating accelerated, posteriorly-directed left ventricular outflow velocities, with a mitral valve susceptible to SAM because of elongated leaflets that are positioned anteriorly by displaced papillary muscles (Figure 10). The use of 3D echocardiography has confirmed that the mitral valve area is increased *in vivo*, and that LVOT obstruction relates to a combination of upper septal hypertrophy, leaflet elongation, and papillary muscle displacement anteriorly and towards each other.¹⁵ This situation creates a long, slack distal residual leaflet portion interposed into the outflow stream at the beginning of ejection, so that ejection pushes this leaflet portion towards the septum (Figure 11).^{64,184,185} Secondary systolic acceleration as SAM narrows the LVOT can further propagate SAM via the Venturi effect.^{185,188} These findings explain why obstructive SAM can accompany modest septal hypertrophy when excessive or segmental leaflet elongation and papillary muscle malposition is present.^{187,192} They also explain why SAM, subaortic obstruction, and mitral regurgitation persist after septal ablation owing to anterior papillary muscle displacement and leaflet malcoaptation;¹⁹³ and why, for similar degrees of SAM, mitral regurgitation is greater if the posterior leaflet is limited in its capacity to move anteriorly to coapt effectively.¹⁴ These mechanistic insights translate into new therapeutic approaches.^{194–200}

The use of echocardiography has clearly demonstrated that mitral valve abnormalities have an important role in determining LVOT obstruction. In patients with HCM, the mitral apparatus can have a number of diverse structural differences. For example, the mitral valve and papillary muscles can be mispositioned;¹⁸⁵ the leaflets and chordae can be elongated, and the papillary muscles hypertrophied;²⁰¹ papillary muscles can be directly inserted onto the anterior leaflet;²⁰² or there can be accessory mitral valve tissue.²⁰³ Furthermore, the

leaflets might be degenerated with increased leaflet or chordal thickness,¹⁹⁴ ruptured chordae,²⁰⁴ and mitral valve prolapse.²⁰⁵ Investigators using MRI have confirmed leaflet elongation that is independent of left ventricular wall thickness, and which correlates with resting and inducible LVOT gradient.²⁰⁶ The pattern of obstruction can vary with mitral valve structure. For example, elongated leaflets sharply bend at their coaptation point and their distal portions move excessively to contact the septum, whereas shorter, thickened leaflets contact the septum primarily through posterior septal motion.²⁰⁷ Mitral valve anomalies must, therefore, be addressed to achieve comprehensive physiological repair in patients with HCM, and limit SAM after mitral valve repair for MVP when elongated leaflets are interposed into the LVOT.^{9,14,188,208}

Pathology

In anatomical examinations of the mitral valve in 94 highly symptomatic patients with HCM undergoing surgery, 66% of patients had intrinsic valve abnormalities, 36% had increased posterior leaflet scallop number, leaflet thickness, and mass (twice that of healthy patients), and 10% had direct papillary muscle insertion into the leaflet that was not directly linked to hypertrophy or degree of obstruction.²⁰⁹ In a histopathological investigation by Klues and colleagues, no myxomatous changes in the spongiosa of excised mitral valves were seen.²⁰⁹ Conversely, myxomatous mitral valve degeneration was found in case reports of patients with HCM who had massive mitral regurgitation owing to chordal rupture,²⁰⁴ and echocardiographic findings in a surgical cohort showed abnormalities that often resembled degenerative changes.¹⁹⁴

Leaflet elongation

Given the central role of LVOT obstruction via systolic anterior motion of the mitral leaflet, how might mutations that lead to HCM cause mitral valve leaflet elongation? Several theories have been proposed. Firstly, leaflet elongation might be governed by the mutated gene that results in HCM. For example, posterior leaflet length is often increased despite the absence of obstruction in healthy patients who carry a morbid HCM mutation^{15,206,210} and in *Mybpc3* knock-out mice.²¹¹ However, sarcomeric proteins are not present within the mitral valve apparatus, except the papillary muscles. Secondly, leaflet elongation might be stimulated by increased endothelial shear rate in the narrowed LVOT, or is an adaptive process due to repeated episodes of SAM, as mechanical stretch can reactivate embryonic growth pathways.²¹² However, many patients with leaflet enlargement do not have SAM (presumably owing to a lack of anterior malposition into the LVOT flow).²⁰⁹ Thirdly, leaflet elongation might be an independent disorder with tightly linked genetic contributions. However, the chromosomal loci reported to date in MVP do not overlap with genes that are associated with HCM.¹¹⁵ Finally, elongation might be the result of paracrine effects from the abnormal left ventricular wall, which is consistent with evidence that valvulogenesis is influenced by molecular signals generated by the surrounding myocardium as well as the valve.²¹³

Another potential mechanism of leaflet elongation might involve pluripotent epicardial-derived cells (EPDC), a subset of extracardiac mesenchymal cells derived from coelomic mesothelium.^{214,215} The cells might abnormally differentiate into fibroblast-like cells rather

than cardiomyocytes, with consequent increased synthesis of periostin that contributes to the increased myocardial fibrosis observed in patients with HCM.^{214,215} Markedly elevated levels of periostin are indeed expressed in the left ventricular walls of HCM mice,^{216–218} secreted by nonmyocyte elements, and periostin promotes VIC proliferation, differentiation, and matrix production, which might drive leaflet elongation.^{68,72,218} Periostin might also be part of a molecular switch that determines whether a mesodermal progenitor cell becomes a cardiomyocyte or fibroblast,⁷² while blocking expression of the serum response factor transcription factor required for initiating contractile protein synthesis.²¹⁹ Patterns of periostin expression in mouse models of HCM exactly correlate with the hypothesis that EPDC-derived cardiomyocytes become fibroblasts when a gene encoding a contractile protein is mutated.²²⁰ Elevated periostin in close proximity to the mitral valve might increase collagen production and elongate the leaflets, perhaps also driven by reawakened EMT.²¹² These correlated molecular studies are consistent with the theme of a living valve with a developmental abnormality beginning in early life and continuing into adulthood, relating to its interactions with an abnormal ventricle. Improved understanding of the responsible adaptive mechanisms might enable studies targeting prevention and treatment of MVD in HCM.

Ischaemic mitral regurgitation

Definition and mechanisms

2D echocardiography has been used to demonstrate that apically restricted leaflet closure in the ventricle, owing to papillary muscle displacement causing tethering on the mitral leaflets, is the mechanism of ischaemic mitral regurgitation (IMR) in patients with either localized inferior wall ischaemia or global dilatation and failure.^{10,17,221,222} The use of 3D echocardiography in experimental studies has proved that global left ventricular dysfunction alone is insufficient to cause IMR.^{223–225} Displacement and abnormal contraction of the left ventricular wall underlying the papillary muscles, along with decreased shortening of the distance between the papillary muscles, causes mitral leaflet tethering and restricted closure that leads to IMR (Figures 12a,b),^{223–226} which is referred to as a Carpentier type IIIb lesion. Isolated annular dilation, such as occurs in patients with lone atrial fibrillation, also impairs coaptation (Carpentier type I), but relatively mildly unless the leaflets are tethered at both annular and ventricular ends.²²⁷ Tethering geometry also predicts progression of IMR.²²⁸ The posterior location of the papillary muscles explains why IMR is seen predominantly with inferior, as opposed to anterior, MIs unless the anterior infarctions are sufficiently large to cause global dilatation or are associated with anterior dyskinesia causing papillary muscle traction.^{229–233} Investigators using 3D imaging have mapped leaflet tenting relative to papillary muscle displacement by echocardiography and CT, and have shown that asymmetric papillary muscle displacement in the apical direction causes asymmetric leaflet tenting, and is most likely to increase the severity of mitral regurgitation.^{234–237} The substantially increased mortality in patients with IMR is independent of left ventricle size and function, and occurs even in those with mild IMR.^{238–243} IMR also reduces exercise capacity.²⁴⁴ IMR can exacerbate left ventricular remodelling, with adverse changes in the noninfarcted myocardium in correlated cellular and molecular studies;^{245–248} the use of cardiac MRI has also quantified this remodelling.²⁴⁹ IMR is a dynamic disease that varies

with multiple factors, such as with progressive LV remodelling and dysfunction,^{250–253} with varied haemodynamic loading under exercise conditions,^{254,255} reduced tethering under anaesthesia,^{256,257} and vasodilating thermal therapy.²⁵⁸ Dynamic increases in IMR can explain reduced survival in patients with apparently mild resting mitral regurgitation,²⁵⁹ and suggest a need for exercise testing in patients with exertional dyspnoea out of proportion to their resting mitral regurgitation.²⁶⁰

The frequent failure of annular ring reduction relates to persistent leaflet tethering by displaced papillary muscles,^{250,261,262} with continued left ventricular remodelling associated with recurrent mitral regurgitation.^{250,263} The NHLBI-supported trial by the CardioThoracic Surgical Network indicated that 32.6% of the 126 patients who received mitral annuloplasty for severe ischaemic mitral regurgitation had recurrent regurgitation within 1 year, with recurrence predicted by the preoperative degree of tethering.^{264,265} Recurrence of mitral regurgitation predicts absence of the favourable reverse ventricular remodelling that occurs in patients without recurrence.²⁶⁴ Some investigators have shown that stenosis after annuloplasty is caused by subannular tethering and funnel formation by displaced papillary muscles,^{266,267} causing important exercise-induced pressure gradients and reduced survival in more severely affected patients.^{267–269} Subannular repair, therefore, provides the most comprehensive and lasting reduction in mitral regurgitation,²⁷⁰ and the best haemodynamics for forward flow as well, guided by computational analysis of this dynamic, interactive system.^{271–273} Importantly, reducing IMR might also slow progression of left ventricular remodelling, ultimately leading to a decrease in volume- overloaded heart failure.^{274–278} Echocardiographic insights into the tethering mechanism and papillary muscle–ventricular unit have led to potential new interventions to resolve postoperative mitral regurgitation by repositioning the papillary muscles. Such interventions include direct surgical left ventricular reconstruction or using an adjustable external pericardial patch or polymer injected into the subpapillary muscle of the ventricular wall, or surgical papillary muscle approximation.^{10,11,226,237,275,277,279–283} Chordal modification can relieve tethering by cutting the secondary chordae that most restrict leaflet motion or by implanting chords to reduce the papillary muscle-to-annulus tethering distance.^{276,278,284–289} Saddle-shaped annuloplasty can also reduce leaflet stress^{48,112} and improve coaptation.^{47,50} Papillary muscle resynchronization and leaflet augmentation are additional options for the reduction of mitral regurgitation.^{290,291} Patients whose IMR will improve with revascularization of ischaemic, but viable, subpapillary muscle myocardium alone²⁹² are being predicted using advanced cardiac perfusion imaging.²⁹³

New evidence for valve adaptation

Ischaemic mitral regurgitation reflects the insufficiency of valve area relative to the remodelling left ventricle (Figure 12b). This raises the question of whether the valve can adapt to tethering by increasing its area under the influence of the increased stress imposed by tethering,²⁹⁴ particularly at the chordal attachment zone.^{272,295–298} Early mitral valve growth is well described: the annular area increases 20-fold from fetal life to adulthood and the mitral valve grows proportionately.²⁹⁹ EMT is central to this adaptation, and is finely regulated by TGF β , BMP, vascular endothelial growth factor, and Notch signalling (Figure

4), to yield leaflets large enough to maintain effective closure, with an excess of leaflet relative to annular area to adapt to increased volume loads without regurgitation.⁷¹

Testing whether the mitral leaflet area increases in response to tethering requires 3D imaging. Initial clinical studies have confirmed that mitral valve area is 35% larger in patients with left ventricular dysfunction than in healthy individuals, and that larger leaflet area in relation to the demands imposed by leaflet tethering corresponds to less mitral regurgitation.¹² The mitral valve area also increases in an animal model involving MI of the inferior myocardial wall.¹¹

Is this mitral valve enlargement caused by passive stretch or active cell proliferation? Kunzelman and colleagues compared mitral valve leaflets from sheep with an infarct and IMR, sheep with an infarct only, and healthy sheep.^{297,298} The investigators reported a dynamic adaptation to increased leaflet stresses by changes in leaflet thickness and stiffness.^{297,298} To study the mechanisms regulating these adaptive leaflet processes *in vivo*, an animal model was developed to separate the effects of tethering forces, the ischaemic environment, and turbulent mitral regurgitation flow.²¹² The effect of isolated tethering was studied in sheep mitral valves *in situ* over 2 months of papillary muscle retraction. Such tethering leads to increases in both valve area and thickness, with reactivation of EMT in which endothelial cells—normally quiescent—express smooth muscle α actin and enter the valve interstitium to deposit ECM (Figures 13 and 14).²¹²

These findings indicate that the mitral valve is a dynamic cellular environment that actively adapts to superimposed stresses and is influenced by ventricular pathology.^{43,60,71,300} Adaptive leaflet growth has been reported in other studies,^{301–303} as further evidence of the cellular plasticity of mitral valves.³⁰⁴ The compensatory adaptation might restore cellular and biomechanical leaflet homeostasis and ultimately valve function, which leads to the question of why leaflet adaptation is often insufficient to alleviate IMR in many patients. Notably, excised leaflets in patients with end-stage heart failure are thicker and stiffer than healthy valves, potentially impairing leaflet coaptation, which requires systolic leaflet stretch and bending.^{13,296,305} Similarly, in the experimental tethering model described above, preliminary results from adding an apical MI distal to the papillary muscles indicate that the area and thickness of the valve increase in the presence of ischaemia, with exaggerated EMT, interstitial cell activation and matrix turnover. By contrast, in patients with compensated aortic insufficiency, the mitral valve enlarges to compensate for left ventricular remodelling and seems to remain supple, making functional mitral regurgitation infrequent.³⁰⁶ Remodelling of the leaflet, which is beneficial at the early stage after MI, might therefore become counterproductive in the chronic setting after MI. Understanding how the valve adapts over time after MI could create opportunities for physiological interventions, for example, by modulating TGF β , which seems to have a major regulatory role in leaflet homeostasis in IMR as well as Marfan syndrome.^{59,148}

Therapeutics and future directions

The most effective treatment for patients with MVD is surgery, and clinicians recognize that the benefits of mitral valve repair exceed those of valve replacement, at least in the case of

organic mitral regurgitation.³⁰⁷ The remaining clinical debate is focused on timing of surgery to preserve left ventricular contractility.^{94–96,308} Although current surgical repairs for organic mitral regurgitation achieve excellent haemodynamic outcomes and long-term durability, their efficacy on the progressive degeneration or remodelling of the mitral valve leaflets remains unknown. If mechanistic interactions and their role in leaflet degeneration are understood, tailoring repair to achieve optimal haemodynamics and also restore physiological biomechanics should be possible. Such a strategy should also benefit the clinically challenging lesion of IMR, in which progressive leaflet fibrosis and restricted leaflet motion, even after mitral annuloplasty, can reduce the durability of the repair.^{13,250,283,305,309} Success has been achieved in this area to some extent with papillary muscle repositioning or banding, by cutting chordal structures to relieve tethering-induced tension, or by using a saddle-shaped annuloplasty ring to reduce tension in the leaflets and chordae as well as leaflet lengthening.^{226,265,278–280,284–289} However, their effect on chronic valve biomechanics at the ECM and cellular levels remains to be investigated. A ventricular approach to reducing mitral regurgitation can be achieved with vasodilators, which alter the ventricular–arterial impedance relationship, and increase the forward flow. Although this approach follows intuitive reasoning, and medications such as angiotensin-converting-enzyme inhibitors and angiotensin-receptor blockers are beneficial for dilated cardiomyopathy, results are conflicting for the treatment of MVD.^{310,311} Medical therapies that limit left ventricular remodelling and damage to the myocardial substrate from increased adrenergic drive, such as β blockers, might be beneficial in patients with functional mitral regurgitation secondary to left ventricular dilatation, but do not address primary mitral valve dysfunction.^{312,313} Similarly, biventricular pacing in patients with dilated cardiomyopathy and dyssynchrony can improve left ventricular morphology and reduce mitral regurgitation.^{291,314,315}

The exploration of novel therapies might be enabled using valve progenitor cells to recapitulate developmental abnormalities caused by genetic mutations or altered early stresses, complemented with *in vivo* studies using explanted endocardial cushions. Although challenging, the derivation of human embryonic stem cells from endothelial cells that are specified to become valve tissue, and which are competent to perform EMT, might be possible.^{74,316–318} One approach to this challenge is first to derive a multipotent cardiovascular progenitor population of human embryonic stem cells.³¹⁷ Using specific substrates and growth factors, these progenitors can then be differentiated into cells expressing markers of endocardial cushion endothelium and capable of undergoing EMT. Deriving such valve-specified progenitors from induced pluripotent stem cells generated from somatic cells of patients harbouring genetic valve disease might provide invaluable early disease models that predict how mutations can lead to MVD.

Conclusions

Cardiac imaging presents valve biologists with the challenge of explaining observed valve abnormalities on the basis of cellular and molecular signalling processes. The mitral valve remains dynamic into adult life and retains the capacity to reactivate early growth processes. Inherited mutations provide clues to the mechanisms of valve growth and, as more genetic contributions are recognized, novel insights into the milieu of abnormal growth factors,

cytoskeletal proteins, and mechanical strain will inevitably lead to improved therapies. We are learning that valve growth in different conditions is influenced by common factors: signals intrinsic to the valve, paracrine signals from the left ventricular wall, activation of the endothelium to populate the interstitium, and potentially engraftment of blood-borne cells. Reciprocal interactions between the endothelial and interstitial cells of the valve can reveal signals that can be used to promote valve homeostasis.⁴⁵ Current areas of investigation are focused on genetic analysis of the disease, small animal models of MVD, the role of growth factor dysregulation, abnormal EMT, and the influence of biomechanical strain in the pathogenesis of these common disorders. Novel molecular therapies have the potential to target the underlying mechanisms and limit disease progression to reduce the burden of heart failure imposed by the adverse effect of mitral regurgitation on myocardial function.

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Key points

- Mitral valve disease is a major cause of heart failure and mortality
- Even in adult life, the mitral valve is a dynamic structure and therapeutically accessible
- Genetic analysis has revealed that regulation of growth signalling and cell migration pathways could potentially be modified to limit progression from developmental defects to clinically evident valve degeneration, such as mitral valve prolapse
- Mitral valve enlargement is an important determinant of left ventricular outflow tract obstruction in hypertrophic cardiomyopathy and might be stimulated by valvular–ventricular interactions
- Mitral valve plasticity allows adaptation to ventricular remodelling, but adverse processes create relative leaflet deficiency in the ischaemic setting, leading to mitral regurgitation with worse prognosis
- Understanding these concepts can create new opportunities to reduce the clinical progression of mitral valve disease through early detection and biological modification

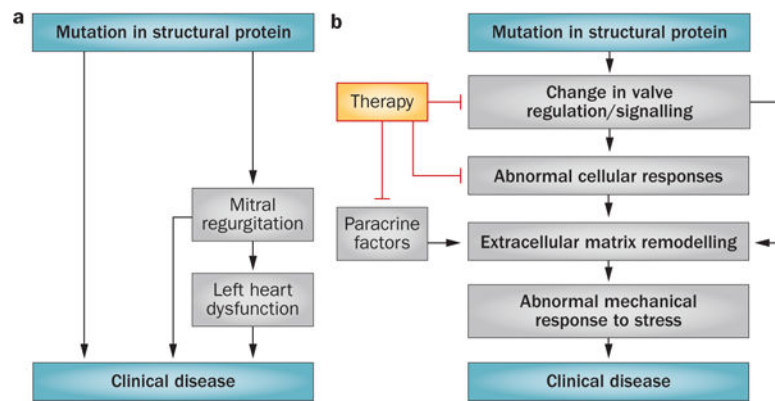


Figure 1.

Models of mitral valve disease. a | Previous conceptual model of mitral valve disease caused by mutations in structural proteins. In this model, the mutations impair valve biomechanical integrity and thereby cause mitral regurgitation and clinical disease. Therapeutic options are limited to surgical valve repair or replacement, as the underlying mechanisms are inaccessible. **b** | Our conceptual model of mitral valve disease caused by mutations in structural proteins. This model is amenable to molecular therapy. In addition to impairing valve biomechanical integrity, mutations in structural proteins can also alter valve regulatory and signalling pathways, leading to abnormal cellular responses and extracellular matrix remodelling. Paracrine factors provide further therapeutic targets.

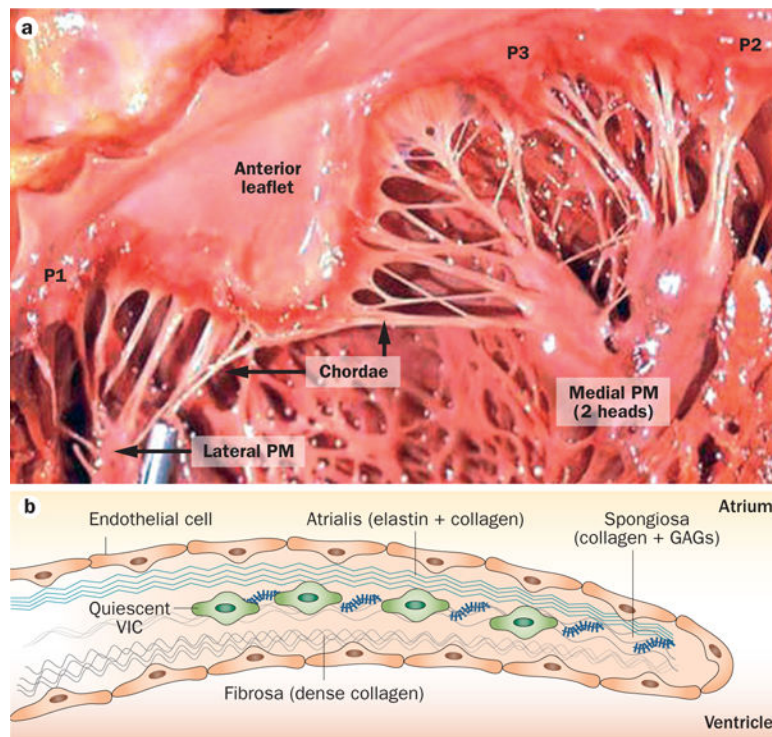


Figure 2.

Mitral valve structure. **a** | The mitral valve has two leaflets—the anterior leaflet below the aortic valve, and the posterior leaflet composed of three scallops: P1, P2, and P3. At the leaflet edges, the chordae tendineae link the leaflets to the PM anchors on the left ventricular wall. **b** | The leaflet structure is well organized, with a dense fibrous layer of collagen, known as the fibrosa, which arises from the mitral annulus and faces the left ventricle. The fibrosa is thicker close to the annulus and thinner at the edge of the leaflet. The fibrosa is covered by the spongiosa, a loose connective tissue layer rich in GAGs. The spongiosa is thicker at the leaflet edge and thinner at the annulus. The atrialis is made up of lamellar collagen and elastin sheets, which extend from the left atrial endocardium into the leaflet. Endothelial cells cover the valve. The deep subendothelial layers contain quiescent VICs—noncontractile, fibroblast-like cells that originate from endocardial endothelial cells. Abbreviations: GAG, glycosaminoglycan; PM, papillary muscle; VIC, valvular interstitial cell.

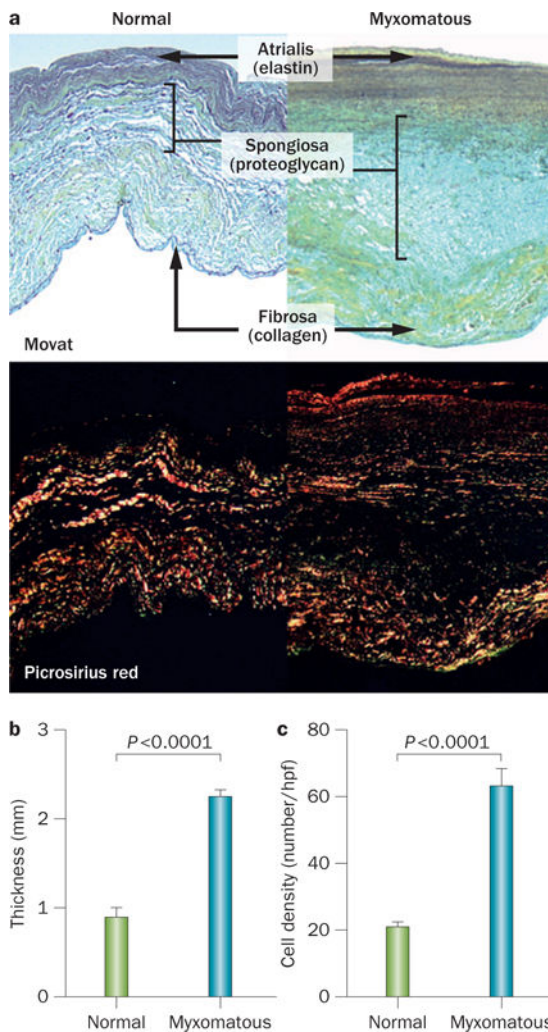


Figure 3.

Morphological features of normal and myxomatous mitral valves. **a** | Normal mitral valves (left) and valves with myxomatous degeneration (right). Myxomatous valves have an abnormal layered architecture: loose collagen in fibrosa, expanded spongiosa strongly positive for proteoglycans, and disrupted elastin in atrialis. Top: Movat pentachrome stain (collagen stains yellow; proteoglycans, blue-green; and elastin, black). Bottom: Picrosirius red staining viewed under polarized light detected disruption and lower birefringence of collagen fibres in myxomatous leaflets. Magnification $\times 100$. **b** | Quantitative analysis of valve thickness, demonstrating thickening of myxomatous valves. **c** | Increased density of interstitial cells in myxomatous spongiosa. Bars indicate SEM. Abbreviation: hpf, high-power field. Reprinted from Rabkin, E. *et al.* Activated interstitial myofibroblasts express catabolic enzymes and mediate matrix remodeling in myxomatous heart valves. *Circulation* **104** (21), 2525–2532 (2001).

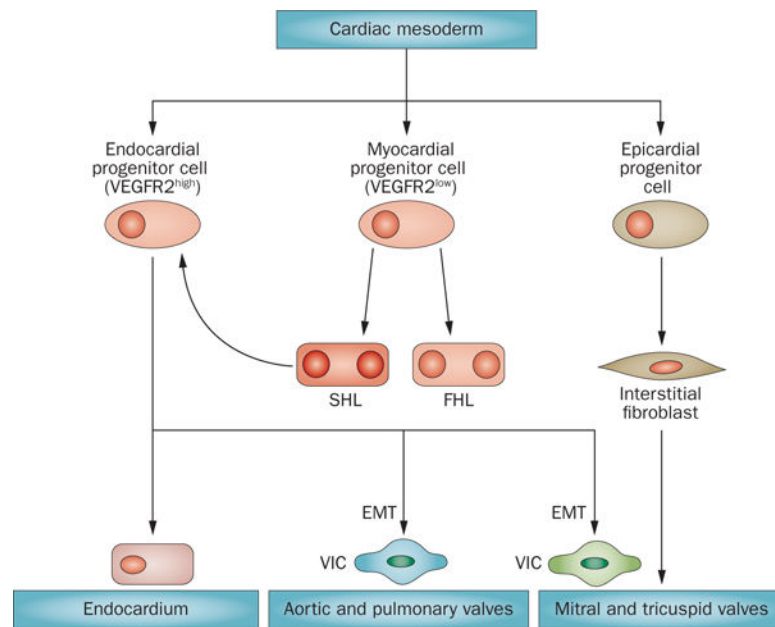


Figure 4.

Cell lineages that contribute to valve formation. The myocardial and endocardial cells are segregated at the onset or just after gastrulation in the E7.5 mouse embryo. This process occurs before, or at the onset of, gastrulation within the cardiac mesoderm. Endocardial progenitor cells emerge highly expressing VEGFR2. These cells undergo EMT to give rise to different cell types that participate in the valve leaflets. Some of these prospective valve cells are also likely to contribute to the endocardium. Myocardial progenitor cells, expressing only low levels of VEGFR2, give rise to both the FHL and SHL. SHL cells increase their expression of VEGFR2 and become endocardial progenitor cells. Epicardial progenitors also give rise to cells found within the valves. Abbreviations: EMT, endothelial-to-mesenchymal transition; FHL, first heart lineage; SHL, second heart lineage; VEGFR2, vascular endothelial growth factor receptor 2; VIC, valve interstitial cell.

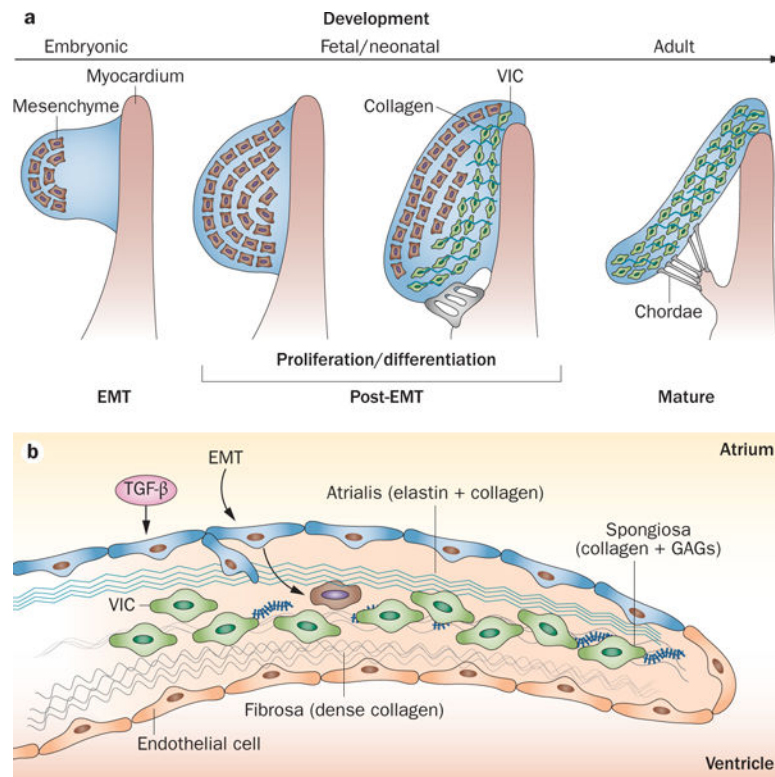


Figure 5.

Mitral valve growth and development. **a** | Mitral valve development from an endocardial cushion. The cells undergo a process of EMT to generate mesenchyme (brown cells) that invades the underlying matrix. These cells undergo proliferative expansion and differentiate into collagen-secreting VICs (green) that become progressively aligned along the proximal–distal tissue axis. Finally, the valve elongates and thins resulting in a mature valve comprised of stratified and aligned cells and matrix. **b** | Valve growth and response to stress. In a growing or stressed valve, or a valve stimulated by TGF- β , endothelial cells undergo EMT, increasing the number of matrix-producing interstitial cells. Abbreviations: EMT, endothelial-to-mesenchymal transition; GAG, glycosaminoglycan; TGF- β , transforming growth factor β ; VIC, valve interstitial cell.

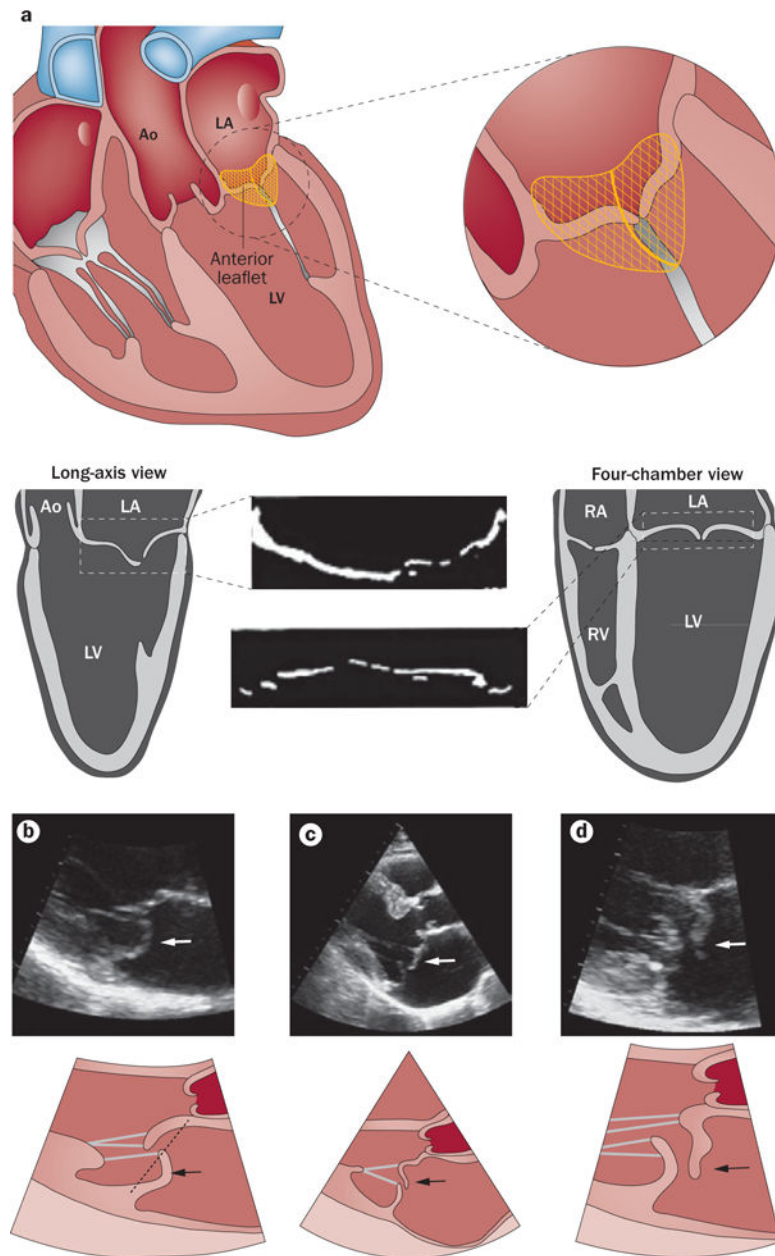


Figure 6.

Echocardiographic diagnosis of mitral valve prolapse. **a** | Diagnosis of mitral valve prolapse must take into account the normal saddle shape of the valve and annulus, which produces opposite leaflet–annular relationships in perpendicular views. Mitral valve prolapse is most specifically diagnosed by leaflet displacement above the annular high points, imaged in long-axis views; and by leaflet misalignment at their point of coaptation. **b** | Parasternal long-axis echocardiographic view of posterior leaflet prolapse (arrows) beyond the annular hinge points (dashed line). **c** | Anterior leaflet prolapse and partial flail (partial eversion of the leaflet tip into the dilated LA; arrows) relative to the posterior leaflet, which is restricted, tethered by the dilated LV.⁹⁰ These opposite leaflet displacements increase the regurgitant gap between the leaflets. **d** | Patient with extensive leaflet thickening and anterior leaflet flail

(arrows). Abbreviations: Ao, aorta; LA, left atrium; LV, left ventricle; RA, right atrium; RV, right ventricle.

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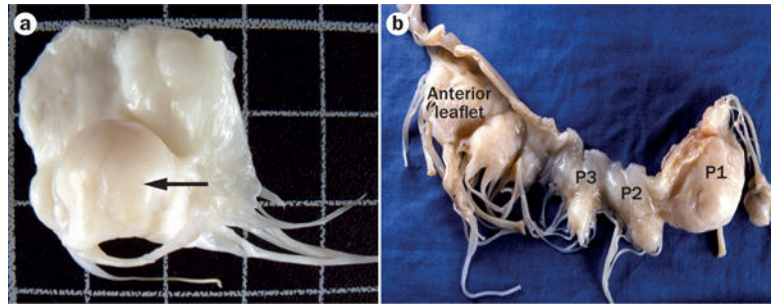


Figure 7.

Anatomy of mitral valve prolapse. **a** | A resected mitral valve with prominent leaflet thickening and opacity and a prolapsing and domed central scallop (P2) of the posterior leaflet (arrow). **b** | Mitral valve removed from a patient with mitral valve prolapse. Note that the lesions, reflected by leaflet bulging towards the observer, are present throughout and most prominent in the medial scallop (P1) of the posterior leaflet.

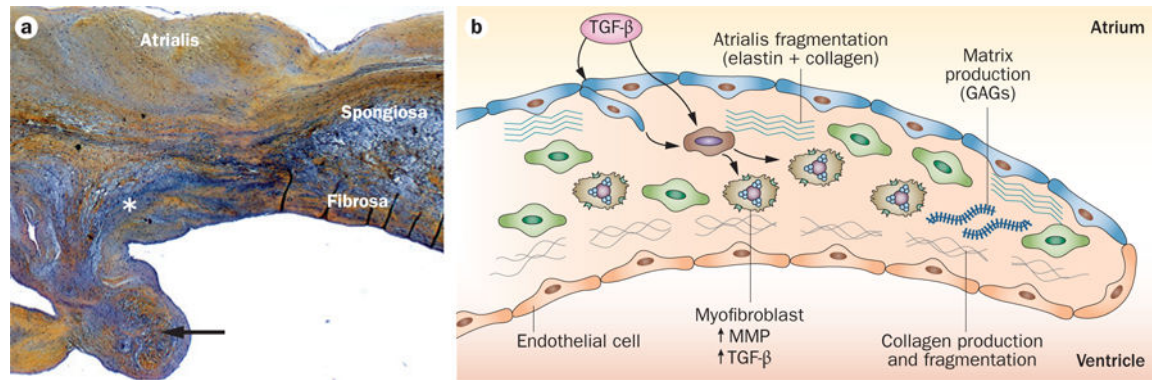


Figure 8.

Mechanisms of mitral valve prolapse. **a** | A mitral valve stained with haematoxylin and eosin to define the lesion of mitral valve prolapse as disruption of the fibrosa by myxoid extracellular matrix (*), which also infiltrates the collagen core of the chordae tendineae, one of which was ruptured (arrow). The elastin lamina beneath the atrialis is also disrupted.

b | A schematic showing the mechanism of myxomatous degeneration, with activation of valve interstitial cells to myofibroblasts that increase matrix production and turnover, secrete MMPs that drive collagen and elastin fragmentation, and release TGF- β that in turn promotes further cell proliferation and myofibroblast differentiation. Abbreviations: GAG, glycosaminoglycan; MMP, matrix metalloprotease; TGF- β , transforming growth factor β .

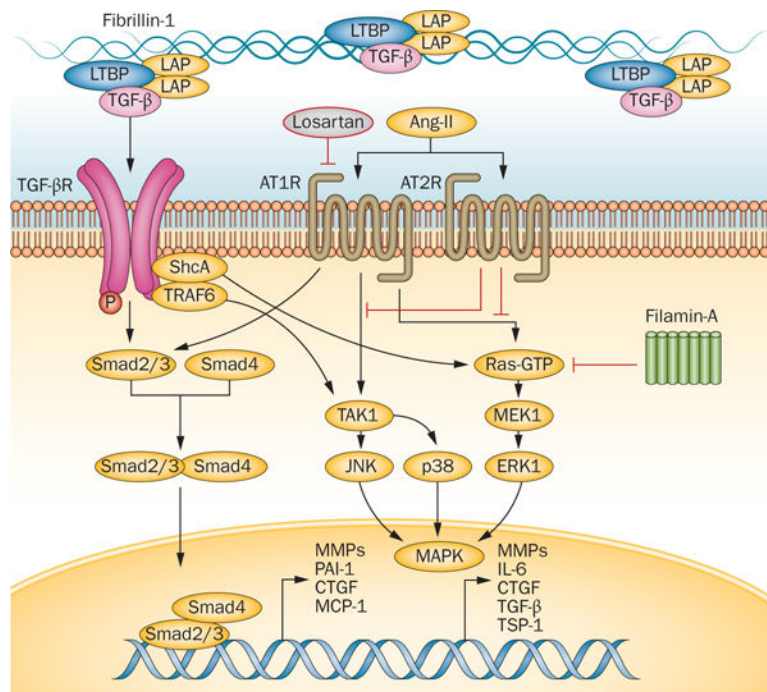


Figure 9.

Proposed mechanism of mitral valve disease in Marfan syndrome. A mutation in fibrillin-1 decreases local binding of the TGF-β LLC, consisting of LAP, LTBP, and TGF-β ligands. TGF-β binds to its receptor (TGFBR1/2), activating canonical (Smad2/3, Smad4) and noncanonical (ERK1, JNK1, MAPK, MEK1, p38, Ras-GTP, ShcA, TAK1, TRAF6) signalling. Activation of these pathways leads to increased nuclear transcription of several downstream products, including CTGF, IL6, MCP1, MMPs, PAI-1, TSP1, and TGF-β itself. Filamin A inhibits activation of ERK1 and MEK1, and its deficiency increases these pathways and might contribute to mitral valve proliferation through this mechanism. Cross-talk between the Ang II receptors (AT1R and AT2R) and TGF-β receptors provides a therapeutic benefit of selective AT1R antagonism for aortic aneurysms in mice with fibrillin deficiency, and this approach is under investigation for treatment of myxomatous mitral valve disease. Abbreviations: Ang-II, angiotensin II; AT1R, angiotensin II type 1 receptor; AT2R, angiotensin II type 2 receptor; CTGF, connective tissue growth factor; ERK1, mitogen-activated protein kinase 3; JNK, mitogen-activated protein kinase 8; LAP, latency-associated peptide; LLC, large latent complex; LTBP, latent TGF-β-binding protein; MAPK, mitogen-activated protein kinase; MCP-1, C-C motif chemokine 2 (also known as monocyte chemoattractant protein 1); MEK1, dual specificity mitogen-activated protein kinase kinase 1; MMP, matrix metalloproteinase; PAI-1, plasminogen activator inhibitor 1; ShcA, SHC-transforming protein 1; TGF-β, transforming growth factor β; TGF-βR, transforming growth factor β receptor; TAK1, TGF-β-activated kinase 1; TRAF6, TNF receptor-associated factor 6; TSP-1, thrombospondin-1.

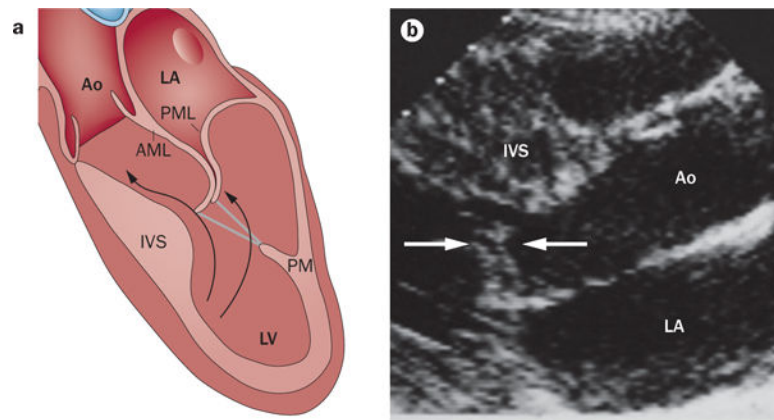


Figure 10.

Mitral valve enlargement in SAM. **a** | SAM relates to a combination of a susceptible mitral valve, which is anteriorly positioned and elongated, encountering a posteriorly shifted left ventricular outflow stream, diverted by hypertrophy of the upper IVS. Anterior PM displacement decreases the posterior restraining force on the leaflets. Leaflet elongation allows greater leaflet excursion to create obstructive SAM. **b** | SAM of elongated leaflets (arrows) approaching the IVS and obstructing left ventricular outflow. Abbreviations: AML, anterior mitral leaflet; Ao, aorta; IVS, interventricular septum; LA, left atrium; LV, left ventricle; PM, papillary muscle; PML, posterior mitral leaflet; SAM, systolic anterior motion.

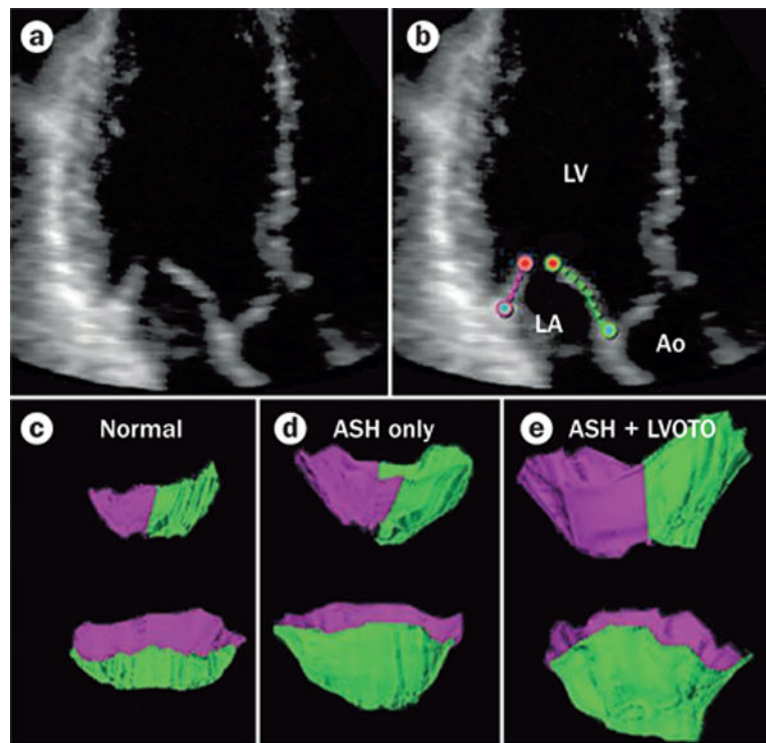


Figure 11.

3D echocardiography showing increase in systolic anterior motion and LVOTO with increased mitral leaflet area. **a,b** | Component view from a healthy control individual with diastolic leaflet traces for 3D reconstruction. **c–e** | Representative open mitral leaflet area measurements in green and purple for anterior and posterior leaflets viewed from the side. Top row: lateral commissure in foreground. Bottom row: left ventricular outflow tract aspect. Smallest values are seen in healthy controls (c), intermediate values with ASH and no resting LVOTO (d), and greatest leaflet areas in patients with ASH and resting LVOTO (e). Abbreviations: Ao, aorta; ASH, asymmetrical septal hypertrophy; LA, left atrium; LV, left ventricle; LVOTO, left ventricular outflow tract obstruction. Reprinted from Kim, D. H. *et al.* *In vivo* measurement of mitral leaflet surface area and subvalvular geometry in patients with asymmetrical septal hypertrophy: insights into the mechanism of outflow tract obstruction. *Circulation* **122** (13), 1298–1307 (2010).

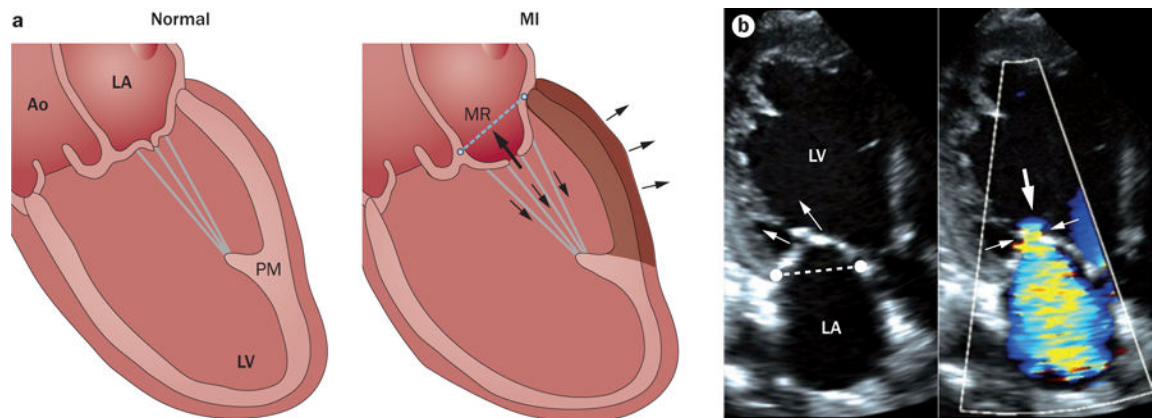


Figure 12.

Mechanism of ischaemic mitral regurgitation. a | Mechanism of ischaemic mitral regurgitation caused by increased mitral leaflet tethering owing to left ventricular remodelling after MI (shaded wall) with outward bulging (three arrows on the outer surface of the heart). Leaflet closure is restricted by increased tethering forces on the leaflets exerted via the chordae (arrows within the heart, exceeding normal on the left). **b** | Echocardiogram from a patient with inferior wall infarction and tethered mitral leaflets (arrows in left panel) with characteristic anterior leaflet bend and concavity towards the LA indicating chordal tethering, mitral regurgitation orifice (small arrows in right panel), and mitral regurgitant flow (large arrow in right panel). Abbreviations: Ao, aorta; LA, left atrium; LV, left ventricle; MI, myocardial infarction; MR, mitral regurgitation; PM, papillary muscle.

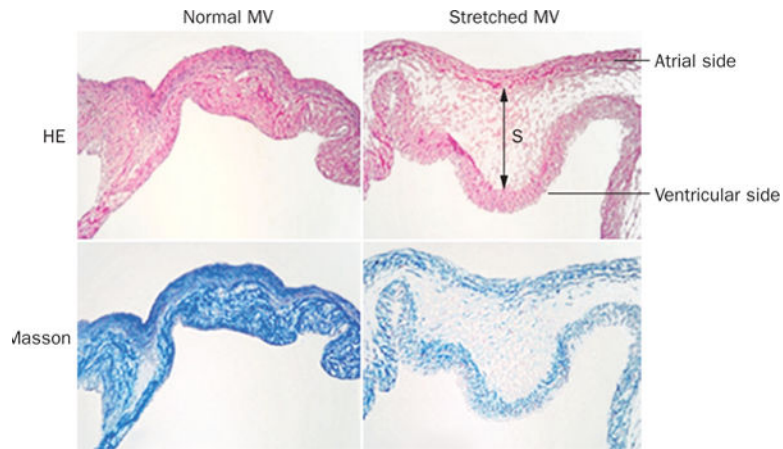


Figure 13.

HE and Masson staining in the normal (left) and stretched (right) MV demonstrating increased leaflet thickness and spongiosa matrix after 2 months of stretching in a sheep model. Blue indicates collagen. Abbreviations: HE, hematoxylin and eosin; MV, mitral valve; S, spongiosa. Reprinted from Dal-Bianco, J. P. *et al.* Active adaptation of the tethered mitral valve: insights into a compensatory mechanism for functional mitral regurgitation. *Circulation* **120** (4), 334–342 (2009).

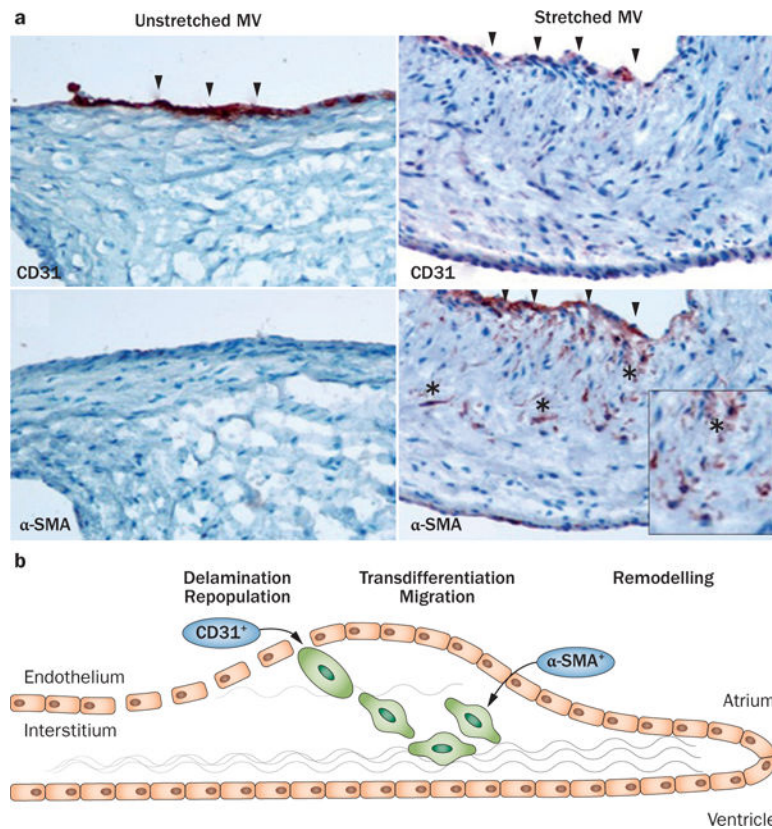


Figure 14.

Evidence for reactivated EMT owing to leaflet stretch. **a** | Staining for the endothelial marker CD31 is reduced in the atrialis, and staining for smooth muscle α-actin is detected, which is normally absent, with positive cells (*) migrating into the interstitium, all features of EMT. Left panels: unstretched MV showing no staining for α-SMA along the CD31⁺ endothelium. Right panels: staining for α-SMA in the atrial endothelium (also CD31⁺) of a stretched MV, with nests of cells positive for α-SMA seeming to penetrate the interstitium (*). Upper panels: arrowheads indicate CD31⁺ endothelial cells. Lower panels: arrowheads indicate α-SMA⁺ endothelial cells, indicative of EMT. **b** | Schematic of active MV adaptation by EMT. Abbreviations: α-SMA, smooth muscle α-actin; EMT, endothelial-to-mesenchymal transition; MV, mitral valve. Reprinted from Dal-Bianco, J. P. *et al.* Active adaptation of the tethered mitral valve: insights into a compensatory mechanism for functional mitral regurgitation. *Circulation* **120** (4), 334–342 (2009). Bottom panel modified from Armstrong, E. J. & Bischoff, J. Heart valve development: endothelial cell signaling and differentiation. *Circ. Res.* **95** (5), 459–470 (2004).

Table 1Echocardiographic differences between so-called Barlow disease and fibroelastic deficiency⁴⁴

Lesion	Barlow disease	Fibroelastic deficiency
Primary effects	Excess valve tissue	No excess valve tissue
	Posterior leaflet height >annulus septal–lateral diameter*	Excess tissue and thickness if present limited to the prolapsing segment
	Anterior leaflet height >38 mm*	Thin leaflets in nonprolapsing segments
	Leaflet thickness >3 mm	No leaflet billowing
	Leaflet billowing	Prolapse often limited to one scallop (usually P2)*
	Prolapse of multiple scallops*	Elongated chordae
	Atrialization of the base of the posterior leaflet* Chordae thickened, elongated, and calcified Chordal rupture infrequent	Chordal rupture*
Secondary effects	Leaflet or annular calcifications*	No calcifications*
	Constant annular dilatation involving preferentially the posterior region	Inconsistent or only moderate annular dilatation

* Inconsistent, but frequent lesions.