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Methodological clues for improving species distribution models in data-poor areas:

example of sub-Antarctic benthic species on the Kerguelen Plateau

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ABSTRACT

Species distribution models (SDM) are essential tools for conservation biologists to evaluate the combined effects of environmental changes and direct human activities on natural habitats and develop relevant conservation plans. However, modeling species distribution in vast and remote regions is often challenging due to poor and heterogeneous datasets and questions the relevance of modeling procedures. In the past few years, there have been many methodological developments in SDM procedures using virtual species and broad datasets but few solutions have been proposed to deal with poor and heterogeneous datasets. In the present work, we address this methodological challenge by studying the performance of different modeling procedures based on four real species, presence-only data compiled from various oceanographic surveys on the Kerguelen Plateau (Southern Ocean). We followed a practical protocol to test for the reliability and the performance of models and to correct for the incompleteness of data, and for spatial and temporal sampling biases. Our results show that producing reliable species distribution models is feasible as long as the number and quality of available data allow testing and correcting for these biases. However, SDM could be

corrected for spatial and temporal heterogeneities in one species only, showing the need to consider all these potential biases when modeling the distribution of species. Finally, we show that model reliability and performance also depend on the interaction between the incompleteness of data and species niches, the distribution of narrow niche species being less sensitive to data gaps than wider niche species.

Keywords:

- Species distribution modeling, model performance, historical datasets, Kerguelen Plateau, presence-only data

INTRODUCTION

- Today, species distribution models (SDM) constitute essential tools for conservation biologists to understand species distribution patterns and their underpinning drivers (see Guillera-Arroita et al. 2015 for a review), assess the combined effects of environmental changes and direct human pressures (i.e. economic activities including tourism) on natural habitats (Gutt et al. 2012), define conservation priorities (Vierod et al. 2014, Greathead et al. 2014), and develop relevant management plans (Reiss et al. 2014, Koubbi et al. 2016). SDM allow to interpolate the known distribution of single species, assemblages or communities (Ferrier and Guisan 2006) to little-accessed or under-sampled areas (Reiss et al. 2011, Robinson et al. 2011) and help improve our knowledge of the distribution of rare species (McCune 2016).
 - In regions subject to fast environmental changes and significant anthropogenic activities, SDM can provide useful tools for conservation purposes (Guisan et al. 2013, Reiss et al. 2014). However, modeling species distribution over vast and remote areas is challenging

and questions the relevance of the method compared to more traditional and qualitative approaches (Koubbi et al. 2016). In such regions, our knowledge of species distribution usually relies on historical and heterogeneous presence-only datasets, which concentrate many gaps and can induce methodological biases altering the level of SDM performance (Loiselle et al. 2008, Costa et al. 2010, Newbold 2010). The use of historical data in SDM has been widely discussed (Reutter et al. 2003, Hortal et al. 2007, 2008), for instance with regards to the spatial and temporal heterogeneities induced by the practice of different sampling strategies. Limitations to SDM performance are mainly due to uncertainties in data location and detection (Costa et al. 2010, Naimi et al. 2014, Tessarolo et al. 2014), to over-estimations of habitat suitability in intensively sampled areas (Guillera-Arroita et al. 2015), and to artefacts in niche descriptions (Hortal et al. 2008). The lack of available data from remote areas also constitutes a limitation to SDM, which are restricted to presence-only data, and are regarded as less reliable and less efficient than presence-absence and abundance-based models (Brotons et al. 2004). Over the past few years, many methodological developments in SDM procedures have been produced to correct for such biases (Dormann 2007, Phillips et al. 2009, Barbet-Massin et al. 2012) but no single procedure emerged (Qiao et al. 2015) and few practical solutions have been proposed to deal with poor and heterogeneous datasets. Our knowledge of Southern Ocean species distribution remains patchy (Koubbi et al. 2016). Therefore, the growing interest of marine biologists and biogeographers for the region has led to the conception of collaborative projects compiling past and present marine biodiversity data in information networks like the SCAR-Marine Biodiversity Information Network (SCAR-MarBIN) (Griffiths et al. 2011), the Biogeographic Atlas of the Southern Ocean (De Broyer et al. 2014) and other open access databases (Danis et al. 2013, Gutt et al.

2013, Van de Putte et al. 2014). However, running species distribution models in the region

still requires a significant effort of data compilation (Guillaumot et al. 2016) to complement

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the existing open access data sources, and check for data quality. In addition, modeling Southern Ocean species distribution poses auxiliary problems due to the paucity of data and model performances that can vary with ecological niche width (Qiao et al. 2015). Recent works have developed methodologies to adapt SDM to rare species and poorly-sampled areas but none was tested for the Southern Ocean (Pokharel et al. 2016, Phillips et al. 2017).

In this work, we analysed the reliability of modeling procedures with regards to the heterogeneous nature of data available and the gaps in our knowledge of species distribution. We compiled echinoid presence-only data collected from several ancient and recent oceanographic campaigns carried out on the Kerguelen Plateau (sub-Antarctic region) for one and a half century. The distribution of four echinoid species with contrasting ecological niches was modeled, and the reliability and the performance of modeling procedures were tested. We propose methodological clues to correct for spatial and temporal biases and assess the sensitivity of modeling procedures to species ecological niche width. This is the first methodological approach to correct for potential biases in SDM in the Southern Ocean. Our objective is to offer useful perspectives for future modeling works along with a practical and transferable protocol to test for the reliability and performance of modeling procedures.

MATERIAL AND METHODS

Biological data

Species occurrence data were taken from Guillaumot et al. (2016) and Pierrat et al. (2012). The dataset includes presence-only data of echinoid species collected during scientific cruises carried out on the Kerguelen Plateau (63°/81°E; -46°/-56°S) since 1872 (Fig. 1). Scientific objectives, dates, sampling efforts, gears, and surveyed areas have differed between cruises, leading to spatial and temporal heterogeneities (Guillaumot et al. 2016). In the dataset, four

echinoid species with contrasting ecological preferences and a high number of presence-only records were selected. Species include two sediment feeders of the family Schizasteridae, one shallow water species, *Abatus cordatus*, and a deeper one, *Brisaster antarcticus*, one carnivorous/detritivorous and eurybathic species of the family Cidaridae, *Ctenocidaris nutrix*, and one omnivorous and eurybathic species of Echinidae, *Sterechinus diadema* (David et al. 2005) (Fig. 1). *A. cordatus* is a coastal species that is endemic to the Kerguelen Plateau, *B. antarcticus* is known in the Kerguelen and Crozet archipelagoes and has broader environmental preferences than *A. cordatus* (Fig. 1). *C. nutrix* and *S. diadema* are widespread in the Southern Ocean and have contrasting environmental preferences (Fig. 1).

Environmental descriptors

Environmental descriptors were taken from Guillaumot et al. (2016). The dataset covers the geographic extent of the Kerguelen Plateau (63°/81°E and -46°/-56°S) and compiles environmental data for six decades included in [1955-2012]. Environmental data are available at a grid cell resolution of 10km precision. Environmental layers include no data pixels, particularly in seafloor related descriptors. Data were not interpolated to avoid the potential biases due to interpolation procedures.

Collinearity between descriptors can alter modeling performances (Phillips et al. 2006) because collinear data may (1) inflate standard errors, (2) induce the violation of residual independency during model validation and (3) generate noise that can be interpreted as a link between descriptors (Dormann et al. 2013). To reduce the collinearity effect, we computed the Variance Inflation Factor (VIF) and the Spearman correlation coefficient (rs) between all available descriptors from Guillaumot et al. (2016). VIF analysis was performed using a stepwise procedure, using the *vifstep* function proposed in the 'usdm' R package (Naimi et al. 2014). Descriptor pairs with high VIF and rs values were omitted based on the commonly

used thresholds of VIF < 5 and rs < 0.85 (Pierrat et al. 2012, Dormann et al. 2013, Duque-Lazo et al. 2016). Environmental descriptors finally selected to model species distribution are displayed in Table 1. Environmental changes were tested between 1955 and 2012. The comparison of pixel values

Environmental changes were tested between 1955 and 2012. The comparison of pixel values between periods was generated using a Wilcoxon signed-rank test with the Bonferroni correction.

Analytical procedures

The flow chart of Figure 2 details the analytical procedure used in the present work.

Model selection

Due to the growing interest of ecologists for species distribution modeling, a large range of modeling techniques is now available (Reiss et al. 2011, Guillera-Arroita et al. 2015, Qiao et al. 2015). Running the most appropriate model involves selecting the best modeling technique for the data under analysis and also involves considering the scientific objectives to be addressed (Reiss et al. 2011, Qiao et al. 2015).

Here we compared several modeling techniques using the 'biomod2' R3.3.0 library (Thuiller et al. 2016) and we tested the performance of these approaches with regards to the chronological addition of new data and to the transferability of models between areas. Several models were generated with an increasing number of occurrence data (Fig. S1). The best modeling techniques were then compared with each other using a non-random cross-validation procedure (Fig. S2, Wengen and Olden 2012) in order to determine the approach

154 with the best accuracy in transferability performances (Randin et al. 2006, Wengen and Olden 155 2012). 156 Results show high performance and stability values for Random Forest (RF) and Boosted 157 Regression Trees (BRT) in our case study (Appendix 1). However, BRT performed better in 158 transferability in comparison with RF (Heikkinen et al. 2012), and previous works showed 159 that RF does not deal correctly with missing values and patchy datasets (Breiman 2001, 160 Barbet-Massin et al. 2012, Qiao et al. 2015, see Table S1 for a review). Therefore, BRT was 161 chosen in the present work to generate the analyses. 162 BRT calibration was realised using the 'gbm' R package (Ridgeway 2015, Elith et al. 2008). 163 The three main parameters (learning rate lr, tree complexity to and bag fraction bg) were 164 selected using the method developed by Elith et al. (2008) to figure out the combination of 165 values minimizing the predicted deviance of the models (Elith and Leathwick 2014). The 166 parameters were finally set at respectively lr= 0.0001, tc=2 and bf= 0.75. 167 Following Barbet-Massin et al. (2012), we sampled the same number of background data as 168 the number of presence data available for computing BRT models. Considering the low 169 number of presence data available, 100 model replicates (i.e. background sampling) were 170 generated for each analysis. Finally, to correct for data aggregation in space, presence 171 duplicates were removed when present in a same 10km resolution pixel. 172 173 Model performance was assessed by measuring AUC score values (Area Under the Curve of 174 the Receiver Operating Curve) of each model replicate using the 'dismo' R library (Hijmans 175 et al. 2016). AUC expresses the relationship between model sensitivity and the commission 176 error (1-specificity). The sensitivity corresponds to the number of « presence » pixels 177 correctly predicted as present and the specificity the number of « absence » pixels correctly 178 predicted as absent (Fielding and Bell 1997). The use of the AUC to evaluate SDM

performance has been repeatedly discussed (Lobo et al. 2007, Peterson et al. 2008) but the AUC remains the most appropriate metric for presence-background models as values stay stable with low-prevalence datasets and are not sensitive to threshold effects (Hand 2009, Proosdij et al. 2016). Following the recommendation of Jimenez-Valverde et al. (2012), we used the AUC to estimate the robustness of models but not for direct comparisons between models that were generated for different species, on different studied areas and with different training samples.

Correcting for sampling bias

Data collected during the various scientific cruises led over the Kerguelen Plateau for the last 145 years present conspicuous spatial heterogeneities. The resulting bias can generate an unequal number of records in the different sectors of the study area and heterogeneous patterns in record distribution. Such heterogeneities might increase the risk of over-estimating the contribution of environmental conditions to the models in the most sampled areas (Araújo and Guisan 2006).

The effect of spatial heterogeneities on the quality of distribution models was tested using a null model approach. A first null model, null model #1, was generated by sampling presence data at random within the total set of sites that were visited during the different campaigns, whether echinoid specimens were collected at these sites or not (Fig. 3). Because absence data are not available, this approach allows us to assess the weight of sampling bias in the models. If a sampling bias is significant, null model #1 is expected to produce distribution maps with higher suitability values in the most sampled areas (Merckx et al. 2011).

A second null model, null model #2, was built by simulating presence data sampled at random over the entire studied area. Null model #2 is expected to produce distribution maps of equal suitability over the entire study area. If sampling is spatially biased, we expect that null model #1 deviates from null model #2 (Raes and ter Steege 2007).

The two null models were generated for the four selected species. The number of presence-only data used in the models was contained between the number of data collected until the MD04 campaign and until the PROTEKER campaign, between 1974 and 2015, which corresponds to periods of high sampling effort (Fig. 1). In each null model, 100 replicates were produced. Time-averaged environmental descriptors [1955-2012] were used for the analysis.

To correct for sampling bias when null models #1 and #2 significantly differ between each other, we used the methodology proposed by Phillips et al. (2009), which has been shown to improve modeling performances (Phillips et al. 2009, Aguirre-Gutiérrez et al. 2013). A grid layer was built using a kernel density estimation (KDE) to represent sampling spatial bias. The layer was calculated from the map of visited sites. The estimated proportion of presence-only data present in each pixel was determined using the 'kde2d' function of 'MASS' R package (Venables and Ripley 2002). Background data were sampled according to the weighting scheme of the KDE layer, to reduce discrepancies between presence-only records and background data (Phillips et al. 2009, Barbet-Massin et al. 2012). In order to test for the efficiency of model correction based on the KDE, the Pearson r correlation was computed between pixel values of the KDE layer (a proxy for the sampling effort) and the predicted probabilities of models, before and after the KDE correction.

Spatial heterogeneities in data collection can also generate spatial autocorrelation (SAC) between presence records, which can violate model calibration assumptions, and affect model accuracy with wrong parameter estimations (Segurado et al. 2006, Dormann 2007, Crase et al. 2012). Several approaches have been developed to take into account SAC in SDM (Crase et al. 2012 for a review). They consist in including an additional term in models (the autocovariate), which represents the influence of neighboring records on modeling predictions. The significance of SAC was tested using the Moran I autocorrelation index computed on model residuals (Luoto et al. 2005, Crase et al. 2012) for both original and corrected models. Models were built using time-averaged environmental descriptors [1955-2012].

Testing for the effect of the chronological addition of new records on model

performance

Our dataset compiles presence-only data collected during various scientific cruises and with distinct sampling protocols, which may alter the performance of the models (Fig. 1). To test for model reliability, we separately analysed (1) the influence of the chronological addition of presence records, (2) the influence of data number alone and (3) the influence of sampling patterns (distribution of data in space). The analyses were performed for *A. cordatus*, *C. nutrix* and *S. diadema*. Not enough data were available for *B. antarcticus*. We used time-averaged environmental descriptors [1955-2012] to generate the models.

To test for the potential effect of the chronological addition of new data on model performance, we followed the protocol proposed by Aguiar et al. (2015). The dataset was split into distinct subsets corresponding to main periods of sampling effort (1975: including Marion Dufresne campaigns, 1993: including ANARE campaigns, 2010: including POKER II campaign, 2015: including PROTEKER campaigns). New presence data were progressively

added to the models, following the chronological collection of new records. The influence of
the chronological addition of data was assessed by measuring the correlation between models
using the Schoener's D statistic. The Schoener's D is a correlation metric adapted to the study
of niche similarities (Warren et al. 2008, Rödder and Engler 2011). It evaluates the similarity
of pixel values between two distribution grids. A D value of 0 means that the two maps are
perfectly different, a D value of 1 means that maps are perfectly similar. Values were
computed using the <i>niche.overlap</i> function of the 'ENMeval' R package (Muscarella et al.
2014).
The significance of correlations was tested following a null model protocol, using 100
replicates, pairwise-compared using the Schoener's D statistic (Raes and ter Steege 2007,
Warren et al. 2008, Ficetola et al. 2009).
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Testing for the effect of temporal variations on model performance

To test for the effect of environmental shifts on models, different distribution models were generated using distinct environmental descriptors for four periods ([1955-1964]; [1965-1974]; [1975-1994]; [2005-2012]) and the complete set of presence data available. Similarities between models were measured using the Schoener's D statistic.

RESULTS

Environmental shifts

Mean sea surface temperature and amplitude, mean seafloor temperature and amplitude, mean sea surface salinity and amplitude were all tested significantly different between all the studied decades (p < 0.001). Seafloor temperature amplitude only was not proved significantly different between decades [2005-2012] and [1955-1964]. These results indicate that significant environmental shifts occurred during the studied time period and may induce important variations in models as the dataset extends over 145 years.

Spatial bias

Null model #1 predicts higher suitability values in areas with most intense sampling effort corresponding to the northern part of the Kerguelen Plateau and the vicinity of the Kerguelen archipelago (Fig. 3A). In contrast, null model #2 predicts equally medium suitability values over the entire Kerguelen Plateau because presence data were sampled at random in the area (Fig. 3B). The difference between null models #1 and #2 was tested significant for the four species (Fig. 3) showing that sampling bias has a significant impact on model outputs, which will over estimate environment suitability in areas with the highest number of sampling sites if no correction is applied.

Correlation between visited areas and predicted probability distribution decreases in models built with the KDE-correction compared to non-corrected models (Table 2), showing that the

- 302 correction is efficient to reduce the influence of sampling bias on modeling performances.
- 303 However, the correction was proved less efficient in models of the coastal and narrow niche
- 304 species A. cordatus for which correlation values after the KDE correction remain high
- 305 (r=0.44) (Table 2).
- 306 Spatial autocorrelation (SAC) was tested significant for non-corrected models (Moran index,
- 307 I_{min}=0.05, I_{max}=0.16) but values were not significant in corrected models (I_{min}=0.04,
- 308 I_{max}=0.06), except for A. cordatus (Table S2, Figure S3). This shows that the KDE procedure
- also corrected for SAC for three of the four studied species.

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Chronological addition of new records

- The different models built with a chronological addition of new data show high AUC values
- 313 for C. nutrix and A. cordatus $(0.814\pm0.018 < AUC_{C.nutrix} < 0.883\pm0.024$ and $0.908\pm0.023 <$
- 314 AUC_{A.cordatus} $< 0.909\pm0.018$ respectively) demonstrating the relevance of all models (Fig. 4,
- 315 Fig. S4). For these two species, Schoener's D correlation values are high $(\overline{D}_{A.cordatus}=$
- 316 0.978 \pm 0.023, $\overline{D}_{C,nutrix}$ =0.968 \pm 0.020) and were tested significant, showing that models are
- 317 similar to each other.
- 318 In contrast, models generated for S. diadema significantly differ between each other with
- lower Schoener's D statistics ($\overline{D}_{S.diadema} = 0.932 \pm 0.036$) (Fig. S5). Therefore, the
- 320 chronological addition of new data has contrasting impacts on model outputs according to the
- 321 studied species, which may be explained by a various sensitivity of models to data addition
- and to sampling patterns.

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Data addition and sampling patterns

- 325 Comparison of models produced with an increasing number of data presents high and
- significant Schoener's D values (minimum=0.979 \pm 0.031 for S. diadema, maximum=0.985 \pm

0.020 for *C. nutrix*), showing that model outputs do not vary significantly with increasing data in our case study (Table 3).

To test for the influence of sampling patterns, models built using subsets with contrasting distribution patterns (radial versus random patterns) were compared. Schoener's D statistics measured between these two types of models present low values. This suggests a significant influence of sampling patterns on model outputs (Table 3).

Environmental change and model performance

The different models generated with contrasting environmental descriptors are highly similar as shown by high Schoener's D and low standard deviation values (\overline{D} = 0.981±0.005). This proves that environmental shifts have no significant impact on model outputs. In addition, the respective contributions of environmental descriptors to models do not vary significantly between periods for the four species. However, *A. cordatus* seems to be less impacted by environmental shifts in comparison with other species (Fig. 5).

Finally, the contribution of time-averaged environmental descriptors over the total studied period [1955-2012] tends to differ from contributions computed for each decade separately (Fig. 5).

Final species distribution models

Sampling bias analyses and model corrections show that reliable distribution models can be built for *C. nutrix* only. It is the only dataset in which spatial and temporal heterogeneities do not impact prediction performances significantly. A final, reliable model was produced for *C. nutrix* over the Kerguelen Plateau (Fig. 6).

DISCUSSION

Data scarcity and heterogeneity

First research surveys of the Kerguelen Plateau date back to the oceanographic campaign of the HMS Challenger in 1872. One and a half century later, our knowledge of benthic species distribution on the Kerguelen Plateau has significantly increased but remains patchy (Koubbi et al. 2016). As in most parts of the Southern Ocean, modeling species distribution on the Kerguelen Plateau faces significant limitations due to data gaps and heterogeneities (Guillaumot et al. 2016). Such limitations seriously question the relevance of modeling procedures, which are required by environmental managers for conservation purposes (Féral et al. 2016, Koubbi et al. 2016). In the present work, we follow a step by step protocol to assess, quantify, and correct the potential effects of data scarcity and heterogeneity on models, a critical issue when considering the growing interest for modeling approaches in Antarctic and Sub-Antarctic regions (Gutt et al. 2012). Our results demonstrate that such approaches can prove feasible and reliable in certain case studies, when data quality and sampling bias can be tested and corrected.

Coping with spatial and temporal bias in presence-only datasets

Spatial bias and spatial autocorrelation (SAC)

Building SDM for remote and little-accessed regions often implies the use of spatially biased datasets conditioned by sampling caveats. Because parts of these regions that are the most easily accessed aggregate most of the available presence data, more weight is given to the most sampled sites and model performance is reduced (Phillips et al. 2009). In the present work, a significant difference was measured between the two null models (that were generated by selecting presence data either from visited stations only or at random over the

total investigated area), highlighting the strong heterogeneity of sampling effort with more data collected in the northern part of the Kerguelen Plateau and in coastal, shallow areas.

The significant spatial autocorrelation (SAC) values that were computed from model residuals also reveal the impact of sampling bias. The significance of SAC on uncorrected model residuals can be partly explained by the relative accumulation and high density of presence data in shallow areas of the Kerguelen Plateau where species presence probability is overpredicted. One could argue that SAC analysis does not apply to SDM as species presence proximities must be considered in the environmental niche space, not in the geography. However, in the present study, the difference between null models constitutes an operational evidence of the impact of sample clumping on model outputs, which is also revealed by significant SAC values.

To correct for sampling bias, we used a background-based correction method (Phillips et al. 2009) that was already used in former studies based on presence-only and limited datasets (Mateo et al. 2010, Pokharel et al. 2016, Phillips et al. 2017). These methods allow to reduce the effect of sample spatial bias on modeling performance by weighting background records according to sampling patterns. In the present study, the correction was proved efficient to correct both for the influence of the uneven sampling effort on predicted distributions (Table 2) and for SAC on all SDM except for models of *A. cordatus*. *A. cordatus* is a coastal, shallow marine species that was mainly sampled in the northern part of the Kerguelen Plateau. Species presence records are strongly conditioned by the location of most important sampling efforts. This is in line with previous studies that already highlighted the difficulties of modeling the distribution of narrow niche species with low prevalence distribution (*i.e.*, corresponding to the proportion of the area where presence records are located) (Barbet Massin et al. 2012, Qiao et al. 2015). In small presence-only datasets, the methodologies used to correct for spatial bias are not as efficient for narrow niche species as

for broader niche species. Reducing the extent of distribution modeling of narrow niche species to the boundaries of their environmental limits could prove a good alternative.

Influence of record addition

The chronological addition of new data has a limited impact on certain model outputs as demonstrated by high similarities between the chronological models generated for *A. cordatus* and *C. nutrix*. In contrast, chronological models of *S. diadema* significantly differ between each other. The detailed analysis of data increment proved that the increasing number of presences has no impact on modeling performance, which is not in line with previous works (Stockwell and Peterson 2002, Wisz et al. 2008). However, these results can be altered by our incomplete knowledge of species full distribution due to the limited number of data available and to sampling bias (Hernandez et al. 2006, Bean et al. 2012). In models of *S. diadema*, differences between the chronological models are due to contrasted spatial patterns between datasets (transect versus random patterns).

Historical data and environmental change

Significant environmental shifts were measured for the descriptors analysed between 1955 and 2012 over the Kerguelen Plateau (*i.e.* mean sea surface temperature and amplitude, mean surface salinity and amplitude). However, for all species, distribution models built for each decade are highly similar between each other. These results confirm that temporal heterogeneities in datasets do not necessarily impact the robustness of models, because species preferences for their environment may be wider than the magnitude of changes in time. Working with both present and historical data to improve the completeness of occurrence records proved reliable when assuming that species niche and distribution have not significantly changed during the studied time period.

Between the five decades, the respective contributions of temperature and salinity to the models did not vary over the range of within-decade variation for *B. antarcticus*, *C. nutrix* and *S. diadema*. Variations between decades are more marked in models produced for *A. cordatus*. This near-shore species is found in shallow waters of Kerguelen and Heard islands, where environmental descriptors include many no data pixels (Guillaumot et al. 2016). Consequently, the varying contributions of temperature and salinity to the models of *A. cordatus* between decades cannot be attributed with certainty to the effect of environmental change but to modeling limitations.

Sea surface temperature and salinity amplitudes have significant contributions to the models, contributing more than averaged parameters (i.e. *A. cordatus* and *B. antarcticus*, Fig. 5). This is in line with the results of Bradie and Leung (2016) who tested for the contribution of several environmental descriptors on a wide panel of taxa. They showed the importance of including seasonal means and extremes in models to further depict species distribution, considering their stronger relationships with species niche width and ecological traits (*i.e.* growth and survival, see Franklin 2009).

Using time-averaged descriptors over the entire period [1955-2012] could have been considered the best approach to produce representative models, independent from short-term environmental variations. Unexpectedly, our results show that for all species, contributions of time-averaged descriptors to the models are much more different than all differences between decadal descriptors (Fig. 5). This shows that using time-averaged descriptors for long time periods does not necessarily improve model reliability in comparison with using descriptors averaged for a shorter time period. This also shows the importance of the descriptor selection

in modeling procedures, a critical issue for improving model performance as already stressed in previous studies (Bradie and Leung 2016). This is particularly relevant for certain regions of the Southern Ocean like the Western Antarctic Peninsula that has experienced among the most significant environmental changes in the world ocean during the last decades (Turner 2015).

Influence of species niche width in modeling performances

Among the four studied species, *A. cordatus* has the narrowest ecological niche and most restricted distribution in the vicinity of coastal areas of the Kerguelen and Heard archipelagoes. Such limited geographic and environmental distributions compared to the total extent of the studied area implies that similar environmental conditions prevail in geographically close occurrence sites. This induces a strong SAC pattern that explains the difficulties to correct for spatial bias in comparison to other species models. Moreover, the limited environmental variability between coastal sampling sites of the different oceanographic surveys can also explain the absence of data addition effect on modeling performances for *A. cordatus*.

In contrast, *C. nutrix* and *S. diadema* have wider ecological niches than *A. cordatus* (Fig.1). For these two species, record data are more widely distributed and show contrasting sampling patterns (*i.e.* transect-like versus random patterns) that have been shown to influence modeling performance in *S. diadema* only (Table 3). This can be explained by the higher number of presence records available for *C. nutrix* (n=114 and n=98 for *C. nutrix* and *S. diadema* respectively) that allowed a more complete survey of *C. nutrix* distribution. Finally, *C. nutrix* dataset only presents a quality and number of occurrence records that fulfill all methodological requirements to produce reliable distribution models.

Considering species niche width to cope with spatial and temporal bias in SDM is important, as already shown by Tessarolo et al. (2014) who studied the influence of survey designs on the performance of distribution models for endemic species with narrow ecological niches. They concluded that survey designs have a low impact on models in comparison with the effect of niche width, data number, and type of modeling technique used. However, they did not generate any analysis of species with broad ecological niche as a comparison. Our results are also in line with other modeling studies in which distribution models of species with broad niche were the least stable (Reiss et al. 2011, Qiao et al. 2015, Guo et al. 2015, Ranc et al. 2016).

Conclusion

The use of SDMs has gained in importance during the last decades. They can provide complementary information for environmental managers. Modeling results can help interpolate species distribution, identify the potential drivers of species distribution and predict the potential effects of environmental changes on habitat suitability. However, modeling species distribution over vast and remote marine areas like the Southern Ocean using poor and heterogeneous datasets remains challenging and improvement of biological and environmental datasets is still required.

In the present study, we showed that reliable species distribution models can be produced in such areas as long as the number and quality of data allow testing and correcting for the effects of biases. Using historical data requires proper environmental descriptors for modeling the effect of environmental changes on species distribution. Using time-averaged predictors over long time periods can generate unrealistic models.

Model selection is also crucial at this stage and the statistical performance of models is not the only criteria to be considered. Modeling procedures must be chosen with regards to the scientific issues that are addressed. Two procedures (BRT and RF) performed best in our case study, but one of them (BRT) was proved to be more relevant because it better dealt with transferability and data patchiness.

Modeling species distribution in data-poor areas poses the practical problem of the minimum number of presence-only data required to run reliable models, although this is not the only and most critical issue. Beforehand, the number of occurrence records must be high enough for testing model robustness and reliability. In regions with limited access, sampling effort may be heterogeneous, which influences model performance. We showed that sampling bias can be corrected, but the efficiency of correction depends on species niche width, narrow niche species models being more troublesome to correct. In our study, *A. cordatus* is a species limited to coastal shallow areas, which implies a strong correlation between species occurrence and sampling patterns. Restricting the model to a more reduced area could allow to correct for spatial bias and improve modeling performances.

There is also a crucial need for improving the quality of datasets (Kennicutt et al. 2014) and running more accurate models to better tackle conservation issues (Guisan et al. 2013, Rodríguez et al. 2007). For the time being, producing uncertainty maps can be an alternative (Rocchini et al. 2011, Tessarolo et al. 2014) and can provide additional information to environmental managers and stakeholders (Addison et al. 2013, Guisan et al. 2013).

Model reliability and performance also rely on the interaction between dataset completeness and species intrinsic ecological properties. Hence, we showed that the type and width of ecological niches were important to consider, distribution of narrow niche species being easier to model and less sensitive to incomplete datasets (Guo et al. 2015, Ranc et al.

2016). However, narrow niches usually imply that species are distributed over small areas for which distribution models will be highly sensitive to extrapolations.

Our protocol shows that reliable SDMs can be produced when enough data are available and dataset bias can be tested and corrected. In the present study, one SDM only (*C. nutrix*) could be corrected for spatial and temporal heterogeneities to generate reliable distribution predictions. However, our results stress the need to consider methodological issues when modeling species distribution based on poor and spatially biased datasets. They should contribute to bring new insights and enhance modeling performance in future studies.

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References

Addison PF, Rumpff L, Bau SS, Carey JM and others (2013) Practical solutions for making models indispensable in conservation decision-making. Divers. Distrib. 19: 490–502.

Aguiar LMDS, Rosa ROL, Jones G and Machado RB (2015) Effect of chronological addition of records to species distribution maps: The case of *Tonatia saurophila maresi* (Chiroptera, Phyllostomidae) in South America. Austral Ecol. 40: 836–844.

Aguirre-Gutiérrez J, Carvalheiro LG, Polce C, van Loon EE and others (2013) Fit-for-purpose: species distribution model performance depends on evaluation criteria-Dutch hoverflies as a case study. PloS one 8: e63708. Araújo MB and Guisan A (2006) Five (or so) challenges for species distribution modelling. J. Biogeogr. 33: 1677–1688. Barbet-Massin M, Jiguet F, Albert CH and Thuiller W (2012) Selecting pseudo-absences for species distribution models: how, where and how many? Method. Ecol. Evol. 3: 327–338. Bradie J and Leung B (2016) A quantitative synthesis of the importance of variables used in MaxEnt species distribution models. J. Biogeogr. 44: 1344-1361. Breiman L (2001) Random forests. Machine learning. 45: 5–32. Brotons L, Thuiller W, Araújo MB and Hirzel AH (2004). Presence-absence versus presence-only modelling methods for predicting bird habitat suitability. Ecography. 27(4): 437-448. Costa GC, Nogueira C, Machado RB and Colli GR (2010) Sampling bias and the use of ecological niche modeling in conservation planning: a field evaluation in a biodiversity hotspot. Biodiv. Conserv. 19: 883-899. Crase B, Liedloff AC and Wintle BA (2012) A new method for dealing with residual spatial autocorrelation in species distribution models. Ecography. 35: 879-888.

574 Danis B, Van de Putte A, Renaudier S and Griffiths H (2013) Connecting biodiversity data 575 during the IPY: the path towards e-polar science. Adapt. Evol. Mar. Env. Springer Berlin Heidelberg. 2: 21–32. 576 577 578 David B, Choné T, Mooi R, De Ridder C (2005) Antarctic Echinoidea – Synopses of the 579 Antarctic benthos. 10: 273. 580 581 De Broyer C, Koubbi P, Griffiths H, Grant A and others (2014) Biogeographic Atlas of the 582 Southern Ocean. Scientific Committee on Antarctic Research, Cambridge. 498pp. 583 Dormann CF (2007) Effects of incorporating spatial autocorrelation into the analysis of 584 585 species distribution data. Glob. Ecol. Biogeogr. 16:129–138. 586 587 Dormann CF, Elith J, Bacher S, Buchmann C and others (2013) Collinearity: a review of 588 methods to deal with it and a simulation study evaluating their performance. Ecography. 36: 589 27–46. 590 591 Douglass LL, Turner J, Grantham HS, Kaiser S, Constable A and others (2014) A hierarchical 592 classification of benthic biodiversity and assessment of protected areas in the Southern Ocean. 593 PloS one. 9: e100551. 594 595 Duque-Lazo J, Van Gils HAMJ, Groen TA and Navarro-Cerrillo RM (2016) Transferability 596 of species distribution models: The case of *Phytophthora cinnamomi* in Southwest Spain and 597 Southwest Australia. Ecol. Model. 320: 62–70.

599 Elith J, Leathwick JR and Hastie T (2008) A working guide to boosted regression trees. J. 600 Anim. Ecol. 77: 802-813. 601 602 Elith J and Leathwick JR (2009) Species distribution models: ecological explanation and 603 prediction across space and time. Annu. Rev. Ecol. Evol. Syst. 40: 677–697. 604 605 Elith J and Leathwick J (2014). Boosted Regression Trees for ecological modeling. At 606 http://www.lcis.com.tw/paper store/paper store/brt(5)-2015115131033846.pdf, accessed 607 02/2016. 608 609 Féral JP et al. (2016) PROTEKER: implementation of a submarine observatory at the 610 Kerguelen Islands (Southern Ocean) – Underwater Technol. 3: 3–10. 611 Ferrier S and Guisan A (2006) Spatial modelling of biodiversity at the community level. J. 612 613 Appl. Ecol. 43: 393-404. 614 615 Ficetola GF, Thuiller W and Padoa-Schioppa E (2009) From introduction to the establishment 616 of alien species: bioclimatic differences between presence and reproduction localities in the 617 slider turtle. Divers. Distrib. 15: 108–116. 618 619 Fielding AH and Bell JF (1997) A review of methods for the assessment of prediction errors 620 in conservation presence/absence models. Environ. Conserv. 24: 38–49. 621 622 Franklin J (2009) Mapping species distributions: spatial inference and prediction. University

623

Press, Cambridge: 320pp.

624 625 Greathead C, González-Irusta JM, Clarke J, Boulcott P and others (2014) Environmental 626 requirements for three sea pen species: relevance to distribution and conservation. ICES J. 627 Mar. Sci. 72: 576-586. 628 629 Griffiths HJ, Danis B and Clarke A (2011) Quantifying Antarctic marine biodiversity: The 630 SCAR-MarBIN data portal. Deep Sea Res. Pt II. 58: 18–29. 631 Guillaumot C, Martin A, Fabri-Ruiz S, Eléaume M and Saucède T (2016) Echinoids of the 632 633 Kerguelen Plateau: occurrence data and environmental setting for past, present, and future 634 species distribution modelling. ZooKeys. 630: 1–17. 635 636 Guillera-Arroita G, Lahoz-Monfort JJ, Elith J, Gordon A and others (2015) Is my species 637 distribution model fit for purpose? Matching data and models to applications. Glob. Ecol. 638 Biogeogr. 24: 276-292. 639 640 Guisan A, Tingley R, Baumgartner JB, Naujokaitis-Lewis I and others (2013) Predicting 641 species distributions for conservation decisions. Ecol. Lett. 16: 1424–1435. 642 643 Guo C, Lek S, Ye S, Li W, Liu J and Li Z (2015) Uncertainty in ensemble modelling of large-644 scale species distribution: Effects from species characteristics and model techniques. Ecol. 645 Model. 306: 67-75.

Gutt J, Zurell D, Bracegridle T, Cheung W and others (2012) Correlative and dynamic species 647 648 distribution modelling for ecological predictions in the Antarctic: a cross-disciplinary concept. Polar Res. 31:11091. 649 650 651 Gutt J, Barnes D, Lockhart SJ and van de Putte A. (2013) Antarctic macrobenthic 652 communities: A compilation of circumpolar information. Nat. Conserv. 4: 1–13. 653 654 Hand D.J. (2009). Measuring classifier performance: a coherent alternative to the area under 655 the ROC curve. Machine learning. 77(1): 103-123. 656 657 Heikkinen RK, Marmion M and Luoto M (2012) Does the interpolation accuracy of species 658 distribution models come at the expense of transferability? Ecography. 35: 276–288. 659 660 Hernandez PA, Graham CH, Master LL and Albert DL (2006). The effect of sample size and 661 species characteristics on performance of different species distribution modeling methods. 662 Ecography. 29(5): 773–785. 663 664 Hijmans RJ, Phillips S, Leathwick J and Elith J (2016) dismo: Species Distribution Modeling. 665 R package version 1.1-1. https://CRAN.R-project.org/package=dismo. 666 667 Hortal J, Lobo JM and Jimenez-Valverde A (2007) Limitations of biodiversity databases: case 668 study on seed-plant diversity in Tenerife, Canary Islands. Conserv. Biol. 21: 853–863.

- Hortal J, Jiménez-Valverde A, Gómez JF, Lobo JM and Baselga A (2008) Historical bias in
- biodiversity inventories affects the observed environmental niche of the species. Oikos. 117:
- 672 847–858.

- Jackson AL, Inger R, Parnell AC and Bearhop S (2011) Comparing isotopic niche widths
- among and within communities: SIBER-Stable Isotope Bayesian Ellipses in R. J. Ann. Ecol.
- 676 80: 595–602.

677

- 678 Jiménez-Valverde A (2012) Insights into the area under the receiver operating characteristic
- 679 curve (AUC) as a discrimination measure in species distribution modelling. Glob. Ecol.
- 680 Biogeogr. 21: 498–507.

681

- 682 Kennicutt MC, Chown SL, Cassano JJ, Liggett D and others (2014) Six priorities for
- Antarctic science. Nature. 512: 23–25.

684

- Koubbi P, Mignard C, Causse R, Da Silva O and others (2016) Ecoregionalisation of the
- Kerguelen and Crozet islands oceanic zone Part I: Introduction and Kerguelen oceanic zone.
- 687 Workshop Paris. 06/6-9/2016.

688

- Lobo JM, Jiménez-Valverde A and Real R (2007). AUC: a misleading measure of the
- 690 performance of predictive distribution models. Glob. Ecol. Biogeog. 17(2): 145-151.

- 692 Loiselle BA, Jørgensen PM, Consiglio T, Jiménez I and others (2008) Predicting species
- distributions from herbarium collections: does climate bias in collection sampling influence
- model outcomes? J. Biogeogr. 35: 105–116.

695 696 Luoto M, Pöyry J, Heikkinen RK and Saarinen K (2005) Uncertainty of bioclimate envelope models based on the geographical distribution of species. Glob. Ecol. Biog. 14: 575–584. 697 698 699 Mateo RG, Croat TB, Felicisimo AM and Munoz J. (2010). Profile or group discriminative 700 techniques? Generating reliable species distribution models using pseudo-absences and target-701 group absences from natural history collections – Divers. Distrib. 16:84–94. 702 703 McCoy FW (1991) Southern Ocean sediments: circum-Antarctic to 30°S. Marine Geological 704 and Geophysical Atlas of the circum-Antarctic to 30°S. (ed. by D.E. Hayes) – Antarctic 705 Research Series. 706 707 McCune JL (2016) Species distribution models predict rare species occurrences despite 708 significant effects of landscape context – J. Appl. Ecol. 53: 1871–1879. 709 710 Merckx B, Steyaert M, Vanreusel A, Vincx M and Vanaverbeke J (2011) Null models reveal 711 preferential sampling, spatial autocorrelation and overfitting in habitat suitability modelling. 712 Ecol. Model. 222: 588-597. 713 714 Muscarella R, Galante PJ, Soley-Guardia M, Boria RA and others (2014) ENMeval: An R 715 package for conducting spatially independent evaluations and estimating optimal model 716 complexity for ecological niche models. Method. Ecol. Evol. 5: 1198–1205. 717 718 Naimi B, Hamm NA, Groen TA, Skidmore AK and Toxopeus AG (2014) Where is positional 719 uncertainty a problem for species distribution modelling? Ecography. 37: 191–203.

- 721 Newbold T (2010) Applications and limitations of museum data for conservation and
- ecology, with particular attention to species distribution models. Prog. Phys. Geogr. 34: 3–22.

- Paradis E, Claude J and Strimmer K (2004) APE: analyses of phylogenetics and evolution in
- 725 R language. Bioinformatics. 20: 289–290.

726

- Peterson AT, Papeş M and Soberón J (2008). Rethinking receiver operating characteristic
- analysis applications in ecological niche modeling. Ecol. Model. 213(1): 63–72.

729

- 730 Phillips SJ, Anderson RP and Schapire RE (2006) Maximum entropy modeling of species
- 731 geographic distributions. Ecol. Model. 190: 231259.

732

- 733 Phillips SJ, Dudík M, Elith J, Graham CH and others (2009) Sample selection bias and
- presence-only distribution models: implications for background and pseudo-absence data.
- 735 Ecol. Appl. 19: 181–197.

736

- Phillips ND, Reid N, Thys T, Harrod C, Payne N and others (2017) Applying species
- distribution modelling to a data poor, pelagic fish complex: the ocean sunfishes. J. Biogeogr.

739

- Pierrat B, Saucède T, Laffont R, De Ridder C and others (2012) Large-scale distribution
- analysis of Antarctic echinoids using ecological niche modelling. Mar. Ecol. Prog. Ser. 463:
- 742 215–230.harel KP, Ludwig T and Storch I (2016). Predicting potential distribution of poorly
- known species with small database: the case of four horned antelope *Tetracerus quadricornis*
- on the Indian subcontinent. Ecol. Evol. 6: 2297-2307.

- Proosdij A.S., Sosef M.S., Wieringa J.J. and Raes N. (2016). Minimum required number of
- specimen records to develop accurate species distribution models. Ecography. 39(6): 542–
- 748 552.

- 750 Qiao H, Soberón J and Peterson AT (2015) No silver bullets in correlative ecological niche
- 751 modelling: insights from testing among many potential algorithms for niche estimation.
- 752 Method. Ecol. Evol. 6: 1126–1136.

753

- Raes N and ter Steege H (2007) A null-model for significance testing of presence-only
- species distribution models. Ecography. 30: 727–736.

756

- Ranc N, Santini L, Rondinini C, Boitani L and others (2016) Performance tradeoffs in target-
- 758 group bias correction for species distribution models. Ecography. 39: 1–12.

759

- Randin CF, Dirnböck T, Dullinger S, Zimmermann NE and others (2006) Are niche-based
- species distribution models transferable in space? J. Biogeogr. 33: 1689–1703.

762

- Reiss H, Cunze S, König K, Neumann H and Kröncke I (2011) Species distribution modelling
- of marine benthos: a North Sea case study. Mar. Ecol. Prog. Ser. 442: 71–86.

765

- Reiss H, Birchenough S, Borja A, Buhl-Mortensen L and others (2014) Benthos distribution
- modelling and its relevance for marine ecosystem management. ICES J. Mar. Sci. 72: 297–
- 768 315.

- Reutter BA, Helfer V, Hirzel AH and Vogel P (2003) Modelling habitat suitability using
- 771 museum collections: an example with three sympatric Apodemus species from the Alps. J.
- 772 Biogeogr. 30: 581–590.

- Ridgeway G (2015) gbm: Generalized Boosted Regression Models. R package version 2.1.1.
- 775 https://CRAN.R-project.org/package=gbm.

776

- 777 Robinson LM, Elith J, Hobday AJ, Pearson RG and others (2011) Pushing the limits in
- 778 marine species distribution modelling: lessons from the land present challenges and
- opportunities. Glob. Ecol. Biogeogr. 20: 789–802.

780

- 781 Rocchini D, Hortal J, Lengyel S, Lobo JM and others (2011) Accounting for uncertainty
- when mapping species distributions: the need for maps of ignorance. Prog. Phys. Geogr. 35:
- 783 211–226.

784

- 785 Rödder D and Engler JO (2011) Quantitative metrics of overlaps in Grinnellian niches:
- advances and possible drawbacks. Glob. Ecol. Biogeogr. 20: 915–927.

787

- Rodríguez JP, Brotons L, Bustamante J and Seoane J (2007) The application of predictive
- modelling of species distribution to biodiversity conservation. Divers. Distrib. 13: 243–251.

790

- 791 Segurado PA, Araújo MB and Kunin WE (2006) Consequences of spatial autocorrelation for
- 792 niche-based models. J. Appl. Ecol. 43: 433–444.

794 Smith WH and Sandwell DT (1997) Global sea floor topography from satellite altimetry and 795 ship depth soundings. Science. 277: 1956–1962. 796 797 Stockwell DR and Peterson AT (2002) Effects of sample size on accuracy of species 798 distribution models. Ecol. Model. 148: 1–13. 799 800 Tessarolo G, Rangel TF, Araújo MB and Hortal J (2014) Uncertainty associated with survey 801 design in Species Distribution Models. Divers. Distrib. 20: 1258–1269. 802 803 Thuiller W, Georges D, Engler R, Breiner F and others (2016) biomod2: Ensemble Platform 804 for Species Distribution Modeling. R package version 3.3-7. https://CRAN.R-805 project.org/package=biomod2. 806 807 Van de Putte A. et al. (2014) Chapter 2.1. Data and Mapping. In De Broyer C., Koubbi P., 808 Griffiths H.J., Raymond B., Udekem d'Acoz C. d', et al. (eds.). Biogeographic Atlas of the 809 Southern Ocean. Scientific Committee on Antarctic Research, Cambridge: 14–15. 810 811 Venables WN and Ripley BD (2002) Modern Applied Statistics with S. Fourth Edition. 812 Springer, New York. ISBN 0-387-95457-0. 813 814 Vierod AD, Guinotte JM and Davies AJ (2014) Predicting the distribution of vulnerable

marine ecosystems in the deep sea using presence-background models. Deep Sea Res. II. 99:

815

816

817

6-18.

818	Warren DL, Glor RE and Turelli M (2008) Environmental niche equivalency versus
819	conservatism: quantitative approaches to niche evolution. Evolution. 62: 2868–2883.
820	
821	Wenger SJ and Olden JD (2012) Assessing transferability of ecological models: an
822	underappreciated aspect of statistical validation. Method. Ecol. Evol. 3: 260–267.
823	
824	Wisz MS, Hijmans RJ, Li J, Peterson AT, Graham CH and Guisan A (2008) Effects of sample
825	size on the performance of species distribution models. Divers. Distrib. 14: 763–773.
826	
827	Zucchetta M, Venier C, Taji MA, Mangin A and Pastres R (2016) Modelling the spatial
828	distribution of the seagrass Posidonia oceanica along the North African coast: Implications
829	for the assessment of Good Environmental Status. Ecol. Indic. 61: 1011–1023.
830	
831	
832	
833	
834	
835	
836	
837	
838	
839	
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Figures legend list

Figure 1: (A) Map showing occurrence data of the four studied echinoid species over the Kerguelen Plateau: *Brisaster antarcticus* (orange diamond), *Ctenocidaris nutrix* (grey circle), *Sterechinus diadema* (green triangle) and *Abatus cordatus* (purple square). (B) Evolution of sampling effort (in presence-only records) through time with the position of main scientific cruises during which the four studied species were collected on the Kerguelen Plateau. (C) Species presence data plotted according to depth (z axis), seafloor salinity (y axis) and seafloor temperature (x axis) on the extent of the Kerguelen Plateau with projection of standardized distribution ellipsoids (see Jackson et al. 2011 for details) on bivariate plots. (D) Species depth range over the Kerguelen Plateau based on occurrence data (solid line: median, box: upper and lower quartiles, whiskers: 75% ± 1.5 interquartile range, dots: outliers).

Figure 2. Tests and procedures carried out in the present work. Arrows indicate the stepwise procedure with statistical validation conducting either to the following step or correction/stepback requirements.

Figure 3: (A) Null model #1 and (B) Null model #2 for the different species under study. Mean AUC value and standard deviation are given for the 100 replicates. Comparisons between models compiled with Pearson r correlation value and associated probabilities.

Figure 4: First row: Distribution models of *Ctenocidaris nutrix* species with increasing number of presence data to build the model, for four periods. Averaged maps of 100 model replicates. Second row: (A) Difference in probability distribution between n=54 and n=46, (B) between n=106 and n=54, (C) between n=114 and n=106.

Figure 5: Mean contributions of environmental descriptors to the models with standard deviation (error bars) for the four time periods and species under study. sst= sea surface temperature, sst amp= sea surface temperature amplitude, sssalinity= sea surface salinity, sst amp= sea surface salinity amplitude, chl a= chlorophyll-a (see Guillaumot et al. 2016 for details). Figure 6: Species distribution model generated for Ctenocidaris nutrix using all presence-only data available and present environmental descriptors (2005-2012). AUC=0.813±0.02

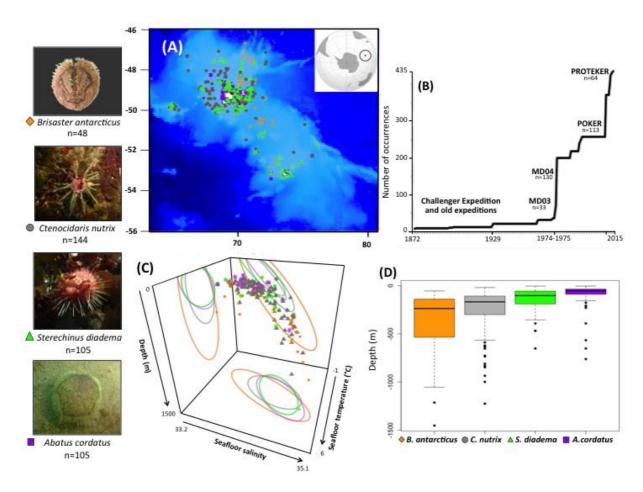


Figure 1: (A) Map showing occurrence data of the four studied echinoid species on the Kerguelen Plateau: *Brisaster antarcticus* (orange diamond), *Ctenocidaris nutrix* (grey circle), *Sterechinus diadema* (green triangle) and *Abatus cordatus* (purple square). (B) Evolution of sampling effort (in presence-only records) through time with the position of main scientific cruises during which the four studied species were collected on the Kerguelen Plateau. (C) Species presence data plotted according to depth (z axis), seafloor salinity (y axis) and seafloor temperature (x axis) on the extent of the Kerguelen Plateau with projection of standardized distribution ellipsoids (see Jackson et al. 2011 for details) on bivariate plots. (D) Species depth range over the Kerguelen Plateau based on occurrence data (solid line: median, box: upper and lower quartiles, whiskers: 75% ± 1.5 interquartile range, dots: outliers).

Table 1: List of environmental descriptors selected for SDM. Asterisks ^(*) indicate that environmental layers are available for the following time periods: [1955-2012], [1955-1964], [1965-1974], [1975-1994], [2005-2012]. Minimum and maximum values are indicated for [1955-2012]. Spatial resolution of layers: 10km resolution grid-cell pixels.

Environmental descriptors	Unit	Description	on Min value M		Source	Spatial extent
Depth	Meters	Bathymetric grid around the Kerguelen Plateau	-4977.0000	-1.0000	This study. Derived from the Biogeographic Atlas of the Southern Ocean (De Broyer et al. 2014)	46_56°S/63_81°E
Sea surface mean temperature*	°Celsius degrees	Mean sea surface temperature	3.0566	7.6223	World Ocean Circulation Experiment 2013	46_56°S/63_81°E
Sea surface temperature amplitude*	°Celsius degrees	Amplitude between mean summer and mean winter sea surface temperature	-3.3036	-1.4108	World Ocean Circulation Experiment 2013	46_56°S/63_81°E
Seafloor mean temperature*	°Celsius degrees	Mean seafloor temperature	-0.2978	4.6422	This study. Derived from World Ocean Circulation Experiment 2013 sea surface temperature layers	46_56°S/63_81°E
Seafloor temperature amplitude*	*Colsius Amplitude between meen summer and This study. Derived from World Oc		This study. Derived from World Ocean Circulation Experiment 2013 sea surface temperature layers	46_56°S/63_81°E		
Sea surface mean salinity*	Sea surface mean salinity* PSS Mean sea surface salinity 33.6849 33.8		33.8251	World Ocean Circulation Experiment 2013	46_56°S/63_81°E	
Sea surface salinity amplitude*	PSS	Amplitude between mean summer and mean winter sea surface salinity	-0.0859	0.3165	World Ocean Circulation Experiment 2013	46_56°S/63_81°E
Seafloor salinity amplitude*	PSS	Amplitude between mean summer and mean winter seafloor salinity	- 169	0.0937	This study. Derived from World Ocean Circulation Experiment 2013 sea surface salinity layers	46_56°S/63_81°E
Mean surface chlorophyll a	ean surface chlorophyll a mg/m³ Surface chlorophyll a concentration. Summer mean over 2002-2009 0.1358 2.7324		2.7324	MODIS AQUA (NASA) 2010	46_56°S/63_81°E	
Sediments	Categorical	Sediment features	14 categories		McCoy (1991), updated by Griffiths 2014 (unpublished)	46_56°S/63_81°E
Geomorphology	Categorical	Geomorphologic features	27 categories		ATLAS ETOPO2 2014 (Douglass et al. 2014)	46_56°S/63_81°E
Slope Degrees		Bathymetric slope	4.8229. 10 ⁻	0.1547	Biogeographic Atlas of the Southern Ocean (De Broyer et al. 2014)	46_56°S/63_81°E
Mean seafloor oxygen concentration	mL/L	Mean seafloor oxygen concentration over 1955-2012	4.0080	7.6223	This study. Derived from World Ocean Circulation Experiment 2013 sea surface oxygen concentration layers	46_56°S/63_81°E

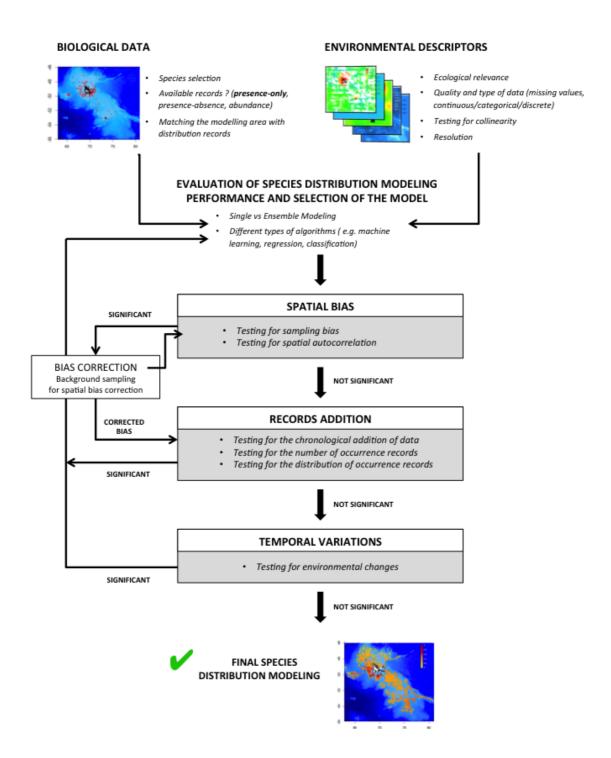


Figure 2. Tests and procedures carried out in the present work. Arrows indicate the stepwise procedure with statistical validation conducting either to the following step or correction/stepback requirements.

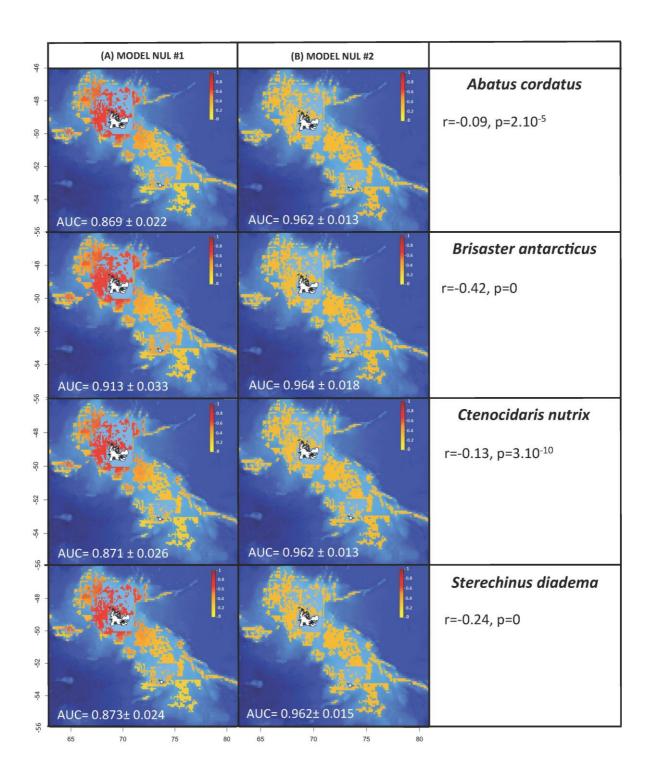


Figure 3: (A) Null model #1 and (B) Null model #2 for the different species under study. Mean AUC value and standard deviation are given for the 100 replicates. Comparisons between models compiled with Pearson r correlation value and associated probabilities.

Table 2: r Pearson correlation of pixel values between the KDE layer and the predicted probability of each species model. Statistic probabilities are all < 0.05.

	Before KDE correction	After KDE correction
Abatus cordatus	0.72	0.44
Brisaster antarcticus	0.60	-0.17
Ctenocidaris nutrix	0.80	0.11
Sterechinus diadema	0.61	0.20

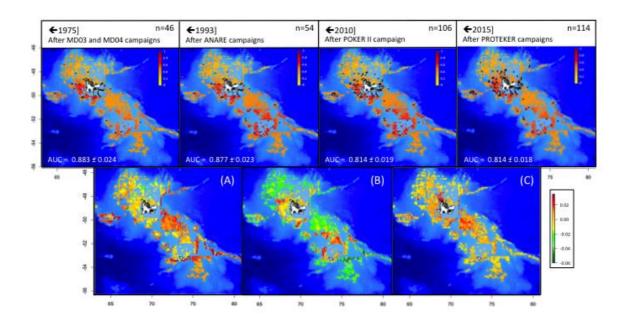


Figure 4: First row: Distribution models of *Ctenocidaris nutrix* species with increasing number of presence data to build the model, for four periods. Averaged maps of 100 model replicates. Second row: (A) Difference in probability distribution between n=54 and n=46, (B) between n=106 and n=54, (C) between n=114 and n=106.

Table 3: Influence of data addition and sampling patterns on models for *Abatus cordatus*, *Ctenocidaris nutrix* and *Sterechinus diadema*. Column 1: mean Schoener's D and associated *p*-value computed between models (100 replicates) produced respectively with {n=54, 76, 95}, {n=46, 54, 106, 114} and {n=54, 66, 98} occurrences randomly sampled from the total dataset. Column 2: mean Schoener's D and associated p-value computed between models (100 replicates) produced with subsets contrasting in data distribution patterns (transect versus random sampling).

Species	occurrenc	e number	sampling pattern		
	Mean Dobs	Mean D _{obs} Mean p-value		p-value	
Abatus cordatus	0.981 ± 0.025	< 0.05		-	
Ctenocidaris nutrix	0.985 ± 0.020	< 0.05	0.941 ± 0.030	0.147	
Sterechinus diadema	0.979 ± 0.031	< 0.05	0.842 ± 0.040	0.941	

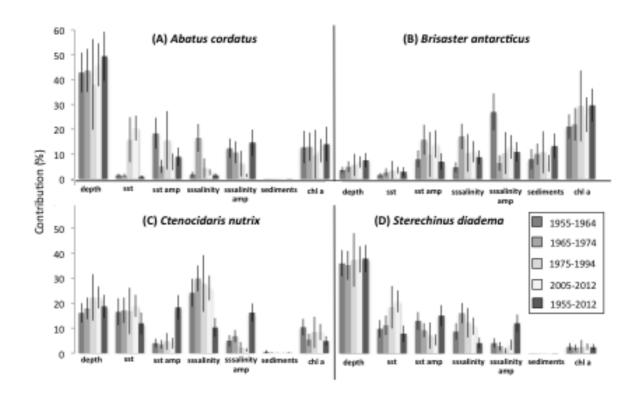


Figure 5: Mean contributions of environmental descriptors to the models with standard deviation (error bars) for the four time periods and species under study. sst= sea surface temperature, sst amp= sea surface temperature amplitude, sssalinity= sea surface salinity, sst amp= sea surface salinity amplitude, chl a= chlorophyll-a (see Guillaumot et al. 2016 for details).

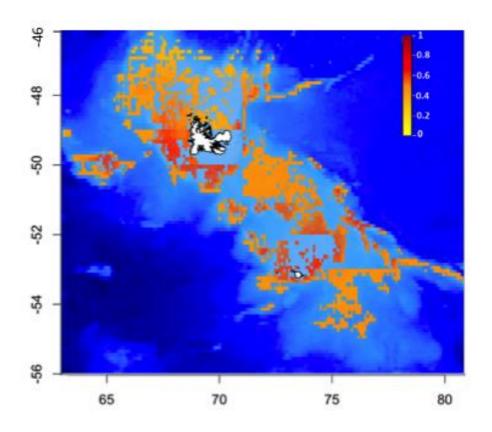


Figure 6: Species distribution model generated for *Ctenocidaris nutrix* using all presence-only data available and present environmental descriptors (2005-2012). AUC=0.813±0.02