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## Functionalization of Reduced Graphene Oxide via Thiol–Maleimide “Click” Chemistry: Facile Fabrication of Targeted Drug Delivery Vehicles

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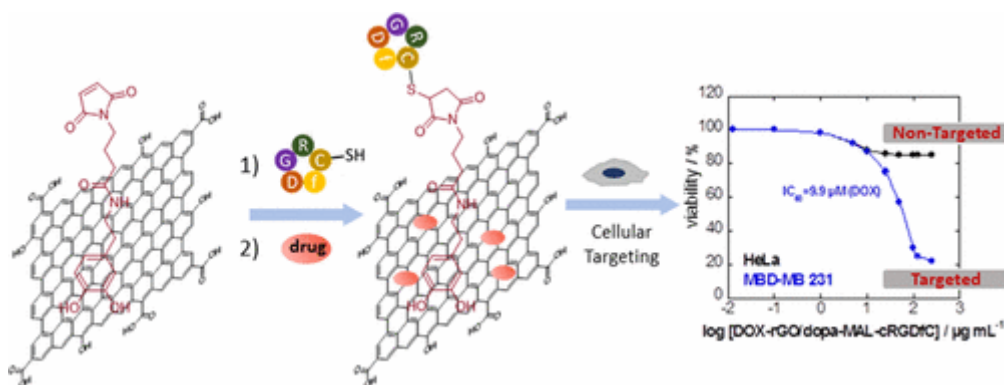
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Abstract.



Materials based on reduced graphene oxide (rGO) have shown to be amenable to noncovalent functionalization through hydrophobic interactions. The scaffold, however, does not provide sufficient covalent linkage given the low number of reactive carboxyl and alcohol groups typically available on the rGO. The integration of clickable groups, particularly the ones that can undergo efficient conjugation without any metal catalyst, would allow facile functionalization of these materials. This study reports on the noncovalent association of a maleimide-containing catechol (dopa-MAL) surface anchor onto the rGO. Thiol–maleimide chemistry allows thereafter the facile attachment of thiol-containing molecules under ambient metal-free conditions. Although the attachment of glutathione and 6-(ferrocenyl)hexanethiol

was used as model thiols, the attachment of a cancer cell targeting cyclic peptide, c(RGDfC), opened the possibility of using the dopa-MAL-modified rGO as a targeted drug delivery system for doxorubicin (DOX). Although free DOX showed to be more effective at killing the human cervical cancer cells (HeLa) over human breast adenocarcinoma cancer cells (MDA-MB-231), the DOX-loaded rGO/dopa-MAL-c (RGDfC) nanostructure showed an opposite effect being notably more effective at targeting and killing the MDA-MB-231 cells. The effect is enhanced upon laser irradiation for 10 min at  $2 \text{ W cm}^{-2}$ . The facile fabrication and functionalization to readily obtain a functional material in a modular fashion make this clickable-rGO construct an attractive platform for various applications.

*Keywords:*

[cellular targeting](#); [click chemistry](#); [DOX delivery](#); [graphene oxide](#); [photothermal effect](#); [thiol–maleimide conjugation](#)

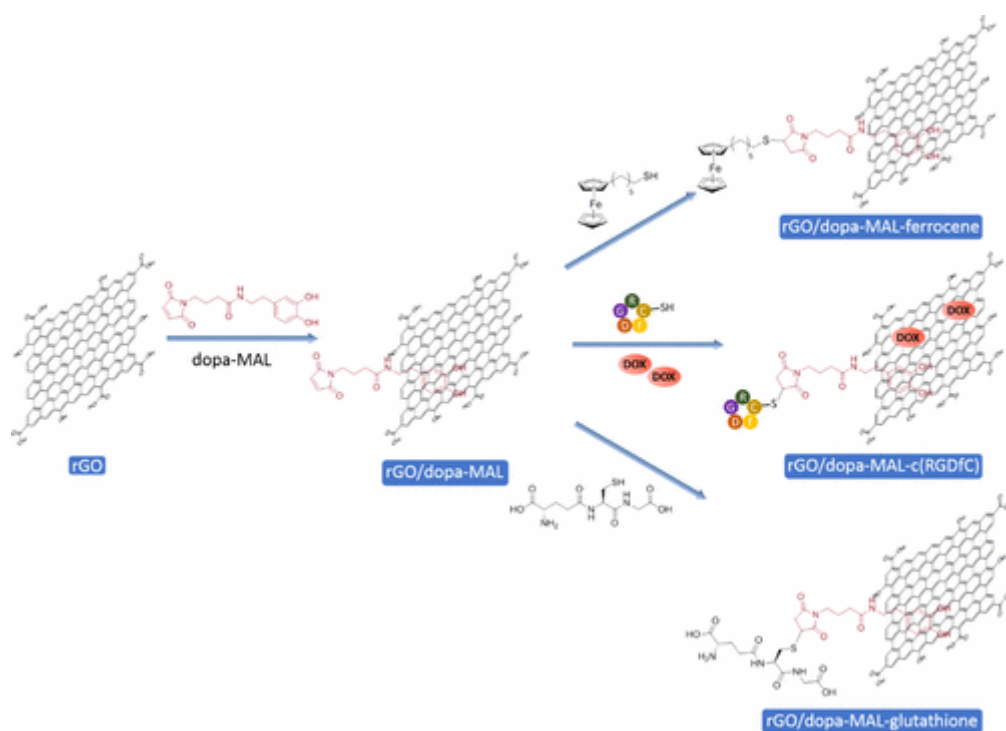
## 1. Introduction

Nanosystems that release cancer drugs and induce toxicity in response to an external stimulus offer unique advantages over the commonly employed conventional carriers that release their payload in a passive fashion. Reduced graphene oxide (rGO) nanosheets with restored  $sp^2$  network have most lately shown to be ideally suited for drug-loading- and drug-release-based applications using electrical(1-3) as well as thermal induced triggers.(4-10) Although the majority of the aromatic scaffold of rGO is devoid of any functional groups, a few carboxylic acid and alcohol groups are present at the edges and other defect points. This limits the effective functionalization of the rGO through covalent transformations. To circumvent this limitation, the noncovalent  $\pi$ – $\pi$  stacking-based functionalization route of the rGO scaffold has been extensively exploited.(11-16) This approach is effective for hydrophobic/aromatic molecules that are able to undergo efficient noncovalent interactions. Conjugation of molecules unable to undergo such an association with the rGO, however, becomes challenging. An approach that would allow facile and effective covalent attachment in a modular manner to the rGO would be rather useful.

Aromatic anchoring groups like pyrene,(11, 17) tetrathiafulvalene(13, 15) and dopamine derivatives(14, 16) have shown to be of high interest for rGO functionalization. Some of us have reported surface functionalization of the rGO with dopamine-containing clickable alkyne and azide groups.(14, 16) The azide-containing rGO could be further modified with alkynyl-terminated molecules such as ferrocene via the Cu(I)-catalyzed azide–alkyne Huisgen-type click reaction. Although the azide–alkyne-based rGO functionalization was effective, the requirement to use copper as a catalyst can be a concern for the biomedical applications because any trace of copper adds to the overall cytotoxicity of the nanostructures. This necessitates the expansion of the tool box to include click reactions that do not need metal catalysts, like the strain-promoted azide–alkyne cycloaddition (SPAAC)(18) and the nucleophilic or radical thiol–ene reactions.(16, 19) Although approaches based on SPAAC have gained interest lately,(20) the multistep synthesis or high cost of the cyclooctyne-based materials limits the widespread utilization of this approach. Nucleophilic thiol–ene-based reactions have been extensively employed in the bioconjugations using the thiol–maleimide functional group dyad and seem advantageous.(21-24) The maleimide–thiol addition reaction is a facile process that does not require any catalyst and can be carried out under mild

conditions. Although thiol groups can be readily incorporated in various molecules, they are also present in the cysteine residue of various peptides and biomolecules, or can be installed using standard protocols during or after synthesis.

Herein, we report the synthesis of a maleimide functional group containing the rGO, which is amenable to facile functionalization by thiol group containing molecules ([Scheme 1](#)). The conjugation of thiol-bearing molecules proceeds through the thiol–ene click-type conjugation reaction, under metal-free conditions. Maleimide functionalized rGO was easily prepared through  $\pi$ – $\pi$  interactions under ultrasonication of the rGO with a maleimide-containing dopamine (dopa-MAL) ligand. To demonstrate the utility of this clickable rGO, we took the advantage of the covalent conjugation of a thiol-bearing targeting peptide, c(RGDfC), and the noncovalent loading of the rGO–c(RGDfC) nanostructures with the anticancer drug doxorubicin (DOX). The two-dimensional structure of graphene sheets serves as an excellent drug carrier due to its high drug loading capacity arising from the  $\pi$ – $\pi$  stacking and other hydrophobic interactions. Attachment of a targeting group on the rGO through covalent conjugation further expands its application to the targeted drug delivery. Selective delivery of anticancer agents to the tumor site has become important for efficient cancer therapy by minimizing the negative side effects. Synthetic cyclic peptides with arginine–glycine–aspartic acid sequences (RGDs) are known to possess high affinity to integrin  $\alpha_v\beta_3$ , a cell adhesion domain highly expressed on the tumor cells.[\(25, 26\)](#) Their integration into the nanostructures and polymers for targeted drug delivery has thus found growing interest.[\(27, 28\)](#)



**Scheme 1.** Schematic Illustration of the Synthesis of Thiol–Ene “Clickable” Maleimide-rGO and its Subsequent Functionalization

Finally, the excellent photothermal properties of DOX-loaded rGO/dopa-MAL-c(RGDfC) allow for chemophototherapy; the combination of photothermal therapy with chemotherapeutic release of DOX is often beneficial specifically for solid tumors.<sup>(29)</sup> The cytotoxicity of targeted and nontargeted rGO under normal and photothermal conditions was assessed on two cancer cell lines, human breast MDA-MB-231 and human cervical cancer cells (HeLa), to highlight the advantages of the construct. Overall, facile fabrication and functionalization of a maleimide group containing rGO-based functional material with potential for applications in many areas is outlined.

## 2. Experimental Section

### 2.1 Materials

*N*-Hydroxysuccinimide (NHS) was purchased from Alfa Aesar. *N,N'*-Dicyclohexylcarbodiimide (DCC), 4-(dimethylamino)pyridine (DMAP), dopamine hydrochloride, triethylamine (TEA),  $\gamma$ -aminobutyric acid, l-glutathione reduced, 6-(ferrocenyl)hexanethiol, graphite powder (<20  $\mu\text{m}$ ) and hydrazine monohydrate were purchased from Sigma-Aldrich. 4-(*exo*-3,5-Dioxo-10-oxa-4-aza-tricyclo[5.2.1.0]dec-8-en-4-yl)-butyric acid (furan-protected maleimide containing acid) was synthesized according to a previous report.<sup>(30)</sup> Bags with regenerated cellulose dialysis membranes (Spectra/Por Biotech, molecular weight cut-offs 10 kDa) were obtained from Spectrum Laboratories. Purification by silica gel column was done using silica gel 60 (Merck). Purity of the compounds was determined using thin-layer chromatography on the silica-coated aluminum plates (Kieselgel 60 F254, Merck). Methanol, tetrahydrofuran, and dimethylformamide (DMF) were bought from Merck. Anhydrous toluene from SciMatCo system was used, and other solvents were stored on the molecular sieves when needed. Graphene oxide was bought from Graphenea, Spain.

### 2.2. Preparation of Reduced Graphene Oxide (rGO)

GO aqueous suspension (150 mg, 3 mg mL<sup>-1</sup>) was sonicated and hydrazine hydrate (50  $\mu\text{L}$ , 1.03 mmol) added to this suspension. The mixture was heated for 24 h at 100 °C and rGO slowly precipitated out. The desired material was obtained by filtering with a poly(vinylidene difluoride) membrane (0.45  $\mu\text{m}$  pore size), rinsed with water (20 mL  $\times$  5) and methanol (20 mL  $\times$  5), followed by drying in an oven.

### 2.3. Synthesis of Maleimide-Containing Dopamine (dopa-MAL)

#### 2.3.1. Protected Maleimide-Containing Dopamine (dopa-pMAL)

Furan-protected maleimide-containing acid (2.45 g, 9.76 mmol), NHS (1.18 g, 10.25 mmol), and DMAP (0.24 g, 1.95 mmol) were dissolved in DMF (15 mL) in a round-bottom flask at 0 °C. DCC dissolved in 5 mL of DMF was subsequently added to this solution dropwise. After stirring at 0 °C for 15 min, the reaction was left to proceed at room temperature for 16 h. The solvent was removed in vacuo and the product (NHS ester of the acid) was extracted with NaHCO<sub>3</sub>. This activated ester was used without further purification. Dopamine, HCl (1.43 g, 7.54 mmol), and TEA (1.09 g, 10.8 mmol) were mixed in 10 mL of DMF with magnetic stirring for 15 min. The DMF solution of the activated ester (2.5 g, 7.18 mmol) was added to the mixture and stirred at room temperature for 24 h. The solvent was partially evaporated

( $\approx 1\text{--}2$  mL) and 1 M HCl solution (50 mL) was added to the concentrated solution. It was extracted with EtOAc. The organic phase was collected and dried over  $\text{Na}_2\text{SO}_4$ . Column chromatography using  $\text{CH}_2\text{Cl}_2$ /methanol (10:1) was employed to obtain the pure product (1.28 g, yield: 82%).

### **2.3.2. Maleimide Group Activation via Retro Diels–Alder Reaction (dopa-MAL)**

Furan–maleimide adduct-containing dopamine (1.0 g) in anhydrous toluene (150 mL) was heated under nitrogen protection at  $110\text{ }^\circ\text{C}$  for 10 h. The solution was then cooled down to ambient temperature and toluene was removed via rotary evaporation, yielding maleimide-appended dopamine (dopa-MAL) (0.80 g, yield: 97%).

### **2.4. Preparation of rGO/dopa-MAL**

To a solution of rGO in DMF ( $1\text{ mg mL}^{-1}$ ), dopa-MAL (6.3 mM) was added and the mixture ultrasonicated for 3 h at room temperature. Maleimide-functionalized rGO was collected by centrifugation at 13 000 rpm for 30 min and washed with DMF to remove all of the unbound dopa-MAL compounds. The functionalized rGO was then dried under vacuum.

### **2.5. Functionalization of rGO/dopa-MAL via Thiol–Ene Michael Addition Reactions**

#### **2.5.1. Glutathione and 6-(Ferrocenyl)hexanethiol Linking**

Dispersion of rGO/dopa-MAL in DMF ( $1\text{ mg mL}^{-1}$ ) was mixed with glutathione (10 mM) and sonicated for 6 h at room temperature. The product rGO/dopa-Mal-glutathione was collected by centrifugation at 13 000 rpm for 30 min. Similarly, 6-(ferrocenyl)hexanethiol (10 mM) was attached to rGO/dopa-MAL ( $1\text{ mg mL}^{-1}$  DMF solution) under ultrasonication for 6 h at room temperature. Purified rGO/dopa-MAL-ferrocene was obtained by centrifugation at 13 000 rpm for 30 min.

#### **2.5.2. c(RGDfC) Attachment to rGO/dopa-MAL**

c(RGDfC) (4 mM) was added to  $1\text{ mg mL}^{-1}$  DMF dispersion of rGO/dopa-MAL and the mixture sonicated for 12 h at room temperature. The nonconjugated peptide was removed by dialyzing (using MWCO 10 000 Da membrane) against water for 48 h to yield rGO/dopa-MAL-c(RGDfC).

### **2.6. Quantification of Peptide and DOX Loading**

#### **2.6.1. Peptide Quantification Using Bicinchoninic Acid (BCA) Assay**

The amount of peptide c(RGDfC) in rGO/dopa-MAL-c(RGDfC) was measured using a BCA kit (ThermoScientific) using a protocol provided by the manufacturer.

## 2.6.2. Drug Loading

Peptide-modified rGO was dispersed in water at  $0.5 \text{ mg mL}^{-1}$ . Doxorubicin hydrochloride (4.2 mM) was added to the dispersion and left to stir for 12 h at ambient temperature. The solution was centrifuged to remove free DOX from the DOX-loaded rGO/dopa-MAL-c(RGDfC) nanoconjugate.

## 2.7. Characterization

Full details of the characterization techniques and instrumentation used in this study can be found in the [Supporting Information](#).

## 2.8. Cell Viability and Internalization

The HeLa cell line derived from cervical carcinoma (ATCC CCL-2, ECACC, Sigma-Aldrich, Saint-Quentin Fallavier, France) and the MDA-MB-231 cell line derived from human breast adenocarcinoma (ATCC HTB-26, ECACC, Sigma-Aldrich, Saint-Quentin Fallavier, France) were used. Dulbecco's modified Eagle's medium (DMEM, Gibco) containing 10% fetal bovine serum (FBS, Gibco) and 1% penicillin–streptomycin (Gibco) were used to culture and maintain the cells in a humidified incubator at  $37 \text{ }^\circ\text{C}$  and 5%  $\text{CO}_2$ .

### 2.8.1. Cell Viability upon Treatment with rGO/dopa-MAL-c(RGDfC)

A 96-well plate was used to seed cells at a density of  $10^4$  cells/well and grown for 24 h prior to assay. A fresh medium containing rGO/dopa-MAL-c(RGDfC) was used to replace the culture medium. After 48 h, the old medium was aspirated, washed with phosphate-buffered saline (PBS), and replaced with new medium. Cellular viability was measured using Uptiblu (Interchim) method. Briefly, 10  $\mu\text{L}$  of the Uptiblu solution were added to the wells containing 100  $\mu\text{L}$  of DMEM with 10% FBS and the plate was incubated for 4 h in the incubator. The fluorescence of each well (exc. 540 nm/em. 590 nm) was obtained with a plate reader. Experimental runs were repeated five times. The average absorbance of the untreated cells was considered as 100% viability.

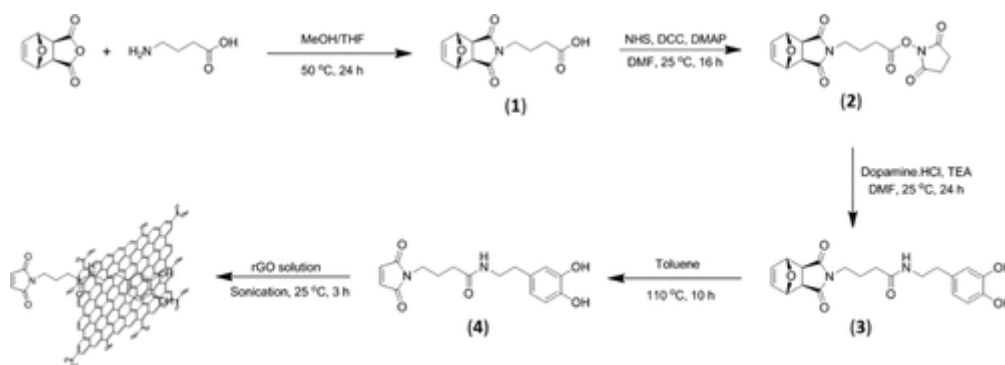
### 2.8.2. Cell Viability of DOX-Loaded rGO/dopa-MAL-c(RGDfC) under Passive and Active Conditions

Cells were seeded in a 24-well plate ( $10^5$  cells/well) and grown for 24 h prior to assay. For passive release of DOX, the cell culture medium was renewed with a fresh medium that contains DOX-loaded rGO/dopa-MAL-c(RGDfC) at pH 7.4. After 48 h, the old medium was removed, washed with PBS, and replaced with a fresh medium. The cell viability was measured using Uptiblu method as described above. For active release of DOX, the culture medium was replaced with the medium containing DOX-loaded rGO/dopa-MAL-c(RGDfC). After 6 h, each well was illuminated with a continuous laser at 980 nm at various power densities ( $1\text{--}4 \text{ W cm}^{-2}$ ) for 10 min. During the same time, thermal images were taken with an IR camera. The cell viability was evaluated after additional 18 h using Uptiblu method as described above.

### 3. Results and Discussions

#### 3.1. Synthesis of Dopamine–Maleimide-Modified reduced graphene oxide (rGO/dopa-MAL)

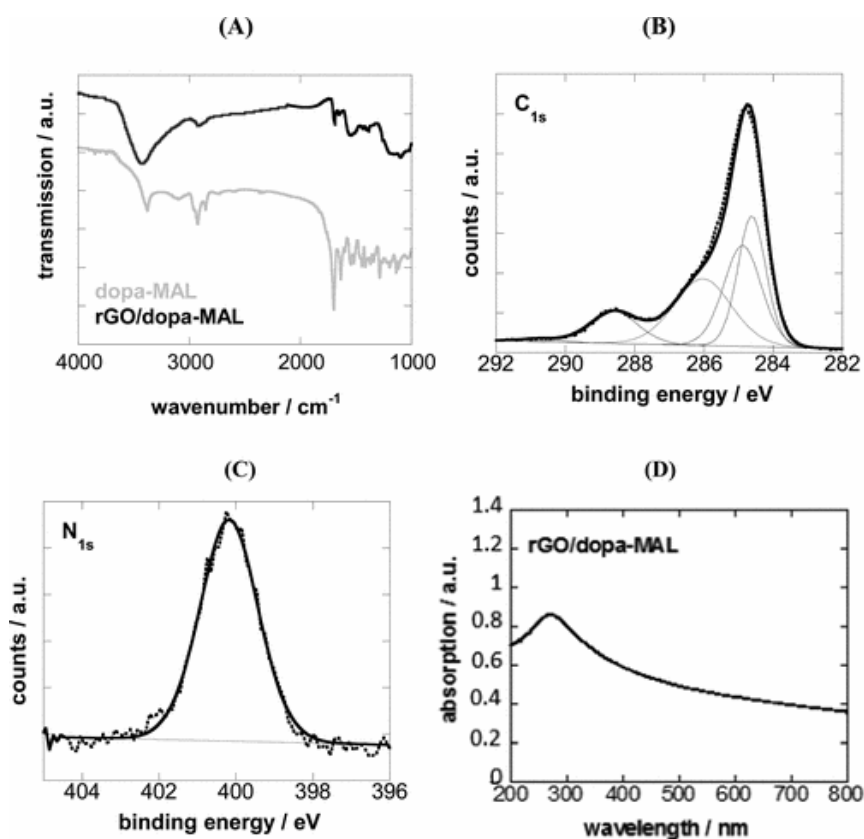
The aromatic structure of catechol units allows dopamine to develop strong  $\pi$ -stacking interactions with other aromatic bearing units such as rGO. Additionally, dopamine can be easily functionalized by exploiting its amine terminal functions as used here for the synthesis of dopa-MAL (Figure 1). 4-(*exo*-3,5-Dioxo-10-oxa-4-aza-tricyclo[5.2.1.0]dec-8-en-4-yl)-butyric acid (**1**) was obtained according to a previously reported literature procedure.<sup>(30)</sup> Dopamine group conjugation was achieved through the reaction between NHS-activated acid of maleimide derivative (**2**) and dopamine. The <sup>1</sup>H NMR spectroscopy of dopa-pMAL (**3**) is seen in Figure S1A. Furan deprotection was achieved by heating dopa-pMAL in toluene followed by NMR spectroscopy (Figure S1B). Loss of proton resonances at 2.84, 5.23, and 6.51 ppm belonging to the bicyclic core and emergence of a new proton resonance at 7.0 ppm belonging to the proton on maleimide ring were indicative of effective furan deprotection. The <sup>13</sup>C NMR spectrum for unmasked maleimide-containing dopamine (dopa-MAL, **4**) showed the expected resonances, particularly those from the two distinct carbonyl groups, one from the amide and the other from the imide group (Figure S1C).



**Figure 1.** Synthesis of dopa-MAL (**4**) and its conjugation onto rGO through  $\pi$ - $\pi$  stacking interactions

Maleimide-functionalized rGO was obtained by mixing DMF dispersion of the rGO with dopa-MAL at room temperature for 3 h using a mass ratio of 2:1 (dopa-MAL/rGO). Fourier transform infrared (FT-IR) spectroscopy (Figure 2A) and X-ray photoelectron spectroscopy (XPS, Figure 2B) were performed for establishing the successful formation of rGO/dopa-MAL. The FT-IR spectrum of rGO/dopa-MAL shows a broad band at  $\sim 3400\text{ cm}^{-1}$  due to the vibration of hydroxyl groups of the dopamine ligand and the remaining OH groups on the rGO. The characteristic vibration band of the maleimide carbonyl group at  $1696\text{ cm}^{-1}$  is clearly observed. The band at  $\sim 1570\text{ cm}^{-1}$  is due to the C=C stretching modes of the aromatic network, whereas the band at  $1223\text{ cm}^{-1}$  is due to the C–O (alkoxy) stretching modes of the rGO and the ligand. The absorption bands at  $2922$  and  $2859\text{ cm}^{-1}$  correspond to the CH and CH<sub>2</sub> stretching vibrations, respectively.





**Figure 2.** (A) FT-IR spectra of rGO/dopa-MAL (black) and dopa-MAL (gray). (B) C<sub>1s</sub> wide-scan XPS spectrum of rGO/dopa-MAL. (C) N<sub>1s</sub> wide-scan XPS spectrum of rGO/dopa-MAL. (D) UV-vis of rGO/dopa-MAL.

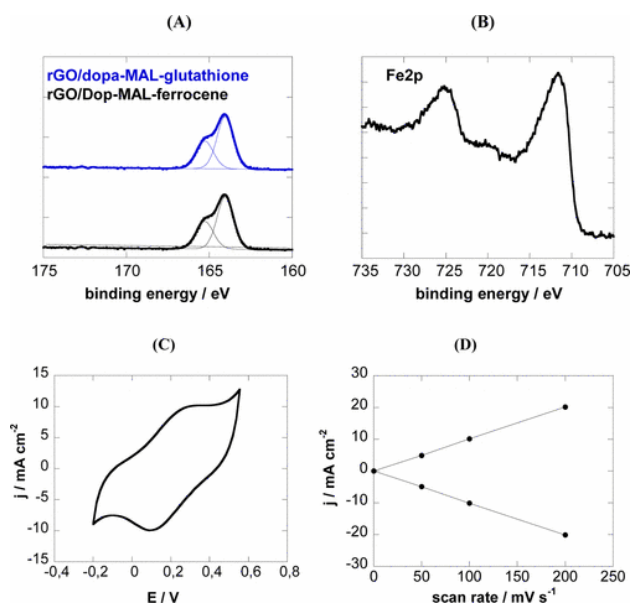
X-ray photoelectron spectroscopy (XPS) analysis was performed in addition to gain additional information on the rGO/dopa-MAL chemical composition (Figure 2B). The XPS survey spectrum displays bands due to C<sub>1s</sub>, O<sub>1s</sub>, and N<sub>1s</sub> (Table 1), in good agreement with the composition of the functionalized rGO. The C<sub>1s</sub> high-resolution spectrum can be deconvoluted into bands at 284.5 eV (C–C sp<sup>2</sup>), 285.0 eV (C–H/C–C), 286.1 eV (C–O), 288.2 eV (N–C=O, C=O), and 290.3 eV (COOH). The presence of N<sub>1s</sub> is attributed to the incorporation of maleimide onto the rGO sheets. The band at 400.2 eV in the N<sub>1s</sub> high resolution (Figure 2C) corresponds to the N–C=O bonds in the rGO/dopa-MAL nanocomposite. The UV-vis spectrum of a water dispersion of rGO/dopa-MAL (Figure 2D) exhibits an absorption maximum at 269 nm, characteristic of the sp<sup>2</sup> hybridized structure together with a prolonged absorption tail in the NIR.

**Table 1. Determination of the Atomic Percentages of Elements by the XPS Analysis**

composite	C <sub>1s</sub> atom %	O <sub>1s</sub> atom %	N <sub>1s</sub> atom %	S <sub>2p</sub> atom %	Fe <sub>2p</sub> atom %
rGO/dopa-MAL	81.84	14.86	3.30	0	0
rGO/dopa-MAL-glutathione	81.03	11.17	6.6	1.2	0
rGO/dopa-MAL-ferrocene	81.6	12.00	3.30	1.5	1.6

### 3.2. Thiol–Ene Michael Addition Reaction on rGO/dopa-MAL

The possibility to integrate thiol-containing ligands onto rGO/dopa-MAL was investigated by the conjugation of glutathione and 6-(ferrocenyl)hexanethiol via Michael addition reaction (Scheme 1). Both conjugations were carried out in DMF at room temperature for 6 h, and the attachment was confirmed by XPS and FT-IR spectroscopy. The presence of  $S_{2p}$  indicates the successful linking of glutathione or ferrocene onto the rGO/dopa-MAL nanocomposite (Table 1). The exact binding constant is not determined for the rGO/dopa-MAL system due to its heterogeneous nature. No significant decrease in nitrogen atom content was observed in the XPS analysis of the product obtained after conjugation with ferrocene thiol, thus indicating that the maleimide-containing ligands do not detach from the rGO surface during these steps (Table 1). Figure 3A depicts the high-resolution XPS spectrum of the  $S_{2p}$  core-level spectra of rGO/dopa-MAL-glutathione and rGO/dopa-MAL-ferrocene. Both spectra can be fitted with two peaks at 164.2 eV ( $S_{2p_{3/2}}$ ) and 165.3 eV ( $S_{2p_{1/2}}$ ) due to S–C bond formation with no oxidized sulfur being present, underlining the covalent integration of the thiol-bearing ligands. As a control experiment, the rGO was reacted with glutathione under the same conditions and the product was analyzed by XPS. The absence of  $S_{2p}$  component in the XPS spectrum clearly indicates that glutathione physisorption does not take place (or very limited) under our experimental conditions. The same is true for 6-(ferrocenyl)hexanethiol-modified rGO/dopa-MAL. The presence of  $Fe_{2p}$  at  $\approx 710$  eV shows in addition the integration of ferrocene units onto the matrix (Figure 3B). The  $Fe_{2p}$  high-resolution XPS spectrum comprises two bands at 711.7 and 725.6 eV due to  $Fe_{2p_{3/2}}$  and  $Fe_{2p_{1/2}}$ , respectively, of the Fe moiety in the (III) oxidation state.<sup>(31)</sup>



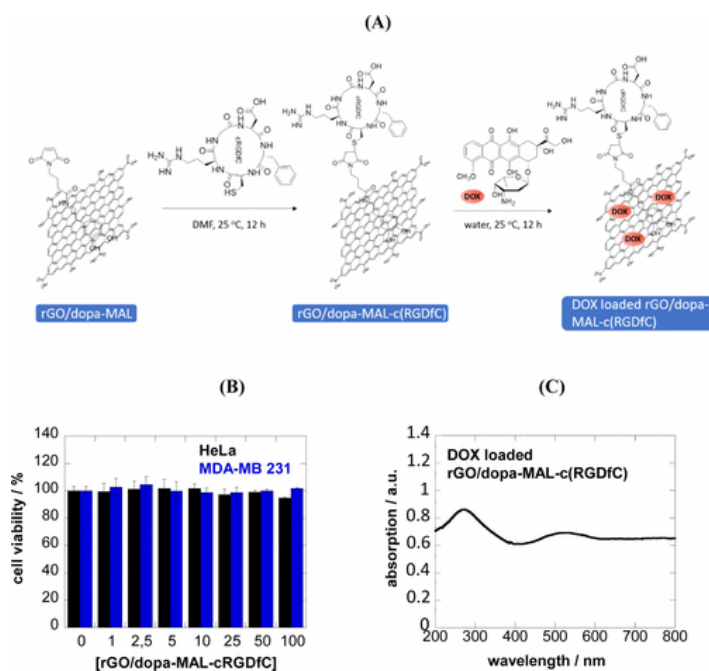
**Figure 3.** (A)  $S_{2p}$  high-resolution XPS spectra of rGO/dopa-MAL-glutathione (blue) and rGO/dopa-MAL-ferrocene (black). (B)  $Fe_{2p}$  high-resolution XPS spectrum of rGO/dopa-MAL-ferrocene. (C) Cyclic voltammogram of rGO/dopa-MAL-ferrocene in tetraethyl ammonium hexafluorophosphate ( $TEAPF_6$ )/DMF (0.1 M), scan rate 100 mV s<sup>-1</sup>. (D) Change in the current density as a function of scan rate.

In addition, the ferrocene-modified rGO/dopa-MAL was probed by cyclic voltammetry (CV) in 0.1 M tetraethyl ammonium hexafluorophosphate (TEAPF<sub>6</sub>) in DMF (Figure 3C). The presence of the redox peak with a peak separation of 164 mV and  $E^0 \approx 0.18$  V versus Ag/AgCl evidences the incorporation of ferrocene groups on the rGO/dopa-MAL. The peak current change recorded as a function of the scan rate suggests that the ferrocene units are immobilized on the surface rather than being adsorbed. Figure 3D shows that the anodic and cathodic peak currents scale linearly with the scan rate, indicating a surface redox process. The loading of attached ferrocene onto the rGO surface via thiol–ene reaction was calculated according to  $\Gamma = Q_A/nFA$ , where the Faraday constant is represented by  $F$ ,  $n$  is the number of exchanged electrons ( $n = 1$ ),  $A$  is the area of the surface ( $A = 0.12$  cm<sup>2</sup>), and  $Q_A$  is the transferred anodic charge. A surface coverage of  $\Gamma = 4.5 \times 10^{-10}$  mol cm<sup>-2</sup> ( $2.71 \times 10^{14}$  molecules cm<sup>-2</sup>) is determined, being similar to that determined for the ferrocene “clicked” to diamond interfaces(31) and 6-(ferrocenyl)hexanethiol immobilized on rGO/alkynyl–dopamine through thiol–yne reaction.(16)

### 3.3. Fabrication of Multifunctional Scaffold: Doxorubicin-Loaded rGO/dopa-MAL-c(RGDfC)

After the successful demonstration of facile covalent functionalization of the maleimide-containing rGO, this scaffold was investigated for its potential to target cancer cells by covalently linking to the maleimide-functionalized rGO a cyclic peptide containing the sequence c(RGDfC), known to target the integrin receptors that are often overexpressed in various types of cancer (Figure 4A). Attachment of the thiol-containing peptide was undertaken in DMF under ambient conditions, followed by dialysis against water to remove any unbound peptide. The peptide attachment was confirmed by using the BCA protein assay kit. The amount of attached peptide on rGO/dopa-MAL was determined to be 263  $\mu\text{g mg}^{-1}$  by using the calibration plot obtained from the UV absorbance data of the peptide solution standards at 562 nm (see SI, Figure S2). The cytotoxicity of rGO/dopa-MAL-c(RGDfC) was assessed using the Uptiblu assay on the human breast MDA-MB-231 and human cervical HeLa cancer cells (Figure 4B). The rGO/dopa-MAL-c(RGDfC) material does not show damage to the two cell lines at the concentration of 0–100  $\mu\text{g mL}^{-1}$  after 48 h incubation, illustrating the suitability of the material as a drug delivery platform.

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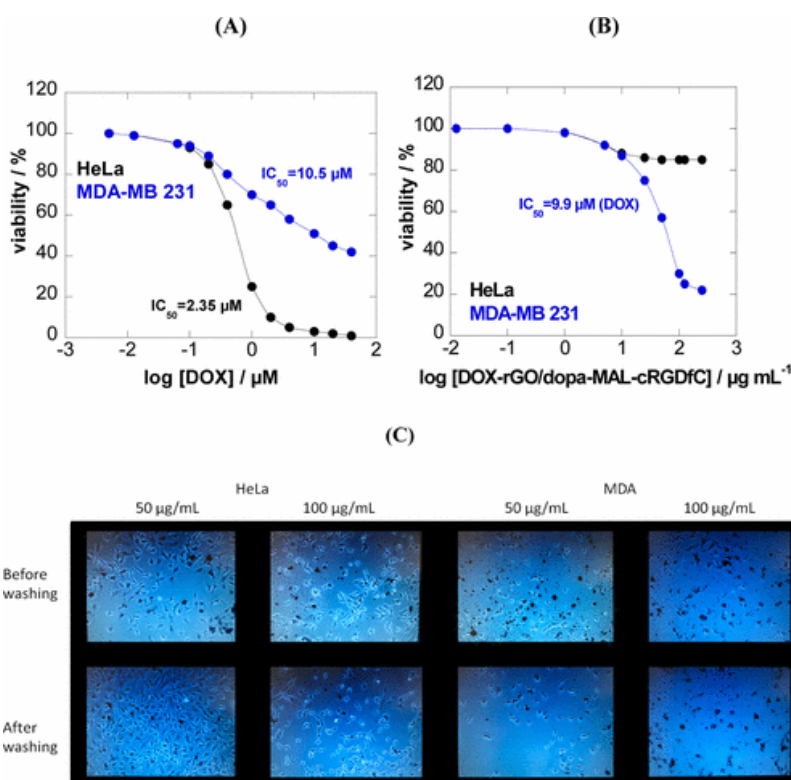


**Figure 4.** Formulation of doxorubicin-loaded rGO/dopa-MAL-c(RGDfC): (A) Schematic of the attachment of targeting peptide to rGO/dopa-MAL via Michael addition followed by the noncovalent integration of doxorubicin. (B) Cell viability of different concentrations ( $\mu\text{g mL}^{-1}$ ) of rGO/dopa-MAL-c(RGDfC) on HeLa and MDA-MB-231 cells after incubation for 48 h. (C) UV-vis spectrum of the DOX-loaded rGO/dopa-MAL-c(RGDfC).

The chemotherapy drug doxorubicin (DOX) was next loaded onto the rGO/dopa-MAL-c(RGDfC) nanosheets by exploiting the various interactions ( $\pi$ -stacking, H-bonding, hydrophobic) between DOX and rGO. The loading was carried out in water in the dark, and the unbound drug was removed through centrifugation of the dispersion at 13 000 rpm (Figure 4A). The successful loading of DOX on rGO/dopa-MAL-c(RGDfC) was evidenced by the reddish color of the solution attributed to DOX showing an absorption peak at 480 nm (Figure 4C). The amount of DOX loaded on the matrix was determined by monitoring the UV-vis absorbance of the supernatant (see SI, Figure S3) and determined to be as 170  $\mu\text{g}$  DOX loaded onto 1 mg of rGO/dopa-MAL-c(RGDfC).

### 3.4. Targeted Cell-Killing Ability of DOX-Loaded rGO/dopa-MAL-c(RGDfC)

We next evaluated the cell-killing ability of DOX-loaded rGO/dopa-MAL-c(RGDfC) on the human cervical HeLa and the human breast MDA-MB-231 cancer cells at pH 7.4. The HeLa cells were used as a negative control, as the nanostructures are designed to target essentially the MDA-MB-231 cancer cell line. The dose-response curves of the free DOX for HeLa and MDA-MB-231 cancer cells (Figure 5A) allow to determine the  $\text{IC}_{50}$  values of 2.35 and 10.5  $\mu\text{M}$  for HeLa and MDA-MB-231 lines, respectively. It is, thus, clear that DOX is much less effective on the MDA-MB-231 cell line than on the HeLa and will preferentially kill HeLa cells.

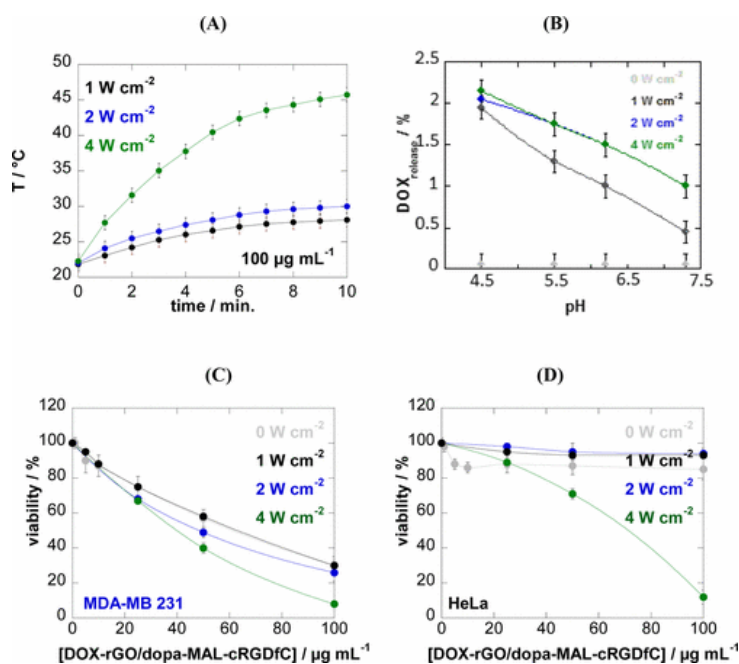


**Figure 5.** Dose–response curves of HeLa (black) and MDA-MB-231 (blue) cancer cells to various concentrations of (A) free DOX for 24 h and (B) DOX-loaded rGO/dopa-MAL-c(RGDfC) nanostructures for 48 h. (C) Optical images showing the targeting effect of DOX-loaded rGO/dopa-MAL-c(RGDfC) at two different concentrations (50 and 100  $\mu\text{g mL}^{-1}$ ) after 6 h incubation before and after washing with PBS.

We next evaluated the viability of HeLa and MDA-MB-231 cells upon the incubation with DOX-loaded rGO/dopa-MAL-c(RGDfC) nanostructure for 48 h. [Figure 5B](#) suggests that under physiological pH, the nanostructure does not show any cell-killing ability for the HeLa cells. However, 100  $\mu\text{g mL}^{-1}$  of rGO/dopa-MAL-c(RGDfC) (corresponding to 17  $\mu\text{g DOX}$ ) induced cell killing with an estimated  $\text{IC}_{50} = 58 \mu\text{g mL}^{-1}$  for DOX-loaded rGO/dopa-MAL-c(RGDfC). Considering that 17  $\mu\text{g DOX}$  is integrated onto 100  $\mu\text{g}$  of the rGO nanosheets, this corresponds to an  $\text{IC}_{50} = 9.9 \mu\text{g mL}^{-1}$  for DOX in the matrix, comparable to the  $\text{IC}_{50} = 10.5 \mu\text{M}$  of free DOX for the same cell line ([Figure 5A](#)). The targeting effect is even more clearly pronounced, as HeLa cells are more sensitive to DOX with free DOX showing a  $\approx 4$  times lower  $\text{IC}_{50}$  value when compared with the MDA-MB-231 cells. The optical images of both cell lines after treatment with DOX-loaded rGO/dopa-MAL-c(RGDfC) (100  $\mu\text{g mL}^{-1}$ ) ([Figure 5C](#)) demonstrate in addition the targeting effect of DOX-loaded rGO/dopa-MAL-c(RGDfC). The cell morphology of HeLa cells is preserved, whereas that of MDA-MB-231 cells is compromised. Interestingly, most DOX molecules remained attached to the rGO/dopa-MAL-c(RGDfC) nanostructure and only  $<1\%$  of DOX was released from the structures after 24 h incubation in the physiological solution with the same ionic strength but different pH values. We were thus intrigued to see if the photothermal properties of the rGO/dopa-MAL-c(RGDfC) nanosheets can further improve the cell-killing ability without losing its targeting benefit.

### 3.5. Targeted Photothermal Cell-Killing Ability of DOX-Loaded rGO/dopa-MAL-c(RGDfC)

Graphene nanosheets, due to their good photothermal properties together with excellent loading capacity of water-insoluble cancer drugs(10) and drugs with aromatic structures(7) such as DOX, are considered as an ideal nanomaterial for the development of combined therapeutic approaches.(32, 33) Particularly, the photothermal properties are appealing, as the generated heat will result in the photothermal ablation of cancer cells, which is known to increase the sensitivity of chemotherapy.(34) Figure 6A depicts the heat profiles of different concentrations of DOX-loaded rGO/dopa-MAL-c(RGDfC) solutions upon irradiation for 10 min with a continuous NIR laser at 980 nm at various laser power densities. The temperature increase is dependent on concentration and laser power, with a  $100 \mu\text{g mL}^{-1}$  DOX-loaded rGO/dopa-MAL-c(RGDfC) heating from 22 to  $45 \text{ }^\circ\text{C}$  in 10 min using a laser power of  $4 \text{ W cm}^{-2}$ , being sufficient to induce cancer cell death. In addition, the generated heat gradient changes the affinity of DOX to rGO/dopa-MAL-c(RGDfC). As seen from Figure 6B, a pH- and laser-power-dependent DOX release takes place. The release is rather moderate from  $0.17 \mu\text{g mL}^{-1}$  DOX (no light) to  $2.1 \mu\text{g mL}^{-1}$  upon illumination at  $4 \text{ W cm}^{-2}$ . Looking at cell viability upon light irradiation (Figure 6C,D), it becomes additionally clear that the laser power densities of  $4 \text{ W cm}^{-2}$  override the targeting approach, resulting in nontargeted cell killing. With a laser power of  $2 \text{ W cm}^{-2}$ , the DOX-loaded rGO/dopa-MAL-c(RGDfC) shows a somewhat improved  $\text{IC}_{50} = 48 \mu\text{g mL}^{-1}$  for MDA-MB-231 cells, corresponding to DOX  $\text{IC}_{50} = 8.2 \mu\text{g mL}^{-1}$  and keeping a complete targeting capability.



**Figure 6.** (A) Photothermal heating profiles of DOX-loaded rGO/dopa-MAL-cRGDfC ( $100 \mu\text{g mL}^{-1}$ ) using a 980 nm laser at different laser power densities. (B) DOX amount released from the DOX-loaded rGO/dopa-MAL-cRGDfC ( $17 \mu\text{g DOX}$  for  $100 \mu\text{g rGO/dopa-MAL-cRGDfC}$ ) matrix. (C) Cell viability of MDA-MB-231 cells after irradiation for 10 min at 980 nm with different laser power densities after 6 h incubation and then additional 18 h incubation. (D) Cell viability of HeLa cells after irradiation at 980 nm for 10 min at different laser power densities after 6 h incubation and then additional 18 h incubation.

## Conclusions

In conclusion, we have demonstrated the interest of a maleimide-containing catechol (dopa-MAL) ligand as a versatile surface anchor onto the rGO nanosheets. The rGO/dopa-MAL-modified nanosheets can be easily modified with the thiol-containing molecules under ambient metal-free conditions. We could show that this approach is not only effective for model molecules such as glutathione and ferrocene containing a thiol group, but it is also adaptable to the covalent linking of thiol-containing tripeptides, such as the cancer cell targeting peptide c(RGDfC). The specific cell targeting and ablation properties of DOX-loaded rGO/dopa-MAL-c(RGDfC) nanosheets was confirmed through the cell viability of human breast MDA-MB-231 adenocarcinoma cells after 48 h incubation with the nanostructures and of the human cervical HeLa cancer cells as the negative control. The HeLa cells remained unaffected by even high concentrations of DOX-loaded rGO/dopa-MAL-c(RGDfC). The nanostructures were, however, effective on MDA-MB-231 cells with an  $IC_{50} = 58 \mu\text{g mL}^{-1}$  for the DOX-loaded rGO/dopa-MAL-c(RGDfC), corresponding to an  $IC_{50} = 9.9 \mu\text{g mL}^{-1}$  for DOX in the matrix. Slight improvements were achieved upon 980 nm irradiation of the DOX-loaded rGO/dopa-MAL-c(RGDfC) in the presence of MDA-MB-231 cells for 10 min at  $2 \text{ W cm}^{-2}$ , whereas high laser power densities proved detrimental for the targeting ability of the c(RGDfC) peptide-containing construct.

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