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SurA Is Involved in the Targeting to the Outer Membrane of a Tat Signal Sequence-Anchored Protein

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The twin arginine translocation (Tat) pathway exports folded proteins from the cytoplasm to the periplasm of bacteria. The targeting of the exported proteins to the Tat pathway relies on a specific amino-terminal signal sequence, which is cleaved after exportation. In the phytopathogen *Dickeya dadantii*, the pectin lyase homologue PnlH is exported by the Tat pathway without cleavage of its signal sequence, which anchors PnlH into the outer membrane. In proteobacteria, the vast majority of outer membrane proteins consists of β -barrel proteins and lipoproteins. Thus, PnlH represents a new kind of outer membrane protein. In *Escherichia coli*, periplasmic chaperones SurA, Skp, and DegP work together with the β -barrel assembly machinery (Bam) to target and insert β -barrel proteins into the outer membrane. In this work, we showed that SurA is required for an efficient targeting of PnlH to the outer membrane. Moreover, we were able to detect an *in vitro* interaction between SurA and the PnlH signal sequence. Since the PnlH signal sequence contains a highly hydrophobic region, we propose that SurA protects it from the hydrophobic periplasm during targeting of PnlH to the outer membrane. We also studied the nature of the information carried by the PnlH signal sequence responsible for its targeting to the outer membrane after exportation by the Tat system.

Proteobacteria are divided into four compartments: the cytoplasm surrounded by the inner membrane (IM), which along with the outer membrane (OM) delimitates the periplasm. In order to maintain this organization, proteobacteria have evolved different processes allowing the targeting of molecules to the appropriate compartment. OM biogenesis may be the most challenging process. Indeed, OM components, synthesized in the cytoplasm, have to cross the inner membrane and the periplasm before being inserted into the OM (38). The large majority of OM proteins are either lipoproteins, anchored in the OM by an amino-terminal lipid moiety, or outer membrane integral proteins (OMPs) forming a transmembrane β barrel in the OM. Like periplasmic or some secreted proteins, OM proteins are synthesized in the cytoplasm as precursors containing an amino-terminal signal sequence specifically targeting them to either the Sec translocon or the twin arginine translocation (Tat) pathway. The bulk mass of OM proteins is exported to the periplasm in an unfolded state by the Sec translocon. However, a few proteins are folded in the cytoplasm and then targeted to the Tat translocon by a particular signal sequence containing two arginine residues in their N-terminal part (11, 32).

After exportation, OMP precursors are matured by the cleavage of their signal sequence by the signal peptidase LepB (34). Due to their propensity to form a β barrel, OMPs are prone to aggregation and have to be kept in a folding competent state during the transport from the exit of the Sec translocon to the OM. This process relies on three periplasmic proteins: (i) SurA, a chaperone that also has two peptidyl-prolyl isomerase domains and that is the main OMP-targeting factor in the periplasm of *Escherichia coli* (5, 39, 48); (ii) DegP, a multimeric protein that has both a chaperone and a protease activity depending on its oligomerization state (49); and (iii) Skp, a chaperone that has been described to bind to unfolded OMPs to protect them from aggregation (54). In *E. coli*, Skp and DegP form a rescue pathway that deals with OMPs falling off the SurA main pathway (39, 48). However, whichever periplasmic chaperone is involved in OMP transport, the last step of OMP biogenesis, folding and insertion into the OM, is catalyzed

by the β -barrel assembly machinery (Bam) complex (56). This complex of five OM proteins consists of a widely conserved OMP from the Omp85 family, BamA, and four lipoproteins, BamB, BamC, BamD, and BamE (47). Interestingly, the Bam complex has been shown to catalyze *in vitro* OMP folding and insertion into proteoliposomes in the presence of SurA, reinforcing the idea of a SurA main pathway (13). Since OMPs are not fully folded before insertion into the OM, the signal allowing their targeting to the OM should reside in the linear sequence of the protein. Consistent with this idea, the periplasmic chaperone SurA has been widely described to exhibit a preferential affinity for Ar-X-Ar sequences, where Ar is an aromatic amino acid, which are more common in OMPs than in soluble proteins because of the presence of an aromatic girdle in β barrels (15, 45). Furthermore, it has been suggested that the *E. coli* BamA could recognize OMPs via the 12 carboxy-terminal residues and notably via a carboxy-terminal phenylalanine (40). However, this signal is not the only one allowing the interaction between BamA and OMPs, and it is not conserved among all bacteria (4, 40).

Lipoprotein maturation relies on a signal peptidase (LspA) and on two enzymes involved in lipoprotein acylation (Lgt, Lnt) (14). Together, these three enzymes are responsible for signal sequence cleavage and addition of three acyl chains to the lipoprotein amino-terminal cysteine residue, resulting in the insertion of the lipoprotein lipid moiety within the IM. Lipoproteins lacking an IM retention signal (or Lol avoidance signal) are then extracted from the IM by the ABC transporter LolCDE and transferred to the chaperone LolA to form a water-soluble complex (24, 26, 58). The

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exact nature of the Lol avoidance signal varies from one organism to another but often relies on the nature of the residues following the amino-terminal cysteine. In *E. coli*, the presence of an aspartate residue at position +2 seems to constitute the strongest Lol-CDE avoidance signal (50). Moreover, the presence of three acyl chains on the amino-terminal cysteine is indispensable for interaction with LolCDE and therefore for transfer to LolA (9). Eventually, lipoproteins are transferred in an energy-independent manner from LolA to the OM lipoprotein LolB that is thought to ensure insertion of the lipoprotein lipid moiety into the OM (25, 55).

Recent work in the gammaproteobacterial phytopathogen *Dickeya dadantii* has shown the existence of a third kind of OM protein. The pectin lyase homologue, PnlH, is anchored in the OM by an uncleaved Tat signal sequence (8). Fusion of the PnlH Tat signal sequence to various proteins allows targeting of these fusion proteins to the outer membrane. This indicates that, contrary to other OM proteins, all the information required for PnlH targeting to the OM relies in its signal sequence. Moreover, PnlH does not share any similarity (transmembrane β barrel or amino-terminal acylation) with OMPs or lipoproteins. Taken together, these data suggest the existence of a new pathway for the targeting of proteins to the OM of *D. dadantii*. Since the expression of PnlH in *E. coli* results in a correct insertion of PnlH into the OM, this pathway should be conserved in other bacteria. The aim of this work was to identify proteins involved in the targeting of this third kind of OM protein. To this end, we analyzed the impact of mutations in the OMP-targeting pathway on the targeting of PnlH to the OM of *D. dadantii*. Surprisingly, we showed that SurA is involved in the targeting of PnlH to the OM. We also studied the nature of the information carried by the PnlH signal sequence.

MATERIALS AND METHODS

Bacterial strains, plasmids, and oligonucleotides. The bacterial strains and plasmids used in this study are described in Table 1. Oligonucleotides are listed in Table S1 in the supplemental material. *D. dadantii* and *E. coli* were grown at 30°C and 37°C, respectively, in LB medium. When required, the following antibiotics were added at the indicated concentrations: ampicillin, 100 μ g/ml; kanamycin, 25 μ g/ml; and chloramphenicol, 25 μ g/ml and 5 μ g/ml for *E. coli* and *D. dadantii*, respectively.

To construct strains A5275, A5276, A5281, A5476, A5477, and A5478, DNA fragments containing the genes *bamE*, *bamC*, *bamB*, *degP*, *skp*, and *surA*, respectively, were amplified with the corresponding primers. The resulting DNA fragments were inserted into the pGEM-T plasmid. The Kan^r cassette was extracted from pHP45 Ω Kan and inserted into the different coding sequences (CDSs), except for *skp*, for which the *uidA*::Kan^r cassette of pL247 was prepared and inserted into its CDS. All the resulting constructs were inserted into the *D. dadantii* A350 chromosome by recombination in low-phosphate medium (41). Recombinations were checked by PCR. Before phenotype analysis, all mutations were transduced into a fresh A350 background (37).

To construct plasmid pPnlH, a DNA fragment containing *pnlH* was amplified with the oligonucleotides PnlHHis⁺ and PnlHHis⁻. The resulting DNA was subcloned into pBluescript (pBS) between the SacI and ApaI sites. The resulting plasmid (pBS*pnlH*) was digested by SacI and KpnI, and the DNA fragment containing *pnlH* was inserted into pACT3. To construct plasmids carrying one or more substitutions in the PnlH-6His signal sequence, site-directed mutagenesis with appropriate primers (see Table S1 in the supplemental material) was performed. To construct pPnlHssPema, a DNA fragment containing the CDS of the PnlH signal sequence fused to the mature form of PemaA (PnlHssPemaA) was amplified from the PnlH-PemaA plasmid (8) with PnlHssPema⁺ and Pnl-

HssPema⁻. The PCR product was digested by SacI and KpnI and then inserted into pACT3. To construct pPnlH Δ 28-41, site-directed mutagenesis with primers Δ 28-41⁺ and Δ 28-41⁻ and pPnlH as the template was performed to add an extra HindIII site at nucleotide 123 of the *pnlH* CDS. The resulting DNA was digested by HindIII and recircularized to remove the DNA fragment between the two HindIII sites. To construct pSurA, the SurAStrep⁻ primer was designed to add a Strep tag at the carboxy terminus of SurA, and a DNA fragment containing *surA* was amplified with SurAStrep⁺ and SurAStrep⁻. The PCR product was inserted into pGEM-T. Insertions allowing expression of SurA with the carboxy-terminal Strep tag under the control of the pGEM-T *lac* promoter were selected. To construct pXss20, the sequence of the PnlH signal sequence was amplified with Xss20⁺ and Xss20⁻. The PCR product was inserted in frame into pGEX-6P3 between the BamHI and SalI sites downstream of the glutathione *S*-transferase (GST)-coding sequence. All constructions were checked by sequencing.

Drug sensitivity assay. The sensitivity to EDTA, SDS, and rifampin of different mutant strains was determined by disk diffusion assay. A 0.1-ml sample from an LB overnight culture was mixed with 3 ml of molten LB top agar and poured over an LB agar plate. Three 6-mm filter paper disks (Dominique Dutscher) were put on the top of molten agar, and 7.5 μ l of EDTA (0.5 M), SDS (10%, wt/wt), or rifampin (20 mg/ml) was added onto a disk. The diameters of the growth inhibition zones were measured after 16 h.

Cell fractionation. To prepare the periplasm, cells were grown overnight at 30°C in LB medium complemented with the appropriate antibiotics. Cells were harvested by centrifugation for 7 min at 5,000 \times g, and the periplasm was prepared by lysozyme treatment and osmotic shock as previously described (2). Whole-cell fractions from steady-state cultures or exponential-phase cultures were obtained by concentrating cells up to an optical density at 600 nm (OD₆₀₀) of 7.5. To prepare membranes, cells from exponential-phase cultures were harvested by centrifugation for 7 min at 5,000 \times g, concentrated up to an OD₆₀₀ of 7.5 in TE (10 mM Tris-HCl, pH 8.0, 1 mM EDTA), 1 mM phenylmethylsulfonyl fluoride (PMSF), and broken with a French press. Debris was removed by centrifugation for 20 min at 7,000 \times g, and the supernatant was ultracentrifuged for 1 h at 100,000 \times g. The supernatant and the pellet were taken as the soluble fraction and membrane fraction, respectively. When necessary, membranes were separated by centrifugation for 48 h at 200,000 \times g on a 38 to 65% sucrose flotation gradient as previously described (46). Fractions were collected from the bottom of the tube and precipitated with ethanol.

To analyze the composition of the different fractions, proteins were separated by SDS-PAGE and transferred onto a PVDF membrane (Millipore). PnlH, KdgR, PemaA, BlaM, and TolA were detected by immunodetection. Anti-PnlH, anti-KdgR, anti-PemaA, anti-BlaM, and anti-TolA antibodies were used at dilutions of 1/3,000, 1/5,000, 1/5,000, 1/3,000, and 1/10,000, respectively. The position of the inner membrane in the sucrose gradient was detected by NADH oxidase activity assay (31) or immunoblotting with TolA antibodies; the position of porins was detected by staining the immunoblot membrane with Coomassie blue.

In vitro interactions with SurA-Strep. To prepare PnlH-6His inclusion bodies, NM522/pPnlH cells were grown at 30°C. At an OD₆₀₀ of 0.6, isopropyl- β -D-thiogalactopyranoside (IPTG) was added at a final concentration of 1 mM and cells were grown for an extra 2 h. Cells were harvested by centrifugation for 7 min at 5,000 \times g, concentrated up to an OD₆₀₀ of 7.5 in TE, 1 mM PMSF, and broken with a French press. The lysate was centrifuged for 20 min at 7,000 \times g, and PnlH inclusion bodies were harvested in the pellet. To get pure GST and derivatives, NM522/pGEX-6P3 or NM522/pXSS20 cells were grown at 30°C. At an OD₆₀₀ of 0.5 mM, IPTG was added at a final concentration of 1 mM and cells were grown for an extra 2 h. Cells were harvested by centrifugation for 7 min at 5,000 \times g, concentrated up to an OD₆₀₀ of 7.5 in TE, 1 mM PMSF, and broken with a French press. Triton X-100 was added at a final concentration of 1% (vol/vol). The lysate was centrifuged for 20 min at 7,000 \times g to

TABLE 1 Strains and plasmids used in this study

Strain or plasmid	Relevant characteristics	Source or reference
Strains		
<i>D. dadantii</i> 3937 derivatives		
A350	<i>rafR ganB</i>	17
A5275	<i>rafR ganB bamE::Kan^r</i>	This study
A5276	<i>rafR ganB bamC::Kan^r</i>	This study
A5281	<i>rafR ganB bamB::Kan^r</i>	This study
A5476	<i>rafR ganB degP::Kan^r</i>	This study
A5477	<i>rafR ganB skp::uidA Kan^r</i>	This study
A5478	<i>rafR ganB surA::Kan^r</i>	This study
<i>Escherichia coli</i>		
NM522	F' <i>proAB lacI^q Δ(lacZ)M15 Δ(lac-proAB) thi hsd-5</i>	Stratagene
SR1458	F– <i>araD139 Δ(argF-lac)U169 φ(PdegP-lacZ)</i>	36
Plasmids		
pGEM-T	Amp ^r ; allows the AT cloning of a PCR product	Promega
pBluescript	pUC <i>ori</i> Amp ^r	Stratagene
pACT3	pACYC <i>ori</i> Cm ^r	23
pL247	Amp ^r <i>uidA::Kan^r</i> cassette	Laboratory collection
pHP45ΩKan	pMB1 <i>ori</i> Amp ^r Kan ^r cassette	7
pGEX-6P3	pBR322 <i>ori</i> Amp ^r , codes for GST	GE Healthcare
pGEM-T <i>surA</i>	Contains the DNA fragment amplified with SurA+ and SurA–	This study
pGEM-T <i>surA::Kan^r</i>	pHP45ΩKan Kan ^r cassette in the HindIII site of pGEM-T <i>surA</i>	This study
pGEM-T <i>skp</i>	Contains the DNA fragment amplified with Skp+ and Skp–	This study
pGEM-T <i>skp::Kan^r</i>	pL247 <i>uidA::Kan^r</i> cassette in the PstI site of pGEM-T <i>skp</i> ΔPstI	This study
pGEM-T <i>degP</i>	Contains the DNA fragment amplified with DegP+ and DegP–	This study
pGEM-T <i>degP::Kan^r</i>	pHP45ΩKan Kan ^r cassette in the MunI site of pGEM-T <i>degP</i>	This study
pGEM-T <i>bamB</i>	Contains the DNA fragment amplified with BamB+ and BamB–	This study
pGEM-T <i>bamB::kan^r</i>	pHP45ΩKan Kan ^r cassette in the BclI site of pGEM-T <i>bamB</i>	This study
pGEM-T <i>bamC</i>	Contains the DNA fragment amplified with BamC+ and BamC–	This study
pGEM-T <i>bamC::kan^r</i>	pHP45ΩKan Kan ^r cassette in the BclI site of pGEM-T <i>bamC</i>	This study
pGEM-T <i>bamE</i>	Contains the DNA fragment amplified with BamE+ and BamE–	This study
pGEM-T <i>bamE::kan^r</i>	pHP45ΩKan Kan ^r cassette between BamHI sites of pGEM-T <i>bamE</i>	This study
pGEM-T <i>pnIH</i>	Contains the DNA fragment amplified with PnlHHis+ and PnlHHis–	This study
pBS <i>pnIH</i>	Insertion of pGEM-T <i>pnIH</i> Apal SacI DNA fragment containing <i>pnIH</i> CDS in pBS	This study
pPnlH	pACT3 containing the gene of PnlH with a C-terminal His tag between SacI and KpnI sites	This study
PnlH-PemA	pBAD ₁₈ containing a fusion of the PnlH signal sequence with the mature form of PemA	8
pPnlHssPemA	pACT3 containing a fusion of the PnlH signal sequence with the mature form of PemA	This study
pD16A	pPnlH containing a D16A substitution in PnlH	This study
pL30SL31T	pPnlH containing L30SL31T substitutions in PnlH	This study
pP32L	pPnlH containing a P32L substitution in PnlH	This study
pF34A	pPnlH containing a F34A substitution in PnlH	This study
pP35A	pPnlH containing a P35A substitution in PnlH	This study
pP32LP35A	pPnlH containing P32LP35A substitutions in PnlH	This study
pY36A	pPnlH containing a Y36A substitution in PnlH	This study
pQ41A	pPnlH containing a Q41A substitution in PnlH	This study
pPnlHA28-41	pPnlH containing a deletion of nucleotides 81 to 123	This study
pSurA	pGEM-T containing SurA with a C-terminal Strep tag under the control of the <i>lac</i> promoter	This study
pXss20	pGEX-6P3 containing of a fusion of GST with the first 41 amino acids of PnlH	This study

removed debris, and the supernatant was loaded onto Protino glutathione agarose 4B (Macherey-Nagel) resin previously equilibrated with TES (50 mM Tris-HCl, pH 7.4, 1 mM EDTA, 100 mM NaCl), 1% (vol/vol) Triton X-100. The resin was then washed 3 times with TES, 1% (vol/vol) Triton X-100. GST and derivatives were eluted with 2× Laemmli buffer. For overlay experiments, samples containing proteins of interest were run on an SDS-polyacrylamide gel and transferred onto a PVDF membrane. The membrane was saturated with 4% gelatin in phosphate-buffered saline (PBS; 81 mM Na₂HPO₄, 19 mM NaH₂PO₄, 150 mM NaCl, pH 7.4). As after all following steps, the membrane was washed 3 times for 5 min each time with PBS containing 1% gelatin (PBS-G). After incubation for 45 min at room temperature with 8.5 ml of PBS-G complemented with 1.5

ml of periplasm containing or not SurA-Strep, the membrane was washed and finally incubated for 45 min with streptavidin conjugated with horseradish peroxidase (HRP; Pierce) at a dilution of 1/30,000 in PBS-G. After the membrane was washed, an Immobilion Western ECL kit (Millipore) was used to reveal the membrane. In order to compare two conditions of overlay, samples were loaded twice on a gel, the resulting membrane was cut in two, and both parts were treated in parallel.

β-Galactosidase assays. Cells from overnight cultures were diluted to an OD₆₀₀ of approximately 0.02 and grown at 33°C to an OD₆₀₀ of 0.5 in LB medium supplemented with the appropriate antibiotic and 1 mM IPTG. Growth was stopped by adding toluene, and β-galactosidase activity was assayed on permeabilized cells as previously de-

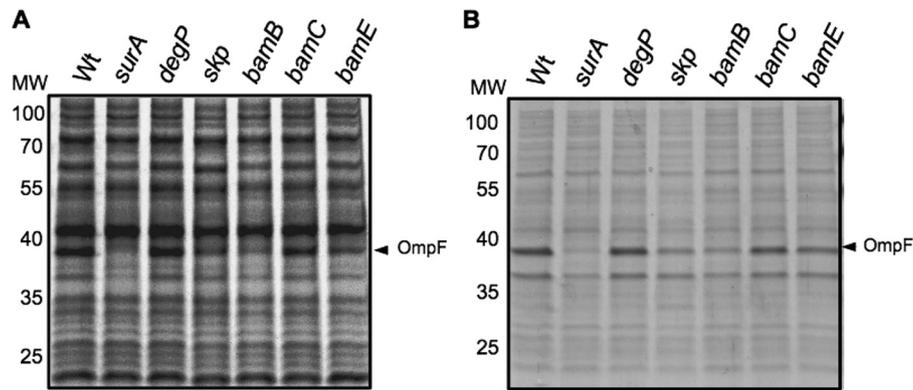


FIG 1 Porin depletion in *D. dadantii* mutants with mutations in the OMP-targeting pathway. Protein profiles of whole cells of *D. dadantii* wild-type (Wt) strain A350 and derivative strains (indicated at the top) carrying the chromosomal mutation (A) and protein profiles of membrane fractions of the same strains (B). Equivalent amounts of cell materials from exponential-phase cultures of the different strains were separated by SDS-PAGE, followed by Coomassie blue staining. The arrowhead indicates the *D. dadantii* major porin OmpF. MW, molecular weight (in thousands).

scribed (28). Three independent replications were averaged to obtain the indicated values.

RESULTS

Analysis of *D. dadantii* mutants in the OMP-targeting pathway.

In *E. coli*, the OMP-targeting pathway mainly relies on the three periplasmic chaperones SurA, Skp, and DegP and on the OM complex BamABCDE. The *D. dadantii* genome contains homologues of *E. coli* *surA*, *skp*, *degP*, *bamA*, *bamB*, *bamC*, *bamD*, and *bamE* encoded by the loci ADN00090, ADM97288, ADM99598, ADM97287, ADM99516, ADM97479, ADM99647, and ADM97110, respectively. Notably, all these loci were found in a genetic context similar to that in *E. coli*. Since *bamA* and *bamD* mutations are lethal in *E. coli*, these genes were excluded from our analysis (22, 51). *D. dadantii* *surA*, *skp*, *degP*, *bamB*, *bamC*, and *bamE* mutant strains were constructed by insertion of a kanamycin resistance cassette. Mutant strains did not exhibit obvious growth defects, indicating that, as in *E. coli*, none of these genes is essential in *D. dadantii*. To check the effect of these mutations on OMP biogenesis, an analysis of the protein profiles of whole-cell and membrane fractions of the corresponding strains was performed (Fig. 1). Compared to the wild-type strain, the *surA* mutant exhibited a strong decrease in the quantity of the major porin OmpF in both whole-cell and membrane fractions. This decrease was also observed to a lower extent for the *skp* and *bamB* mutants. This indicates that the OMP-targeting pathway is at least partially affected in these three mutants. Accordingly, *surA*, *skp*, and *bamB* mutants also presented an increased sensitivity to SDS, EDTA, and rifampin (Table 2), indicating that the integrity of their outer membrane is affected (43). The *bamE* mutant presented a weak but reproducible decrease in the quantity of OmpF and an increased sensitivity to EDTA and, to a lower extent, to rifampin, indicating an alteration of its OM biogenesis. Taken together, these data suggest that in *D. dadantii*, as in *E. coli*, SurA is the major periplasmic factor for OMP targeting. Moreover, the relative importance of each component of the Bam complex in OM biogenesis seems to be conserved between *D. dadantii* and *E. coli*.

Efficient targeting of PnlH to the OM depends on SurA but not on Skp and DegP. To determine if the periplasmic chaperones SurA, Skp, and DegP could be involved in PnlH targeting to the OM, we checked PnlH localization in corresponding *D. dadantii*

mutant strains. Since PnlH is not expressed in strain A350 and derivatives under the conditions used in this study (8), we expressed PnlH with a carboxy-terminal 6His tag (PnlH-6His) from a pACT3 plasmid (pPnlH). Additionally, expression of PnlH-6His from this plasmid allowed us to avoid any possible feedback regulatory effect due to the activation of the σ^E stress response, as described for the *E. coli* *surA* mutant (42). Cell and membrane separation experiments showed that PnlH-6His is mostly located in the OM, when expressed in the wild-type *D. dadantii* A350 strain (see Fig. S1 in the supplemental material). To analyze PnlH-6His localization in the different genetic backgrounds, appropriate strains were transformed with the pPnlH plasmid. The amount of PnlH-6His in whole-cell fractions of *skp* and *degP* mutants was equivalent to that for the wild-type strain but was strongly decreased for the *surA* mutant (Fig. 2A). Accordingly, the same diminution was observed in the membrane and soluble fractions of the last mutant (Fig. 2A). This phenomenon was not due to a global protein synthesis/stability decrease, since the level of the IM protein TolA was identical in the membrane fraction of all the tested strains (Fig. 2A). To determine if the small amount of PnlH-6His present in the *surA* mutant membrane fraction was in the IM or the OM, we performed membrane separation on a flotation sucrose gradient. However, we were not able to properly sepa-

TABLE 2 Growth inhibition of *D. dadantii* mutants for the OMP targeting pathway^a

Strain	Characteristic or mutated gene	Inhibition zone diam (mm)		
		SDS (10%)	EDTA (0.5 M)	Rifampin (20 mg/ml)
A350	Wild type	11 ± 0	19 ± 2	16 ± 1
A5275	<i>bamE</i>	14 ± 1	29 ± 1	21 ± 1
A5276	<i>bamC</i>	13 ± 1	20 ± 1	17 ± 1
A5281	<i>bamB</i>	19 ± 1	32 ± 1	22 ± 0
A5476	<i>degP</i>	12 ± 1	24 ± 2	16 ± 1
A5477	<i>skp</i>	18 ± 1	30 ± 1	25 ± 1
A5478	<i>surA</i>	17 ± 0	31 ± 1	22 ± 1

^a Sensitivity to SDS (10%), EDTA (0.5 M), and rifampin (20 mg/ml) was determined by measuring the growth inhibition zone on LB agar plates topped with filter paper disks containing these compounds, as described in Materials and Methods. The values indicated are the means of 3 experiments.

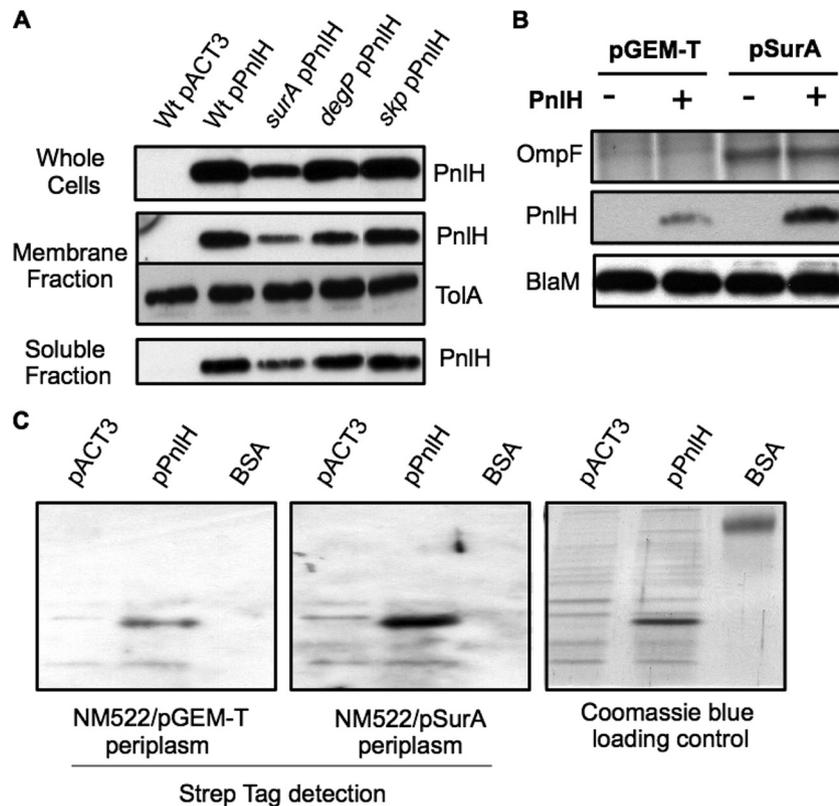


FIG 2 Role of periplasmic chaperones SurA, Skp, and DegP in the targeting of PnlH-6His to the OM of *D. dadantii*. (A) Immunodetection of PnlH in whole-cell, membrane, and soluble fractions of *D. dadantii* wild-type strain and derivative strains carrying a chromosomal mutation (indicated at the top) and either empty pACT3 vector or pPnlH. Equivalent amounts of cell material from exponential-phase cultures of the different strains were loaded in each well. Immunodetection of the inner membrane protein TolA was used as a loading control. (B) Complementation of a *surA* mutant carrying either empty pACT3 vector or pPnlH with either empty pGEM-T vector or pSurA. Equivalent amounts of whole cells of steady-state cultures were analyzed. Proteins were separated on an SDS-polyacrylamide gel and transferred onto a PVDF membrane for immunodetection of PnlH and BlaM (loading control). OmpF was detected by SDS-PAGE, followed by Coomassie blue staining. (C) The *in vitro* interaction between PnlH-6His and SurA-Strep was assayed by overlay. BSA and cell lysates of NM522 carrying either empty pACT3 vector or pPnlH were separated by SDS-PAGE and transferred onto a PVDF membrane. The membrane was next incubated with the periplasm of NM522 carrying either empty pGEM-T vector or pSurA. SurA-Strep bound to proteins on the membrane was detected with streptavidin conjugated with HRP. The amounts of protein loaded on the gel were estimated by Coomassie blue staining.

rate the IM from the OM of the *surA* mutant. This observation is consistent with the decreased OM density described for an *E. coli surA* mutant strain (48). Since *surA* is part of an operon, we tested whether the decrease in PnlH-6His level in the *surA* mutant was due to a polar effect. When SurA carrying a carboxy-terminal Strep tag (SurA-Strep) was expressed from plasmid pSurA, wild-type quantities of OmpF and PnlH-6His were restored in the *surA* mutant (Fig. 2B). These results indicate that the effect of the *surA* mutation on OmpF and PnlH stability is not due to a polar effect.

Our data suggest that SurA could be involved directly or indirectly in the stabilization of PnlH in the periplasm. Overlay experiments have previously been used to demonstrate the interaction of SurA with the passenger domain of the *E. coli* autotransporter EspP (44). We performed such an experiment to assess whether PnlH can interact directly with SurA *in vitro*. To this end, PnlH-6His was immobilized on a PVDF membrane and used as a matrix to detect a possible interaction with SurA-Strep. The membrane was incubated with either the periplasm of an *E. coli* strain expressing SurA-Strep or the periplasm of an *E. coli* control strain. The use of streptavidin conjugated with HRP allowed us to detect SurA-Strep bound on the immobilized PnlH-6His (Fig. 2C). No

interaction between SurA-Strep and the control, bovine serum albumin (BSA), could be detected in this way, indicating that SurA-Strep specifically binds to PnlH-6His. Taken together, these data indicate that SurA is able to bind to PnlH *in vitro*. Moreover, since a *surA* mutation leads to a strong decrease in PnlH stability, it is likely that SurA is involved in the targeting of PnlH to the OM by preventing its periplasmic degradation.

SurA is able to interact with the PnlH signal sequence. SurA is thought to ensure a specific targeting of OMPs by binding to Ar-X-Ar motifs, which are particularly enriched in OMPs (15). To ensure a specific targeting of PnlH to the OM, SurA should also recognize and bind a specific signal within this protein. Since a fusion of the PnlH signal sequence to the mature form of the *D. dadantii* pectin methylesterase Pema (PnlHssPema) is sufficient to target it to the OM of *D. dadantii* (8), we hypothesized that the PnlH signal sequence could contain the SurA recognition site. To test this hypothesis, we examined whether PnlHssPema is dependent on SurA for its targeting to the OM. First, we checked the expression and the localization of PnlHssPema in the wild-type strain A350 carrying the pPnlHssPema plasmid. Antibody directed against Pema detected two bands in the cells producing this fusion (Fig. 3A). After cell fractionation, the upper band (indi-

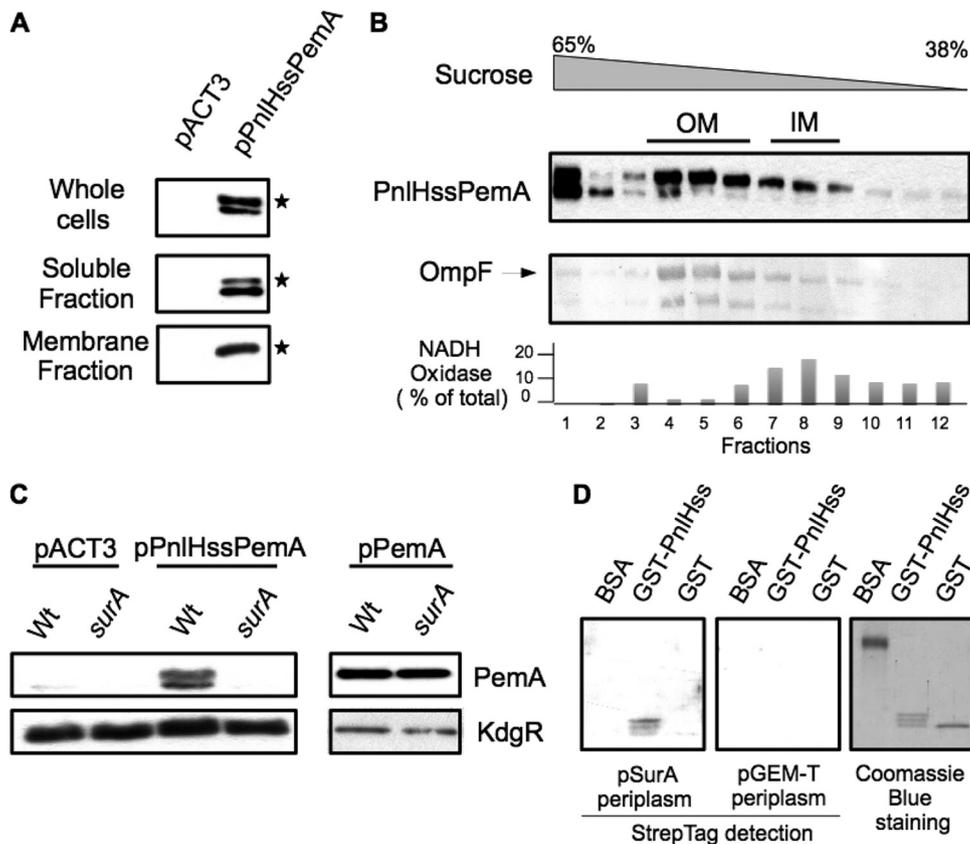


FIG 3 Role of the PnlH signal sequence in the interaction with SurA. (A) Cell fractionation of *D. dadantii* A350 strains carrying either empty pACT3 vector or pPnlHssPemA. PnlHssPemA was detected with antibodies against PemA. Equivalent amounts of cell materials from exponential-phase cultures of the different strains were loaded in each well. (B) The membrane fraction from strain A350 carrying pPnlHssPemA was separated by flotation sucrose gradient centrifugation and analyzed by immunoblotting with PemA antibodies or stained with Coomassie blue to detect the major porin, OmpF, which reflects the position of the outer membrane. NADH oxidase activity indicates the position of the inner membrane. (C) Immunodetection of PemA in whole cells of *D. dadantii* wild-type strain A350 and a *surA* mutant carrying either empty pACT3 vector or pPnlHssPemA. Equivalent amounts of cell material from steady-state cultures were loaded in each well. The cytoplasmic protein KdgR was used as a loading control. (D) The *in vitro* interaction between the PnlH signal sequence and SurA-Strep was assayed by overlay. Purified BSA, GST, and GST fused to the PnlH signal sequence (GST-PnlHss) were run on an SDS-polyacrylamide gel and transferred onto a PVDF membrane. The membrane was next incubated with the periplasm of NM552 carrying either empty pGEM-T vector or pSurA. SurA-Strep bound to proteins on the membrane was detected with streptavidin conjugated with HRP. The amounts of protein loaded on the gel were estimated by Coomassie blue staining.

cated by a star) was mainly found in the membranes. The smallest one was found in the soluble fraction and could be a degradation product. Membrane separation on a sucrose gradient showed that the upper band was associated with the OM, while the lower band remained at the bottom of the gradient (Fig. 3B). Contrary to the wild-type strain, no PnlHssPemA was detected in the *surA* mutant (Fig. 3C), suggesting that its stability was affected in the absence of SurA. Since a *surA* mutation has no effect on the amount of the native PemA expressed from pACT3 (Fig. 3C), we concluded that the PnlH signal sequence is responsible for the degradation of PnlHssPemA in the *surA* mutant and that it could be the part of PnlH that interacts with SurA. Since SurA interacts *in vitro* with full-length PnlH in an overlay experiment, we verified if SurA could bind the PnlH signal sequence. For this purpose, the first 41 amino acids of PnlH were fused to the carboxy-terminal end of the GST (GST-PnlHss). When GST-PnlHss was produced in NM522, it was mostly found in inclusion bodies, indicating a high propensity of the PnlH signal sequence to aggregate. Purification of GST-PnlHss on glutathione agarose generated three forms of the protein (Fig. 3D, right). Apparently, the upper band corresponds to

the full-length GST-PnlHss and the smaller bands correspond to degradation products. Overlay experiments showed interactions between SurA-Strep and the different forms of GST-PnlHss (Fig. 3D). Since a larger amount of SurA-Strep bound to the full-length GST-PnlHss and since SurA-Strep did not bind to GST or BSA, we concluded that the interaction between SurA-Strep and GST-PnlHss was due to the PnlH signal sequence. Altogether, these data suggest that SurA is able to recognize and bind the PnlH signal sequence to protect PnlH from degradation in the periplasm.

The secondary structure of the PnlH signal sequence could be an important factor for the targeting of PnlH to the OM. If it is clear that the PnlH signal sequence is responsible for the targeting of PnlH to the OM, the exact nature of the information carried by this signal sequence remains unknown. The only known homologue of the *D. dadantii* PnlH can be found in *D. zea*, and alignment of the two signal sequences reveals that besides the Tat consensus, the carboxy-terminal part of the signal sequence is the most conserved (Fig. 4A). To check whether this part of the signal sequence could carry a signal for the targeting of PnlH, we con-

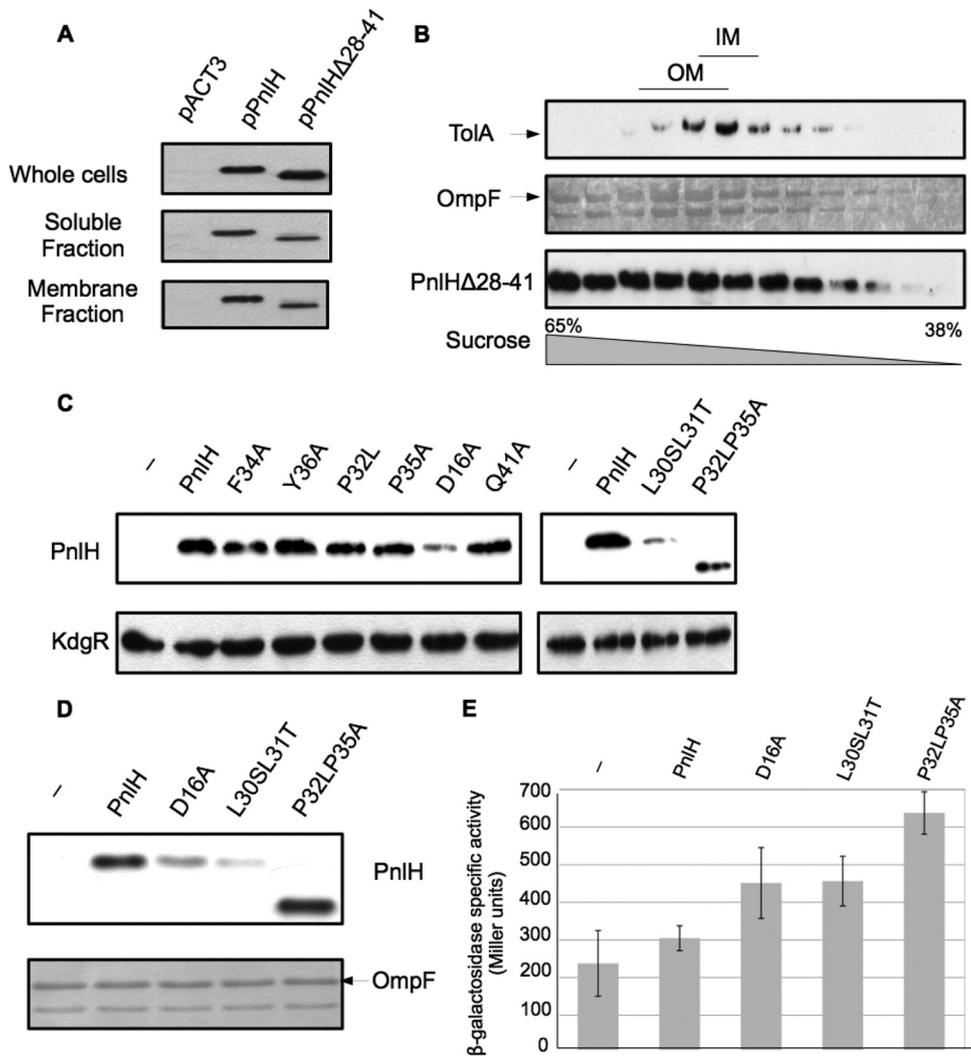


FIG 5 Study of the PnlH signal sequence by directed mutagenesis. (A) Cell fractionation of *D. dadantii* A350 strains carrying either empty pACT3 vector, pPnlH, or pPnlH Δ 28-41. PnlH and PnlH Δ 28-41 were detected with antibodies against PnlH. Equivalent amounts of cell materials from exponential-phase cultures of the different strains were loaded in each well. (B) The membrane fraction from strain A350 carrying pPnlH Δ 28-41 was separated by flotation sucrose gradient centrifugation and analyzed by immunoblotting with Pema antibodies or stained with Coomassie blue to detect the major porin OmpF, which reflects the position of the outer membrane. Immunoblotting with TolA antibodies indicates the position of the inner membrane. (C) Immunodetection of PnlH in whole-cell lysates of *D. dadantii* A350 strains carrying either empty pACT3 vector (-) or pPnlH (PnlH) or pPnlH derivatives carrying one or two substitutions in the PnlH signal sequence. Equivalent amounts of cell material from steady-state cultures were loaded in each well. Immunoblotting with KdgR antibodies was used as a loading control. (D) Membrane fraction of A350 strains carrying either empty pACT3 vector (-), pPnlH (PnlH), pD16A (D16A), pL30SL31T (L30SL31T), or pP32LP35A (P32LP35A). PnlH and derivatives were detected with antibodies against PnlH. Equivalent amounts of membrane fractions from exponential-phase cultures were loaded in each well. OmpF was visualized by the coloration of the PVDF membrane with Coomassie blue and served as a loading control. (E) Induction of periplasmic stress in *E. coli* SR1458 expressing PnlH and derivatives. *E. coli* SR1458 carrying either empty pACT3 vector (-), pPnlH (PnlH), pD16A (D16A), pL30SL31T (L30SL31T), or pP32LP35A (P32LP35A) was grown at 33°C in LB medium supplemented with chloramphenicol and 1 mM IPTG. The value for β -galactosidase specific activity is the average of three independent determinations.

through its carboxy-terminal His tag (see Fig. S2C in the supplemental material), we concluded that this extremity was intact and that the degradation should have occurred at the amino-terminal part of the protein. Expression of PnlHP32LP35A in *E. coli* strain SR1458 provoked a strong increase in the expression of the *degP-lacZ* fusion (approximately 2-fold) (Fig. 5E), indicating that PnlHP32LP35A is able to induce a periplasmic stress response and is therefore exported to the periplasm. Altogether, these data suggest that the double substitution P32L P35A strongly affects the stability of the PnlH signal sequence without impairing its targeting to the OM.

The predicted alpha helix of the PnlH signal sequence is rich in hydrophobic residues (residues 24 to 31). To decrease the global hydrophobicity of the predicted alpha helix, leucines 30 and 31 were replaced by serine and threonine (PnlHL30SL31T). When PnlHL30SL31T was expressed in the *D. dadantii* A350 strain, it was barely detectable in either the whole-cell (Fig. 5C) or the membrane (Fig. 5D) fraction. Moreover, expression of PnlHL30SL31T in *E. coli* strain SR1458 provoked a significant increase in β -galactosidase activity (approximately 1.5-fold) (Fig. 5E), indicating that PnlHL30SL31T is able to induce a periplasmic stress response. These data suggest that the reduction of the hydrophobicity in the

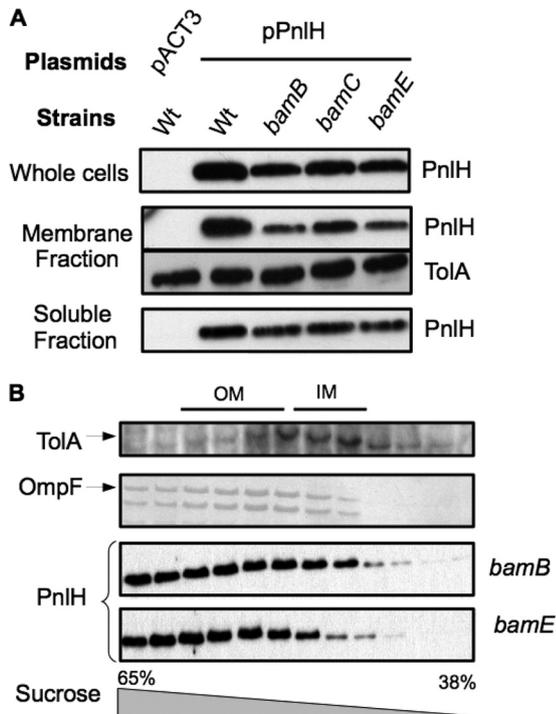


FIG 6 Effect of *bamB*, *bamC*, and *bamE* mutations on the targeting of PnlH-6His to the OM of *D. dadantii*. (A) Immunodetection of PnlH in whole-cell, membrane, and soluble fractions of *D. dadantii* wild-type strain A350 and derivative strains carrying chromosomal mutations (indicated at the top) and either empty pACT3 vector or plasmid pPnlH. Equivalent amounts of cell materials from exponential-phase cultures of the different strains were loaded in each well. Immunodetection of the inner membrane protein TolA was used as a loading control. (B) The membrane fractions from the *bamB* and *bamE* mutant strains carrying pPnlH were separated by flotation sucrose gradient centrifugation and analyzed by immunoblotting with PnlH antibodies or stained with Coomassie blue to detect the major porins, which reflect the position of the outer membrane. Immunoblotting with TolA antibodies indicates the position of the inner membrane. For clarity, porins and TolA profiles are presented only for the *bamB* mutant, but the profiles were equivalent for the *bamE* mutant.

predicted alpha helix of the PnlH signal sequence significantly impairs the stability of the protein and could alter its targeting to the OM.

***bamB* and *bamE* null mutations affect the stability of PnlH in *D. dadantii*.** By interacting with SurA, PnlH seems to follow the main pathway used by OMPs. Therefore, PnlH could also depend on a functional Bam system to be inserted into the membrane. To assess this question, we studied the targeting of PnlH to the membranes of *D. dadantii* lacking different components of the Bam system. In comparison to wild-type strain A350, a somewhat smaller quantity of PnlH was detected in the cells of the *bamB* and *bamE* strains (Fig. 6A, top). This difference became more evident in the membrane fraction prepared from these strains (Fig. 6A, middle). Since similar quantities of the IM protein TolA were detected in all these strains (Fig. 6A, bottom), the observed diminution of PnlH quantity was specific. However, separation of the *bamB* and *bamE* mutant membranes on a sucrose gradient showed that PnlH-6His is still efficiently associated with the OM of these mutants (Fig. 6B). Moreover, the level of PnlH-6His-targeting defect in the three mutants was proportional to the level of OMP-targeting defect observed in the same mutants (Fig. 1).

Taken together, these data show that *bamB* and *bamE* null mutations slightly affect the stability of PnlH without clearly affecting its targeting to the OM.

DISCUSSION

The large majority of proteobacteria outer membrane proteins are either lipoproteins or β -barrel proteins (OMPs), and two distinct specific pathways ensure their targeting to the OM. Lipoproteins are sorted and targeted by the Lol pathway, while OMP targeting relies on the combined action of three periplasmic chaperones, SurA, Skp, and DegP, together with the Bam complex (30, 38). In this work, we have studied the targeting of PnlH, a *D. dadantii* atypical outer membrane protein which is anchored in the OM by an uncleaved Tat signal sequence (8). Since PnlH is not acylated at its amino-terminal extremity, it is unlikely that it directly interacts with the Lol system (8, 9). Thus, we focused our study on the possible role of the OMP-targeting pathway in the targeting of PnlH to the OM.

After exportation by the Tat system, PnlH is released in a folded state in the periplasm. Since the pectin lyase portion of PnlH exhibits 43% sequence identity with a *Pseudomonas marginalis* pectin lyase, PNL, that has been described to be a soluble protein (33), it should be soluble in the periplasm. However, the PnlH signal sequence is highly hydrophobic, and its fusion to a soluble protein (GST) makes it nonsoluble. Thus, the PnlH signal sequence should be buried in the course of the periplasmic transit. Our data show that a *D. dadantii surA* mutant exhibits a strong PnlH stability defect. The same defect was observed for a fusion of the PnlH signal sequence to the mature form of the pectin methyl-esterase PemA but not for the native PemA, indicating that the role of SurA in the stability of PnlH is linked to the signal sequence. Since OMP-targeting defects, such as the one provoked by the absence of SurA, lead to a strong activation of the periplasmic stress response in *E. coli* (42, 53, 10), the PnlH stability defect observed in the *D. dadantii surA* mutant could be an indirect effect due to the activation of the periplasmic stress response by mistargeted OMPs. However, we favor a direct role of SurA on the stability of PnlH because (i) a *skp* mutant also presents a OMP-targeting defect that should activate a periplasmic stress response (38) but does not exhibit any effect on PnlH stability and (ii) SurA binds to the Tat signal sequence of PnlH in an overlay experiment. Therefore, a possible role for SurA could be to hide the hydrophobic PnlH signal sequence and to prevent the periplasmic degradation of PnlH. SurA has been shown to be involved in the transport of the passenger domain of the IcsA autotransporter in the periplasm of *Shigella flexneri* (35). Moreover, direct binding of SurA on the unfolded passenger domain of the *E. coli* autotransporter EspP has been demonstrated (44). Altogether our results and these observations indicate that SurA specificity and chaperone activity are not restricted only to unfolded β -barrel proteins. SurA specificity for OMPs has been linked to a strong preference for aromatic amino acid-rich sequences containing the Ar-X-Ar motif that is particularly frequent in OMPs (1, 15). Such a motif is present in the PnlH signal sequence (phenylalanine 34 and tyrosine 36). Surprisingly, single substitution of each of these aromatic residues did not obviously affect PnlH targeting, indicating that an Ar-X-Ar motif on the PnlH signal sequence could be dispensable for the interaction with SurA. Consistent with this observation, it has been shown that SurA can strongly bind peptides that do not fit the Ar-X-Ar consensus by adapting its tertiary

and quaternary structure (57). Moreover, SurA can fold to form a nonspecific conduit for unfolded polypeptide, as can the domain III of the trigger factor, a general cytoplasmic chaperone (21, 57).

SurA has been described to ensure the delivery of OMPs to the Bam system by interacting with BamA (3, 47, 52). Therefore, it is possible that SurA delivers PnlH to the OM, but how the PnlH signal sequence is inserted in the OM remains unknown. Our data show that *bamB* and *bamE* null mutations slightly affect the stability of PnlH. However, the PnlH insertion in the OM is not disrupted in the corresponding mutants, and the effect of these null mutations on the level of PnlH is proportional to the effect on the level of the β -barrel protein OmpF. This suggests that the role of BamB and BamE on the stability of PnlH could be indirect, provoked by the activation of a periplasmic stress response (38) or the misinsertion of a protein involved in the targeting of PnlH. However, it is worth noting that a model of spontaneous insertion in the OM has been proposed for some bacterial secretins and for mitochondrial signal-anchored proteins (12, 16, 27). Therefore, further work is necessary to determine how the PnlH signal sequence is inserted in the OM and if this process requires proteins other than those of the Bam complex.

Since the fusion of the PnlH signal sequence to PemA is able to target it to the OM, this signal sequence should carry all the information necessary to the targeting of PnlH. The first step of PnlH targeting is exportation through the Tat system, and the PnlH signal sequence thus has the same organization as other Tat signal sequences with a polar amino-terminal part, a Tat consensus, a hydrophobic region, and a polar carboxy-terminal part (32). In this work, we tried to identify the information that ensures the targeting of PnlH to the OM after exportation by the Tat system, and the difficulty was sorting between information allowing the targeting to the Tat system (32) and information allowing the transit in the periplasm and the insertion in the OM. We showed that the substitution PnlHD16A strongly impairs the stability of PnlH in *D. dadantii* without blocking its exportation, as shown by the activation of a periplasmic stress by PnlHD16A in *E. coli*. Since negatively charged residues are not a compulsory characteristic of the amino-terminal part of the Tat signal sequence (32) and since aspartate 16 is conserved in *D. zea*, it is possible that it constitutes a specific signal for the targeting of PnlH to the OM. We could also identify a region of the PnlH signal sequence contained between residues 28 and 41 that could be specifically involved in the targeting of PnlH to the OM after exportation since PnlH with deletion of this region is present in both the IM and the OM of *D. dadantii*. Consistent with this experimental observation, this part of the signal sequence is well conserved in the *D. zea* PnlH. However, single amino acid substitutions in this part of the protein did not lead to an obvious targeting defect, indicating that the signal may not rely on a single amino acid. Interestingly, structure prediction of the PnlH signal sequence suggests the existence of an alpha helix between residues 14 and 32, followed by a coil region formed by prolines 32 and 35. Substitution of these two prolines (PnlHP32LP35A) leads to a degradation of the amino-terminal part of the PnlH signal sequence but does not impair the targeting of PnlH to the OM of *D. dadantii*. Since PnlHP32LP35A induces a strong periplasmic stress in *E. coli*, it is likely that the coil region and thus the length of the predicted alpha helix are important factors for the stability of the PnlH signal sequence in the

periplasm but not for its insertion in the OM. The PnlH signal sequence-predicted alpha helix contains a highly hydrophobic region between residues 24 and 31. Replacement of leucines 30 and 31 by serine and threonine (PnlHL30SL31T), respectively, led to a strong decrease in the PnlH stability in *D. dadantii* and activation of a periplasmic stress in *E. coli*. This could indicate that the decrease of the hydrophobicity of the predicted alpha helix of PnlHL30SL31T is not sufficient to impair the exportation but could impair the targeting to the OM after exportation. Since the PnlH signal sequence anchors PnlH in the OM, it is tempting to think that its predicted hydrophobic alpha helix is responsible for the anchoring of PnlH in the OM. The presence of a hydrophobic alpha helix in an OM protein seems surprising, since the presence of such a structure in membrane proteins constitutes a strong IM retention signal. However, it has been shown that the lipoproteins of the outer membrane auxiliary family PelC of *Pseudomonas aeruginosa* and Wza of *E. coli* are anchored in the OM by a carboxy-terminal amphipathic alpha helix (6, 20), indicating that alpha helices can be inserted in the OM. Another surprising point is the presence of a cluster of hydrophobic residues in the PnlH signal sequence. Indeed, β -barrel proteins do not have such clusters in their primary sequences, but their folding in the OM forms a hydrophobic surface allowing the insertion into the OM (38). Similarly, the oligomerization of Wza forms in the OM a helical barrel that exposes the hydrophobic residues of the carboxy-terminal amphipathic alpha helices to the membrane, while polar residues are localized in the central cavity of the barrel (6). Such an oligomerization has not been reported for PnlH. Thus, the presence and the importance of the hydrophobic cluster in the PnlH signal sequence could be attributed to the requirement for a hydrophobic surface to anchor proteins in the OM.

The *pnlH* is located in a 13-gene cluster, bordered by tRNA genes, that has been acquired by lateral transfer (8). This cluster is not present in most sequenced *Dickeya* strains. No gene that could encode a dedicated targeting system is present in this cluster. OM Tat-anchored proteins, such as PnlH, do not seem to be widespread, and they probably have to rely on existing targeting systems. As shown by this study, SurA could be versatile enough to ensure the targeting of noncanonical OM proteins. This could, at least in part, explain how newly acquired OM proteins can be targeted to the OM to ensure their functions. Such versatility in an OM-targeting pathway could, in short, ensure the acquisition of new functions by lateral gene transfer.

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REFERENCES

- Alcock FH, et al. 2008. Conserved substrate binding by chaperones in the bacterial periplasm and the mitochondrial intermembrane space. *Biochem. J.* 409:377–387.
- Allen JWA, Tomlinson EJ, Hong L, Ferguson SJ. 2002. The *Escherichia coli* cytochrome c maturation (Ccm) system does not detectably attach heme to single cysteine variants of an apocytochrome c. *J. Biol. Chem.* 277:33559–33563.

3. Bennion D, Charlson ES, Coon E, Misra R. 2010. Dissection of β -barrel outer membrane protein assembly pathways through characterizing Bama POTRA 1 mutants of *Escherichia coli*. *Mol. Microbiol.* 77:1153–1171.
4. de Cock H, Struyv  M, Kleerebezem M, van der Krift T, Tommassen J. 1997. Role of the carboxy-terminal phenylalanine in the biogenesis of outer membrane protein PhoE of *Escherichia coli* K-12. *J. Mol. Biol.* 269:473–478.
5. Denoncin K, Schwalm J, Vertommen D, Silhavy TJ, Collet J-F. 2012. Dissecting the *Escherichia coli* periplasmic chaperone network using differential proteomics. *Proteomics* 12:1391–1401.
6. Dong C, et al. 2006. Wza the translocon for *E. coli* capsular polysaccharides defines a new class of membrane protein. *Nature* 444:226–229.
7. Fellay R, Frey J, Krisch H. 1987. Interposon mutagenesis of soil and water bacteria: a family of DNA fragments designed for *in vitro* insertional mutagenesis of Gram-negative bacteria. *Gene* 52:147–154.
8. Ferrandez Y, Condemine G. 2008. Novel mechanism of outer membrane targeting of proteins in Gram-negative bacteria. *Mol. Microbiol.* 69:1349–1357.
9. Fukuda A, et al. 2002. Aminoacylation of the N-terminal cysteine is essential for Lol-dependent release of lipoproteins from membranes but does not depend on lipoprotein sorting signals. *J. Biol. Chem.* 277:43512–43518.
10. Gerken H, Leiser OP, Bennion D, Misra R. 2010. Involvement and necessity of the Cpx regulon in the event of aberrant β -barrel outer membrane protein assembly. *Mol. Microbiol.* 75:1033–1046.
11. Gralnick JA, Vali H, Lies DP, Newman DK. 2006. Extracellular respiration of dimethyl sulfoxide by *Shewanella oneidensis* strain MR-1. *Proc. Natl. Acad. Sci. U. S. A.* 103:4669–4674.
12. Guilvout I, et al. 2008. *In vitro* multimerization and membrane insertion of bacterial outer membrane secretin PulD. *J. Mol. Biol.* 382:13–23.
13. Hagan CL, Kim S, Kahne D. 2010. Reconstitution of outer membrane protein assembly from purified components. *Science* 328:890–892.
14. Hayashi S, Wu HC. 1990. Lipoproteins in bacteria. *J. Bioenerg. Biomembr.* 22:451–471.
15. Hennecke G, Nolte J, Volkmer-Engert R, Schneider-Mergener J, Behrens S. 2005. The periplasmic chaperone SurA exploits two features characteristic of integral outer membrane proteins for selective substrate recognition. *J. Biol. Chem.* 280:23540–23548.
16. Hoang HH, et al. 2011. Outer membrane targeting of *Pseudomonas aeruginosa* proteins shows variable dependence on the components of Bam and Lol machineries. *mBio* 2:e00246–11. doi:10.1128/mBio.00246-11.
17. Hugouvieux-Cotte-Pattat N, Charaoui-Boukerza S. 2009. Catabolism of raffinose, sucrose, and melibiose in *Erwinia chrysanthemi* 3937. *J. Bacteriol.* 191:6960–6967.
18. Hunke S, Betton J-M. 2003. Temperature effect on inclusion body formation and stress response in the periplasm of *Escherichia coli*. *Mol. Microbiol.* 50:1579–1589.
19. Kelley LA, Sternberg MJE. 2009. Protein structure prediction on the Web: a case study using the Phyre server. *Nat. Protoc.* 4:363–371.
20. Kowalska K, et al. 2010. The C-terminal amphipathic α -helix of *Pseudomonas aeruginosa* PelC outer membrane protein is required for its function. *Biochimie* 92:33–40.
21. Ludlam AV, Moore BA, Xu Z. 2004. The crystal structure of ribosomal chaperone trigger factor from *Vibrio cholerae*. *Proc. Natl. Acad. Sci. U. S. A.* 101:13436–13441.
22. Malinverni JC, et al. 2006. YfiO stabilizes the YaeT complex and is essential for outer membrane protein assembly in *Escherichia coli*. *Mol. Microbiol.* 61:151–164.
23. Maneewannakul S, Maneewannakul K, Ippen-Ihler K. 1994. The pKSM710 vector cassette provides tightly regulated lac and T7lac promoters and strategies for manipulating N-terminal protein sequences. *Plasmid* 31:300–307.
24. Masuda K, Matsuyama S, Tokuda H. 2002. Elucidation of the function of lipoprotein-sorting signals that determine membrane localization. *Proc. Natl. Acad. Sci. U. S. A.* 99:7390–7395.
25. Matsuyama S, Yokota N, Tokuda H. 1997. A novel outer membrane lipoprotein, LolB (HemM), involved in the LolA (p20)-dependent localization of lipoproteins to the outer membrane of *Escherichia coli*. *EMBO J.* 16:6947–6955.
26. Matsuyama S, Tajima T, Tokuda H. 1995. A novel periplasmic carrier protein involved in the sorting and transport of *Escherichia coli* lipoproteins destined for the outer membrane. *EMBO J.* 14:3365–3372.
27. Merklinger E, et al. 2012. Membrane integration of a mitochondrial signal-anchored protein does not require additional proteinaceous factors. *Biochem. J.* 442:381–389.
28. Miller JH. 1992. A short course in bacterial genetics: a laboratory manual and handbook for *Escherichia coli* and related bacteria. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
29. Miot M, Betton J-M. 2004. Protein quality control in the bacterial periplasm. *Microb. Cell Fact.* 3:4.
30. Okuda S, Tokuda H. 2011. Lipoprotein sorting in bacteria. *Annu. Rev. Microbiol.* 65:239–259.
31. Osborn MJ, Gander JE, Parisi E, Carson J. 1972. Mechanism of assembly of the outer membrane of *Salmonella typhimurium*. Isolation and characterization of cytoplasmic and outer membrane. *J. Biol. Chem.* 247:3962–3972.
32. Palmer T, Berks BC. 2012. The twin-arginine translocation (Tat) protein export pathway. *Nat. Rev. Microbiol.* 10:483–496.
33. Papi R, Kyriakidis D. 2003. Overexpression of the pectin lyase gene of *Pseudomonas marginalis* in *Escherichia coli* and purification of the active enzyme. *Biotechnol. Appl. Biochem.* 37:187–194.
34. Pugsley AP. 1993. The complete general secretory pathway in gram-negative bacteria. *Microbiol. Rev.* 57:50–108.
35. Purdy GE, Fisher CR, Payne SM. 2007. IcsA surface presentation in *Shigella flexneri* requires the periplasmic chaperones DegP, Skp, and SurA. *J. Bacteriol.* 189:5566–5573.
36. Raina S, Missiakas D, Georgopoulos C. 1995. The rpoE gene encoding the sigma E (sigma 24) heat shock sigma factor of *Escherichia coli*. *EMBO J.* 14:1043–1055.
37. Resibois A, Colet M, Faelen M, Schoonejans E, Toussaint A. 1984. phiEC2, a new generalized transducing phage of *Erwinia chrysanthemi*. *Virology* 137:102–112.
38. Ricci DP, Silhavy TJ. 2012. The Bam machine: a molecular cooper. *Biochim. Biophys. Acta* 1818:1067–1084.
39. Rizzitello AE, Harper JR, Silhavy TJ. 2001. Genetic evidence for parallel pathways of chaperone activity in the periplasm of *Escherichia coli*. *J. Bacteriol.* 183:6794–6800.
40. Robert V, et al. 2006. Assembly factor Omp85 recognizes its outer membrane protein substrates by a species-specific C-terminal motif. *PLoS Biol.* 4:e377. doi:10.1371/journal.pbio.0040377.
41. Roeder DL, Collmer A. 1985. Marker-exchange mutagenesis of a pectate lyase isozyme gene in *Erwinia chrysanthemi*. *J. Bacteriol.* 164:51–56.
42. Rouviere PE, Gross CA. 1996. SurA, a periplasmic protein with peptidyl-prolyl isomerase activity, participates in the assembly of outer membrane porins. *Genes Dev.* 10:3170–3182.
43. Ruiz N, Falcone B, Kahne D, Silhavy TJ. 2005. Chemical conditionality: a genetic strategy to probe organelle assembly. *Cell* 121:307–317.
44. Ruiz-Perez F, et al. 2009. Roles of periplasmic chaperone proteins in the biogenesis of serine protease autotransporters of Enterobacteriaceae. *J. Bacteriol.* 191:6571–6583.
45. Schulz GE. 2000. beta-Barrel membrane proteins. *Curr. Opin. Struct. Biol.* 10:443–447.
46. Shevchik VE, Condemine G, Hugouvieux-Cotte-Pattat N, Robert-Baudouy J. 1996. Characterization of pectin methylesterase B, an outer membrane lipoprotein of *Erwinia chrysanthemi* 3937. *Mol. Microbiol.* 19:455–466.
47. Sklar JG, et al. 2007. Lipoprotein SmpA is a component of the YaeT complex that assembles outer membrane proteins in *Escherichia coli*. *Proc. Natl. Acad. Sci. U. S. A.* 104:6400–6405.
48. Sklar JG, Wu T, Kahne D, Silhavy TJ. 2007. Defining the roles of the periplasmic chaperones SurA, Skp, and DegP in *Escherichia coli*. *Genes Dev.* 21:2473–2484.
49. Subrini O, Betton J-M. 2009. Assemblies of DegP underlie its dual chaperone and protease function. *FEMS Microbiol. Lett.* 296:143–148.
50. Terada M, Kuroda T, Matsuyama SI, Tokuda H. 2001. Lipoprotein sorting signals evaluated as the LolA-dependent release of lipoproteins from the cytoplasmic membrane of *Escherichia coli*. *J. Biol. Chem.* 276:47690–47694.
51. Voulhoux R, Bos MP, Geurtsen J, Mols M, Tommassen J. 2003. Role of a highly conserved bacterial protein in outer membrane protein assembly. *Science* 299:262–265.
52. Vuong P, Bennion D, Mantei J, Frost D, Misra R. 2008. Analysis of YfgL and YaeT interactions through bioinformatics, mutagenesis, and biochemistry. *J. Bacteriol.* 190:1507–1517.
53. Walsh NP, Alba BM, Bose B, Gross CA, Sauer RT. 2003. OMP peptide signals initiate the envelope-stress response by activating DegS protease via relief of inhibition mediated by its PDZ domain. *Cell* 113:61–71.

54. Walton TA, Sandoval CM, Fowler CA, Pardi A, Sousa MC. 2009. The cavity-chaperone Skp protects its substrate from aggregation but allows independent folding of substrate domains. *Proc. Natl. Acad. Sci. U. S. A.* **106**:1772–1777.
55. Watanabe S, Oguchi Y, Yokota N, Tokuda H. 2007. Large-scale preparation of the homogeneous LolA lipoprotein complex and efficient in vitro transfer of lipoproteins to the outer membrane in a LolB-dependent manner. *Protein Sci.* **16**:2741–2749.
56. Wu T, et al. 2005. Identification of a multicomponent complex required for outer membrane biogenesis in *Escherichia coli*. *Cell* **121**: 235–245.
57. Xu X, Wang S, Hu Y-X, McKay DB. 2007. The periplasmic bacterial molecular chaperone SurA adapts its structure to bind peptides in different conformations to assert a sequence preference for aromatic residues. *J. Mol. Biol.* **373**:367–381.
58. Yakushi T, Masuda K, Narita S, Matsuyama S, Tokuda H. 2000. A new ABC transporter mediating the detachment of lipid-modified proteins from membranes. *Nat. Cell Biol.* **2**:212–218.