

Generation of photoluminescent ultrashort carbon nanotubes through nanoscale exciton localization at sp3 -defect sites

Noémie Danné, Mijin Kim, Antoine G. Godin, Hyejin Kwon, Zhenghong Gao, Xiaojian Wu, Nicolai F Hartmann, Stephen K Doorn, Brahim Lounis, Yuhuang Wang, et al.

▶ To cite this version:

Noémie Danné, Mijin Kim, Antoine G. Godin, Hyejin Kwon, Zhenghong Gao, et al.. Generation of photoluminescent ultrashort carbon nanotubes through nanoscale exciton localization at sp3 -defect sites. 2018. hal-01727325

HAL Id: hal-01727325 https://hal.science/hal-01727325

Preprint submitted on 9 Mar 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Generation of photoluminescent ultrashort carbon nanotubes through nanoscale exciton localization at sp³-defect sites

Noémie Danné^{1,2}, Mijin Kim³, Antoine G. Godin^{1,2}, Hyejin Kwon³, Zhenghong Gao^{1,2}, Xiaojian Wu³, Nicolai F. Hartmann⁴, Stephen K. Doorn⁴, Brahim Lounis^{1,2}, YuHuang Wang^{3,5} and Laurent Cognet^{1,2,*}

¹Univ. Bordeaux, Laboratoire Photonique Numérique et Nanosciences, UMR 5298, F-33400 Talence, France ²Institut d'Optique & CNRS, LP2N UMR 5298, F-33400 Talence, France

³Department of Chemistry and Biochemistry, University of Maryland, College Park, MD 20742, United States

⁴Center for Integrated Nanotechnologies, Materials Physics and Applications Division,, Los Alamos National Laboratory, Los Alamos, NM 87545 USA

⁵Maryland NanoCenter, University of Maryland, College Park, MD 20742, United States

*Laurent.cognet@u-bordeaux.fr.

ABSTRACT

The intrinsic near-infrared photoluminescence observed in long single walled carbon nanotubes is systematically quenched in ultrashort single-walled carbon nanotubes (usCNTs, below 100 nm length) due to their short dimension as compared to the exciton diffusion length. It would however be key for number of applications to have such tiny nanostructure displaying photoluminescence emission to complement their unique physical, chemical and biological properties. Here we demonstrate that intense photoluminescence can be created in usCNTs (~40 nm length) upon incorporation of emissive sp^3 -defect sites in order to trap excitons. Using super-resolution imaging at <25 nm resolution, we directly reveal the localization of excitons at the defect sites on individual usCNTs. They are found preferentially localized at nanotube ends which can be separated by less than 40 nm and behave as independent emitters. The demonstration and control of bright near-infrared photoluminescence in usCNTs through exciton trapping opens the possibility to engineering tiny carbon nanotubes for applications in various domains of research including quantum optics and bioimaging.

The introduction of chemical or structural defects in carbon nanostructures is a powerful route to shape and expand their optical properties. For instance, in nanodiamonds, nitrogen

vacancies generate stable photoluminescence leading to the realization of quantum light sources, sensors, and of promising biological applications^{1.4}. More recently, the incorporation of fluorescent quantum defects in single-walled carbon nanotubes (SWCNTs) created the opportunity to enhance SWCNT near-infrared photoluminescence (PL) properties⁵⁻⁸ thus generating a growing interest for the realization of single photon sources or bioimaging^{1-4,6,9,10}. Indeed, pristine semiconducting SWCNTs display low luminescence quantum yield, since their photophysical properties^{5-8,11} are mainly imposed by non-radiative mechanisms, associated with structural and environmental defects¹²⁻¹⁶. In this framework, exciton diffusion, which spans typically one to several hundreds of nanometers along the nanotube^{13,17}, plays a key role since an exciton scans the integrity of the nanotube backbone before it may eventually emit a photon. As a consequence, ultrashort nanotubes (usCNTs)-semiconducting SWCNTs with lengths significantly shorter than the exciton diffusion range-display dominant non-radiative exciton decay mechanisms at nanotube ends which are efficient PL quenching sites and thus imperceptible photoluminescence^{18,19}. Due to their exceptional small sizes and near-infrared optical properties, luminescent usCNTs would however be a key asset for a variety of applications including nano-electronics²⁰, biology^{21,22} and nanomedicine²³.

In sp³-defect functionalized SWCNTs, evidences for exciton localization at defects were obtained^{5,24,26} and wide-field PL imaging showed emission from isolated defect sites²⁷. However, the extent of localization could not be resolved due to the limited spatial resolution of standard microscopy (~ 450 nm for high numerical aperture (NA) objectives and emission wavelength of ~1100 nm) as compared to exciton diffusion lengths. Here we demonstrate that sp³-defect functionalization can be used to trap excitons at emissive defect sites that are intentionally incorporated into usCNTs, and thus efficiently brightens usCNTs so that single nanotube detection of usCNTs becomes possible by fluorescence microscopy. Through the application on single usCNTs, well below the diffraction limit, usCNT lengths and free exciton diffusion lengths. Through this direct visualization, we could then demonstrate that defect sites are preferentially localized at usCNTs ends, and display independent emission properties.

We prepared length-sorted CoMoCAT usCNTs with median length of 43 nm following a previously published procedure¹⁹. The ultrashort lengths were determined by atomic force microscopy (AFM) measurement (N=88, **Figure 1a**) and single particle photothermal imaging (PhI). The photothermal signals are directly linked to nanoparticle absorption so that this distribution mirrors the AFM length distribution albeit with better statistics¹⁹ (N= 472, **Figure 1b**). Ensemble absorption spectra confirmed the predominant presence of (6,5) nanotubes in the sample as expected from SG65i CoMoCAT nanotubes (Supporting Information **Figure S1**). On the other hand, an excitation-emission 2D PL map displayed imperceptible PL signals (**Figure 1c**) which is not surprising as it has been well established that usCNTs do not fluoresce^{18,19,28}. Consistent with the negligible PL in the bulk samples, no luminescent usCNTs were detected at the single nanotube level even using long integration times (several seconds, **Figure 1d**).



Figure 1. Brightening of ultrashort SWCNTs through fluorescent quantum defects. (a) Cumulative distribution of usCNT lengths measured by AFM (N=88, median= 43 nm, $1^{st}/3^{rd}$ quartile= 24/69 nm). (b), Distribution of usCNT absorption signals measured by PhI (N=472) which also reflects the nanotube lengths distribution but with better statistics. (c) 2D PL map of usCNTs shows no luminescence. (d) single molecule imaging of usCNTs is not observed. (e) 2D PL map of f-usCNT reveals bright luminescence. (f) single molecule imaging of f-usCNT is clearly observed. (g) Distribution of f-usCNT PL signals (N=265) mirrors nanotube length distributions in (a) and (b).

We next covalently attached perfluorinated-hexyl chain $(-C_6F_{13})$ to the usCNTs producing functionalized usCNTs (f-usCNTs). A 2D PL map of the f-usCNTs solution reveals a bright and stable PL peak, E₁₁* around 1160 nm (Figure 1e). In C₆F₁₃-functionalized, long (6,5)-SWCNTs, this red-shifted peak ($E_{11} - E_{11}^*$, 183 meV) has been attributed to PL arising from the chemical functionalization⁸. Interestingly, weak yet photostable E_{11} exciton emission also became observable from f-usCNTs, in contrast to the unfunctionalized usCNT samples. fusCNTs that were spin-coated on a microscope glass coverslip pre-coated with polyvinylpyrrolidone were next observed at the single tube level. We used a dual detection fluorescence microscope in order to simultaneously detect the E_{11} and/or E_{11} * PL bands of (6,5) f-usCNTs upon 568 nm excitation (Supporting Information Figure S2). Unambiguously, bright f-usCNT PL spots could be observed in the E_{11}^* channel at the video frame rates (Figure 1f). All detected spots were diffraction limited as expected from single nanoscale emitters. In addition, we built a distribution of intensities measured from 265 luminescent spots and obtained a monomodal distribution (Figure 1g) in agreement with usCNT length and PhI signal distributions (Figure 1a-b). These observations reveal that the detected spots stem from individual f-usCNTs and thus that luminescent usCNTs are bright enough to be detected at the single tube level owing to C₆F₁₃-functionalization. Noteworthy, on occasion a few dim PL spots were detected in the E₁₁ channel and co-localized with E₁₁* PL spots from single f-usCNTs (Supporting Information Figure S3) confirming the observation in Figure 1e.

Because usCNT lengths (median length of 43 nm, **Figure 1a**) are significantly shorter than the exciton diffusion length (≥ 100 nm), the efficient PL of f-usCNTs is an unambiguous signature of E_{11}^* exciton localization (**Figure 2**). Indeed, the prevailing interpretation for the vanishing E_{11} PL in usCNTs is that excitons systematically decay non-radiatively at nanotube ends acting as efficient quenching defects^{15,18,29,30} (**Figure 2a**). C₆F₁₃ functionalization thus prevents diffusion-related end-quenching of E_{11} excitons through the creation of localized E_{11}^* excitons (**Figure 2b**). Interestingly, the observation of weak yet stable E_{11} PL in f-usCNTs in contrast to usCNTs might indicate that thermal detrapping of E_{11}^* excitons can occur from localized sites.



Figure 2. Schematics of photo-excited E_{11} exciton fate in usCNTs. (a) exciton diffusion and quenching in unfunctionalized usCNTs. In an usCNT, exciton decay is not diffusion-limited but nanotube-lengthdependent. End quenching imposes extremely low fluorescence quantum yields of usCNTs. (b) Schematics of exciton diffusion and trapping at defects in f-usCNTs. Mobile excitons in an f-usCNT can be trapped at fluorescent quantum defects and therefore efficiently luminesce (top: one luminescent defect, bottom: multiple defects at different positions).

In order to investigate the detrapping efficiency, we measured the temperature dependence of the ratio of E_{11} PL intensity to that of E_{11}^* PL measured on bulk spectra. This ratio follows an Arrhenius type relation (**Figure 3a**) with a fitted thermal detrapping energy of 109 meV, which is smaller than the optical gap (183 meV). This energy difference can be understood in terms of the vibrational reorganization energy associated with defect sites²⁴. From the Arrhenius low, we estimated that ~2 % of the E_{11}^* excitons can be transferred to the E_{11} population through thermal detrapping at room temperature (**Figure 3b**). However, thermal detrapping cannot fully account for the total observed E_{11} PL in the f-usCNTs spectra. Another mechanism could be that the formation of trapped E_{11}^* excitons from photo-created E_{11} excitons requires crossing a small potential barrier²⁵ (**Figure 3c**). Finally, all these effects might be reinforced by an E_{11}^* state filling effect previously suggested³¹. Indeed, we observed at the single nanotube level that in f-usCNTs, E_{11}^* PL saturates at lower laser intensities than E_{11} PL measured on longer SWCNTs (**Figure 3 d-e**). Saturation at lower intensities is also consistent with the observation of longer PL decays in sp³ defect functionalized SWCNTs as

compared to unfunctionalized SWCNTs²⁶. Efficient E_{11}^* state filling would then enhance the subpopulation of E_{11} excitons in f-usCNTs and thus the observed E_{11} emission (**Figure 3f**). We note that in all these three scenarios (**Figure 3d-f**), the presence of at least two defect sites is required to compete with diffusive quenching at the nanotube ends (**Figure 2b**).



Figure 3. Trapping and detrapping of E_{11} * excitons. (a) The van 't Hoff plot for f-usCNTs as derived from the integrated PL intensity ratio of E11 and E11* at the corresponding temperature (black dots). The slope of the linear fitting (red line) provides the detrapping energy of 109 meV. (b) Schematic representation of thermal detrapping of a E_{11} * exciton to generate a mobile E_{11} exciton. (c) Schematic representation of E_{11} * exciton trapping in the presence of a potential barrier leading to the presence of a small population of E_{11} excitons. (d) Mean E_{11} * PL signal of individual f-usCNTs as a function of cw laser intensity (N = 8 usCNTs; circles: experimental data; solid line: fit using a saturation profile yielding a saturation intensity of 0.8 kW/cm²). (e) Mean E_{11} PL signal of individual long CNTs from the same preparation as usCNTs as a function of cw laser intensity (N = 12 long nanotubes; circles: experimental data; solid line: fit using a saturation profile yielding a saturation intensity of 4.3 kW/cm²). (f) Schematic representation of E_{11} * exciton state filling leading to the presence of a small population of E_{11} excitons.

Conversely, by preventing E_{11}^* excitons from reaching and quenching at the nanotube ends, a single isolated site of sp³-defect localization can in principle brighten usCNTs through

emission at 1160 nm, while the incorporation of several sites, including at the usCNT ends, should further enhance f-usCNT E_{11}^* photoluminescence (**Figure 2b**). To determine the relative localization of the defects along the usCNTs, the position of emitting sites separated by $\Delta d < 43$ nm must be imaged at a resolution much below the diffraction limit (1,22 λ /2NA ~ 500 nm, for $\lambda = 1160$ nm). This requires a fluorescence imaging modality having at least a ten-fold resolution improvement over standard diffraction-limited fluorescence microscopy. We also wish to investigate whether luminescent defects behave as independent emitters in f-usCNTs or whether their close proximity on the nanotube backbone would condition their PL properties.

To achieve the required resolution and considering that f-usCNT emission occurs in the near-infrared (i.e. at long wavelengths), we implemented a super-resolution microscopy strategy based on single emitter localization for its adequacy with f-usCNT photophysics — several super-resolution imaging techniques have been widely used in biology to achieve sub-wavelength resolution imaging in fluorescence microscopy³². Localization microscopy was previously demonstrated on pristine SWCNTs where the nanotubes were exposed to quenching moieties³³. The presence of local extrinsic PL, defects were localized as missing PL signals at sub-wavelength resolution which also provided a direct visualization of the ~100 nm exciton excursion range in pristine SWCNTs. Yet, neither usCNTs which do not fluoresce nor defect tailored SWCNTs were super-resolved previously.

In the following, f-usCNTs were directly spincoated on glass coverslips and single fusCNTs PL imaging was performed. In these conditions, the majority of nanotubes showed blinking behavior as similarly observed in long functionalized SWCNTs²⁷. **Figure 4a-d** displays PL intensity traces acquired over several minutes revealing the frequent presence of multiple intensity levels for single f-usCNTs. This is quantified by analyzing the amplitude of 64 intensity steps on different f-usCNTs, presented in the distribution of **Figure 4e**. The observed distribution of intensity steps is multimodal, with a factor of two separating the two sub-populations that we observe, demonstrating that individual sites are imaged in the first subpopulation. Importantly, the presence of the second blinking population indicates that the PL behaviors from two defect sites can be uncorrelated on single (sub 50 nm) f-usCNTs. To unambiguously establish the independent localization of defect sites at a scale on f-usCNTs and rule out the possibility that nanotube bundles or outliers in nanotube length distribution were at the origin of the second intensity step population, defect localization was investigated by superlocalization analysis on the f-usCNTs that displayed multiple intensity steps.



Figure 4. Evidence of E_{11}^* exciton localization. (a-d) E_{11}^* PL intensity traces of 4 different single blinking f-usCNTs as a function of time. Red (resp. green) triangles represent negative (resp. positive) unitary steps. (e) Histogram of E_{11}^* PL step intensities observed in single blinking f-usCNTs (N=64). The histogram is multimodal which reveals the presence of several individual emitting sites in f-usCNTs. The presence of independent intensity levels in f-usCNTs having lengths shorter than the diffusion length of E_{11} excitons in long CNTs is a signature of E_{11}^* exciton.

For each intensity step event, differential images were computed, consisting of the differences between successive images before and after the step, in order to isolate the appearance or disappearance of the PL stemming from an individual luminescent site (**Figure 5a** and Supporting Information **Figure S5**). When steps were negative, the absolute value of the pixel intensities in the differential image was considered to obtain images with positive pixel values. Images preceding and following the intensity step over time are averaged in order to improve signal-to-noise ratio in the differential images and subsequent localization precision (see Supporting Information methods and **Figure S5**). Individual spots in differential images were then fitted by a Gaussian approximating the point-spread function as commonly performed in localization microscopy³⁴. Indeed, these fits retrieve the localization were created for several f-usCNTs showing more than one intensity level (i.e. at least two independent emitting sites). When background noise can be neglected relative to the number of detected photons *N*, the localization precision can be approximated by ~ *s*/*N*^{1/2}, where *s* is the standard deviation of the point spread function. In practice, camera pixelization and noise should be taken into account³⁶

(see Supporting Information methods) leading to localization precisions that could be estimated for each individual defect site. Taking into account localization precision, on each single tube, defect detections that were below the localization precision were considered as one single defect (see Supporting Information methods and **Figure S6**). The mean localization precision for individual defect sites was ultimately determined to be 22 nm in our experiments (see Supporting Information methods). Subsequent defect localizations are displayed for several fusCNTs, revealing the presence of at least two unmistakably resolved defects on **Figure 5a-e** with localization precision displayed as dotted circles.



Figure 5. Super-resolved images of E_{11}^* exciton localization in f-usCNTs. (a-e) Image representation of single blinking E_{11}^* exciton localizations in f-usCNTs by colored 2D Gaussian curve rendering. For this, each localization is displayed as a two-dimensional Gaussian of unit amplitude and width equal to the localization precision as commonly used in localization microscopy³⁴. The mean E_{11}^* exciton localization of each single site is represented by blue spots and the corresponding localizaton precision by dotted circles (see supporting information methods and Figure S5). (f) Histogram of super-resolved distances ΔL between E_{11}^* exciton localizations (18 f-usCNTs analyzed). (g) Schematic of defect sites preferentially localized at nanotube ends.

These images constitute a direct visualization of exciton localization at defect sites to locations below the measured precision, which is significantly smaller than the exciton diffusion range in pristine nanotubes. Interestingly, the distribution of distances between independently localized defects supports this finding (**Figure 5f**), but also indicates that typical distances between defects match the f-usCNT lengths (**Figure 1a-b**) and PL signals (**Figure 1f**). This observation indicates that functionalization preferentially occurred at nanotube ends (**Figure 5g**). We suspect that the functionalization occurs propagatively from the end defects,^{37,38} creating fluorescent quantum defects^{7,8} that block excitons from being trapped by the otherwise quenching ends.

In this work, we achieved dramatic PL brightening of ultrashort carbon nanotubes upon defect functionalization, which otherwise do not luminesce. Brightening is so efficient that ultrashort carbon nanotubes can be detected at the single nanotube level. This brightening of sub-100 nm nanotubes is a direct consequence of exciton localization at intentionally introduced defects in the functionalized tubes. To directly support this mechanism, we developed a super-resolution imaging methodology to resolve the different emission sites with <25 nm resolution on single tubes. Super-resolution imaging unambiguously reveals that emitting excitons are localized at nanotube ends. Brightening of ultrashort nanotubes not only constitutes a direct evidence for exciton localization in defect functionalized tubes, it also opens a promising route for several applications based on carbon nanotubes. For instance, luminescent ultra-short nanotubes with a controlled number of single emitters might be highly valuable for the development of high quality single photon emitters for quantum information¹⁰. We also foresee that luminescent ultra-short nanotubes will constitute a milestone for biological imaging where ultra-small, bright photostable emitters in the near-infrared biological window are vividly required³⁹.

ACKNOWLEDGEMENTS

This work was supported by CNRS, the French Ministry of Higher Education, Research and Innovation, the Agence Nationale de la Recherche (ANR-14-OHRI-0001-01), IdEx Bordeaux (ANR-10-IDEX-03-02), and Conseil Regional Nouvelle-Aquitaine (2015-1R60301-00005204). YHW gratefully acknowledges the National Science Foundation of U.S. for financial support through grant no. 1507974 for the development of the defect chemistry. The authors thank having had access to the Center for Integrated Nanotechnologies, a U.S. Department of Energy Office of Science user facility.

REFERENCES

- Maze, J. R.; Stanwix, P. L.; Hodges, J. S.; Hong, S.; Taylor, J. M.; Cappellaro, P.; Jiang, L.; Dutt, M. V. G.; Togan, E.; Zibrov, A. S.; *et al.* Nanoscale Magnetic Sensing with an Individual Electronic Spin in Diamond. *Nature* 2008, 455, 644–647.
- (2) Vlasov, I. I.; Shiryaev, A. A.; Rendler, T.; Steinert, S.; Lee, S.-Y.; Antonov, D.; Vörös, M.; Jelezko, F.; Fisenko, A. V.; Semjonova, L. F.; *et al.* Molecular-Sized Fluorescent Nanodiamonds. *Nat. Nanotechnol.* **2013**, *9*, 54–58.
- Dolde, F.; Jakobi, I.; Naydenov, B.; Zhao, N.; Pezzagna, S.; Trautmann, C.; Meijer, J.; Neumann, P.; Jelezko, F.; Wrachtrup, J. Room-Temperature Entanglement Between Single Defect Spins in Diamond. *Nat. Phys.* 2013, *9*, 139–143.
- (4) Mochalin, V. N.; Shenderova, O.; Ho, D.; Gogotsi, Y. The Properties and Applications of Nanodiamonds. *Nat. Nanotechnol.* **2011**, *7*, 11–23.
- Miyauchi, Y.; Iwamura, M.; Mouri, S.; Kawazoe, T.; Ohtsu, M.; Matsuda, K.
 Brightening of Excitons in Carbon Nanotubes on Dimensionality Modification. *Nat. Photon.* 2013, 7, 715–719.
- Ghosh, S.; Bachilo, S. M.; Simonette, R. A.; Beckingham, K. M.; Weisman, R. B.
 Oxygen Doping Modifies Near-Infrared Band Gaps in Fluorescent Single-Walled Carbon Nanotubes. *Science* 2010, *330*, 1656–1659.
- Piao, Y.; Meany, B.; Powell, L. R.; Valley, N.; Kwon, H.; Schatz, G. C.; Wang, Y. Brightening of Carbon Nanotube Photoluminescence Through the Incorporation of Sp3 Defects. *Nat. Chem.* 2013, *5*, 840–845.
- Kwon, H.; Furmanchuk, A.; Kim, M.; Meany, B.; Guo, Y.; Schatz, G. C.; Wang, Y.
 Molecularly Tunable Fluorescent Quantum Defects. *J. Am. Chem. Soc.* 2016, *138*, 6878–6885.
- (9) Ma, X.; Hartmann, N. F.; Baldwin, J. K. S.; Doorn, S. K.; Htoon, H. Room-Temperature Single-Photon Generation From Solitary Dopants of Carbon Nanotubes. *Nat. Nanotechnol.* 2015, 10, 671–675.
- (10) He, X.; Hartmann, N. F.; Ma, X.; Kim, Y.; Ihly, R.; Blackburn, J. L.; Gao, W.; Kono, J.; Yomogida, Y.; Hirano, A.; *et al.* Tunable Room-Temperature Single-Photon Emission at Telecom Wavelengths From Sp3 Defects in Carbon Nanotubes. *Nat. Photon.* 2017, *11*, 577–582.
- O'Connell, M. J.; Bachilo, S. M.; Huffman, C. B.; Moore, V. C.; Strano, M. S.;
 Haroz, E. H.; Rialon, K. L.; Boul, P. J.; Noon, W. H.; Kittrell, C.; *et al.* Band Gap
 Fluorescence From Individual Single-Walled Carbon Nanotubes. *Science* 2002, 297, 593–596.
- (12) Perebeinos, V.; Avouris, P. Phonon and Electronic Nonradiative Decay Mechanisms of Excitons in Carbon Nanotubes. *Phys. Rev. Lett.* **2008**, *101*, 057401.
- (13) Cognet, L.; Tsyboulski, D. A.; Rocha, J.-D. R.; Doyle, C. D.; Tour, J. M.; Weisman, R. B. Stepwise Quenching of Exciton Fluorescence in Carbon Nanotubes by Single-Molecule Reactions. *Science* 2007, *316*, 1465–1468.
- (14) Berciaud, S.; Cognet, L.; Lounis, B. Luminescence Decay and the Absorption Cross Section of Individual Single-Walled Carbon Nanotubes. *Phys. Rev. Lett.* 2008, 101, 077402.
- (15) Crochet, J. J.; Duque, J. G.; Werner, J. H.; Lounis, B.; Cognet, L.; Doorn, S. K. Disorder Limited Exciton Transport in Colloidal Single-Wall Carbon Nanotubes. *Nano Lett.* 2012, *12*, 5091–5096.
- (16) Crochet, J. J.; Duque, J. G.; Werner, J. H.; Doorn, S. K. Photoluminescence Imaging of Electronic-Impurity-Induced Exciton Quenching Insingle-Walled Carbon

Nanotubes. Nat. Nanotechnol. 2012, 7, 126–132.

- (17) Siitonen, A. J.; Tsyboulski, D. A.; Bachilo, S. M.; Weisman, R. B. Surfactant-Dependent Exciton Mobility in Single-Walled Carbon Nanotubes Studied by Single-Molecule Reactions. *Nano Lett.* **2010**, *10*, 1595–1599.
- (18) Rajan, A.; Strano, M. S.; Heller, D. A.; Hertel, T.; Schulten, K. Length-Dependent Optical Effects in Single Walled Carbon Nanotubes †. J. Phys. Chem. B 2008, 112, 6211–6213.
- (19) Gao, Z.; Oudjedi, L.; Faes, R.; Moroté, F.; Jaillet, C.; Poulin, P.; Lounis, B.; Cognet, L. Optical Detection of Individual Ultra-Short Carbon Nanotubes Enables Their Length Characterization Down to 10 Nm. *Sci. Rep.* 2015, *5*, 17093.
- (20) Franklin, A. D.; Luisier, M.; Han, S.-J.; Tulevski, G.; Breslin, C. M.; Gignac, L.; Lundstrom, M. S.; Haensch, W. Sub-10 Nm Carbon Nanotube Transistor. *Nano Lett.* 2012, 12, 758–762.
- (21) Kolosnjaj-Tabi, J.; Hartman, K. B.; Boudjemaa, S.; Ananta, J. S.; Morgant, G.; Szwarc, H.; Wilson, L. J.; Moussa, F. In VivoBehavior of Large Doses of Ultrashort and Full-Length Single-Walled Carbon Nanotubes After Oral and Intraperitoneal Administration to Swiss Mice. ACS Nano 2010, 4, 1481–1492.
- (22) Geng, J.; Kim, K.; Zhang, J.; Escalada, A.; Tunuguntla, R.; Comolli, L. R.; Allen, F. I.; Shnyrova, A. V.; Cho, K. R.; Munoz, D.; *et al.* Stochastic Transport Through Carbon Nanotubes in Lipid Bilayers and Live Cell Membranes. *Nature* 2014, *514*, 612–615.
- (23) Donkor, D. A.; Tang, X. S. Tube Length and Cell Type-Dependent Cellular Responses to Ultra-Short Single-Walled Carbon Nanotube. *Biomaterials* 2014, 35, 3121–3131.
- Kim, M.; Adamska, L.; Hartmann, N. F.; Kwon, H.; Liu, J.; Velizhanin, K. A.; Piao, Y.; Powell, L. R.; Meany, B.; Doorn, S. K.; *et al.* Fluorescent Carbon Nanotube Defects Manifest Substantial Vibrational Reorganization. *J. Phys. Chem. C* 2016, *120*, 11268–11276.
- Ma, X.; Adamska, L.; Yamaguchi, H.; Yalcin, S. E.; Tretiak, S.; Doorn, S. K.; Htoon, H. Electronic Structure and Chemical Nature of Oxygen Dopant States in Carbon Nanotubes. ACS Nano 2014, 8, 10782–10789.
- (26) Hartmann, N. F.; Velizhanin, K. A.; Haroz, E. H.; Kim, M.; Ma, X.; Wang, Y.; Htoon, H.; Doorn, S. K. Photoluminescence Dynamics of Aryl Sp 3Defect States in Single-Walled Carbon Nanotubes. ACS Nano 2016, 10, 8355–8365.
- (27) Hartmann, N. F.; Yalcin, S. E.; Adamska, L.; Haroz, E. H.; Ma, X.; Tretiak, S.; Htoon, H.; Doorn, S. K. Photoluminescence Imaging of Solitary Dopant Sites in Covalently Doped Single-Wall Carbon Nanotubes. *Nanoscale* **2015**, *7*, 20521–20530.
- (28) Cherukuri, T. K.; Tsyboulski, D. A.; Weisman, R. B. Length- and Defect-Dependent Fluorescence Efficiencies of Individual Single-Walled Carbon Nanotubes. ACS Nano 2012, 6, 843–850.
- (29) Hertel, T.; Himmelein, S.; Ackermann, T.; Stich, D.; Crochet, J. Diffusion Limited Photoluminescence Quantum Yields in 1-D Semiconductors: Single-Wall Carbon Nanotubes. ACS Nano 2010, 4, 7161–7168.
- (30) Oudjedi, L.; Parra-Vasquez, A. N. G.; Godin, A. G.; Cognet, L.; Lounis, B. Metrological Investigation of the (6,5) Carbon Nanotube Absorption Cross Section. J. Phys. Chem. Lett. 2013, 4, 1460–1464.
- (31) Iwamura, M.; Akizuki, N.; Miyauchi, Y.; Mouri, S.; Shaver, J.; Gao, Z.; Cognet, L.; Lounis, B.; Matsuda, K. Nonlinear Photoluminescence Spectroscopy of Carbon Nanotubes with Localized Exciton States. ACS Nano 2014, 8, 11254–11260.
- (32) Hell, S. W.; Sahl, S. J.; Bates, M.; Zhuang, X.; Heintzmann, R.; Booth, M. J.;

Bewersdorf, J.; Shtengel, G.; Hess, H.; Tinnefeld, P.; *et al.* The 2015 Super-Resolution Microscopy Roadmap. *J. Phys. D: Appl. Phys.* **2015**, *48*, 443001.

- (33) Cognet, L.; Tsyboulski, D. A.; Weisman, R. B. Subdiffraction Far-Field Imaging of Luminescent Single-Walled Carbon Nanotubes. *Nano Lett.* **2008**, *8*, 749–753.
- (34) Godin, A. G.; Lounis, B.; Cognet, L. Super-Resolution Microscopy Approaches for Live Cell Imaging. *Biophys. J.* **2014**, *107*, 1777–1784.
- (35) Bobroff, N. Position Measurement with a Resolution and Noise-Limited Instrument. *Rev. Sci. Instrum.* **1986**, *57*, 1152.
- (36) Mortensen, K. I.; Churchman, L. S.; Spudich, J. A.; Flyvbjerg, H. Optimized Localization Analysis for Single-Molecule Tracking and Super-Resolution Microscopy. *Nat. Methods* **2010**, *7*, 377–381.
- (37) Deng, S.; Zhang, Y.; Brozena, A. H.; Mayes, M. L.; Banerjee, P.; Chiou, W.-A.; Rubloff, G. W.; Schatz, G. C.; Wang, Y. Confined Propagation of Covalent Chemical Reactions on Single-Walled Carbon Nanotubes. *Nat. Commun.* 2011, 2, 382.
- (38) Zhang, Y.; Valley, N.; Brozena, A. H.; Piao, Y.; Song, X.; Schatz, G. C.; Wang, Y. Propagative Sidewall Alkylcarboxylation That Induces Red-Shifted Near-IR Photoluminescence in Single-Walled Carbon Nanotubes. J. Phys. Chem. Lett. 2013, 4, 826–830.
- (39) Godin, A. G.; Varela, J. A.; Gao, Z.; Danné, N.; Dupuis, J. P.; Lounis, B.; Groc, L.; Cognet, L. Single-Nanotube Tracking Reveals the Nanoscale Organization of the Extracellular Space in the Live Brain. *Nat. Nanotechnol.* 2017, *12*, 238–243.

SUPPORTING INFORMATION

METHODS

Ultrashort SWCNT preparation. usCNTs were prepared following the protocol described in ref¹. In brief, 30 mg of raw CoMoCAT nanotubes were dispersed in 10 mL of milli-Q water containing 0.3 wt% sodium deoxycholate (DOC) by pulsed tip sonication in an ice bath (output at 45 W, pulse 0.5 s and pause 0.2 s) for 5 h. After sonication, gentle centrifugation (30 min, 3500 rpm, Eppendorf Centrifuge 5804 R) was performed to remove the majority of bundles and insoluble materials. The supernatant (9 mL) was diluted to 20 mL with 0.3 wt% DOC solution and an ultracentrifugation was performed to remove any remaining bundles and catalyst particles (45000 rpm, i.e. 184000 g, 2 h, Thermo Scientific Sorvall WX+ Ultracentrifuge). This process was repeated twice, and the supernatant was collected for nanotube length separation by density gradient ultracentrifugation $(DGU)^2$. A density gradient solution was prepared via subsequently stacking four layers of 5 mL of 60 wt%, 10 wt%, 7.5 wt%, and 5 wt% iodixanol solution (OptiPrep, Sigma-Aldrich) in 0.3 wt% DOC solution, from bottom to top in a polycarbonate centrifugal tube (Beckman Coulter) with a volume of 26.3 mL. Iodixanol (60 wt%) was diluted in 0.3 wt% DOC solution to reach required concentrations. A layer of 5 mL freshly prepared sonication-cut SWCNT suspension was placed on top of the density-gradient iodixanol solution and centrifuged at 184,000g for 3h at 4 °C. After ultracentrifugation, SWCNTs with different lengths were distributed along the density gradient with the shortest usCNTs near the top. Fractioned samples were collected from top to bottom. Their lengths were characterized by AFM and photothermal imaging (Figure 1). For long-term storage, the concentration of DOC in all received samples was raised to 1 wt% to maintain individual dispersion of nanotubes.

Covalent functionalization of usCNTs. The usCNTs were covalently functionalized by perfluorinated hexyl groups³. $(-C_6F_{13})^3$. To an aqueous solution of usCNTs in 1% w/v sodium dodecyl sulfate-D₂O, 7.6 mM of NaHCO₃ (EMD chemicals, HPLC grade), 0.16 %v/v CH₃CN (Acros organics, HPLC grade, 99.9 %) and perfluoro hexyl iodide (Sigma Aldrich, 99%) were added. 3.6 mM of Na₂S₂O₄ (Sigma Aldrich, 85 %) was then added to the mixture and stirred at room temperature with protection from ambient light. After 24 hours of reaction, the SWCNT solutions were characterized by absorption and fluorescence spectroscopy. The absorption spectra were taken on a Lambda 1050 UV-vis-NIR spectrophotometer (Perkin Elmer) equipped

with both a photomultiplier tube and an extended InGaAs detector. The PL of f-usCNT solutions was collected using a NanoLog spectrofluorometer (HORIBA Jobin Yvon). The samples were excited with a 450 W Xenon source dispersed by a double-grating monochromator. The slit width of the excitation and emission beams were 10 nm. The PL spectra were collected using a liquid-N₂ cooled linear InGaAs array detector.

Near-infrared single nanotube PL imaging. Single SWCNT PL imaging was performed with an inverted microscope equipped with a 1.40 NA 60x objective and two detection arms separated by a 50/50 beam splitter (see Supplementary **Figure** S2). SWCNTs were excited in a widefield geometry with a 568 nm cw solid-state laser (Coherent Saphirre) at 0.5 kW/cm² otherwise stated (**Figure** 3). The excitation beam was circularly polarized light. A dichroic mirror (FF875-Di01, Semrock) in combination with a long-pass emission filter (RazorEdge 1064, Semrock) for (6,5) functionalized SWCNTs was used to illuminate and select E_{11} * PL. E_{11} PL of (6,5) SWCNTs was detected with an emCCD camera (Princeton Instruments ProEM), while E_{11} * PL of (6,5) functionalized SWCNTs was detected by an InGaAs camera (XEVA 1.7 320 TE3). For imaging, usCNTs were suspended in 1% w/v sodium dodecyl sulfate (SDS) and spin-coated on microscope slides at 3000 rpm for 3 minutes.

Photoluminescence intensity traces of blinking f-usCNTs. Movies of nanotubes blinking were recorded for up to 2000 images (integration times of 100 to 200 ms per frame). Intensity traces correspond to the integrated intensity of a given nanotube spot in the movie frames. For display, intensity traces were smoothed by the Savitsky-Golay filter to increase signal-to-noise ratios. Intensity steps displayed in **Figure** 4e were calculated by 2D Gaussian fitting of the spots appearing in image differences corresponding to the subtraction of images immediately before and after the step. Intensity steps correspond to the integral of the Gaussian fits using the software ImageJ.

Super-resolution imaging of E_{11}^* excitons in f-usCNTs. In order to localize E_{11}^* excitons in f-usCNTs, f-usCNTs displaying PL blinking were analyzed using the following procedure (see Supplementary Figure S5): (i) N images immediately before and after an intensity step were averaged to enhance signal-to-noise ratio (N varied between 5 and 30 depending on intensity steps). (ii) the resulting averaged images were subtracted to create a differential image isolating a blinking event from one emitting defect. The absolute PL values are taken into account in case of a positive blinking event to equally analyze positive and negative steps. (iii) Each spot

in a differential image was then fitted by a 2D Gaussian profile with the plug-in GDSC SMLM (ImageJ), as used in single molecule localization microscopy,⁴⁻⁶ to retrieve the emitter localization with sub-diffraction precision. (iv) for visualization (**Figure** 5 and Supplementary **Figure** S6b-c), super-resolved images are produced from the individual defect localizations displayed as color-coded two-dimensional Gaussians of unit amplitude and width equal to the precision of localization. Localization precision of each event was calculated taking into account signal-to-noise ratio of the blinking event fitted by the 2D Gaussian at step (iii) in the initial differential image, as well as the pixel size in the image as described in ref.⁷. In Supplementary **Figure** S6c, each individual defect detection is also displayed as a red spot. (v) In order to identify individual emitting defects that might be detected several times through multiple blinking events, adjacent detections that were situated within the range of neighboring localization precisions (white dotted circle in Supplementary **Figure** S6e) and displayed with the corresponding localization precision in final images (blue circles in **Figure** S6e) and displayed with the corresponding localization precision in final images (blue circles in **Figure** S6a). The average localization precision was equal to 22 nm.

- Gao, Z.; Oudjedi, L.; Faes, R.; Moroté, F.; Jaillet, C.; Poulin, P.; Lounis, B.; Cognet, L. Optical Detection of Individual Ultra-Short Carbon Nanotubes Enables Their Length Characterization Down to 10 Nm. Sci. Rep. 2015, 5, 17093.
- (2) Sun, X.; Zaric, S.; Daranciang, D.; Welsher, K.; Lu, Y.; Li, X.; Dai, H. Optical Properties of Ultrashort Semiconducting Single-Walled Carbon Nanotube Capsules Down to Sub-10 Nm. J. Am. Chem. Soc. 2008, 130, 6551–6555.
- Kwon, H.; Furmanchuk, A.; Kim, M.; Meany, B.; Guo, Y.; Schatz, G. C.; Wang, Y.
 Molecularly Tunable Fluorescent Quantum Defects. J. Am. Chem. Soc. 2016, 138, 6878–6885.
- Cognet, L.; Tsyboulski, D. A.; Weisman, R. B. Subdiffraction Far-Field Imaging of Luminescent Single-Walled Carbon Nanotubes. *Nano Lett.* 2008, 8, 749–753.
- (5) Godin, A. G.; Lounis, B.; Cognet, L. Super-Resolution Microscopy Approaches for Live Cell Imaging. *Biophys. J.* 2014, 107, 1777–1784.
- Sage, D.; Kirshner, H.; Pengo, T.; Stuurman, N.; Min, J.; Manley, S.; Unser, M. Quantitative Evaluation of Software Packages for Single-Molecule Localization Microscopy. *Nat. Methods* 2015, *12*, 717–724.
- Mortensen, K. I.; Churchman, L. S.; Spudich, J. A.; Flyvbjerg, H. Optimized Localization Analysis for Single-Molecule Tracking and Super-Resolution Microscopy. *Nat. Methods* 2010, 7, 377–381.



Figure S1 | The absorption spectra of 43 nm long SG65i CoMoCAT usCNTs before (black) and after functionalization by perfluorinated hexyl defects (red).



Figure S2 | Single nanotube luminescence microscope. Dual-detection single nanotube microscope for the detection of E_{11} and E_{11}^* PL emission of (6,5) f-usCNTs. The spectral bands of each detection arm are indicated.



Figure S3 | Dual-detection of single (6,5) f-usCNTs in E_{11}^* (a) and E_{11} (b) PL detection channels. On occasion (e.g. second row), weak E_{11} PL is detected and is colocalized with E_{11}^* PL as shown in composite images in (d). Red: E_{11}^* and green: E_{11} channels. For creating composite images in (d), images from the E_{11} channel have been rescaled in (c) to match pixel sizes of images in (a). Integration times: 200 ms. Scale bar = 5 μ m.



Figure S4 | Characterization of the SWCNTs used in Fig. 3e. a, Cumulative distribution of SWCNT lengths measured by AFM (N=104). b, Distribution of SWCNT absorption signals measured by PhI (N=244) which also reflects the SWCNT length distribution. c, E_{11} PL image of single SWCNTs. Integration time: 200ms. Scale bar: 5µm. See Fig. 1a,b&d for comparison to usCNTs.



Figure S5 | Super-localization of a single defect (see methods).



Figure S6 | Super-resolved imaging of E_{11}^* exciton localization in f-usCNTs. a, Differential image revealing a blinking event from E_{11}^* PL emission of a single (6,5) f-usCNT. b, Corresponding superresolved image produced from 6 individual defect localizations displayed as color-coded twodimensional Gaussian distributions of unit amplitude and widths equal to the precision of each localization. c, Same as b. but with zoomed scale. The localization of each of the 6 detections is indicated by a red spot. d, The 6 localizations are represented with the precision of their localization. e, The blue dots represent the two emitting defect sites at the origin of the blinking events detected in a-d. The localization precisions of the emitting defects are indicated by blue circles. f, Super-resolved image of the two defect sites with their localizations as revealed by the procedure presented in a-e.