

## Innovative strategies for intervertebral disc regenerative medicine: From cell therapies to multiscale delivery systems

Nina Henry, Johann Clouet, Jean Le Bideau, Catherine Le Visage, Jérôme Guicheux

### ► To cite this version:

Nina Henry, Johann Clouet, Jean Le Bideau, Catherine Le Visage, Jérôme Guicheux. Innovative strategies for intervertebral disc regenerative medicine: From cell therapies to multiscale delivery systems. Biotechnology Advances, 2018, 36 (1), pp.281 - 294. 10.1016/j.biotechadv.2017.11.009 . hal-01718005

## HAL Id: hal-01718005 https://hal.science/hal-01718005

Submitted on 3 Mar 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Innovative strategies for intervertebral disc regenerative medicine: from cell therapies to multiscale delivery systems

Nina Henry<sup>a,b,c</sup>, Johann Clouet<sup>a,c,d,e</sup>, Jean Le Bideau<sup>b</sup>, Catherine Le Visage<sup>a,c\*</sup>, Jérôme Guicheux<sup>a,c,f\*</sup>.

- a. Inserm, UMR 1229, RMeS, Regenerative Medicine and Skeleton, Université de Nantes, ONIRIS, Nantes, France
- <sup>b.</sup> Institut des Matériaux Jean Rouxel (IMN), Université de Nantes, CNRS, 2 rue de la Houssinière, BP 32229, 44322 Nantes cedex 3, France
- <sup>c.</sup> Université de Nantes, UFR Odontologie, Nantes, F-44042, France.
- d. CHU Nantes, Pharmacie Centrale, PHU 11, Nantes, France.
- e. Université de Nantes, UFR Sciences Biologiques et Pharmaceutiques, Nantes, France.
- f. CHU Nantes, PHU 4 OTONN, Nantes, France.

#### \* These authors equally contributed to this work.

Correspondence to:

C. Le Visage and J. Guicheux

INSERM U1229, 1 place Alexis Ricordeau, 44042 Nantes cedex 1, France.

Catherine.levisage@inserm.fr; jerome.guicheux@inserm.fr

1. Abstract	
2. Outlook section	
3. Introduction	
4. IVD degeneration physiopathology	
4.1. IVD structure	4
4.2. Extracellular matrix anabolism/catabolism homeostasis	5
5. From physiopathology to IVD specific biotherapies	6
5.1. Cell-based therapies	6
5.2. Molecular therapies	7
5.3. Gene and interfering RNA therapies	9
6. Engineering biomaterials for IVD delivery	10
6.1. Natural and synthetic polymers	
6.2. In situ forming hydrogels as macro-scaled delivery systems	11
6.3. Micro-scale delivery systems	
6.4. Nano-scale delivery systems	
6.5. Multiscale systems for complex regenerative strategies	
7. Conclusion	15
8. Acknowledgements	15
9. Figures legends	15
10. References	16

#### 1 1. Abstract

2 As our understanding of the physiopathology of intervertebral disc (IVD) degeneration has improved, 3 novel therapeutic strategies have emerged, based on the local injection of cells, bioactive molecules, and 4 nucleic acids. However, with regard to the harsh environment constituted by degenerated IVDs, protecting 5 biologics from in situ degradation while allowing their long-term delivery is a major challenge. Yet, the 6 design of the optimal approach for IVD regeneration is still under debate and only a few papers provide a 7 critical assessment of IVD-specific carriers for local and sustained delivery of biologics. In this review, we 8 highlight the IVD-relevant polymers as well as their design as macro-, micro-, and nano-sized particles to 9 promote endogenous repair. Finally, we illustrate how multiscale systems, combining in situ-forming 10 hydrogels with ready-to-use particles, might drive IVD regenerative medicine strategies towards 11 innovation.

12

16

17

#### 13 2. Outlook section

- Intervertebral disc (IVD) regenerative medicine could benefit from the emergence of cell,
   molecule-, gene- and siRNA-based strategies.
  - Design of IVD-specific carriers to protect biologics from in situ degradation, while allowing their local and sustained delivery, is a major challenge.
- A critical assessment of IVD-relevant natural and synthetic polymers, and their design as macro-,
   micro-, and nano-sized particles is provided.
- Multiscale systems, combining in situ forming hydrogels with ready-to-use particles, are introduced as new opportunities to stimulate IVD regeneration.

22

#### 23 3. Introduction

24 Low back pain (LBP) is a common health condition and it is now well established that over 80% of adults

25 will experience LBP during their lifetime. LBP has become the second most frequent cause of hospital

visits and the leading cause of years lived with disability (Maher et al., 2017; Vos et al., 2016). Thanks to

a better understanding of the etiology of LBP, the involvement of intervertebral disc (IVD) degeneration,

28 referred to as disc degenerative disease (DDD) (Vos et al., 2016), was identified.

29 Current treatments of LBP include nonpharmacological, pharmacological, or surgical approaches, all

30 aimed at controlling pain. For early degenerative stages, pharmacotherapies are mainly used (Abdel

31 Shaheed et al., 2016) in association with rehabilitation or education of the patient (Latimer et al., 2004).

32 For more advanced cases, surgical approaches are considered if conservative methods fail. Spine fusion

aims to directly remove the IVD and immobilize the adjacent vertebrae. However, this intervention
 reduces spine mobility and neighboring segments can be affected by an accelerated degenerative process

35 (Baliga et al., 2015). Arthroplasty, with *Nucleus pulposus* or IVD total replacement, aims at conserving

the spine motion notably at the affected levels. These interventions, although generally clinically effective

in alleviating pain (Galbusera et al., 2008), are very invasive and often include a long postoperative

38 recovery and a non negligible risk of complications (Baliga et al., 2015).

39 Recent progress in our understanding of IVD physiopathology has led to new approaches for IVD

40 regenerative medicine. In view of the decreasing number of cells in degenerating IVD, cell-based

41 therapies were studied with growing interest. However, owing to the restrictive regulatory framework and

42 the difficult clinical translatability of cell-based therapies, the use of biological factors targeting IVD

degenerative processes was also contemplated. More recently, the role of gene mutations or microRNA
 dysregulation was identified at the onset of DDD, leading to the development of a third therapeutic

dysregulation was identified at the onset of DDD, leading to the development of a third therapeutic
 approach based on the delivery of genes or small interfering RNAs (siRNA). In this context, this review

46 will first briefly give recent insights into the pathophysiological mechanisms of DDD and then focus on

47 describing cell-, molecule-, gene-, and siRNA-based strategies for tackling DDD as well as the scaffolding

- 48 biomaterials used for their respective delivery.
- 49
- 50

#### 51 4. IVD degeneration physiopathology

#### 52 **4.1. IVD structure**

53 IVDs are fibro-cartilaginous tissues located between the vertebrae that are avascular and non-innervated

54 (Whatley and Wen, 2012). They act as shock absorbers by distributing the mechanical loads along the

spine and facilitate trunk mobility (Whatley and Wen, 2012; Zhou et al., 2014). IVDs are generally

56 presented as a tripartite anatomical entity composed of the following parts (Figure 1).

57 The *Nucleus pulposus* (NP) is the inner part and the most hydrated region of the IVD (Fontana et al.,

58 2015). NP cell density in human is the lowest of the IVD (Maroudas et al., 1975). These cells can be

59 classified into two types: notochordal cells and nucleopulpocytes (Colombier et al., 2014). Human adult

60 NP cells produce a highly hydrated extracellular matrix (ECM), rich in proteoglycans (mainly aggrecan)

61 and type II collagen.

62 The Annulus fibrosus (AF) is the outer part of the IVD. It can be divided into the outer AF containing a

63 oriented lamellar array of densely packed collagen fibers, while the inner AF is less dense and less

organized (Raj, 2008; Whatley and Wen, 2012). AF cells have a fibroblastic morphology (Sakai and Grad,

65 2015). The AF is composed of water, types I and II collagen fibers, proteoglycans, and other proteins

66 (Roughley, 2004). The AF retains the pressurized NP and allows for disc compression/decompression

67 (Fontana et al., 2015).

68 **The endplates** (EPs), composed of hyaline cartilage at the top and bottom of the vertebral bodies,

69 interface the IVDs and the adjacent vertebrae. EPs cells are chondrocytes that synthesize an ECM rich in

70 proteoglycans, types I and II collagens, and water. Mechanically, EPs support significant loads during

71 daily-living activities and distribute the intradiscal pressures onto the adjacent vertebrae. They represent 72 the main route for glucose and oxygen to reach the disc cells through the vertebral capillaries and for

73 waste products to be eliminated from the disc (Fontana et al., 2015; Lotz et al., 2013).

74

96

#### 4.2. Extracellular matrix anabolism/catabolism homeostasis

97 DDD has a complex and multifactorial etiology. Genetics, environmental causes, mechanical factors, 98 aging, as well as systemic and toxic factors (Elmasry et al., 2015; Hadjipavlou et al., 2008) were identified 99 as risk factors. Interestingly, recent data highlighted that the concomitant presence of notochordal cells 100 and nucleopulpocytes was a pivotal factor for IVD ECM homeostasis (Erwin et al., 2006). Indeed, these 101 two cell types interact via synthesized molecules and the early disappearance of notochordal cells is 102 associated with a downregulation of nucleopulpocyte survival. This decrease in nucleopulpocyte density

103 leads to a misbalance in the ECM anabolism/catabolism ratio that culminates in the alteration of the

- 104 mechanical properties of IVDs (Erwin et al., 2011). In the advanced stages of IVD degeneration,
- 105 proteoglycans, hydration, and the type-II/type-I collagen ratio are decreased(Russo et al., 2016; W.-J.
- 106 Wang et al., 2015), leading to a dehydrated and fibrotic NP with a solid-like mechanical behavior (Russo
- 107 et al., 2016). Additionally, the AF structure is disorganized and EPs calcify (Fontana et al., 2015; W.-J.

108 Wang et al., 2015). As the degeneration worsens, innervation and vascularization of the disc occur

109 (Whatley and Wen, 2012), leading to discogenic pain (Alkhatib et al., 2014).

110 From a molecular point of view, IVD degeneration is associated with an upregulation of pro-inflammatory

- 111 cytokines such as interleukin-1 $\beta$  (IL-1 $\beta$ ) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) (Sakai and Grad, 2015).
- These two molecules trigger the over-expression of matrix metalloproteinases (MMPs) notably MMP-1, -2, -3, -7, -8, and -13. These MMPs are known for their proteolytic action toward collagen and
- proteoglycans (Kepler et al., 2013b; W.-J. Wang et al., 2015). Similarly to MMPs, there is a marked
- 115 increase in the a disintegrin and metalloprotease with thrombospondin motifs (ADAMTS) 4 and 5.
- ADAMTS 4 and 5 act as aggrecanases, thereby contributing to worsening ECM degradation (Molinos et
- al., 2015; W.-J. Wang et al., 2015). As a pro-inflammatory cytokine, IL-1β up-regulates the expression of
- 118 vascular endothelial growth factor (VEGF), nerve growth factor (NGF), and brain-derived neurotrophic
- factor (BDNF), thereby triggering the neo-vascularization and neo-innervation of IVD that ultimately
- result in inflammation and pain (Alkhatib et al., 2014; Whatley and Wen, 2012). Finally, in the presence of TNF- $\alpha$  or IL-1 $\beta$ , NP cells tend to secrete chemo-attractant molecules including (C-C motif) ligand
- 121 of TNF-a of TL-TP, NP cens tend to secrete chemo-attractant molecules including (C-C motif) ligand 122 5/regulated on activation of normal T cells expressed and secreted (CCL5/RANTES) or chemokine (C-X-
- 123 C motif) ligand 6 (CXCL6) (Grad et al., 2016), thus increasing the migration of mesenchymal stem cells
- 124 (MSCs) (Pattappa et al., 2014). This increased secretion of CCL5 was correlated with painful degenerated
- 125 IVDs (Kepler et al., 2013a). In addition, stromal cell-derived factor-1 (SDF-1) was over-expressed in
- degenerated IVDs and involved in NP cell apoptosis (Z. Liu et al., 2016). Interestingly, SDF-1 also
- demonstrated a chemo-attracting effect toward MSC migration in an ex vivo IVD culture model (Pereira
- 128 et al., 2014).
- 129 In contrast to these catabolic factors, anabolic factors stimulate the biological cascade involved in
- 130 regenerative processes. Among them, Transforming Growth Factor- $\beta$  (TGF- $\beta$ ) is known to play a pivotal
- role in collagen and proteoglycan synthesis and in ECM homeostasis (S. Wang et al., 2015). TGF- $\beta$  is also
- 132 one of the most potent inhibitors of  $TNF-\alpha$ -induced upregulation of MMPs as well as of aggrecan and
- type-II collagen degradation (Yang et al., 2015). In parallel, Bone Morphogenic Proteins (BMPs) are
- known to increase proteoglycan and type-II collagen synthesis as well as cell proliferation and
- differentiation (S. Wang et al., 2015). Among them, Growth and Differentiation Factor-5 (GDF-5 or
- BMP-14) plays a crucial role during musculoskeletal development and postnatal IVD maintenance (Feng

137 et al., 2015). GDF-5 gene mutations notably lead to disc abnormalities in mice (Li et al., 2004). GDF-5

also upregulates the proliferation and secretory activities of NP and AF cells (Chujo et al., 2006) and

139 modulates MMPs, IL-1 $\beta$ , or TNF- $\alpha$  expression levels (Enochson et al., 2014; Gruber et al., 2014a).

140 Altogether, these data highlight the pivotal role of GDF-5 in IVD matrix homeostasis.

141 The ECM homeostasis of IVDs and the consequences associated with a homeostasis rupture are illustrated142 in Figure 2.

143

#### 144 5. From physiopathology to IVD specific biotherapies

Regarding the multifactorial nature of DDD, three main therapeutic strategies were considered in the pastfew years: cell-, molecular-, and nucleic acid-based therapies (Figure 3).

#### 147 **5.1. Cell-based therapies**

148 A converging body of evidence suggested that IVD degeneration originates from the early disappearance 149 of IVD resident cells. Supplementation of the IVD with regenerative cells thus naturally emerged as a 150 strategy. Repopulating a degenerated IVD with healthy cells aims at either (i) restoring tissue homeostasis 151 by transplanting cells able to secrete a functional ECM or (ii) tackling in situ inflammation by transplanting stem cells with immune-suppressive and anti-inflammatory properties. To achieve these 152 ambitious and clinically relevant objectives, various cells sources were considered. Logically, NP cells 153 154 harvested from surgically removed IVDs were the first to be used, with promising results in a canine study 155 and in human clinical trials (Meisel et al., 2007). However, their further use was dramatically hampered 156 by the low cell number harvested (Sakai and Grad, 2015). In parallel, pluripotent stem cells such as embryonic stem cells could be of great interest in view of their theoretical potential to differentiate into 157 158 NP cells. Nevertheless, legal and ethical controversies limit the use of these cells for clinical application. 159 Induced pluripotent stem (iPS) cells were also considered for various applications in regenerative 160 medicine (Hirshi et al., 2014) and their suitability for IVD repair is being studied (Chen et al., 2013) but further studies are needed before their potential can be judged. Last but not least, adipose- (ASCs) or bone 161 162 marrow-derived (MSCs) mesenchymal stem cells in their undifferentiated state are likely the most widely 163 used in the DDD context. In addition to providing us with promising results (see next section), the 164 extensive use of MSCs raised the question of their allogeneic or autologous origin (Richardson et al., 165 2016). Indeed, MSCs are considered hypo-immunogenic and immune-evasive cells able to be transplanted across major histocompatibility barriers (Ankrum et al., 2014), making the use of allogeneic MSCs a 166 167 credible alternative to autologous ones.

168 ASCs are considered particularly advantageous because of their abundance in fat tissue, their accessibility, 169 and their more potent immuno-modulatory effects in comparison with MSC isolated from bone marrow 170 (Li et al., 2015). Of interest, human ASCs exhibit a nucleopulpogenic potential when committed with TGF-β1 and GDF-5 (Colombier et al., 2016). When implanted in mice, these cells also demonstrated their 171 172 ability to secrete GAGs and type II collagen, consistently with native NP cell capacities (Colombier et al., 2016). In rabbits, undifferentiated MSCs injected in the NP inhibited disc degeneration by reducing MMP-173 174 2, MMP-3, and MMP-13 expression and by promoting type-II collagen synthesis, resulting in the 175 maintenance of the IVD structure (Miyamoto et al., 2010). The beneficial outcomes of injecting 176 undifferentiated MSCs were further demonstrated in other animal models, such as rat, mouse, dog, or pig models (Henriksson et al., 2009; Hiyama et al., 2008; Jeong et al., 2009; Marfia et al., 2014; Richardson et 177 178 al., 2016). In these studies, IVD degeneration was significantly slowed down, disc height and MRI signal 179 intensity improved, and nucleopulpogenic ECM neo-synthesis was observed. In humans, the intradiscal 180 injection of MSCs could lower pain and disability (Orozco et al., 2011). This relief was associated with 181 the anti-inflammatory (Maria et al., 2016; Sakai and Grad, 2015) and pro-anabolic properties of MSCs 182 (Shabbir et al., 2009). Several ongoing clinical trials are conducted worldwide to determine the optimal cell source (autologous or allogeneic) to be used in DDD (Sakai and Andersson, 2015). 183

184 Despite their global promise, the intradiscal injection of "naked" MSCs remains hampered by the harsh

185 environmental condition the cells experience when injected into a degenerated IVD (low oxygen tension,

low nutrient level, metabolic product accumulation, acidic pH, high osmolarity, and high intradiscal
 pressure). These stringent conditions are responsible for the poor survival rate and altered activity of the

188 injected cells. To overcome this hurdle, tissue engineers proposed to use cytoprotective 3D scaffolds able

- 189 to create a biomimetic environment that could support cell survival. These scaffolds could additionally
- 190 promote cell retention at the injection site thereby limiting the risk of off-target effects related to cell
- 191 effusion. These scaffolds will be detailed in the last section of this review.
- 192

#### **5.2. Molecular therapies**

More recently, and facing the numerous limitations of cell transplantation, an alternative to cell-based therapies emerged. It consists in exploiting the ability of chemoattractant molecules to mobilize and recruit endogenous progenitor cells at the degenerated site (Illien-Jünger et al., 2012). This strategy, based on the capability of the endogenous body to self-regenerate, could free clinicians from the restrictive regulatory framework and cost ineffectiveness of cell transplantation. With respect to the myriad of factors

198 regulatory framework and cost memetriveness of cen transplantation. With respect to the mynad of fact 199 playing a crucial role in IVD physiopathology, many chemoattractant, pro-anabolic, and anti-catabolic

200 molecules were contemplated for the stimulation of resident cells or the attraction/recruitment of

- 201 endogenous progenitors.
- 202

203 One key factor for improving the endogenous repair of IVDs via cell attraction/recruitment is the presence 204 of endogenous progenitors with stem cell properties in the IVD (Risbud et al., 2007). These IVD-derived 205 progenitor cells (either from NP or EPs) are very similar to MSCs isolated from the bone marrow of the 206 same patient (Blanco et al., 2010). Interestingly, analyses of the migration capacity of progenitors located 207 in the NP, AF, and EP indicate that EP progenitors have the highest migration and invasion potency (S. 208 Liu et al., 2016). To favor cell migration in degenerative IVDs, chemokines were studied since, as 209 chemotactic cytokines, they are involved in cell mobilization, attraction, and homing. Regarding IVDs, the 210 three main chemokines expressed are SDF-1, CCL5, and CXCL16, both in healthy or degenerated IVDs. Interestingly, the expression of SDF-1 and CXCR4 was upregulated in degenerated IVDs, notably in 211 212 MSCs isolated from the EPs (S. Liu et al., 2016; Zhang et al., 2014). SDF-1 also increased MSC attraction 213 in a nucleotomized disc model (Pereira et al., 2014). The implication of SDF-1 in stem cell attraction was

also demonstrated in many tissues including myocardium or periodontal ligament (Kaku et al., 2017;

- 215 Purcell et al., 2012). Recently, IVD cells stimulated with IL-1 $\beta$  and TNF- $\alpha$  were found to secrete a higher
- 216 concentration of CCL5 compared with IVD cells cultured under physiological conditions (Gruber et al.,
- 217 2014b). The presence of CCL5 in degenerated IVDs as well as herniated and scoliotic discs was further
- 218 demonstrated by proteomic assays in other in vitro studies in humans (Kawaguchi et al., 2002; Kepler et al., 2013a). Finally, in AF cells, CXCL16 is up-regulated by IL-1 $\beta$ , suggesting a role for the
- 220 CXCL16/CXCR6 system (Gruber et al., 2016).
- Altogether, these data highlight the role of chemokines in the degenerative process of IVDs and strengthen
   their relevance in developing new strategies to attract progenitor cells and stimulate endogenous repair
- 223 (Illien-Jünger et al., 2012).
- 224
- In line with our understanding of DDD, anabolic factors can contribute to IVD regeneration. A large panel
   of anabolic factors was contemplated for IVD regeneration. Here, we mainly focus on the factors that
   demonstrated promising results in several reports.
- 228 TGF-β contributes toward maintaining the expression levels of connective tissue growth factor (CCN2),
- one of the most potent factors able to stimulate aggrecan synthesis, in NP cells (Tran et al., 2014). In
  - addition, CCN2 suppressed the inductive effect of IL-1 $\beta$  on catabolic genes (Tran et al., 2014).

231 Considering that pro-inflammatory cytokines decreased CCN2 expression, this makes TGF- $\beta$  an anabolic

- 232 factor able to counteract the effects of pro-inflammatory cytokines (Bedore et al., 2014). TGF- $\beta$  was also
- 233 used in association with dexamethasone to stimulate the proliferation of degenerated human NP cells and
- 234 exhibit an anti-catabolic gene expression profile (Liang et al., 2013).

235 GDF-5 is a major factor involved in early IVD development, and mutations of its related gene lead to

236 severe structural impairment (Feng et al., 2015). Noteworthy, GDF-5 is down-regulated by IL-1 $\beta$  and

237 TNF- $\alpha$ . In vitro, GDF-5 favors cell proliferation and stimulates the synthesis of proteoglycan and type II

- 238 collagen, thus demonstrating its anabolic activity (Chujo et al., 2006). GDF-5 was used to differentiate human ASCs into nucleopulpocytes, in association with TGF-B1 (Colombier et al., 2016). Interestingly, 239
- intradiscal injection of GDF-5 in rats and rabbits was shown to increase disc height. Consequently, clinical 240
- 241 trials were conducted to evaluate the safety and efficacy of intradiscal injection of GDF-5 in patients with
- DDD (NCT01158924, NCT00813813, NCT01182337, and NCT01124006 on clinicaltrials.gov). As far as 242
- 243 we know, no peer-reviewed publication is currently available. While a modest improvement in clinical
- parameters was observed, some adverse events were also reported suggesting that despite the relevance of 244
- 245 GDF-5 in DDD, its direct intradiscal injection may not be the most suitable route.

246 Other anabolic factors were investigated and their anti-apoptotic activity toward NP cells was evidenced. 247 Indeed, aside from promoting angiogenesis, it is now clear that in IVD, vascular endothelial growth factor

248 (VEGF) also prevents NP cell apoptosis (Tran et al., 2010). Additionally, insulin-like growth factor (IGF-

249 1), platelet-derived growth factors (PDGFs), epidermal growth factor (EGF), and fibroblast growth factor

250 (FGF-2) stimulate cell proliferation and inhibit cell apoptosis in human and animal cells (Vasiliadis and

Pneumaticos, 2014). Together with the aforementioned CCN2 (Tran et al., 2014), these factors 251 252 demonstrated their anabolic activity.

253 Regarding the role of anabolic factors in DDD treatment, the use of platelet-rich plasma (PRP), which

254 contains a myriad of multifunctional growth factors, was recently proposed. Two clinical trials suggested 255

the efficacy of PRP in DDD (Kristin et al., 2017; Tuakli-Wosornu et al., 2016). Further double-blind and

- 256 controlled clinical trials are now required to confirm these first results.
- 257
- 258 Recent studies highlighted the key role of both TNF- $\alpha$  and interleukins in inflammation and further IVD 259 degeneration, notably via the up regulation of MMPs and aggrecanases.

260 Thus, anti-TNF- $\alpha$  treatments (infliximab) were proposed and showed a significant decreased expression of

- 261 IL-1β and IL-6 in NP cells (Walter et al., 2015), thereby demonstrating their anti-catabolic activity. In
- addition, the local administration of anti-IL-6 receptor, such as atlizumab or tocilizumab, lowered 262
- 263 inflammation-related pain in humans (Sainoh et al., 2016). Interestingly, human IVD cells overexpressing 264 IL-1 receptor antagonist were able to lower an IL-1-dependent increase of MMPs when injected into IVD
- 265 degenerative explants, demonstrating their anti-catabolic potential (Le Maitre et al., 2007). In parallel,
- 266 because NF-kB increases IVD degeneration via the upregulation of pro-inflammatory cytokines (Fontana
- 267 et al., 2015), studies examined its inhibition. Indeed, human NP cell exposure to NF-κB inhibitor led to
- 268 significantly decreased IL-1β-dependent MMPs and ADAMTS upregulation (Orita et al., 2013; Zhongyi
- et al., 2015). In addition, inhibition of other cytokines such as BDNF, NGF, VEGF, or TNF-α was studied 269
- 270 and showed promising results in the context of inflammation-associated DDD (Orita et al., 2011; Walter et
- al., 2015; Wu et al., 2011) 271
- 272 In parallel to the previously described IVD degenerative processes, aging-associated accumulation of
- 273 oxidative damage is of great interest. Collagen is subject to glycoxidation reactions that form advanced-
- 274 glycation end-products (AGEs). The accumulation of these AGEs molecules contributes to IVD
- 275 degeneration by increasing TNF- $\alpha$ , MMPs, and aggrecanases expression, and to a greater extent AGEs
- 276 participate in the alteration of structural and mechanical properties of the ECM (Colombini et al., 2008).
- 277 Interestingly, administration of an anti-inflammatory drug (pentosan-polysulfate) associated with anti-

278 AGE molecules (pyridoxamine) alleviated NP degeneration by lowering MMPs and ADAMTS production 279 in diabetic mice (Illien-Junger et al., 2013).

280

281 With the discovery of molecules involved in IVD degenerative processes, new therapeutic approaches 282 were proposed, either through stimulating anabolism or inhibiting catabolism. Despite encouraging 283 results, the short half-life of such molecules does not allow for long-term activity, and they may not be sufficient to reactivate the physiological IVD machinery. Whether embedding these molecules in 284 285 biomaterials to allow their in situ sustained and controlled delivery, while protecting them from 286 proteolytic degradation, is a good option will be discussed in the next section.

287

#### 5.3. Gene and interfering RNA therapies 288

289 An alternative to biotherapies may be the genetic modification of resident cells through gene transfer, for 290 which two strategies can be envisioned: (i) The vector containing the gene can be introduced in the body 291 (direct or in vivo gene therapy) or (ii) target cells can be harvested, genetically modified in vitro, and re-292 implanted (indirect or ex vivo gene therapy) (Goldspiel et al., 1993). Considering the decreased number of 293 cells in degenerative IVD, direct gene therapy could be of interest for early degeneration, while the 294 indirect approach seems more suitable for more severe cases. Two carriers were used for gene delivery: 295 non-viral and viral vectors. Non-viral gene therapy refers to the use of naked DNA or cationic compounds 296 (liposomes, polyplexes, lipopolyplexes) (Daraee et al., 2014; Rezaee et al., 2016). Such gene carriers help 297 avoid the risks of mutagenicity, viral infections, and viral immunogenicity (Mintzer and Simanek, 2009). 298 However, their application in IVD degeneration treatment remains limited. Regarding viral vectors, while 299 retroviruses are the most commonly used, they have a more potent action on cells with a high dividing rate 300 making them poorly adapted for NP-cell-mediated IVD gene therapy (Miller et al., 1990). Many authors 301 thus chose adenoviruses despite their potential infectious and immunogenic risk owing to their higher 302 transduction efficiency notably in quiescent or slowly dividing cells. Therapeutic genes encoding for antiinflammatory agents or MMPs inhibitors (e.g., TIMP-1) were considered as they could drastically reduce 303 304 the expression of MMPs involved in ECM degradation (W.-J. Wang et al., 2015). It was found that the use of adenoviral vectors coding for BMP-2 and TIMP-1 on human degenerated disc cells increased 305 306 proteoglycan synthesis (Wallach et al., 2003). Adenoviruses were also used to deliver genes coding for 307 growth factors such as GDF-5 (X.-W. Luo et al., 2016) or TGF- $\beta$  (Nishida et al., 1999) and they were 308 shown to efficiently stimulated anabolism.

309 In parallel to classic gene therapies, there is growing interest in the use of siRNAs. SiRNAs are short non-

310 coding sequences of 20-25 nucleotides that control gene expression by binding to their target mRNA and

311 causing the repression of RNA translation or degradation (Bartel, 2004). Many miRNAs deregulations

312 emerged as key players in IVD degeneration by triggering NP cell apoptosis, ECM degradation, or NP cell proliferation and inflammatory response (see (Zhou et al., 2017) for review) and can thus be envisioned as

- 313
- 314 new therapeutic targets.
- 315 Gene- and RNA interfering-based therapies seem to be particularly interesting for restoring the

316 anabolic/catabolic balance in IVDs. However, as NP cell density drastically decreases during

317 degeneration, the effectiveness of such approaches needs to be clarified. Finally, adenoviruses or siRNAs

318 are not integrative tools and do not lead to extended gene expression. In addition, as siRNAs need to be

319 protected from endonuclease degradation, the use of specifically designed biomaterial-based systems, as

320 mentioned previously for peptidic molecules, could be a reasonable option.

321

- 322 As stated above, the broadening of our understanding of IVD degenerative processes led to new
- 323 therapeutic strategies based on the local administration of cells, molecules, genes, or siRNA and several
- 324 clinical trials assessing these three main therapeutic strategies have been reported (Table 1). As of

November 2017, eight clinical trials have been published using autologous or allogenic cells, four trials

have reported the injection of anabolic factor GDF-5 and only one trial is based on PRP. To the best of our

327 knowledge, there is no ongoing clinical trial based on gene or RNA interfering therapies for IVD

328 regeneration. Despite encouraging results, these therapies need further improvement with regard to the

harsh environment in which these biologics are injected. Thus, biomaterials were considered so as to

improve cell survival and protect molecules and nucleic acids from degradation. These biomaterials could additionally allow the biologics to be delivered in a spatiotemporally controlled manner thereby promoting

- 332 long-term IVD regeneration.
- 333

#### 334 6. Engineering biomaterials for IVD delivery

Successful delivery of fragile biologics into IVDs is highly challenging, mostly because of their short in
 vivo half-life. With the aim of targeting this poorly vascularized tissue, delivery routes are limited and

thus minimally invasive direct injection received attention. While there are reviews extensively describing

pharmaceutical innovations, including self-setting hydrogels, microspheres, and liposomes, for the oral

(Diab et al., 2017), the intra-vitreal (Bochot and Fattal, 2012), and the transdermal (Naves et al., 2017)

340 routes, only few papers are available providing a critical assessment of IVD-specific approaches for local 341 and sustained delivery. Here, we highlight the rational selection of natural or synthetic polymers, we

exemplify the design of delivery systems based on macro-, micro-, and nano- sized properties, and we

343 illustrate how multiscale combined systems might help direct IVD regenerative medicine strategies.

344

#### **6.1. Natural and synthetic polymers**

One key parameter in the design of delivery systems is the selection of the constitutive polymer, since the chemical functionalities of a polymer affect all aspects of its performance, including encapsulation efficiency as well as polymer degradation rate and release (Mitragotri et al., 2014). As such, recent advances in delivery systems are based on polymers tailored for a specific tissue and engineered to exert distinct biological functions.

351 Among natural polymers, collagen, hyaluronic acid (HA), alginate, and chitosan were extensively

352 studied (Sionkowska, 2011). Extracted from natural sources or obtained by bacterial fermentation, these 353 macromolecules offer various advantages such as large-scale production, low toxicity, and similarity to

354 ECM components.

355 Collagen, a major component of healthy IVD ECM, exhibits biodegradability, weak antigenicity, and

356 superior biocompatibility compared with other natural proteins. The widespread biomedical use of

collagen is based on its ability to be shaped into films, sponges, and micro- or nanoparticles (Liu et al.,

358 2013). Gelatin, derived from collagen hydrolysis, is biodegradable in a physiological environment, with

the formation of harmless metabolic products. In degenerated IVD, injectable gelatin microspheres

360 impregnated with PRP reduced NP cells apoptosis and slowed down degeneration in adult rabbits

361 (Sawamura et al., 2009). HA, another leading constituent of the NP ECM, was also studied as a scaffold

362 for tissue engineering (Sionkowska et al., 2016), but less so for IVD strategies.

363 Although they cannot be found in the human body, alginate, a cell wall polysaccharide constituent of

- brown algae, and chitosan, a polysaccharide synthesized by crustaceans, were used for multiple
- biomedical applications (Bernkop-Schnürch and Dünnhaupt, 2012). Alginate microparticles were
- designed for the in vitro delivery of PRP in the treatment of degenerated IVDs (Sawamura et al., 2009);
- however, their ionic gelation-dependent low stability in the body might constitute a drawback. Similarly,
- cationic chitosan was considered for IVD regenerative applications, as a cell or molecule carrier (Cheng et al., 2010; Roughley et al., 2006).
  - 10

370 Synthetic polymers are obtained through reproducible industrial processes, with tunable physicochemical
 371 and mechanical properties. The most commonly used polymers for drug delivery systems are poly(lactic 372 co-glycolic acid) (PLGA) and poly-ε-caprolactone (PCL) (Kondiah et al., 2016).

- 373 PLGA microspheres were widely used for the release of TGF-β3 or anti-inflammatory molecules
- 374 (Bhardwaj et al., 2007; Liang et al., 2013). They were used for recombinant human GDF-5 delivery in
- vivo in a rat model and increased the synthesis of aggrecan and type II collagen (Yan et al., 2013). PLGA
- 376 microspheres were also used for the sustained delivery of IL-1 $\beta$  receptor antagonist in a 3D culture model,
- and this treatment effectively attenuated the IL-1 $\beta$ -mediated inflammatory response of the 3D cultured NP calls (Gorth et al. 2012)
- 378 cells (Gorth et al., 2012).
- 379 PCL, a semi-crystalline and hydrophobic polymer, is currently used in FDA-approved sutures for surgery
- and in commercially available contraceptive devices (Wei et al., 2009). PCL is often associated with
   hydrophilic polymers, either natural or synthetic ones (e.g., PEG, PAA or pNIPAAM), to form di- or
- hydrophilic polymers, either natural or synthetic ones (e.g., PEG, PAA or pNIPAAM), to form di- or
   triblock amphiphilic copolymers (Fu et al., 2014; Li and Tan, 2015). Moreover, PCL was extensively used
- for IVD tissue engineering, especially for AF repair (Wismer et al., 2013). Moreover, PCL was extensively used
- microparticles were successfully designed for the sustained delivery of bioactive molecules, although not
- for IVD applications (Jung et al., 2015; Z. Luo et al., 2016).
- 386 In the context of IVD regenerative medicine, the usefulness of these natural and synthetic polymers is
- 387 broadened further by our ability to create systems with multiple shapes and sizes that deliver drugs,
- 388 cytokines, growth factors, or stem cells in a controlled manner. They are reviewed in the following
- 389 sections.
- 390

#### 391 6.2. In situ forming hydrogels as macro-scaled delivery systems

- 392 Hydrogels, networks of hydrophilic polymers capable of absorbing large amounts of water, are of
- particular interest for highly hydrated NP and AF tissue regeneration. Besides water, they provide
- 394 mechanical support and a 3D microenvironment adapted for cell survival and proliferation. Attention was 395 focused on the development of stimuli-responsive polymer solutions that could be injected in a minimally
- 396 invasive manner prior to in situ solidifying, ensuring a perfect fit to the desired tissue. The in situ
- 397 formation of hydrogels occurs through chemical covalent bonds or physical bonds (electrostatic or
- 398 hydrophobic interactions).
- 399 **Among chemical crosslinking**, photo-induced crosslinking requires the production of radical initiating 400 species and a precursor solution of a photoreactive polymer, usually a polyethylene glycol (PEG) acrylate or methacrylate derivative (Francisco et al., 2014; Li et al., 2014). While the use of photo-crosslinked 401 402 polymers allows for time-controlled gelation, the limited access of the light apparatus to the target tissue 403 weakens this strategy for IVD regeneration. Click chemistry, Schiff's base chemistry, Michael-type 404 reactions, and enzymatic, protein, or pH modification-based crosslinking approaches might be better suited for IVD in situ gelation (Blanquer et al., 2014). These processes are generally compatible with cell 405 406 or bioactive molecule loading as they often take place in mild conditions (37°C, physiological pH, short 407 period). In the early 2000s, a silanized cellulose-based hydrogel, in which crosslinking is triggered by pH,
- 408 was developed (Bourges et al., 2002). The importance of this hydrogel has been demonstrated for the
- 409 regeneration of articular cartilage (Vinatier et al., 2009) and myocardium (Mathieu et al., 2012) and it is 410 currently studied for IVD regeneration (Henry et al., 2017).
- 411 **Physically crosslinked** hydrogels are formed via non-covalent interactions in response to an external
- stimulus (pH, temperature, molecule level, light, pressure, electric current) (Qiu and Park, 2012). Thermo-
- 413 responsive polymers spontaneously undergo a sol-gel transition when exposed to physiological
- 414 temperature after in vivo injection. Recombinant polypeptides [e.g., elastin-like polypeptide (Nettles et al.,
- 415 2010)], naturally derived polysaccharides [e.g., chitosan (Cheng et al., 2013)], and synthetic polymers
- 416 [e.g., block copolymers of PEG and PLGA (Tran et al., 2012), poly (N-isopropylacrylamide) (p-

NIPAAM) (Willems et al., 2015)] are the most commonly reported thermo-responsive polymers for IVD
 regeneration.

419 In situ-forming hydrogels exhibit many advantages for IVD treatment. They provide an aqueous 420 environment that can rehydrate the degenerated microenvironment, they protect biologics, and they can 421 restore, at least partially, the mechanical properties of IVDs. The permeability of hydrogels facilitates 422 good transport of nutrients to encapsulated cells. Nevertheless, hydrogels can present some restrictions 423 due to, for example, their weak mechanical properties. One of the main drawbacks of in situ-forming bulk 424 hydrogels is the difficulty to tune their release characteristics. Indeed, while the diffusion of entrapped 425 molecules can be slightly modified by adjusting the polymer network mesh, exchanges with the targeted microenvironment are limited to the interface hydrogel/tissue, usually in the order of a few cm<sup>2</sup>/g. By 426 427 comparison, micro- and nano-scale delivery systems can develop areas greater than  $1,000 \text{ m}^2/\text{g}$  as 428 measured by N<sub>2</sub> adsorption-desorption experiments.

429

#### 430 **6.3. Micro-scale delivery systems**

In parallel to stimuli-responsive polymer solutions ensuring a perfect fit to the IVD defect following in situ gelation, injectable micro-sized particles, with diameters varying from 20 µm to 200 µm, have release kinetics depending on their size and shape. Their release properties can be modified by tuning parameters such as the size, chemical composition, crosslinking degree, or molecular weight of the polymer (Balmert and Little, 2012). Moreover, chemical modification of the microparticle surface can allow for covalent coupling of factors such as antibodies and receptors. Microparticles are able to deliver biological factors

437 over a long period while protecting them from inactivation (Wenk et al., 2009).

438 Many inorganic and polymer-based microparticles can be prepared by simple or double emulsion

439 methods, in which a solution of the material is emulsified in water, followed by the formation of particles

440 during the evaporation of the organic solvent. A recent modified protocol combines a crosslinking process 441 and the emulsion step in an oil phase (Phromsopha and Baimark, 2014). Other production methods are

441 and the emulsion step in an on phase (Phromsopha and Balmark, 2014). Other production methods are 442 spray drying, gelation, coacervation, electrospray, supercritical fluid mixing, and microfluidic technique.

- 442 spray drying, geration, coacervation, electrospray, supercritical fund mixing, and incromulate technique 443 While their main advantage over classic emulsion is the narrower particle size distribution, the yield is
- 444 often disappointing.

445 Recently, GDF-5-loaded PLGA microspheres, with an in vitro sustained delivery for 42 days, were

446 injected into degenerated rat IVDs. Restoration of the disc height to almost 90% of the non-punctured disc
 447 was achieved (Yan et al., 2013). Moreover, the injection significantly increased aggrecan and type II

447 was achieved (1 an et al., 2013). Moreover, the injection significantly increased aggregation and type in448 collagen mRNA expression, whereas type I collagen mRNA expression was significantly decreased,

suggesting that a micro-sized carrier could be of interest for IVD regeneration (Yan et al., 2013).

#### 450 **6.4. Nano-scale delivery systems**

451 Alongside microparticles, many materials were used to design nanoparticles for biomedical applications,

452 as nanocarriers are easily taken up by cells (Wilczewska et al., 2012). Polymeric nanoparticles, liposomes,

dendrimers, polymers, silicon or carbon materials, and magnetic nanoparticles are examples of nano-

454 carriers used as drug delivery systems notably for cancer (Sun et al., 2014) or bone regeneration (van Rijt

and Habibovic, 2017). In this section, we highlight the particles that are feasible for use in the context of

456 DDD.

457 **Polymeric nanoparticles:** Bioactive molecules can be either encapsulated or immobilized onto the

surface of the nanoparticles, then released by desorption, diffusion, or nanoparticle erosion. Thus,

degradable nanocarriers are widely used in biomedical applications as they can be hydrolyzed in situ

460 (Kohane et al., 2006) and often considered to be biocompatible with tissues and cells, since they do not

induce pro-inflammatory or immune reactions (des Rieux et al., 2006). An upregulation of aggrecan and

462 type II collagen was achieved by using chitosan/Poly-(c-glutamic acid) nanoparticles loaded with the anti-

inflammatory drug diclofenac. These nanoparticles were injected into a needle-punctured bovine IVD
cultured in vitro with IL-1β. Additionally, down-regulation of IL-6, IL-8, MMP1, and MMP3 was noted
(Teixeira et al., 2016).

466 Self-organized spherical liposomes, usually 80-300 nm in diameter, contain at least one lipid bilayer 467 surrounding an aqueous core (Daraee et al., 2014). They are thus able to encapsulate both hydrophobic 468 and hydrophilic molecules within the lipid bilayer or the aqueous core, respectively. Liposomes were first used to improve the solubility and therapeutic index of chemotherapeutic agents, by allowing targeted 469 470 delivery and enhanced tissue penetration. Since liposomes interact with cells by adsorption, fusion, 471 endocytosis, or lipid transfer, they were studied as gene and siRNA carriers, especially cationic liposomes 472 (Majzoub et al., 2016). The release kinetics can be tuned by the liposome composition, pH, osmotic 473 gradient, incorporation of temperature- or pH-sensitive lipids (dos Santos Giuberti et al., 2011), and the 474 surrounding environment. Liposomes were thus used for drug delivery in various applications such as in 475 anti-cancer or anti-inflammation therapies or to reach specific organs (lungs, liver, spleen, brain). However, to date, their use for tackling IVD degeneration has not been reported. Interestingly, lipid 476 477 nanocapsules were successfully designed for nucleic acid delivery (Huynh et al., 2009) and their

478 application in siRNA delivery for IVD regeneration could be of interest.

479 **Dendrimers** are branched polymers whose toxicity is mainly related to functional groups presented at the 480 surface layer and can be lowered by functionalization, e.g., by PEGylation (Zeng et al., 2016). This 481 functionalization improves the surface activity of dendrimers as well as their biological and 482 physicochemical properties. Drugs are entrapped in the internal structure of the dendrimer, adsorbed or 483 chemically bonded onto the dendrimer surface according to its properties (sensitivity to temperature or 484 pH, toxicity, solubility). One of the most commonly used in biomedical applications is poly-(amino 485 amide; PAMAM-) as it was proven to be non-toxic and safe (Chauhan et al., 2009) and its capacity for 486 gene or drug delivery was extensively explored (Zhao et al., 2011). Indeed, PAMAM dendrimers can 487 modulate cytokine and chemokine release (Seelbach et al., 2015) and exhibit anti-inflammatory behavior 488 by inhibiting proinflammatory cytokine secretion (Bosch, 2011). Thereby, dendrimers demonstrated their 489 suitability in rheumatoid arthritis and musculoskeletal diseases and their application in IVD regeneration

490 could be envisioned.

491 **Carbon nanotubes** are the most commonly used carbon nanoparticles and were widely studied for drug

delivery in biomedical applications (Beg et al., 2011). They are characterized by a single or multiwall

493 graphene-based tube with a high specific surface area and possibly excellent electronic and thermal 494 conductivity (Beg et al., 2011). They have exceptional mechanical properties and are thus of interest as

- 494 conductivity (Beg et al., 2011). They have exceptional mechanical properties and are thus of interest as 495 fillers in composite materials (Harrison and Atala, 2007). Toxicity due to their dimensions and released
- 496 impurities was reported, but their biocompatibility can be enhanced with chemical surface engraftment of
- 497 PEG, PAMAM, or albumin (Harrison and Atala, 2007). Molecules are encapsulated inside carbon
- 498 nanotubes or chemically adsorbed onto their surface. Carbon nanotubes improve the mechanical properties
- 499 of hydrogels (Dong et al., 2013) and can be of potential interest for IVD regenerative medicine.
- 500 Mesoporous silica nanoparticles are a widely studied biomaterial for their drug delivery capacity in
- particular for cancer therapies (Yang and Yu, 2016) or nerve growth (Sun et al., 2016). Indeed,
- 502 mesoporous colloidal silica exhibit several advantages such as cytocompatibility, a high specific surface
- area, and ease of functionalization (Moritz and Geszke-Moritz, 2015). Additionally, the size, shape,
- 504 surface pattern, porosity, and pore size of silica nanoparticles are easily tunable (Hoffmann et al., 2006). 505 In the biomedical field, the best known measurements are supersticles are MCM 41 and SDA 15 D
- 505 In the biomedical field, the best known mesoporous silica nanoparticles are MCM-41 and SBA-15. Drug 506 loading into mesoporous silica might occur via chemical or physical adsorption, and release is usually
- 500 loading into mesoporous silica might occur via chemical or physical adsorption, and release is usually 507 controlled by diffusion throughout the mesopores. The potential of these particles was demonstrated for
- 507 controlled by diffusion inroughout the mesopores. The potential of these particles was demonstrated for 508 biomedical applications (Vallet-Regi and Balas, 2008) and more recently for the IVD (Henry et al., 2017).

#### 509 **6.5. Multiscale systems for complex regenerative strategies**

510 Over the past few years, composite materials, composed of at least two distinct components, drew

511 attention in biomedical engineering since they possess additional properties that each constituent alone

512 does not exhibit. They were widely studied for bone regeneration, and successful clinical trials were

513 recently reviewed (Jahan and Tabrizian, 2016).

514 Regarding specific IVD applications, we would like to direct the reader's attention to the multiscale

515 systems illustrated in Figure 4 that emphasize the integration of technologies such as in situ-forming

516 hydrogels with ready-to-use micro- or nanoparticles. The assembly of specific functionalities obtained by 517 combining a bulk matrix, to restore hydration and mechanical properties, with a long-term reservoir of

cells or molecules, to provide biological cues, could constitute a breakthrough in IVD regenerative

519 therapies. The main outcome of this review is to highlight the crucial role of biomaterials in IVD

regenerative medicine strategies. With regard to the aforementioned device needs, we would like to stress

521 that biomaterials are considered the driving force of the IVD regenerative medicine train (Figure 5).

522 Biomaterials are essential for the design and development of hydrogels capable of restoring load-bearing

523 function while supporting cells until an appropriate ECM is synthesized. Biomaterials are also essential

524 for engineering efficient and robust delivery devices that can be tailored to specific tissues.

525 For example, we recently reported on a multiscale system where an Si-HPMC hydrogel supplemented

with rod-shaped silica nanofibers provided sustained release of TGF- $\beta$ 1 and GDF-5, compared with the

527 plain hydrogel, while maintaining their in vitro biological activity (Henry et al., 2017). Recently, TGF- $\beta$ 3-528 loaded PLGA nanoparticles dispersed in a dextran/gelatin hydrogel, although not injectable, increased the

gene expression of both aggrecan and type II collagen of MSCs in vitro (Gan et al., 2016). We believe that

530 promising engineering technologies such as interpenetrating network-strengthened hydrogels (Gan et al.,

531 2017), self-assembling nanofibrous peptide hydrogels (Wan et al., 2016), and programmed

532 sequential bioactive factor delivery to target multiple repair steps (Bayer et al., 2017) could lead to

533 significant advances in IVD regenerative strategies. Finally, an ambitious strategy would aim at delivering

534 chemo-attractants from micro- or nanoparticles to attract progenitor cells toward a hydrogel-based

535 microenvironment that would promote endogenous repair.

536

537

538

#### 539 7. <u>Conclusion</u>

540 IVD degeneration is a major health concern and today, thanks to recent discoveries, the physiopathology 541 of this disease is better understood. Building on this deeper knowledge, various biotherapies were 542 proposed in the past few years, including cell-based therapies, molecular therapies, and nucleic acid-based 543 therapies. These therapies aim at locally injecting biologics including cells, proteins, or DNA as mentioned earlier. Even if promising results have been obtained in vitro, the harsh environment of 544 545 degenerated IVDs is a major concern that has hampered the clinical translatability of such concepts. The 546 main outcome of this review is to highlight the essential role of biomaterials in strategies for IVD 547 regenerative medicine. Indeed, injectable in situ crosslinking hydrogels could restore IVD height and 548 hydration and would be particularly suited for IVD regenerative strategies. Additionally, hydrogels can 549 improve cell viability and protect bioactive molecules from in situ degradation. Finally, hydrogels can be used as scaffolds, supplemented or not with micro- or nanoparticles. These so-formed multiscale systems 550 551 would allow for a sustained delivery of the biologics promoting long-term IVD regenerative processes.

552 The design of the optimal system for IVD regeneration is still under debate as disc degeneration is a

553 multifactorial disease. Cell-based therapies were widely considered in view of the major decrease in cell

numbers that is observed with degeneration. However, the regulatory framework and the costs associated

with these strategies may restrict their use. We believe that an effective approach would target the early

degenerative processes and involve multiscale biomaterials allowing molecules to be sustainably delivered

- in situ and enabling a prolonged stimulation of endogenous repair.
- 558

#### 559 8. <u>Acknowledgements</u>

The authors acknowledge financial support from: Fondation de l'Avenir pour la Recherche Médicale
Appliquée (AP-RMA-2015-018), Région Pays de la Loire (Paris Scientifique BIO2, LMA project and
DIVA project), Agence Nationale pour la Recherche (REMEDIV, JC/JC STIMUDISC) and Fondation

- 563 pour la Recherche Médicale (DBS20131128442).
- 564

#### 565 9. Figures legends

566

567 Figure 1: Healthy intervertebral disc (IVD) local vascularization, innervation, and internal organization

A. Located between vertebrae, the IVD is composed of two vertebral endplates, an *Annulus fibrosus* 

569 (purple) and a *Nucleus pulposus* (NP, blue). Since the IVD is non-innervated and non-vascularized,

570 oxygen and glucose supply as well as metabolite excretion occur by diffusion through the vertebral

- endplates. This particular property induces specific conditions of osmolarity and pH, leading the IVD
- 572 resident cells to adapt their metabolism. B. In AF tissue, fibroblast-like cells are organized parallel to
- 573 collagen concentric lamellae. In NP tissue, large vacuolated notochordal cells and nucleopulpocytes
- 574 coexist and ensure homeostasis of the extracellular matrix.
- 575
- 576 Figure 2: NP homeostasis rupture leads to IVD degeneration
- 577 In a healthy IVD, regulation of anabolic and catabolic processes (i.e., homeostasis) enables the tissue to be
- 578 hydrated and maintains its mechanical properties. Under various factors such as genetics, aging,
- 579 environmental causes, or mechanical stress, the cellular dialogue underlying the extracellular matrix
- 580 homeostasis is disrupted. This rupture causes lower anabolism and increased catabolism, ultimately

- 581 leading to ECM degradation, tissue inflammation, vascularization, fibrosis, and nerve ingrowth.
- 582 Altogether, these phenomena are the key players in IVD degeneration.
- 583

#### 584 Figure 3: Current IVD regenerative medicine is based on three approaches

585 Cell-based therapies. Considering the pivotal role of the decreased cell number in IVD degeneration, cell-based therapies have been increasingly considered. Autologous or allogeneic cells have been 586 proposed and stem cells have been the subject of major investigations. Stem cells have been mainly 587 588 isolated from adipose tissue or bone marrow and more recently pluripotent stem cells have been contemplated. It is also largely debated whether the optimal cells for IVD repair should be differentiated 589 590 or undifferentiated. Molecular therapies. The broadened understanding of the phenomena involved 591 during IVD degeneration has made it possible to consider molecular therapies via the direct injection of 592 various factors to promote anabolism, to manage inflammation, or to attract regenerative cells. Nucleic 593 acid-based therapies. Recent approaches target DNA and miRNA sequences and mostly aim to 594 downplay catabolic factor expression. Finally, combined approaches such as genetically modified cells 595 that could contribute to decreasing MMP local expression and activity could potentiate the efficacy of 596 these therapies.

597

#### 598 Figure 4: Multiscale systems associate in situ-forming scaffolds with ready-to-use particles

599 In the context of IVD regenerative medicine, we propose to combine in situ-forming systems with ready-

to-use micro- or nanoparticles, thereby forming multiscale systems. We hypothesize that injectable
 polymers that undergo local gel formation will restore IVD hydration and mechanical properties, while

portine is that and ergo rocal get formation will restore i v D injulation and incontainear properties, while
 micro- or nano-sized carriers loaded with cells or molecules will provide supportive cues for regenerative
 processes.

604

#### 605 Figure 5: Biomaterials: the driving force of the IVD regenerative medicine train

Biomaterials are essential for the design and development of delivery systems for IVD regenerative

- medicine. They can protect and deliver nucleic acids, bioactive molecules, or cells for a long period and
   may thus contribute to regenerating the degraded ECM of IVDs.
- 609
- 610

#### 611 **10.** <u>References</u>

- Abdel Shaheed, C., Maher, C.G., Williams, K.A., Day, R., McLachlan, A.J., 2016. Efficacy,
  Tolerability, and Dose-Dependent Effects of Opioid Analgesics for Low Back Pain. JAMA
  Intern. Med. 176, 958.
- Alkhatib, B., Rosenzweig, D.H., Krock, E., Roughley, P.J., Beckman, L., Steffen, T., Weber,
   M.H., Ouellet, J.A., Haglund, L., Scoliosis, M., 2014. Acute mechanicalt injury of the
- human intervertebral dics Link to degeneration and pain. Eur. Cell. Mater. 28, 98–111.
- Ankrum, J.A., Ong, J.F., Karp, J.M., 2014. Mesenchymal stem cells: immune evasive, not
   immune privileged. Nat Biotechnol 32, 252–260.
- Baliga, S., Treon, K., Craig, N.J.A., 2015. Low back pain: Current surgical approaches. Asian
  Spine J. 9, 645–657.
- Balmert, S.C., Little, S.R., 2012. Biomimetic Delivery with Micro- and Nanoparticles. Adv.

- 623 Mater. 24, 3757–3778.
- Bartel, D.P., 2004. MicroRNAs: Genomics, Biogenesis, Mechanism, and Function. Cell 116,
   281–297.
- Bayer, E.A., Jordan, J., Gottardi, R., Fedorchak, M., Kumta, P.N., Little, S., 2017. Programmed
   Platelet Derived Growth Factor-β and Bone Morphogenetic Protein-2 Delivery from a
   Hybrid Calcium Phosphate/Alginate Scaffold. Tissue Eng. Part A Ahead of print.
- Bedore, J., Leask, A., Séguin, C.A., 2014. Targeting the extracellular matrix: Matricellular
   proteins regulate cell-extracellular matrix communication within distinct niches of the
   intervertebral disc. Matrix Biol. 37, 124–130.
- Beg, S., Rizwan, M., Sheikh, A.M., Hasnain, M.S., Anwer, K., Kohli, K., 2011. Advancement in
  carbon nanotubes: Basics, biomedical applications and toxicity. J. Pharm. Pharmacol. 63,
  141–163.
- Bernkop-Schnürch, A., Dünnhaupt, S., 2012. Chitosan-based drug delivery systems. Eur. J.
  Pharm. Biopharm. 81, 463–469.
- Bhardwaj, U., Sura, R., Papadimitrakopoulos, F., Burgess, D.J., 2007. Controlling acute
  inflammation with fast releasing dexamethasone-PLGA microsphere/pva hydrogel
  composites for implantable devices. J. diabetes Sci. Technol. 1, 8–17.
- Blanco, J.F., Graciani, I.F., Sanchez-Guijo, F.M., Muntion, S., Hernandez-Campo, P.,
  Santamaria, C., Carrancio, S., Barbado, M.-V., Cruz, G., Gutierrez-Cosio, S., Herrero, C.,
  San Miguel, J.F., Brinon, J.G., del Canizo, M.-C., 2010. Isolation and characterization of
  mesenchymal stromal cells from human degenerated nucleus pulposus: comparison with
  bone marrow mesenchymal stromal cells from the same subjects. Spine (Phila. Pa. 1976).
  35, 2259–2265.
- Blanquer, S.B.G., Grijpma, D.W., Poot, A. a., 2014. Delivery systems for the treatment of
   degenerated intervertebral discs. Adv. Drug Deliv. Rev. 84, 172–187.
- Bochot, A., Fattal, E., 2012. Liposomes for intravitreal drug delivery: A state of the art. J.
  Control. Release 161, 628–634.
- Bosch, X., 2011. Dendrimers to treat rheumatoid arthritis. ACS Nano 5, 6779–6785.
- Bourges, X., Weiss, P., Daculsi, G., Legeay, G., 2002. Synthesis and general properties of
  silated-hydroxypropyl methylcellulose in prospect of biomedical use. Adv Colloid Interface
  Sci 99, 215–228.
- Chauhan, A.S., Jain, N.K., Diwan, P. V., 2009. Pre-clinical and behavioural toxicity profile of
   PAMAM dendrimers in mice. Proc. R. Soc. A Math. Phys. Eng. Sci. 466, 1535–1550.
- Chen, J., Lee, E.J., Jing, L., Christoforou, N., Leong, K.W., Setton, L.A., 2013. Differentiation of
  mouse induced pluripotent stem cells (iPSCs) into nucleus pulposus-like cells in vitro. PLoS
  One 8, e75548.
- Cheng, Y.H., Yang, S.H., Liu, C.C., Gefen, A., Lin, F.H., 2013. Thermosensitive hydrogel made
   of ferulic acid-gelatin and chitosan glycerophosphate. Carbohydr. Polym. 92, 1512–1519.
- 661 Cheng, Y.H., Yang, S.H., Su, W.Y., Chen, Y.C., Yang, K.C., Cheng, W.T., Wu, S.C., Lin, F.H.,
  662 2010. Thermosensitive chitosan-gelatin-glycerol phosphate hydrogels as a cell carrier for
  663 nucleus pulposus regeneration: an in vitro study. Tissue Eng Part A 16, 695–703.

- 664 Chujo, T., An, H.S., Akeda, K., 2006. Effects of Growth Differentiation Factor-5 on the
  665 Intervertebral Disc <sup>'</sup>. In Vitro Bovine Study and In Vivo Rabbit Disc Degeneration Model
  666 Study. Spine (Phila Pa 1976) 31, 2909–2917.
- Colombier, P., Camus, A., Lescaudron, L., Clouet, J., Guicheux, J., 2014. Intervertebral disc
   regeneration: a great challenge for tissue engineers. Trends Biotechnol. 32, 433–5.
- Colombier, P., Clouet, J., Boyer, C., Ruel, M., Bonin, G., Lesoeur, J., Moreau, A., Fellah, B.H.,
  Weiss, P., Lescaudron, L., Camus, A., Guicheux, J., 2016. TGF- b 1 and GDF5 Act
  Synergistically to Drive the Differentiation of Human Adipose Stromal Cells toward
  Nucleus Pulposus -like Cells. Stem Cells 34, 653–667.
- Colombini, A., Lombardi, G., Corsi, M.M., Banfi, G., 2008. Pathophysiology of the human
  intervertebral disc. Int. J. Biochem. Cell Biol. 40, 837–842.
- 675 Daraee, H., Etemadi, A., Kouhi, M., Alimirzalu, S., Akbarzadeh, A., 2014. Application of
- liposomes in medicine and drug delivery. Artif. Cells, Nanomedicine, Biotechnol. 1401, 1–
  11.
- des Rieux, A., Fievez, V., Garinot, M., Schneider, Y.J., Préat, V., 2006. Nanoparticles as
  potential oral delivery systems of proteins and vaccines: A mechanistic approach. J. Control.
  Release 116, 1–27.
- Diab, R., Canilho, N., Pavel, I.A., Haffner, F.B., Girardon, M., Pasc, A., 2017. Silica-based
  systems for oral delivery of drugs, macromolecules and cells. Adv. Colloid Interface Sci. In
  press.
- Dong, W., Huang, C., Wang, Y., Sun, Y., Ma, P., Chen, M., 2013. Superior mechanical
  properties of double-network hydrogels reinforced by carbon nanotubes without organic
  modification. Int. J. Mol. Sci. 14, 22380–22394.
- dos Santos Giuberti, C., de Oliveira Reis, E.C., Ribeiro Rocha, T.G., Leite, E.A., Lacerda, R.G.,
   Ramaldes, G.A., de Oliveira, M.C., 2011. Study of the pilot production process of long circulating and pH-sensitive liposomes containing cisplatin. J. Liposome Res. 21, 60–9.
- Elmasry, S., Asfour, S., De Rivero Vaccari, J.P., Travascio, F., 2015. Effects of tobacco smoking
   on the degeneration of the intervertebral disc: A finite element study. PLoS One 10, 1–22.
- Enochson, L., Stenberg, J., Brittberg, M., Lindahl, a, 2014. GDF5 reduces MMP13 expression in
   human chondrocytes via DKK1 mediated canonical Wnt signaling inhibition. Osteoarthritis
   Cartilage 22, 566–77.
- Erwin, W.M., Ashman, K., O'Donnel, P., Inman, R.D., 2006. Nucleus pulposus notochord cells
   secrete connective tissue growth factor and up-regulate proteoglycan expression by
   intervertebral disc chondrocytes. Arthritis Rheum 54, 3859–3867.
- Erwin, W.M., Islam, D., Inman, R.D., Fehlings, M.G., Tsui, F.W., 2011. Notochordal cells
   protect nucleus pulposus cells from degradation and apoptosis: implications for the
   mechanisms of intervertebral disc degeneration. Arthritis Res Ther 13, R215.
- Feng, C., Liu, H., Yang, Y., Huang, B., Zhou, Y., 2015. Growth and Differentiation Factor-5
  Contributes to the Structural and Functional Maintenance of the Intervertebral Disc. Cell.
  Physiol. Biochem. 35, 1–16.
- Fontana, G., See, E., Pandit, A., 2015. Current trends in biologics delivery to restore
   intervertebral disc anabolism. Adv. Drug Deliv. Rev. 84, 146–158.

- Francisco, A.T., Hwang, P.Y., Jeong, C.G., Jing, L., Chen, J., Setton, L.A., 2014.
   Photocrosslinkable laminin-functionalized polyethylene glycol hydrogel for intervertebral disc regeneration. Acta Biomater. 10, 1102–1111.
- Fu, W., Liu, Z., Feng, B., Hu, R., He, X., Wang, H, Meng, Y., Huang, H., Zhang, H., Wang, W.,
  2014. Electrospun gelatin / PCL and collagen / PLCL scaffolds for vascular tissue
  engineering. Int. J. Nanomedicine 9, 2335–2344.
- Galbusera, F., Bellini, C.M., Zweig, T., Ferguson, S., Raimondi, M.T., Lamartina, C., BraydaBruno, M., Fornari, M., 2008. Design concepts in lumbar total disc arthroplasty. Eur. Spine
  J. 17, 1635–1650.
- Gan, Y., Li, P., Wang, L., Mo, X., Song, L., Xu, Y., Zhao, C., Ouyang, B., Tu, B., Luo, L., Zhu,
  L., Dong, S., Li, F., Zhou, Q., 2017. An interpenetrating network-strengthened and
  toughened hydrogel that supports cell-based nucleus pulposus regeneration. Biomaterials
  136, 12–28.
- Gan, Y., Li, S., Li, P., Xu, Y., Wang, L., Zhao, C., Ouyang, B., Tu, B., Zhang, C., Luo, L., Luo,
   X., Mo, X., Zhou, Q., 2016. A Controlled Release Codelivery System of MSCs
   Encapsulated in Dextran/Gelatin Hydrogel with TGF- β 3-Loaded Nanoparticles for Nucleus
- 722 Pulposus Regeneration. Stem Cells Int. 2016.
- Goldspiel, B.R., Green, L., Calis, K.A., 1993. Human gene therapy. Clin. Pharm. 12, 488–505.
- Gorth, D.J., Mauck, R.L., Chiaro, J.A., Mohanraj, B., Hebela, N.M., Dodge, G.R., Elliott, D.M.,
  Smith, L.J., 2012. IL-1ra delivered from poly(lactic-co-glycolic acid) microspheres
  attenuates IL-1β-mediated degradation of nucleus pulposus in vitro. Arthritis Res. Ther. 14,
  R179.
- Grad, S., Bow, C., Karppinen, J., Luk, K.D., Cheung, K.M., Alini, M., Samartzis, D., 2016.
   Systemic blood plasma CCL5 and CXCL6: Potential biomarkers for human lumbar disc
   degeneration. Eur. Cell. Mater. 30, 1–10.
- Gruber, H., Marrero, E., Ingram, J., Hoelscher, G., Hanley, E., 2016. The chemokine, CXCL16,
  and its receptor, CXCR6, are constitutively expressed in human annulus fibrosus and
  expression of CXCL16 is up-regulated by exposure to IL-1β in vitro. Biotech. Histochem.
  295, 1–8.
- Gruber, H.E., Hoelscher, G.L., Ingram, J.A., Bethea, S., Hanley, E.N., 2014a. Growth and
  differentiation factor-5 (GDF-5) in the human intervertebral annulus cells and its modulation
  by IL-1b and TNF-b in vitro. Exp. Mol. Pathol. 96, 225–229.
- Gruber, H.E., Hoelscher, G.L., Ingram, J.A., Bethea, S., Norton, H.J., Hanley, E.N., 2014b.
   Production and expression of RANTES (CCL5) by human disc cells and modulation by IL 1-β and TNF-α in 3D culture. Exp. Mol. Pathol. 96, 133–138.
- 741 Hadjipavlou, a G., Tzermiadianos, M.N., Bogduk, N., Zindrick, M.R., 2008. The
- pathophysiology of disc degeneration: a critical review. J. Bone Joint Surg. Br. 90, 1261–
  1270.
- Harrison, B.S., Atala, A., 2007. Carbon nanotube applications for tissue engineering.
  Biomaterials 28, 344–353.
- Henriksson, H.B., Svanvik, T., Jonsson, M., Sci, L.M., Hagman, M., Horn, M., Lindahl, A.,
  Brisby, H., 2009. Transplantation of Human Mesenchymal Stems Cells Into Intervertebral

- 748 Discs in a Xenogeneic Porcine Model. Spine (Phila. Pa. 1976). 34, 141–148.
- Henry, N., Clouet, J., Le Visage, C., Weiss, P., Gautron, E., Humber, B., Terrisse, H., Guicheux,
  J., Le Bideau, J., 2017. Silica nanofibers as a new drug delivery system: a study of proteinsilica interactions. J Mater Chem B 5, 2908–2920.
- Hirshi, K.K., Li, S., Roy, K., 2014. Induced pluripotent stem cells for regenerative medicine.
  Annu Rev Biomed Eng 16, 277–294.
- Hiyama, A., Mochida, J., Iwashina, T., Omi, H., Watanabe, T., Serigano, K., Tamura, F., Sakai,
  D., 2008. Transplantation of mesenchymal stem cells in a canine disc degeneration model. J.
  Orthop. Res. 26, 589–600.
- Hoffmann, F., Cornelius, M., Morell, J., Froba, M., 2006. Silica based mesoporous organic inorganic hybrid materials. Angew Chem Int Ed 45, 3216–51.
- Huynh, N.T., Passirani, C., Saulnier, P., Benoit, J.P., 2009. Lipid nanocapsules: A new platform
  for nanomedicine. Int. J. Pharm. 379, 201–209.
- Illien-Junger, S., Grosjean, F., Laudier, D.M., Vlassara, H., Striker, G.E., Iatridis, J.C., 2013.
   Combined Anti-Inflammatory and Anti-AGE Drug Treatments Have a Protective Effect on Intervertebral Discs in Mice with Diabetes. PLoS One 8.
- Illien-Jünger, S., Pattappa, G., Peroglio, M., Benneker, L.M., Stoddart, M.J., Sakai, D., Mochida,
  J., Grad, S., Alini, M., 2012. Homing of Mesenchymal Stem Cells in Induced Degenerative
  Intervertebral Discs in a Whole Organ Culture System. Spine (Phila. Pa. 1976). 37, 1865–
  1873.
- Jahan, K., Tabrizian, M., 2016. Composite biopolymers for bone regeneration enhancement in
   bony defects. Biomater. Sci. 4, 25–39.
- Jeong, J.H., Jin, E.S., Min, J.K., Jeon, S.R., Park, C.S., Kim, H.S., Choi, K.H., 2009. Human
  mesenchymal stem cells implantation into the degenerated coccygeal disc of the rat.
  Cytotechnology 59, 55–64.
- Jung, S.-M., Yoon, G.H., Lee, H.C., Shin, H.S., 2015. Chitosan nanoparticle/PCL nanofiber
   composite for wound dressing and drug delivery. J. Biomater. Sci. Polym. Ed. 26, 252–263.
- Kaku, M., Kitami, M., Rosales Rocabado, J.M., Ida, T., Akiba, Y., Uoshima, K., 2017.
   Recruitment of bone marrow-derived cells to the periodontal ligament via the stromal cell derived factor-1/C-X-C chemokine receptor type 4 axis. J. Periodontal Res. Ahead of print.
- Kawaguchi, S., Yamashita, T., Katahira, G., Yokozawa, H., Torigoe, T., Sato, N., 2002.
  Chemokine profile of herniated intervertebral discs infiltrated with monocytes and macrophages. Spine (Phila. Pa. 1976). 27, 1511–6.
- Kepler, C.K., Markova, D.Z., Dibra, F., Yadla, S., Vaccaro, A.R., Risbud, M. V, Albert, T.J.,
  Anderson, D.G., 2013a. Expression and relationship of proinflammatory chemokine
  RANTES/CCL5 and cytokine IL-1β in painful human intervertebral discs. Spine (Phila. Pa.
  1976). 38, 873–80.
- Kepler, C.K., Ponnappan, R.K., Tannoury, C. a, Risbud, M. V, Anderson, D.G., 2013b. The
   molecular basis of intervertebral disc degeneration. Spine J. 13, 318–30.
- Kohane, D.S., Tse, J.Y., Yeo, Y., Padera, R., Shubina, M., Langer, R., 2006. Biodegradable
   polymeric microspheres and nanospheres for drug delivery in the peritoneum. J. Biomed.

- 789 Mater. Res. Part A 77, 351–361.
- Kondiah, P.J., Choonara, Y.E., Kondiah, P.P.D., Marimuthu, T., Kumar, P., Du Toit, L.C., Pillay,
   V., 2016. A review of injectable polymeric hydrogel systems for application in bone tissue
   engineering. Molecules 21.
- Kristin, C., Robert, S., Michelle, P., 2017. Effects of the intradiscal implantation of stromal
   vascular fraction plus platelet rich plasma in patients with degenerative disc disease. J.
- 795 Transl. Med. 15, 12.
- Latimer, J., Maher, C., Refshauge, K., 2004. The attitudes and beliefs of physiotherapy students
   to chronic back pain. Clin. J. Pain 20, 45–50.
- Le Maitre, C.L., Hoyland, J.A., Freemont, A.J., 2007. Interleukin-1 receptor antagonist delivered directly and by gene therapy inhibits matrix degradation in the intact degenerate human intervertebral disc: an in situ zymographic and gene therapy study. Arthritis Res. Ther. 9, R83.
- Li, C., Wang, T., Hu, L., Wei, Y., Liu, J., Mu, X., Nie, J., Yang, D., 2014. Photocrosslinkable
  bioadhesive based on dextran and PEG derivatives. Mater. Sci. Eng. C 35, 300–306.
- Li, C., Wu, X., Tong, J., Yang, X., Zhao, J., Zheng, Q., Zhao, G., Ma, Z., 2015. Comparative
  analysis of human mesenchymal stem cells from bone marrow and adipose tissue under
  xeno-free conditions for cell therapy. Stem Cell Res. Ther. 6, 55.
- Li, X., Leo, B.M., Beck, G., Balian, G., Anderson, G.D., 2004. Collagen and proteoglycan
  abnormalities in the GDF-5-deficient mice and molecular changes when treating disk cells
  with recombinant growth factor. Spine (Phila. Pa. 1976). 29, 2229–34.
- Li, Z., Tan, B.H., 2015. Towards the development of polycaprolactone based amphiphilic block
   copolymers: Molecular design, self-assembly and biomedical applications. Mater. Sci. Eng.
   C 45, 620–634.
- Liang, C.Z., Li, H., Tao, Y.Q., Peng, L.H., Gao, J.Q., Wu, J.J., Li, F.C., Hua, J.M., Chen, Q.X.,
  2013. Dual release of dexamethasone and TGF-β3 from polymeric microspheres for stem
  cell matrix accumulation in a rat disc degeneration model. Acta Biomater. 9, 9423–9433.
- Liu, S., Liang, H., Lee, S., Li, Z., Zhang, J., Fei, Q., 2016. Isolation and identification of stem
  cells from degenerated human intervertebral discs and their migration characteristics. Acta
  Biochim. Biophys. Sin. (Shanghai). 49, 101–109.
- Liu, Y., Ren, L., Wang, Y., 2013. Crosslinked collagen-gelatin-hyaluronic acid biomimetic film
   for cornea tissue engineering applications. Mater. Sci. Eng. C 33, 196–201.
- Liu, Z., Ma, C., Shen, J., Wang, D., Hao, J., Hu, Z., 2016. SDF-1/CXCR4 axis induces apoptosis
- of human degenerative nucleus pulposus cells via the NF-κB pathway. Mol. Med. Rep. 14,
  783–789.
- Lotz, J.C., Fields, A.J., Liebenberg, E.C., 2013. The role of the vertebral end plate in low back
  pain. Glob. spine J. 3, 153–64.
- Luo, X.-W., Liu, K., Chen, Z., Zhao, M., Han, X.-W., Bai, Y.-G., Feng, G., 2016. Adenovirusmediated GDF-5 promotes the extracellular matrix expression in degenerative nucleus
  pulposus cells. J. Zhejiang Univ. Sci. B 17, 30–42.

- Luo, Z., Jin, L., Xu, L., Zhang, Z.L., Yu, J., Shi, S., Li, X., Chen, H., 2016. Thermosensitive
   PEG–PCL–PEG (PECE) hydrogel as an *in situ* gelling system for ocular drug delivery of
   diclofenac sodium. Drug Deliv. 23, 63–68.
- Maher, C., Underwood, M., Buchbinder, R., 2017. Non-specific low back pain. Lancet 389, 736–
  47.
- Majzoub, R.N., Ewert, K.K., Safinya, C.R., 2016. Cationic liposome-nucleic acid nanoparticle
  assemblies with applications in gene delivery and gene silencing. Philos. Trans. A. Math.
  Phys. Eng. Sci. 374.
- Marfia, G., Campanella, R., Navone, S.E., Zucca, I., Scotti, A., Figini, M., Di Vito, C.,
  Alessandri, G., Riboni, L., Parati, E., 2014. Potential use of human adipose mesenchymal
  stromal cells for intervertebral disc regeneration: a preliminary study on biglycan-deficient
  murine model of chronic disc degeneration. Arthritis Res. Ther. 16, 457.
- Maria, A.T.J., Toupet, K., Maumus, M., Fonteneau, G., Le Quellec, A., Jorgensen, C., Guilpain,
  P., Noël, D., 2016. Human adipose mesenchymal stem cells as potent anti-fibrosis therapy
  for systemic sclerosis. J. Autoimmun. 70, 1–9.
- Maroudas, A., Stockwell, R.A., Nachemson, A., Urban, J., 1975. Factors involved in the nutrition
  of the human lumbar intervertebral disc: cellularity and diffusion of glucose in vitro. J. Anat.
  120, 113–30.
- Mathieu, E., Lamirault, G., Toquet, C., Lhommet, P., Rederstorff, E., Sourice, S., Biteau, K.,
  Hulin, P., Forest, V., Weiss, P., Guicheux, J., Lemarchand, P., 2012. Intramyocardial
  Delivery of Mesenchymal Stem Cell-Seeded Hydrogel Preserves Cardiac Function and
  Attenuates Ventricular Remodeling after Myocardial Infarction. PLoS One 7, e51991.
- Meisel, H.J., Siodla, V., Ganey, T., Minkus, Y., Hutton, W.C., Alasevic, O.J., 2007. Clinical
  experience in cell-based therapeutics: Disc chondrocyte transplantation. A treatment for
  degenerated or damaged intervertebral disc. Biomol. Eng. 24, 5–21.
- Miller, D.G., Adam, M.A., Miller, A.D., 1990. Gene transfer by retrovirus vectors occurs only in
   cells that are actively replicating at the time of infection. Mol. Cell. Biol. 10, 4239–42.
- Mintzer, M.A., Simanek, E.E., 2009. Nonviral vectors for gene delivery. Chem. Rev. 109, 259–
  302.
- Mitragotri, S., Burke, P. a., Langer, R., 2014. Overcoming the challenges in administering
  biopharmaceuticals: formulation and delivery strategies. Nat. Rev. Drug Discov. 13, 655–
  672.
- Miyamoto, T., Muneta, T., Tabuchi, T., Matsumoto, K., Saito, H., Tsuji, K., Sekiya, I., 2010.
  Intradiscal transplantation of synovial mesenchymal stem cells prevents intervertebral disc
  degeneration through suppression of matrix metalloproteinase-related genes in nucleus
  pulposus cells in rabbits. Arthritis Res. Ther. 12, R206.
- Molinos, M., Almeida, C.R., Caldeira, J., Cunha, C., Goncalves, R.M., Barbosa, M.A., 2015.
  Inflammation in intervertebral disc degeneration and regeneration. J R Soc Interface 12, 20141191.
- Moritz, M., Geszke-Moritz, M., 2015. Mesoporous materials as multifunctional tools in
   biosciences: Principles and applications. Mater. Sci. Eng. C 49, 114–151.
- 870 Naves, L., Dhand, C., Almeida, L., Rajamani, L., Ramakrishna, S., Soares, G., 2017. Poly(lactic-

- co-glycolic) acid drug delivery systems through transdermal pathway: an overview. Prog.
  Biomater. 6, 1–11.
- Nettles, D.L., Chilkoti, A., Setton, L.A., 2010. Applications of Elastin-like Polypeptides in Tissue
   Engineering. Adv. Drug Deliv. Rev. 62, 1479–1485.
- Nishida, K., Kang, J.D., Gilbertson, L.G., Moon, S., Suh, J., Vogt, M.T., Robbins, P.D., Evans,
  C.H., 1999. Modulation of the biologic activity of the rabbit intervertebral disc by gene
  therapy: An in vivo study of adenovirus-mediated transfer of the human transforming
  growth factor beta 1 encoding gene. Spine (Phila. Pa. 1976). 24, 2419–2425.
- Orita, S., Eguchi, Y., Kamoda, H., Arai, G., Ishikawa, T., Miyagi, M., Inoue, G., Suzuki, M.,
  Toyone, T., Aoki, Y., Takahashi, K., Ohtori, S., 2011. Brain-Derived Neurotrophic Factor
  Inhibition at the Punctured Intervertebral Disc Downregulates the Production of Calcitonin
  Gene-Related Peptide in Dorsal Root Ganglia in Rats. Spine (Phila. Pa. 1976). 36, 1737–
  1743.
- Orita, S., Miyagi, M., Kobori, S., Gemba, T., Ishikawa, T., Inoue, G., Toyone, T., Aoki, Y.,
  Eguchi, Y., Takahashi, K., Ohtori, S., 2013. IkB kinase b inhibitor downregulates painrelated neuropeptide production in the sensory neurons innervating injured lumbar
  intervertebral discs in the dorsal root ganglia of rats. Spine J. 13, 284–288.
- Orozco, L., Soler, R., Morera, C., Alberca, M., Sanchez, A., Garcia-Sancho, J., 2011.
  Intervertebral disc repair by autologous mesenchymal bone marrow cells: a pilot study.
  Transplantation 92, 822–828.
- Pattappa, G., Peroglio, M., Sakai, D., Mochida, J., Benneker, L.M., Alini, M., Grad, S., Science,
  S., 2014. Ccl5 / Rantes Is a Key Chemoattractant Released By Degenerative Intervertebral
  Discs in Organ Culture. Eur Cell Mater 1, 124–136.
- Pereira, C.L., Gonçalves, R.M., Peroglio, M., Pattappa, G., D'Este, M., Eglin, D., Barbosa, M. a,
  Alini, M., Grad, S., 2014. The effect of hyaluronan-based delivery of stromal cell-derived
  factor-1 on the recruitment of MSCs in degenerating intervertebral discs. Biomaterials 35,
  8144–53.
- Phromsopha, T., Baimark, Y., 2014. Preparation of starch/gelatin blend microparticles by a
  water-in-oil emulsion method for controlled release drug delivery. Int. J. Biomater. 2014, 1–
  6.
- 901 Purcell, B.P., Elser, J.A., Mu, A., Margulies, K.B., Burdick, J.A., 2012. Synergistic effects of
  902 SDF-1a chemokine and hyaluronic acid release from degradable hydrogels on directing bone
  903 marrow derived cell homing to the myocardium. Biomaterials 33, 7849–7857.
- Qiu, Y., Park, K., 2012. Environment-sensitive hydrogels for drug delivery. Adv. Drug Deliv.
   Rev. 64, 49–60.
- Raj, P.P., 2008. Intervertebral disc: Anatomy-physiology-pathophysiology-treatment. Pain Pract.
  8, 18–44.
- 908 Rezaee, M., Oskuee, R.K., Nassirli, H., Malaekeh-Nikouei, B., 2016. Progress in the
- development of lipopolyplexes as efficient non-viral gene delivery systems. J. Control.
  Release 236, 1–14.
- Richardson, S.M., Kalamegam, G., Pushparaj, P.N., Matta, C., Memic, A., Khademhosseini, A.,
  Mobasheri, R., Poletti, F.L., Hoyland, J.A., Mobasheri, A., 2016. Mesenchymal stem cells in

- regenerative medicine: Focus on articular cartilage and intervertebral disc regeneration. 913 914 Methods 99, 69-80.
- 915 Risbud, M. V, Guttapalli, A., Tsai, T.-T., Lee, J.Y., Danielson, K.G., Vaccaro, A.R., Albert, T.J., Gazit, Z., Gazit, D., Shapiro, I.M., 2007. Evidence for skeletal progenitor cells in the 916 degenerate human intervertebral disc. Spine (Phila. Pa. 1976). 32, 2537-2544. 917 doi:10.1097/BRS.0b013e318158dea6
- 918
- 919 Roughley, P., Hoemann, C., DesRosiers, E., Mwale, F., Antoniou, J., Alini, M., 2006. The 920 potential of chitosan-based gels containing intervertebral disc cells for nucleus pulposus supplementation. Biomaterials 27, 388-396. 921
- 922 Roughley, P.J., 2004. Biology of intervertebral disc aging and degeneration: involvement of the 923 extracellular matrix. Spine (Phila. Pa. 1976). 29, 2691–2699.
- 924 Russo, F., Hartman, R.A., Bell, K.M., Vo, N., Sowa, G.A., Kang, J.D., Vadala, G., Denaro, V., 925 2016. Biomechanical Evaluation of Transpedicular Nucleotomy with Intact Annulus Fibrosus. Spine (Phila Pa 1976) 42, E193-E201. 926
- Sainoh, T., Orita, S., Miyagi, M., Inoue, G., Yamauchi, K., Suzuki, M., Sakuma, Y., Kubota, G., 927 Oikawa, Y., Inage, K., Sato, J., Nakata, Y., Aoki, Y., Takahashi, K., Ohtori, S., 2016. Single 928 intradiscal injection of the interleukin-6 receptor antibody tocilizumab provides short-term 929 930 relief of discogenic low back pain; prospective comparative cohort study. J. Orthop. Sci. 21, 931 2–6.
- 932 Sakai, D., Andersson, G.B.J., 2015. Stem cell therapy for intervertebral disc regeneration: 933 obstacles and solutions. Nat. Rev. Rheumatol. 11, 243-256. doi:10.1038/nrrheum.2015.13
- 934 Sakai, D., Grad, S., 2015. Advancing the cellular and molecular therapy for intervertebral disc 935 disease. Adv. Drug Deliv. Rev. 84, 159–171.

Sawamura, K., Ikeda, T., Nagae, M., Okamoto, S., Mikami, Y., Hase, H., Ikoma, K., Yamada, T., 936 937 Sakamoto, H., Matsuda, K., Tabata, Y., Kawata, M., Kubo, T., 2009. Characterization of in 938 vivo effects of platelet-rich plasma and biodegradable gelatin hydrogel microspheres on degenerated intervertebral discs. Tissue Eng. Part A 15, 3719–3727. 939

- Seelbach, R.J., Fransen, P., Pulido, D., D'Este, M., Duttenhoefer, F., Sauerbier, S., Freiman, 940 941 T.M., Niemeyer, P., Albericio, F., Alini, M., Royo, M., Mata, A., Eglin, D., 2015. Injectable 942 Hyaluronan Hydrogels with Peptide-Binding Dendrimers Modulate the Controlled Release of BMP-2 and TGF-b1. Macromol. Biosci. 15, 1035-1044. 943
- 944 Shabbir, A., Zisa, D., Suzuki, G., Lee, T., 2009. Heart failure therapy mediated by the trophic 945 activities of bone marrow mesenchymal stem cells: a noninvasive therapeutic regimen. AJP 946 Hear. Circ. Physiol. 296, H1888-H1897.
- Sionkowska, A., 2011. Current research on the blends of natural and synthetic polymers as new 947 948 biomaterials: Review. Prog. Polym. Sci. 36, 1254-1276.
- 949 Sionkowska, A., Kaczmarek, B., Lewandowska, K., Grabska, S., Pokrywczyńska, M., Kloskowski, T., Drewa, T., 2016. 3D composites based on the blends of chitosan and 950 951 collagen with the addition of hyaluronic acid. Int. J. Biol. Macromol. 89, 442–448.
- 952 Sun, B., Taing, A., Liu, H., Nie, G., Wang, J., Fang, Y., Liu, L., Xue, Y., Shi, J., Liao, Y.-P., Ku, 953 J., Xia, T., Liu, Y., 2016. Nerve Growth Factor-Conjugated Mesoporous Silica
- 954 Nanoparticles Promote Neuron-Like PC12 Cell Proliferation and Neurite Growth. J.

- 955 Nanosci. Nanotechnol. 16, 2390–3.
- Sun, T., Zhang, Y.S., Pang, B., Hyun, D.C., Yang, M., Xia, Y., 2014. Engineered nanoparticles
   for drug delivery in cancer therapy. Angew. Chemie Int. Ed. 53, 12320–12364.
- 958 Teixeira, G.Q., Leite Pereira, C., Castro, F., Ferreira, J.R., Gomez-Lazaro, M., Aguiar, P.,
- Barbosa, M.A., Neidlinger-Wilke, C., Goncalves, R.M., 2016. Anti-inflammatory
- 960 Chitosan/Poly-gamma-glutamic acid nanoparticles control inflammation while remodeling
   961 extracellular matrix in degenerated intervertebral disc. Acta Biomater. 42, 168–179.
- Tran, C.M., Markova, D., Smith, H.E., Susarla, B., Ponnappan, R.K., Anderson, D.G., Symes, A.,
  Shapiro, I.M., Risbud, M. V., 2010. Regulation of CCN2/connective tissue growth factor
  expression in the nucleus pulposus of the intervertebral disc: Role of smad and activator
  protein 1 signaling. Arthritis Rheum. 62, 1983–1992.
- Tran, C.M., Schoepflin, Z.R., Markova, D.Z., Kepler, C.K., Anderson, D.G., Shapiro, I.M.,
  Risbud, M. V, 2014. CCN2 Suppresses Catabolic Effects of Interleukin-1β through α5β1
  and αVβ3 Integrins in Nucleus Pulposus Cells. J. Biol. Chem. 289, 7374–7387.
- Tran, V.T., Karam, J.P., Garric, X., Coudane, J., Benoit, J.P., Montero-Menei, C.N., Venier Julienne, M.C., 2012. Protein-loaded PLGA-PEG-PLGA microspheres: a tool for cell
   therapy. Eur J Pharm Sci 45, 128–137.
- Tuakli-Wosornu, Y.A., Terry, A., Boachie-Adjei, K., Harrison, J.R., Gribbin, C.K., LaSalle, E.E.,
  Nguyen, J.T., Solomon, J.L., Lutz, G.E., 2016. Lumbar Intradiskal Platelet-Rich Plasma
  (PRP) Injections: A Prospective, Double-Blind, Randomized Controlled Study. PM R 8, 1–
  10.
- Vallet-Regi, M., Balas, F., 2008. Silica materials for medical applications. Open Biomed Eng J 2,
   1–9.
- van Rijt, S., Habibovic, P., 2017. Enhancing regenerative approaches with nanoparticles. J. R.
   Soc. Interface 14.
- Vasiliadis, E., Pneumaticos, S., 2014. Biologic treatment of mild and moderate intervertebral disc
   degeneration. Mol. Med. 20, 1.
- Vinatier, C., Gauthier, O., Fatimi, A., Merceron, C., Masson, M., Moreau, A., Moreau, F., Fellah,
  B., Weiss, P., Guicheux, J., 2009. An injectable cellulose-based hydrogel for the transfer of
  autologous nasal chondrocytes in articular cartilage defects. Biotechnol. Bioeng. 102, 1259–
  1267.
- Vos, T., Allen, C., Arora, M., Barber, R.M., Bhutta, Z.A., Brown, A., Carter, A., Casey, D.C.,
  Charlson, F.J., Chen, A.Z., Coggeshall, M., Cornaby, L., Dandona, L., Dicker, D.J., Dilegge,
  T., Erskine, H.E., Ferrari, A.J., Fitzmaurice, C., Fleming, T., Forouzanfar, M.H., Fullman,
- 989 N., Gething, P.W., Goldberg, E.M., Graetz, N., Haagsma, J.A., Hay, S.I., Johnson, C.O.,
- 990 Kassebaum, N.J., Kawashima, T., Kemmer, L., Khalil, I.A., Kinfu, Y., Kyu, H.H., Leung, J.,
- 991 Liang, X., Lim, S.S., Lopez, A.D., Lozano, R., Marczak, L., Mensah, G.A., Mokdad, A.H.,
- 992 Naghavi, M., Nguyen, G., Nsoesie, E., Olsen, H., Pigott, D.M., Pinho, C., Rankin, Z.,
- 993 Reinig, N., Salomon, J.A., Sandar, L., Smith, A., Stanaway, J., Steiner, C., Teeple, S.,
- Thomas, B.A., Troeger, C., Wagner, J.A., Wang, H., Wanga, V., Whiteford, H.A., Zoeckler,
- 995 L., Abajobir, A.A., Abate, K.H., Abbafati, C., Abbas, K.M., Abd-Allah, F., Abraham, B.,
- 996 Abubakar, I., Abu-Raddad, L.J., Abu-Rmeileh, N.M.E., Ackerman, I.N., Adebiyi, A.O.,
- 997 Ademi, Z., Adou, A.K., Afanvi, K.A., Agardh, E.E., Agarwal, A., Kiadaliri, A.A.,

998 Ahmadieh, H., Ajala, O.N., Akinyemi, R.O., Akseer, N., Al-Aly, Z., Alam, K., Alam, 999 N.K.M., Aldhahri, S.F., Alegretti, M.A., Alemu, Z.A., Alexander, L.T., Alhabib, S., Ali, R., 1000 Alkerwi, A., Alla, F., Allebeck, P., Al-Raddadi, R., Alsharif, U., Altirkawi, K.A., Alvis-1001 Guzman, N., Amare, A.T., Amberbir, A., Amini, H., Ammar, W., Amrock, S.M., Andersen, 1002 H.H., Anderson, G.M., Anderson, B.O., Antonio, C.A.T., Aregay, A.F., Arnlöv, J., Artaman, A., Asayesh, H., Assadi, R., Atique, S., Avokpaho, E.F.G.A., Awasthi, A., 1003 Quintanilla, B.P.A., Azzopardi, P., Bacha, U., Badawi, A., Balakrishnan, K., Banerjee, A., 1004 1005 Barac, A., Barker-Collo, S.L., Bärnighausen, T., Barregard, L., Barrero, L.H., Basu, A., 1006 Bazargan-Hejazi, S., Beghi, E., Bell, B., Bell, M.L., Bennett, D.A., Bensenor, I.M., Benzian, H., Berhane, A., Bernabé, E., Betsu, B.D., Beyene, A.S., Bhala, N., Bhatt, S., Biadgilign, S., 1007 1008 Bienhoff, K., Bikbov, B., Biryukov, S., Bisanzio, D., Bjertness, E., Blore, J., Borschmann, 1009 R., Boufous, S., Brainin, M., Brazinova, A., Breitborde, N.J.K., Brown, J., Buchbinder, R., Buckle, G.C., Butt, Z.A., Calabria, B., Campos-Nonato, I.R., Campuzano, J.C., Carabin, H., 1010 Cárdenas, R., Carpenter, D.O., Carrero, J.J., Castañeda-Orjuela, C.A., Rivas, J.C., Catalá-1011 1012 López, F., Chang, J.-C., Chiang, P.P.-C., Chibueze, C.E., Chisumpa, V.H., Choi, J.-Y.J., Chowdhury, R., Christensen, H., Christopher, D.J., Ciobanu, L.G., Cirillo, M., Coates, 1013 1014 M.M., Colquhoun, S.M., Cooper, C., Cortinovis, M., Crump, J.A., Damtew, S.A., Dandona, 1015 R., Daoud, F., Dargan, P.I., das Neves, J., Davey, G., Davis, A.C., Leo, D. De, Degenhardt, L., Gobbo, L.C. Del, Dellavalle, R.P., Deribe, K., Deribew, A., Derrett, S., Jarlais, D.C. Des, 1016 Dharmaratne, S.D., Dhillon, P.K., Diaz-Torné, C., Ding, E.L., Driscoll, T.R., Duan, L., 1017 1018 Dubey, M., Duncan, B.B., Ebrahimi, H., Ellenbogen, R.G., Elyazar, I., Endres, M., Endries, 1019 A.Y., Ermakov, S.P., Eshrati, B., Estep, K., Farid, T.A., Farinha, C.S. e S., Faro, A., Farvid, M.S., Farzadfar, F., Feigin, V.L., Felson, D.T., Fereshtehnejad, S.-M., Fernandes, J.G., 1020 Fernandes, J.C., Fischer, F., Fitchett, J.R.A., Foreman, K., Fowkes, F.G.R., Fox, J., Franklin, 1021 R.C., Friedman, J., Frostad, J., Fürst, T., Futran, N.D., Gabbe, B., Ganguly, P., Gankpé, 1022 1023 F.G., Gebre, T., Gebrehiwot, T.T., Gebremedhin, A.T., Geleijnse, J.M., Gessner, B.D., Gibney, K.B., Ginawi, I.A.M., Giref, A.Z., Giroud, M., Gishu, M.D., Giussani, G., Glaser, 1024 E., Godwin, W.W., Gomez-Dantes, H., Gona, P., Goodridge, A., Gopalani, S.V., Gotay, 1025 1026 C.C., Goto, A., Gouda, H.N., Grainger, R., Greaves, F., Guillemin, F., Guo, Y., Gupta, R., Gupta, R., Gupta, V., Gutiérrez, R.A., Haile, D., Hailu, A.D., Hailu, G.B., Halasa, Y.A., 1027 Hamadeh, R.R., Hamidi, S., Hammami, M., Hancock, J., Handal, A.J., Hankey, G.J., Hao, 1028 Y., Harb, H.L., Harikrishnan, S., Haro, J.M., Havmoeller, R., Hay, R.J., Heredia-Pi, I.B., 1029 Heydarpour, P., Hoek, H.W., Horino, M., Horita, N., Hosgood, H.D., Hoy, D.G., Htet, A.S., 1030 Huang, H., Huang, J.J., Huynh, C., Iannarone, M., Iburg, K.M., Innos, K., Inoue, M., Iyer, 1031 V.J., Jacobsen, K.H., Jahanmehr, N., Jakovljevic, M.B., Javanbakht, M., Jayaraman, S.P., 1032 Jayatilleke, A.U., Jee, S.H., Jeemon, P., Jensen, P.N., Jiang, Y., Jibat, T., Jimenez-Corona, 1033 1034 A., Jin, Y., Jonas, J.B., Kabir, Z., Kalkonde, Y., Kamal, R., Kan, H., Karch, A., Karema, C.K., Karimkhani, C., Kasaeian, A., Kaul, A., Kawakami, N., Keiyoro, P.N., Kemp, A.H., 1035 Keren, A., Kesavachandran, C.N., Khader, Y.S., Khan, A.R., Khan, E.A., Khang, Y.-H., 1036 Khera, S., Khoja, T.A.M., Khubchandani, J., Kieling, C., Kim, P., Kim, C., Kim, D., Kim, 1037 1038 Y.J., Kissoon, N., Knibbs, L.D., Knudsen, A.K., Kokubo, Y., Kolte, D., Kopec, J.A., Kosen, S., Kotsakis, G.A., Koul, P.A., Koyanagi, A., Kravchenko, M., Defo, B.K., Bicer, B.K., 1039 Kudom, A.A., Kuipers, E.J., Kumar, G.A., Kutz, M., Kwan, G.F., Lal, A., Lalloo, R., 1040 1041 Lallukka, T., Lam, H., Lam, J.O., Langan, S.M., Larsson, A., Lavados, P.M., Leasher, J.L., 1042 Leigh, J., Leung, R., Levi, M., Li, Y., Li, Y., Liang, J., Liu, S., Liu, Y., Lloyd, B.K., Lo, 1043 W.D., Logroscino, G., Looker, K.J., Lotufo, P.A., Lunevicius, R., Lyons, R.A., Mackay, M.T., Magdy, M., Razek, A. El, Mahdavi, M., Majdan, M., Majeed, A., Malekzadeh, R., 1044

1045 Marcenes, W., Margolis, D.J., Martinez-Raga, J., Masiye, F., Massano, J., McGarvey, S.T., McGrath, J.J., McKee, M., McMahon, B.J., Meaney, P.A., Mehari, A., Mejia-Rodriguez, F., 1046 1047 Mekonnen, A.B., Melaku, Y.A., Memiah, P., Memish, Z.A., Mendoza, W., Meretoja, A., 1048 Meretoja, T.J., Mhimbira, F.A., Millear, A., Miller, T.R., Mills, E.J., Mirarefin, M., Mitchell, P.B., Mock, C.N., Mohammadi, A., Mohammed, S., Monasta, L., Hernandez, 1049 J.C.M., Montico, M., Mooney, M.D., Moradi-Lakeh, M., Morawska, L., Mueller, U.O., 1050 Mullany, E., Mumford, J.E., Murdoch, M.E., Nachega, J.B., Nagel, G., Naheed, A., Naldi, 1051 L., Nangia, V., Newton, J.N., Ng, M., Ngalesoni, F.N., Nguyen, O. Le, Nisar, M.I., Pete, 1052 P.M.N., Nolla, J.M., Norheim, O.F., Norman, R.E., Norrving, B., Nunes, B.P., Ogbo, F.A., 1053 Oh, I.-H., Ohkubo, T., Olivares, P.R., Olusanya, B.O., Olusanya, J.O., Ortiz, A., Osman, M., 1054 Ota, E., PA, M., Park, E.-K., Parsaeian, M., de Azeredo Passos, V.M., Caicedo, A.J.P., 1055 1056 Patten, S.B., Patton, G.C., Pereira, D.M., Perez-Padilla, R., Perico, N., Pesudovs, K., Petzold, M., Phillips, M.R., Piel, F.B., Pillay, J.D., Pishgar, F., Plass, D., Platts-Mills, J.A., 1057 Polinder, S., Pond, C.D., Popova, S., Poulton, R.G., Pourmalek, F., Prabhakaran, D., Prasad, 1058 1059 N.M., Qorbani, M., Rabiee, R.H.S., Radfar, A., Rafay, A., Rahimi, K., Rahimi-Movaghar, V., Rahman, M., Rahman, M.H.U., Rahman, S.U., Rai, R.K., Raisic, S., Ram, U., Rao, P., 1060 1061 Refaat, A.H., Reitsma, M.B., Remuzzi, G., Resnikoff, S., Reynolds, A., Ribeiro, A.L., 1062 Blancas, M.J.R., Roba, H.S., Rojas-Rueda, D., Ronfani, L., Roshandel, G., Roth, G.A., Rothenbacher, D., Roy, A., Sagar, R., Sahathevan, R., Sanabria, J.R., Sanchez-Niño, M.D., 1063 Santos, I.S., Santos, J.V., Sarmiento-Suarez, R., Sartorius, B., Satpathy, M., Savic, M., 1064 1065 Sawhney, M., Schaub, M.P., Schmidt, M.I., Schneider, I.J.C., Schöttker, B., Schwebel, D.C., 1066 Scott, J.G., Seedat, S., Sepanlou, S.G., Servan-Mori, E.E., Shackelford, K.A., Shaheen, A., Shaikh, M.A., Sharma, R., Sharma, U., Shen, J., Shepard, D.S., Sheth, K.N., Shibuya, K., 1067 Shin, M.-J., Shiri, R., Shiue, I., Shrime, M.G., Sigfusdottir, I.D., Silva, D.A.S., Silveira, 1068 D.G.A., Singh, A., Singh, J.A., Singh, O.P., Singh, P.K., Sivonda, A., Skirbekk, V., Skogen, 1069 J.C., Sligar, A., Sliwa, K., Soljak, M., Søreide, K., Sorensen, R.J.D., Soriano, J.B., Sposato, 1070 L.A., Sreeramareddy, C.T., Stathopoulou, V., Steel, N., Stein, D.J., Steiner, T.J., Steinke, S., 1071 Stovner, L., Stroumpoulis, K., Sunguya, B.F., Sur, P., Swaminathan, S., Sykes, B.L., 1072 Szoeke, C.E.I., Tabarés-Seisdedos, R., Takala, J.S., Tandon, N., Tanne, D., Tavakkoli, M., 1073 Taye, B., Taylor, H.R., Ao, B.J. Te, Tedla, B.A., Terkawi, A.S., Thomson, A.J., Thorne-1074 Lyman, A.L., Thrift, A.G., Thurston, G.D., Tobe-Gai, R., Tonelli, M., Topor-Madry, R., 1075 Topouzis, F., Tran, B.X., Truelsen, T., Dimbuene, Z.T., Tsilimbaris, M., Tura, A.K., Tuzcu, 1076 E.M., Tyrovolas, S., Ukwaja, K.N., Undurraga, E.A., Uneke, C.J., Uthman, O.A., van Gool, 1077 C.H., Varakin, Y.Y., Vasankari, T., Venketasubramanian, N., Verma, R.K., Violante, F.S., 1078 Vladimirov, S.K., Vlassov, V.V., Vollset, S.E., Wagner, G.R., Waller, S.G., Wang, L., 1079 Watkins, D.A., Weichenthal, S., Weiderpass, E., Weintraub, R.G., Werdecker, A., 1080 Westerman, R., White, R.A., Williams, H.C., Wiysonge, C.S., Wolfe, C.D.A., Won, S., 1081 Woodbrook, R., Wubshet, M., Xavier, D., Xu, G., Yadav, A.K., Yan, L.L., Yano, Y., 1082 Yaseri, M., Ye, P., Yebyo, H.G., Yip, P., Yonemoto, N., Yoon, S.-J., Younis, M.Z., Yu, C., 1083 Zaidi, Z., Zaki, M.E.S., Zeeb, H., Zhou, M., Zodpey, S., Zuhlke, L.J., Murray, C.J.L., 2016. 1084 1085 Global, regional, and national incidence, prevalence, and years lived with disability for 310 diseases and injuries, 1990–2015: a systematic analysis for the Global Burden of Disease 1086 Study 2015. Lancet 388, 1545–1602. 1087

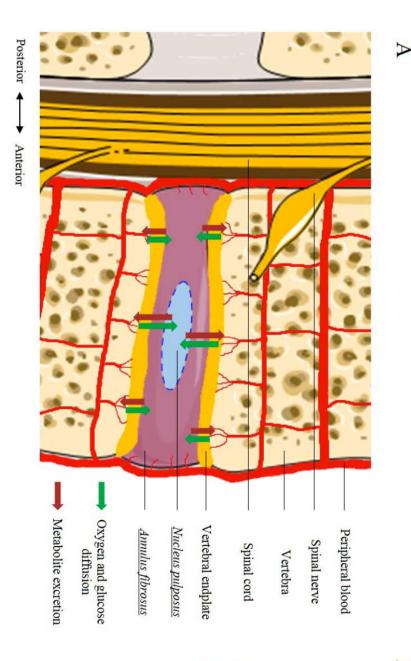
Wallach, C.J., Sobajima, S., Watanabe, Y., Kim, J.S., Georgescu, H.I., Robbins, P., Gilbertson,
 L.G., Kang, J.D., 2003. Gene transfer of the catabolic inhibitor TIMP-1 increases measured
 proteoglycans in cells from degenerated human intervertebral discs. Spine (Phila. Pa. 1976).

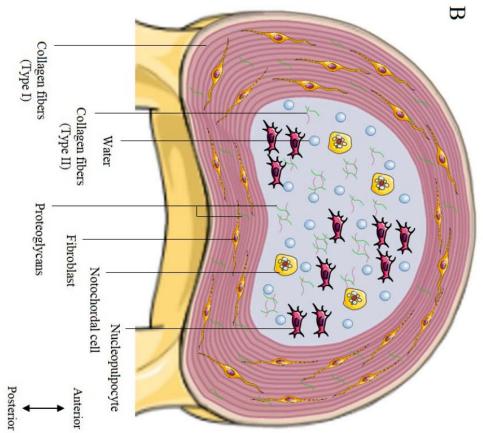
- 1091 28, 2331–2337.
- Walter, B. a, Purmessur, D., Likhitpanichkul, M., Weinberg, A., Cho, S.K., Qureshi, S. a., Hecht,
   A.C., Iatridis, J.C., 2015. Inflammatory Kinetics and Efficacy of Anti-inflammatory
   Treatments on Human Nucleus Pulposus Cells. Spine (Phila. Pa. 1976). 40, 1.
- Wan, S., Borland, S., Richardson, S.M., Merry, C.L.R., Saiani, A., Gough, J.E., 2016. Selfassembling peptide hydrogel for intervertebral disc tissue engineering. Acta Biomater. 46,
  29–40.
- Wang, S., Chang, Q., Lu, J., Wang, C., 2015. Growth factors and platelet-rich plasma: promising
   biological strategies for early intervertebral disc degeneration. Int. Orthop. 39, 927–934.
- Wang, W.-J., Yu, X.-H., Wang, C., Yang, W., He, W.-S., Zhang, S.-J., Yan, Y.-G., Zhang, J.,
  2015. MMPs and ADAMTSs in intervertebral disc degeneration. Clin. Chim. Acta. 448.
- Wei, X.W., Gong, C.Y., Gou, M.L., Fu, S.Z., Guo, Q.F., Shi, S., Luo, F., Guo, G., Qiu, L.Y.,
  Qian, Z.Y., 2009. Biodegradable poly(e-caprolactone)-poly(ethylene glycol) copolymers as
  drug delivery system. Int. J. Pharm. 381, 1–18.
- Wenk, E., Meinel, A.J., Wildy, S., Merkle, H.P., Meinel, L., 2009. Microporous silk fibroin
   scaffolds embedding PLGA microparticles for controlled growth factor delivery in tissue
   engineering. Biomaterials 30, 2571–2581.
- Whatley, B.R., Wen, X., 2012. Intervertebral disc (IVD): Structure, degeneration, repair and
   regeneration. Mater. Sci. Eng. C 32, 61–77.
- Wilczewska, A.Z., Niemirowicz, K., Markiewicz, K.H., Car, H., 2012. Nanoparticles as drug
  delivery systems. Pharmacol Rep 64, 1020–1037.
- Willems, N., Yang, H.-Y., Langelaan, M.L.P., Tellegen, A.R., Grinwis, G.C.M., Kranenburg, H.J.C., Riemers, F.M., Plomp, S.G.M., Craenmehr, E.G.M., Dhert, W.J.A., Papen-Botterhuis,
  N.E., Meij, B.P., Creemers, L.B., Tryfonidou, M.A., 2015. Biocompatibility and intradiscal
  application of a thermoreversible celecoxib-loaded poly-N-isopropylacrylamide MgFelayered double hydroxide hydrogel in a canine model. Arthritis Res. Ther. 17, 214.
- Wismer, N., Grad, S., Fortunato, G., Ferguson, S.J., Alini, M., Eglin, D., 2014. Biodegradable
  electrospun scaffolds for annulus fibrosus tissue engineering: effect of scaffold structure and
  composition on annulus fibrosus cells in vitro. Tissue Eng. Part A 20, 672–82.
- Wu, J., Guan, T., Zheng, S., Grosjean, F., Liu, W., Xiong, H., Gordon, R., Vlassara, H., Striker,
  G.E., Zheng, F., 2011. Inhibition of inflammation by pentosan polysulfate impedes the
  development and progression of severe diabetic nephropathy in aging C57B6 mice. Lab.
  Investig. 91, 1459–1471.
- Yan, J., Yang, S., Sun, H., Guo, D., Wu, B., Ji, F., Zhou, D., 2013. Effects of releasing
  recombinant human growth and differentiation factor-5 from poly(lactic-co-glycolic acid)
  microspheres for repair of the rat degenerated intervertebral disc. J. Biomater. Appl. 29, 72–
  80.
- Yang, H., Gao, F., Li, X., Wang, J., Liu, H., Zheng, Z., 2015. TGF-β1 antagonizes TNF-α
  induced up-regulation of matrix metalloproteinase 3 in nucleus pulposus cells: role of the
  ERK1/2 pathway. Connect. Tissue Res. 8207, 1–8.
- 1131 Yang, Y., Yu, C., 2016. Advances in silica based nanoparticles for targeted cancer therapy.
- 1132 Nanomedicine 12, 317–32.

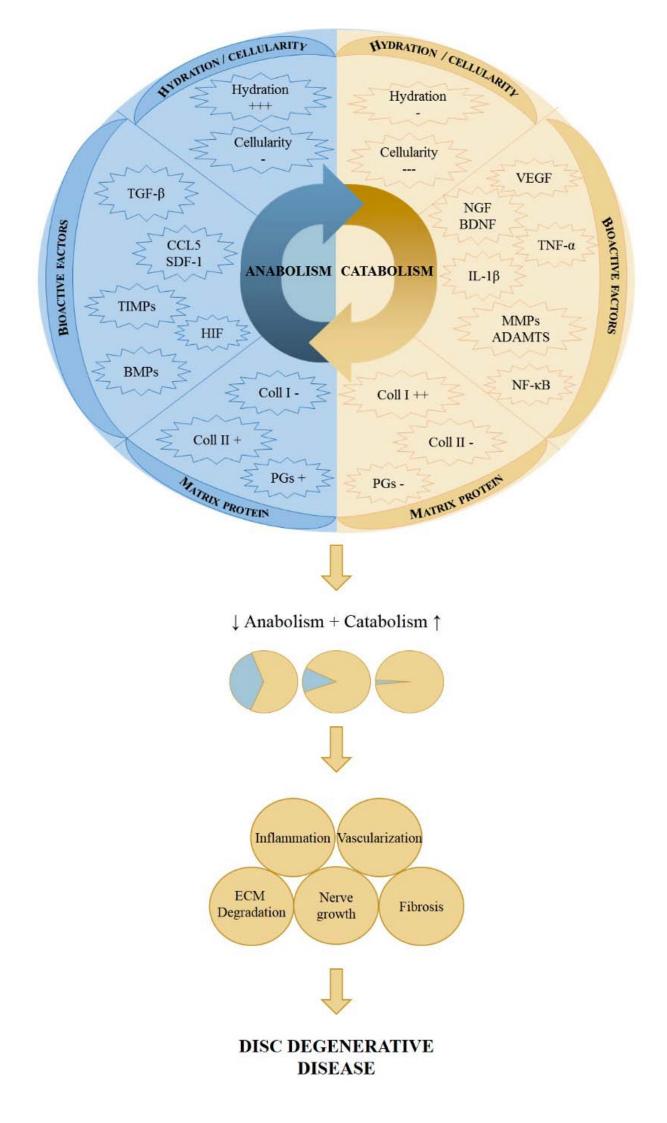
- Zeng, Y., Kurokawa, Y., Win-Shwe, T.-T., Zeng, Q., Hirano, S., Zhang, Z., Sone, H., 2016.
  Effects of PAMAM dendrimers with various surface functional groups and multiple
  generations on cytotoxicity and neuronal differentiation using human neural progenitor
  toxicol. Sci. 41, 351–70.
- Zhang, H., Zhang, L., Chen, L., Li, W., Li, F., Chen, Q., 2014. Stromal cell-derived factor-1 and
  its receptor CXCR4 are upregulated expression in degenerated intervertebral discs. Int. J.
  Med. Sci. 11, 240–245.
- Zhao, Y., Fan, X., Liu, D., Wang, Z., 2011. PEGylated thermo-sensitive poly(amidoamine)
  dendritic drug delivery systems. Int. J. Pharm. 409, 229–236.
- Zhongyi, S., Sai, Z., Chao, L., Jiwei, T., 2015. Effects of nuclear factor kappa B signaling
  pathway in human intervertebral disc degeneration. Spine (Phila. Pa. 1976). 40, 224–32.
- Zhou, X., Chen, L., Grad, S., Alini, M., Pan, H., Yang, D., Zhen, W., Li, Z., Huang, S., Peng, S.,
   2017. The roles and perspectives of microRNAs as biomarkers for intervertebral disc
- 1146 degeneration. J. Tissue Eng. Regen. Med. Ahead of print.
- Zhou, Z., Gao, M., Wei, F., Liang, J., Deng, W., Dai, X., Zhou, G., Zou, X., 2014. Shock
  absorbing function study on denucleated intervertebral disc with or without hydrogel
  injection through static and dynamic biomechanical tests in vitro. Biomed Res. Int. 2014,
- 1150

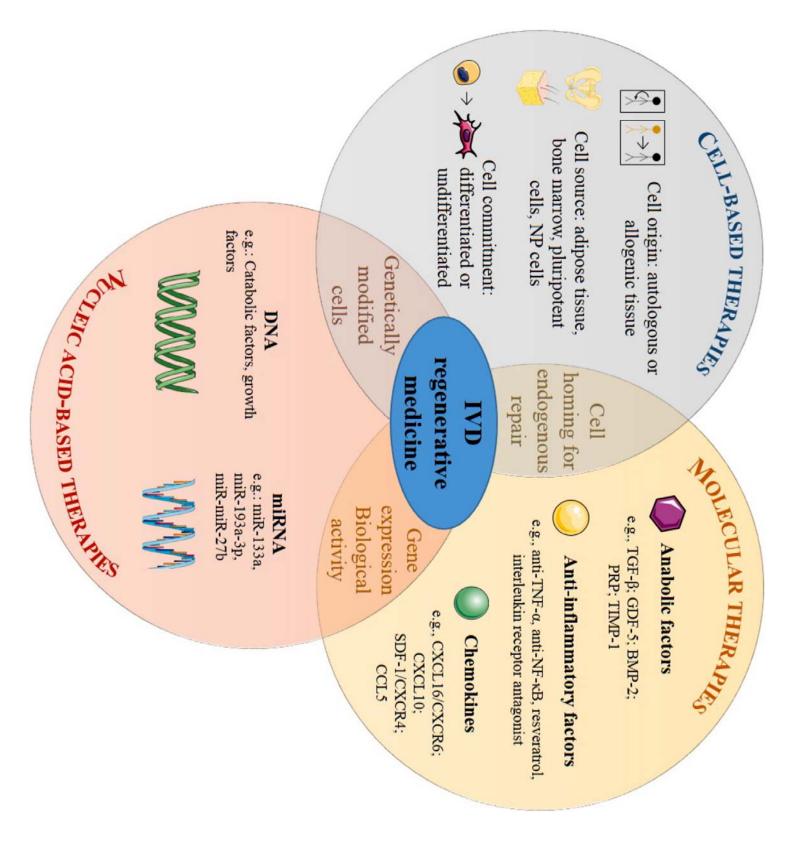
461724.

1152









MULTISCALE SYSTEMS

