

Natural radionuclides 210Po and 210Pb in the Delaware and Chesapeake Estuaries: modeling scavenging rates and residence times

D. Marsan, T. Church, Sylvain Rigaud

▶ To cite this version:

D. Marsan, T. Church, Sylvain Rigaud. Natural radionuclides 210Po and 210Pb in the Delaware and Chesapeake Estuaries: modeling scavenging rates and residence times. Journal of Environmental Radioactivity, 2014, 138, pp.447-455. 10.1016/j.jenvrad.2014.08.014. hal-01717777

HAL Id: hal-01717777

https://hal.science/hal-01717777

Submitted on 26 Feb 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Natural Radionuclides ²¹⁰Po and ²¹⁰Pb in the Delaware and Chesapeake Estuaries; Modeling scavenging rates and residence times Marsan, D., Rigaud, S., and Church, T. School of Marine Science and Policy, University of Delaware, Newark, DE 19716-3501 USA. Journal of Environmental Radioactivity Corresponding author David Marsan dmarsan@udel.edu Phone: (908) 268-9204 Present Address: 013Lammot DuPont Laboratory Newark, DE 19711 USA Sylvain Rigaud s.rigaud@epoc.u-bordeaux1.fr Thomas Church tchurch@udel.edu

50		l Radionuclides ²¹⁰ Po and ²¹⁰ Pb in the Delaware and Chesapeake Estuaries; Modeling ging rates and residence times							
52									
54									
56		ghts Estuarine ²¹⁰ Pb and ²¹⁰ Po data reveal key biogeochemical processes and rates Delaware Bay displays regional differentiation due to dominant particle reaction							
58	3) 4)	Chesapeake Bay displays vertical differentiation from deep sub-oxic redox cycling Parent (210Pb) grand-daughter (210Po) disequilibria evidence principle processes							
60	5)	Net scavenging residence times calculated weeks (Delaware) to months (Chesapeake)							
62									
64									
66									
68									
70									
72									
74									
76									
78									
80									
82									
84									
86									
88									
90									
92									
94									
96									

Abstract

98 During the spring and summer months of 2012, ²¹⁰Po and ²¹⁰Pb activity were measured in the dissolved and particulate phases from the Delaware and upper Chesapeake 100 estuaries. Both nuclides were also determined in the dissolved phase during a tidal cycle from a lower Delaware salt marsh. The upper Delaware estuary, near the freshwater end 102 member, was characterized by high-suspended matter concentrations that scavenged dissolved ²¹⁰Po and ²¹⁰Pb. The intertidal marsh acted as a source for dissolved ²¹⁰Po and 104 ²¹⁰Pb to the Delaware estuary, while deeper within the marsh evidence of bioaccumulation and remineralization of ²¹⁰Po was observed. Box models were applied using mass balance 106 calculations to assess the nuclides residence times in each estuary. A consecutively sectioned (upper, mid, and lower) box-model of the Delaware estuary found the mean 108 dissolved scavenging residence time of ²¹⁰Po to be relatively constant (20-25 days) throughout all of the sections. However, dissolved ²¹⁰Pb scavenging residence times 110 decreased from 21 to 10 days from the upper to lower estuary. Only 60% of the dissolved ²¹⁰Po and 55% of the dissolved ²¹⁰Pb from the Delaware estuary were exported to coastal 112 waters. A large fraction of soluble ²¹⁰Po and ²¹⁰Pb within the estuary was either reversibly adsorbed onto suspended particles or trapped in sediment accumulation zones (such as 114 intertidal marshes), or bioaccumulated by phytoplankton and discharged to the coastal ocean. The upper Chesapeake estuary was largely characterized by sub-oxic bottom waters 116 that contained higher concentrations of dissolved ²¹⁰Po and ²¹⁰Pb, hypothesized to be subjected to redox cycling of manganese. The Delaware and Chesapeake estuary mean 118 residence times for ²¹⁰Po differed significantly at 87 and 126 days respectively, while they were similar for ²¹⁰Pb (53-67 days). The difference in residence times corresponds to the 120 greater extent of biogeochemical scavenging and regeneration processes within the upper Chesapeake.

Keywords

122

126

124 ²¹⁰Po; ²¹⁰Pb; estuaries; salt marsh; Anoxic; Chesapeake Bay; Delaware Bay

1 Introduction

Estuaries are among the most important regions for the transfer of continental dissolved and particulate elements to the oceans (Millward and Turner, 1995). These elements enter the estuarine system via rivers from natural weathering of continental material, anthropogenic discharges from urban areas, groundwater, and atmospheric fallout. In the estuarine region, the elements are subjected to a series of biogeochemical processes in the water and sediments, which include adsorption, transformation, uptake/release by estuarine microorganisms, and removal by biogeochemical scavenging. These processes can result in significant modification of the transfer fluxes from the continent to the oceans. Such transfers associated within the estuarine river and seawater-mixing zone are well established (Church, 1986). However, the rates are difficult to establish without the use of radionuclide tracers to help understand and quantify the changes.

Over the past several decades, the naturally occurring radionuclide pair of ²¹⁰Po and ²¹⁰Pb has been widely used to examine dissolved and particle dynamics in open ocean ecosystems (Bacon et al., 1976; Nozaki, et al. 1976; Thomson and Turekian, 1976; Cochran and Masque, 2003; Rutgers van der Loeff and Moore, 2007). The systems controlling ²¹⁰Po and ²¹⁰Pb dynamics in the ocean are relatively well constrained and mainly associated with sorption onto particles (²¹⁰Pb) and biogenic material cycling (²¹⁰Po). However, in coastal and estuarine systems these dynamics are less constrained due to high temporal and spatial variability. Indeed, estuarine and coastal waters are subjected to seasonal changes of water flow, particle re-suspension/transport from different origins, eutrophication, phytoplankton bloom events, exchange with the ocean, and anthropogenic inputs that influence ²¹⁰Po and ²¹⁰Pb dynamics. In addition, ²¹⁰Po and ²¹⁰Pb radionuclides have several origins in coastal environments. Sources include riverborne sediment (Chung and Chang, 1995; Smoak et al., 1996; Santschi et al., 1979), boundary scavenging of ²¹⁰Pb (Bacon et al., 1976; Chung and Chang, 1995; Smoak et al., 1996), or regeneration of ²¹⁰Po from rapidly degrading biogenic particles or upwelling (Heussner et al., 1990).

Several studies have used the ²¹⁰Pb and ²¹⁰Po nuclide pair in the estuarine marine environment as quantitative tracers (Benninger, 1978; Santschi et al., 1979; Church, 1986; Church and Sarin, 1995; Carvalho, 1997). From this limited research, it has been verified that removal of these nuclides are primarily controlled by the particulate matter concentration (Santschi et al., 1979) and with nuclide disequilibria imposed along the seasonal salinity gradient, which is due to particle scavenging along these gradients (Church

and Sarin, 1995). Estuarine environments are also subject to hypoxic/anoxic conditions with specific biogeochemical dynamic at redox interfaces that clearly impact the ²¹⁰Po and
 ²¹⁰Pb dynamics in the ocean (Bacon et al., 1980; Wei and Murray, 1993; Kim et al., 2005). It was shown that within anoxic basins scavenging of radionuclides by Fe and Mn rich
 particulate matter and by bacteria affects distribution in the water column.

168

170

172

174

176

178

180

182

184

186

188

190

192

194

This paper presents a comprehensive estuarine data set on the radionuclides ²¹⁰Po and ²¹⁰Pb for two of the largest U.S. estuaries, the Delaware and Chesapeake (Figure 1). Both estuaries receive major fresh water input, the Delaware and Susquehanna Rivers, which host the metropolitan regions of Philadelphia and Baltimore, respectively. The Delaware River is shallow, with an average depth of 10m, and bordered by numerous intertidal creeks and salt marshes. The upper Delaware estuary has a well-defined turbidity maximum, and during the spring, bloom events occur within the lower estuary. In contrast, the Chesapeake Bay is a composite of several smaller river-estuarine systems, each with individual lithologies and drainage characteristics. The section north of the Potomac (Figure 1) is a deep drowned river channel sided by clay cliffs that often contain basins of suboxic to anoxic conditions at depth during the summer time. On average, the Delaware estuary has a flushing time averaging 3 months, while the upper Chesapeake flushing can range from 6-12 months. Both estuaries are commensurate with the ²¹⁰Po half-life (138days) and thus compatible with the ²¹⁰Po/²¹⁰Pb radionuclide parent-daughter pair (Church and Sarin, 1995). These estuaries present two distinct biogeochemical systems and provide an overall rationale to use the ²¹⁰Po and ²¹⁰Pb radionuclides to better understand and quantify processes controlling the fate and behavior of other proxy trace elements in estuarine environments.

Here we report data on 210 Po and 210 Pb activities in the dissolved and particulate phases along the salinity gradients of the Delaware and upper Chesapeake estuary. This data is supported by physiochemical water parameters: temperature, salinity, dissolved oxygen, fluorescence, suspended matter, particulate organic carbon (POC) and nitrogen (PON), and major nutrients (NO₃-, NH₄+, PO₄³⁻ and Si) concentrations. The main objectives of this paper are to 1) improve understanding of 210 Po and 210 Pb cycling in the estuarine environment, 2) use the radionuclide pair to quantify dissolved-particulate exchange rates that translate to scavenging residence times 3) assess the potential role of pervasive intertidal salt marshes that may contribute to the nuclide distribution of the lower Delaware Bay, and 4) trace the redox processes in deep waters of the upper Chesapeake.

Quantitatively, this study uses mass balance calculations in order to model the mean scavenging residence times, defined by the mean dissolved 210 Po and 210 Pb adsorptive exchange rates onto particles. Such reaction sorption rates (λ_s ,) are defined as the scavenging rate constant for dissolved 210 Po and 210 Pb onto suspended particles and can be transposed to residence times. The results are compared between the two estuaries, as well as regionally for the Delaware estuary. These rates can be employed as proxies for other trace elements with similar biogeochemical behavior in order to determine their behavior and fate in the estuarine environment.

2 Material and Methods

Water samples for ²¹⁰Po and ²¹⁰Pb analysis were collected during April and August of 2012 for the Delaware and Chesapeake estuaries, respectively. The samples were collected aboard the R/V Sharp along the entire salinity gradient (0-31psu) in the Delaware and the upper Chesapeake estuaries (2-17psu) (Figure 1). At 34 stations surface (2m) and bottom samples at depth were collected using Niskin bottles, supplemented by the surface ship pump (SMS). Additional water samples were taken from intertidal waters of a lower Delaware salt marsh to investigate whether a salt marsh can act as a source or sink for ²¹⁰Po and ²¹⁰Pb radionuclides. This collection occurred over a 12-hour tidal cycle on 01 June 2012 at two sites, near the Roosevelt Inlet (RI) and within Canary Creek (CC) (Figure 1). A single end member sample was collected from Red Mill Pond, a primary freshwater source for the marsh (RMP, Figure 1).

A total of 34 Delaware estuary, 17 intertidal Delaware salt marsh, and 17 Chesapeake dissolved and particulate samples were collected. They were dispersed into 20L Cubitainers® filled with filtered seawater using Gelman Sciences sterilized Mini Capsule Filter membranes (<0.45µm), and acidified to pH 2 using HCl. The cubitainers were sealed and stored in the dark until they were transferred into cold dark storage once ashore. The filtered and acidified seawater samples were assumed to be isolated from storage artifacts based on earlier findings (Chung and Craig, 1983). Separate subsamples were filtered through pre-weighed 0.45µm polycarbonate membrane filters for ²¹⁰Po and ²¹⁰Pb particulates. Separate samples were collected on ashed glass fiber filters for POC and PON and placed in an -80°C freezer until processing. Polycarbonate membrane filters were subjected to HNO₃ digestion at 90°C for a day. Filtrate from POC and PON samples were collected and stored using HCl cleaned vials in order to analyze dissolved bay nutrients

(NO₃-, NH₄+, PO₄³- and Si). Physiochemical parameters such as temperature, salinity, dissolved oxygen, conductivity, fluorimetry, transmissometer were measured in-situ using a
 CTD probes in the estuary and manual probes in the marsh (Seabird and YSI, Inc.).
 Suspended particulate matter (SPM) was determined by weighed mass on the
 polycarbonate filter.

234

236

238

240

242

244

246

248

250

252

254

256

258

260

Dissolved seawater samples were processed for ²¹⁰Po and ²¹⁰Pb using an established protocol (Fleer and Bacon 1984) and modified according to Rigaud et al. (2013). Weighed additions of ²⁰⁹Po isotope spike (1 dpm) and stable Pb carrier (20 mg) were added to each sample for isotopic dilution analysis and chemical yield. The ²⁰⁹Po spike was calibrated using two certified reference materials (IAEA-RGU-1 and NIST-SRM983) and found to be 2.12±0.05 dpm/g (35.3 ±0.8 mBq/g). Stable Pb carrier was calibrated using atomic absorption. Nuclides were extracted by co-precipitation with Fe(OH)₃ (Thomson and Turekian, 1976). The precipitate was recovered by filtration (0.45µm polycarbonate Nucelpore) and then dissolved in a 0.5M HCl solution. In the case of particulate samples, the solid phase is dried, weighed and spiked with ²⁰⁹Po and lead carrier before being completely dissolved using a mixture of strong acids (including HF). The solution digest is evaporated to near-dryness, any residue oxidized with nitric acid, reconstituted after several evaporations in strong HCl, and recovered in 0.5M HCl solution. The ²¹⁰Po and ²⁰⁹Po were plated in this solution by spontaneous deposition onto a silver disc heated in this solution with about 200 mg of ascorbic acid (Flynn, 1968) and their activities measured by alpha spectroscopy. A small aliquot of the solution was taken from this solution in order to measure Pb recovery from the time of collection. The remaining Po in solution was removed in 9 M HCl using AG-1X8 anion exchange resin as described by Sarin et al., (1992). After separation, another small aliquot was taken from the final eluate solution containing the ²¹⁰Pb to gauge the Pb yield during subsequent in-growth of ²¹⁰Po from ²¹⁰Pb and both measured using Flame atomic absorption to determine Pb recovery. The final eluate solution was re-spiked with ²⁰⁹Po and stored at least 6 months to allow in-growth of ²¹⁰Po from ²¹⁰Pb. The ²¹⁰Pb activity of the samples was then determined by plating the eluate solution on another silver disc, via the in-growth of ²¹⁰Po. Finally, the determination of the initial activities of ²¹⁰Po and ²¹⁰Pb at the time of collection was calculated using the equations and corrections accounting for decay and ingrowth of the nuclides, which are detailed in Rigaud et al., (2013). High errors on a few ratios are due to low activity concentrations, as is normal when sample volumes are limited. Overall,

errors were <10% for most samples. Duplicate blanks were determined by processing a
liter of distilled water in the same way as the samples to correct the calculated activities
(<4% sample activity). Detector background was individually subtracted from each sample
activity (<3%). The reproducibility of the method was tested based on duplicate analysis of
five different stations with the relative variations found to be 3.3% between duplicate
analyses.

3. Results

The radionuclide activities and associated errors are presented in Table 1 for the Delaware and Chesapeake estuaries and in Table 2 for the Delaware salt marsh. Two dimensional distribution plots of 210 Po, 210 Pb, and 210 Po/ 210 Pb activity ratios in the dissolved and particulate phases for the Chesapeake and Delaware estuaries were created using Ocean Data View (ODV)(Schlitzer, 2013) (Figures 2 and 4). All of the physiochemical data, which includes temperature, salinity, dissolved oxygen, conductivity, suspended matter and chemical analysis of POC, PON, NO_3 , NH_4 , PO_4 3- and Si, are reported in the Supplemental Materials (including Figures 2 and 4 in color).

3.1 Delaware Estuary

Throughout the Delaware estuary, the salinity and temperature display well mixed features from the surface to within a few meters of the bottom. Increased temperatures were observed in the upper estuary and gradually decreased towards the high salinity coastal waters. Dissolved oxygen (DO) concentrations were constant (93% DO saturation) in the upper estuary, increased to 96% in the surface waters of the mid estuary, and ultimately decreased to concentrations of <90% in the lower estuary. The maximum oxygen and fluorometer signal, along with a decrease in dissolved nutrient concentrations of $NO_{3^{\circ}}$, $NH_{4^{\circ}}$, are indicative of a phytoplankton bloom in the mid to lower estuary. This occurrence is common in the Delaware estuary, as reported by Voynova and Sharp (2012). The upper estuary exhibits low concentrations of suspended particulate matter (3.91mg/L) that dramatically increase to 35 mg/l in the mid estuary, where a turbidity maximum is found.

Dissolved ²¹⁰Po concentrations in the Delaware estuary ranged from 1.3 dpm/100L in the upper estuary to 4.7 dpm/100L in the lower estuary (Table 1 and Figure 2). Dissolved ²¹⁰Pb concentrations varied from 1.0 to 4.0 dpm/100L, with the greatest variation occurring

in the lower estuary deep waters (0.91 dpm/100L) and in the mid estuary where the phytoplankton bloom occurred. The dissolved ²¹⁰Po/²¹⁰Pb activity ratios in the upper estuary (0-50km transect distance) were less than one, while an excess (>1) was present in the mid to lower estuary.

Particulate concentrations of 210 Po and 210 Pb ranged from 1.1 to 18 and 0.42 to 60 dpm/100L, respectively. Particulate 210 Po concentrations were highest in the turbidity maximum, defined by the transmissometer readings and weighed filters of particulate matter. The particulate 210 Pb maximum was located in the intermediate waters of the upper estuary, while the minimum was found at the surface and bottom samples throughout the lower estuary. Of the total concentration in 210 Po, 60% was found in the dissolved phase at the headwaters (sample DB01) and decreased within the turbidity maximum to 11% before increasing again in the lower bay to 68%. Of the total concentration, dissolved 210 Pb activity comprised 12% in the headwaters and dropped to 3% in the turbidity maximum.

3.2 Lower Delaware Salt Marsh (Canary Creek)

Intertidal sampling occurred at two locations, Roosevelt Inlet (entrance) and Canary Creek (upstream). The salinity concentrations over the tidal cycle ranged between 27-29 and 20-24 psu for the Roosevelt Inlet and Canary Creek sampling sites, respectively (Supplemental Materials). Here is noted that little difference between the temperature, dissolved oxygen, and NO_3 - concentrations for each sampling site and over the tidal cycle. POC, PON, Si and PO_4 ³- concentrations were greatest at high tide in Canary Creek, while Roosevelt Inlet concentrations were constant over the tidal cycle (Supplemental Materials).

Dissolved 210 Po ranged from 2.8 to 9.9 dpm/100L at Roosevelt Inlet and 3.9 to 14 dpm/100L at Canary Creek (Table 2 and Figure 3). In the sample collected at Red Mill Pond, the fresh water end member of the creek, 210 Po activity was 9.0 dpm/100L. Over the tidal cycle, 210 Po activities in Canary Creek increased during the flooding tide, while activities increased during low tide at Roosevelt Inlet. Dissolved 210 Pb ranged from 1.2 to 12 dpm/100L at Roosevelt Inlet and from 4.6 to 14 dpm/100L at Canary Creek, while it was higher (19 dpm/100L) at Red Mill Pond (Table 2). The maximum dissolved 210 Po and 210 Pb for Roosevelt Inlet were both observed during low tide. Interestingly, at Canary Creek, farthest from tidal influence, 210 Po reached a maximum at flooding tide and 210 Pb at ebbing tide. Dissolved 210 Po/ 210 Pb activity ratios at Canary Creek indicated a similar to greater proportion of 210 Pb versus 210 Po (\leq 1) during high, ebbing and low tide, while a dissolved

²¹⁰Po excess (>1) was measured during flooding tide (up to 2.3±0.3). In Roosevelt Inlet, ²¹⁰Po/²¹⁰Pb ratio presented the opposite trend, with significant excess at high to ebbing tide and values close to equilibrium during and following low and flooding tide.

3.3. Upper Chesapeake Estuary

During late summer, the upper Chesapeake estuary can be divided into two main zones: an upper zone (from 0-10m) that is vertically well mixed along the upper extent (0-50km), and with deeper waters (10-25m) isolated by a strongly stratified water column along a lower zone (60-120km) according to temperature and salinity (Supplemental Materials). In the vertically well-mixed upper zone, salinity ranged from 3 to 10 psu with a constant temperature of 27°C. In the upper well-mixed zone of the estuary, a turbidity maximum was present with suspended particulate matter concentrations greater than 60mg/L (Supplemental Materials). These high suspended sediment concentrations were constant until midway down the estuary and decreased to <10 mg/L in the lower estuary. Dissolved nutrient concentrations of NO_3 - and Si at the mouth of the Susquehanna River reached a maximum and decreased down the estuary, while the opposite trend was observed for NH_4 + and PO_4 3-. In the upper stratified zone the water column exhibited well-oxygenated surface water (90% DO above 12 m depth) and suboxic/anoxic (O_2 <1% DO) bottom waters below (Supplemental Material).

Dissolved radionuclide concentrations along the upper Chesapeake ranged from 0.8 - 4.4 dpm/100L and 0.4 - 4.3 dpm/100L for ²¹⁰Po and ²¹⁰Pb, respectively (Table 1 and Figure 4). The first sample near the headwaters of the Chesapeake estuary consisted of high dissolved ²¹⁰Po concentrations (4 dpm/100L). These concentrations decreased to 1- 2.5dpm/100L seaward (to 40 km), with a mid-depth water column increase to 5 dpm/100L at 120km (Figure 4). The dissolved ²¹⁰Pb activity was generally constant at 1 dpm/100L throughout the upper Chesapeake estuary, except for two samples: 60km (at surface) and 120km (at depth) transect distance (2-5 and 3-5 dom/100L, respectively). At 60km transect distance, an enrichment of 4.0 dpm/100L of dissolved ²¹⁰Pb was observed in the vicinity of the Baltimore Harbor. North of the Potomac, a suboxic zone was associated with increases in dissolved ²¹⁰Pb and ²¹⁰Po concentrations. Here between 7m and 12m the dissolved ²¹⁰Po (4.4 dpm/100L) and ²¹⁰Pb (3.0 dpm/100L) concentrations were at a maximum before decreasing to 1 dpm/100L in lower oxygenated waters (below 12m). An excess of dissolved

²¹⁰Po/²¹⁰Pb activity ratio (>1) was observed over most of the Chesapeake estuary, and at depth two locations (55km and 120km) reached a ratio of 5 (Table 1 and Figure 4).

Particulate concentrations ranged from 0.3 to 31 dpm/100L and from 0.3 to 67 dpm/100L for ²¹⁰Po and ²¹⁰Pb, respectively. In the upper estuary, the radionuclide particulate concentrations were associated with similar trends to the suspended matter at the turbidity maximum (0-20 km), as noted by Chuch and Sarin, 1995. The highest particulate activity for ²¹⁰Pb was located (120km transect distance) at 15m in the sub-oxic bottom waters (67 dpm/100L), while the highest particulate activity for ²¹⁰Po (31 dpm/100L) was located at 10m depth (15km). The proportion of total ²¹⁰Po in the dissolved phase at the freshwater end member (CB01) was 6% and increased gradually to 26% until 60km transect distance. ²¹⁰Pb increased from 3% to 28% until 60km transect distance. At this point the percent of ²¹⁰Po in the dissolved phase changed to 83% while, the ²¹⁰Pb increased to 58%.

4 Discussion

The ²¹⁰Po and ²¹⁰Pb dynamics will be discussed in association with the main characteristics observed in both estuaries. These characteristics include, the turbidity maximum, phytoplankton blooms, exchanges with surrounding salt marshes along the Delaware estuary, and redox chemistry at the oxic/anoxic transition in the Chesapeake estuary. Finally, box-model calculations for both estuaries will be applied to determine dissolved-particulate sorption rates that translate to scavenging residence times.

4.1 ²¹⁰Po and ²¹⁰Pb cycling in the maximum turbidity zone

Maximum turbidity zones were found in both the upper Delaware and Chesapeake estuaries and corresponded to a decrease of dissolved nuclides and an increase of particulate nuclide activities. This is clearly illustrated when reporting the differential removal of dissolved versus total nuclides. Indeed, outside the turbidity maximum zone, 210 Po is mainly present under dissolved form $59 \pm 6\%$ in the Delaware and $55 \pm 5\%$ in the Chesapeake. However this proportion decreased to $11 \pm 1\%$ and $7 \pm 1\%$ in the turbidity zone for the Delaware and the Chesapeake, respectively. Similarly, 210 Pb is mainly present in the dissolved form $66 \pm 5\%$ in the Delaware and $51 \pm 5\%$ in the Chesapeake outside of the turbidity zone. Within the turbidity zone, the proportion decreases to $3 \pm 1\%$ in the Delaware and $3 \pm 1\%$ in the Chesapeake.

Such observations suggest that ²¹⁰Po and ²¹⁰Pb undergo marked chemical transfers from the dissolved to the particulate phases in the turbidity zone, as ²¹⁰Pb has a strong surface particle affinity (Benninger, 1978). It is suggested that ²¹⁰Pb transfer from the dissolved to particulate phase is due to surface adsorption of ²¹⁰Pb onto surface particles. This is further enhanced in the turbidity zone due to the increase of the available surface particulate sorption sites. However, in the case of ²¹⁰Po, adsorption process and biological uptake may both be involved in ²¹⁰Po transfer, although this is unlikely in the turbidity maximum zone.

These observations in two distinct estuarine systems give evidence that most 210 Po enters estuaries in the dissolved phase. However, most 210 Pb enters the estuaries in the particulate phase or is scavenged further upriver in each system. Within the turbidity maximum, both nuclides are rapidly scavenged as previously reported in estuarine systems (Church and Sarin, 1986; Benninger, 1978). Both studies reported that 210 Pb and 210 Po become largely associated with suspended particles and tightly correlated in the low salinity turbidity maximum. This is in agreement with our findings of nuclide scavenging within the turbidity maximum for both the Delaware and Chesapeake estuaries. The increased 210 Po and 210 Pb activity ratios in the particulate phase (Po/Pb >1) indicating higher scavenging of 210 Po, most likely the result of bioaccumulation.

4.2 ²¹⁰Po and ²¹⁰Pb cycling with phytoplankton bloom

In each estuary, a phytoplankton bloom was present and led to scavenging and subsequent remineralization of nuclides. Within the entire Delaware water column a bloom was observed (75-90km). Correspondingly in the Chesapeake surface waters (0-5m) the bloom was evident from 50-120km. This was indicated by a higher fluorimetry signal coupled with corresponding nutrient data (Supplemental Materials). In these areas the 210 Po and 210 Pb activity ratios in the particulate (Po/Pb >1) indicates higher scavenging of 210 Po from the possibility that bioaccumulation may have occurred. The Delaware bloom displays these conditions with a particulate Po/Pb ratio that ranged from 4 to 6, while within the Chesapeake bloom a ratio of 2.5-3 is observed (Figure 2 and 4). Another illustration to support the observation of bioaccumulation is the dissolved nuclides versus particulate nuclides. The proportion of total dissolved 210 Po in the surface water at the point of the proposed bloom was $17\pm3\%$ in the Delaware and $38\pm4\%$ in the Chesapeake. Notably greater was, the proportion of total dissolved 210 Pb within the bloom area in the Delaware

at 50% and 46% in the Chesapeake. This suggests that biogeochemical processes in the bloom act as efficiently to either solubilize both nuclides or allow greater particulate removal.

Remineralization of nuclides down estuary within the bloom (Delaware) and at depth within the water column (Chesapeake) also leads to increased distribution of dissolved versus particulate \$^{210}\$Po activities. In these areas estuarine conditions resulted in the dissolved ratio \$^{210}\$Po/\$^{210}\$Pb to range from 2-2.5 and 3.5-5 for the Delaware and Chesapeake, respectively (Figure 2 and 4). Here too the proportion of dissolved versus particulate nuclide for dissolved \$^{210}\$Po was 68% in the whole Delaware and 73% in the Chesapeake. Such differential behavior implies that the \$^{210}\$Po has been regenerated in the mid-salinity surface water region (100-150km) within the Delaware and at depth within the Chesapeake. It is important to note that this trend does not continue at deeper depths (>10m), as the waters become suboxic.

Previous studies within the Delaware estuary (Church and Sarin, 1995) included only surface samples and thus were not able to observe the regeneration processes occurring at bottom depths during times of high productivity in the Delaware estuary. Thus it is proposed that the scavenging and regeneration of ²¹⁰Po and ²¹⁰Pb occurs in a similar manner to that at depth in the open ocean. In the open ocean there is a preferential transfer of ²¹⁰Po from the dissolved phase to the particulate phase by bioaccumulation in surface waters and export to the subsurface by the settling of particles (i.e., scavenging) (Stewart, et al., 2007). In a vertically stratified trench, this sorption step appears to be very fast (hours) and becomes evident during times of high productivity, such as during spring and summer months. In deeper ocean waters, the remineralization of particulate matter results in a release of ²¹⁰Po from the particles to the dissolved phase (Bacon et al., 1976, 1978; Cochran et al., 1983; Stewart and Fisher, 2003a; Stewart et al., 2007). It appears that these processes can occur on time scales comparable to larger estuaries as documented in this study.

 $4.3\,^{210}Po$ and ^{210}Pb cycling in salt marshes (and exchange with the Delaware estuary)

The dissolved concentrations for both radionuclides in the Delaware intertidal marsh are generally higher than those measured in the estuary and coastal waters (Table 2). The radionuclides also present strong variations over tidal cycles and furthermore differ between the two sampling sites, located at the entrance of the salt marsh (Roosevelt Inlet; RI) and within the salt march (Canary Creek; CC) (Figure 3). These variations, which

458 transpire over a 12hr tidal cycle, are a result of rather rapid biogeochemical and physical processes occurring within the salt march.

At RI, both dissolved radionuclides follow the same trend throughout the tidal cycle with a maximum activity (10 dpm/100L) present at low tide. During low tide, the dissolved proportion of the water from the salt march at RI is greatest. The high activity values also found at low tide suggest the salt marsh as a source for both ²¹⁰Po and ²¹⁰Pb to Delaware Bay. At low tide, the activity ratio is about 1, which indicates similar and older sources of ²¹⁰Po and ²¹⁰Pb. Conversely, an activity ratio of >1 is present during high and ebbing tide, which evidence a greater proportion of water from the Delaware estuary.

On the other hand, the interior (CC) concentration of dissolved ²¹⁰Po reached a maximum (14 dpm/100L) at flooding tide, while the dissolved concentration of ²¹⁰Pb reached a maximum (14 dpm/100L) during ebbing tide. At flooding tide, ²¹⁰Po activity at CC is notably higher than in the Delaware Bay and RI. In addition, the increase in activity ratio of ²¹⁰Po/²¹⁰Pb >1 during the flooding tidal transit from RI to CC indicates remineralization of ²¹⁰Po-bearing particles, which originated in the Delaware Bay. During high tide in CC, the ²¹⁰Po decreased and reached equilibrium with ²¹⁰Pb, which indicates that the ²¹⁰Po is released then scavenged within the marsh at high tide. Contrarily, the maximum dissolved ²¹⁰Pb activity during ebbing tide may be caused by ²¹⁰Pb inputs within the marsh, such as redox mobilization during groundwater discharge. This input is then conserved during the ebbing tidal transit from CC to RI when a ²¹⁰Pb maximum occurred at low tide, supporting a continuous groundwater discharge.

The difference in nuclide behavior between salt marsh sites indicates that the dissolved ²¹⁰Po from the Delaware Bay entering the marsh is removed during flooding tide. The ²¹⁰Po removal processes within the marsh could also be associated with the ²¹⁰Po bioaccumulation by vegetation (Church, 1986), and ²¹⁰Pb scavenging by sedimentation with other proxy trace elements such as stable Pb (Kim, et al. 2004). Alternatively, dissolved ²¹⁰Po could be precipitated by the marsh sediments during the reduction of sulfur, its elemental analogue (Luther and Church, 1988). In any case, the data suggests that the salt marsh is a probable net source of dissolved ²¹⁰Pb to the surrounding estuary. Such processes would support our findings of higher levels of dissolved ²¹⁰Pb and ²¹⁰Po are found within the marsh compared to the surrounding estuary.

4.4 ²¹⁰Po and ²¹⁰Pb cycling at oxic/suboxic interface

In order to investigate processes in an estuarine redox environment, ²¹⁰Po and ²¹⁰Pb nuclides can be an important tool. The upper Chesapeake Bay contains a pervasive suboxic and anoxic deeper zone, which occurs at depths as shallow as 5 to 10m. However, during the August 2012 cruise only sub-oxic waters were evident below 12m in the mid-estuary (75-120km) (Supplemental Materials). In surface waters (0-10m) from 100-125km, an elevated ²¹⁰Po/²¹⁰Pb activity ratio in the particulate phase of 4 indicates preferential scavenging of ²¹⁰Po from bioaccumulation (Figure 4). The proportion of total particulate ²¹⁰Po in the surface water was 68±5%. As the waters approach suboxic levels, a pycnocline (10m) arises and can be identified by high dissolved ²¹⁰Po and ²¹⁰Pb activity, 5 and 4 dpm/100L, respectively (Table 1). The proportion of total dissolved ²¹⁰Po and ²¹⁰Pb increased at the pycnocline to 86±5% and 63±4%, respectively; indicating that regeneration of the nuclides has taken place. Interestingly, below the pycnocline (12-20m) the activity of both radionuclides in the particulate phase largely increased,10 dpm/100L in ²¹⁰Pb and 80 dpm/100L in ²¹⁰Pb. As such, the proportion of nuclides in the particulate phase increased to 80±7% and 98±6% for ²¹⁰Po and ²¹⁰Pb, respectively.

We hypothesize that the solubilized 210 Po and 210 Pb seen at 12m depth is due to sub-oxic reduction and dissolution of MnO₂. The newly released dissolved 210 Po and 210 Pb diffuse upward in the water column towards the sub-oxic boundary for scavenging by MnO₂. Alternatively, the nuclides could diffuse downward towards anoxic sediments where 210 Pb and 210 Po could be permanently removed from the water column by sulfide co-precipitation. Thus, we propose that in the upper Chesapeake Bay, the water column profiles of 210 Po and 210 Pb in sub-oxic areas are influenced by the redox cycling of Fe/Mn oxides at the water column pycnocline, or Fe sulfides at depth. This is analogous to the vertically stratified anoxic oceanic systems as follows.

There are limited investigations of the aqueous geochemistry of 210 Po and 210 Pb at sub-oxic and anoxic interfaces in the open ocean (Bacon et al., 1980; Todd et al., 1986; Wei and Murray, 1994), and none exist for an estuarine environment. From these few studies we know the following; at the pycnocline particulate 210 Po and 210 Pb show evidence of enrichment relative to mid-depth water column values (Todd, et al., 1986). In the region above the redox interface where MnO₂ begins aerobic reduction and dissolution, MnO₂-bound 210 Po and 210 Pb becomes solubilized (Bacon et al., 1980; Wei and Murray, 1994). In contrast, FeOOH-bound 210 Po and 210 Pb are not released here because Fe does not undergo reduction and dissolution until the anoxia is reached.

Observations from the Chesapeake estuary data agree with the oceanic studies and suggest similar behavior of 210 Po and 210 Pb across the oxic/anoxic interface in an estuarine environment. Previous studies of the Chesapeake Bay characterized the chemical structure of the water column during periods of anoxia (Luther et al., 1994). Prominent features include the presence of a redox front, defined by the O_2/H_2S interface, and the presence of a well defined manganese-rich particulate layer above the interface. In this region particulate Fe (III) and Mn (III, IV) are reduced to their divalent states, thus solubilized with reduced electron activity (Luther et al., 1994). As the dominant carrier phase of many metals and radionuclides, Fe and Mn oxide dissolution and precipitation at the O_2/H_2S interface can determine the fate of such particle reactive elements. Furthermore, redox reaction involving sulfur species in anoxic waters can form aqueous polysulfide ions to efficiently scavenge many free metals, such as Pb^{+2} , to form highly insoluble phases such as galena, PbS (Stumm and Morgan, 1981; Luther and Church, 1988).

4.5 Residence times

Under different conditions within both estuaries (phytoplankton bloom, max turbidity zone, anoxic/oxic interface) both ²¹⁰Po and ²¹⁰Pb are primarily scavenged on particles, inducing disequilibrium, which is used to estimate the net scavenging residence time. A scavenging rate constant for transfer of dissolved ²¹⁰Po and ²¹⁰Pb onto suspended matter is based on a simple box model (Figure 5). This rate can then be translated into scavenging residence times. The Delaware Bay geomorphology matches that of a typical estuary with a broad inner bay bound by two cape entrances, with partially mixed characteristics. As such the Delaware Bay meets the general requirements of a simple box-model based on a mass balance of the radionuclides between phases. From this we assume that measured concentrations of radionuclides (Table 1) are representative of yearly spring and summer annual periods. The estuaries may be characterized by a single (upper Chesapeake estuary) or three successive (upper, mid, and lower Delaware estuary) box-models, each assuming steady-state conditions are maintained for several months (Figure 5).

The balance of dissolved ²¹⁰Po concentrations in the estuary depends upon the following: discharges of dissolved ²¹⁰Po into the estuary, ²¹⁰Po produced through radioactive decay of dissolved ²¹⁰Pb, ²¹⁰Po removal from solution, and ²¹⁰Po exported with the water discharged to the coastal sea. Therefore, the mass balance equation for dissolved ²¹⁰Po in the estuary is

 $\lambda_{Po}A_{Pb} + I_r + I_{smPo} + \lambda_{Po}I_{smPb} + I_{aPo} + \lambda_{Po}I_{aPb} = \lambda_{Po}A_{Po} + \lambda_sA_{Po} + \lambda_wA_{Po} \tag{1}$

All parameters and rates as noted are specified in Figure 5 and Table 3. Substituting the data into the equation (1) and solving for λ_s, the scavenging rate constant in the
 Delaware estuary for ²¹⁰Po onto suspended matter and sediment particles, is 4.2 a⁻¹. A corresponding residence time of ²¹⁰Po can be translated for the whole estuary as 86 days.
 The ²¹⁰Po is exported at a rate that is consistent with about 60% of the annual delivery of dissolved ²¹⁰Po into the estuary. Therefore, only 40% of the dissolved ²¹⁰Po, entering or
 generated in the estuary from the decay of ²¹⁰Pb, is removed by sorption onto sedimentary particles and trapped in the estuary and intertidal marshes.

566

568

570

572

574

576

586

588

In the Delaware estuary sufficient data allows an upper, mid, and lower region to be readily defined (on the basis of salinity) into three box-models zonally distributed to distinguish successive estuarine processes. Similarly, the same equations can be abridged using values for each specific region of the Delaware. Using the concentration of dissolved ²¹⁰Po broken down by region (i.e. upper, mid, and lower) results in scavenging residence times of 22 days, 20 days, and 25 days, with errors of less than one day per region. Though the residence times do not display a large range between regions, they differentiate between different scavenging processes: onto suspended particles (turbidity maximum, upper estuary), biogenic scavenging during phytoplankton blooms (mid estuary), and a combination of regeneration and salt marsh exchange (lower estuary).

An identical mass balance equation may be written for dissolved ^{210}Pb in the Delaware with:

λ_{Pb}A_{Ra} + I_r + I_{smPb} + λ_{Pb}I_{sm} + I_a + λ_{Pb} = λ_{Pb}A_{Pb} + λ_sA_{Pb} + λ_wA_{Pb} (2) where, λ_{Pb} is the radioactive decay constant ²¹⁰Pb, λ_{Pb}A_{Ra} corresponds to the ²¹⁰Pb
 ingrowth from dissolved ²²⁶Ra in the estuary and the other variables similar to those already introduced (Figure 5 and Table 3). The contribution of ²²⁶Ra from atmospheric deposition is assumed to be negligible. The input of dissolved ²²⁶Ra through radioactive decay as a source of total dissolved ²¹⁰Pb also is also assumed to be minimal, compared with other ²¹⁰Pb sources primarily weathering of particles in the watershed or atmospheric deposition.

Solving equation (2), λ_s , defined as the scavenging rate constant for dissolved ^{210}Pb onto suspended sediment particles, is 5.5 a-1, and as transposed inversely, a mean residence time of soluble ^{210}Pb in the whole Delaware estuary of 67 days. This discharge of soluble ^{210}Pb to coastal waters accounts for 55% of the annual input received by the estuary, which

includes atmospheric deposition. Therefore, most of the ²¹⁰Pb entering the estuary in the soluble phase is either absorbed onto suspended particles and trapped in sediment accumulation zones of the estuary, such as intertidal marshes, or discharged via binding to suspended matter into the coastal ocean. As with ²¹⁰Po, the ²¹⁰Pb residence time for the Delaware estuary can be differentiated by zonal region resulting in times of 21 days (upper), 14 days (mid), and 10 days (lower) that notably decrease down estuary.

Utilizing a similar approach, mean residence times for surface waters of the upper portions of the Chesapeake estuary in the summer were calculated using a single box-model. As the sampling focused only on the upper Chesapeake, a zonal approach was not applied. The upper estuary of the Chesapeake is defined here as the oxic region extending from the mouth of the Susquehanna River, seaward to the mouth of the Potomac Estuary and from 0-5 meters in depth. This region encompassing the upper bay includes the eastern and western tributaries, which include 1484 km² and 14% of the total Chesapeake Bay area (Ranasinghe et al., 1998). Unlike the Delaware estuary, there are essentially no significant intertidal marshes surrounding the upper bay, thus no marsh exchange is considered. Using the data from Table 1, the scavenging rate constant for dissolved ²¹⁰Po onto suspended matter and sediment particles is 2.91a-1. The mean residence time of ²¹⁰Po in solution relative to adsorption on sediment particles is translated to be 125 days, while dissolved ²¹⁰Pb scavenging rate is 6.6a-1, with the mean residence time translated as 55 days.

Comparing the two single-box model residence time results of ²¹⁰Po for the entire estuary, Delaware (87 days) and the Chesapeake (126 days), suggest that biogenic cycling is greater in the upper Chesapeake estuary. However, ²¹⁰Pb scavenging residence times of the Delaware (66 days) and the Chesapeake (53 days) suggest that the physicochemical adsorption on particles is slightly greater in the Delaware. It is hypothesized that other trace elements with similar chemical characteristics as ²¹⁰Pb (Type A) and ²¹⁰Po (Type B) will present similar sorption rates. "Type A" trace metals tend to form cations, which react with negatively charged ions and suspended particles, including elements such as Fe, Mn, Co and Cr (Church, 1986). However, "Type B" metals tend to form stronger complexes with organic matter, which include elements such as Zn, Ni, Cu, Cd, Ag, and Hg.

5.1 Conclusions

The following conclusions and implications of geochemical cycling of ²¹⁰Po and ²¹⁰Pb in the estuarine waters of the Delaware and Chesapeake Bays can be made. Both upper

estuaries are influenced by maximum turbidity zones, which correspond to a decrease in dissolved and an increase of particulate nuclide activities. Outside the turbidity maximum zone, 210 Po is mainly present in the dissolved form at $59 \pm 6\%$ in the Delaware and $55 \pm 5\%$ in the Chesapeake, while this proportion decreased to $11 \pm 1\%$ and $7 \pm 1\%$ in the turbidity zone for the Delaware and the Chesapeake, respectively.

Phytoplankton blooms in both estuaries scavenge ²¹⁰Po in surface waters and are regenerated at depth similar to processes seen in the open ocean. The scavenging by biological particles is one of the main processes that affect nuclide distribution found in lower sections of the estuaries. Regeneration occurs at depth for more stratified systems in the Chesapeake and further down the estuary in the Delaware. In a lower Delaware salt marsh sources of ²¹⁰Pb and ²¹⁰Po are evident during tidal cycling. Within the Delaware salt marsh scavenging and regeneration of ²¹⁰Po occurs, whereby ²¹⁰Po is released by biological particles during low tide and scavenged during high tide. In the Chesapeake estuary redox chemistry at the oxic/suboxic transition show a release of radionuclides at the pycnocline and scavenging as the waters become more anoxic, proposed evidence of redox cycling from Fe/Mn oxides. In the Delaware, box-model calculations result in ²¹⁰Po residence times for three successive regions similar at 20-25 days, while ²¹⁰Pb decreased from 20 to 10 days from the upper to lower estuary. The upper Chesapeake has a longer residence time, 125 days for ²¹⁰Po and 55 days for ²¹⁰Pb due to greater biological scavenging and stratified waters than the Delaware. The use of the radionuclides 210Po and 210Pb to determine residence times can also be applied as proxies for other Type A (210Po) or Type B (210Pb) trace elements and thus potential pollutant impacts such elements have on estuarine systems.

646

624

626

628

630

632

634

636

638

640

642

644

648

650

652

654

656

658

Acknowledgements -Dr. Luther for providing research cruise time in the Delaware and Chesapeake supported by NSF/OCE grants 1031272 and 1059892. -Graduate support for D. Marsan was provided by NSF Grant- HRD-1139870- Greater Philadelphia Region LSAMP Bridge to the Doctorate (Cohort IX) -Post-Doctoral support for S. Rigaud was provided by NSF/OCE grant 0961653.

REFERENCES

708	ALL LALLACED
710	Bacon, MP., Spencer, D.W., Brewer, P.G. 1976. 210Pb/226Ra and 210Po/210Pb disequilibria in seawater and suspended particulate matter. Earth Planet. Sci. Lett 32, 277-296.
712	Bacon, M.P., Brewer, P.G., Spencer, D.W., Murrary, J.W., 1980. Lead-210, polonium-210 manganese and iron in the Cariaco Trench. Deep Sea Res. 27, 119-135.
714716	Benninger, L.K.,(1978) 210Pb balance in Long Island Sound. Geochimica et Cosmochimica Acta 42 (8), 1165-1174.
718	Carvalho, F., 1997. Distribution, cycling and mean residence time of 226Ra, 210Pb and 210Po in the Tagus estuary. The Science of the Total Environment 196, 151-161.
720 722	Chung, Y., Chang, W.C., 1995. Pb-210 fluxes and sedimentation rates on the lower continental slope between Taiwan and south Okinawa Trough. Continental Shelf Research
724	15, 149–164. Chung, Y., and Craig, H. 1983. 210Pb in the Pacific: the GEOSECS measurements of
726	particulate and dissolved concentrations. Earth and Planetary Sci., 65 406-432.
726 728	Church, T.M., 1986. Biogeochemical factors influencing the residence time of microconstituents in a large tidal estuary, Delaware Bay. Mar. Chem. 18, 393-406.
730	Church, T.M., M. Bernat & P. Sharma, 1987. Distribution of natural uranium, thorium, lead and polonium radionuclides in tidal phases of a Delaware salt marsh. Estuaries 9: 2-8.
732 734	Church, T., Sarin, M 1995. Natural U-Th series radionuclide studies of estuarine processes,in: Dyer, K.R., Orth, R.J. (Eds.), Changes in Fluxes in Estuaries: Implication From Science to Management. Proceedings of the ECSA22/ERF Symposium, Plymouth, England, September 13-18,1992, pp.91-95.
736738	Cochran, JK., Masqué,, P.,. 2003. Short-lived U/Th Series Radionuclides in the Ocean: Tracers for Scavenging Rates, Export Fluxes and Particle Dynamics. Reviews in Mineralogy and Geochemistry 52, 461-492.
740	Fleer, A.P., Bacon, M.P., 1984. Determination of 210Pb and 210Po in seawater and marine particulate matter. Nuclear Instruments and Methods in Physics Research 223, 243-249.
742744	Flynn, W. W., 1968. The determination of low levels of polonium-210 in environmental materials. Anal. Chim. Acta 43, 221-227.
746	Heussner, S., Cherry, R.D., Heyraud, M., 1990. ²¹⁰ Po, ²¹⁰ Pb in sediment trap particles on a Mediterranean continental margin. Continental Shelf Research 10, 989–1004.
748	Kim, G., Hussain, N. Church, T., 2000. Excess 210Po in the coastal atmosphere. Tellus. 52B, 74-80.
750	

- Kim, G., Alleman, L.Y., and T.M Church. 2004. Accumulation records of radionuclides
- and trace metals in two contrasting Delaware salt marshes." Marine Chemistry, 87: 87-96.
- 754
 - Kim, G., Kim, S., Harada, K., Schultz, M., Burnett, W. 2005. Enrichment of Excess 210Po in
- Anoxic Ponds. Environ. Sci. Technol. 39, 4894-4899.
- Luther, G.W. III, Church, T.M., 1988. Seasonal cycling of sulfur and iron in porewaters of a Delaware salt marsh. Mar. Chem. 23, 295-310.
- 760
- Luther, G.W. III, Church, T.M., 1988. Seasonal cycling of sulfur and iron in porewaters of a
- 762 Delaware salt marsh. Mar. Chem. 23, 295–310.
 - Luther, G.W. III, Nuzzio, D.B., Wu, J., 1994. Speciation of manganese in Chesapeake Bay
- 764 waters by volumetric methods. Anal. Chim. Acta 284, 473-480.
- McKee, B.A., Todd, J.F., 1993. Uranium behavior in a permanently anoxic fjord: microbial control? Limnol. Oceanogr. 38, 408-414.
- 768
- Millward, G.E., Turner, A., 1995. Trace elements in estuaries. In: Salbu, B., Steinnes, E. (Eds.),
- 770 Trace Metals in Natural Waters. CRC Press, Boca Raton, FL, pp. 223–245.
- Nozaki, Y., 1986. 226Ra-222Rn-210Pb systematics in seawater near the bottom of the
- ocean. Earth and Planetary Science Letters 80, 36-40.
- Nozaki, Y., J. Thomson, and K. K. Turekian. 1976. The distribution of 210Pb and 210Po in the surface waters of the Pacific Ocean. Earth Planet. Sci. Lett 32, 304-312.
- 776
- Ranasinghe, J.A., Scott, L.C. Kelly, F.S., 1998. Chesapeake Bay Water Quality Monitoring
- Program Long-term benthic monitoring and assessment component. July 1984-Dec 1997. Versar, Inc., Columbia, MD.
- 780
- Rigaud, S., Puigcorbé, V., Camara-Mor, P., Casacuberta, N., Roca-Martí, M., Garcia-Orellana, J.,
- Benitez-Nelson, C.R., Masque, P., Church, T. submitted, 2013. An assessment of the methods, calculation and uncertainties in the determination of 210Po and 210Pb activities in
- 784 seawater. Limnol. Oceanogr. Methods.
- Rutgers van Der Loeff, M. M., Moore, W.S., 2007. Determination of natural radioactive tracers, p. 365-397, in: Methods of Seawater Analysis. Wiley-VCH Verlag GmbH.
- 788
- Santschi, P.H., Li, Y.H., Bell, J., 1979. Natural radionuclides in the water of Narragansett Bay.
- 790 Earth Planet. Sci. Lett., 45, 201-213.
- Sarin, M. M., Bhushan, R., Rengarajan, R., Yadav, D.N. 1992. The simultaneous determination of 238U series nuclides in seawater: results from the Arabian Sea and Bay of Bengal. Indian
- 794 J. Mar. Sci. 21, 121-127.
- Schlitzer, R., 2013. Ocean Data View, http://odv.awi.de.

798	Sharp, J. Coulberson, C. Church, T., 1982. The chemistry of the Delaware estuary. General Considerations. Limnol. Oceanogr., 27(6), 1015-1028.
800	Smoak, J.M., DeMaster, D.J., Kuehl, S.A., Pope, R.H., McKee, B.A., 1996. The behavior of
802	particle-reactive tracers in a high turbidity environment: 234 Th and 210 Pb on the Amazon continental shelf. Geochimica et Cosmochimica Acta 12, 2123–2137.
804	Stewart, G., Cochran, J., Xue, J., Lee, C., Wakeham, S., Armstrong, R., Masque, P., Miquel, J. 2007. Exploring the connection between 210Po and organic matter in the northwestern
806	Mediterranean. Deep Sea Research I. 54, 415-427.
808	Stewart G.M. and Fisher, N.S. 2003a. Experimental Studies on the accumulation of polonium 210 by marine phytoplankton. Limnology and Oceanography. 48(3): 1192-1201.
810	Stewart, G., Fowler, S., and Fisher, N. 2007. Chapter 8: The Bioaccumulation of U- and Th-
812	Series Radionuclides in Marine Organisms. Radioactivity in the Environment, Volume 13.
814	Stumm, W., Morgan, J.J., 1981. Aquatic Chemistry. Wiley, New York, 780 pp.
816	Thomson, J., Turekian, K.K., 1976. 210Po and 210Pb distributions in ocean water profiles from the Eastern South Pacific. Earth Planet. Sci. Lett 32, 297-303.
818	Todd, J.F., Wong, G.T.F., Reid, D., 1986. The geochemistries of 210Po and 210Pb in waters
820	overlying and within the Orca Basin, Gulf of Mexico. Deep Sea Res. 33, 1293-1306.
822	Voynova, Y. and Sharp, J. 2012. Anomalous Biogeochemical Response to a Flooding Event in the Delaware Estuary: A Possible Typology Shift Due to Climate Change. Estuaries and Coasts 35:943-958.
824	
826	Wei, C, and Murray, J. 1993. The behavior of scavenged isotopes in marine anoxide environments: 210Pb and 210Po in the water column of the Black Sea. Geochmica et al. 1995. The state of the scale of t
828	Cosmochimica Acta, Vol. 58, No. 7, 1795-1811.
830	
832	
834	
836	
838	

Table and Figure Captions

840	
842	Table 1 : Dissolved and particulate ²¹⁰ Po and ²¹⁰ Pb activities (dpm/100L) for the Delaware Bay (DB) and salt marsh (DBSMS), and Chesapeake Bay (CB).
844	Table 2 : Dissolved ²¹⁰ Po and ²¹⁰ Pb activities (dpm/100L) for Roosevelt Inlet (RI) and Canary Creek (CC) lower Delaware Bay salt marsh.
846 848	Table 3: Box model units, rates, and residence times for the Delaware Bay and upper Chesapeake Bay estuaries
850 852 854	Figure 1: Location of the sampling sites in the Delaware Bay and Chesapeake Bay estuaries The dashed line delineates the regions modeled. Samples were collected as follows Delaware estuary 2012 April 1-3, Delaware salt marsh two locations over a 12hi tidal cycle 2012 June 1, and Chesapeake estuary 2012 August 20-25. The shaded area denotes deeper estuarine basins.
856 858	Figure 2 : Distribution of dissolved and particulate ²¹⁰ Po and ²¹⁰ Pb activities (dpm/100L) and ²¹⁰ Po/ ²¹⁰ Pb activity ratios distribution in the Delaware Bay estuary. Contour plots were carried out using the DIVA gridding tool of Ocean Data View software Dashed lines delineate the regions modeled (upper, mid and lower estuary).
860 862	Figure 3 : Dissolved ²¹⁰ Po, ²¹⁰ Pb activities (dpm/100L) and ²¹⁰ Po/ ²¹⁰ Pb activity ratios in Roosevelt inlet and Canary Creek over a 12-hr tidal cycle.
864 866 868	Figure 4: Distribution of dissolved and particulate ²¹⁰ Po and ²¹⁰ Pb activities (dpm/100L) and ²¹⁰ Po/ ²¹⁰ Pb activity ratios distribution in the upper Chesapeake Bay estuary Contour plots were carried out using the DIVA gridding tool of Ocean Data View software. Dashed lines delineate the regions modeled (upper, mid and lower estuary).
870 872	Figure 5: Box model depiction and symbol notation for dissolved ²¹⁰ Po in the Delaware Bay and upper Chesapeake Bay estuary. The Delaware Bay utilized a regional (3 box model) estuary model, while the Chesapeake used a single box model for the entire upper estuary.
874	
876	
878	
880	
882	

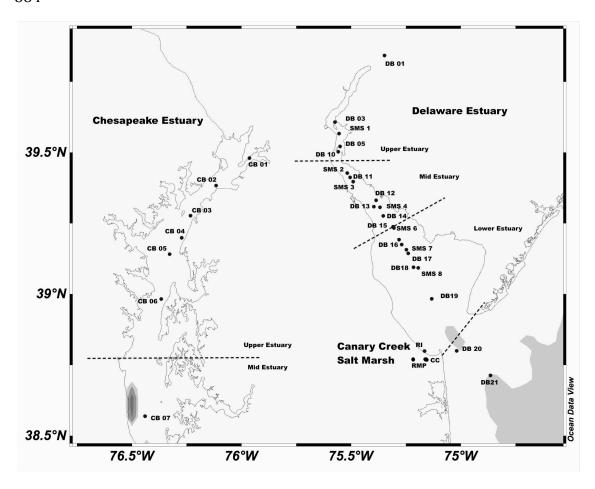


Figure 1

Table 1

Station	Salinity	Oxygen	Depth	2	¹⁰ Po _{di}	iss	21	⁰ Po _{pa}	ert	21	¹⁰ Pb _{di}	iss	21	⁰ Pb _{pa}	art	²¹⁰ Pc	o/ ²¹⁰ P	b _{diss}	²¹⁰ Po/ ²¹⁰ Pb _{part}		
	(psu)	(µmol/L)	(m)		m/10			m/10			m/10		(dpm/100L)								
DB 01	0.1	323	11.8	1.8	±	0.2	1.2	±	0.7	1.9	±	0.3	13.1	±	1.6	0.9	±	0.2	0.1	±	0.0
DB 01	0.1	325	2.4	1.4	±	0.2	1.3	±	1.1	2.2	±	0.4	15.2	±	2.1	0.7	±	0.1	0.1	±	0.0
DB 03	2.0	319	10.4	1.6	±	0.1	4.7	±	1.0	3.0	±	0.2	16.7	±	2.0	0.5	±	0.1	0.3	±	0.1
DB 03	2.0	321	2.8	1.7	±	0.2	4.8	±	1.0	3.2	±	0.4	16.8	±	2.1	0.5	±	0.1	0.3	±	0.1
DB 05	7.2	320	2.0	2.6	±	0.2	18.4	±	1.9	1.4	±	0.3	15.9	±	3.2	1.9	±	0.4	1.2	±	0.3
DB 10	6.7	327	1.8	1.9	±	0.2	14.8	±	2.0	2.1	±	0.3	57.8	±	5.0	0.9	±	0.2	0.3	±	0.1
DB 10	8.5	319	9.1	2.0	±	0.2	15.4	±	1.0	2.1	±	0.4	60.1	±	4.9	0.9	±	0.2	0.3	±	0.1
DB 11	8.2	337	2.0	2.3	±	0.2	4.9	±	1.7	3.2	±	0.4	9.1	±	2.7	0.7	±	0.1	0.5	±	0.1
DB 11	12.4	315	10.0	2.0	±	0.2	6.0	±	1.0	1.7	±	0.3	14.7	±	2.9	1.2	±	0.2	0.4	±	0.1
DB 12	16.5	315	12.9	2.5	±	0.2	11.0	±	1.7	3.8	±	0.4	20.1	±	2.0	0.7	±	0.1	0.5	±	0.2
DB 12	10.9	342	2.2	1.8	±	0.2	8.4	±	2.6	1.0	±	0.3	10.8	±	2.9	1.8	±	0.6	0.8	±	0.2
DB 13	12.3	353	2.2	2.2	±	0.2	6.8	±	1.3	1.3	±	0.3	10.7	±	2.4	1.7	±	0.4	0.6	±	0.2
DB 13	15.4	300	10.1	2.3	±	0.2	13.1	±	1.5	1.3	±	0.3	14.4	±	2.8	1.8	±	0.5	0.9	±	0.2
DB 14	21.5	308	2.1	2.5	±	0.2	16.1	±	2.3	1.6	±	0.3	14.5	±	3.0	1.6	±	0.3	1.1	±	0.2
DB 14	16.2	347	15.4	2.2	±	0.2	13.3	±	2.6	2.4	±	0.3	14.1	±	4.3	0.9	±	0.2	0.9	±	0.3
DB 15	18.8	330	2.0	4.7	±	0.4	4.0	±	1.0	3.9	±	0.4	0.6	±	0.3	1.2	±	0.2	6.8	±	2.3
DB 15	23.7	307	14.3	2.1	±	0.2	2.1	±	0.9	2.0	±	0.3	0.6	±	0.3	1.1	±	0.2	3.9	±	1.4
DB 16	25.5	306	2.1	3.9	±	0.3	1.8	±	0.9	2.8	±	0.3	0.4	±	0.3	1.4	±	0.2	4.5	±	1.2
DB 16	21.5	318	11.7	2.6	±	0.2	1.4	±	1.1	1.6	±	0.3	1.2	±	0.9	1.6	±	0.3	1.2	±	0.8
DB 17	24.1	310	1.9	3.2	±	0.3	1.6	±	1.0	3.1	±	0.4	0.5	±	0.3	1.0	±	0.2	3.2	±	1.0
DB 17	26.0	301	12.1	3.4	±	0.3	2.1	±	0.9	2.9	±	0.4	0.6	±	0.3	1.2	±	0.2	3.5	±	1.1
DB 18	26.3	306	2.0	4.5	±	0.4	3.4	±	1.4	2.2	±	0.3	1.2	±	1.0	2.1	±	0.3	2.7	±	0.8
DB 19	28.2	297	2.0	3.3	±	0.3	4.4	±	1.2	1.6	±	0.3	2.1	±	1.3	2.0	±	0.4	2.1	±	1.5
DB 20	31.8	297	1.6	2.5	±	0.2	1.6	±	0.8	2.2	±	0.3	2.4	±	1.2	1.1	±	0.2	0.6	±	0.3
DB 21	30.4	295	2.0	2.2	±	0.2	2.3	±	1.3	1.8	±	0.2	2.4	±	2.0	1.3	±	0.1	1.0	±	0.3
DB 21	31.0	292	15.3	2.7	±	0.2	1.1	±	0.4	1.2	±	0.3	1.0	±	0.4	2.2	±	0.5	1.1	±	0.3
DBSMS 1	5.2	326	2.0	1.5	±	0.2	15.1	±	1.8	1.3	±	0.3	14.2	±	2.3	1.1	±	0.3	1.1	±	0.1
DBSMS 2	10.3	323	2.0	2.0	±	0.2	9.1	±	2.2	1.6	±	0.3	14.0	±	2.5	1.2	±	0.3	0.7	±	0.2
DBSMS 3	13.2	315	2.0	2.4	±	0.2	10.0	±	2.0	1.3	±	0.3	11.8	±	1.8	1.9	±	0.5	0.9	±	0.2
DBSMS 4	15.1	311	2.0	1.3	±	0.2	7.5	±	1.3	1.0	±	0.3	10.5	±	2.5	1.3	±	0.4	0.7	±	0.3
DBSMS 5	17.6	308	2.0	2.5	±	0.3	3.7	±	1.1	3.8	±	0.5	0.8	±	0.2	0.6	±	0.1	4.8	±	1.9
DBSMS 6	21.2	303	2.0	1.5	±	0.2	1.4	±	0.9	1.7	±	0.3	0.8	±	0.4	0.9	±	0.2	1.8	±	0.6
DBSMS 7	23.3	302	2.0	4.3	±	0.3	1.7	±	0.9	2.1	±	0.4	0.6	±	0.2	2.0	±	0.4	2.9	±	1.0
DBSMS 8	25.9	301	2.0	3.9	±	0.3	3.3	±	1.2	2.0	±	0.3	1.4	±	0.9	2.0	±	0.4	2.4	±	1.0
DESIVIS	23.3	301	2.0	3.5		0.5	3.3	_		2.0		0.5	1.7		0.5	2.0		0.4		_	1.0
CB 01	12.7	188	4.8	2.1	±	0.2	0.8	±	0.1	1.2	±	0.3	1.8	±	0.3	1.8	±	0.5	0.4	±	0.1
CB 01	14.2	130	10.4	1.4	±	0.2	0.5	±	0.2	0.6	±	0.3	3.1	±	0.5	2.2	±	1.1	0.2	±	0.1
CB 01	15.9	34	17.7	2.8	±	0.2	0.5	±	0.2	0.9	±	0.3	1.0	±	0.4	3.2	±	1.2	0.5	±	0.2
CB 01	17.1	2	22.0	2.2	±	0.2	1.3	±	0.1	0.7	±	0.3	0.7	±	0.3	3.0	±	1.2	1.8	±	0.5
CB 02	13.1	153	3.0	1.5	±	0.2	2.4	±	0.1	0.6	±	0.3	0.7	±	0.3	2.5	±	1.2	3.5	±	1.4
CB 02	14.2	71	8.2	4.4	±	0.4	5.3	±	0.1	3.0	±	0.5	1.9	±	0.3	1.5	±	0.3	2.8	±	0.1
CB 02	15.5	41	13.9	2.7	±	0.4	10.7	±	0.1	1.0	±	0.5	67.1	±	0.9	2.6	±	1.3	0.2	±	0.6
CB 02	17.0	1	21.9	1.1	±	0.3	2.6	±	0.2	4.3	±	1.0	0.3	±	0.3	0.2	±	0.1	8.3	±	2.5
CB 02	17.0	1	21.5	1.3	±	0.2	1.6	±	0.1	0.4	±	0.3	0.3	±	0.2	3.2	±	2.7	4.3	±	1.8
CB 02	10.8	142	0.8	1.6	±	0.3	0.3	±	0.2	2.7	±	0.3	1.9	±	0.2	0.6	±	0.1	0.2	±	0.0
CB 03	13.3	23	9.1	2.5	±	0.2	6.9	±	0.1	0.5	±	0.4	2.8	±	0.1	4.7	±	2.7	2.5	±	0.0
CB 03	8.0	188	1.1	1.4	±	0.2	5.5	±	0.1	0.5	±	0.3	1.8	±	0.2	2.0	±	0.9	3.1	±	0.2
CB 04 CB 05	6.6	177	1.1	2.0	±	0.2	3.3	±	0.1	2.4	±	0.5	3.3	±	0.2	0.8	±	0.9	1.0	±	0.3
CB 05	10.0	96	9.1	2.0	±	0.3	9.6	±	0.1	0.9	±	0.5	3.3 10.7	±	0.2	2.1	±	1.1	0.9	±	0.0
CB 05	3.2	175	0.5	0.8	±	0.3	10.0	±	0.1	0.9	±	0.3	7.6	±	0.2	1.9	±	1.1	1.3	±	0.0
CB 00	2.1	173	0.5	2.6	±	0.2	3.5	±	0.2	1.1	±	0.5	34.8	±	0.5	2.3	±	1.1	0.1	±	0.0
CB 07	3.1	155	9.0	2.0	±	0.4	31.3	±	0.2	0.9	±	0.5	34.8 27.6	±	0.5	2.3	±	0.9	1.1	±	0.0
CB U/	3.1	132	5.0	۷.1	Ξ	U.Z	51.5	Ξ	ט.ט	0.9	Ξ	U.3	۷./۵	Ξ	U./	2.3	Ξ	0.9	1.1	Ξ	U.U

DB: Delaware Bay Stations DBSMS: Delaware Bay Surface Mapping System

CB: Chesapeake Bay Stations

Table 2:

Station	tion Salinity Tide Height			^{l0} Po _{di}	ss	210	Pb _{di}	ss	²¹⁰ Po,	²¹⁰ Po/ ²¹⁰ Pb _{diss}			
(psu)				m/10			n/10						
RI 01	26.8	high tide	4.3	±	0.3	1.7	±	0.4	2.5	±	0.5		
RI 02	27.4	ebbing	3.5	±	0.3	1.2	±	0.4	2.8	±	0.5		
RI 03	27.6	ebbing	5.8	±	0.5	3.8	±	0.4	1.5	±	0.2		
RI 04	27.4	ebbing	2.8	±	0.2	1.9	±	0.4	1.5	±	0.3		
RI 05	25.8	low tide	9.9	±	0.8	11.6	±	1.1	0.9	±	0.1		
RI 06	25.7	flooding	5.0	±	0.4	4.2	±	0.5	1.2	±	0.2		
RI 07	27.9	flooding	3.5	±	0.3	2.5	±	0.5	1.4	±	0.3		
RI 08	28.1	flooding	3.3	±	0.3	2.6	±	0.7	1.2	±	0.4		
CC 01	21.7	high tide	6.7	±	0.6	6.4	±	0.7	1.0	±	0.1		
CC 02	23.2	ebbing	3.9	±	0.4	7.2	±	0.8	0.5	±	0.1		
CC 03	21.2	ebbing	6.1	±	0.7	6.4	±	0.7	1.0	±	0.2		
CC 04	20.2	ebbing	6.7	±	1.1	13.8	±	1.5	0.5	±	0.1		
CC 05	19.5	low tide	4.8	±	0.7	5.5	±	0.7	0.9	±	0.2		
CC 06	18.2	flooding	8.3	±	0.7	4.6	±	0.6	1.8	±	0.3		
CC 07	19.3	flooding	10.1	±	0.9	8.7	±	0.9	1.2	±	0.2		
CC 08	20.5	flooding	13.9	±	1.0	6.0	±	0.7	2.3	±	0.3		
RMP	0.4	low tide	9.0	±	1.4	18.8	±	2.5	0.5	±	0.1		

RI: Roosevelt Inlet Marsh

RMP: Red Mill Pond end member

CC: Canary Creek Marsh

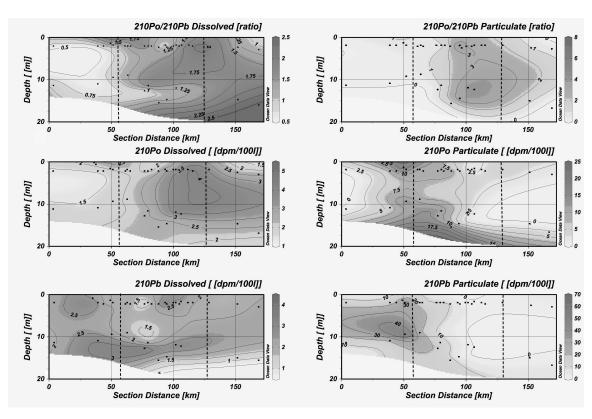


Figure 2

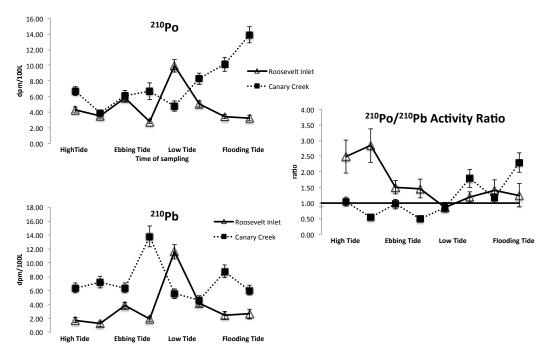


Figure 3

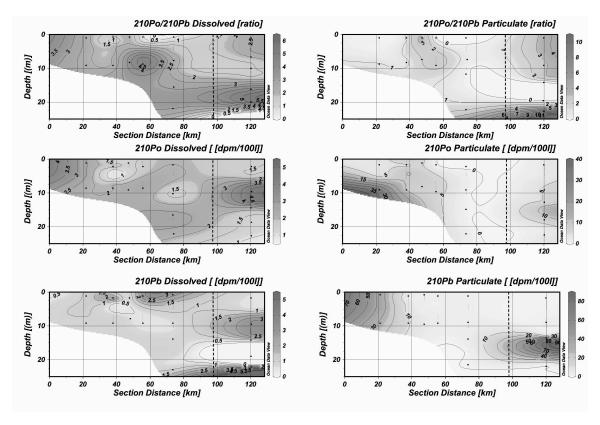


Figure 4

Full estuary Regional estuary I_{a2} I_{a1} I_{sm1} I_{sm2} A_{Po3} A_{Po1} A_{Po2} I_{r1} I_{r3} I_{r4} Upper estuary Mid-estuary Lower estuary $\mathsf{A}_{\mathsf{Ra1}}$ A_{Pb1} $\mathsf{A}_{\mathsf{Pb3}}$ $\mathbf{A}_{\mathrm{Ra2}}$ $\mathsf{A}_{\mathsf{Ra3}}$ \mathbf{A}_{Pb2} $\begin{array}{c} \lambda_{Po/Pb} \\ (a^{\text{-}1}) \end{array}$ Radioactive decay Annual input of dissolved constant Ingrowth from dissolved estuary $\begin{array}{c} A_{Po/Pb/Ra} \\ \text{(Bq m}^{\text{-3}}) \end{array}$ parent or grandparent λ_w (a⁻¹) Constant for water mass Annual input of dissolved outflow and influx from ${\rm I}_{\rm r(Po/Pb/Ra)} \\ {\rm (Bq~m^{-3})}$ nuclide higher salinity waters I_{sm(Po/Pb/Ra)} Annual input of (Bq m⁻³) dissolved nuclida $\begin{matrix} \lambda_s \\ (a^{\text{-}1}) \end{matrix}$ Scavenging rate constant dissolved nuclide from intertidal salt marsh

Figure 5

974

976

978

980

982

984

986

Table 3

	λ(a ⁻¹)	A(dpm/100L)	I,(dpm/100L)	I _{sm} (dpm/100L)	I _a (dpm/100L/a ¹)	λ _s (a ⁻¹)	λ _w (a ⁻¹)	Residence Time (d)
	man manusan	240 (²¹⁰ Po)	480 (²¹⁰ Po)	480 (²¹⁰ Po)	The transfer of the second			*** ** participation on the co
	1.8 (²¹⁰ Po)	180 (210 Pb)	240 (²¹⁰ Pb)	780 (²¹⁰ Pb)	1.3x10 ⁻⁵ (²¹⁰ Po)	4.2 (²¹⁰ Po)		87(²¹⁰ Po)
Delaware	3.1x10 ⁻² (²¹⁰ Pb)	540 (²²⁶ Ra)	180 (²²⁶ Ra)	1080 (226Ra)	1.3x10 ⁻³ (²¹⁰ Pb)	5.5 (²¹⁰ Pb)	2.9	66 (²¹⁰ Pb)
	Marian Marian	180 (²¹⁰ Po)	240 (²¹⁰ Po)	1750 (670)	15> 41 100> 16> 41	1 1000000		1000 mm
	1.8 (²¹⁰ Po)	120 (²¹⁰ Pb)	120 (²¹⁰ Pb)		1.3x10 ⁻⁵ (²¹⁰ Po)	2.9 (²¹⁰ Po)		126 (²¹⁰ Po)
Chesapeake	3.1x10 ⁻² (²¹⁰ Pb)	1860 (²²⁶ Ra)	600 (²²⁶ Ra)	-	1.3x10 ⁻³ (²¹⁰ Pb)	6.6 (²¹⁰ Pb)	1.0	53 (²¹⁰ Pb)