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## Transient biogeochemistry in intertidal sediments: new insights from tidal pools in Zostera noltei meadows of Arcachon Bay (France)

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#### Abstract

Several studies highlighted the occurrence of circular pools in intertidal flats of different coastal systems and their transient water chemistry over both tidal and diurnal cycles. However, little is known about (1) the response of benthic biogeochemical reactions and fluxes at the sediment-water interface over such short time scales, and (2) the role of these tidal pools in the biogeochemical functioning of coastal systems. Based on *in situ* microprofiles and water sampling, we investigated the dynamics of dissolved oxygen (O<sub>2</sub>), nutrients, sulfides and metals, and the associated fluxes at the sediment-water interface in tidal pools from the Arcachon Bay (Atlantic coast of France). Our integrative approach included several tidal and diurnal cycles over two different seasons in the presence and absence of Zostera noltei. The results show that water temperature and light irradiance were the main factors driving the biogeochemical functioning of the tidal pools, as they controlled the physiological activity of the microphytobenthos. Changes in light radiations induced diurnal fluctuations of  $O_2$  concentrations within surficial sediment, thus resulting in fluctuations of the  $O_2$ diffusive fluxes at the sediment-water interface and of the O<sub>2</sub> penetration depth in sediment. At high tide, the increase in turbulence above the sediment induced the advection of oxygenated water within the first millimeters of sediment, resulting in a significant increase in porewater  $O_2$  concentrations and sediment O2 penetration depth. Porewater sulfide concentrations and apparition depth were concomitant with the  $O_2$  dynamic over both diurnal and tidal cycles, indicating that intermediate redox diagenetic processes were impacted by O<sub>2</sub> dynamic over such short time-scale. The rapid changes in redox processes in the sediment column are confirmed by a significant flux of dissolved manganese toward the water column during nighttime. The consumption of nitrate and the release of ammonium and phosphate, associated to the mineralization of the organic matter in the surface sediment did not appeared related however to such short time cycles. The efflux of dissolved silica from the sediment was most likely associated with the enhanced dissolution of Si-bearing particles in surface sediment at higher temperatures, although silica uptake by Z. noltei was also noted. This study clearly shows that tidal pools function as natural incubators of transient biogeochemical processes. A rough assessment of the nutrient budget at the scale of the bay indicates the tidal pools may contribute significantly to the biogeochemical functioning of Arcachon Bay.

Keywords: tidal pools; biogeochemistry; sediment-water interface; fluxes; Zostera noltei

#### 1. Introduction

In shallow coastal areas, the functioning of pelagic and benthic compartments is tightly coupled (Soetaert et al., 2000). Physical and chemical properties, such as temperature light, salinity, concentration of dissolved oxygen and nutrients, strongly vary over time scales ranging from hours to years in the coastal water column due to tidal, diurnal and seasonal cycles. These environmental evolutions profoundly impact the sediment biogeochemistry and chemical exchanges at the sediment-water interface, which in turn impact the water column chemistry. Assessing the transient dynamics of sedimentary biogeochemical processes is of primary importance to better understand the functioning of coastal ecosystems and to predict their evolution in a changing world.

Intertidal zones are particularly suitable to investigate the transient dynamics of sedimentary biogeochemical processes as they undergo strong changes in light, temperature, nutrient availability and hydrodynamic conditions over time, from tidal to seasonal time scales (e.g., Taillefert et al., 2007; Jansen et al., 2009). Recent works showed that the intensity of light irradiance was the main factor controlling the short-term O<sub>2</sub> dynamic in intertidal sediments over diurnal and tidal cycles (Jansen et al., 2009; Denis et al., 2012; Delgard et al. 2012). During the daytime, O<sub>2</sub> generally presents higher concentrations and penetration depths in sediments due to the photosynthetic activity of microphytobenthos, which may result in a net  $O_2$  production in surface sediments and  $O_2$  flux from the sediment to the water column (Bottcher et al., 2000; De Beer et al., 2005; Werner et al., 2006; Jansen et al., 2009; Delgard et al., 2012). Moreover, Denis et al. (2012) indicated that benthic photosynthetic production was generally higher during emersion periods than during inundation periods, as the light irradiance could be strongly attenuated through the water column. Previous studies also reported a vertical migration of microphytobenthos during tidal cycles in intertidal sediments that significantly modified the photosynthetic activity in surficial sediments (Consalvey et al., 2004; Migné et al., 2009; Denis et al., 2012; Delgard et al. 2012). The changes in  $O_2$  concentration in surficial sediments directly impact the fate of redox species through modifications of early diagenetic processes such as denitrification, nitrification, and metal (e.g., Fe and Mn) and sulfide oxidation (Deflandre et al., 2002; Dalsgaard, 2003; Taillefert et al., 2007; Glud, 2008; Rigaud et al., 2013). Indeed, the increase in O<sub>2</sub>

penetration depth in sediments lowers the sulfate reduction rates (Billerbeck et al., 2006) and intensifies the oxidation of reduced species produced in the anoxic part of the sediment ( $NH_4^+$ ,  $Fe^{2+}$ , sulfides S(-II) and FeS; Taillefert et al., 2007; Delgard et al., 2012), thereby directly modifying the exchanges of nutrients and metals at the sediment-water interface (Rigaud et al., 2013).

The main limitation in understanding the biogeochemical processes and chemical transfers at the sediment-water interface in intertidal environments is the specific conditions occurring at low tide, notably the emersion periods that interrupt the exchanges at the sediment-water interface and induce perturbations such as the evaporation of porewater and desiccation of the top layer of sediments. Interestingly, intertidal flats may present topographic depressions where water can be trapped during low tide (e.g., Van der Laan and Wolff, 2006). These environments, referred as tidal pools, are particularly exposed to temporal variations in light and temperature because of their small volumes and shallow depths (e.g., Morris and Taylor, 1983; Clavier et al., 2011). Tidal pools have transient chemical compositions over tidal cycles due to chemical exchanges at the sediment-water interface (Lillebø et al., 2004; Murray et al., 2006). Thus, tidal pools can be considered as natural incubators, where it is possible to investigate the dynamics of sedimentary biogeochemical processes in response to changes in the environmental conditions in the overlying water. Indeed, the benthic-pelagic coupling is expected to be amplified due to the reduced volume of the pelagic compartment. In the present study, we characterized the transient biogeochemistry of sediments in response to the variation of external forcing (temperature, light, water depth) over tidal and diurnal cycles in tidal pools from the Arcachon Bay. Biogeochemical processes and fluxes at the sediment-water interface were characterized and quantified from continuous in situ measurement of O<sub>2</sub>, H<sub>2</sub>S and pH in porewaters using an autonomous miniprofiler and from temporal variations of major biogeochemical species concentration  $(O_2, NO_3^-, tCO_2, PO_4^{3-}, NH_4^+, Si_d, Mn_d and Fe_d)$  in the overlying water trapped in the pool at low tide. The contribution of driving factors was evaluated by statistical treatments. A chemical budget at the scale of the bay is proposed to estimate the role of tidal pools to the biogeochemical functioning of Arcachon Bay.

#### 2. Material and Methods

#### 2.1. Study area and sampling

The tidal pools were located in an intertidal mudflat of Arcachon Bay (44°42.737N; 1°08.097W), a macrotidal lagoon along the French Atlantic coast characterized by a semi-diurnal tide with an amplitude ranging from 1.2 m to 4.4 m (Figure 1A). The pools at the study site were topographic depressions, a few meters wide and up to 20 cm depth (Figure 1B). Lagoon water was trapped in the pools for approximately 4-5 hours around low tide, until the rising tide reached the study site. The intertidal flat consisted of silty loam sediment colonized by a dense meadow of the dwarf eelgrass Zostera noltei. Although their origin is still not known (i.e., hydrodynamic vs. biological structure; see discussion in Van der Laan and Wolff, 2006; Takeuchi and Tamaki, 2014), the tidal pools presented consistent features in Arcachon Bay. Aerial photos of the sampling site enabled to estimate a surface coverage of 20% for the tidal pools (Figure 1C). Arcachon lagoon pools were similar to those previously described by van der Laan and Wolff (2006) in the Banc d'Arguin (Mauritania). Three field campaigns were carried out in July 2010 and August and October 2013. In July 2010, four different pools were studied during the same day: two pools densely covered by Zostera noltei and two unvegetated pools (Table 1). The water trapped in the four pools was sampled in triplicate every hour throughout an emersion period in the daytime (between 13:30 and 16:20). Pools studied in August and October 2013 were both unvegetated. In August 2013, the pool water was sampled during two consecutive periods of emersion, including one daytime (between 14:40 and 18:40) and one night (between 02:20 and 06:30). In October 2013, the pool water was sampled during emersion periods occurring during three consecutive days (between 12:00 and 18:00). Both in August and October 2013, sampling of the pool water started approximately one hour before the complete disconnection of the pool during the ebb tide and finished about one hour after the pools reconnected with the bay water during the flood tide. Water samples were collected in the central part of the pool using a 250 mL Nalgene beaker fixed on a long rod with a sampling frequency of 20-30 minutes. Samples were immediately filtered using acetate cellulose 0.2 µm syringe filters and were partitioned in either plastic or glass vials according to the type of analysis to be performed. The subsamples were stored untreated in gastight glass vials for total dissolved inorganic carbon ( $tCO_2 = CO_2 + HCO_3^2 + CO_3^2$ ) analysis or

collected in polyethylene vials and acidified with a 1% equivalent volume of concentrated HNO<sub>3</sub> for dissolved Fe and Mn analysis. The subsamples for dissolved nutrients (NO<sub>3</sub>, NH<sub>4</sub>, PO<sub>4</sub>, Si<sub>d</sub>) analysis were transferred into two 12 mL polyethylene vials. Samples for tCO<sub>2</sub>, metals and Si<sub>d</sub> were kept refrigerated until analysis, while those for other nutrients were immediately frozen. The analysis of these samples was completed within a month after sampling.

At the end of 2013 campaigns, three 10 cm long and 2.5 cm diameter sediment cores were collected and sliced at 2 mm vertical resolution for porosity assessment. Porosity was obtained on each sediment slice by mass differences between fresh and freeze-dried sediment following correction for salt content, and assuming a sediment particle density of 2.65 g.cm<sup>-3</sup>.

#### 2.2. In situ measurements in the pool water

Temperature, salinity, dissolved  $O_2$  concentration, photosynthetically active radiation (PAR) reaching the sediment surface and water pressure (i.e., depth) were continuously monitored in the pools using *in situ* autonomous probes. Oxygen concentration and temperature were recorded using a SDOT300 data logger (*NKE Instruments*) equipped with an *Aanderaa* optode 3835. Salinity, temperature and water pressure were recorded using a STPS 100-SI data logger (*NKE Instruments*). PAR was monitored using a SPAR data logger (*NKE Instruments*) equipped with a flat LI-192 sensor (*LI-COR Corporate*). The SDOT and STPS probes were mounted on a foot of the miniprofiler MP6 system (see below) and positioned to be immerged in the pool water, while the SPAR probe was directly inserted into the pool sediment. During the 2010 campaign, the SDOT and STPS sensors were deployed in only one of the four pools, while temperature was manually recorded in the other at each sampling time. Oxygen concentrations were compensated for salinity, temperature and depth using the Interactive TD 280 spreadsheet (AADI, *Aanderaa*). The precisions of measurements were  $\pm$  5% for oxygen, 0.1°C for temperature, 0.1 for salinity and 0.1 m for water depth. Water depth within the pools was precisely measured in the field with a ruler. All parameters were recorded every 5 minutes.

#### 2.3. In situ microelectrode measurements

The vertical distribution and temporal evolution of temperature, pH, O<sub>2</sub> and H<sub>2</sub>S at the sediment-water

interface were measured using an autonomous *in situ* miniprofiler MP6 system (Unisense A/S). The miniprofiler was deployed at high tide during the two campaigns of 2013, but only the data obtained during the October campaign was exploitable. The benthic MP6 miniprofiler is a 6-channel system equipped with a temperature sensor (200 µm tip), a pH sensor (500 µm tip) with a reference electrode, a  $H_2S$  sensor (100 µm tip) and three oxygen sensors (100 µm tips). The tips of the temperature,  $H_2S$ and pH sensors were positioned at the same level, which was 25 mm below the oxygen sensors tips, and manually positioned on field at about 10 mm above the sediment-water interface. The reference electrode was mounted on the frame of the profiler to be continuously emerged. Two sets of measurements were carried out sequentially. The first set consisted of measurements along vertical profiles from +10 mm to -45 mm (temperature, pH, and H<sub>2</sub>S sensors) and from +35 mm to -20 mm (oxygen sensors) across the sediment-water interface. The vertical resolution was 1 mm for the upper 25 mm, and 0.1 mm below. Once the profiles were completed (i.e., approximately 100 minutes), the sensors returned to their initial position, moved horizontally by approximately 1 cm and were put on hold for 30 minutes before starting the next profiling sequence. The profiling experiment lasted approximately 48 hours, providing 29 depth profiles of temperature, pH and  $H_2S$  and 87 depth profiles of O<sub>2</sub>. Then, the sensors were introduced into the sediment for a time-series recording for 24 hours. The three oxygen sensors were precisely  $(\pm 0.1 \text{ mm})$  located at 0.5, 1.1 and 1.7 mm depth below the sediment-water interface, while the temperature,  $H_2S$  and pH sensors were at  $25 \pm 1$  mm below the interface. Measurements were carried out every 5 seconds. The pH sensor was calibrated just before deployment at the in situ temperature with reference NBS standards (Metrohm Ltd.) and corrected from the *in situ* temperature shift during deployment. The H<sub>2</sub>S sensor was calibrated in the laboratory before and after each deployment using a N<sub>2</sub> bubbled pH4 buffer solution in which successive volumes of a precisely titrated Na<sub>2</sub>S stock solution were added. Oxygen sensors were calibrated using the in situ O<sub>2</sub> concentration in the pool water measured with an Aanderaa optode and the zero O<sub>2</sub> concentration of the anoxic sediment.

#### 2.4. Laboratory analyses

Water samples were analyzed for NO<sub>3</sub><sup>-</sup> (Hansen and Koroleff, 2007), NH<sub>4</sub><sup>+</sup> (Koroleff, 1976), PO<sub>4</sub><sup>3-</sup>

(Murphy and Riley, 1962) and Si<sub>d</sub> (Truesdale and Smith, 1976) using a Quattro-AXFLOW autoanalyzer. Concentration of dissolved Mn was determined by flame atomic absorption spectrometry (AAnalyst 300, Perkin Elmer). Concentration of dissolved Fe was determined by spectrophotometry (Stookey, 1970). tCO<sub>2</sub> was analyzed by flow injection analysis (FIA) as described by Hall and Aller (1992). The limits of quantification associated with these techniques are 0.2  $\mu$ M for nutrients, 0.2  $\mu$ M for Mn<sub>d</sub>, and 0.5  $\mu$ M for Fe. The relative uncertainties estimated from replicates were better than 5%.

#### 2.5. Computation of benthic exchanges in tidal pools

Benthic exchanges in the tidal pools were obtained using two methods. The diffusive oxygen flux at the sediment-water interface  $(J_{dif})$  was first calculated from the oxygen concentration gradient measured with microelectrodes during the profiling experiment. Oxygen profiles were treated, and diffusive fluxes were calculated with a modified version of the PRO<sub>2</sub>FLUX program software (Deflandre and Duchêne, 2010) using Fick's law:

$$J_{dif} = -\phi D_s \frac{\delta O_2}{\delta z} \qquad (1)$$

where  $\phi$  is the porosity, D<sub>s</sub> is the oxygen molecular diffusion coefficient corrected for sediment temperature and tortuosity (Boudreau, 1997), and  $\frac{\delta o_2}{\delta z}$  is the measured oxygen concentration gradient at the sediment-water interface.

In the second approach, flux calculation was based on temporal changes in the concentrations of chemical species in the pool water. Total fluxes at the sediment-water interface  $(J_T \text{ in } \mu \text{mol.m}^{-2}.\text{h}^{-1})$  were calculated from the slope of the linear concentration change over time ( $\propto$  in  $\mu$ mol.m<sup>-3</sup>.h<sup>-1</sup>) and the water depth (h in m) in the pool water according to:

$$J_T = \propto h \tag{2}$$

This allowed the estimation of the total flux during the period of complete disconnection of the pool from the bay water. Field observations showed that for approximately one hour after the pool water was disconnected from the bay water, water still flowed through the pool from the surrounding tidal

flat and from upstream connected tidal pools. Consequently, the pool was a closed system only when this flow ceased. Total flux calculation was thus only applied for this specific period of time. Pelagic processes that can induce chemical changes in the water column are integrated in this approach. However, given the relatively low height of water column (i.e., <12 cm), the effect of pelagic processes on the chemical composition of the pool water are expected to be negligible in comparison to the effect of benthic exchanges. Atmospheric deposition was not considered here, as no rain events occurred during the field campaigns. In addition, the influence of evaporation was shown to be negligible (see discussion below). Fluxes of  $O_2$  and t $CO_2$  at the sediment-water interface could not be assessed as exchanges at the pool water-atmosphere interface may occur.

#### 2.6. Statistical treatments

The significance level of slopes used for flux calculations in the pool water was tested. Only slopes that significantly differed from 0 (t-test, p<0.05) were considered as indicative of existing fluxes. In addition, the differences in each pair of slopes leading to fluxes calculation for a given chemical species were tested using slope homogeneity tests (after concentration data was standardized by the pool water height). The obtained p-values were therefore corrected for multiple comparison using the Holm-Bonferonni method. Two slopes with corrected p-value <0.05 were thus considered as significantly different.

The contributions of environmental factors explaining the temporal variability in (1) the pool water  $O_2$  and tCO<sub>2</sub> concentrations in each investigated pool during disconnection from the bay water, (2) processes at the water-sediment interface (diffusive  $O_2$  fluxes,  $O_2$  penetration depth and  $H_2S$  appearance depth) in the pool continuously monitored for two days in October 2013, and (3) sediment chemistry (dissolved  $O_2$  and  $H_2S$  concentrations and pH in the sediment) in the pool continuously monitored for 24 hours in October 2013, were investigated using a distance-based redundancy analysis (dbRDA) performed with the DistLM option in PERMANOVA + add-on for PRIMER (Anderson et al., 2008). Forward selection was used to build models using AIC selection criterion. Pool water  $O_2$  and tCO<sub>2</sub> concentrations and  $O_2$  concentrations at 0.5, 1.1 and 1.7 mm depth in the sediment were respectively used as response multivariate data cloud defined using Euclidean distance amongst

samples. Oxygen diffusive fluxes and penetration depth,  $H_2S$  appearance depth in the sediment and pH and  $H_2S$  concentration at 2.5 cm depth in the sediment were used separately as single response variable. Tested predictor variables included pool water temperature, PAR, water depth and sediment temperature at 2.5 cm depth in the sediment.

#### 3. Results

#### 3.1. Biogeochemical dynamics in the overlying water of the tidal pool

The biogeochemical composition of the water trapped in the pools at low tide exhibited strong temporal changes (Figures 2 and 3). Water temperature fluctuated by >10°C amplitude over tidal and diurnal cycles, ranging between 41°C in July 2010 during the day and 12°C in October 2013 during nightime (Figure 2). The salinity remained constant  $(32.5 \pm 0.3)$  when the pools were connected to the bay water. After disconnection, the salinity increased over time by up to 3.4 and 1.0 units during daytime in July 2010 and in 2013, respectively, while the salinity was steady ( $\pm 0.2$  unit) during the night. During the July 2010 campaign, when conditions for evaporation were optimum, the salinity increased by 11%. Not surprisingly, the PAR reaching the sediment surface in the pool was higher when the pool was disconnected from the bay water (i.e., when the water depth was the lowest). Oxygen concentration was relatively constant around 190-220 µM (i.e., 80-95% air saturation) when the pools were connected to the bay waters (Figure 2). Once disconnected during daytime and when the water from the intertidal flat ran off through the pool, O<sub>2</sub> concentration strongly increased up to 250-480  $\mu$ M (i.e., 120-210% air saturation). After the run-off stopped, O<sub>2</sub> concentration slightly decreased over time. Conversely, O<sub>2</sub> content decreased down to 35-50% air saturation (i.e., 80-150  $\mu$ M) during the night. The concentration of tCO<sub>2</sub> showed the opposite trend, with a strong decrease during daytime and an increase during the night while being relatively constant when pools were connected to the bay waters (Figure 3).

Dissolved inorganic nitrogen and phosphate remained at low concentrations in pool waters:  $\leq 5 \ \mu M$  for NO<sub>3</sub><sup>-</sup>,  $\leq 6 \ \mu M$  for NH<sub>4</sub><sup>+</sup> and  $\leq 0.6 \ \mu M$  for PO<sub>4</sub><sup>3-</sup> (Figure 3). NO<sub>3</sub><sup>-</sup> and NH<sub>4</sub><sup>+</sup> concentrations were even below the quantification limit in July 2010. The PO<sub>4</sub><sup>3-</sup> concentration increased over time in the pool water during the disconnection periods. NO<sub>3</sub><sup>-</sup> concentration decreased when the pools were

disconnected from the bay waters, while the concentration of  $NH_4^+$  increased with a more scattered evolution and the presence of peaks. The concentration of  $Si_d$  always decreased when the water of the intertidal flat ran off through the pools and then steadily increased with time until the bay water reinundated the pools (Figure 3). Dissolved Mn content in pool water remained low during the daytime. During the night, a marked and regular increase in  $Mn_d$  was observed in August 2013. Dissolved Fe was always below the quantification limit (<0.5  $\mu$ M; data not shown).

#### 3.2. Biogeochemical dynamic in the tidal pool sediment

The vertical and temporal changes in pH, temperature, and O<sub>2</sub> and H<sub>2</sub>S concentrations in pool sediments measured in October 2013 are presented in Figures 4 and 5. Temperature exhibited strong fluctuations in the upper 5 cm of sediment concomitant with temperature changes in the overlying water, although the amplitude of thermal shift decreased with sediment depth. As an example, temperature varied between 16 and 24°C at the interface, while it varied between 18 and 22°C at 4 cm depth (Figure 4).

When the pool was connected to the bay water,  $O_2$  concentration decreased from 80-200  $\mu$ M in the overlying water to 0  $\mu$ M at 1.7±0.3 mm (Figure 4). During daytime, when the pool was disconnected from the bay water, the  $O_2$  concentration increased in the surficial sediment to values up to 600  $\mu$ M, resulting in a significant increase in  $O_2$  penetration depth up to  $2.6 \pm 0.4$  mm. Similar trends were observed at the beginning of the time-series experiment during daytime with  $O_2$  concentrations of ~500  $\mu$ M at 0.5 mm depth and ~450  $\mu$ M at 1.1 mm depth (Figure 5). Deeper at 1.7 mm depth, the  $O_2$  concentration fluctuated between 0 and 150  $\mu$ M. During the night,  $O_2$  concentration decreased to values <150  $\mu$ M at 0.5 mm depth and close to 0  $\mu$ M deeper. Surficial sediment  $O_2$  concentration increased after the re-connection between the pool and the bay waters during both daytime and night flood tides (Figure 5).

The apparition depth of  $H_2S(Z_{H2S})$ , which refers here to a concentration  $\ge 1 \mu M$ , varied between -5 and -20 mm (Figure 4). The maximum concentration measured at -4.5 cm fluctuated between 25 and 175  $\mu M$ . The minimum  $Z_{H2S}$  and the highest  $H_2S$  concentrations were observed during the night, and the maximum  $Z_{H2S}$  and lower  $H_2S$  concentrations occurred during the daytime. In addition,  $H_2S$ 

concentration at -25 mm depth was significantly lower at high tide when the pool was connected to the bay water (Figure 5).

In pool water, pH reached 9.7 during daytime, 8.0 during the night, and was intermediate at high tide (Figure 4). The pH decreased in surface sediment, with minimum values at depths between -5 and -10 mm depth. A minimum pH value of 7.3 was measured in this layer when the pool was disconnected during the night. Deeper, pH values were between 7.6 and 8.2. The pH was approximately 8.15 during the 24-h monitoring at -25 mm depth (Figure 5).

#### 3.3. Chemical exchanges at the sediment-water interface

Diffusive  $O_2$  flux  $(J_{02})$  at the sediment-water interface exhibited strong fluctuations over time (Figure 4).  $J_{02}$  was from the overlying water to the sediment, and relatively constant when the pool was connected to the bay water (-375 ± 100 µmol.m<sup>-2</sup>.h<sup>-1</sup>).  $J_{02}$  slightly decreased (-230 ± 70 µmol.m<sup>-2</sup>.h<sup>-1</sup>) when the pool was disconnected from the bay water during the night. During daytime disconnection,  $J_{02}$  was directed from the sediment to the overlying water, reaching up to 1290 µmol.m<sup>-2</sup>.h<sup>-1</sup>. When significant, the flux of NO<sub>3</sub><sup>-</sup> calculated from the evolution of concentration in the pool waters over time, was always negative while the fluxes of NH<sub>4</sub><sup>+</sup>, PO<sub>4</sub><sup>-3</sup>. Si<sub>d</sub> and Mn<sub>d</sub> were always positive (Table 1). The calculated fluxes of NO<sub>3</sub><sup>-</sup>, NH<sub>4</sub><sup>+</sup> and PO<sub>4</sub><sup>-3</sup> did not exhibit significant differences between conditions. Fluxes of Si<sub>d</sub> were significantly different between daytime and night conditions and between seasons (Table 1). The flux of Si<sub>d</sub> in unvegetated pools exhibited a positive correlation with the average temperature in the pool water (Figure 6). For similar temperature conditions in July 2010, the flux of Si<sub>d</sub> in vegetated pools (339-426 µmol.m<sup>-2</sup>.h<sup>-1</sup>) appeared lower than in unvegetated pools (605-620 µmol.m<sup>-2</sup>.h<sup>-1</sup>). A high flux of Mn<sub>d</sub> was obtained during night conditions in August, whereas the flux was one order of magnitude lower during daytime conditions in both August and October 2013.

#### 4. Discussion

Several studies have shown the occurrence of circular pools in intertidal flats of different coastal systems (e.g., Smith and Able, 1994; Van der Laan and Wolff, 2006; Takeuchi and Tamaki, 2014,

Wilson et al., 2014). However, little is known about the role of tidal pools in the biogeochemical dynamic of coastal sediments (Jensen and Muller-Parker, 1994) because reports on the diagenetic evolution of redox species in tidal pool sediments are scarce (Lillebø et al., 2004; Murray et al., 2006). The dataset obtained from the tidal pools in the *Zostera sp.* meadows of Arcachon Bay gives a unique opportunity to better understand the biogeochemical dynamics of these small-scale ecosystems and their potential impact at a larger scale. In the following discussion, we present the environmental factors that control the transient biogeochemical functioning of coastal ecosystems, and their potential contribution to the water quality of Arcachon Bay.

#### 4.1. Dynamics of physicochemical conditions in the tidal pool water

Temperature and light irradiance greatly changed in the pools over the course of the study (Figure 2). Changes in temperature in the pool water over tidal and diurnal cycles reached an amplitude of up to 10°C, similar to the variations reported in tidal rock pools by Morris and Taylor (1983). Although with a lower amplitude, temperature changes also occurred in surface sediment. In contrast, salinity in the pools remained constant ( $\pm$  0.5 unit) when connected to the bay water and slightly increased in the pools during periods of disconnection. The maximum increase in salinity was observed during the daytime, likely as a consequence of the evaporation of pool water. Assuming that the salt or water exchanges were negligible between the water pools and the sediment or surrounding area during pool complete isolation, the relative volume of water evaporated from the pools was  $\leq$  3% for the daytime and  $\leq$  1% for the night in August and October 2013 and  $\leq$  11% in July 2010, which suggests that the consequence of evaporation on biogeochemical processes can be neglected.

#### 4.2. Transient biogeochemistry of O2 and tCO2

Changes in temperature and light irradiance in aquatic ecosystems primarily impact the photosynthetic and respiration activity of organisms, and hence the dynamic of  $O_2$  and t $CO_2$ . As expected, we observed opposite fluctuations of  $O_2$  and t $CO_2$  concentrations at diurnal scale (Figures 2 and 3). The dynamics of  $O_2$  and t $CO_2$  appeared strongly governed by both light and temperature, as between 75.0

and 94.7% of the variance in O<sub>2</sub> and tCO<sub>2</sub> concentration are explained by PAR and temperature (Table A1). The photosynthetic activity in the pool is mainly attributed to the activity of microphytobenthos. Indeed, the highest O<sub>2</sub> concentrations were always recorded in the uppermost sediment layer during the daytime (Figures 4 and 5). The  $O_2$  production in surface sediment induced  $O_2$  fluxes from the sediment to the water column (Figure 4), which is in agreement with previous field studies (de Beer et al., 2005; Jansen et al., 2009; Denis et al., 2012; Delgard et al., 2012). The control of benthic photosynthetic activity on the O<sub>2</sub> dynamic in the pool water is supported by the fact that changes in PAR and temperature significantly accounted together for 72.5 % of the temporal variation of the oxygen concentrations in the surface sediment during the time-series experiment and changes in PAR alone for 80.7 % of the temporal variations of the O<sub>2</sub> diffusive fluxes at the sediment-water interface during the profiling experiment (Table A2). During nightime, the decrease in O<sub>2</sub> concentration in the pool water (Figure 2) may be attributed to the respiration of the benthic fauna living in the pool and O<sub>2</sub> consumption by the sediment for microbial respiration and oxidation reactions. Benthic O<sub>2</sub> uptake estimated by the diffusive transport of  $O_2$  at the sediment-water interface was -230±70 µmol.m<sup>-2</sup>.h<sup>-1</sup> (n=16) during the night in October 2013. Assuming that this flux was homogeneous over the pool surface, the resulting decrease in water column  $O_2$  concentration ( $\Delta O_2$ ) due to diffusive benthic uptake is given by the following expression:

$$\Delta O_2 = \frac{JO_2 \cdot t}{h}$$

where t is the time of pool isolation (4 h),  $J_{O2}$  is the mean  $O_2$  diffusive flux, and h is the water height (12 cm). The calculated  $\Delta O_2$  value is 7.7±2.4  $\mu$ M, which is 2-10 times lower than the effective decrease in  $O_2$  measured during the night (i.e., 15-70  $\mu$ M). This suggests that  $O_2$  uptake in the pool water was mostly due to fauna activity, in agreement with *ex situ* core incubations previously carried out in the same area (Delgard et al., 2013; Delgard et al., 2016a).

In the pool sediment, the  $O_2$  concentration was shown to be transient with variations of several tens of  $\mu M$  within minutes in relation to both diurnal and tidal cycles. Two major processes may be involved in the changes in  $O_2$  concentrations and penetration depths in pool sediments. The first one is

associated with the variations in the activity of microphytobenthos in surface sediment over diurnal and tidal cycles as previously described. The production of O<sub>2</sub> in surface sediment during the daytime when pools were disconnected from the bay water allowed a deeper  $O_2$  diffusion into the sediment. This process is supported by the high proportion (63.4%) of the temporal variation of O<sub>2</sub> penetration depth in sediment that could be explained by changes in temperature and PAR together during the profiling experiment (Table A2). The second process involved in the observed changes in  $O_2$  in the pool sediments is associated with tides. Indeed, O2 concentrations increased in the surface sediment when the pool became connected to the bay water (Figure 5). The water depth during immersion also contributed significantly in explaining changes in O<sub>2</sub> concentration in surface sediment during the time-series experiment (Table A2). This coupling may be due to porewater circulation induced by waves and currents during the increasing height of the water column at rising tide (De Beer et al., 2005; Billerbeck et al., 2006; Werner et al., 2006; Jansen et al., 2009; Delgard et al., 2012), causing a higher hydrostatic pressure, which drives porewater movements through the sediments (Precht et al. 2004). This phenomenon is well known to have a significant impact in permeable sediments (Huettel and Webster 2001; Charbonnier et al., 2016) but is limited in cohesive sediments. Although hydrodynamics is quite reduced in intensity in this area of Arcachon Bay (Ganthy et al. 2013), our results indicate that advective flushing may act upon benthic biogeochemistry at a millimeter scale. Such observation could not have been possible *in situ* outside a tidal pool. The cumulated effect of PAR, temperature and water level are thus able to explain 74.0% of the total variance of oxygen concentration in surface sediment in the studied tidal pool (Table A2).

#### 4.3. Transient diagenetic processes in the tidal pool

The sulfide profiles measured in October 2013 revealed that the  $H_2S$  concentrations in sediment porewaters also exhibited strong vertical and temporal dynamics (Figure 4). Dissolved sulfide production resulted from bacterial sulfate reduction coupled to organic carbon oxidation when other oxidants (i.e.,  $O_2$ ,  $NO_3^-$ , Mn and Fe oxides) are depleted (Jørgensen, 1982). These fluctuations were in phase with the  $O_2$  concentration variations: the lowest  $H_2S$  apparition depth and the highest  $H_2S$ concentration generally corresponded to the lowest oxygen concentrations (i.e., during the night when

the pools were isolated). In addition, the water depth could significantly explain 78.7% of the temporal variation of  $H_2S$  concentrations at -2.5 cm during the time-series experiment (Figure 5, Table A2). This indicates that the  $O_2$  production and the flushing of oxygenated water in surface sediment were an efficient forcing controlling the dynamic of porewater H<sub>2</sub>S concentrations. However, we always noticed a significant gap of  $9 \pm 4$  mm between the O<sub>2</sub> penetration depth and the H<sub>2</sub>S apparition depth in sediment (Figure 4). This suggests that O<sub>2</sub> was not the primary oxidant of H<sub>2</sub>S. It is worth noting that a rapid and direct H<sub>2</sub>S oxidation by O<sub>2</sub> through cable bacteria may be involved here, as reported by Nielsen and Risgaard-Petersen (2015) and Malkin and Meysman (2015), although we could not prove that this process was occurring in the studied tidal pools. Intermediate redox processes with a rapid kinetic (within minutes time-scales) involving nitrogen, manganese or iron species may possibly act as electron transfers between O<sub>2</sub> and H<sub>2</sub>S within this depth interval of a few millimeters. Interestingly, our results show that the Mn<sub>d</sub> concentration increased in the pool water during isolation from the bay water (Figure 3), with corresponding effluxes one order of magnitude higher during nightime than during daytime (Table 1). The buildup of dissolved Mn in the pool water can be directly associated with the release of Mn from sediments due to the reductive dissolution of Mnoxyhydroxides below the thin oxic layer of the sediment. The oxic layer is enriched in Mn oxides, acting as a diffusion barrier for the dissolved Mn (Kristiansen et al., 2002; Pakhamova et al., 2007; Rigaud et al., 2013). As the oxic layer became thinner during night, porewater Mn<sub>d</sub> was produced closer to the surface sediment, facilitating its release to the pool water. This indicates that redox processes involving at least Mn species may act as an intermediate redox processes for electron transfer between O2 and H2S over diurnal timescales. The absence of measurable content of Fe in the pool waters indicates that most of the Fe diffusing up from deeper sediments was re-oxidized within the oxic layer, preventing its release and accumulation in the pool water (Sundby et al., 1986; Kristiansen et al., 2002; Pakhamova et al., 2007; Rigaud et al., 2013).

The uptake of  $NO_3^-$  and release of  $NH_4^+$  and  $PO_4^{3-}$  indicate that denitrification and OM (i.e., organic P and N) mineralization processes controlled the dynamic of those nutrients at the sediment-water

interface of tidal pools. However, no clear trend was identified over the diurnal and tidal cycles, suggesting that these ongoing redox processes occurred over longer time scale.

The Si<sub>d</sub> fluxes in unvegetated pools was shown to be positively related with average temperature (Figure 6). This may be partly attributed to the increase in molecular diffusivity of Si<sub>d</sub> with temperature. However, the Si<sub>d</sub> diffusion coefficient increases by 45% between 20 and 35°C (Boudreau, 1997), whereas we observed an increase in Si<sub>d</sub> flux by more than 600%. As the quartz solubility is between 122 and 240  $\mu$ M of H<sub>4</sub>SiO<sub>4</sub> and between 1475 and 2380  $\mu$ M for amorphous silica in the range of temperature measured in pools (Gunnarsson and Arnorsson, 2000), Si<sub>d</sub> in pool waters was far below equilibrium with respect to quartz and biogenic silica. This suggests that the Si<sub>d</sub> release in the tidal pools was likely related to the dissolution of Si-bearing particles or biogenic silica debris in surface sediment, its accumulation in porewaters and diffusion to the water column, all processes being further enhanced under higher temperature (Feuillet-Girard et al., 1997). In vegetated pools, the release of Si<sub>d</sub> appeared lower than in unvegetated pools (Table 1) suggesting Si<sub>d</sub> uptake by *Zostera noltei*. Although aquatic macrophytes contain significant amounts of biogenic silica (Schoelynck et al., 2010), only a few works have reported on the Si uptake by marine macrophytes (Herman et al., 1996; Gié et al., 2008; Querné et al., 2012).

<u>4.4. Importance of the tidal pools ecosystems in the biogeochemical functioning of Arcachon Bay</u> Environmental conditions prevailing in the tidal pools induced the release and accumulation of  $PO_4^{3-}$ ,  $NH_4^+$ ,  $Si_d$  and  $Mn_d$  during the period of isolation. Then, tidal pool waters mix with the lagoon waters at rising tide contributing to the input of chemical species into the bay. The intertidal flats of the bay have a surface area of 115 km<sup>2</sup> (Plus et al., 2009). The surface area of tidal pools represents 17 km<sup>2</sup>, assuming that pools cover 20% of the intertidal flats. Considering the range of fluxes measured in the unvegetated tidal pools (Table 1) and an average emersion period of 6 h at each tidal cycle, we calculated net fluxes up to 450 kmol.y<sup>-1</sup> for  $PO_4^{3-}$ , 12000 kmol.y<sup>-1</sup> for  $NH_4^+$ , 62000 kmol.y<sup>-1</sup> for  $Si_d$  and 4000 kmol.y<sup>-1</sup> for  $Mn_d$ . Phosphate inputs from tidal pools are of the same magnitude as the dissolved

inorganic phosphorus supplied by the continental waters that flow to the Arcachon Lagoon (Buquet et al., 2017). The flux of ammonium from tidal pools corresponds up to 18% of the average annual inputs of dissolved inorganic nitrogen from the drainage basin (Canton et al. 2012), and the tidal pool flux of Si<sub>d</sub> is less than 3% of the river flux (Buquet et al., 2017). Annual discharge of Mn<sub>d</sub> is not available in Arcachon Bay. The uptake of NO<sub>3</sub><sup>-</sup> observed in the tidal pool may account for a total NO<sub>3</sub><sup>-</sup> removal up to 5000 kmol.y<sup>-1</sup> that correspond to about 45% of the released NH<sub>4</sub><sup>+</sup>. It is worth noting that the presence of *Zostera noltei* within the pool induced an uptake of Si<sub>d</sub> that reduced their net release fluxes to the pool water by 30% (Table 1). Tidal pools were not colonized by *Zostera noltei* in 2013, the seagrass only being present at the rim of the pool. Therefore, it is most likely that the seagrass decline observed in the bay since 1989 (Plus et al., 2010) has caused an additional input of Si<sub>d</sub> from the tidal pools to the bay waters during the last decades.

#### Conclusion

This study describes the coupling between water and sediment biogeochemical dynamic in mud flat tidal pools of the Arcachon Bay. Tidal pools are dynamic natural ecosystems where the light irradiance and temperature variations are largely amplified due to the reduced size of the water body, while the effect of evaporation can be neglected. We show that the diagenetic processes are transient in response to tidal and diurnal changes of the physicochemical characteristics in the overlying water. Benthic biogeochemistry is primarily governed by the rapid (minutes time-scale) O<sub>2</sub> variations in surface sediment due to microphytobenthos activity and superficial porewater flushing. Diagenetic processes implying highly sensitive redox species such as H<sub>2</sub>S and Mn responded rapidly to these O<sub>2</sub> fluctuations with however a smoother temporal trend attributed to the delay for electron transfer within the chain of redox reactions and chemical transport in sediment. Nutrient (NO<sub>3</sub><sup>-'</sup>, NH<sub>4</sub><sup>+'</sup>, PO<sub>4</sub><sup>-3</sup><sup>-</sup>) recycling at the sediment water interface did not appear to be closely related to the rapid O<sub>2</sub> dynamic, but likely to longer time-scale cycle associated to the variation in organic matter oxidation processes. The dynamic of Si was mainly driven by the dissolution of biogenic silica, which was directly controlled by temperature. Si releases from the sediment appeared to be lowered in vegetated pools

because of its uptake by *Zostera noltei*. This study showed that tidal pools are easily accessible natural incubators that allow us to apprehend at *in situ* conditions transient biogeochemical processes that occur at the sediment water interface. Tidal pool may also contribute significantly to tidal bay budget of nutrient. Therefore, an all-inclusive budget of nutrient in the Arcachon Bay should take into account benthic fluxes in tidal pools ecosystems.

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### TABLES AND TABLES CAPTIONS

Table 1: General information and total fluxes of nutrients  $(NO_3, NH_4, PO_4^{3-} and Si_d)$  and dissolved Mn  $(Mn_d)$  in intertidal pools studied the July 1<sup>st</sup> 2010 (two vegetated and two unvegetated pools), 27-28 August (same pool, day and night) and 8-10 October (same pool, 3 consecutive days). Fluxes were calculated from concentrations change in the isolated pool water. No flux of dissolved Fe was reported as Fe concentration was always below the detection limit. The uncertainties associated to fluxes correspond to the standard error associated to the slope of the linear regression and n is the number of data point used for the linear regression. Fluxes associated with the same letter correspond to those for which no significant (adjusted p>0.05) differences in the slopes were detected using pairwise slope homogeneity test (p values adjusted using Holm-Bonferroni method)

									Fluxes $(\mu mol.m^{-2}.h^{-1})$				
Date	Pool numb er	Substrat e	Pool diame ter (m)	Po ol dep th (cm )	Condit ion	Т (° С)	Salin ity	n	NO <sub>3</sub>	NH4 <sup>+</sup>	PO4 <sup>3-</sup>	Sid	Mn <sub>d</sub>
July 2010	1	Zostera sp.	3.6	2.5	Day	38 .0 ± 2.0	32.2 ± 0.7	1 2	<lq< td=""><td><lq< td=""><td>0.53±0. 14<sup>a</sup></td><td>426±6 3<sup>ab</sup></td><td>n.d.</td></lq<></td></lq<>	<lq< td=""><td>0.53±0. 14<sup>a</sup></td><td>426±6 3<sup>ab</sup></td><td>n.d.</td></lq<>	0.53±0. 14 <sup>a</sup>	426±6 3 <sup>ab</sup>	n.d.
	3	Zostera sp.	1.4	1.0	Day	38 .1 ± 0.8	n.d.	8	<lq< td=""><td><lq< td=""><td>0.67±0. 17<sup>ab</sup></td><td>339±1 3<sup>a</sup></td><td>n.d.</td></lq<></td></lq<>	<lq< td=""><td>0.67±0. 17<sup>ab</sup></td><td>339±1 3<sup>a</sup></td><td>n.d.</td></lq<>	0.67±0. 17 <sup>ab</sup>	339±1 3 <sup>a</sup>	n.d.
	2	Unveget ated	2.5	2.5	Day	38 .0 ± 2.0	n.d.	8	<lq< td=""><td><lq< td=""><td>n.s.</td><td>605±1 05<sup>a</sup></td><td>n.d.</td></lq<></td></lq<>	<lq< td=""><td>n.s.</td><td>605±1 05<sup>a</sup></td><td>n.d.</td></lq<>	n.s.	605±1 05 <sup>a</sup>	n.d.
	4	Unveget ated	3.6	10	Day	38 .0 ± 2.0	n.d.	8	<lq< td=""><td><lq< td=""><td>n.s.</td><td>620±1 12<sup>a</sup></td><td>n.d.</td></lq<></td></lq<>	<lq< td=""><td>n.s.</td><td>620±1 12<sup>a</sup></td><td>n.d.</td></lq<>	n.s.	620±1 12 <sup>a</sup>	n.d.
Augu st 2013	5	Unveget ated	4.0	12	Day	27 .0 ± 1.0	33.1 ± 0.2	8	n.s.	119± 32 <sup>a</sup>	n.s.	182±2 5 <sup>b</sup>	4.1±1 .4 <sup>a</sup>
		40			Night	19 .3 ± 0.5	32.7 4 ± 0.04	8	- 15±2 a	81±2 9 <sup>ab</sup>	2.8±0.7	49±6 <sup>d</sup>	41±4 b
Octo ber 2013	6	Unveget ated	4.5	12	Day	21 .7 ± 0.3	31.6 ± 0.2	1 2	- 33±5 a	33±3 b	4.5±0.3	68±5 <sup>cd</sup>	n.s.
					Day	22 .9 ± 0.6	32.5 ± 0.2	1 3	n.s.	52±9 ab	3.4±0.7 abc	83±5 <sup>c</sup>	4.6±2 .3 <sup>a</sup>
					Day	20 ± 1	33.2 ± 0.3	9	53±1 5 <sup>a</sup>	n.s.	3.1±0.8 abc	61±2 <sup>cd</sup>	6.8±2 .1 <sup>a</sup>

<lq: concentrations lower than the limit of quantification / n.d.: not determinated / n.s.: not significant (p<0.05) slope detected.

SPECIAL SCRAME





Figure 1: (a) Map of the Arcachon Bay (France) showing the location of the sampling site Germanan. (b) Photo of an intertidal pool studied in the *Zostera noltei* meadow of Germanan with the MP6 miniprofiler system. (c) Example of an aerial photo of the intertidal flat studied showing the numerous tidal pools (Image @2017 Google).



Figure 2: Evolution of  $O_2$  concentrations and physicochemical properties in the intertidal pools studied (a) during daytime in one of the four pools the July 1<sup>st</sup> 2010, (b) during daytime and the following night the 27-28 August 2013 and (c) during three consecutive days the 8-10 October 2013. Note the different scales for temperature and salinity in July 2010. The three vertical dashed lines indicate the beginning of the pool isolation (left), the time when the water of the intertidal flat ceased to flow through the pool (set at 0 h), and the time of pool inundation at flood tide (right).



Figure 3. Evolution of chemical composition of pool water (a) in four pools during daytime the July 1<sup>st</sup> 2010, (b) in one pool during daytime and the following night the 27-28 August 2013 and (c) in one pool during three consecutive days the 8-10 October 2013. Note the different scales for Si<sub>d</sub> and PO<sub>4</sub><sup>3-</sup> in July 2010. The three vertical dashed lines indicate the beginning of the pool isolation (left), the time when the water of the intertidal flat ceased to flow through the pool (set at 0 h), and the time of pool inundation at flood tide (right). Note that dissolved Fe was always below the detection limit.



Figure 4: Vertical distribution of temperature,  $O_2$  and  $H_2S$  concentrations and pH in pool sediment measured by microsensors for a 48h period the 8-10 October 2013. The 2D mapping was carried out using the DIVA gridding mode of the Ocean Data View software (Schlitzer, 2014). The  $O_2$  penetration depths ( $Z_{O2}$ ),  $H_2S$  apparition depths ( $Z_{H2S}$ ) and diffusive  $O_2$  fluxes at the sediment-water interface (JO<sub>2</sub>) obtained from the depth profiles are also reported. Negative JO<sub>2</sub> values correspond to fluxes from the water to the sediment. Values reported for  $J_{O2}$  and  $Z_{O2}$  correspond to the average of 3 profiles and the error bars correspond to  $\pm 1$  standard deviation. The error bars for  $H_2S$  apparition depths correspond to  $\pm 1$  mm assumed to be the uncertainty on the interface detection on  $H_2S$  profiles. The horizontal white dashed lines correspond to the sediment-water interface. PAR and water depth are

also reported with the periods when the pool was disconnected from the bay water (dashed area) and the daytime period (light shaded areas). The absence of pH values is due to a broken electrode during the profiling sequence.



Figure 5. Evolution of  $O_2$  and  $H_2S$  concentrations, pH and temperature in the sediment measured using microsensors for a 24h period the 10-11 October 2013. Temperature and oxygen concentration in the pool water as well as light (PAR) and water depth are also reported.



Figure 6.  $Si_d$  fluxes vs. average temperature in the unvegetated intertidal pools. Fluxes were calculated from concentrations change in the pool water at low tide. The error bars associated to fluxes correspond to the uncertainty associated to the slope of the linear regression. The error bars associated to the temperature correspond to the standard deviation around the average during pool disconnection.

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### **APPENDICES**

Table A1: Distance Based Linear Model results between predictors (Temperature and Photosynthetically Active Radiation, PAR) and the concentration patterns of O<sub>2</sub> and tCO<sub>2</sub> (in a multivariate space defined using Euclidean distance) in the pool water during disconnection from the bay water. Marginal tests show the proportion of variation explained by predictor variables fitted individually. Sequential tests (using a Forward selection and AIC selection criteria) determine the variance explained by predictor variables when fitted sequentially. AIC (Akaike Information Criteria) is the criteria used for model selection, the lower the AIC, the better the model (conversely to R<sup>2</sup>, AIC is not affected by the number of fitted variables). pseudo-F indicate the statistic (using permutations) used to test the significativity of the model selected against the hypothesis of no relationship between the response data cloud and the model chosen (predictors).

Tidal pool campaign	Test	Variable	AIC	Pseudo-F	p value	Prop. of variance explained
August 2013 - Night	Marginal tests	T(°C)		106.8	0.001	94.7%
August 2013 - Day	Marginal tests	$T (^{\circ}C)$ PAR (µmol.m <sup>-2</sup> .s <sup>-1</sup> )	0	3.8 31.3	0.104 0.002	43.1% 86.2%
	Sequential tests	PAR (µmol.m <sup>-2</sup> .s <sup>-</sup> <sup>1</sup> ) + T (°C)	-6.1 -11.2	31.3 7.06	0,001 0.021	86.2% 95.0%
October 2013 - Day 1	Marginal tests	T (°C) PAR (μmol.m <sup>-2</sup> .s <sup>-1</sup> )	k	64.8 14.1	0.001 0.003	87.8% 61.1%
	Sequential tests	T (°C)	-12.6	64.8	0,001	87.8%
October 2013 - Day 2	Marginal tests	$T (^{\circ}C)$ PAR (µmol.m <sup>-2</sup> .s <sup>-1</sup> )		7.2 4.3	0.017 0.026	47.3% 35.1%
	Sequential tests	T (°C) +PAR (μmol.m <sup>-</sup> <sup>2</sup> .s <sup>-1</sup> )	3.5 -2.0	7.2 7.8	0.02 0.03	47.3% 75.0%
October 2013 - Day 3	Marginal tests	T (°C) PAR (μmol.m <sup>-2</sup> .s <sup>-1</sup> )		11.7 10.7	0.002 0.006	62.6% 60.3%
	Sequential tests	T (°C) +PAR ( $\mu$ mol.m <sup>-</sup> <sup>2</sup> .s <sup>-1</sup> )	0.3 -4.5	11.7 6.8	0.001 0.017	62.6% 82.5%

Table A2: Distance Based Linear Model results between predictors variables (abiotic drivers) and the temporal variation of sediment processes. Marginal tests show the proportion of temporal variation explained by predictor variables fitted individually. Sequential tests (using a Forward selection and AIC selection criteria) determine the variance explained by predictor variables when fitted sequentially. AIC (Akaike Information Criteria) is the criteria used for model selection, the lower the AIC, the better the model (conversely to R<sup>2</sup>, AIC is not affected by the number of fitted variables). pseudo-F indicate the statistic (using permutations) used to test the significativity of the model selected against the hypothesis of no relationship between the response data cloud and the model chosen (predictors).

Tidal pool campaign	Response variables	Test	Variable	AIC	Pseudo- F	p value	Prop. of variation explained
October 2013 – profiling experiment	Oxygen diffusive fluxes	Marginal tests	Water depth (m) T (°C) PAR (µmol.m <sup>-2</sup> .s <sup>-1</sup> )	Q	10.6 8.2 104.5	0.001 0.012 0.001	29.9% 24.7% 80.7%
		Sequential tests	PAR ( $\mu$ mol.m <sup>-2</sup> .s <sup>-1</sup> ) +Water depth (m)	76.5 69.9	104.5 9.0	0.001 0.006	80.7% 86.0%
October 2013 – profiling experiment	Oxygen penetration depth	Marginal tests	Water depth (m) T (°C) PAR (µmol.m <sup>-2</sup> .s <sup>-1</sup> )		4.9 32.2 16.6	0.036 0.001 0.002	16.3% 56.3% 39.9%
		Sequential tests	T (°C) +PAR (μmol.m <sup>-</sup> <sup>2</sup> .s <sup>-1</sup> )	-59.3 -62.1	32.2 4.6	0.001 0.047	56.3% 63.4%
October 2013 – profiling experiment	H <sub>2</sub> S appearance depth	Marginal tests	Water depth (m) T (°C) PAR ( $\mu$ mol.m <sup>-2</sup> .s <sup>-1</sup> ) O <sub>2</sub> penetration depth		2.1 7.4 5.7 1.8	0.155 0.006 0.018 0.188	7.8% 22.9% 18.5% 6.7%
		Sequential tests	T (°C) +Water depth (m) +PAR ( $\mu$ mol.m <sup>-</sup> <sup>2</sup> .s <sup>-1</sup> )	68.6 64.9 61.3	7.4 5.6 5.3	0.012 0.027 0.023	22.9% 37.5% 49.3%
October 2013 – time- series experiment	Oxygen concentration in surface sediment (-0.5, -1.1, -1.7 mm)	Marginal tests	T (°C) Water depth (m) Sediment T (°C) PAR ( $\mu$ mol.m <sup>-2</sup> .s <sup>-1</sup> )		150.5 8.2 103.0 603.5	0.001 0.005 0.001 0.001	35.4% 28.8% 27.2% 68.7%
	×	Sequential tests	PAR ( µmol.m <sup>-2</sup> .s <sup>-1</sup> ) +T (°C) +Water depth (m)	2419.4 2385.9 2372.0	603.5 37.4 16.1	0.001 0.001 0.001	68.7% 72.5% 74.0%
October 2013 –time- series experiment	H <sub>2</sub> S concentration in sediment (- 2.5 cm)	Marginal tests	T (°C) Water depth (m) Sediment T (°C) PAR ( μmol.m <sup>-2</sup> .s <sup>-1</sup> )		14.8 1013.4 9.5 28.8	0.003 0.001 0.001 0.001	50.9% 78.7% 33.3% 9.4%
		Sequential tests	Water depth (m)	616.1	1013.4	0.001	78.7%
October 2013 -time-	pH in sediment (-2.5 cm)	Marginal tests	T (°C) Water depth (m)		67.7 15.8	0.001 0.001	19.8% 5.4%

series experiment		Sediment T (°C) PAR ( µmol.m <sup>-2</sup> .s <sup>-1</sup> )		96.0 109.3	0.001 0.001	25.9% 28.4%
	Sequential tests	PAR ( µmol.m <sup>-2</sup> .s <sup>-1</sup> ) +Sediment T (°C) +T (°C) +Water depth (m)	2380.6 - 2419.5 - 2477.4 - 2479.6	109.3 43.6 65.9 4.1	0.001 0.001 0.001 0.035	28.4% 38.2% 50.3% 51.0%

r (c) depth (m) 2479.6

#### HIGHLIGHTS

- Tidal pools are natural and very dynamic small-scale ecosystems •
- They are suitable to investigate the transient dynamic of benthic biogeochemical processes •
- Temperature, light and hydrodynamic are the main driving factors
- O<sub>2</sub>, CO<sub>2</sub>, sulphide and metals fluctuate at diurnal and tidal timescales •
- Tidal pools play a significant role in the biogeochemical functioning of Arcachon Bay •

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