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Abstract:

Among nonhuman primates, the evolutionary underpinnings of variation in social structure remain debated, with both ancestral relationships and adaptation to current conditions hypothesized to play determining roles. Here we assess whether interspecific variation in higher-order aspects of female macaque (genus: *Macaca*) dominance and grooming social structure show phylogenetic signals, i.e. greater similarity among more closely-related species. We use a social network approach to describe higher-order characteristics of social structure, based on both direct interactions and secondary pathways that connect group members. We also ask whether network traits covary with each other, with species-typical social style grades, and/or with sociodemographic characteristics, specifically group size, sex-ratio, and current living condition (captive vs. free-living). We assembled 34-38 datasets of female-female dyadic aggression and allogrooming among captive and free-living macaques representing 10 species. We calculated dominance (transitivity, certainty), and grooming (centrality coefficient, Newman's modularity, clustering coefficient) network traits as aspects of social structure. Computations of K statistics and randomization tests on multiple phylogenies revealed moderate-strong phylogenetic signals in dominance traits, but moderate-weak signals in grooming traits. GLMMs showed that grooming traits did not covary with dominance traits and/or social style grade. Rather, modularity and clustering coefficient, but not centrality coefficient, were strongly predicted by group size and current living condition. Specifically, larger groups showed more modular networks with sparsely-connected clusters than smaller groups. Further, this effect was independent of variation in living condition, and/or sampling effort. In summary, our results reveal that female dominance networks were more

phylogenetically conserved across macaque species than grooming networks, which were more labile to sociodemographic factors. Such findings narrow down the processes that influence interspecific variation in two core aspects of macaque social structure. Future directions should include using phylogeographic approaches, and addressing challenges in examining the effects of socioecological factors on primate social structure.

Key words:

Social networks, phylogenetic signals, social style, group size, macaques

Introduction:

In group-living animals, diversity in sociality may be characterized by variation in social structure, i.e. the patterning and distribution of different types of interactions (e.g. competitive, affiliative, cooperative) and relationships among group members (Hinde, 1976; Kappeler & Van Schaik, 2002). Several factors have been found to govern social structure, including but not restricted to kinship, age similarity, sex and dominance status (Hinde, 1976; Kappeler & Van Schaik, 2002). However, our understanding of the evolutionary and ecological factors that produce diverse social structures remains limited. Specifically, the extent to which such social structure has been influenced by species' ancestral relationships versus variation in current conditions remains debated (Clutton-Brock & Janson, 2012; Kasper & Voelkl, 2009; Koenig et al., 2013; Thierry, 2004). Here, we use Social Network Analysis to assess whether variation in aspects of social structure across multiple species of macaques (genus: Macaca) is related to species' phylogenetic relatedness (Blomberg et al., 2003; Thierry, 2004), and hypothesized tendencies for social traits to co-evolve with each other in suites or packages (de Waal & Luttrell, 1989; Thierry, 2007). We also investigate the extent to which such covariation may be influenced by

group-specific sociodemographic characteristics (Kasper & Voelkl, 2009; Sterck et al., 1997; Sueur et al., 2011b)

A major explanatory framework for the evolution of variation in primate social structure is related to species' ancestry or phylogenetic closeness (Blomberg et al., 2003). According to this framework, aspects of sociality are expected to show phylogenetic signals, i.e. to exhibit greater similarity in social structure among more closely-related than distantly-related species (Blomberg et al., 2003). Ecological adaptation is not ruled out but rather hypothesized to occur minimally, or in response to novel, drastic environmental changes (Blomberg et al., 2003; Chan, 1996; Matsumura, 1999). In the past, proponents of phylogenetic signals often referred to them as outcomes of phylogenetic constraints or phylogenetic inertia (Blomberg et al., 2003). However, more recent theory recognizes that phylogenetic signal refers to the observed patterns of trait similarity, rather than one or more evolutionary processes – e.g. random genetic drift, stabilizing selection, pleiotropy, or low rates of evolution -- by which such patterns may arise (reviewed in Kamilar & Cooper, 2015). Rather than attempt to distinguish between possible processes or the rates at which they occur, here we simply aim to identify strong and weak phylogenetic signals in order to determine the extent to which social network structure covaries with macaques' phylogenetic closeness.

Across animal societies, behavioral traits are more labile to evolutionary change in comparison to morphological and/or physiological characteristics (Blomberg et al., 2003). Primates are no exception, with traits like brain size, body mass and canine dimorphism exhibiting stronger phylogenetic signals than those related to social organization and activity budgets (Kamilar & Cooper, 2015). As such, phylogenetic signals for aspects of

primate social network traits have been hard to identify. For instance, Kasper and Voelkl

(2009) established that several aspects of affiliative social networks – e.g. community sub-
group formation or modularity, mean centrality in the connectedness of individuals, and the
density of network connections (see Methods for precise definitions) all showed greater
variation between closely related than between distantly related species, i.e. weak
phylogenetic signals and a substantial effect of socioecological or demographic factors on
network structure. Similarly, Pasquaretta et al. (2014) found no effect of phylogenetic
signals on the efficiency of information flow through affiliative social networks of
primates, including humans. Tests for phylogenetic signals in aspects of sociality within
specific primate families and genera have also revealed inconsistencies. In the genus
Eulemur, group size and sex-ratio both strongly correlated with phylogenetic distances
(Ossi & Kamilar, 2006). Yet among both baboons (genus: Papio) and squirrel monkeys
(genus: Saimiri), interspecific variation in multiple forms of social organization seem to be
strongly linked to ecological factors like food distribution, predation pressure, and
intergroup feeding competition (Barton et al., 1996; Boinski, 1999; Henzi & Barrett, 2003).
Nevertheless, among all primate genera, phylogenetic signals in social traits have
been most consistently detected within the genus Macaca. Macaques constitute 23 species
of Old World primates, with wide-spread geographic distributions in nature (Cords, 2013).
Although all species show a predominantly similar social organization (multi-male, multi-
female social groups, female philopatry and male dispersal), they show broad inter- and
intraspecific variation in several aspects of female social structure (Thierry, 2007).

Specifically, post-conflict affiliation, rates of affiliative contact (Thierry et al., 2008), the

steepness of dominance hierarchies (Balasubramaniam et al., 2012a), and asymmetry in

aggression (Balasubramaniam et al., 2012a; Thierry et al., 2008) all show phylogenetic signals, whereas grooming kin bias shows a weak signal (Thierry et al., 2008). These findings have led some proponents of phylogenetic explanations to posit that core aspects of macague social structure may also be structurally linked, covarying with each other at the species level (Petit et al., 1997; Thierry et al., 2008). Thierry (2007) has characterized this tendency to covary by assigning macaque species to positions on a four-grade 'social style' scale that is broadly consistent with phylogenetic lineage splits (Thierry, 2007). At one extreme, 'grade-1' species (e.g. rhesus macaques (Macaca mulatta). Japanese macaques (M. fuscata)) of the Fascicularis lineage are hypothesized to show 'despotic' social structure, characterized by steep and asymmetric dominance hierarchies, low levels of post-conflict affiliation, intense affiliative kin-bias, and highly modular or sub-structured social networks in which grooming is both centralized among dominant individuals and is concentrated among more preferred partners. At the other extreme, 'grade-4' species (e.g. Tonkean macaques (M. tonkeana), crested macaques (M. nigra)) belonging to the Sulawesi lineage are hypothesized to be more 'tolerant' or 'egalitarian', showing shallower dominance hierarchies with more frequent counter-aggression from subordinates towards dominants, high proportions of post-conflict affiliation, less grooming kin bias, and denser, less centralized or modular grooming social networks (Balasubramaniam et al., 2012a; Berman & Thierry, 2010; de Waal & Luttrell, 1989; Sueur et al., 2011b; Thierry, 2007; Thierry et al., 2008). Some versions of the covariation hypothesis posit that a species that shows a single behavioral trait associated with a particular social style will display all other traits (Castles et al., 1996; Petit et al., 1997). However, this extent of covariation has not always been found. Rather, most studies (but see Thierry et al. 2008) thus far have revealed

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marked differences between species at the extreme ends of the social style scale (grade-1 and grade-4 macaques, e.g. Sueur et al., 2011b), but found inconsistencies among macaques in intermediate grades (reviewed in Balasubramaniam et al., 2012b).

In the present study, we expand on the above findings by assessing phylogenetic signals and covariation in dominance and grooming social network traits across 10 species of macaques representing all four social style grades, and groups from both captive and free-living conditions. In accordance with the predictions of socioecological models, we also examine whether sociodemographic factors, specifically group size and current living condition (Kasper & Voelkl, 2009; Pasquaretta et al., 2014), influence social network traits in addition to, or instead of, social style or phylogenetic closeness. Broadly, socioecological models differ from phylogenetic explanations by positing that social structure is relatively more labile to changes in current conditions, and that such changes are independent of phylogenetic relatedness (Koenig et al., 2013; Sterck et al., 1997; van Schaik, 1989). Specifically, some versions of the model predict that among free-living primates, larger group size should lead to the evolution of more despotic social network structures, characterized by steeper, asymmetric dominance hierarchies, and highly centralized, modular grooming networks with less dense connections/ redundant ties. This may be because larger groups are likely to display heightened levels of contest competition for resources (Koenig et al., 2013; Sterck et al., 1997; van Schaik, 1989) and/or increased temporal constraints (Berman & Thierry, 2010; Dunbar, 1992; Korstjens et al., 2010). On the other hand, smaller groups may be expected to show greater within-group social tolerance characterized by more reversals in dominance interactions, and de-centralized grooming networks that show greater density/ redundancy of ties. Such effects may also be

influenced by the spatial constraints imposed by captive housing that may result in more frequent social interactions among individuals with a potentially wider range of partners (Duboscq et al., 2013; McCowan et al., 2008).

Following the pioneering applications of social network analysis on macaque social structure by Sade (1972), comparative studies of nonhuman primate social structure have increasingly used social network approaches to compare various types of societies (Griffin & Nunn, 2012; Kasper & Voelkl, 2009; Pasquaretta et al., 2014; Sueur et al., 2011b). SNA tools improve on earlier methods by constructing aspects of higher-order group structure from a combination of both direct interactions and indirect or secondary connections among group members, and by identifying consistent patterns of interaction among sub-sets of individuals (Brent et al., 2011; Croft et al., 2008; Farine & Whitehead, 2015; Kasper & Voelkl, 2009; Krause et al., 2007; Lusseau & Newman, 2004; McCowan et al., 2008; Newman, 2004; Sueur et al., 2011a; Wey et al., 2008). Yet in comparison to applied animal behavioral research (e.g. infectious disease transmission (reviewed in Drewe & Perkins, 2015), quantifying group social stability (reviewed in McCowan et al., 2008)), SNA has been used less extensively to examine the evolutionary bases for variation in primate social structure (see Kasper & Voelkl, 2009; Griffin & Nunn, 2012; Sueur et al., 2011b).

We first ask whether aspects of female dominance and grooming networks show strong phylogenetic signals (Q1) and whether grooming networks covary with dominance networks (Q2) and/or hypothesized social style grades across macaques (Q3). In accordance with the covariation and social style hypotheses, we predict that macaque species that have the most transitive, decided and/or certain dominance relationships will also show (a) the greatest tendencies for high-ranking individuals to occupy more central positions in more modular grooming networks, and (b) networks that are more modular

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with sparser, less redundant ties. In comparison, those with the least transitive and/or uncertain dominance relationships are expected to show the opposite characteristics. Further, we predict that transitions from more despotic to more tolerant social styles will be correlated with systematic decreases in the centralization of grooming among dominants and modularity, and increases in network density or redundancy of ties. Finally, we also examine (Q4) the potential impact of sociodemographic factors on networks and patterns of covariation. Specifically, we predict that group size will be positively associated with the centralization of grooming among dominants and with grooming modularity, and negatively associated with the density/ redundancy of ties in grooming network clusters. We also explore the effects of living condition on such relationships, and the impact of a third sociodemographic characteristic -- female-to-male sex ratio -- on such covariation.

Methods:

This research adhered to the American Society of Primatologists principles for the ethical treatment of primates. We first assembled behavioral and phylogenetic datasets from various sources – published literature, our own unpublished data, and personal donations from other researchers (Table 1). Below we describe these data, the computation of social network measures, and statistical analyses.

Datasets – macaque social behaviors and phylogenetic trees:

To compute dominance network measures, we assembled a total of 38 behavioral datasets on dyadic aggressive interactions among adult female macaques (individuals ≥ 3.5 - 5 years of age, depending on species-typical ages of maturity) from groups living in captive and free-living (free-ranging and wild) conditions (Table 1). These encompassed 20 populations representing 10 macaque species from all four social style grades. Except for

stumptailed macaques (*M. arctoides*) and bonnet macaques (*M. radiata*), we acquired at least two datasets for each species; these were either from different social groups or from the same social group during different stable study-periods. To compute grooming network measures, we obtained data from 34 (out of the 38) datasets from nine species (Table 1).

The datasets had been collected using different types of sampling approaches, and varied in sampling effort per subject (Table 1). The majority (aggression: 33/38, or 87%; grooming: 25/34, or 74%) had been collected using focal-animal and/or all-occurrences sampling methods (Altmann, 1974; see citations provided in Table 1). For these datasets, all subjects within a dataset had been observed for roughly equal amounts of time (see corresponding citations; information available on request). A minority used either scan sampling (aggression: 2/38, or 5%; grooming: 7/34, or 20%) or ad-libitum sampling along with focal animal sampling (aggression: 3/38, or 8%; grooming: 2/34, or 6%). The scan sampling datasets were included based on a previous study on Japanese macaques revealing that for a given number of scans, this approach yields identical amounts of behavioral data to focal sampling for the same duration (in minutes) as the number of scans (Romano et al., 2016). Datasets that used ad libitum sampling were included after ensuring that they had been (a) used in previous comparative studies (Balasubramaniam et al., 2012a; Schino & Aureli, 2008), and (b) conducted for overall durations that were comparable to those of the focal- or all-occurrences sampled datasets (6 - 12 months).

All datasets included had been collected as part of long-term studies, and during periods when the groups were socially stable in terms of group composition, size, dominance relationships, age-class and/or sex ratios. Out of 38 datasets, the majority (27, or 71%) was from free-living groups of macaques. We found significant differences in group

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0.03

size (captive = 28.4 ± 19.1 ; free-living = 48.3 ± 23.9 ; ANOVA: $F_{I, 32} = 6.04$, $p = 0.02$) and
sex ratios (captive = 0.50 ± 0.21 ; free-living = 0.33 ± 0.09 ; ANOVA: $F_{I, 32} = 12.07$, $p <$
0.01) between captive vs. free-living macaques. Hence, we either conducted separate
assessments for only free-living groups (Q1), or directly examined the effect of living
condition in our analyses (Q2 - Q4).

In order to assess and control for the effects of phylogenetic signals, we extracted a Bayesian consensus tree (Supplementary Figure 1) and a tree-block of 10 trees from 10Ktrees, an online resource for primate phylogeny based on mitochondrial and nuclear genetic datasets (Arnold et al., 2010). All 11 phylogenies showed proportional branchlengths, and identical topologies that were also consistent with previously constructed macaque topologies using parametric approaches (e.g. Chatterjee et al., 2009; Tosi et al., 2003). We hence report the results of tests performed using the consensus phylogeny, with those from the tree-block submitted as supplementary material (Supplementary Table 4). *SNA – Dominance and Grooming Measures:*

Table 2 provides the definitions and details of the dominance and grooming network traits that we computed. We computed dominance network measures from winner-loser dominance matrices in which non-diagonal cells contained data on the frequencies of dyadic contact and non-contact aggression from animals in the rows to animals in the columns. Acts of bi-directional or counter-aggression were scored twice, once on either side of the diagonal for each interactant. Bidirectional and counter-aggression were included to produce representative measures of dominance network traits, particularly for grade-3 and grade-4 macaque species that typically experience high proportions of such interactions (Balasubramaniam et al., 2012a; Thierry et al., 2008). From each dominance

matrix, we computed two traits – triangle transitivity (Shizuka & Mcdonald, 2012) and dominance certainty (Fujii et al., 2013; Fushing et al., 2011) – that have not been used in previous comparative studies of primate social networks (see Table 2 for definitions). Whereas most methods for describing group level dominance hierarchies consider only dvadic wins and losses from agonistic encounters (e.g., dominance gradient (hierarchical steepness: de Vries et al., 2006), and dominance asymmetry (Directional Consistency Index: de Vries, 1998)), the computation of network-based measures, particularly dominance certainty, uses both direct dominance interactions and indirect pathways of imputed wins and losses (Fujii et al., 2013; Fushing et al., 2011). Thus, they capture more global aspects of a dominance hierarchy than steepness or DCI, and are less susceptible than steepness to error due to the numbers of non-interacting dyads present in the dataset. Specifically, dominance certainty incorporates the presence of non-interacting dyads in its calculation (Table 2) by weighting both the total number of direct wins by an individual 'A' against an individual 'D' (A→D), as well as imputed wins incurred via the likelihood of traversing longer paths (e.g. $A \rightarrow B \rightarrow C \rightarrow D$) through the dominance network. In doing so, the certainty measure characterizes the network-level flow-structure of dominance information, thereby accounting for hidden elements in dominance structure such as

We constructed grooming networks (Figure 1) from weighted, undirected networks, i.e. where individuals represented by *nodes* were connected by *edges* that represent the normalized values of grooming given and received for each pair of nodes. Weighted networks were used to account for the potential impact of differences in observation times across comparative datasets (Griffin & Nunn, 2012; Newman, 2004). Specifically, we

avoidance between individuals (Fujii et al., 2013; Fushing et al., 2011) (Table 2).

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calculated three measures – centrality coefficient, Newman's modularity, and global clustering coefficient -- that were all (a) not collinear (see below), (b) captured biologically different aspects of network structure, and (c) mathematically independent of the number of nodes (or group size: Nunn et al., 2015). First, we calculated *centrality coefficient* as the correlation coefficient between individuals' dominance statuses (computed using normalized David's scores computed from dominance matrices: Gammell et al., 2003) see below) and their centralities within grooming networks (as in Sueur et al., 2011b; see Table 2 for definitions). For the latter, we used standardized eigenvector centrality, i.e. the number and strength of an individuals' direct and indirect grooming connections indicating each individual's access to social capital or support (Farine & Whitehead, 2015; Newman, 2004; Whitehead, 2008). Eigenvector centrality is among the most biologically meaningful of the centrality measures; it is more indicative of individuals' social status than measures of betweenness and closeness centrality in taxa such as primates that have more complete network graphs (Brent et al., 2010; Farine & Whitehead, 2015; Kasper & Voelkl, 2009; McCowan et al., 2008; Sueur et al., 2011a). We also calculated Newman's eigenvectorbased modularity, which is a global measure of network sub-structuring into communities (Newman, 2006; Nunn et al., 2015; see Table 2 for a detailed definition). Finally, as a measure of the density/ redundancy of ties in the network, we calculated the global clustering coefficient, which is an unweighted measure of how well-connected nodes are to their immediate neighbors (Table 2). Clustering coefficient captures the underlying redundancy of node connectivity at a more local level than alternative measures like network density, making it mathematically independent of the number of nodes (Table 2; Nunn et al., 2015). However, since increased localized clustering may also be a

characteristic of more modular networks (Nunn et al., 2015), we confirmed that clustering coefficient was positively correlated with network density measures across our datasets (n = 34, r = 0.91, p < 0.01). Grooming measures were calculated using the *statnet* (for eigenvector centrality) and *igraph* (for Newman's modularity and global clustering coefficient) R packages (Csardi & Nepusz, 2006; Handcock et al., 2006). Individuals' dominance David's scores were calculated from the aggression matrices using the *steepness* R package (de Vries et al., 2006; Leiva & de Vries, 2014).

Supplementary Table 1 shows the values of the various network measures computed for each dataset. To avoid potentially pseudoreplicating our results owing to network measures being collinear to each-other, we confirmed that collinearity diagnostics were met for both dominance measures (Pearson's r correlation = 0.61; variance inflation factors (VIF) < 2), and grooming measures (r < 0.78 for all pairs; generalized variance inflation factors (GVIF) < 3) using the *car* package in R (Fox & Weisberg, 2011). Given the susceptibility of network measures to variation in sampling effort (Croft et al., 2008; Croft et al., 2011; Farine & Whitehead, 2015; Lusseau et al., 2008), we performed diagnostic tests to ascertain that these did not impact our analyses. For each grooming dataset, we generated 1000 permuted networks using a bootstrapping algorithm (Lusseau et al., 2008), by randomly sampling increasingly smaller subsets (100%, 80%, 60%, 40%, and 20%) of edges from the original network. Plots of percent sampling effort and mean grooming network measure computed across 200 permuted networks under each category of sampling effort revealed asymptotic relationships (Supplementary Figure 2). Further, Wilcoxon ranksum tests revealed that network measures computed from the original data were not significantly different to those computed from the bootstrapped networks that used smaller

subsets of data (80% - 40% sampling effort; but see clustering coefficient at 20% sampling effort: Supplementary Table 2). Finally, we confirmed that across datasets, sampling effort calculated as the mean observation times spent per subject (Table 1) was unrelated to group-level dominance and/or grooming network measures (Supplementary Table 3; Supplementary Figure 3).

Statistical Analyses:

To examine evidence for phylogenetic signals (Q1) in dominance and grooming network traits at the species level, we used the Phytools R package (Revell, 2012). For each trait, we computed Blomberg's Kappa (K) coefficient (Blomberg et al., 2003) as the ratio between the Mean-Squared Errors (or MSE) of the raw, species-means of behavioral traits over the MSEs of traits that have been phylogenetically transformed using tree-specific branch-length information. The K coefficient therefore indicates the strength of the phylogenetic signal and further, is scaled by a similar ratio of MSEs calculated assuming a Brownian motion model of evolution where trait change is hypothesized to be directly proportional to the shared evolutionary histories, or branch length distances between taxa (Blomberg et al., 2003; Kamilar & Cooper, 2015). We considered K > 1 from the randomization test as indicative of a "strong" signal that exceeds the amount of signal expected under a Brownian motion model of evolution (Blomberg et al., 2003). Values of 0.8 < K < 1 were considered to be indicative of a "moderate" signal, while values that are << 0.8 indicate a weak signal, i.e. greater plasticity due to adaptation to current conditions in a trait (Blomberg et al., 2003). These values were based on previous comparative studies of nonhuman primate behavioral traits that also examined phylogenetic signals among smaller numbers of species (Balasubramaniam et al., 2012a; Thierry et al., 2008). We

avoided splitting terminal species' nodes into multiple nodes to represent groups within species. Although doing so offers the potential advantage of increased statistical power, it would also artificially inflate phylogenetic signals (Blomberg et al., 2003; Thierry et al., 2008). Blomberg's K generates identical results to Pagel's Lambda (Pagel, 1997, 1999), an alternative indicator of phylogenetic signal. However, we chose to report K since this statistic is more useful in informing about trait variation relative to Brownian evolution (Kamilar & Cooper, 2015). In addition, we conducted randomization tests to compare the MSE of phylogenetically transformed data to those computed from 1000 randomized datasets generated by scattering the behavioral scores randomly on a phylogeny. In addition to handling the intrinsic dependency of the data, such tests indicate whether any observed phylogenetic signal is greater than that expected by chance. Although they had low statistical power of 20-35% (Blomberg et al., 2003), randomization tests have nonetheless been used in previous studies of similarly small sample sizes (7-10 species: Balasubramaniam et al., 2012a; Thierry et al., 2008).

To examine whether grooming network traits were linked to dominance traits (Q2) or social style scale (Q3) across species, and to assess the potential impact of sociodemographic factors (Q4) on such covariations, we constructed General Linear Mixed-effects Models (GLMMs: Bates et al., 2016). We implemented an Information-Theoretical approach (Burnham et al., 2011; Grueber et al., 2011) using the *lme4* and *MuMIn* R packages. Given the relatively small number of species, we used neither Phylogenetic Least Squares regression (PGLS: Nunn, 2011), nor permutation-based MCMC models (Croft et al., 2011; Farine & Whitehead, 2015) that are recommended for examining variation in phylogenetic distances and network sampling effort respectively.

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Instead, we indirectly tested for the effects of phylogeny by assessing covariation with social style grade, since social style and phylogeny have been shown to strongly covary across macaques (Thierry et al., 2000). To rule out the effect of network sampling effort on our results, we replicated our GLMMs using mean network measures computed from permuted datasets of varying degrees of sampling effort (20% - 80% of edges: see above).

We first generated 3 global models, one for each grooming network trait treated as the outcome variable. While eigenvector coefficient and Newman's modularity were both normally distributed (modularity: Shapiro-wilcoxon test: w = 0.95, p = 0.13), clustering coefficient was arcsine-transformed to meet normality assumptions (w = 0.98, p = 0.65). As recommended by I-T approaches (Burnham et al., 2011; Grueber et al., 2011), our global models were constructed by the inclusion of just those covariates that catered to testing our specific predictions (Q2-Q4). Specifically, dominance traits (transitivity and dominance certainty), social style category, and sociodemographic factors (group size, sex ratio, and living condition category) were included as fixed effects. Since the effect of group size on grooming networks could be more pronounced among free-living groups (Griffin & Nunn, 2012; Sueur et al., 2011b), we also examined a potential interaction between group size and living condition. Species was set as a random effect to avoid pseudoreplication issues. We next used the arm package in R to z standardize ($\mu = 0$, SD =1) the predictors in the global model, which is essential for interpreting parameter estimates after model averaging (Gelman et al., 2009). We then used the automated 'dredge' function in the MuMIn R package to generate a 'complete' set of 80 models from multiple possible combinations of predictors from the global models (Grueber et al., 2011) (Supplementary Tables 4-6). This included a 'null' or an intercept-only model with just species as a random

effect for the sake of comparison. From each full model set, we identified candidate models that included only models whose difference in AICc scores were < 4 from the model with the lowest AICc score (Burnham et al., 2011) (models in bold font in Supplementary Tables 4-6). We confirmed that model diagnostics criteria were met for each candidate model, and ruled out multicollinearity issues for models with multiple predictors (r < 0.8 for all pairs of predictors; GVIF < 4 for each: the *car* package in R: Fox & Weisberg, 2011). We then determined parameter estimates by model-averaging each set of candidate models using the zero-averaging approach (Burnham & Anderson, 2002; Grueber et al., 2011). Since this approach weakens the effect sizes of predictors that only appear in a minority of models, it is more useful (than the natural averaging method) in cases where the aim is to determine which (among multiple) factors have the strongest effect on a response variable (Grueber et al., 2011; Nakagawa & Freckleton, 2010). For all parameter interpretations, we set the alpha level at 0.05.

Results:

(Q1) Tests for phylogenetic signals:

Mean species-level scores of both dominance network traits -- transitivity and dominance certainty -- showed moderate phylogenetic signals (Table 3). When we repeated the analyses including only free-living groups (27 datasets across 7 species), the results showed a strong phylogenetic signal for dominance traits (K statistics >>1: Table 3). In comparison, grooming traits -- especially eigenvector and clustering coefficient -- showed weak phylogenetic signals (Table 3). Although Newman's modularity showed a moderate signal as indicated by the K statistic, randomization tests neither reached significance nor showed a non-significant trend for this trait (Table 3). Given the relatively small number of

species, our tests of phylogenetic signals lacked power (~ 20-35%). Yet, results from the		
randomization significance tests were mostly consistent with our estimations of the strength		
of the signals from the K statistic (Table 3). Moreover, they were also robust to		
phylogenetic branch length variation, with results from the consensus phylogeny being		
largely consistent with those across 10 phylogenetic trees extracted from the 10KTrees		
website (Supplementary Table 3).		
(Q2-4) The effect of dominance traits, social style, and sociodemographic factors on		
grooming traits:		

Table 4 shows the model-averaged coefficients and parameter estimates from candidate GLMM sets shortlisted using our dAICc criterion, from the complete model sets for grooming Newman's modularity and clustering coefficient. Our criterion shortlisted 8 models for modularity, and 7 for clustering coefficient. Neither candidate model set included the 'random effect only' model, which had a dAICc > 12 from the candidate sets (Supplementary Table 6, 7). Model averaged coefficients revealed that group size strongly influenced modularity and clustering coefficient in predicted directions. Larger groups showed more modular, but less dense clusters or redundant ties than smaller groups, which showed the opposite characteristics (Table 4; Figure 2). Further, group size was both present, and the most important predictor in all candidate models for both modularity and clustering coefficient (Table 4). We also detected a significant effect of living condition, with captive groups showing less modular networks and greater density/ redundancy of cluster ties than free-living groups (Table 4; Figure 2). However, there was no interaction between group size and living condition (Table 4), suggesting that the effect of group size were independent of living condition. Sex ratio also showed no effect. Finally, neither

dominance traits nor social style grade strongly influenced these grooming network traits in predicted directions (Table 4). In fact, social style did not make the candidate models for modularity.

In contrast to Newman's modularity and clustering coefficient, candidate model selection for grooming eigenvector centrality coefficient failed to eliminate the 'random effect only' model (Supplementary Table 5). Zero-based model averaging revealed that none of the predictors had a strong effect on these indices (Supplementary Table 8).

Finally, replications of our GLMMs using network measures computed from permuted datasets revealed that our findings, particularly for group size, were highly consistent across varying degrees of sampling effort. As per findings from the complete data, group size showed positive associations with modularity, and negative associations with clustering coefficient for all degrees of sampling effort (e.g. at 60% and 20% effort: Supplementary Tables 9, 10). On the other hand, the effect of living condition on grooming clustering coefficient weakened at lower sampling effort (Supplementary Tables 9, 10).

Discussion:

In group-living animals, the evolutionary origins of variation in aspects of social structure remain much debated. Here we used a social network approach to examine the impact of phylogenetic relatedness, specifically phylogenetic signals and trait covariation, as well as sociodemographic factors like group size and living condition on interspecific variation in higher-order aspects of dominance and grooming social structure across macaques. We found that while dominance social network traits showed strong phylogenetic signals, grooming network traits showed weak signals and were not strongly covariant with social style or with dominance traits. Rather, two aspects of grooming

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network structure – modularity and clustering coefficient – were strongly influenced by group size independently of current living condition. Below we discuss our findings in depth, and offer avenues for future comparative assessments of primate social network structure.

Previous studies among macaques have established strong phylogenetic signals in some aspects of social structure (e.g. dominance steepness and counter-aggression: Balasubramaniam et al., 2012a), but moderate-to-weak signals in others (e.g. kin-bias: Thierry et al., 2008). Consistent with this pattern, we found moderate to strong phylogenetic signals for aspects of female dominance network traits, but moderate or weak signals for grooming networks. Specifically, both transitivity and dominance certainty exhibited degrees of signal that are close in strength to what is expected under a Brownian motion model of evolution (Blomberg et al., 2003). In other words, the covariance between dominance trait values of adjacent taxa strongly proportional to their shared evolutionary histories indicated by their phylogenetic branch length distances (Kamilar & Cooper, 2015). This finding extends earlier findings of strong phylogenetic signals on group-level aspects of dominance structure that were based on dyadic interactions (e.g., hierarchical steepness, counter-aggression: Balasubramaniam et al., 2012a) to broader patterns of triadic motifs (transitivity) and network-wide flow patterns (dominance certainty). It also extends it to indirect connections that reflect the higher-order structure of the flow of dominance information through pathways that connect both interacting and non-interacting individuals (dominance certainty: Fujii et al., 2013; Fushing et al., 2011). Phylogenetic signals were even stronger when we analyzed only free-living groups of macaques, despite a reduction in sample size from 10 to 7 species (Blomberg et al., 2003). This may be due to the

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differences in living conditions between groups of closely related species. In captivity, spatial constraints and/or management strategies may force animals to interact that might otherwise avoid one another after one or two decided outcomes (Duboscq et al., 2013; McCowan et al., 2008). This may result in greater than expected degrees of reversals in agonistic success and hence, greater uncertainty in dominance relationships. Unfortunately, limited datasets prevented an analysis of phylogenetic signals among captive groups alone (11 out of 38 datasets: 5 species). Nevertheless, the detection of strong phylogenetic signals among free-living macaques (27 out of 38 datasets: 7 species) builds on previous studies that have either included both captive and free living groups (Balasubramaniam et al., 2012a) or only captive groups (Thierry et al., 2008). Since previous comparative studies across primates have shown a general lack of correspondence between phylogeny and socioecology (Koenig et al., 2013; Ménard, 2004; Okamoto & Matsumura, 2002), it is unlikely that the observed phylogenetic signals in dominance traits are 'artifacts' of correlations between phylogeny and current socioecological factors. Further, the signals were detected despite the fact that a fraction of the free-living groups in our dataset were exposed to food provisioning by humans (Table 1), although predation pressure, another socioecological factor (van Schaik, 1989), was anecdotally either uniformly low or absent across the study populations. Taken together, our findings suggest that phylogeny predicts a major component of the interspecific variance in dominance structure among free-living macaques. Nevertheless, these findings do not entirely rule out an adaptive component. For instance, the concept of social reaction norms proposes that aspects of social structure may change dynamically by responding to socioecological and environmental factors, but within the limits posed by a species- or phylogenetic lineage-typical range of responses that may

in turn be linked to phylogenetic closeness (Berman & Thierry, 2010). Moreover, phylogenetic signals, rather than being linked to ancestral genetic splits, may be outcomes of adaptations by ancestral species dispersing into historically similar environments (Kamilar & Cooper, 2015). We suggest that future assessments of the bases of phylogenetic signals in social network traits focus on phylogeographic approaches (Lemey et al., 2009; Ree & Smith, 2008). Specifically, a comparison of the reconstructed social networks and geographic ranges of ancestral primate taxa may provide better insights into whether phylogenetic signals in primate social networks are indeed 'artifacts' of adaptive responses in their evolutionary past.

In comparison to dominance traits, grooming networks, particularly eigenvector and clustering coefficients, showed moderate (modularity) or weak (eigenvector and clustering coefficient) phylogenetic signals. Further, grooming traits did not covary with dominance traits in the directions predicted. A possible explanation is that covariation between social network traits is not always linear (but see Thierry et al.'s (2008) findings on covariation between traits associated with conciliatory tendencies across macaques). Further, it may be discernible at levels of organization other than species, for instance, across phylogenetic lineages (Thierry, 2000). Yet the lack of a strong effect of species-typical social style on grooming networks may be seen as evidence to refute this argument given that social style and phylogeny are correlated (Thierry et al., 2000). A more likely explanation is that grooming traits may come under the additional influence of current socioecological and/or sociodemographic factors.

Among all the primate genera, macaques are presently unique in the extent to which interspecific variation in social traits is consistent with phylogeny, and covariant with social

style (Balasubramaniam et al., 2012a; Sueur et al., 2011b; Thierry et al., 2008; Thierry et al., 2000). Yet we found that group size had a stronger influence on grooming modularity and clustering coefficient than either dominance traits or social styles. Specifically, larger groups of macaques showed more modular grooming networks, as well as less density/

redundancy in connections among localized, triadic clusters.

It is conceivable that the effect of group size on grooming traits may have been an artifact of intergroup differences in (1) living condition (McCowan et al., 2008; Duboscq et al., 2013) and/or (2) chance interactions (Griffin & Nunn, 2012; Nunn et al., 2015). In our dataset, for instance, captive groups were both significantly smaller (see Methods) and, as expected due to spatial constraints, showed less modular grooming networks with more densely connected clusters (see Results) than free-living groups. Yet we found no effect of an interaction between group size and living condition on grooming traits, suggesting that the effect of group size on grooming traits was independent of the effect of living condition. Moreover, the permuted datasets revealed that both the grooming traits themselves, and the strong relationships between group size and grooming traits, were robust to variation in sampling effort. Together, these findings suggest that the effect of group size on grooming traits, rather than being artefactual, may more likely have socioecological underpinnings. Specifically, one school of thought is that living in larger groups may heighten levels of within-group contest competition (or WGC) for resources (Koenig et al., 2013; Sterck et al., 1997; van Schaik, 1989). This may lead to despotic, nepotistic primate societies characterized by both steep and transitive dominance structure (Majolo et al., 2009), and more modular, sparsely connected grooming community structures (the social bottleneck hypothesis: Griffin & Nunn, 2012). In comparison, smaller groups may show weaker WGC

and the opposite characteristics that are indicative of greater social tolerance. An alternative perspective is that individuals in large groups may face spatial or time constraints on their abilities to interact consistently with all other group members (Berman & Thierry, 2010; Dunbar, 1992; Korstjens et al., 2010; Puga & Sueur, 2017). This may in turn result in modular or clustered but sparser social networks in larger groups, in which individuals may come into contact with each other less frequently (Griffin & Nunn, 2012). Given the lack of a clear association between group size and dominance networks in our dataset (e.g. Group size vs. dominance certainty: n = 38, r = 0.02, p = 0.88), our findings point to a spatiotemporal rather than a resource-competition based explanation. Yet final conclusions await the expansion of our dataset to include additional groups and species, and comparable operational measures of resource distribution (see below).

Unlike grooming modularity and clustering coefficient, grooming centrality coefficient was not influenced by dominance traits, social style, or sociodemographic factors. One reason for this lack of concordance could be due to the way this index was computed. For instance, the exclusion of males from our grooming networks, unlike in Sueur et al.'s (2011b) study, may have masked skew in grooming centrality towards dominants. In despotic macaques, high-ranking males may be among the most attractive grooming partners for females who may gain benefits such as access to social support and/or resources (Schino, 2001; Silk et al., 2003; Sueur et al., 2011b). It is possible that female dominance status could be related to centralization in the grooming network that is based on *specific types* of grooming connections rather than their overall connectedness. These could be grooming connections with non-kin, grooming that is directed up the hierarchy in exchange for access to rank-related benefits such as resource tolerance and

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support in conflicts, and/or grooming that is reciprocated in the short or long-term (reviewed in Henzi & Barrett, 1999). Should comparative data be available, such assessments may generate centralization indices that better reflect female social structure.

Alternatively, it is conceivable that intergroup variation in centrality coefficients may be influenced by additional socioecological variables (Griffin & Nunn, 2012; Sterck et al., 1997), and/or intraspecific variation (Borgeaud et al., 2016; Romano et al., 2016). Yet assessments of the socioecological bases of variation in sociality have proved to be complicated. For instance, we currently lack a consistent operational measure of resource abundance and distribution that may be used in comparative contexts across a wide range of taxa (Clutton-Brock & Janson, 2012). Another challenge is assessing the effects of infectious disease risk on social networks. For instance, increased parasite infection risk may lead to the evolution of more modular networks because individuals engage fewer social partners (Griffin & Nunn, 2012; Nunn et al., 2015). At the same time, possessing more partners can also reduce infection risk via socially buffering individuals against susceptibility to infectious agents (Balasubramaniam et al., 2016; Duboscq et al., 2016; Young et al., 2014). Third, the spatial distribution of females may also influence male reproductive tactics, i.e. the extent to which males show reproductive skew by monopolizing mating opportunities which may in turn influence female social networks (Schülke & Ostner, 2008, 2013). More definitive conclusions await the ability to draw interspecies links between reproductive skew and female social networks, and indeed malemale social relationships. Finally, in comparison to interspecific variation, few comparative studies have assessed the evolutionary origins of inter-individual or intergroup variation (Clutton-Brock & Janson, 2012). In addition to impacting group-level outcomes, selection

pressures may act on the spatial orientation, behavioral strategies, and personalities of
individuals, any or all of which may in turn affect social networks (Borgeaud et al., 2016;
Capitanio, 1999; Krause et al., 2010; Lusseau & Newman, 2004; Puga & Sueur, 2017; Sih
et al., 2004). Although beyond the scope of this study, adopting similar comparative,
evolutionary approaches to assess the influence of intraspecific variation in spatial and
social strategies on social network structure may be a logical next step.
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900 Table 1: Macaque datasets analyzed in the study

Species (Macaca)	Social Style	Study Location	Source or Citation	Group Size	Adult Females	Sampling Effort ^b	Living Condition ^c
fuscata ^a	1	Yakushima, Japan	Ventura et al. (2006)	25	8	25 (NA)	Free-living
fuscata ^a	1	Yakushima, Japan	Ventura et al. (2006)	50	21	7 (NA)	Free-living
Fuscata	1	Koshima, Japan	Duboscq et al. (2016); Romano et al. (2016)	45	21	10	Free-living
Fuscata	1	Koshima, Japan	Duboscq et al. (2016); Romano et al. (2016)	45	20	5	Free-living
Fuscata	1	Koshima, Japan	Duboscq et al. (2016); Romano et al. (2016)	45	20	16	Free-living
Fuscata	1	Rome Zoo, Italy	Schino et al. (2007)	57	23	291 (280)	Captive
Fuscata	1	Rome Zoo, Italy	Schino et al. (2007)	57	23	229 (224)	Captive
Fuscata	1	Paris Zoo, France	Petit et al. (1997)	22-24	9	180 (8)	Captive
mulatta	1	University of Cambridge, Madingley, UK	Desportes & Thierry (unpub)	16	7	72 (8)	Captive
mulatta	1	Cayo Santiago, Puerto Rico	Balasubramaniam et al. (2014)	135	30	18	Free-living d
mulatta	1	Cayo Santiago, Puerto Rico	Balasubramaniam et al. (2014)	33	10	16	Free-living d
mulatta ^a	1	Yerkes Primate Center, USA	de Waal & Luttrell (1989)	51-62	13	675 (NA)	Captive
assamensis	2	Tukreshwari Temple, Assam, India	Cooper & Bernstein (2008)	28	14	-	Free-living d
assamensis	2	Tham Pla Temple, Thailand	Ogawa (unpub)	47	14	9	Free-living d
assamensis	2	Shivapuri Nagarjun National Park, Nepal	Ogawa & Koirala (unpub)	25	8	10	Free-living
fascicularis	2	Primatological Station, Kassel University, Germany	Butovskaya et al. (1995)	9	7	47 (3)	Captive
fascicularis	2	Primatological Station, Kassel University, Germany	Butovskaya et al. (1995)	13	9	50 (3)	Captive
thibetana	2	Mt. Huangshan, China	Balasubramaniam et al. (2011)	39	8	47	Free-living d

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thibetana	2	Mt. Huangshan, China	Balasubramaniam et al. (2011)	42	9	47	Free-living ^d
thibetana	2	Mt. Huangshan, China	Balasubramaniam et al. (2011)	42	9	38	Free-living d
thibetana	2	Mt. Huangshan, China	Balasubramaniam et al. (2011)	52	10	11	Free-living d
thibetana	2	Mt. Huangshan, China	Balasubramaniam et al. (2011)	49	10	12	Free-living d
thibetana	2	Mt. Huangshan, China	Balasubramaniam et al. (2011)	37	10	20	Free-living d
arctoides ^a	3	Yerkes Primate Center, USA	de Waal & Luttrell (1989)	14	12	225 (NA)	Captive
radiata	3	Mysore, India	Cooper et al. (2007)	41-49	13	-	Free-living d
sylvanus	3	Middle Atlas Mountains, Morocco	McFarland & Majolo (2011)	19	8	32	Free-living
sylvanus	3	Middle Atlas Mountains, Morocco	McFarland & Majolo (2011)	29	10	10	Free-living
sylvanus	3	Middle Atlas Mountains, Morocco	Molesti & Majolo (2013)	18	7	- (12)	Free-living
sylvanus	3	Rocamadour, France	Sosa (2014)	55	24	14	Free-living d
sylvanus	3	Rocamadour, France	Sosa (2014)	55	24	8	Free-living d
sylvanus	3	Rocamadour, France	Sosa (2014)	55	24	12	Free-living d
nigra	4	Tangkoko Reserve, Indonesia	Duboscq et al. (2013)	50	18	24	Free-living
nigra	4	Tangkoko Reserve, Indonesia	Duboscq et al. (2013)	80	21	45	Free-living
nigra	4	Tangkoko Reserve, Indonesia	Duboscq et al. (2013)	80	21	43	Free-living
nigra	4	Tangkoko Reserve, Indonesia	Duboscq et al. (2013)	80	24	22	Free-living
tonkeana	4	Primatology Center, Strasbourg, France	Thierry (1985); Demaria & Thierry (2001)	16	9	104 (8)	Captive
tonkeana	4	Orangerie Zoo, Strasbourg, France	De Marco et al. (2010)	21	7	110 (5)	Captive
tonkeana	4	Primatology Center, Strasbourg, France	Thierry (1985); Demaria & Thierry (2001)	29	11	87 (6)	Captive

901 Groups excluded from computation and analyses of grooming network metrics on account of either (a) non-availability of data, 902r (b) too few (< 7) individuals

⁹⁰³ Calculated as the mean observation time per subject in hours, for datasets collected using focal animal (mean focal observation 904 ime), all-occurrences (total observation time of the group), and/or scan sampling (number of scans per subject times the

⁹⁰⁵ Juration of the scan (derived from Romano et al. (2016)). Blank entries represent datasets in which ad-libitum sampling was

⁹⁰⁶ sed. Entries in parentheses represent sampling effort for grooming datasets, where different from aggression

⁹⁰⁷Criteria for designation of group's living-condition: Free-living: within their natural geographic distribution, or an unfenced 908rea outside of their natural geographic distribution; Captive: housed within a fenced enclosure

⁹⁰⁹Free-living groups that were regularly food-provisioned by humans

^{91@}odes for Social Style (Thierry, 2007): (1) Highly Despotic, (2) Mildly Despotic, (3) Mildly Tolerant, (4) Highly Tolerant

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911 Table 2: Definitions of Social Network Traits

<u>Trait</u>	<u>Description</u>	Data Source	Code Source
Dominance		Source	
Triangle transitivity	A transitive triangle is 3 individuals connected in the following manner: a wins its encounters with b , b with c , and a with c (Snijders, 2012). Group-level triangle transitivity is number of such "triangles" divided by all possible triangles within a group	Aggression network matrix	Perc R package (Fujii et al. 2015)
Dominance certainty	Based on a tally of all directed network paths between <i>a</i> and <i>b</i> , dyadic dominance certainty is the degree of consistency in direction of all dominance paths. It reflects the probability that <i>a</i> will win against <i>b</i> . Group-level dominance certainty is the mean of the row-wise averages of just the win-probabilites for each individual	Aggression network matrix	Perc R package (Fujii et al. 2015)
Grooming			
Centrality Coefficient	The Pearson's R correlation coefficient between the eigenvector centrality scores and David's scores of all the individuals in the group. This metric reveals the extent to which dominant individuals are more central and have extended secondary grooming connections compared to subordinates	Groom network string	Statnet R package (Handcock et al., 2006)
Newman's Modularity	The extent to which a network can be divided into communities of individuals which interact more among themselves (than expected by chance), than with members from another community. Higher values (approaching 1) indicate greater division into distinct sub-groups than lower values (closer to 0), which indicate little or no sub-divisioning	Groom network string	Igraph R package (Csardi & Nepusz, 2006)
Global clustering coefficient	Similar in calculation to triangle transitivity, but with edge direction not considered. The proportion of triplets (i.e. sets of 3 individuals) that are completely connected to each other (or form a closed triangle) relative to the total number of <i>potential</i> triplets.	Groom network string	Igraph R package (Csardi & Nepusz, 2006)

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Table 3: Results of tests for phylogenetic signals among macaque dominance and grooming social network traits. K: Blomberg's Kappa coefficient; P: significance from Randomization tests (Blomberg et al. 2003)

Behavioral Metric	All Datasets (10 species)		Free-living Groups (7 species)						
	K	P	K	P					
<u>Dominance Metrics</u>									
Transitivity	0.892	0.081(*)	1.704	0.022*					
Dominance Certainty	0.893	0.078(*)	1.634	0.023*					
	Grooming N	etwork Metrics							
Eigenvector Coefficient	0.654	0.205	0.722	0.238					
Newman's Modularity	0.872	0.110	0.959	0.110					
Clustering coefficient	0.406	0.567	0.836	0.165					

Bold font: K values (> 1) indicate strong phylogenetic signals

Phylogenetic tree used was the Bayesian MCMC consensus tree from the 10ktrees website (Arnold et al. 2010)

^{*} P < 0.05; (*) 0.05 < P < 0.1

Table 4: Summary of zero-averaged coefficients from the candidate models for Newman's modularity and clustering coefficient of grooming networks.

/ 4 \ 3.7	1 3 5	1.1.1: //	1.1	111		92
(A) New	man's Mo	odularity (8	s candida	te models)		93 93
	<u>B</u>	<u>Adj SE</u>	<u>z</u>	Pr(> z)	<u>Impor</u>	ta 93
T	0.10		5.00	0.01 44		93 93
Intercept	0.19	0.03	5.82	<0.01**		93
Living Condition (Captive vs. Free living)	0.15	0.04	4.16	<0.01**	1	93 93 93
Sex Ratio (proportion of females)	0.04	0.05	0.86	0.39	0.57	93 94
Group Size	0.09	0.03	3.27	<0.01**	1	94 94 94
Dominance Certainty	0.00	0.02	0.21	0.84	0.15	94 94
Transitivity	0.00	0.01	0.15	0.88	0.13	94 94
Group Size:	0.00	0.03	0.08	0.94	0.13	94 94
Living Condition						95 95
(B) Clus	tering Co	efficient (7	candida	te models)		95
', '			<u></u>			95 95
Intercept	0.92	0.14	6.71	<0.01**		95
Living Condition (Captive vs. Free living)	-0.44	0.20	2.25	0.02*	0.93	95 95 95
Dominance Certainty	0.34	0.19	1.75	0.08	0.83	95 96
Group Size	-0.21	0.09	2.43	0.02*	1	96 96
Group Size: Living Condition	0.03	0.12	0.26	0.79	0.14	96 96 96
Social Style (Grade 1 vs. Grade 2)	0.02	0.11	0.18	0.85	0.12	96 96 96
Social Style (Grade 1 vs. Grade 3)	0.06	0.20	0.31	0.76		96 97 97
Social Style (Grade 1 vs. Grade 4)	0.09	0.28	0.34	0.73		97 97 97
Sex Ratio (proportion of females)	0.01	0.04	0.14	0.89	0.09	97 97 97
Γransitivity	0.00	0.03	0.02	0.98	0.08	97 97 98

^a Effect sizes are standardized, comparable values (Gelman et al., 2009)

Data are from 34 groups of 9 macaque species

^{*} $p \le 0.05$ ** p < 0.01

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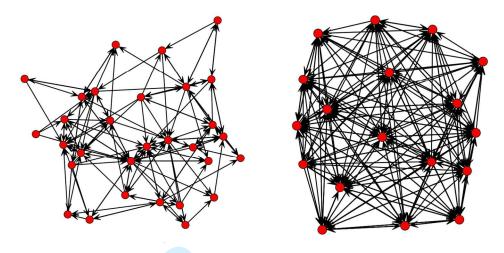


Figure 1: Grooming social networks from a (A) rhesus macaque (M. mulatta)

(Balasubramaniam et al. 2014; n = 29 individuals), and a (B) Sulawesi crested macaque (M.

nigra) (Duboscq et al. 2013; n = 19 individuals) group. Circles represent individual adult

females, and lines connecting them represent edges (unweighted) of grooming relationships

990 between females

. . .

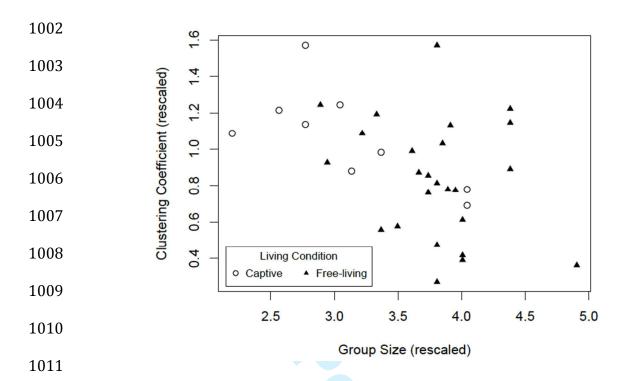
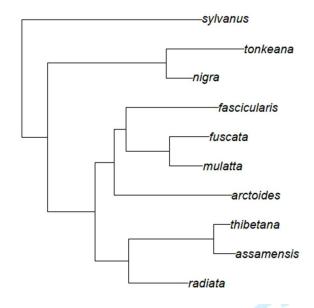


Figure 2: Influence of group size on grooming clustering coefficient across 34 datasets of captive and free-living macaque groups representing 9 species

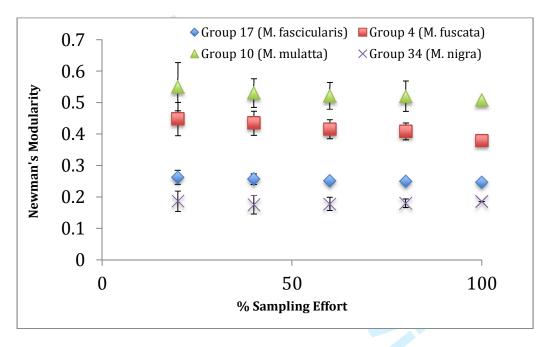
- We assessed the bases for variation in macaque social network structure
- Dominance traits showed phylogenetic signals, but grooming traits were linked to group size
- Social networks are influenced by both ancestral and sociodemographic factors

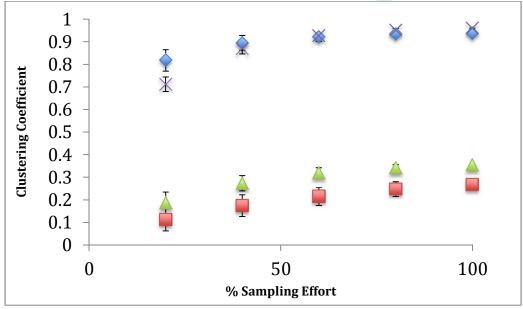


Supplementary Figure 1: Consensus tree of phylogenetic relationships among 10 species of macaques extracted from the 10ktrees website (Arnold et al., 2010), on which tests of phylogenetic signals were conducted

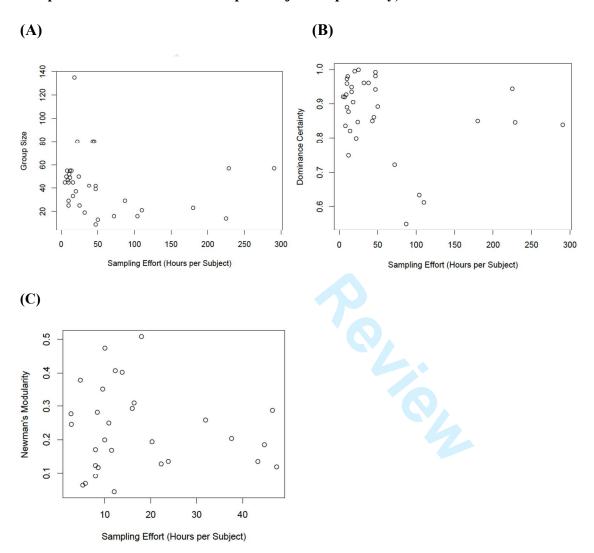


Supplementary Figure 2: Plots of grooming network measures against % sampling effort. Data-points represent means computed from values across 200 networks permuted from a natural dataset, and error bars represent standard deviations. Data-points at 100% sampling effort represent the value of the original network measure. The four groups selected were largest in size (e.g. Group 10: 135 individuals), or had the lowest observation times (e.g. Group 4: 5 hours/ subject)





Supplementary Figure 3: Scatterplots between sampling effort and (A) group size, (B) dominance certainty, and (C) grooming Newman's modularity across the datasets. Datasets that used *Ad libitum* sampling (B: 3/38; C: 2/34), and/or were outliers (B: 1/38 dataset sampled for 675 hours per subject; C: 2/34 datasets sampled for 224 and 280 hours per subject respectively) have been omitted



Supplementary Table 1: Measures of dominance and grooming network traits, computed for each macaque dataset

			ninance easures	<u>Gr</u>	ires	
Dataset	<u>Species</u>	Transit -ivity	Dominance Certainty	Centrality Coefficient	Newman's Modularity	Clustering Coefficient
1	M. fuscata	1.000	0.999	NA	NA	NA
2	M. fuscata	1.000	0.920	NA	NA	NA
3	M. fuscata	1.000	0.973	0.533	0.352	0.273
4	M. fuscata	1.000	0.921	0.429	0.379	0.732
5	M. fuscata	1.000	0.949	0.522	0.309	0.544
6	M. fuscata	0.983	0.839	0.649	0.202	0.297
7	M. fuscata	0.975	0.846	0.528	0.202	0.362
8	M. fuscata	1.000	0.850	-0.440	0.123	0.229
9	M. mulatta	1.000	0.992	NA	NA	NA
10	M. mulatta	0.714	0.722	0.618	0.170	0.093
11	M. mulatta	0.957	0.905	0.393	0.508	0.646
12	M. mulatta	1.000	0.935	0.493	0.293	0.455
13	M. assamensis	1.000	0.955	0.341	0.182	0.071
14	M. assamensis	0.972	0.927	0.449	0.117	0.142
15	M. assamensis	1.000	0.959	0.071	0.199	0.115
16	M. fascicularis	1.000	0.981	0.033	0.277	0.115
17	M. fascicularis	0.988	0.892	-0.303	0.246	0.063
18	M. thibetana	1.000	0.942	0.849	0.287	0.234
19	M. thibetana	1.000	0.992	0.532	0.119	0.245
20	M. thibetana	1.000	0.961	-0.091	0.204	0.309
21	M. thibetana	1.000	0.980	0.676	0.250	0.3
22	M. thibetana	1.000	0.877	-0.038	0.169	0.297
23	M. thibetana	1.000	0.995	0.675	0.194	0.164
24	M. arctoides	0.976	0.944	NA	NA	NA
25	M. radiata	0.984	0.962	-0.281	0.086	0
26	M. sylvanus	1.000	0.961	0.581	0.258	0.2
27	M. sylvanus	0.917	0.890	-0.644	0.474	0.471
28	M. Sylvanus	1.000	0.979	0.825	0.043	0.053
29	M. sylvanus	0.881	0.821	0.350	0.402	0.425

30	M. sylvanus	0.935	0.836	0.385	0.282	0.619
31	M. sylvanus	0.902	0.749	0.308	0.407	0.595
32	M. nigra	0.958	0.847	-0.468	0.136	0.095
33	M. nigra	0.906	0.861	0.027	0.185	0.06
34	M. nigra	0.898	0.850	0.065	0.135	0.089
35	M. nigra	0.941	0.799	0.055	0.128	0.222
36	M. tonkeana	1.000	0.633	-0.075	0.091	0
37	M. tonkeana	1.000	0.612	-0.477	0.063	0.053
38	M. tonkeana	0.750	0.550	0.370	0.068	0.168

NA: Grooming network measures unavailable or not calculated owing to dataset restrictions (see Table 1)

Supplementary Table 2: Wilcoxon rank-sum tests comparing grooming network measures -- (A) modularity and (B) clustering coefficient – between the original dataset (n=34), and permuted datasets with reduced sampling effort (80% - 20% of the total number of edges). Cells contain W coefficients and p values within parentheses

(A) Modularity

			% Sam	pling Effort		
		Original Dataset	80	60	40	20
Ħ	Original	X				
££C	Dataset					
ng E	80	560 (0.83)	X			
Sampling Effort	60	552 (0.76)	565 (0.88)	X		
Sar	40	527 (0.54)	531 (0.57)	539 (0.64)	X	
%	20	458 (0.14)	463 (0.16)	472 (0.20)	498 (0.33)	X

(B) Clustering Coefficient

% Sar	40 20	718 (0.02) 924 (<0.01*)	731 (0.06) 893 (<0.01*)	683 (0.20) 848 (<0.01*)	x 774 (0.02)	Х
ildu	60	688 (0.18)	644 (0.42)	X		
ng E	80	625 (0.57)	Х			
Sampling Effort	Original Dataset	х				
		Original Dataset	80	60	40	20
			% Samp	oling Effort		
			% Samp	oling Effort		

^{*}p (Bonferroni-corrected alpha = 0.005) < 0.005

Supplementary Table 3: Correlations between mean sampling effort (mean observation hours per subject) and group size, dominance network measures, and grooming network measures

	1				
			$\underline{N^1}$	Pearson's R	$\underline{p} \ (\underline{p}_{crit} = 0.02)$
Sampling Effort	Group	size	34	-0.17	0.32

Sampling Effort vs. Dominance Network Measures

Sampling Effort Group size 34 -0.17 0.32

Sampling Effort Dominance certainty 34 -0.27 0.12

Sampling Effort Transitivity 34 0.00 0.99

Sampling Effort vs. Grooming Network Measures

		$\underline{N^1}$	Pearson's R	$\underline{p \ (p_{crit} = 0.01)}$
Sampling Effort	Group Size	30	0.36	0.06
Sampling Effort	Eigenvector coefficient	30	0.18	0.35
Sampling Effort	Newman's Modularity	30	-0.10	0.59
Sampling Effort	Sparseness Coefficient	30	-0.13	0.50

Datasets that used *Ad libitum* sampling (dominance measures: 3/38; grooming measures: 2/34), and/or were outliers (dominance measures: 1/38 dataset sampled for 675 hours per subject; grooming measures: 2/34 datasets sampled for 224 and 280 hours per subject respectively) were omitted from the analyses.

 $p_{\text{crit}}.$ critical p value set after implementing a Bonferroni correction for multiple comparisons

Supplementary Table 4: Results of tests for phylogenetic signals among macaque dominance and grooming social Network traits, across a tree block of 10 phylogenetic trees extracted from the 10ktrees website (Arnold et al., 2010). K: Blomberg's Kappa coefficient; P: significance from Randomization tests (Blomberg et al. 2003)

Behavioral Metric	All Datasets (1	0 species)	Free-living (7 spec	
	K (m±sd)	P (m)	K (m±sd)	P (m)
	<u>Domin</u>	ance Metric	<u>es</u>	
Transitivity	0.84 ± 0.07	0.10	1.64 ± 0.06	0.03*
Dominance Certainty	0.90 ± 0.08	0.10	$\boldsymbol{1.58 \pm 0.07}$	0.02*
	Grooming	Network M	etrics	
Eigenvector Coefficient	0.66 ± 0.05	0.21	0.73 ± 0.05	0.23
Newman's Modularity	0.85 ± 0.03	0.10	0.94 ± 0.06	0.19
Sparseness Coefficient	0.39 ± 0.03	0.58	0.82 ± 0.05	0.16

Bold font: K values (> 1) indicate strong phylogenetic signals

^{*} P < 0.05

Supplementary Table 5: Model parameters and coefficients for all 80 sub-models within the full model set derived using the dredge function for the outcome variable groom eigenvector coefficient. Species was set as a random effect. Models in bold represent the candidate set (dAICc < 4 from the best-fit) selected for model averaging and interpretation.

Model	(Int)				B Co	efficien	<u>ts</u>		<u>df</u>	AICc	<u>A</u>	<u>w</u>
		LC	SS	DC	SR	GS	T	GS:				
		LC	33	DC	SK	GS	1	LC				
1^	0.21								3	40.50	0.00	0.11
2	0.23			0.22					4	40.60	0.10	0.11
3	0.22			0.38			-0.28		5	41.00	0.49	0.09
4	0.22	0.18							4	41.80	1.33	0.06
5	0.41		+						6	42.00	1.51	0.05
6	0.21					0.10			4	42.60	2.07	0.04
7	0.21						-0.08		4	42.90	2.37	0.03
8	0.22				-0.06				4	42.90	2.42	0.03
9	0.23			0.21		0.09			5	43.10	2.62	0.03
10	0.23			0.21	-0.05				5	43.30	2.80	0.03
11	0.30	-0.05				0.13		-0.81	6	43.40	2.85	0.03
12	0.45	0.20	+						7	43.40	2.92	0.03
13	0.22	0.07		0.17					5	43.50	2.97	0.03
14	0.21			0.37		0.08	-0.27		6	43.80	3.27	0.02
15	0.41		+				-0.15		7	44.00	3.47	0.02
16	0.21	0.22					-0.13		5	44.10	3.55	0.02
17	0.21	0.04		0.36			-0.28		6	44.10	3.56	0.02
18	0.21			0.38	-0.01		-0.28		6	44.10	3.58	0.02
19	0.40		+			0.13			7	44.30	3.77	0.02
20	0.42		+		-0.12				7	44.30	3.83	0.02
21	0.22	0.15				0.05			5	44.60	4.04	0.02
22	0.46	0.25	+				-0.20		8	44.70	4.17	0.01
23	0.21	0.20			0.04				5	44.70	4.19	0.01
24	0.21					0.10	-0.08		5	45.20	4.65	0.01
25	0.41		+	0.06					7	45.40	4.84	0.01
26	0.21				0.01	0.11			5	45.50	4.94	0.01
27	0.30	-0.15		0.16		0.16		-0.76	7	45.60	5.05	0.01
28	0.21				-0.05		-0.08		5	45.60	5.05	0.01
29	0.30	0.00				0.11	-0.15	-0.86	7	45.70	5.21	0.01
30	0.30	-0.20		0.35		0.15	-0.29	-0.82	8	45.80	5.29	0.01

	0.20	0.00			0.04	0.04		0.00		46.00		0.01
31	0.30	0.00		0.10	0.24	0.24		-0.98	7	46.20	5.65	0.01
32	0.22	0.01		0.19		0.09			6	46.30	5.74	0.01
33	0.47	0.30	+	-0.17	0.01	0.10			8	46.30	5.80	0.01
34	0.22			0.20	0.01	0.10	0.15		6	46.30	5.83	0.01
35	0.39		+		0.10	0.13	-0.15		8	46.40	5.93	0.01
36	0.42	0.06	+	0.15	-0.12		-0.15		8	46.50	6.00	0.01
37	0.22	0.06		0.17	-0.02	0.04			6	46.60	6.05	0.01
38	0.44	0.17	+			0.04			8	46.80	6.31	0.01
39	0.45	0.18	+		-0.03				8	46.90	6.34	0.01
40	0.21	0.20				0.03	-0.13		6	47.00	6.49	0
41	0.20	0.27			0.09		-0.15		6	47.00	6.50	0
42	0.21	-0.01		0.38		0.08	-0.28		7	47.10	6.56	0
43	0.20			0.39	0.07	0.11	-0.29		7	47.20	6.72	0
44	0.21	0.06		0.35	0.02		-0.28		7	47.40	6.92	0
45	0.41		+		-0.07	0.09			8	47.50	6.97	0
46	0.21	0.18			0.08	0.08			6	47.60	7.08	0
47	0.40		+	0.00		0.13			8	47.70	7.22	0
48	0.31	0.09			0.32	0.24	-0.20	-1.09	8	47.80	7.25	0
49	0.42		+	0.02	-0.11				8	47.80	7.27	0
50	0.40		+	0.26			-0.25		8	47.90	7.37	0
51	0.45	0.01	+			0.11		-0.61	9	48.00	7.51	0
52	0.20				0.03	0.11	-0.08		6	48.20	7.73	0
53	0.45	0.24	+			0.01	-0.20		9	48.40	7.91	0
54	0.46	0.25	+		0.01		-0.20		9	48.40	7.91	0
55	0.46	0.25	+	-0.01			-0.20		9	48.40	7.91	0
56	0.47	0.07	+			0.09	-0.24	-0.73	10	48.70	8.14	0
57	0.30	-0.07		0.09	0.23	0.25		-0.94	8	49.40	8.89	0
58	0.30	-0.08		0.32	0.30	0.27	-0.32	-1.02	9	49.50	8.94	0
59	0.41		+	0.17	-0.09		-0.22		9	49.60	9.05	0
60	0.22	0.03		0.18	0.04	0.10			7	49.70	9.21	0
61	0.40		+		-0.07	0.09	-0.15		9	49.90	9.40	0
62	0.47	0.28	+	-0.17		0.03			9	50.00	9.51	0
63	0.47	0.30	+	-0.17	0.00				9	50.10	9.54	0
64	0.20	0.25			0.12	0.07	-0.15		7	50.10	9.59	0
65	0.44	0.16	+		-0.02	0.03			9	50.50	10.03	0
66	0.20	0.03		0.38	0.09	0.11	-0.29		8	50.80	10.26	0
67	0.39		+	0.21		0.09	-0.24		9	51.00	10.49	0
68	0.41		+	0.00	-0.07	0.09			9	51.20	10.71	0
69	0.48	0.12	+	-0.18		0.10		-0.63	10	51.30	10.83	0
70	0.45	0.25	+		0.01	0.01	-0.20		10	52.50	11.97	0
71	0.45	0.25	+	0.00		0.01	-0.20		10	52.50	11.98	0
72	0.46	0.26	+	-0.01	0.01		-0.20		10	52.50	11.98	0

73	0.47	0.06	+	0.01		0.09	-0.25	-0.73	11	53.10	12.57	0
74	0.47	0.28	+	-0.17	0.01	0.03			10	54.10	13.57	0
75	0.49	0.05	+		0.26	0.22		-0.95	10	55.10	14.55	0
76	0.50	0.15	+		0.28	0.21	-0.26	-1.04	11	55.10	14.59	0
77	0.45	0.25	+	-0.01	0.01	0.01	-0.20		11	56.90	16.41	0
78	0.50	0.14	+	-0.14	0.23	0.20		-0.92	11	58.20	17.64	0
79	0.36		+	0.40	0.15	0.14	-0.30		10	58.80	18.27	0
80	0.47	0.02	+	0.31	0.38	0.28	-0.34	-1.12	12	62.20	21.66	0

^{^ &#}x27;null' model with just the intercept and random effect term

Random term: 1|Species

Int: Intercept; DC: Dominance Certainty; SR: Sex Ratio; GS: Group Size; LC: Living Condition; SS: Social Style; T:

Transitivity

Supplementary Table 6: Model parameters and coefficients for all 80 sub-models within the full model set derived using the dredge function for the outcome variable groom Newman's Modularity. Species was set as a random effect. Models in bold represent the candidate set (8 models: dAICc < 4 from the best-fit) selected for model averaging and interpretation.

Model	(Int)				B Co	efficien	<u>ts</u>		df	AICc	<u>A</u>	W
		LC	SS	DC	SR	GS	— Т	GS:			_	_
		LC	SS	De	SIC	GB	1	LC				
1	0.19	0.15			0.07	0.10			6	-72.40	0.00	0.29
2	0.20	0.14				0.08			5	-71.70	0.65	0.21
3	0.19	0.16		-0.03	0.07	0.10			7	-69.60	2.77	0.07
4	0.19	0.15			0.07	0.10	-0.01		7	-69.30	3.06	0.06
5	0.19	0.15			0.07	0.10		0.00	7	-69.10	3.30	0.06
6	0.20	0.16		-0.03		0.08			6	-69.10	3.31	0.06
7	0.19	0.15				0.08		0.04	6	-68.90	3.43	0.05
8	0.20	0.15				0.08	-0.01		6	-68.90	3.46	0.05
9	0.29	0.15	+		0.07	0.10			9	-66.40	6.00	0.02
10	0.19	0.16		-0.03	0.07	0.10	0.00		8	-66.00	6.35	0.01
11	0.19	0.16		-0.03	0.07	0.10		-0.01	8	-66.00	6.35	0.01
12	0.19	0.16		-0.02		0.07		0.03	7	-66.00	6.42	0.01
13	0.30	0.14	+			0.08			8	-65.90	6.49	0.01
14	0.19	0.15				0.08	-0.01	0.03	7	-65.80	6.54	0.01
15	0.20	0.16		-0.02		0.08	0.00		7	-65.80	6.57	0.01
16	0.19	0.15			0.07	0.10	-0.01	0.00	8	-65.70	6.65	0.01
17	0.30	0.18	+	-0.06	0.07	0.09			10	-65.50	6.84	0.01
18	0.30	0.17	+	-0.05		0.07			9	-64.20	8.20	0.01
19	0.19	0.17							4	-63.90	8.48	0
20	0.29	0.15	+			0.07		0.05	9	-62.70	9.64	0
21	0.29	0.15	+		0.07	0.10	-0.01		10	-62.50	9.90	0
22	0.19	0.16		-0.02		0.07	0.00	0.03	8	-62.40	9.98	0
23	0.19	0.21		-0.05					5	-62.30	10.06	0
24	0.30	0.15	+			0.08	-0.01		9	-62.30	10.09	0
25	0.29	0.15	+		0.07	0.10		0.01	10	-62.20	10.15	0
26	0.19	0.16		-0.03	0.07	0.10	0.00	-0.01	9	-62.10	10.26	0
27	0.32	0.21	+	-0.09					8	-61.90	10.43	0
28	0.19	0.18					-0.01		5	-61.40	10.99	0
29	0.30	0.19	+	-0.08	0.07	0.09	0.01		11	-61.40	11.00	0
30	0.19	0.17			0.00				5	-6 1.10	11.31	0

_													
=	31	0.31	0.17	+						7	-61.00	11.34	0
	32	0.30	0.18	+	-0.06	0.07	0.09		0.00	11	-60.90	11.48	0
	33	0.30	0.18	+	-0.05		0.07		0.04	10	-60.60	11.81	0
	34	0.32	0.23	+	-0.10	0.03				9	-60.30	12.08	0
	35	0.31	0.17	+	-0.06		0.07	0.01		10	-60.20	12.17	0
	36	0.19			0.07		0.11			5	-60.20	12.21	0
	37	0.19					0.12			4	-59.80	12.57	0
	38	0.19	0.21		-0.05	0.01				6	-59.30	13.05	0
	39	0.19	0.20		-0.05			0.00		6	-59.30	13.09	0
	40	0.29	0.15	+			0.07	-0.01	0.04	10	-58.70	13.68	0
	41	0.32	0.22	+	-0.11			0.02		9	-58.60	13.79	0
	42	0.19			0.07	0.05	0.13			6	-58.40	13.99	0
	43	0.19	0.18			0.00		-0.01		6	-58.30	14.02	0
	44	0.29	0.15	+		0.07	0.10	-0.01	0.01	11	-57.90	14.50	0
	45	0.19				0.05	0.13			5	-57.90	14.51	0
	46	0.31	0.18	+				-0.01		8	-57.80	14.55	0
	47	0.19					0.11	0.02		5	-57.70	14.63	0
	48	0.32	0.17	+		0.01				8	-57.70	14.71	0
	49	0.27		+			0.11			7	-57.60	14.78	0
	50	0.19			0.08		0.11	-0.01		6	-57.20	15.19	0
	51	0.33	0.23	+	-0.12	0.03		0.02		10	-56.40	15.92	0
	52	0.30	0.18	+	-0.06		0.07	0.01	0.04	11	-56.30	16.10	0
	53	0.30	0.19	+	-0.08	0.07	0.09	0.01	0.00	12	-56.20	16.13	0
	54	0.19	0.21		-0.06	0.01		0.00		7	-56.00	16.33	0
	55	0.19				0.05	0.13	0.03		6	-55.70	16.66	0
	56	0.19			0.08	0.05	0.13	-0.01		7	-55.10	17.23	0
	57	0.31	0.18	+		0.02		-0.01		9	-54.20	18.15	0
	58	0.27		+			0.11	0.02		8	-54.10	18.27	0
	59	0.27		+	0.06		0.11			8	-53.40	19.00	0
	60	0.27		+		0.04	0.13			8	-53.30	19.05	0
	61	0.27		+	0.08	0.06	0.13			9	-49.90	22.50	0
	62	0.27		+		0.04	0.12	0.02		9	-49.70	22.65	0
	63	0.27		+	0.06		0.11	0.00		9	-49.50	22.82	0
	64^	0.19								3	-49.20	23.12	0
	65	0.19			0.07					4	-49.00	23.33	0
	66	0.19				-0.05				4	-47.70	24.7	0
	67	0.19						0.03		4	-47.30	25.03	0
	68	0.19			0.07	-0.04				5	-47.10	25.25	0
	69	0.29		+						6	-47.10	25.29	0
	70	0.19			0.08			0.00		5	-46.20	26.12	0
	71	0.29		+		-0.05				7	-46.20	26.2	0
	72	0.27		+	0.08	0.06	0.13	0.00		10	-45.70	26.7	0

73	0.19			-0.05	0.03	5	-45.50	26.89	0
74	0.29	+	0.07			7	-44.70	27.63	0
75	0.19		0.08	-0.05	0.00	6	-44.10	28.26	0
76	0.29	+			0.03	7	-44.00	28.33	0
77	0.29	+		-0.05	0.03	8	-42.30	30.07	0
78	0.29	+	0.07	-0.04		8	-42.10	30.26	0
79	0.29	+	0.07		0.00	8	-41.20	31.18	0
80	0.29	+	0.06	-0.04	0.00	9	-38.20	34.13	0

^{^ &#}x27;null' model with just the intercept and random effect term

Random term: 1|Species

Int: Intercept; DC: Dominance Certainty; SR: Sex Ratio; GS: Group Size; LC: Living Condition; SS: Social Style; T:

Transitivity

Supplementary Table 7: Model parameters and coefficients for all 80 submodels within the full model set derived using the dredge function for the outcome variable groom clustering coefficient. Species was set as a random effect. Models in bold represent the candidate set (7 models: dAICc < 4 from the best-fit) selected for model averaging and interpretation.

Mod	(Int)				D.Co	efficient	~		4t	AICo	<u> </u>	***
Mod el	(Int)				<u> </u>	emcient	<u>s</u>		<u>df</u>	<u>AICc</u>	Δ	$\underline{\mathbf{W}}$
<u> </u>		LC	SS	DC	SR	GS	T	GS:				
								LC				
1	0.97	-0.49		0.37		-0.20			6	7.50	0.00	0.28
2	0.94	-0.49		0.41		-0.22		0.23	7	9.70	2.17	0.10
3	0.62	-0.54	+	0.46		-0.19			9	10.00	2.44	0.08
4	0.97	-0.51		0.39	0.07	-0.18			7	10.50	2.95	0.06
5	0.97	-0.49		0.36		-0.20	0.01		7	10.70	3.18	0.06
6	0.97	-0.21				-0.26			5	10.90	3.40	0.05
7	0.97					-0.31			4	11.10	3.61	0.05
8	0.97	-0.63		0.46					5	11.70	4.16	0.04
9	0.97	-0.29				-0.26	0.11		6	12.10	4.56	0.03
10	0.57	-0.67	+	0.55					8	12.20	4.66	0.03
11	0.96	-0.63		0.50	0.19				6	12.70	5.15	0.02
12	0.94	-0.50		0.42	0.05	-0.21		0.22	8	13.00	5.49	0.02
13	0.94	-0.49		0.40		-0.22	0.01	0.23	8	13.10	5.61	0.02
14	0.60	-0.54	+	0.49		-0.21		0.23	10	13.40	5.90	0.02
15	0.97					-0.31	0.04		5	13.70	6.16	0.01
16	0.97			0.04		-0.31			5	13.80	6.27	0.01
17	0.62	-0.54	+	0.48		-0.19	-0.02		10	13.80	6.28	0.01
18	0.96	-0.20				-0.27		0.07	6	13.80	6.29	0.01
19	0.97	-0.22			-0.03	-0.27			6	13.90	6.32	0.01
20	0.97	-0.50		0.38	0.06	-0.19	0.00		8	13.90	6.39	0.01
21	0.97				0.00	-0.31			5	13.90	6.40	0.01
22	0.62	-0.55	+	0.47	0.07	-0.17			10	14.50	6.99	0.01
23	0.97	-0.63		0.48			-0.02		6	14.60	7.08	0.01
24	0.96	-0.27				-0.27	0.11	0.11	7	15.10	7.55	0.01
25	0.97	-0.29			-0.01	-0.26	0.11		7	15.30	7.74	0.01
26	0.56	-0.67	+	0.59			-0.04		9	15.60	8.03	0.01
27	0.96	-0.63		0.51	0.19		-0.02		7	15.80	8.29	0
28	0.57	-0.67	+	0.56	0.18				9	16.20	8.71	0
29	0.73		+			-0.31			7	16.50	8.98	0

•				0.00		0.01			_	46.50	0.4.4	_
30	0.97			0.02		-0.31	0.03		6	16.70	9.14	0
31	0.97				0.00	-0.31	0.04		6	16.70	9.15	0
32	0.97	-0.32							4	16.70	9.17	0
33	0.94	-0.50		0.41	0.04	-0.21	0.01	0.22	9	16.80	9.24	0
34	0.97			0.05	0.01	-0.31			6	16.80	9.26	0
35	0.96	-0.20			-0.04	-0.28		0.10	7	16.90	9.37	0
36	0.69	-0.23	+			-0.26			8	17.30	9.79	0
37	0.60	-0.54	+	0.50		-0.21	-0.02	0.22	11	17.70	10.13	0
38	0.97	-0.39					0.12		5	17.70	10.19	0
39	0.60	-0.55	+	0.49	0.05	-0.19		0.22	11	18.20	10.67	0
40	0.73		+	0.13		-0.31			8	18.40	10.83	0
41	0.95	-0.27			-0.04	-0.28	0.11	0.13	8	18.50	10.93	0
42	0.62	-0.55	+	0.49	0.06	-0.17	-0.02		11	18.70	11.16	0
43	0.96	-0.29			0.11				5	19.00	11.48	0
44	0.69	-0.31	+			-0.25	0.12		9	19.00	11.51	0
45	0.74		+			-0.31	0.05		8	19.50	11.93	0
46^	0.97								3	19.60	12.11	0
47	0.56	-0.66	+	0.60	0.18		-0.04		10	19.80	12.31	0
48	0.97			0.02	0.01	-0.31	0.03		7	19.90	12.35	0
49	0.96	-0.37			0.12		0.12		6	20.20	12.65	0
50	0.73		+		0.02	-0.30			8	20.20	12.71	0
51	0.97				0.18				4	20.60	13.07	0
52	0.62	-0.33	+						7	20.70	13.16	0
53	0.68	-0.23	+			-0.26		0.06	9	21.00	13.50	0
54	0.69	-0.23	+		-0.01	-0.26			9	21.10	13.61	0
55	0.73		+	0.14		-0.31	-0.01		9	22.00	14.48	0
56	0.97						0.03		4	22.10	14.58	0
57	0.98			0.03					4	22.20	14.70	0
58	0.73		+	0.13	0.04	-0.30			9	22.40	14.86	0
59	0.62	-0.41	+				0.13		8	22.40	14.90	0
60	0.60	-0.54	+	0.51	0.04	-0.19	-0.02	0.21	12	22.80	15.28	0
61	0.68	-0.30	+			-0.26	0.12	0.12	10	23.00	15.46	0
62	0.97				0.18		0.03		5	23.20	15.68	0
63	0.69	-0.31	+		0.01	-0.25	0.12		10	23.20	15.72	0
64	0.97			0.06	0.19				5	23.30	15.75	0
65	0.67		+						6	23.40	15.87	0
66	0.74		+		0.02	-0.30	0.05		9	23.50	15.94	0
67	0.62	-0.32	+		0.11				8	24.70	17.21	0
68	0.97	0.52		0.01	0.11		0.02		5	24.90	17.39	0
69	0.68	-0.23	+		-0.02	-0.27	3.0 2	0.07	10	25.10	17.52	0
70	0.67	J. _ J	+		0.18	J. _ /		J.J/	7	25.50	17.92	0
71	0.97		•	0.05	0.19		0.01		6	26.20	18.67	0
, 1	0.71			0.05	0.17		0.01		<u> </u>	20.20	10.07	

72	0.67		+	0.11					7	26.30	18.76	0
73	0.67		+				0.03		7	26.30	18.77	0
74	0.73		+	0.14	0.04	-0.30	-0.01		10	26.40	18.84	0
75	0.62	-0.40	+		0.13		0.13		9	26.80	19.30	0
76	0.68	-0.30	+		-0.01	-0.26	0.12	0.12	11	27.40	19.88	0
77	0.67		+	0.15	0.21				8	28.00	20.44	0
78	0.67		+		0.18		0.04		8	28.50	21.01	0
79	0.67		+	0.12			-0.02		8	29.80	22.22	0
80	0.66		+	0.17	0.21		-0.03		9	31.70	24.13	0

^{^ &#}x27;null' model with just the intercept and random effect term

Random term: 1|Species

Int: Intercept; DC: Dominance Certainty; SR: Sex Ratio; GS: Group Size; LC: Living Condition; SS: Social Style; T:

Transitivity

Supplementary Table 8: Summary of zero-averaged coefficients of the predictors from the candidate models (dAICc < 4 from the model with the lowest AICc score) for groom eigenvector coefficient

	В	Adj SE	Z	Pr (> z)	Importance
		3		(1 1)	1
(Intercept)	0.22	0.09	2.48	0.01*	
Dominance Certainty	0.06	0.14	0.43	0.67	0.23
Transitivity	0.02	0.09	0.23	0.82	0.14
Living Condition (captive vs free living)	0.01	0.05	0.17	0.86	0.09
Sex Ratio	0.00	0.04	0.09	0.93	0.07
Group Size	-0.02	0.09	0.26	0.80	0.13
Living Condition: Group Size	-0.01	0.11	0.10	0.92	0.01
Social Style (grade 1 vs grade 2)	0.00	0.03	0.07	0.94	0.02
Social Style (grade 1 vs grade 3)	0.00	0.04	0.09	0.93	
Social Style (grade 1 vs grade 4)	-0.01	0.07	0.12	0.90	

^{*}p < 0.05

Supplementary Table 9: Summary of zero-averaged coefficients from the candidate models for Newman's modularity and clustering coefficient of permuted grooming networks of 60% sampling effort

(A) Newman's Modularity (8 candidate models)							
	<u>B</u>	<u>Adj SE</u>	<u>z</u>	<u>Pr (> z)</u>	<u>Importance</u>		
Intercept	0.22	0.04	6.026	< 0.01			
Living Condition (Captive vs. Free living)	0.15	0.03	4.64	<0.01**	1		
Sex Ratio (proportion of females)	0.07	0.05	1.47	0.14	0.82		
Group Size	0.09	0.03	3.40	<0.01**	1		
Dominance Certainty	0.00	0.03	0.17	0.87	0.12		
Transitivity	0.00	0.01	0.16	0.87	0.12		
Group Size:	0.00	0.01	0.05	0.96	0.1		
Living Condition							
(B) Clustering Coefficient (7 candidate models)							
Intercept	0.87	0.11	7.73	<0.01**			
Group Size	-0.24	0.09	2.59	0.01*	1		
Living Condition (Captive vs. Free living)	-0.23	0.23	0.96	0.34	0.57		
Dominance Certainty	0.14	0.19	0.73	0.47	0.39		
Group Size: Living Condition	0.03	0.11	0.24	0.81	0.08		
Sex Ratio (proportion of females)	0.00	0.02	0.05	0.96	0.02		
Transitivity	0.00	0.01	0.05	0.96	0.01		

^a Effect sizes are standardized, comparable values (Gelman et al., 2009) Data are from 34 groups of 9 macaque species

^{*} $p \le 0.05$ ** p < 0.01

Supplementary Table 10: Summary of zero-averaged coefficients from the candidate models for Newman's modularity and clustering coefficient of permuted grooming networks of 20% sampling effort

(A) Newman's Modularity (8 candidate models)							
	<u>B</u>	<u>Adj SE</u>	<u>z</u>	<u>Pr (> z)</u>	<u>Importance</u>		
Intercept	0.25	0.04	6.25	<0.01**			
Living Condition (Captive vs. Free living)	0.14	0.03	4.52	<0.01**	1		
Sex Ratio (proportion of females)	0.03	0.04	0.72	0.47	0.82		
Group Size	0.10	0.02	4.31	<0.01**	1		
Dominance Certainty	0.00	0.02	0.19	0.85	0.12		
Transitivity	0.00	0.01	0.20	0.85	0.12		
Group Size: Living Condition	0.00	0.03	0.17	0.86	0.1		
Living Condition							
(B) Clustering Coefficient (7 candidate models)							
Intercept	0.61	0.10	5.88	<0.01**			
Group Size	-0.20	0.07	2.82	0.00**	1		
Living Condition (Captive vs. Free living)	-0.23	0.15	1.48	0.14	0.83		
Dominance Certainty	0.19	0.16	1.22	0.22	0.74		
Group Size: Living Condition	0.14	0.21	0.66	0.51	0.41		
Sex Ratio (proportion of females)	0.01	0.05	0.21	0.83	0.18		
Transitivity	-0.01	0.04	0.34	0.74	0.14		

^a Effect sizes are standardized, comparable values (Gelman et al., 2009) Data are from 34 groups of 9 macaque species

^{*} $p \le 0.05$ ** p < 0.01