

# The importance of mass accuracy in selected ion monitoring analysis of branched and isoprenoid tetraethers

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- 1 The importance of mass accuracy in selected ion monitoring
- 2 analysis of branched and isoprenoid tetraethers
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#### ABSTRACT

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Among the new proxies based on the distribution of glycerol dialkyl glycerol 14 tetraethers (GDGTs), the BIT index (Branched and Isoprenoid Tetraether 15 16 index) is one of the most difficult to determine accurately, as shown by two round-robin GDGT studies. Sensitivity to mass spectrometer settings and 17 tuning, and a diversity of mass spectrometry techniques may explain the 18 relatively large observed interlaboratory scatter. However, the mass defect 19 difference between crenarchaeol and branched GDGTs (brGDGTs) has never 20 been specifically scrutinized. In this study, we analyzed five sediment 21 samples with contrasting BIT values using about 60 m/z values to assess the 22 shape of GDGT peaks using selected ion monitoring. We then assessed the 23 biases on relative GDGT signals and mass spectrometry-derived BIT values 24 under two scenarios which ignore the systematic mass defect difference 25 between crenarchaeol and brGDGTs. Our results show that approximate 26 27 mass selection for GDGT analysis using selected ion monitoring generates losses of relative GDGT signals of up to 36%. The observed effects on BIT 28 values are maximal for intermediate BIT values, with shifts of BIT values of 29 30 ±0.1 unit. The shifts of BIT values due to approximate mass selection are thus not negligible compared to the interlaboratory scatter evidenced by the 31 32 latest round-robin GDGT study.

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- 34 Keywords: Mass defect difference, selected ion monitoring, HPLC-APCI-MS,
- 35 GDGTs, BIT index.

#### 1. Introduction

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The development of normal phase high performance liquid 38 chromatography coupled to positive ion atmospheric pressure chemical 39 40 ionization mass spectrometry (HPLC-APCI-MS) has greatly facilitated the analysis of polar lipids with high molecular weights (Hopmans et al., 2000). 41 Following this analytical development, a number of new proxies based on 42 the relative distributions of glycerol dialkyl glycerol tetraethers (GDGTs) 43 44 have been developed (Schouten et al., 2013b and References therein). Among the new GDGT-based proxies, the BIT index (Branched and 45 Isoprenoid Tetraether index; Hopmans et al., 2004) has been proposed as a 46 tracer for terrestrial organic carbon in sediments. The BIT index is a ratio 47 that compares the abundance of crenarchaeol, an isoprenoid GDGT 48 produced by Thaumarchaeota (e.g., Sinninghe Damsté et al., 2002), with 49 those of branched GDGTs (brGDGTs), which are produced by (acido)bacteria 50 51 (Weijers et al., 2009; Sinninghe Damsté et al., 2011, 2014). It is noteworthy that the BIT index, while widely determined, needs to be carefully 52 53 interpreted in order to reconstruct terrigenous fluxes (Schouten et al., 2013b 54 and References therein). The two international intercomparisons of GDGT measurements 55 (round-robin) have shown specific difficulties associated with calculation of 56 the BIT index (Schouten et al., 2009, 2013a). Escala et al. (2009) have shown 57 that the BIT index, which considers two different families of GDGTs, is 58 59 particularly sensitive to APCI conditions. Such sensitivity may explain part

of the relatively large interlaboratory scatter of the BIT index, especially for samples with intermediate BIT values (Schouten et al., 2009, 2013a). However, the approximate mass selection for GDGT analysis using selected ion monitoring (SIM) has not yet been explored as a reason for the poor reproducibility between labs of the BIT index. It is noteworthy that while many types of MS and GDGT analysis techniques exist, most laboratories analyzed GDGTs using SIM with single quadrupole mass spectrometers (QMS) during both round-robin GDGT studies (Schouten et al., 2009, 2013a). 

Following a review of about three hundred manuscripts where GDGTs were analyzed using SIM with QMS, we have found only a dozen manuscripts (ca. 4%) where m/z values are written with at least one decimal place (e.g., Herfort et al., 2006; Yang et al., 2011; Basse et al., 2014). In the other manuscripts, the m/z values are either systematically written as integer numbers (e.g., Cao et al., 2017; Freymond et al., 2017; Naafs et al., 2017), or not indicated at all (e.g., Keisling et al., 2017; Ruan et al., 2017; Woelders et al., 2017). This means that only a few manuscripts clearly reported exact m/z values rounded up to at least one decimal rather than approximate, integer m/z values for GDGT analysis using SIM.

In the present study, we analyzed five sediment samples with contrasting BIT values. We selected about  $60 \, m/z$  values for GDGT detection using SIM to obtain the relative GDGT signals depending on mass selection. We then assessed the biases on relative GDGT signals and on BIT values

under two scenarios that ignore the systematic mass defect difference between crenarchaeol and brGDGTs.

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#### 2. Materials and methods

The mass defect is defined as the exact molecular mass minus the nominal molecular mass – i.e., Mass defect =  $M_{exact}$  –  $M_{nominal}$ . GDGTs are long hydrogenated molecules and their mass defect values are around 1 or more. It should be noted that the mass defect is independent of the protonation – which adds about 1 Da to nominal and exact molecular masses. In this study, the assumption of a unique mass defect for all compounds is referred to as a "unique mass defect scenario".

Taking into account the newest brGDGT isomers (De Jonge et al., 2013; Ding et al., 2016), the BIT index equation of Hopmans et al. (2004) has been redefined as follows:

$$BIT = \frac{Ia + IIa_5 + IIIa_5 + IIa_6 + IIIa_7 + IIIa_7}{Cren + Ia + IIa_5 + IIIa_5 + IIIa_6 + IIIa_6 + IIIa_7 + IIIa_7}$$
(1)

Roman numerals refer to brGDGT isomers following the nomenclature of
Ding et al. (2016). Equation (1) can be simplified as BIT = A/(A+B), with A
and B the brGDGTs and crenarchaeol, respectively. In addition, A and B are
distinguished by their different mass defects (Table 1).

Five previously analyzed sediment samples with contrasting BIT values were selected: a modern marine sediment from the Bay of Marseille (Mediterranean Sea, 43.26°N, 5.29°E; about 60 m water depth), a modern lacustrine sediment from Lake Chad (LT17, 13°N, 14°E) and three sediment

samples from core MD04-2790. The core was retrieved from the continental shelf of the Black Sea (44.21°N, 30.99°E) during the ASSEMBLAGE 1 cruise of R.V. Marion Dufresne. The BIT values of the core and modern marine sediments were previously determined by Ménot and Bard (2012) and Sanchi et al. (2013), respectively. The lacustrine sediment LT17 and the marine sediment from the Bay of Marseille were selected and analyzed as in-house standard sediments to check the absence of instrumental drift in GDGT-derived indices.

New aliquots of the modern sediment samples were extracted at CEREGE with DCM:MeOH (9:1, v:v) using an accelerated solvent extractor ASE 350 (Dionex) at 120 °C and 10<sup>7</sup> Pa. Total lipid extracts were separated following the automated procedure established by Sanchi et al. (2013). The polar fractions of the five selected samples were then dissolved in hexane:isopropanol (98.2:1.8, v:v) prior to GDGT analysis on an Agilent 1260 Infinity HPLC coupled to a 6120 single QMS installed at CEREGE. The GDGTs were separated using similar chromatographic columns and solvents to Hopmans et al. (2016), as well as the same elution program, but with a 24 min re-equilibration time.

The APCI source conditions were: nebulizer pressure, 40 psi; vaporizer temperature, 325 °C; drying gas (N<sub>2</sub>) at 4 L/min and 325 °C; capillary voltage, 5 kV (positive mode); corona, 4  $\mu$ A; fragmentor, 170 V and 280 V for the internal standard C<sub>46</sub> (Huguet et al., 2006) and natural GDGTs, respectively. About 60 m/z values (42 of them being reported in

Supplementary Table S1) were considered for GDGT detection using SIM with a dwell time of 16 ms to assess the shapes of GDGT peaks (Fig. 1). The samples were analyzed five times each. All the analyses were carried out within a few days after an APCI autotune in positive mode, so the APCI source and QMS are assumed to be properly calibrated. The QMS was calibrated by monitoring the m/z values of four APCI calibration compounds: 121.1, 622.0, 922.0 and 1522.0. Because we considered the correct and incorrect m/z values at the same time, the cleanliness and stability conditions of the APCI source and quadrupole ion optics were not the limiting factors for our study as confirmed by the coherent and systematic differences described below.

The mass spectra at the apex of each GDGT peak were averaged over the five replicates of the five selected samples and then normalized to 100% (Fig. 1; Supplementary Table S1). The 5-, 6- and 7-methyl brGDGT isomers were treated as separate compounds due to the improved GDGT separation. The exact BIT values were calculated using equation (1) based on the GDGT peak areas at the m/z values of 1292.3, 1050.0, 1036.0 and 1022.0 (after, for instance, Herfort et al., 2006; Supplementary Table S2).

Two unique mass defects were tested: (i) a unique mass defect of 1.0, which favors brGDGTs over crenarchaeol, and (ii) a unique mass defect of 1.3, which favors crenarchaeol over brGDGTs (Fig. 1). The modified BIT values using the unique mass defects of 1.0 and 1.3 were calculated using equation (1) based on the GDGT peak areas at the m/z values of 1292.0,

1050.0, 1036.0 and 1022.0, and at the m/z values of 1292.3, 1050.3, 1036.3 and 1022.3, respectively (Supplementary Table S2). All single-ion chromatograms were obtained using m/z windows restricted to, for instance, 1050.3–1050.3 to extract the ion at m/z 1050.3 only. The shifts in BIT values were then determined for each unique mass defect and each sample as the modified BIT values minus the exact BIT values – i.e.,  $\Delta$ BIT = Modified BIT value – Exact BIT value. Errors on GDGT signals and BIT values were calculated based on the five replicates per sample and propagated to the shifts in BIT values using the variance formula. The biases on GDGT signals and BIT values were assessed with paired Student's t-tests, which were considered significant if p < 0.05.

#### 3. Results and discussion

Under the two unique mass defect scenarios, the relative losses of GDGT signals were calculated using the GDGT peak areas (Table 2). A deviation of 0.2–0.3 Da from the exact molecular mass generated losses of relative GDGT signals between 35 and 36% for crenarchaeol, and between 18 and 27% for brGDGTs (Table 2; all paired Student's t-tests on GDGT peak areas with p < 0.01). Therefore, for any given sample processing technique, chromatographic method, properly calibrated MS, mass setting and MS technique (e.g., Escala et al., 2009; Schouten et al., 2009, 2013), not taking into account the mass defect difference between crenarchaeol and brGDGTs is indeed sufficient to obtain different GDGT relative responses

and thus different BIT values. It is noteworthy that this caveat applies not only to the BIT index (Fig. 2; Table 3), but also to all GDGT-based indices that consider both isoprenoid GDGTs and brGDGTs.

Shifts in BIT values varied between 0.013 and 0.093 using a unique mass defect of 1.0, and between -0.055 and -0.011 using a unique mass defect of 1.3 (Fig. 2; Table 3; all paired Student's t-tests on BIT values with p < 0.01). Under both unique mass defect scenarios, the highest shifts in BIT values occurred for the sediment samples from the core MD04-2790, which also had intermediate BIT values (Fig. 2; Table 3). Our results are consistent with those from the two round-robin GDGT studies, which gave higher BIT index scatter for samples with intermediate BIT values than they did for samples with extreme BIT values (Schouten et al., 2009, 2013a). Theoretically, the observed shifts in BIT values are explained by a reduction by 35% of the relative crenarchaeol signal using the unique mass defect of 1.0, and by a general reduction by 21% of the relative brGDGT signal using the unique mass defect of 1.3 (Fig. 2).

To assess the severity of the observed and estimated shifts in BIT values due to approximate mass selection, these shifts are compared with the 95% confidence intervals of interlaboratory means for single QMS only based on the latest round-robin GDGT study (Schouten et al., 2013a). The observed biases on BIT values for the five selected sediment samples were generally larger than the interlaboratory scatter of the relevant round-robin samples for single QMS only (Fig. 2). The observed biases on BIT values are

also not negligible compared to the interlaboratory scatter for all MS types (Schouten et al., 2013a). This suggests that approximate mass selection probably contributes to interlaboratory differences.

Laboratories 16, 18, 23 and 25 summed [M+H]<sup>+</sup> and [M+H+1]<sup>+</sup> ions – protonated molecule and first isotope peak with one <sup>13</sup>C atom – for BIT index calculations instead of [M+H]<sup>+</sup> ions only during the latest round-robin GDGT study (Schouten et al., 2013a), which leads to another bias due to the various numbers of carbon atoms between the different GDGTs. To assess this bias, the abundances of [M+H+1]<sup>+</sup> ions relative to [M+H]<sup>+</sup> ions were calculated for the GDGTs included in the BIT index (Table 1) and then used to theoretically estimate the shifts in BIT values depending on the initial BIT index value. The calculated shifts reach a maximal value of –0.03 for an initial BIT index value close to 0.5 (Fig. 2), which means that the most likely sources of bias in addition to approximate mass selection are, among others, the diversity of APCI parameters and MS types (e.g., Escala et al., 2009; Schouten et al., 2009, 2013).

#### 4. Conclusions and perspectives

The results obtained from five sediment samples showed that not taking into account the systematic mass defect difference between crenarchaeol and brGDGTs generated shifts in BIT values of up to  $\pm 0.1$  unit. The observed shifts in BIT values represent a non-negligible proportion of the interlaboratory scatter. A round-robin GDGT study focusing on mass

220	selection for 51M experiments would greatly help to assess the contribution					
221	of approximate mass selection to the interlaboratory scatter of BIT values.					
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### Figure captions

Fig. 1. Examples of averaged mass spectra normalized to 100% of (A) crenarchaeol and of (B) brGDGT-Ia for the sediment sample at 2860 cm depth in core MD04-2790. The error bars correspond to the standard deviations (o) of the averaged relative GDGT signals per mass (mean value  $\pm 2\sigma$ , n=5). The blue and brown dashed lines describe the scenarios that consider a unique mass defect ( $\Delta$ M) of 1.0 and 1.3, respectively. The black curves represent the Gaussian functions fitted to the averaged mass spectra so that the shapes of GDGT peaks are more apparent.

Fig. 2. Shifts in BIT values ( $\Delta$ BIT) vs exact mass spectrometry-based BIT values according to the unique mass defect (1.0 or 1.3). The blue and brown symbols indicate the results from five sediment samples. The horizontal error bars correspond to the standard deviations (o) of the exact BIT values indicated in Table 3 (mean value  $\pm$  2 $\sigma$ , n = 5). The vertical error bars correspond to the standard deviations of the  $\Delta$ BIT values, which were determined by propagating the standard deviations shown in Table 3 using the variance formula (mean value  $\pm$  2 $\sigma$ , n = 5). The blue and brown solid curves indicate the theoretical results for given losses of crenarchaeol and brGDGT signals, respectively. The dark red solid curves indicate the theoretical results when [M+H]<sup>+</sup> and [M+H+1]<sup>+</sup> ions – protonated molecule and first isotope peak with one  $^{13}$ C atom, respectively – are summed for BIT index calculations instead of [M+H]<sup>+</sup> ions only. The colored double arrows

indicate the 95% confidence intervals ( $\pm t \times \mathrm{SD}/\sqrt{n}$ , with t=2.12 the t-statistic value for a two-sided 95% confidence interval with 16 degrees of freedom, SD the standard deviation and n=17) of interlaboratory means for single quadrupole mass spectrometers only from the latest round-robin GDGT study (Schouten et al., 2013a). The x-axis positions of the colored double arrows correspond to the averaged interlaboratory BIT values for single quadrupole mass spectrometers only.

#### Table captions

#### Table 1

Molecule names, nominal and exact masses of [M]<sup>+</sup> and [M+H]<sup>+</sup> ions – non-protonated and protonated molecules, respectively – mass defects of [M+H]<sup>+</sup> ions and abundances of [M+H+1]<sup>+</sup> ions – first isotope peak with one <sup>13</sup>C atom – relative to [M+H]<sup>+</sup> ions. The compounds are indicated following the nomenclature of Ding et al. (2016). All masses and mass defects shown here assume GDGTs with only <sup>12</sup>C, <sup>1</sup>H and <sup>16</sup>O isotopes. Note that the mass defect is independent of the protonation, which adds about 1 Da to both nominal and exact molecular masses.

#### Table 2

Relative losses of GDGT signals (in %) of five sediment samples using the unique mass defects of 1.0 and 1.3 for crenarchaeol and brGDGTs, respectively. The errors correspond to the standard deviations (o) per

407 compound and per sample (mean value  $\pm$  2 $\sigma$ , n = 5). The compounds are 408 indicated following the nomenclature of Ding et al. (2016).

409

## 410 **Table 3**

Exact and modified mass spectrometry-based BIT values of five sediment samples according to the unique mass defect (1.0 or 1.3). The errors correspond to the standard deviations (o) per scenario and per sample (mean value  $\pm 2\sigma$ , n = 5).

Figure 1

(A)

Solution

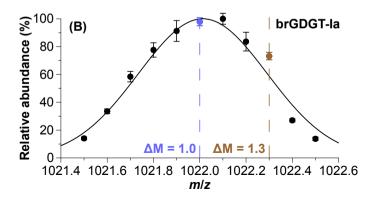
Crenarchaeol

AM = 1.0

AM = 1.3

1291.9

1292.1



1292.3 1292.5 1292.7 1292.9

Figure 2 0.1 -Observations and theoretical estimations Unique mass defect = 1.0 Unique mass defect = 1.3 -35% crenarchaeol area 0.05 -21% brGDGTs area Summing [M+H]<sup>+</sup> and [M+H+1]<sup>+</sup> ions (cases with only brGDGTs-la or -Illa) ΔBIT 0 95% confidence limits round-robin study 2011 Single quadrupole mass spectrometers only ←→ Sediment A -0.05 → Sediment B ←→ Sediment C ← Mixture D <→ Mixture E -0.1 -0.2 0.4 0.6 8.0 BIT exact value

## Table 1 - revised

Molecule name	Nominal mass for [M]+	Nominal mass for [M+H]+	Exact mass for [M+H]+	Mass defect for [M+H]+	[M+H+1]+/[M+H]+ ions
Crenarchaeol/Crenarchaeol regio-isomer	1290	1291	1292.2444	1.2444	0.946
brGDGT-IIIa5/brGDGT-IIIa6/brGDGT-IIIa7	1048	1049	1050.0488	1.0488	0.748
brGDGT-IIa5/brGDGT-IIa6/brGDGT-IIa7	1034	1035	1036.0325	1.0325	0.737
brGDGT-Ia	1020	1021	1022.0166	1.0166	0.726

# Table 2

Sample	Ia	$IIa_5$	$IIa_6$	IIa <sub>7</sub>	$IIIa_5$	$IIIa_6$	$IIIa_7$	Cren
Bay of Marseille	$27 \pm 1$	$24 \pm 6$	$20 \pm 5$	$21 \pm 8$	$20 \pm 1$	$20 \pm 4$	$25 \pm 4$	$35 \pm 2$
Lake Chad LT17	$25 \pm 2$	$21 \pm 2$	$21 \pm 4$	$23 \pm 7$	$20 \pm 4$	$21 \pm 2$	$21 \pm 3$	$35 \pm 2$
MD04-2790 2851 cm	$26 \pm 2$	$21 \pm 4$	$22 \pm 2$	$23 \pm 7$	$19 \pm 1$	$20 \pm 2$	$21 \pm 5$	$36 \pm 3$
MD04-2790 2860 cm	$27 \pm 2$	$22 \pm 5$	$22\pm4$	$25 \pm 10$	$19 \pm 3$	$18 \pm 4$	$18 \pm 8$	$35 \pm 1$
MD04-2790 2870 cm	$26 \pm 3$	$22 \pm 2$	$20 \pm 2$	$26 \pm 3$	$19 \pm 2$	$20 \pm 4$	$23 \pm 11$	$35 \pm 2$

# Table 3

Sample	Exact value	Unique mass defect = $1.0$	Unique mass defect = 1.3
Bay of Marseille	$0.174 \pm 0.007$	$0.246 \pm 0.004$	$0.140 \pm 0.006$
Lake Chad LT17	$0.963 \pm 0.002$	$0.976 \pm 0.001$	$0.952 \pm 0.002$
$MD04\text{-}2790\ 2851\ cm$	$0.661 \pm 0.014$	$0.753 \pm 0.013$	$0.607 \pm 0.012$
$\mathrm{MD04}\text{-}2790\ 2860\ \mathrm{cm}$	$0.635 \pm 0.011$	$0.728 \pm 0.007$	$0.580 \pm 0.012$
$\mathrm{MD04}\text{-}2790\ 2870\ \mathrm{cm}$	$0.651 \pm 0.007$	$0.743 \pm 0.003$	$0.596 \pm 0.010$