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**Modeling and optimization of lipid accumulation by *Yarrowia lipolytica* from glucose
under nitrogen depletion conditions[†]**

Running Title: Modeling and optimization of lipid accumulation by *Yarrowia lipolytica*

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ABSTRACT

Oleaginous yeasts have been seen as a feasible alternative to produce the precursors of biodiesel due to their capacity to accumulate lipids as triacylglycerol having profiles with high content of unsaturated fatty acids. The yeast *Yarrowia lipolytica* is a promising microorganism that can produce lipids under nitrogen depletion conditions and excess of the carbon source. However, under these conditions, this yeast also produces citric acid (overflow metabolism) decreasing lipid productivity. This work presents two mathematical models for lipid production by *Yarrowia lipolytica* from glucose. The first model is based on Monod and inhibition kinetics, and the second one is based on the Droop quota model approach, which is extended to yeast. The two models showed good agreements with the experimental data used for calibration and validation. The quota based model presented a better description of the dynamics of nitrogen and glucose dynamics leading to a good managing of N/C ratio, which makes this model interesting for control purposes. Then, quota model was used to evaluate, by means of simulation, a scenario for optimizing lipid productivity and lipid content. For that, a control strategy was designed by approximating the flow rates of glucose and nitrogen with piecewise linear functions. Simulation results achieved a productivity of $0.95 \text{ g.L}^{-1}.\text{h}^{-1}$ and a lipid content fraction of 0.23 g.g^{-1} , which indicate a promising alternative for the optimization of lipids production. This article is protected by copyright. All rights reserved

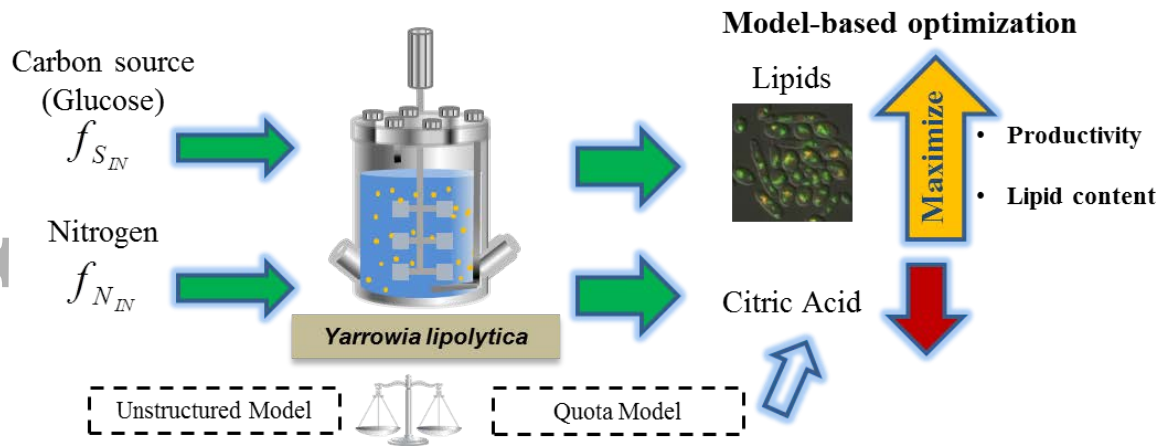
Keywords: Biodiesel; Droop Model; Nitrogen quota; Unstructured modeling; *Yarrowia lipolytica*.

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HIGHLIGHTS

- Two mathematical models for lipid production by *Yarrowia lipolytica* are developed.
- The Droop model for microalgae is extended for yeast.
- The models are calibrated and validated with independent data sets.
- Multi objective optimization of lipid accumulation is studied with the quota model.
- A simple control strategy for critical quota is promising to optimize lipid productivity.

Graphical Abstract



This work presents two mathematical models of the metabolism of *Yarrowia lipolytica*. The first model is based on Monod and inhibition kinetics, and the second one is based on the Droop quota model approach. The quota based model was used for maximizing lipid productivity and lipid content in simulation.

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1. INTRODUCTION

The use of microbial lipids as precursors of biodiesel production has outlasted in the last years as an attractive and sustainable alternative (Papanikolaou and Aggelis, 2009). These lipids are produced by oleaginous microorganisms up to 20% of their dry weight (Ratledge, 2004). An interesting oleaginous microorganism is the yeast *Yarrowia lipolytica*. This yeast is able to use several substrates such as: glucose, fructose, lactose, sucrose, whey, glycerol, fatty acids, molasses, glucose/glycerol, glucose/stearin and glycerol/stearin (Nicaud *et al.*, 2002; Papanikolaou and Aggelis, 2002; Papanikolaou and Aggelis, 2003a; Papanikolaou and Aggelis, 2011a; Rakicka *et al.*, 2015). *Yarrowia lipolytica* has been identified as a potential candidate for the synthesis of biodiesel due to its production of high quantity of triacylglycerols (TAG) (>90% with respect to its total accumulation) with a rich fatty acid profile in linoleic acid (C18:2) (56%), oleic acid (C18:1) (28%), and palmitic acid (C16:0) (11%) (Beopoulos *et al.*, 2009; Papanikolaou and Aggelis, 2011b). Furthermore, this yeast is the only oleaginous yeast with a complete sequenced genome (Barth and Gaillardin, 1997), making it a suitable candidate for genetic and metabolic engineering aiming the production of large amounts of lipids with specific fatty acids composition (Nicaud 2012). When *Yarrowia lipolytica* grows in nitrogen depletion conditions with substrates as glucose, ethanol, and starch hydrolysates; citric acid is secreted in significant amounts (Papanikolaou *et al.*, 2006). This issue hampers the optimization of the carbon flux into lipids. The outbreak of lipid accumulation follows the exhaustion of a nutrient in the medium, mainly nitrogen (Papanikolaou and Aggelis, 2011a). This exhaustion reduces cell growth, but increases lipid content. The main challenge of this trade-off relationship is thus focused on increasing lipid content while maintaining an adequate cell growth through the manipulation of nitrogen fluxes (Yoo *et al.*, 2014).

Few mathematical models have been proposed to study lipid accumulation by oleaginous

yeasts and fungus in different conditions and substrates (Economou *et al.*, 2011; Fakas *et al.*, 2009; Meeuwse *et al.*, 2011; Meeuwse *et al.*, 2012; Ykema *et al.*, 1986). For example, the model developed by (Economou *et al.*, 2011) for the fungus *Mortierella isabellina*, assumes that high substrate concentration inhibits microbial growth and that high nitrogen concentration decreases lipid concentration. Meeuwse *et al.*, (2012) reported a model for the fungus *Umbelopsis isabellina* in three phases: exponential growth phase, lipid accumulation phase, and lipid turnover phase. This was accomplished by the definition of certain fixed times to switch phases. Models with a particular interest on *Yarrowia lipolytica* have considered the mechanisms of lipid accumulation (Papanikolaou and Aggelis, 2002; Papanikolaou and Aggelis, 2003b), and citric acid production (Arzumanov *et al.*, 2000; Papanikolaou and Aggelis, 2003a), separately. All these models represented acceptably the dynamics of growth and accumulated lipids concentration from fat substrates. However, there is only one existing model for the concomitant production of lipids and citric acid (Papanikolaou *et al.*, 2006), which fails to predict lipid accumulation since it was applied for the production of structural lipids (membrane lipids). Other modeling approaches have considered variant yields, such as the model proposed by Droop (Droop, 1968), where biomass production depended on a variable cell quota defined as the fraction of internal nutrient per unit of biomass (Lemesle and Mailleret, 2008). These models have only been presented for microalgae on: autotrophic (Mairet *et al.*, 2011), heterotrophic (Abdollahi and Dubljevic, 2012; De la Hoz Siegler *et al.*, 2011; Surisetty *et al.*, 2010b), and mixotrophic (Adesanya *et al.*, 2014; Yoo *et al.*, 2014) conditions. Nevertheless, lipid accumulation in those models for microalgae was also triggered by the depletion of nitrogen, and thus this similarity between microalgae and yeast is encouraging to propose new models based on quota definitions. In this context, this paper proposes a quota based model for yeast in order to expose its efficiency on modeling and optimization problems. The present paper is organized as follows: Section 2 describes the

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experimental data used in this study, and it details the developed mathematical models. Section 3 presents the calibration and the validation of the models, and it proposes the optimization of lipid accumulation by finding the optimal input flow rates. Finally, conclusions are drawn in section 4.

2. MATERIALS AND METHODS

2.1. Strain and medium

Three independent experiments (culture A, B and C) were performed with the strain *Yarrowia lipolytica* W29 obtained from the Microbiology and Molecular genetics laboratory (Paris-Grignon, France). The optimal growth conditions of this yeast are: 28 °C and pH of 5.6 (Faure, 2002).

The strain was maintained at -80°C in yeast extract peptone dextrose (YPD) medium (10 g.L⁻¹ yeast extract, 10 g.L⁻¹ peptone and 10 g.L⁻¹ glucose) supplemented with 30% (v/v) glycerol. For each culture, one glycerol stock was streaked on a Petri dish (YPD agar medium containing 10 g.L⁻¹ agar) and incubated at 30°C for 48 h. Pre-cultivations were carried out first in a 5 mL tube of YPD medium at 30°C for 16 h inoculated with one colony from Petri dish. Then, two successive steps of 16 h pre-cultures were carried out in baffled-shake flasks inoculated with a 10 % (v/v) ratio with increasing culture volumes and containing mineral medium with the following composition: K₂HPO₄: 3 g.L⁻¹; (NH₄)₂SO₄: 3 g.L⁻¹; NaH₂PO₄.H₂O: 3 g.L⁻¹; MgSO₄.7H₂O: 1 g.L⁻¹; ZnSO₄.7H₂O: 0.04 g.L⁻¹; FeSO₄.7H₂O: 0.0163 g.L⁻¹; MnSO₄.H₂O: 0.0038 g.L⁻¹; CoCl₂.6H₂O: 0.0005 g.L⁻¹; CuSO₄.5H₂O: 0.0009 g.L⁻¹; Na₂MoSO₄.2H₂O: 0.00006 g.L⁻¹; CaCl₂.2H₂O: 0.023 g.L⁻¹; H₃BO₃: 0.003 g.L⁻¹; and 10 mL of vitamins solution (Ochoa-Estopier, 2012). The reactor, which contained the same composition of mineral medium without (NH₄)₂SO₄, was inoculated with a 10% (v/v) ratio.

2.2. Analytical methods

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The data from the three independent experiments were determined with the same protocols. Yeast concentration was quantified by spectrophotometry at 600 nm in a spectrophotometer HITACHI U-1100. For dry cell weight, culture samples (5–10 mL) were harvested by filtration on a 0.45 μm membrane (Sartorius) and dried at 200 mm Hg and 60 °C during 48 h until a constant weight was achieved. Determination of alcohols, organic acids, and sugar concentrations from supernatants was performed by HPLC using a column Aminex HPX-87H (300 mm*7.8 mm) with the following conditions: Temperature of 50 °C, with 5 mM H₂SO₄ as eluent (flow rate of 0.5 mL.min⁻¹) and dual detection (refractometer and UV at 210 nm). Compounds were identified and quantified with respect to standards. An ammonium ion electrode (PH/ISE meter model 710A + Ammonia Gas Sensing Electrode Model 95-12, Orion Research Inc. Boston USA) was used to quantify the residual ammonia concentration in the culture medium. The quantification of lipids was carried out as described by Browse et al. (1986) 20 mg of lyophilized cells were added to 1 mL of methanol solution containing 25 mL.L⁻¹ of sulfuric acid (95%) and internal standard (C9:0). The mix in sealed tubes was placed in a 80°C bath for 90 minutes. After cooling, 450 μL hexane and 1.5 mL water were added and vortexed. The lipidic phase was then sampled and the composition was analyzed by Gas Chromatography as mentioned in Cescut *et al.*, (2011).

2.3. Experimental data

Culture A (Figure 1 a) is a fed-batch reported by Cescut, (2009) where *Yarrowia lipolytica* was grown in a 20 L bioreactor (Biostat E. Braun, Melsungen, Germany). The dissolved oxygen was maintained above 20% of saturation by modulating the stirring rate and/or the air flow rate in order to avoid oxygen limitation. Inoculation volume was of 800 mL. Operating conditions were fixed and controlled to a temperature of 28°C and a pH value of 5.5 regulated by addition of a 10 M NH₃ solution (growth phase) or 5 M KOH solution (after nitrogen limitation). Input air flow rates and composition of input and output air, as well as the

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bioreactors parameters (*e.g.* Temperature, pO₂, pH, agitation) were measured online and acquired by custom made software. The flow rates of nitrogen (N_{IN} , ammonia 5 M) and carbon (S_{IN} , glucose 730 g.L⁻¹) sources varied continuously (Figure 1 a) to induce different levels of nitrogen limitation.

Culture B (Figure 1 b) corresponds to a sequential batch culture under nitrogen starvation. This was obtained from a 5 L stirred tank bioreactor (Biostat B. Braun Biotech International, Sartorius AG, Germany) with the acquisition software MFCS/win 2.1[®]. The inoculation volume was 300 mL. The dissolved oxygen was maintained above 20% of saturation. The working volume of the reactor was 3 L, where the operating conditions were set to a controlled temperature of 28°C and a pH value of 5.6 regulated by a 5 M KOH solution. The initial medium contained 100 g.L⁻¹ of glucose and 4.14 g.L⁻¹ of (NH₄)₂SO₄. The carbon source (glucose) was fed by two pulses at 35 h and 55 h (Figure 1 b). The first half of this culture was used for model calibration and the second half for model validation.

Culture C (Figure 1 c) is a sequential batch culture under nitrogen starvation, where the carbon source (glucose) was fed by one pulse at 19.3 h (Figure 1 c). The initial medium contained 6.6 g.L⁻¹ of (NH₄)₂SO₄ and 100 g.L⁻¹ of glucose. The reactor and operating conditions were the same as the reported for culture B.

2.4. Model assumptions

Our models assume that the metabolism and cell growth of *Yarrowia lipolytica* are driven by the uptake of two extracellular nutrients: carbon (*i.e.* glucose) and nitrogen (*i.e.* ammonia). Figure 2 displays the different physiological states occurring in the cell. These phenomena have as first priority the production of functional biomass (catalytically active biomass), which has an elementary composition of CH_{1.744}O_{0.451}N_{0.132} (Ochoa-Estopier, 2012). Under intracellular nitrogen deficient or limited conditions, the yeast accumulates lipids (mainly TAGs) and storage compounds (*e.g.* carbohydrates). However, under these conditions and carbon excess

citric acid is produced and excreted (Papanikolaou *et al.*, 2006 ; Ochoa-Estopier and Guillouet, 2014). This production is the result of overflow metabolism leading to by-product formation (Amribt *et al.*, 2013; Santos *et al.*, 2012). These by-products are often inhibitory or competing products (*i.e.* citric acid) that could decrease the production of the target metabolite (TAGs in our case). Sugars can also have an inhibitory effect and it was thus considered as suggested by Economou *et al.* (2011). The dynamics of *Yarrowia lipolytica* metabolism under discontinued operation can be described by balance equations considering the concentrations (g.L^{-1}) of Glucose (S), Nitrogen (N), Functional biomass (X_f), Lipid biomass (X_L), Citric Acid (CIT), and volume (V , expressed in L) undertaking homogenous well-mixed conditions with non-limited oxygen requirements. It is noteworthy that the functional biomass contains a part of lipids, which are components of the membrane. These lipids are called structural lipids X_{LS} and are intrinsic to the functional biomass in a constant fraction γ equivalent to $0.06 \text{ g}X_L.\text{g}X_f^{-1}$, (Robles-Rodriguez *et al.*, 2017). This definition allows to accounting for a separate term of accumulated lipids X_{LA} . For simplicity and consistency with experimental data, only Total lipids ($X_L = X_{LS} + X_{LA}$ with $X_{LS} = \gamma.X_f$) are described in the mass balances.

$$\frac{d}{dt}S = S_{IN} \cdot \frac{f_{S_{IN}}}{V} - \left(\mu \frac{1}{Y_{X/S}} + \pi_{LIP} \frac{1}{Y_{LIP/S}} + \pi_{CIT} \frac{1}{Y_{CIT/S}} \right) X_f - \frac{S}{V} F_{IN} \quad (1)$$

$$\frac{d}{dt}N = N_{IN} \cdot \frac{f_{N_{IN}}}{V} - \rho_N X_f - \frac{N}{V} F_{IN} \quad (2)$$

$$\frac{d}{dt}X_f = \mu.X_f - \frac{X_f}{V} F_{IN} \quad (3)$$

$$\frac{d}{dt}X_L = (\pi_{LIP} + \gamma.\mu)X_f - \frac{X_L}{V} F_{IN} \quad (4)$$

$$\frac{d}{dt}CIT = \pi_{CIT}.X_f - \frac{CIT}{V} F_{IN} \quad (5)$$

$$\frac{dV}{dt} = F_{IN} - F_{Sample} \quad (6)$$

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S_{IN} and N_{IN} (g.L^{-1}) hold for the input concentration of glucose and nitrogen, respectively.

The term F_{IN} (L.h^{-1}) considers the inputs to the reactor which is mathematically defined as the sum of the input flow rate of glucose $f_{S_{IN}}$ and nitrogen $f_{N_{IN}}$ as $F_{IN} = f_{S_{IN}} + f_{N_{IN}}$. The volume equation (6) includes a term F_{Sample} to take into account sampling volumes. The terms $Y_{X/S}$, $Y_{LIP/S}$, and $Y_{CIT/S}$ are the yield coefficients (g.g^{-1}) for functional biomass, lipids, and citric acid production with respect to glucose, respectively. Growth rate, lipid production rate, and the citric acid production rate, are represented by μ , π_{LIP} , and π_{CIT} respectively, whereas the nitrogen uptake rate is ρ_N . For consistency with experimental data, total Biomass concentration (X_T) is accounted as the sum of the functional biomass (without the structural lipid part) and lipid biomass as,

$$X_T = X_f(1 - \gamma) + X_L \quad (7)$$

Growth rate μ was defined to take into account the dependency of yeast growth on carbon and nitrogen sources. The impact of nitrogen on growth and lipid accumulation is further detailed. This nitrogen effect has been studied on yeast and fungus by kinetic models (Economou *et al.*, 2011; Papanikolaou *et al.*, 2006; Ykema *et al.*, 1986), and in microalgae by the definition of intracellular quotas as in the Droop model (Droop, 1968; Mairet *et al.*, 2011). In this context, two macroscopic models are here developed based on equations (1 – 7).

2.6. Unstructured Model

The unstructured model assumes that external nitrogen is taken up directly for the production of biomass. Therefore, in the growth rate (μ), the uptake of carbon and nitrogen is described by a double substrate limitation with sugar inhibition as,

$$\mu = \mu^{\max} \left(\frac{S}{K_{S,1} + S + S^2/k_{I,1}} \right) \left(\frac{N}{K_N + N} \right) \quad (8)$$

where $K_{S,1}$, and K_N are the saturation constants for glucose and nitrogen, respectively, μ^{\max} is the specific maximum growth rate, and $k_{I,1}$ is the glucose inhibition constant. In addition, the nitrogen uptake rate (ρ_N) is considered to be proportional to growth rate,

$$\rho_N = \mu \frac{1}{Y_{X/N}} \quad (9)$$

where $Y_{X/N}$ is the yield coefficient for functional biomass production with respect to nitrogen.

The specific lipid accumulation rate π_{LIP} was assumed to follow Andrew's equation for glucose. Nitrogen depletion was considered by an inhibition constant (k_1) ensuring that lipid accumulation is low at high concentrations of nitrogen as developed by (Economou *et al.*, 2011). Moreover, an inhibition by citric acid was added to include a competition with the production of citric acid.

$$\pi_{LIP} = \pi_{LIP}^{\max} \left(\frac{S}{K_{S,2} + S + S^2/k_{I,2}} \right) \left(\frac{k_1}{k_1 + N} \right) \left(\frac{k_2}{k_2 + CIT} \right) \quad (10)$$

The term π_{LIP} is the maximum lipid production rate. $K_{S,2}$, $k_{I,2}$, and k_2 represent the saturation and inhibition constants for glucose, and the inhibition constant due to citric acid concentration, respectively.

Citric acid production occurs as overflow metabolism under nitrogen starvation or limitation conditions and excess of carbon substrate (*i.e.* glucose). This implies a decrease on the production rate of functional biomass (μ) and a stabilization of lipid production rate (π_{LIP}). Therefore, it can be assumed that when the Glucose Uptake Rate for lipids and biomass (GUR_X) decreases to a minimum point Of_U , cells switch to overflow metabolism.

$$GUR_X = \mu \frac{1}{Y_{X/S}} + \pi_{LIP} \frac{1}{Y_{LIP/S}} \quad (11)$$

Hence, glucose uptake rate and the saturation point Of_U can be used to deduce an indicator (Ind_U) of overflow metabolism (Amribt *et al.*, 2013).

$$Ind_U = \begin{cases} 0, & GUR_x > Of_U \\ 1, & GUR_x \leq Of_U \end{cases} \quad (12)$$

The indicator Ind_U is defined as Boolean for which $Ind_U = 1$ assumes overflow. Citric acid production rate (π_{CIT}) is therefore described by this indicator as,

$$\pi_{CIT} = \pi_{CIT}^{\max} (Ind_U) \left(\frac{S}{K_{S,3} + S + S^2/k_{I,3}} \right) \left(\frac{k_3}{k_3 + CIT} \right) \quad (13)$$

Additionally, the citric acid production rate also considers an inhibition due to citric acid.

The maximum citric acid production rate is indicated by π_{CIT}^{\max} , whereas the saturation and inhibition constants for glucose, and the inhibition constant for citric acid are $K_{S,3}$, $k_{I,3}$, and k_3 , respectively.

2.7. Intracellular Quota Model

In this work, it is assumed that the Droop model (Droop, 1968) used for microalgae on heterotrophic conditions can also be applied for yeasts. The main characteristic of this model is the inclusion of variable yield coefficients called quotas. In this way, the Droop model provides regulation where the internal quota of a limiting substrate drives the production of functional biomass (Mairet *et al.*, 2011).

Microbial growth is modulated by the internal nitrogen quota q_N (Droop, 1968), defined by

$$\frac{d}{dt} q_N = \rho_N - \mu q_N \quad (14)$$

where ρ_N is the nitrogen uptake rate, and μ is the growth rate. In this model, the uptake of carbon for growth is described by the Andrew's equation, while the nitrogen effect is described by the Droop model. Hence, growth rate is denoted as,

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$$\mu = \mu^* \left(1 - \frac{Q_0}{q_N} \right) \left(\frac{S}{K_{S,1} + S + S^2/k_{I,1}} \right) \quad (15)$$

where Q_0 is the minimum nitrogen quota at which cell growth is possible, and μ^* is the theoretical maximum growth rate. The nitrogen uptake rate ρ_N is given by

$$\rho_N = \rho_N^{\max} \cdot \left(\frac{N}{K_N + N} \right) \quad (16)$$

where K_N is the nitrogen saturation constant, and ρ_N^{\max} is the maximum nitrogen uptake rate.

The lipid accumulation rate, π_{LIP} , was also described by Andrew's equation for glucose. An inhibitory effect on citric acid was also included as in (10). Moreover, it is assumed that lipid production decreases as the lipid content reaches its theoretical maximum (De la Hoz Siegler *et al.*, 2011; Surisetty *et al.*, 2010a; Yoo *et al.*, 2014).

$$\pi_{LIP} = \pi_{LIP}^{\max} \cdot \left(\frac{S}{K_{S,2} + S + S^2/k_{I,2}} \right) \left(1 - \frac{1}{\alpha} \frac{X_L}{X_T} \right) \left(\frac{k_2}{k_2 + CIT} \right) \quad (17)$$

The term α represents the maximum lipid content, which is $0.36 \text{ gL.gX}_T^{-1}$ for *Yarrowia lipolytica* W29 (Beopoulos *et al.*, 2009; Ratledge and Wynn, 2002). The overflow to produce citric acid is the result of an excessive substrate supply. In this context, the carbon uptake for lipids and functional biomass must encounter a saturation point. It is known that lipid accumulation in yeast depends on the N/C ratio of the inflow rate of nitrogen and carbon (Beopoulos *et al.*, 2009; Cescut, 2009; Ochoa-Estopier and Guillouet, 2014; Sitepu *et al.*, 2014; Ykema *et al.*, 1986). The critical values for the N/C ratio at which citric acid is produced concomitantly with lipids have been reported at $0.021 \text{ molN.Cmol}^{-1}$ (Ochoa-Estopier and Guillouet, 2014) for D-stat experiments, and identified at $0.077 \text{ molN.Cmol}^{-1}$ (Robles-Rodriguez *et al.*, 2017) after model identification. In addition to the input flow rates, some studies have focused on the changes of the consumption rate of the N/C ratio, which is related to the intracellular change of N/C. Based on these ideas, it is straightforward to think that there

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exists a critical quota (q_N^*), where the process shifts towards citric acid overflow. Therefore, it is possible to assume that citric acid is produced when q_N is lower than a critical value q_N^* .

In order to keep consistency with the formulation of (13), a Boolean indicator (Ind_Q) is also proposed as,

$$Ind_Q = \begin{cases} 0, & q_N > q_N^* \\ 1, & q_N \leq q_N^* \end{cases} \quad (18)$$

Therefore, citric acid production occurs when $q_N \leq q_N^*$. Citric acid production rate, π_{CIT} , was then taken as in (13) in which the Boolean indicator Ind_U was replaced by Ind_Q ,

$$\pi_{CIT} = \pi_{CIT}^{\max} \cdot (Ind_Q) \left(\frac{S}{K_{S,3} + S + S^2/k_{I,3}} \right) \left(\frac{k_3}{k_3 + CIT} \right) \quad (19)$$

The difference to the unstructured model relies on the manner of computing the indicator.

Summarizing, the unstructured model describes 6 state variables whose dynamics are represented by equations (1 – 6), and their kinetics by equations (8 – 13). The resulting model has 18 parameters. The quota based model has also 18 parameters with 7 mass balance equations: 6 state variables (equations 1 – 6), and the intracellular nitrogen quota (equation 14). Kinetics are disclosed on equations (15 – 19). The total biomass is evaluated by equation (7) for both models. The calibration and validation of the models are presented in the next section.

3. RESULTS AND DISCUSSIONS

3.1. Model calibration

Parameter calibration was carried out by a Particle Swarm Optimization (PSO) algorithm (Eberhart and Kennedy, 1995), and further refined by Pattern Search (Lewis and Torczon, 1996) implemented in MATLAB®. The data of Culture A and Culture B ($t=0 - 38h$) were used to calibrate the models, whose results are displayed in Figure 3. The unstructured model and the quota model depict similar fit in both cultures. However, it is remarkable that nitrogen concentration is overestimated by 2 g.L^{-1} before 10 h with the unstructured model for culture

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A, whereas the estimation with the quota model is in better agreement with the experimental data. Both models display small underestimation in lipids and citric acid concentration at the end of culture A. This is due to the estimation trade-off between the data from both cultures where lipids and citric acid concentrations are smaller for culture B than for culture A. In Figure 3 b, it is observed that when nitrogen was depleted (21 h for the quota and 23 h the unstructured model) functional biomass was no longer produced. This could be explained regarding some of the assumptions of the models. The unstructured model assumed that growth was based on glucose and nitrogen consumption; thus when nitrogen was depleted the total biomass would have increased due to lipids accumulation. On the other hand, the quota model assumed that there is a minimum nitrogen quota Q_0 to assure growth, which was reached subsequently to the extracellular nitrogen depletion causing a stabilization of functional biomass. Nevertheless, the results indicate that functional biomass concentration continued to rise probably due to the storage of other compounds that were not measured (*i.e.* carbohydrates). The overflow indicators (Ind_U or Ind_Q) are displayed for both cultures (Figure 3), where it is shown that the unstructured model started before due to the differences on the depleted nitrogen for both cultures.

The estimated parameters for both models are presented in Table I and compared with previous values reported in the review of Papanikolaou and Aggelis, (2011a). The values of μ^{\max} , π_{LIP}^{\max} , and $Y_{LIP/S}$ were also experimentally identified for culture A. The theoretical maximum growth rate μ^* for the quota model is higher than the one obtained experimentally, as expected since it depends on the maximum and minimum values of the quota. In the unstructured model, μ^{\max} is close but also higher than the experimental value due to the glucose inhibition term. With respect to the values of the specific production rates π_{LIP}^{\max} , they were consistent with the parameters estimated by (Fakas *et al.*, 2009), and the experimental

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values. Moreover, lipid yields $Y_{LIP/S}$ were also consistent with the experimental values. With respect to the parameters affecting citric acid, the estimated maximum specific production rate π_{CIT}^{max} and the yields $Y_{CIT/S}$ were higher than the reported in literature (Papanikolaou et al., 2008), which could be explained by the high production of citric acid when glucose was added continuously and the differences on the kinetic equations. Regardless their differences in structure, both models reported similar results in the non-growth related parameters. The main distinction was observed on the saturation constant for nitrogen K_N , where the higher value on the quota model is compensated by the nitrogen quota and it is almost equivalent to $Y_{X/N}$. This was expected since the equations of nitrogen uptakes for both approaches (Unstructured (9) and Quota model (16)) are different. The values of the inhibition constants (Table I) show that the glucose inhibition effect differs in magnitude between citric acid production, lipid accumulation, and growth. The estimated value of the critical quota (q_N^*) was interestingly found to be similar to the critical value of the N/C ratio reported by (Robles-Rodriguez et al., 2017).

The determination coefficient R^2 , and the Root Mean Squared Errors ($RMSE$) were calculated to prove the quality and the enforcement of each model (Table II). The results for nitrogen in culture B (nitrogen starvation culture) are not reported since nitrogen data was not available. The results of the $RMSE$ and R^2 demonstrate similar conclusions for the different variables, which are also consistent with Figure 3. With respect to the $RMSE$ for culture A, it is seen that the unstructured model presented smaller values for citric acid and glucose, whereas the quota model was better for functional biomass, nitrogen and lipid concentrations. For culture B, the $RMSE$ values for glucose and citric acid concentrations were smaller for the quota model with similar values for functional biomass and lipid concentrations.

3.2. Model Validation

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Culture B ($t = 38 - 80\text{h}$) and culture C were used to validate the model, considering sequential carbon feeding cultures under nitrogen starvation. The validation results of the two models are displayed in Figure 4. Both models presented good agreement with the experimental data where a similar performance is observed for both validation cultures. It is worth noting that experimental data for nitrogen concentration was not available for culture B, and only the initial point of culture C was known from the initial medium composition.

In Figure 4, functional biomass concentration was stable after nitrogen depletion due to the growth dependency on the nitrogen source availability. The main differences between models were observed on nitrogen concentrations, for which no comparison could be made due to the lack of data. However, this difference was reflected on the shift to overflow metabolism identified by the indicators in equations (12) and (18) for the unstructured and the quota model, respectively. The switching times for overflow in culture B (Figure 4 a) were at 21 h for the quota model and at 23 h for the unstructured model. The difference between switching times were also observed for culture C (Figure 4 b) and culture A (Figure 3 a). However, this difference is bigger on culture A. This can be explained by the two different phenomena acting on the cell metabolism: nitrogen limitation or nitrogen starvation.

Statistical analysis was also performed for both models with the validation cultures, whose results are depicted in Table III. The values of the *RMSE* showed that the quota model performed better estimation for glucose and functional biomass concentrations, whilst citric acid was lower with the unstructured model, and similar results were reported for lipid concentrations in culture B. These results for *RMSE* were maintained for culture C.

To strengthen the validity of our model, *in silico* simulations were challenged against published experimental data of different strains of *Yarrowia lipolytica* growing on glucose under batch conditions (Kavšček *et al.*, 2015; Papanikolaou *et al.*, 2006; Papanikolaou *et al.*, 2008; Papanikolaou *et al.*, 2009). The comparison of results is displayed in Figure 5. Our model

was able to reproduce accurately the experiments reported in the literature. However, the model predicted higher lipid concentrations when initial glucose concentration was high (150 g.L^{-1}). Additionally, the model was tested to represent the dynamics of a mutant strain of *Y. lipolytica* where results demonstrated the strong predictive capabilities of the model (See Supplementary Material D).

The proposed models captured the underlying dynamics of *Yarrowia lipolytica* keeping track of the variations in concentrations of two different types of cultures and two phenomena on the cell metabolism. The models were capable of identifying overflow metabolism, which depends on both the carbon and nitrogen source. Overflow is not occurring if extracellular nitrogen is not depleted or if the critical intracellular nitrogen quota is not reached. However, the level of nitrogen limitation directly affects lipid accumulation. Special care should be paid to the feeding profiles of carbon and nitrogen to have a proper value of N/C ratio allowing maximization of lipids. Therefore, a control strategy to define the optimal feeding rates is needed. In this context, the quota model was selected to evaluate a control strategy aiming at maximizing lipid accumulation by manipulating the feeding profiles of nitrogen and glucose. This choice was based on the lower *RMSE* values for glucose concentrations with the quota model for all the cultures, and the better description of extracellular nitrogen for Culture A, which are essential for regulating the N/C ratio.

3.3. Optimization of lipid accumulation

In fed-batch conditions, the optimization problem focusses on determining the feed rate policies for the substrates during the entire period of operation maximizing an objective function defined in terms of the status of the bioreactor. Until now, there has been a considerable amount of research about yeast cell culture optimization to avoid overflow metabolism (*e.g.* ethanol formation) (Renard and Vande Wouwer, 2008; Valentinotti *et al.*, 2003) while maximizing biomass. However, it has never been applied for other yeasts

experiencing overflow, such as *Yarrowia lipolytica*, where it is necessary to avoid citric acid formation to favor lipid accumulation. Hereof, maximizing lipid production could be achieved by a careful determination of the feeding profiles of glucose and nitrogen to maintain the quota q_N near to the identified value of the critical quota q_N^* . Optimizing lipid accumulation in yeast fed-batch cultures can be seen as a two-fold objective at a final fermentation time (t_f). The first objective is the maximization of the lipid productivity at a final time of operation,

$$P_L = \frac{X_L(t_f)}{t_f} \quad (20)$$

The second objective is the production of high lipid content biomass. In order to do it, let us consider the lipid content fraction of biomass as $w_L(t) = \frac{X_L}{X_T}$, and a target lipid content $w_{L,max}$

, which are put together into a new normalized variable P_W as,

$$P_W = \frac{w_L(t_f)}{w_{L,max}} \quad (21)$$

where the target lipid fraction was set to $w_{L,max} = 0.36$ (g.g⁻¹), which is the maximum theoretical value (Beopoulos *et al.*, 2009). The multi-objective problem is defined as,

$$\begin{aligned} & \max_{f_{S_{IN}}, f_{N_{IN}}, t_f} w_p \cdot P_L + P_W \\ \text{Subject to: } & \begin{cases} Eq (1-7) \\ Eq (14) \\ S(t_f) < S_{max}, N(t_f) < N_{max} \\ V_0 < V < V_{max}, t_{min} \leq t_f \leq t_{max}, \\ 0 \leq f_{S_{IN}} \leq f_{S_{IN,max}}, 0 \leq f_{N_{IN}} \leq f_{N_{IN,max}}, n_{PWL} = n_f \end{cases} \end{aligned} \quad (22)$$

The parameter w_p is a weight coefficient to favor a high lipid content and it was identified from constant flow rates and fixed to 0.25. The quota model was used to determine the optimal inputs and to predict the response of the state variables. Even if the model allows to expressing the accumulated lipids X_{LA} , the term of total lipids $X_L = X_{LS} + X_{LA}$ was preferred to keep

consistency with literature values. Additionally, the results of using X_{LA} or X_L were the same since the structural lipids $X_{LS} = \gamma.X_f$ are proportional to the functional biomass. Initial conditions of the state variables were taken from Culture A (Supplementary Material C). The final fermentation time was set between 20 and 90 h. The maximum allowed residual concentrations of glucose S_{\max} and nitrogen N_{\max} at the end of the culture were set to 2 g.L⁻¹, and 0.1 g.L⁻¹, respectively. The volume was bounded the initial value $V_0 = 8$ L to the maximum volume of the bioreactor $V_{\max} = 15$ L. The upper limits for flow rates of carbon $f_{S_{IN \max}}$ and nitrogen $f_{N_{IN \max}}$ sources were fixed to 0.3 L.h⁻¹. A number n_{PWL} of piecewise linear (PWL) functions were used to parameterize the control $f_{S_{IN}}$ and $f_{N_{IN}}$. The optimization was performed with the Pattern Search algorithm of MATLAB[®]. For the sake of comparison, the optimization problem was solved for two cases: (i) constant optimal flow rates along the complete culture ($n_f = 1$); and (ii) five piecewise linear functions ($n_f = 5$). The PWL functions were defined with open time intervals to optimize the time length t_f of each linear function between t_{\min} and t_{\max} for a given n_f .

Figure 6 shows the optimization results concerning the dynamics of the input variables to maximize the objective functions and the prediction of the state variables from the quota model. The constant input flow rates were: 0.162 L.h⁻¹ for glucose, and 0.021 L.h⁻¹ for nitrogen. The use of these constant optimal flow rates (dashed lines) led to a constant feeding with N/C ratio of 0.027 molN.Cmol⁻¹, which allowed the production of citric acid. Although the value was constant, nitrogen limitation occurred around 11.5 h, and was maintained until the end of the culture (30.3 h) where citric acid concentration achieved 1.28 g.L⁻¹. Concerning the optimization with 5 PWL functions (dark line), glucose feeding was increasing for 4 h, with a further small decrease of 13 h to assure growth and to moderate the concentration of residual glucose. After 17 h, the feeding profile increased again for 8 h to provide the necessary carbon

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for lipid accumulation. Finally, the glucose feeding decreased with two different slopes to fulfill the condition of residual glucose. Nitrogen feeding had a similar trend to glucose, where the second piecewise decreases the nitrogen feeding to induce nitrogen limitation, which is necessary for enhancing lipid accumulation. After 10 h the feeding rate was increased again at two different slopes for 19h to regulate the N/C ratio and to avoid citric acid production. The nitrogen flow dropped at the end of the culture to assure a low value of residual nitrogen. Concerning the quota, it is observed that for both cases the final values were around the critical quota q_N^* , which is reflected on the low citric acid concentrations.

The summary of the results of the control strategy are displayed on Table IV to clarify the enforcement of this strategy. Another case was also studied with two piecewise linear functions ($n_f = 2$) to analyze the impact of the quantity of PWL functions (See supplementary material B). Table IV shows that the three compared cases reported high biomass content, which is important for harvesting the cells at the end of the culture. The use of piecewise linear functions increased 1.1 times productivity and 1.04 times lipid content fraction with respect to an operation at constant flow rates. These results suggested that a constant flow rate could be used, but should be avoided if citric acid production is to be maintained at low values. The performance of 2 PWL functions was slightly better to the constant flow rates, which supports the use of a higher number of piecewise linear function for optimizing both objectives. Literature values of lipid content fraction and volumetric productivity were reported from 0.17 to 0.25 g.g⁻¹ and from 0.08 to 0.27 g.L⁻¹.h⁻¹, respectively (Aggelis and Komaitis, 1999; Beopoulos *et al.*, 2009; Cescut, 2009; Ochoa-Estopier, 2012). Even if our results are encouraging for being competitive with the literature results, the real implementation of the developed control strategy is needed to have a fair comparison. For that, the development of software sensors, which are challenging and time demanding, is required to reconstruct the

state variables of the model. This will allow the proper monitoring of system and the correction of model uncertainties.

4. CONCLUSIONS

An unstructured model based on Monod and inhibition kinetics and a model based on Droop quotas were developed to describe the accumulation of lipids and the production of citric acid on *Yarrowia lipolytica* on fed-batch and sequential batch cultures. The results of both models showed good agreement with the calibration and validation experimental data sets. Parameters from both models were consistent with literature values. Even if both models presented similar goodness-of-fit, the quota model was selected to develop a model based optimization of lipid accumulation due to a better description of residual glucose and extracellular nitrogen concentrations, which was important for the accurate manipulation of the N/C ratio. Two objectives were maximized: lipid productivity and lipid content fraction. Optimization results indicated that it would be possible to produce higher values than the literature proving the effectiveness of the control strategy. Further work need to deal with the experimental implementation to validate the simulation results of the presented control strategy.

SUPPLEMENTARY MATERIAL

- A.** Figures of the Internal Quota for the different cultures.
- B.** Information on optimization results.
- C.** Initial conditions of Experiments for calibration and validation.
- D.** Assessment of the predictive capabilities of the quota model to represent the dynamics of the mutant strain *Yarrowia lipolytica* JMY3501

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NOMENCLATURE

CIT	Citric acid concentration [g.L ⁻¹]
$f_{N_{IN}}$	Input flow rate of nitrogen [L.h ⁻¹]
$f_{N_{INmax}}$	Maximum input flow rate of nitrogen [L.h ⁻¹]
$f_{S_{IN}}$	Input flow rate of glucose [L.h ⁻¹]
$f_{S_{INmax}}$	Maximum input flow rate of glucose [L.h ⁻¹]
F_{IN}	Input flow rate [L.h ⁻¹]
F_{Sample}	Output flow rate [L.h ⁻¹]
GUR_X	Glucose uptake rate [gS.gX _f ⁻¹ h ⁻¹]
Ind_Q	Overflow indicator in Quota model [-]
Ind_U	Overflow indicator in Unstructured model [-]
$K_{S,i}$	Saturation constants of glucose [gS.L ⁻¹]
K_N	Saturation constant of N [gN.L ⁻¹]
k_i	Inhibition constants [g.L ⁻¹]
n	Number of data [-]
n_f	Number of Piecewise linear functions
N	Nitrogen concentration [g.L ⁻¹]
N_{max}	Maximum residual nitrogen concentration [g.L ⁻¹]
N_{IN}	Nitrogen input concentration [g.L ⁻¹]
Of_U	Overflow saturation constant [gS.gX _f ⁻¹ h ⁻¹]
P_L	Lipid productivity [g.(L-h) ⁻¹]
P_W	Normalized lipid fraction [g.g ⁻¹]
Q_0	Minimal quota for growth [gN.gX _T ⁻¹]

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q_N	Internal nitrogen quota [$gN.gX_T^{-1}$]
q_N^*	Critical quota for citric acid formation [$molN.Cmol^{-1}$]
S	Glucose concentration [$g.L^{-1}$]
S_{max}	Maximum residual glucose concentration [$g.L^{-1}$]
t_f	Final optimal time [h]
t_{min}	Minimum allowed time for the optimization [h]
t_{max}	Maximum allowed time for the optimization [h]
S_{IN}	Glucose input concentration [$g.L^{-1}$]
V	Volume [L]
V_0	Initial volume [L]
V_{max}	Maximal volume [L]
w_L	Lipid content fraction [$g.g^{-1}$]
$w_{L,max}$	Maximum theoretical value of lipid content fraction [$g.g^{-1}$]
w_p	weight coefficient to favor a high lipid fraction [-]
X_T	Total biomass concentration [$g.L^{-1}$]
X_f	Functional biomass concentration [$g.L^{-1}$]
X_L	Total Lipid biomass concentration [$g.L^{-1}$]
X_{LA}	Accumulated lipid biomass concentration (TAGs) [$g.L^{-1}$]
X_{LS}	Structural lipid biomass concentration [$g.L^{-1}$]
$Y_{X/S}$	Yield coefficient for functional biomass production with respect to glucose [$gX_f.gS^{-1}$]
$Y_{LIP/S}$	Yield coefficient for lipid production with respect to glucose [$gLIP.gS^{-1}$]
$Y_{CIT/S}$	Yield coefficient for citric acid production with respect to glucose [$gCIT.gS^{-1}$]

$Y_{X/N}$ Yield coefficient for functional biomass production with respect to nitrogen
[g X_f .g N^{-1}]

Abbreviations

PSO Particle Swarm Optimization

PWL Piecewise linear functions

R^2 Correlation Coefficient

RMSE Root Mean Square Error

TAG Triacylglycerol

Greek symbols

α maximum lipid content [g LIP .g X_T^{-1}]

γ constant fraction of the functional biomass [g X_L .g X_f^{-1}]

μ Growth rate [h $^{-1}$]

μ^{\max} Maximal growth rate on carbon and nitrogen [h $^{-1}$]

μ^* Theoretical maximal growth rate on carbon and nitrogen [h $^{-1}$]

π_{LIP} Specific rate of lipid accumulation [g LIP . g $X_f^{-1}h^{-1}$]

π_{LIP}^{\max} Maximal specific rate of lipid accumulation [g LIP .g $X_f^{-1}h^{-1}$]

π_{CIT} Specific rate of citric acid production [g CIT .(g X_f .h) $^{-1}$]

π_{CIT}^{\max} Maximal specific rate of citric acid production [g CIT . g $X_f^{-1}h^{-1}$]

ρ_N Specific uptake rate of nitrogen [g N . g $X_f^{-1}h^{-1}$]

ρ_N^{\max} Maximal specific uptake rate of nitrogen [g N . g $X_f^{-1}h^{-1}$]

Superscripts

max Maximum

Subscripts

i Identification of number of constants

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max Maximum

Operators

$\frac{d}{dt}$ Derivative of a variable with respect to time

max Maximization

Table I. Estimated parameters for the fed-batch cultures of *Yarrowia lipolytica*.

Parameter	This work		References	
	Unstruc	Quota		
μ^{\max} [h ⁻¹]	0.415	-	0.26	Culture A ^a
μ^* [h ⁻¹]	-	0.755	0.18-0.36	(Papanikolaou and Aggelis, 2003b) ^b
			0.566	(Economou <i>et al.</i> , 2011) ^d
π_{LIP}^{\max} [gLIP.(gX _f -h) ⁻¹]	0.030	0.039	0.034	Culture A ^a
			0.028	(Fakas <i>et al.</i> , 2009) ^c
			0.0027	(Papanikolaou <i>et al.</i> , 2006) ^b
π_{CIT}^{\max} [gCIT.(gX _f -h) ⁻¹]	0.301	0.314	0.035-0.13	(Papanikolaou and Aggelis, 2003a) ^b
			0.013	(Papanikolaou <i>et al.</i> , 2008) ^b
$Y_{X/S}$ [gX _f .gS ⁻¹]	0.603	0.519	0.78 – 0.86	(Papanikolaou and Aggelis, 2003b) ^b
$Y_{LIP/S}$ [gLIP.gS ⁻¹]	0.276	0.272	0.23	Culture A ^a
			0.63-0.86	(Papanikolaou and Aggelis, 2003b) ^b
$Y_{CIT/S}$ [gCIT.gS ⁻¹]	0.88	0.89	0.63	(Papanikolaou and Aggelis, 2003a) ^b
			0.45-0.82	(Papanikolaou <i>et al.</i> , 2008) ^b
			31.9-33.6	(Papanikolaou and Aggelis, 2003a) ^b
$Y_{X/N}$ [gX _f .gN ⁻¹]	23.23	-	39.3	(Fakas <i>et al.</i> , 2009) ^c
			18.21	(Economou <i>et al.</i> , 2011) ^d
$K_{S,1}$ [gS.L ⁻¹]	1.517	1.116	20	(Fakas <i>et al.</i> , 2009) ^c
			1.256	(Economou <i>et al.</i> , 2011) ^d
$K_{S,2}$ [gS.L ⁻¹]	0.978	1.482	69.27	(Economou <i>et al.</i> , 2011) ^d
$K_{S,3}$ [gS.L ⁻¹]	11.26	12.22	-	
			0.196	(Papanikolaou and Aggelis, 2003a) ^b
K_N [gN.L ⁻¹]	0.074	23.08	0.179	(Arzumanov <i>et al.</i> , 2000) ^b
			0.085	(Economou <i>et al.</i> , 2011) ^d
k_1 [gN.L ⁻¹]	924	-	835	(Economou <i>et al.</i> , 2011) ^d
k_2 [gCIT.L ⁻¹]	8.3	9.75	-	
k_3 [gCIT.L ⁻¹]	40.44	37.63	-	
$k_{I,1}$ [gS.L ⁻¹]	83.14	83.64	20	(Economou <i>et al.</i> , 2011) ^d
$k_{I,2}$ [gS.L ⁻¹]	35.2	44.17	0.399	(Economou <i>et al.</i> , 2011) ^d
$k_{I,3}$ [gS.L ⁻¹]	28.8	40.11		
Overflow:				
Of_U [gS.(gX _f -h) ⁻¹]	0.268	-	-	
Quotas:				
ρ_N^{\max} [gN.(gX _f -h) ⁻¹]	-	0.716	-	
Q_0 [gN.(gX _T) ⁻¹]	-	0.045	-	
q_N^* [molN.Cmol ⁻¹]	-	0.084	-	

^aExperimental values of Culture A with *Yarrowia lipolytica*. ^b*Yarrowia lipolytica*. ^c*Thamnidium elegans*. ^d*Mortierella isabellina*

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Table II. Statistical evaluation of the models on the calibration data sets.

Model	Criterion	Culture A				
		S	N	CIT	X_f	X_L
Unstructured Model	R^2	0.845	0.027	0.987	0.963	0.966
	RMSE	2.195	0.944	5.236	8.722	1.142
Quota Model	R^2	0.811	0.307	0.992	0.963	0.958
	RMSE	2.359	0.341	6.754	7.845	1.245
Culture B ($t=0 - 38 h$)						
Unstructured Model	R^2	0.916	NA	0.984	0.957	0.835
	RMSE	9.433		4.091	3.044	1.063
Quota Model	R^2	0.904		0.995	0.966	0.836
	RMSE	8.398		2.472	3.182	1.144

NA: Not available

Table III. Statistical evaluation of the models on the validation data set.

Model	Criterion	Culture B ($t=38 - 80 h$)				
		S	N	CIT	X_f	X_L
Unstructured Model	R^2	0.392	NA	0.989	0.622	0.363
	RMSE	10.293		7.731	10.368	1.041
Quota Model	R^2	0.479		0.991	0.476	0.361
	RMSE	9.617		9.439	9.836	0.985
Culture C						
Unstructured Model	R^2	0.645	NA	0.854	0.973	0.918
	RMSE	23.285		2.855	7.303	0.575
Quota Model	R^2	0.719		0.894	0.949	0.902
	RMSE	20.545		3.174	7.695	0.884

NA: Not Available

Table IV. Performance of the control strategy.

	t_f (h)	$V(t_f)$ (L)	$CIT(t_f)$ (g.L ⁻¹)	$X_L(t_f)$ (g.L ⁻¹)	$X_f(t_f)$ (g.L ⁻¹)	$P_L(t_f)$ (g.L ⁻¹ .h ⁻¹)	$w_L(t_f)$ (g.g ⁻¹)	$w_P \cdot P_L + P_W$
<i>Constant flow rate</i>	30.64	13.91	1.266	26.41	100.22	0.861	0.218	0.821
<i>2 PWL functions</i>	30.15	14.46	0	27.78	105.59	0.921	0.218	0.836
<i>5 PWL functions</i>	31.62	14.91	0.098	29.98	108.31	0.948	0.227	0.867

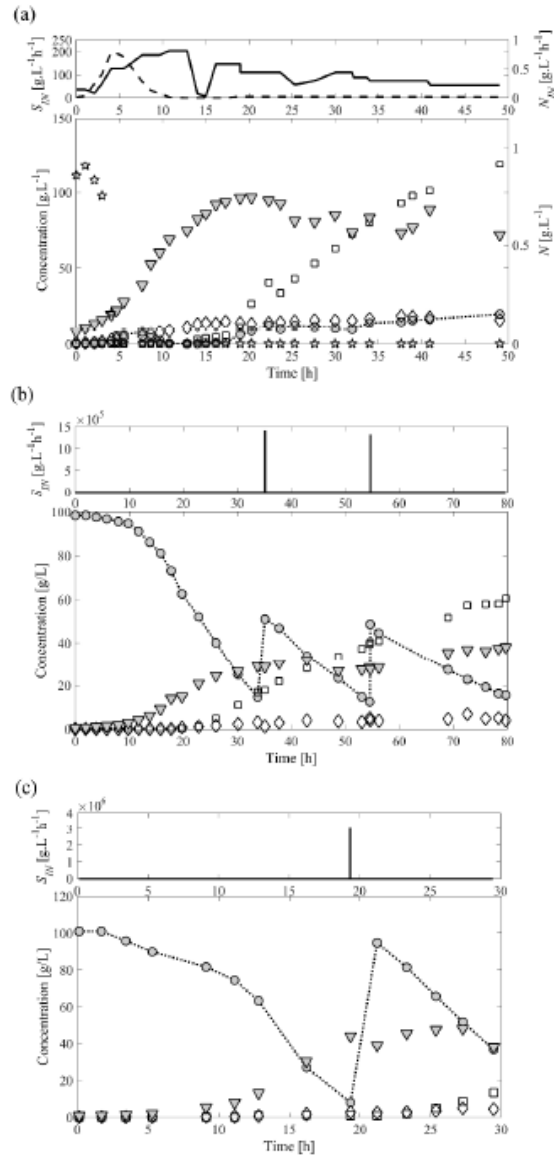


Figure 1. Experimental data (a) Culture A: Fed-Batch culture (Cescut, 2009). (b) Culture B: Sequential-batch culture. (c) Culture C: Sequential-batch culture for model validation. The solid line (-) represents the input flow rate of glucose and (- -) the input flow rate of ammonia. (-●-) Glucose, (□) Citrate, (◇) Lipids, (▼) Biomass, (☆) Nitrogen.

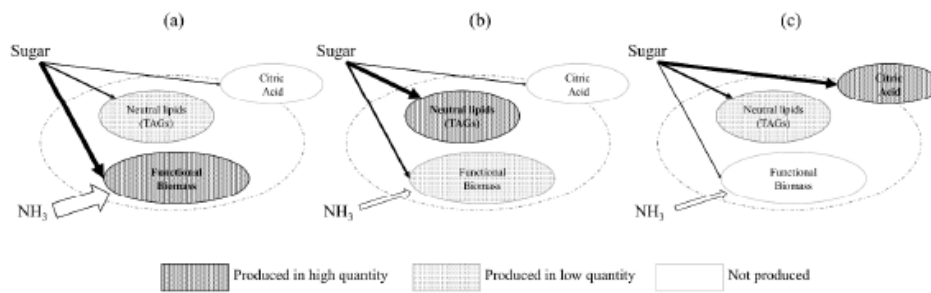


Figure 2. Distribution of carbon flux in the metabolism of *Yarrowia lipolytica* for different physiological states: (a) Production of functional biomass. (b) Lipid accumulation and growth. (c) Excretion of citric acid.

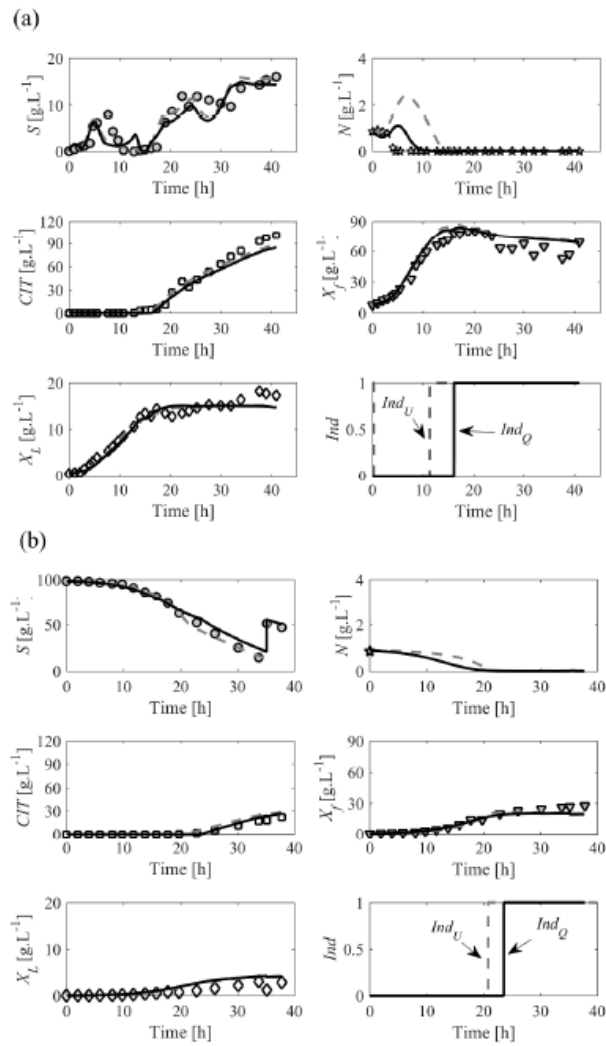


Figure 3. Model calibration: Performance of the two models with the experimental data of (a) Culture A and (b) Culture B ($t = 0 - 38h$). (---) Unstructured model. (—) Quota model.

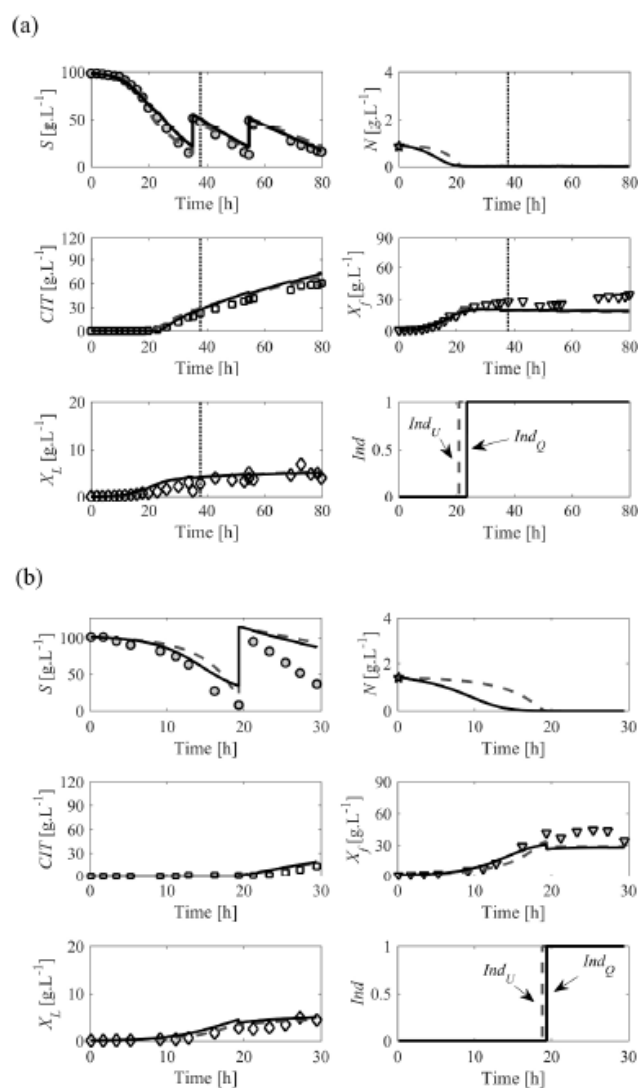


Figure 4. Model validation: Prediction of the models with the experimental data of sequential batch cultures under nitrogen deficiency. (a) Culture B (t = 38 - 80h). (b) Culture C. (---) Unstructured model. (—) Quota model

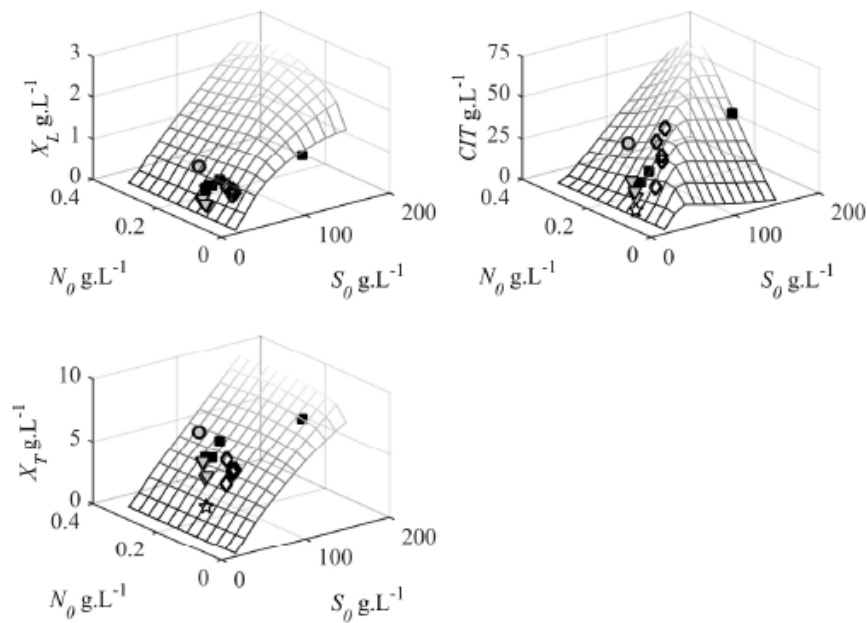


Figure 5. Model validation with different experimental data from the literature in Batch conditions with glucose as substrate. (■) (Papanikolaou et al., 2006). (●) (Papanikolaou et al., 2008) (☆) (Kavšček et al., 2015). (▼) C/N=100 (Papanikolaou et al., 2009). (◇) C/N=200 (Papanikolaou et al., 2009).

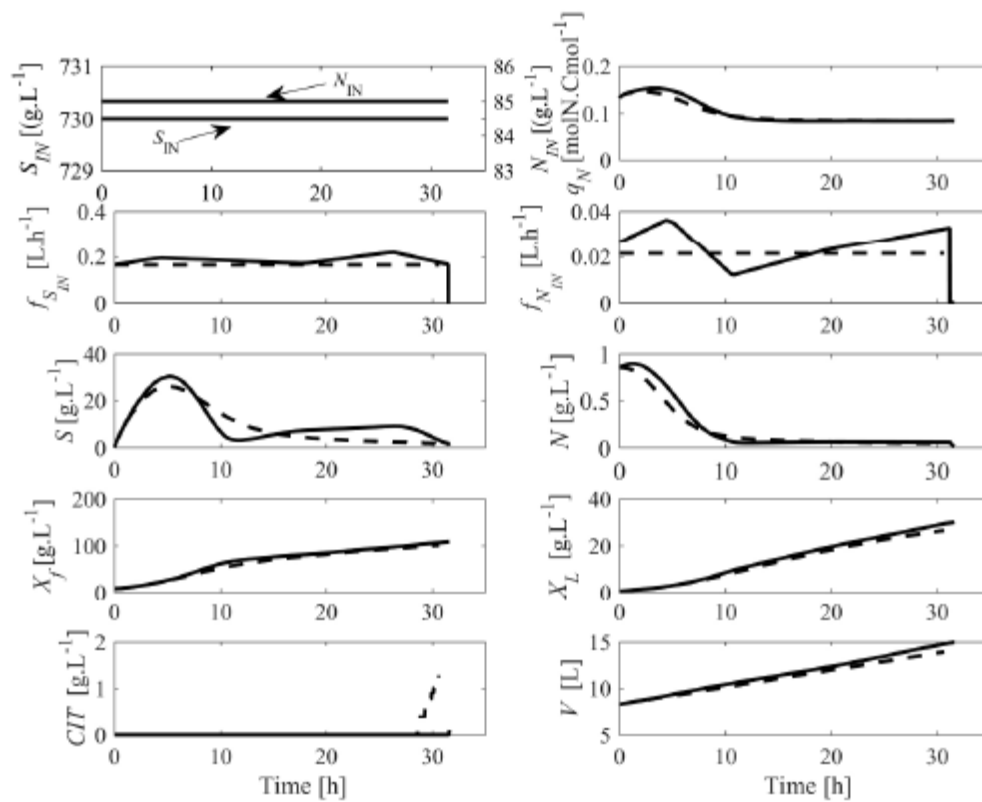


Figure 6. Results of the control strategy. (—) Constant flow rates (—) 5 PWL.