

Patterns of diversity, endemism and specialization in the root symbiont communities of alder species on the island of Corsica

Adrien C Pozzi, Mélanie Roy, Mélissande Nagati, Guillaume P Schwob, Sophie Manzi, Monique Gardes, Pierre-Arthur Moreau, Maria P. Fernandez

▶ To cite this version:

Adrien C Pozzi, Mélanie Roy, Mélissande Nagati, Guillaume P Schwob, Sophie Manzi, et al.. Patterns of diversity, endemism and specialization in the root symbiont communities of alder species on the island of Corsica. New Phytologist, In press, 219 (1), pp.336-349. 10.1111/nph.14996 . hal-01698095

HAL Id: hal-01698095 https://hal.science/hal-01698095v1

Submitted on 31 Jan 2018 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Early View. The definitive version is available at http://onlinelibrary.wiley.com/journal/10.1111/(ISSN)1469-8137.

Patterns of diversity, endemism and specialization in the root symbiont communities of alder species on the island of Corsica

Adrien C. Pozzi^{1,2}* (**b**), Mélanie Roy³*, Mélissande Nagati³, Guillaume Schwob¹, Sophie Manzi³, Monique Gardes³, Pierre-Arthur Moreau⁴ and Maria P. Fernandez¹

¹Laboratoire d'Ecologie Microbienne, UMR5557, CNRS, INRA, VetAgro Sup, UCBL, Université de Lyon, Villeurbanne 69622, France; ²Biométrie et Biologie Evolutive, UMR5558, CNRS, INRIA, VetAgro Sup, HCL, UCBL, Université de Lyon, Villeurbanne 69622, France; ³Laboratoire Evolution et Diversité Biologique, UMR5174, Université Paul Sabatier – CNRS, 118 route de Narbonne, Toulouse Cedex F-31062, France; ⁴Laboratoire IMPECS EA 4483, Fac. Pharma. Lille, Université de Lille, Lille F-59000, France

Authors for correspondence: Adrien C. Pozzi Tel: +33 472 448 200 Email: adrien.c.pozzi@gmail.com

Mélanie Roy Tel: +33 561 556 420 Email: melanie.roy@univ-tlse3.fr

Received: *17 June 2017* Accepted: *7 December 2017*

New Phytologist (2018) **doi**: 10.1111/nph.14996

Key words: Alnus (alders), community ecology, Corsica, diversity, ectomycorrhiza (EM), endemism, *Frankia*, specialization.

Summary

• We investigated whether the diversity, endemicity and specificity of alder symbionts could be changed by isolation in a Mediterranean glacial refugium. We studied both ectomycorrhizal (EM) fungi and nitrogen-fixing actinobacteria associated with alders, and compared their communities in Corsica and on the European continent.

• Nodules and root tips were sampled on the three alder species present in Corsica and continental France and Italy. Phylogenies based on internal transcribed spacer (ITS) and a multilocus sequence analysis approach were used to characterize fungal and *Frankia* species, respectively. Patterns of diversity, endemism and specialization were compared between hosts and regions for each symbiont community.

• In Corsica, communities were not generally richer than on the mainland. The species richness per site depended mainly on host identity: *Alnus glutinosa* and *Alnus cordata* hosted richer *Frankia* and EM communities, respectively. Half of the *Frankia* species were endemic to Corsica against only 4% of EM species.

• Corsica is not a hotspot of diversity for all alder symbionts but sustains an increased frequency of poor-dispersers such as hypogeous fungi. Generalist EM fungi and host-dependent profusely sporulating (Sp+) *Frankia* were abundantly associated with Corsican *A. cordata*, a pattern related to a more thermophilic and xerophylic climate and to the co-occurrence with other host trees.

Introduction

Alders are involved in a tripartite symbiosis with ectomycorrhizal (EM) fungi and nitrogen-fixing actinobacteria belonging to the genus Frankia. Compared to other trees, alders associate with few EM genera, some of them being strictly or mainly specific (Molina, 1981; Kennedy et al., 2015). Similarly, alders associate with a specific clade of Frankia whose strains are shared with a limited number of other actinorhizal host plants (Normand et al., 1996; Dawson, 2007). Among alders, the two subgenera Alnus and Alnobetula associate mainly with distinct EM (Rochet et al., 2011; Roy et al., 2013) and Frankia species (Pozzi et al., 2015; Cotin-Galvan et al., 2016). Few EM or Frankia species associate strictly with a single alder species and rather the host species influences the composition of its symbiont community (Lipus & Kennedy, 2011; Põlme et al., 2013; Roy et al., 2013). The reasons behind alder specificity have been intensely studied and tested, especially for EM fungi. Studies on European alders have demonstrated that the specificity pattern resulted from several events of host-tracking and host shifts (Rochet *et al.*, 2011). Other studies also demonstrated that alder phylogeny was strongly correlated with EM and *Frankia* community structures at a global scale (Põlme *et al.*, 2013, 2014). Apart from the host plant, other factors such as soil parameters, altitude and climate may also select adapted symbionts and explain some of alder's specificity. For instance, EM associated with alders are able to grow in high nitrate and low pH conditions, while other EM fungi show limited growth in these conditions (Huggins *et al.*, 2014). Soil type, acidity and moisture also shape part of *Frankia* diversity (Schwintzer, 1990; Markham & Chanway, 1998). Finally, co-migration or co-invasion events have probably maintained this specificity, as suggested for EM fungi in North America and New Zealand, and for *Frankia* globally (Kennedy *et al.*, 2011; Põlme *et al.*, 2014; Bogar *et al.*, 2015).

Several exceptions to the specificity pattern between alders and their symbionts have been identified in recent years. Alders can also associate with very generalist EM taxa that are often scarce but always present (Roy *et al.*, 2013), such as *Cenococcum geophilum* (Põlme *et al.*, 2013; Roy *et al.*, 2017). The study of co-occurring seedlings of *Alnus rhombifolia* and

^{*}These authors contributed equally to this work.

Betula occidentalis also revealed a few shared EM fungi, including Alnicola sp., and a significant effect of neighboring plants (Bogar & Kennedy, 2013). Similarly, the study of isolated A. glutinosa subsp. barbata populations in Georgia, a Tertiary and glacial refugium, revealed nonspecific species of Inocybe and Tuber spp. (Roy et al., 2017). However, the detection of EM taxa shared with other host species remains rare, and is mainly limited to cases where alders are isolated or planted outside of their natural distribution range. These observations suggest that isolation or colonization of a new habitat may relax alder specificity and select new associations. Compared with EM fungi, no generalist Frankia has been described so far, as Frankia taxa associated with alders have never been detected on other host trees in the wild. The number of genera of actinorhizal plants able to associate with Frankia is also far lower and less diverse than genera of host plants associated with EM fungi. Thus, the co-occurrence of alders with another actinorhizal host is limited, although host-shifts of Alnusinfective Frankia have been described experimentally (Bosco et al., 1992; Lumini et al., 1996). As a consequence, isolated alder stands do not recruit generalist Frankia strains, as shown in Mexico and Georgia, where unique Frankia species have been detected (Higgins & Kennedy, 2012; Roy et al., 2017). Moreover, Frankia sequences from Georgia were phylogenetically distant from other Frankia sampled worldwide. These observations suggest that refugia, or at least isolated populations, might host endemic Frankia species and show a strong biogeographic pattern of distribution, as shown for bacteria specialized toward a specific niche (Angel et al., 2010; Brown et al., 2012). Conversely, EM fungal species have a broader distribution, with only a few exceptions reported for Corsica (Rochet et al., 2011).

The biogeographic pattern of symbiotic fungi and bacteria may also depend on their biology. Indeed, the ability to migrate with or without the host is impacted by life-history traits, particularly dispersal and host-dependency. Sporulation is a major component of microbe dispersal (Huang & Hull, 2017). Both EM fungi and Frankia are able to sporulate, yet with major differences. Among EM fungi, hypogeous taxa such as Alpova or Melanogaster sp. can be dispersed by small mammals and are less likely to migrate than wind-dispersed epigeous fungi. Because actinorhizal nodules closely resemble hypogeous structures, Frankia would also have more limited dispersal abilities as compared to epigeous fungi. Among Frankia, dispersal may also depend on contrasting sporulation traits. Indeed, some strains sporulate profusely within nodular cells (Sp+), while other strains do not (Sp-). Host-dependency also differs: EM fungi and Sp+ Frankia are considered to be obligate symbionts, while Sp- Frankia strains can be cultured and live as saprobes in soil (Schwintzer, 1990; Pozzi et al., 2015; Cotin-Galvan et al., 2016). These life history traits could lead to distinct symbiont distributions and impact alder symbiont communities, particularly where alders have been isolated, such as in glacial refugia or on islands.

In Europe, pollen records and phylogeographic studies have revealed several southern glacial refugia for alders, such as the

Iberian peninsula, the Apennine mountains, Corsica (an island), North Africa, and the Balkan and Anatolian Peninsulas (King & Ferris, 1998; Douda et al., 2014; Havrdová et al., 2015; Mandák et al., 2016), together with a few northern refugia (Douda et al., 2014; Mandák et al., 2016). Corsica was isolated from the continent (c. 80 km) 20-23 Myr ago (Ma) (Blondel & Aronson, 1999) and is known as a hotspot of plant diversity and endemism (Medail & Quezel, 1997), where boreo-arctic plants took refuge from the Tertiary glaciations and now cooccur with Mediterranean and temperate elements (Jeanmonod & Gamisans, 2007). Three alder species are present in Corsica and show distinct degrees of isolation from the mainland. Alnus cordata (Loisel.) Duby is native both to Corsica and southern Italy, where it grows from 150 to 2000 m (Gamisans, 1999). Alnus glutinosa (L.) Gaertn is a widespread Eurasian species and has colonized Corsican riparian forests, marshes and peat bogs from sea level up to 1300 m (Gamisans, 1999). Alnus glutinosa and A. cordata can co-occur and their distribution ranges partly overlap. Finally, the subalpine shrub Alnus alnobetula subsp. suaveolens (Req.) Lambinon & Kerguélen is endemic to Corsica, and diverged from the continental alpine subspecies A. alnobetula subsp. alnobetula (Ehrh.) K. Koch c. 1.1 Ma (Rochet et al., 2011). These features invite study of alder symbiont communities in Corsica, and we investigated if they are particularly diverse and possibly endemic, and if host shifts could have occurred between the three alder species. No reports were available for Frankia. Previous studies on EM fungi of alder that partly investigated Corsican populations revealed only two endemic EM species, Alpova corsicus and Melanogaster rivularis, and highlighted the heterogeneity of EM communities and the abundance of generalist taxa such as ascomycetes on A. cordata roots (Moreau et al., 2011; Roy et al., 2013). In Corsica, A. cordata forms very typical mixed woods with Castanea sativa at mid-altitudes (Gamisans, 1999), while it forms rather monodominant stands along the Italian coast (Ducci & Tani, 2009). While mixed woods may affect the symbiont communities of alders in Corsica, no particular link was detected between C. sativa and A. cordata EM communities, probably due to the limited number of sites investigated (Taudiere et al., 2015). This study also suggested the possible sharing of EM fungi between the three alder hosts. However, this situation was not compared with the mainland.

The aim of this study was to investigate the diversity, endemism and specialization patterns of alder symbionts in Corsica. We sampled both *Frankia* and EM fungi on the three alder species and compared Corsican communities with communities sampled from the continent, in France and Italy. To discuss species distributions for the two symbionts, we chose the same evolutionary species concept and used the most appropriate, yet different, molecular markers to delineate species for each symbiont. We hypothesized that alder symbiont communities in Corsica might be more species-rich and include more endemics, and that co-occurring alder or other host tree species may favor symbiont sharing. These questions will be discussed in the context of host history and current distribution, and of symbiont life history traits.

Materials and Methods

Sampled sites

Seventy sampling sites were investigated on Corsica and in continental France and Italy, from lowland to subalpine habitats (Fig. 1; Supporting Information Table S1). Sampling was carried out in November 2011 in Corsica and in May 2012 in Italy. Data from sites previously studied were included as the sampling protocol was identical (Roy *et al.*, 2013; Pozzi *et al.*, 2015). At each site, one or two species of alder were present, either monodominant or scattered in mixed forests. Relevant climatic data were retrieved from the Worldclim database (http://worldclim.org) using the RASTER package in R (Hijmans & van Etten, 2012).

Symbiont and soil sampling

For each site, four to six trees – separated from each other by at least 10 m – were sampled from two opposing sampling points. Approximately 50 cm of roots harboring both EM root tips and nodules were collected per tree. For the most recent sampling sites (Table S2) a soil core of 3 cm diameter and 10 cm depth was driven into the soil close to the tree roots. Roots, nodules and soils were kept at 4°C until processing in the lab. Soils from the same sampling site were pooled, sifted at 2 mm and dried at 60°C before physiochemical analysis. Soil analyses were provided by ECOLAB (Toulouse, France) or retrieved from previous studies (Rochet *et al.*, 2011; Roy *et al.*, 2013). Approximately 16 EM root tips and four nodule lobes per tree were preserved, as previously described (Roy *et al.*, 2013; Pozzi *et al.*, 2015).

Molecular biology and sequencing of symbionts

For fungi, DNA extraction was performed on each EM root tip using Promega's Wizard genomic DNA purification kit, following the manufacturer's instructions. The fungal internal transcribed spacer (ITS) region was amplified as in Rochet et al. (2011). Putative new symbionts for alders were confirmed by sequencing the plant ITS to check the host identity. Sequencing of the amplification product was done by MilleGen (Labège, France). Sequences were checked and cleaned using GENEIOUS 6.1.8 (Kearse et al., 2012). All new fungal sequences produced in this study were deposited with GenBank under accession numbers KU924024-KU925159 and KY131803 (Table S3). For Frankia, DNA extraction was performed on each lobe individually, as previously described (Pozzi et al., 2015). For the three housekeeping genes, pgk, dnaA and ftsZ, portions were amplified and sequenced with specific primers following Pozzi et al. (2015). Sequencing of the amplification product was done by Biofidal-DTAMB (Villeurbanne, France). Sequence quality was verified using 4PEAKS v.1.7.2 (Griekspoor & Groothuis, 2007). All Frankia sequences were deposited with EMBL under accession numbers LT555594-LT555681 (dnaA), LT555682-LT555771 (ftsZ) and LT555772–LT555865 (pgk).

Identification of symbionts

We chose the same evolutionary species concept for both symbionts where species are sequence-based monophyletic clades of strains sharing a common ancestor. The molecular markers used to call operational taxonomic units (OTUs) were chosen based on their accuracy, reliability and appropriateness for each symbiont and were thus different. Yet, all the OTUs were delineated at the same taxonomic rank – namely species – enabling a valid comparison between EM and *Frankia* species. Sequences were assigned to a Linnaean name after phylogenetic analyses. They were designated by an arbitrary numbering system when several unnamed species were characterized in the same genus, particularly within the genus *Frankia* for which only a small portion of the existing genomospecies has been named.

For fungi, using BLAST on GenBank and UNITE, it was possible to assign sequences to putative genera and to UNITE species hypotheses (Table S3). The closest sequences (97-100% identical) and some more distant sequences (with a reliable Linnaean name) were used for the alignment. Alnicola badiofusca and A. umbrina, two species supported by morphology and ecology but not differentiated by ITS (Moreau et al., 2013), were treated as a species complex and discarded from the analyses. Alignments were made using MAFFT (Katoh & Standley, 2013). Phylogenies of each EM genus were constructed using maximum likelihood analysis under a GTR+G model of evolution in RAxML (Stamatakis et al., 2008). The robustness of phylogenies was tested by rapid bootstrapping (1000 pseudoreplicates) on the CIPRES Science Gateway (Miller et al., 2010). We compared the ITS sequences with data obtained from sporocarps and the phylogenies developed for several genera associated with alders (Rochet et al., 2011), as recommended to delineate accurate fungal species (Taylor et al., 2000) corresponding to unified species (De Queiroz, 2007). For Tomentella spp., sequences associated with a Linnean name were often missing, and OTUs were delineated as monophyletic clusters of sequences, supported by a > 80% bootstrap value and sharing ITS sequences of > 97% similarity.

For Frankia, we used multilocus sequence analysis (MLSA) widely recommended to delineate prokaryotic species (Gevers et al., 2005). Housekeeping genes were chosen as, for Frankia, they better correlate with phylogenetic and whole-genome data (Sen et al., 2014; A. C. Pozzi et al., unpublished). Amino acid sequences of the three loci were aligned separately with MUSCLE v.3.8.31 (Edgar, 2004) and concatenated as nucleotide sequences in SEAVIEW v.4.4.2 (Gouy et al., 2010) for a subset of 94 nodules including 15 reference strains. The Bayesian inference of phylogeny was computed as in Pozzi et al. (2015) on the CIPRES Science Gateway (Miller et al., 2010). OTU delineation was based on an enlarged dataset (one to three loci for 224 root nodules, Table S4) and a similar phylogenetic analysis, enabling a comparison of new sequences with lineages described worldwide (Pozzi et al., 2015). We computed a histogram of pairwise phylogenetic distances (Fig. S1) and determined the threshold of 0.05 substitutions per site as the best barcode gap value (Kekkonen & Hebert, 2014) to delimit OTUs corresponding to unified species (De Queiroz, 2007). An OTU cladogram was also constructed from the phylogenetic distances.

Detection of spatial autocorrelation and geographic structure

4 Research

To test whether symbiotic communities were spatially structured, a multivariate spatial autocorrelation test was performed (SPDEP package; Smouse & Peakall, 1999). Isolation by distance was tested through correlation between geographic distances (calculated from GPS coordinates; see Fig. 1 and Table S1) and Bray–Curtis distances through a Mantel test (Mantel, 1967).

Host and regional effects on alpha, beta and gamma diversities

For all diversity indices, we tested the influence of (1) host identity, with two factors: host subgenus and host species nested in subgenus, and (2) region, with two factors: Continent vs Corsica as the first regional level, and the difference between France and Italy nested in Continent as a second regional level. To compare the gamma (total) diversity between host and regions, species accumulation curves were computed. To estimate alpha diversity, species richness (number of species per site) and Shannon diversity indices were computed (diversity function of the VEGAN package in R; Dixon, 2003). The species richness was square-root-



Fig. 1 Map of sampled populations. (a) Continental alder populations sampled in France and Italy. (b) Alder populations sampled in Corsica. (c) Elevation of sampled population (in metres above sea level (m asl)) for the three sampled alder species and number of sampled populations per symbiont (Continent|Corsica). (a–c) Open symbols, Continental populations; closed symbols, Consican populations. Alder species: *Alnus glutinosa* (AG, squares); *A. cordata* (AC, diamonds); *A. alnobetula* (AA, triangles). EM fungi, ectomycorrhizal fungi. Population numbers refer to Supporting Information Tables S1 and S2.

transformed to meet the assumption of homoscedasticity. An analysis of variance (ANOVA) was used to test the effect of all factors on alpha diversity indices (Table 1). To estimate beta diversity (dissimilarity between communities), Bray-Curtis distances were calculated per site (Bray & Curtis, 1957). Variance analysis of beta diversity was performed through a multivariate permutation test (Adonis, VEGAN package in R; Table 2). A subsequent test was performed to check if beta diversity variance was homogeneous between hosts or regions (betadisper test followed by an ANOVA). The effect of host species and regional factors on site ordination was illustrated using nonmetric multidimensional scaling (NMDS, meta-MDS function, VEGAN package). The correlation between NMDS, host and regional factors as well as geographic and climatic variables was tested through a permutation analysis (envfit function, VEGAN package in R). Only significant factors or variables (P > 0.05) were represented.

Species distribution and detection of symbiont endemism and specificity

To illustrate the distribution of shared symbiont species between regions and hosts in our dataset, and detect putative endemic or specific species, the abundance of *Frankia* and EM fungi was calculated per region (Corsica vs Continent) and per host species to build a Venn diagram and hierarchical clustering, ranked by Bray–Curtis distances (Bray & Curtis, 1957).

Phylogenetic approaches and taxonomic knowledge helped to detect true endemics and determine the specificity level of each

Symbiont

ΕM

Diversity index

Species richness

(square root)

Shannon

symbiont species. Classes of specialization were established as follows: (I) species-specialist: 100% of the species records found with a single Alnus species (A. alnobetula, A. cordata or A. glutinosa); (II) subgenus-specialist: 100% of the records found within the subgenus Alnus (i.e. A. cordata and A. glutinosa in our dataset, and mainly A. incana (L.) Moench in additional data); (III) alderspecialist: 100% of the records found with Alnus spp. with the exclusion of other trees and environmental samples without alders; (IV) generalist: at least one reliable record of species not related to alders; and (V) undetermined. Classes I, II and III were common to EM fungi and Frankia, the others only to fungi. To compare the respective specificity patterns of EM fungi and Frankia, the species abundance matrix was recoded with the five classes of specialization for a restricted number of sites where both symbionts were sampled (Table S5). Site ordination (NMDS) and correlation between NMDS, host and regional factors as well as geographic and climatic variables were tested as described above. Four soil variables were also tested (KCl pH, %N, %C and C : N ratio).

Distribution of short-distance dispersal symbionts

Sum of

squares

0.148

0.523

1.747

6.448

19.488

0.0331

0.0386

df

1

1

1

1

39

1

1

Host subgenus

Region, level 1

Region, level 2

Host subgenus

Host species

Host species

Residuals

The effects of short-distance dispersal of Sp+ *Frankia* and hypogeous fungi were tested between Corsica and the Continent. For *Frankia*, the *in planta* sporulation phenotype was determined microscopically on stained nodule sections (Pozzi *et al.*, 2015). A nodule was considered Sp+ when at least one sporangium was observed out of 50 infected plant cells (Schwintzer, 1990). The proportion of Sp+ nodules was calculated per host and per

F-statistics P-value

0.589109

0.312620

0.069050

0.000907

0.4230

0.3875

0.297

1.046

3.496

0.656

0.764

12.904

	Region, level 1	1	0.1717	3.401	0.0728		
	Region, level 2	1	1.3532	26.806	7.14e-6	***	
	Residuals	39	1.9688				
Species richness	Host subgenus	1	0.488	2.956	0.0937		
(square root)	Host species	1	0.086	0.519	0.4757	ns	
	Region, level 1	1	0.178	1.080	0.3053	ns	
	Region, level 2	1	0.641	3.881	0.0562		
	Residuals	38	6.275				
Shannon	Host subgenus	1	0.653	2.939	0.0946		
	Host species	1	0.000	0.000	0.99446	ns	
	Region, level 1	1	0.095	0.429	0.5167	ns	
	Region, level 2	1	1.255	5.652	0.0226	*	
	Residuals	38	8.436				
of variance (ANOVA each symbiont speci <i>Frankia</i> alone (22 4 utlier populations w	() was performed es pool. Symbiont 3). For EM and Fr vere removed, res	on the (spec <i>ankia</i> , pectiv	e extrapola cies popula , two (nos. ely. df, deg	ted species tions): ector 43 and 59) grees of free	richness and mycorrhiza (E) and four (no edom. Bioregi	Shannon M) alone s. 14, 15, on factor	24 is
	Species richness (square root) Shannon of variance (ANOVA each symbiont speci <i>Frankia</i> alone (22 4 utlier populations w	Region, level 1 Region, level 2 Residuals Species richness (square root) Host subgenus Host subgenus Host subgenus Host subgenus Host subgenus Host subgenus Host subgenus Host species Region, level 2 Residuals Shannon Host subgenus Host species Region, level 1 Region, level 2 Residuals Shannon Host subgenus Host species Region, level 1 Region, level 2 Residuals	Region, level 1 1 Region, level 2 1 Residuals 39 Species richness Host subgenus 1 (square root) Host species 1 Region, level 1 1 Region, level 2 1 Residuals 38 Shannon Host subgenus 1 Host species 1 Region, level 2 1 Residuals 38 Shannon Host subgenus 1 Host species 1 Region, level 2 1 Residuals 38 Shannon Species 1 Region, level 2 1 Residuals 38 Shannon Lost subgenus 1 Host species 1 Region, level 2 1 Residuals 38	Region, level 1 1 0.1717 Region, level 2 1 1.3532 Residuals 39 1.9688 Species richness Host subgenus 1 0.488 (square root) Host species 1 0.086 Region, level 1 1 0.178 Region, level 2 1 0.641 Residuals 38 6.275 Shannon Host subgenus 1 0.653 Host species 1 0.000 Region, level 1 1 0.095 Region, level 2 1 1.255 Residuals 38 8.436 of variance (ANOVA) was performed on the extrapola tach symbiont species pool. Symbiont (species popular Frankia alone (22 43). For EM and Frankia, two (nos. utlier populations were removed, respectively. df, deg	Region, level 1 1 0.1717 3.401 Region, level 2 1 1.3532 26.806 Residuals 39 1.9688 Species richness Host subgenus 1 0.488 2.956 (square root) Host species 1 0.086 0.519 Region, level 1 1 0.178 1.080 Region, level 2 1 0.641 3.881 Residuals 38 6.275 Shannon Host subgenus 1 0.653 2.939 Host subgenus 1 0.095 0.429 Region, level 1 1 0.095 0.429 Region, level 2 1 1.255 5.652 Residuals 38 8.436	Region, level 1 1 0.1717 3.401 0.0728 Region, level 2 1 1.3532 26.806 7.14e-6 Residuals 39 1.9688 7.14e-6 Species richness Host subgenus 1 0.488 2.956 0.0937 (square root) Host species 1 0.078 1.080 0.3053 Region, level 1 1 0.178 1.080 0.3053 Region, level 2 1 0.641 3.881 0.0562 Residuals 38 6.275 5 Shannon Host subgenus 1 0.653 2.939 0.0946 Host species 1 0.000 0.000 0.99446 Region, level 1 1 0.095 0.429 0.5167 Region, level 2 1 1.255 5.652 0.0226 Residuals 38 8.436 0.429 0.5167 Region, level 2 1 1.255 5.652 0.0226 Residuals 3	Region, level 1 1 0.1717 3.401 0.0728 . Region, level 2 1 1.3532 26.806 7.14e-6 *** Residuals 39 1.9688 . . . Species richness Host subgenus 1 0.488 2.956 0.0937 . (square root) Host species 1 0.086 0.519 0.4757 ns Region, level 1 1 0.178 1.080 0.3053 ns Region, level 2 1 0.641 3.881 0.0562 . Residuals 38 6.275 . . . Shannon Host subgenus 1 0.653 2.939 0.0946 . Host species 1 0.095 0.429 0.5167 ns Region, level 1 1 0.095 0.429 0.5167 ns Region, level 2 1 1.255 5.652 0.0226 * Residuals 38 8.436 . . . of variance (ANOVA) was performed on the extrapo

Alnus). Significance: ns, not significant; ., P<0.1; *, P<0.05; **, P<0.01; ***, P<0.001.

Table 1 ANOVA on alpha-diversity indices

Signifiance

ns

ns

ns

ns

ns

Symbiont	Factor tested	df	Sum of squares	F-statistic	R ²	P-value	Signifiance
EM	Host subgenus	1	1.1119	3.4300	0.07055	0.001	***
	Host species	1	1.1409	3.5195	0.07239	0.001	***
	Region, level 1	1	0.3573	1.1024	0.02267	0.331	ns
	Region, level 2	1	0.5083	1.5682	0.03225	0.053	
	Residuals	39	12.6423		0.80214		
Frankia	Host subgenus	1	3.3736	13.5854	0.21078	0.001	***
	Host species	1	1.8202	7.3301	0.11373	0.001	***
	Region, level 1	1	0.6829	2.7499	0.04267	0.016	*
	Region, level 2	1	0.6920	2.7867	0.04324	0.011	*
	Residuals	38	9.4362		0.58958		

Multivariate permutation test was performed on dissimilarities of Bray–Curtis distance matrices for each of the symbiont communities. Symbiont (species|populations): ectomycorrhiza alone (110|44), *Frankia* alone (22|43). Symbiont: for EM and *Frankia*, two (nos. 43 and 59) and four (nos. 14, 15, 24 and 47) outlier populations were removed, respectively. df, degrees of freedom. Host factor is nested: host species (*Alnus alnobetula, A. cordata, A. glutinosa*) into host subgenus (*Alnobetula, Alnus*). Region factor is nested: region level 2 (Corsica, France, Italy) into region level 1 (island, continental). *P*-values assess the overall significance of all terms together. Significance: ns, not significant; ., P < 0.01; **, P < 0.001.

region. The equality of proportions was tested using a twosample test with continuity correction (t.test function, STAT package in R). We also tested the correlation between the proportion of Sp+ nodules and *Frankia* alpha diversity by a linear model and a Spearman rank-order correlation test (cor.test function, STAT package in R). For EM fungi, we estimated the frequency of sequences identified as hypogeous fungi. The effect of host and region on their frequency at each site was tested by an ANOVA and completed with a Mann–Whitney test for pairwise differences, both computed in R.

Results

Molecular biology and sequencing of symbionts

For this study, 1139 EM root tips were collected in Corsica and 769 in Italy, of which 595 and 115 produced an EM sequence, respectively. In Italy, 45% of root tips were colonized by several fungi or were difficult to amplify and 33% of amplicons produced poor quality sequences. Sequences attributed to nonalder roots, that is 30 sequences belonging to Helotiales, Amanitaceae, Sebacinaceae, Boletaceae, Cantharellaceae and a Pezizales, as well as sequences attributed to alders but considered as endophytic rather than EM (Tedersoo *et al.*, 2009), that is 17 Helotiales, were discarded from the dataset. Our final dataset included 1343 EM and 224 *Frankia* sequences (Tables S3, S4).

Identification of EM symbionts

For fungi, 110 species were recognized (mean 99.1% similarity for the best BLAST, Table S3). The genera *Tomentella* and *Alnicola* were the most diverse (23 and 12 species, respectively), and the most abundant (351 and 186 sequences, respectively). Thirtyfour species were not yet recorded on alders and half of them could not be named with certainty at the species level. In the genus *Lactarius*, the species identified as *L. lepidotus* (a specialist of *A. alnobetula* subsp. *alnobetula*) needed re-evaluation, as all sequences were clustered in two well-supported lineages (arbitrarily named '*L. lepidotus* 1' and '*L. lepidotus* 2'), both distinct from the two sister species *L. lilacinus* and *L. cuspidatoaurantiacus* (Fig. 2).

Identification of Frankia symbionts

For Frankia, a single concatenation of up to 2040 nucleotide positions was aligned and used to infer the phylogeny (Fig. 3b). The phylogenetic tree topology was highly supported by posterior probabilities. All Corsican symbionts belonged to the monophyletic lineage of Alnus-infective Frankia and grouped into five divergent clades (1, 5-8), of which three clades (6, 7 and 8) have not been described before. Clade 6 and subclade 8b were strictly Corsican. Strains associated with Corsican A. alnobetula subsp. suaveolens clustered in a monophyletic lineage (1a-AAS) close to two sister clades associated with the other subspecies, either alnobetula in Europe (1a-AAA) or fruticosa and crispa in North America (1a-AAF/AAC). Strains isolated from A. cordata were often shared between Corsica and Italy, except subclade 8b. For A. glutinosa, only clade 6 was endemic to Corsica. Twenty-four species were delineated among 224 sequences, 11 were singletons and three -03, 20 and 23 - grouped together in c. 2/3 of the sequences (Fig. 3a). All the OTUs had species rank, as deduced from the reference strain positions in the phylogeny (Fig. 3b). As alder-infective Frankia species are mostly unnamed, the same is true for their phylogenetic clades and OTUs.

Autocorrelation test between sites and geographical structure

For EM fungi, the autocorrelation test was not significant (P=8e-04) and the spatial weight was not included in further analyses. For 70 out of 73 species (37 singletons were removed), the distribution was not spatially aggregated and the Geary–

Table 2 Multivariate permutation test on ectomycorrhizal (EM) and *Frankia* community dissimilarities (beta-diversity)

New Phytologist

Research 7



Fig. 2 Ectomycorrhizal (EM) fungi phylogenies. Phylogenies of EM genera associated with alders, based on the fungal internal transcribed spacer (ITS) regions and estimated using maximum likelihood: (a) genus *Alnicola*, (b) genus *Lactarius*, (c) genus *Alpova*, (d) genus *Tomentella*. (a–d) Strain origin (host plant): *Alnus glutinosa* nodules (AG, squares), *A. alnobetula* nodules (AA, triangles), *A. cordata* (AC, diamonds). Strain origin (region): continental France and Italy (open symbols) or Corsica (closed symbols). Node reliability was assessed using rapid bootstrapping (1000 pseudoreplicates). The scale represents the average number of substitutions per nucleotide.



0.02

Fig. 3 *Frankia* operational taxonomic unit (OTU) cladogram and multilocus sequence analysis (MLSA) phylogeny. (a) Cladogram of *Frankia* molecular taxonomic units (OTUs) calculated from the hierarchical clustering of *dnaA*, *ftsZ* and/or *pgk* marker alignment distances at the 0.05 threshold. Branch lengths are proportional to alignment distances and the angle sector of an OTU is proportional to its abundance. OTUs reported outside of the circle were singletons. (b) MLSA phylogeny of *Alnus*-infective *Frankia* based on concatenated sequences of *dnaA*, *ftsZ* and *pgk* markers and estimated by Bayesian inference. Strain origin (host plant): *Alnus glutinosa* nodules (squares), *A. incana* nodules (circles), *A. alnobetula* nodules (triangles), *A. cordata* (diamonds), reference strains (stars). Strain origin (region): continental France and Italy (open symbols) or Corsica (closed symbols). Phenotype of *in-planta* sporulation: spore-positive (+), spore-negative (-). Clades 6–8 are defined in the present study; clades 1–5 were defined in a previous study (Pozzi *et al.*, 2015). G1–G4, genomospecies of *Frankia* cluster 1 (Fernandez *et al.*, 1989). Clusters 1–4, as defined in Normand *et al.* (1996). Node reliability was assessed using posterior probabilities. The scale represents the average number of substitutions per nucleotide.

New Phytologist (2018) www.newphytologist.com Moran test was only significant (P < 0.05) for three species, Alnicola salabertii, Lactarius obscuratus and Russula sp. 2. No correlation was detected between geographic distance and beta diversity for EM (Mantel test, r = 0.06, P = 0.12). The Frankia dataset was not spatially autocorrelated, based on a multivariate randomization test (P = 0.0905). At the species level, only OTUs 3, 4 and 23 showed a spatially aggregated distribution (Geary–Moran test, P < 0.05). However, beta diversity and geographic distance were correlated (Mantel test, r = 0.11, P = 0.02).

Host and regional effects on gamma, alpha and beta diversities

Accumulation curves of Frankia reached a saturation plateau, while EM fungi did not (Fig. S2a). In Corsica, more EM species were associated with A. cordata compared to continental communities (Fig. S2b), the opposite pattern being observed for EM communities associated with A. glutinosa, while A. alnobetula subsp. suaveolens and subsp. alnobetula species accumulation curves did not differ and were instead influenced by sampling size. Overall, we did not detect a significant difference between Corsica and the Continent, nor a significant host effect on EM species richness and alpha diversity (Shannon index, Table 1). At the site level, EM species richness was not significantly higher in Corsica, and only Italian communities were less diverse and less species-rich. For Frankia, accumulation curves per host and per region followed a different trend compared to EM fungi. More species were associated with A. glutinosa in Corsica compared to the Continent and the opposite pattern was observed for A. cordata (Fig. S2c). Compared to these two host species, A. alnobetula subsp. alnobetula hosted fewer species than other alders on the Continent, and too few sites were investigated for A. alnobetula subsp. suaveolens to compare its accumulation curve. At the site level, Frankia communities hosted significantly more species in France compared to Italy (Table 1). The Shannon diversity index followed the same pattern as EM fungi, and only Italian communities were less diverse.

Beta diversity was not significantly different between EM communities in Corsica and the Continent, but slightly distinct when comparing Corsica, Italy and continental France (Tables 2, S6). Differences between communities were partly explained by the host subgenus (7%) and host species (7%) factors, yet 80% of the variation remained unexplained (Table 2). Communities associated with A. cordata were particularly distinct between Corsica and Italy (Fig. 4a). For Frankia, 4% of the beta diversity variation was explained by a Corsica vs Continent difference, 4% by the difference between Italy and France, 11% by the host species and 21% by the subgenus. In all, 41% of beta diversity variation was explained by these factors (Table 2). The strongest differences illustrated by the NMDS were between A. cordata communities in Italy and Corsica, and between A. alnobetula subspecies and the two other alder species (Fig. 4c). Subsequent tests confirmed that A. alnobetula hosted less variable communities than other hosts (ANOVA on betadisper test, P=0.047 and 0.008699 for EM fungi and Frankia, respectively) and that communities were

as variable in Corsica as on the continent (ANOVA on betadisper test on region level 1, P=0.897 and 0.8518 for EM and *Frankia*, respectively).

Climatic variables had different effects on EM fungi and *Frankia* communities, the NMDS structure being significantly correlated with twice as many variables for *Frankia* than for EM (Table S6; Fig. 4b,d). The effects of longitude and elevation were significant for EM and *Frankia*, and latitude for EM only (Table S6). For *Frankia*, bioclimatic and geographic variables were correlated with the host subgenus and species factors (Fig. 4d). Soil variables were not included, due to the lack of data available for several sites (Table S2).

Species distribution and detection of endemism and specificity

For fungi, 37 out of 110 species were restricted to Corsica - of which 12 were observed only once - and six were unique to Italy. For A. cordata, 33 of 44 species were exclusive to Corsica (Fig. 5). Two of these (Clavulina sp. 1 and Russula sp. 7) can be considered as potential endemics of Corsica as they have never been found before. For A. alnobetula subsp. suaveolens, six out of 17 species were restricted to Corsica (Fig. 5). Two hypogeous species, already described in Corsica, Alpova corsicus and Melanogaster rivularis, were true endemics. For A. glutinosa, only seven out of 24 species were restricted to Corsica in our dataset but they were mostly unnamed and therefore their endemism remains to be proved. For Frankia, four species were restricted to continental France, three to Italy and 10 were endemic to Corsica, of which seven were observed only once (Table S4). A. glutinosa hosted all the endemic Frankia species and shared three species with A. cordata, while A. alnobetula subsp. suaveolens did not host any endemic species (Fig. 5). Hierarchical clustering showed that the distribution of EM fungi and Frankia species was explained first by host and then by region.

The specialization class of each species is reported in Tables S3 and S4 for EM fungi and Frankia, respectively. Over the whole dataset, 20% of EM species were quoted as undetermined and 35% as generalists. All Frankia species were strictly associated with alders, and not considered as generalists - a term applied here only to fungi. Eight per cent of Frankia and 15% of EM species were alder-specialists, 25% of Frankia and 17% of EM species were subgenus Alnus-specialists, 42% of Frankia and 2% of EM species were A. glutinosa-specialists, 16% of Frankia and 4% of EM species were A. cordataspecialists and 8% of Frankia and 7% of EM species were A. alnobetula-specialists. Over the sites studied, the positions of specificity class factors were similar for both symbionts except for alder-specialists (Fig. 4e). Communities associated with A. glutinosa in Corsica did not appear more specialized than those on the Continent. By contrast, both Frankia and EM specificity was reduced in Corsica compared to Italy (Fig. 4e). The higher proportion of symbionts specific to the Alnus subgenus was significantly correlated with several bioclimatic variables related to warmer and dryer conditions. None of the soil variables tested was significant (Fig. 4f; Table S6).



Fig. 4 Community structures and environment fitting for ectomycorrhizal (EM) fungi and *Frankia*. Community structure of (a) EM fungi, (c) *Frankia* and (e) both symbionts. Fitting of environmental variables and factors for (b) EM fungi, (d) *Frankia* and (f) both symbionts. (a, c, e) The effect of alder host species (*Alnus alnobetula*, AA; *A. cordata*, AC; *A. glutinosa*, AG) and regions (Corsica vs Continent) on site ordination is illustrated by nonmetric multidimensional scaling (NMDS). For (a) EM fungi and (c) *Frankia*, two (nos. 43 and 59) and four (nos. 14, 15, 24 and 47) outlying populations were removed, respectively; for (e) both symbionts, only alder populations in common were included except three *Alnus alnobetula* populations that were discarded. (b, d, f) The correlation between NMDS, host and regional factors as well as geographic and climatic variables was tested through a permutation analysis (envfit function). (f) Four soil variables were also tested. Only significant factors (*P* > 0.05) are represented (see Table S6 for details).

Distribution of short-distance dispersal symbionts

Spore-positive (Sp+) *Frankia* were identified in all regions and host species. In Corsica and on the Continent, *Frankia* nodules were 100% Sp+ vs 69.2% for *A. alnobetula*, 12.8% Sp+ vs 45.7% for *A. glutinosa*, and 86.1% Sp+ vs 35.3% for *A.* cordata, respectively. These proportions were significantly different between Corsica and the Continent (two-sample test with continuity correction, all P < 0.01). Species richness was partly explained by the Sp+ frequency (linear model, P < 0.1; adjusted $R^2 = 25-50\%$ according to the regression method) and these two parameters were negatively correlated (Spearman rank-order correlation, P < 0.01; $\rho = -0.90$).

Seven genera of hypogeous fungi were detected: *Alpova, Hydnobolites, Melanogaster, Octaviania, Pachyphlodes* and *Tuber* spp. The frequency of the hypogeous fungi group (based on the number of sequences) was higher in Corsica than on the Continent (ANOVA, P=0.03). However, the predominance of

hypogeous fungi in Corsica, as compared to Italy, was significant for *A. cordata* (Mann–Whitney test on hypogeous frequency, P=0.02, Fig. S3), but not for the two other alder species (Mann–Whitney tests on hypogeous frequency, P=0.62 for both). Of interest, *Cenococcum geophilum*, a common and widespread ascomycete that does not produce fruiting bodies, was found only twice in Corsica.

Discussion

Based on classical hypotheses on islands and ice-age refugia, we expected to detect higher alpha and gamma diversity of symbiont communities on Corsican alders, and following recent studies on alders, we also expected more endemic species of *Frankia*. Interestingly, Corsica harbours a high level of diversity in some cases, but the patterns of distribution were far more complex than assumed. This was because geographic isolation had a different effect on each symbiont



Fig. 5 Distribution of ectomycorrhizal (EM) fungi and Frankia species among regions and hosts. The clustering was based on community similarity (Bray-Curtis distance). The Venn diagrams show, for each symbiont and each host, the number of species in Corsica and on the Continent, as well as the number of species shared between the two regions. Hosts, Alnus alnobetula (AA), A. cordata (AC), A. glutinosa (AG).

community, which in turn also depended on the host species. This suggests that the theory of island biogeography may not apply in every case.

Diversity of symbiont communities

Corsican Frankia communities were more diverse on A. glutinosa, as previously reported on A. glutinosa subsp. barbata from the Colchis region, another glacial and Tertiary refugium for alders (Roy et al., 2017). Despite a similar sampling effort in Corsica, the number of Frankia species on A. cordata was three times lower than on A. glutinosa. This difference can be partly explained by the sporulation phenotype. Most strains isolated from A. glutinosa in Corsica were Spwhile most strains from A. cordata were Sp+, confirming that the percentage of Sp+ nodules on alder stands negatively correlates with Frankia diversity at a regional scale, as already shown in Europe and North America (Pozzi et al., 2015). As Sp+ strains are highly host-dependent and produce a huge number of clonal offspring, they would invade the roots locally (Cotin-Galvan et al., 2016) and could prevent hosts from associating with other Frankia strains, creating a bottleneck effect in Frankia diversity. Conversely, Corsican EM communities were richer on A. cordata, while it was not the case in a previous study (Roy et al., 2013). The number of EM species on A. cordata was twice that on A. glutinosa. This difference was due to an increased recruitment of generalist EM fungi rather than a recruitment of unique alder specialist EM fungi by A. cordata, a situation which would be easily achieved in Corsica where the ecological range of A. cordata includes mixed forests. In summary, communities were distinct between regions, and shaped by their geographic position, confirming that the spatial structure of alder symbiont communities could be relatively strong at the regional scale, as previously demonstrated for EM (Peay et al., 2007, 2010) and for both symbionts, especially in refugia (Roy et al., 2017). However, regional differences were not necessarily explained by the recruitment of endemic symbionts.

EM and Frankia endemism

While 37 EM species were detected only in Corsica, only four can be considered to be true endemics after database queries: the two hypogeous fungi Alpova corsicus and Melanogaster rivularis (Moreau et al., 2011), one unidentified Russula and one Clavulina species - 'Clavulina sp. 1', specific to A. cordata. Interestingly, these last two species are epigeous (our unpublished observations for Clavulina sp. 1) and supposedly wind-dispersed, but they have not been found associated with A. cordata on the continent. They were not found in native stands in Calabria either, unlike the most common A. cordata-specialist, Alnicola salabertii, which is present wherever its host can be found (Moreau & Garcia, 2005). The remaining 33 EM species were generalist fungi previously recorded outside of Corsica, such as Amanita rubescens or Russula delica. These data confirm that alder stands in Corsica host no more than a few endemic EM species (Taudiere et al., 2015). Similarly, a study on polypores in Corsica also failed to detect endemic taxa (Nemergut et al., 2001). According to a review on EM fungi, isolation by distance was only detected for wind-dispersed populations separated by thousands of kilometres (Douhan et al., 2011). All these observations suggest that geographic distance and time of separation between Corsica and the European continent were not sufficient to induce speciation for EM fungi, with the exception of a few hypogeous species which might be paleoendemics rather than allospeciated (Moreau et al., 2011).

As Frankia may be more dispersal-limited than EM fungi, we expected unique Frankia species and richer communities in Corsica. Indeed, 50% of the recovered Frankia species have never been recorded on the continent or in previous samples (Pozzi et al., 2015; Roy et al., 2017) and were therefore considered to be endemic to the island. Surprisingly, the more widespread was the alder species, the more endemic Frankia it hosted in Corsica: A. glutinosa hosted more endemic species than A. cordata, while the strictly Corsican A. alnobetula subsp. suaveolens hosted no endemic species (Fig. 5). However, although all A. alnobetula strains constituted a unique species (clade 1a) in the phylogeny (Fig. 3), Corsican strains clustered together (clade 1a AAS), suggesting the existence of an endemic lineage at a taxonomic rank lower than species. The divergence between Corsican and continental A. alnobetula populations only 1.1 Ma (Rochet et al., 2011) was probably not sufficient to allow speciation of Frankia and detection of endemic species. At the opposite end of the spectrum, the oldest divergence in Frankia phylogeny corresponded to clade 8. This group is entirely associated with A. cordata, a species that diverged from other alders 22.9 Ma (Rochet et al., 2011). The position of Italian strains in this clade suggests a later colonization from Corsica, possibly across the terrestrial bridge connecting Corsica to Italy during the Messinian crisis 5 Ma (Clauzon et al., 1996). This past exchange between Corsica and Italy may explain the current reduced endemism found on A. cordata in Corsica. For A. glutinosa, the presence of numerous endemic species of Frankia found in Corsica confirms the pattern recently described in another Tertiary and glacial refugium, the Colchis region (Roy et al., 2017). However, for Corsica, *A. cordata* and *A. glutinosa* occur sympatrically at midaltitudes, where they could hybridize (Lhote, 1985; King & Ferris, 2000; Gryta *et al.*, 2017), explaining why endemic strains can be shared between the two hosts.

Patterns of specialization

In Corsica, the co-occurrence of three alder species and the growth of A. cordata in mixed woods represent a rare situation for EM fungi, as alders generally contribute little to networks (Kennedy et al., 2015; Taudiere et al., 2015). We indeed detected shared symbionts in Corsica, but also on the continent and not necessarily at the scale of alder stands. Interestingly, we detected Frankia species shared between A. glutinosa and A. cordata both in Corsica and on the continent (for instance, OTU 20). This pattern of Frankia co-occurrence was relatively rare, as 66% of Frankia species are only associated with a single alder species in our dataset. This was considerably more than the 13% for EM fungi. Only 8% of Frankia species were found to be associated with the three studied alder species, compared to 17% in EM fungi. Thus, Frankia species appeared to be more host-specific than EM species. Among EM fungi, 10 species belonging to the genera Alnicola, Lactarius and Tomentella were shared between alder species both on the continent and in Corsica. Only two EM species were shared only in Corsica, Cenococcum geophilum - a generalist - and Pachyphlodes sp. Therefore, the distribution of alders in Corsica might not promote recent host shifts or sharing, in line with observations on the continent. The analysis of EM specificity confirmed that A. cordata hosted 'more generalist' communities in Corsica, compared with pure A. cordata stands like those in Italy. Although the sequencing rate was lower on Italian populations than in Corsica, the species accumulation curve of A. cordata in Italy was saturated, suggesting that sampling bias may not explain the difference in specificity. We did not detect a correlation between community specialization and any soil variable tested in our study, consistent with the literature (Põlme et al., 2013; Rochet et al., 2011). Our results suggest rather that a warmer and drier climate correlates with less specialized, more generalist symbiont communities. Numerous ascomycetes were isolated from A. subcordata (C.A.Mey) Regel in Iran (Põlme et al., 2013) and A. glutinosa subsp. barbata (C.A.Mey) Yalt. in Georgia (Roy et al., 2017). These two alder species are Tertiary relics of the Hyrcanian forest and the Colchis region, and colonize relatively dry and warm habitats, like A. cordata in Corsica. Interestingly, Corsican A. cordata hosted more ascomycetes and hypogeous taxa than in Italian populations or other hosts. Hypogeous taxa are also supposedly well adapted to drought (Richard et al., 2011; Herzog et al., 2013) and are abundant in Mediterranean or dry habitats (for a review see Zambonelli et al., 2014). Sequences of Tuber sp. were detected on these three alder species, and also on A. glutinosa introduced to New Zealand (Bogar & Kennedy, 2013; Bogar et al., 2015). None of these sequences is identical and their detection on a few sites suggests local recruitment rather than true specialization on alders.

Conclusion

Study of the Frankia-EM-alder symbiosis in Corsica is an interesting model, permitting finer analysis at the community level. We detected a high level of endemism for Frankia but not for EM fungi. Our results also revealed that symbionts which are host-specialists or poor dispersers have distinct biogeographic patterns. Yet, it is difficult to fully understand the biogeography of this tripartite symbiosis at a regional scale, perhaps because several dispersal events contributed to the colonization of Corsica and because gene flow with populations on the continent remains important at least for epigeous fungi. While Corsica is well known as a hotspot of plant diversity, comparison with Italy clearly showed that other alder stands can also host diverse Frankia. Finally, specificity patterns of both symbionts followed a common trend and did not increase particularly in Corsica, suggesting that specificity rarely varies at these geographic and temporal scales. Only A. cordata clearly associated with more generalist EM fungi and more host-dependent Sp+ Frankia, which raises new questions about the ecology of this peculiar host.

Acknowledgements

We thank Jean Alessandri (Société Mycologique d'Ajaccio), Laëtitia Hugot (Conservatoire Botanique de Corse), Marc-André Selosse (MHNH, Paris), Sonia Ravera (Orto Botanico di Roma), Alessandro Bellino and Domenico Puntillo for their help with sampling in Corsica or Italy. We also thank Ian Dickie, Peter Kennedy and two anonymous reviewers for their useful comments that helped to improve this paper, and Benjamin Durrington and Alain Roy for editing the text. M.R. was funded by an EC2CO MicrobiEn grant (Centre National de la Recherche Scientifique, France), and A.C.P. by a doctoral fellowship (Ministère de l'Education Nationale, de l'Enseignement Supérieur et de la Recherche, France).

Author contributions

All authors contributed to this work. A.C.P., M.R., P-A.M., M.G. and M.P.F. designed the research and collected new samples in Corsica. A.C.P., M.N., G.S. and S.M. performed the laboratory work. A.C.P., M.R. and M.N. analyzed the data. A.C.P., M.R., P-A.M. and M.P.F. interpreted the data and wrote the manuscript. A.C.P. and M.R. contributed equally to this work.

ORCID

Adrien C. Pozzi (D) http://orcid.org/0000-0001-6765-4293

References

 Angel R, Soares MIM, Ungar ED, Gillor O. 2010. Biogeography of soil archaea and bacteria along a steep precipitation gradient. *The ISME Journal* 4: 553–563.
Blondel J, Aronson J. 1999. *Biology and wildlife of the Mediterranean region*. Oxford, UK: Oxford University Press. Bogar LM, Dickie IA, Kennedy PG. 2015. Testing the co-invasion hypothesis: ectomycorrhizal fungal communities on *Alnus glutinosa* and *Salix fragilis* in New Zealand. *Diversity and Distributions* 21: 268–278.

Bogar LM, Kennedy PG. 2013. New wrinkles in an old paradigm: neighborhood effects can modify the structure and specificity of *Alnus*associated ectomycorrhizal fungal communities. *FEMS Microbiology Ecology* 83: 767–777.

Bosco M, Fernandez MP, Simonet P, Materassi R, Normand P. 1992. Evidence that some *Frankia* sp. strains are able to cross boundaries between *Alnus* and *Elaeagnus* host specificity groups. *Applied Environmental Microbiology* 58: 1569–1576.

Bray JR, Curtis JT. 1957. An ordination of the upland forest communities of southern Wisconsin. *Ecological Monographs* 27: 325–349.

Brown MV, Lauro FM, DeMaere MZ, Muir L, Wilkins D, Thomas T, Riddle MJ, Fuhrman JA, Andrews-Pfannkoch C, Hoffman JM et al. 2012. Global biogeography of SAR11 marine bacteria. *Molecular Systems Biology* 8: 595.

Clauzon G, Suc JP, Gautier F, Berger A, Loutre MF. 1996. Alternate interpretation of the Messinian salinity crisis: controversy resolved? *Geology* 24: 363–366.

Cotin-Galvan L, Pozzi AC, Schwob G, Fournier P, Fernandez MP, Herrera-Belaroussi A. 2016. *In-planta* sporulation capacity enhances infectivity and rhizospheric competitiveness of *Frankia* strains. *Microbes and Environments* 31: 11–18.

Dawson JO. 2007. Ecology of actinorhizal plants. In: Pawlowski K, Newton WE, eds. *Nitrogen-fixing actinorhizal symbioses*. Dordrecht, the Netherlands: Springer, 199–234.

De Queiroz K. 2007. Species concepts and species delimitation. *Systematic Biology* 56: 879–886.

Dixon P. 2003. VEGAN, a package of R functions for community ecology. *Journal of Vegetation Science* 14: 927–930.

Douda J, Doudová J, Drašnarová A, Kuneš P, Hadincová V, Krak K, Zákravský P, Mandák B. 2014. Migration patterns of subgenus *Alnus* in Europe since the Last Glacial Maximum: a systematic review. *PLoS ONE* 9: e88709.

Douhan GW, Vincenot L, Gryta H, Selosse MA. 2011. Population genetics of ectomycorrhizal fungi: from current knowledge to emerging directions. *Fungal Biology* 115: 569–597.

Ducci F, Tani A. 2009. EUFORGEN Technical Guidelines for genetic conservation and use of Italian alder (Alnus cordata). Rome, Italy: Bioversity International.

Edgar RC. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* 32: 1792–1797.

Fernandez MP, Meugnier H, Grimont PA, Bardin R. 1989. Deoxyribonucleic acid relatedness among members of the genus *Frankia*. International Journal of Systematic and Evolutionary Microbiology 39: 424–429.

Gamisans J. 1999. La végétation de la Corse (2nd edn). Aix-en-Provence, France: EdiSud.

Gevers D, Cohan FM, Lawrence JG, Spratt BG, Coenye T, Feil EJ, Stackebrandt E, Van de Peer Y, Vandamme P, Thompson FL *et al.* 2005. Opinion: re-evaluating prokaryotic species. *Nature Reviews Microbiology* 3: 733–739.

Gouy M, Guindon S, Gascuel O. 2010. SeaView version 4: a multiplatform graphical user interface for sequence alignment and phylogenetic tree building. *Molecular Biology and Evolution* 27: 221–224.

Griekspoor A, Groothuis T. 2007. *4Peaks, version 1.7.1.* [WWW document] URL https://nucleobytes.com/4peaks/index.html [accessed 19 January 2018].

Gryta H, Van de Paer C, Manzi S, Holota H, Roy M, Besnard G. 2017. Genome skimming and plastid microsatellite profiling of alder trees (*Alnus* spp., Betulaceae): phylogenetic and phylogeographical prospects. *Tree Genetics* & *Genomes* 13: 118.

Havrdová A, Douda J, Krak K, Vít P, Hadincová V, Zákravský P, Mandák B. 2015. Higher genetic diversity in recolonized areas than in refugia of *Alnus glutinosa* triggered by continent-wide lineage admixture. *Molecular Ecology* 24: 4759–4777.

Herzog C, Peter M, Pritsch K, Günthardt-Goerg MS, Egli S. 2013. Drought and air warming affects abundance and exoenzyme profiles of *Cenococcum geophilum* associated with *Quercus robur*, *Q. petraea* and *Q. pubescens. Plant Biology* 15: 230–237. Higgins LM, Kennedy PG. 2012. Symbiotic *Frankia* bacteria in *Alnus* forests in Mexico and the United States of America: is geographic location a good predictor of assemblage structure? *Botany-Botanique* 90: 423–431.

Hijmans RJ, van Etten J. 2012. raster: geographic analysis and modeling with raster data. R package version 2.0–12. Vienna, Austria: R Foundation for Statistical Computing. [WWW document] URL https://cran.r-project.org/web/packages/ raster/index.html [accessed 19 January 2018].

Huang M, Hull CM. 2017. Sporulation: how to survive on planet Earth (and beyond). Current Genetics. 18: 1–8.

Huggins JA, Talbot J, Gardes M, Kennedy PG. 2014. Unlocking environmental keys to host specificity: differential tolerance of acidity and nitrate by *Alnus*-associated ectomycorrhizal fungi. *Fungal Ecology* 12: 52–61.

Jeanmonod D, Gamisans J. 2007. Flora Corsica. Aix-en-Provence, France: Edisud.

Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution* 30: 772–780.

Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28: 1647–1649.

Kekkonen M, Hebert PD. 2014. DNA barcode-based delineation of putative species: efficient start for taxonomic workflows. *Molecular Ecology Resources* 14: 706–715.

Kennedy PG, Garibay-Orijel R, Higgins LM, Angeles-Arguiz R. 2011. Ectomycorrhizal fungi in Mexican *Alnus* forests support the host co-migration hypothesis and continental-scale patterns in phylogeography. *Mycorrhiza* 21: 559–568.

Kennedy PG, Walker JK, Bogar LM. 2015. Interspecific mycorrhizal networks and non-networking hosts: exploring the ecology of the host genus *Alnus*. In: Horton TR, ed. *Mycorrhizal networks*. Dordrecht, the Netherlands: Springer, 227–254.

King RA, Ferris C. 1998. Chloroplast DNA phylogeography of *Alnus glutinosa* (L.) Gaertn. *Molecular Ecology* 7: 1151–1161.

King RA, Ferris C. 2000. Chloroplast DNA and nuclear DNA variation in the sympatric alder species, *Alnus cordata* (Lois.) Duby and *A. glutinosa* (L.) Gaertn. *Biological Journal of the Linnean Society* 70: 147–160.

Lhote P. 1985. *Etude écologique des aulnes dans leur aire naturelle en France*. Besançon, France: IDF-ENGREF- Université de Franche-Comté.

Lipus A, Kennedy PG. 2011. *Frankia* assemblages associated with *Alnus rubra* and *Alnus viridis* are strongly influenced by host species identity. *International Journal of Plant Sciences* 172: 403–410.

Lumini E, Bosco M, Fernandez MP. 1996. PCR-RFLP and total DNA homology revealed three related genomic species among broad-host-range *Frankia* strains. *FEMS Microbiology Ecology* 21: 303–311.

Mandák B, Havrdová A, Krak K, Hadincová V, Vít P, Zákravský P, Douda J. 2016. Recent similarity in distribution ranges does not mean a similar postglacial history: a phylogeographical study of the boreal tree species *Alnus incana* based on microsatellite and chloroplast DNA variation. *New Phytologist* 210: 1395–1407.

Mantel N. 1967. Ranking procedures for arbitrarily restricted observation. *Biometrics* 23: 65–78.

Markham J, Chanway CP. 1998. Alnus rubra (Bong.) nodule spore type distribution in southwestern British Columbia. Plant Ecology 135: 197–205.

Medail F, Quezel P. 1997. Hot-spots analysis for conservation of plant biodiversity in the Mediterranean Basin. *Annals of the Missouri Botanical Garden* 1: 112–127.

Miller MA, Pfeiffer W, Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. Gateway Computing Environments Workshop (GCE), IEEE, New Orleans, LA, USA, 1–8.

Molina R. 1981. Ectomycorrhizal specificity in the genus Alnus. Canadian Journal of Botany 59: 325–334.

Moreau PA, Garcia G. 2005. Alnicola salabertii, sp. nov., mycorhizique d'Alnus cordata, et deux autres Alnicola à petites spores. Bulletin Trimestriel de la Société Mycologique de France 120: 273–292.

Moreau PA, Rochet J, Richard F, Chassagne F, Manzi S, Gardes M. 2011. Taxonomy of *Alnus*-associated hypogeous species of *Alpova* and *Melanogaster* (Basidiomycota, Paxillaceae) in Europe. *Cryptogamie Mycologie* **32**: 33–62. Moreau PA, Rochet J, Welti S, Peintner U, Manzi S, Courtecuisse R, Gardes M. 2013. Agarics of alders 2 – Three new species of *Alnicola* (Agaricales, Hymenogastraceae) with a key to species associated with *Alnus alnobetula* in Europe. *Cryptogamie Mycologie* 34: 149–173.

Nemergut DR, Costello EK, Hamady M, Lozupone C, Jiang L, Schmidt SK, Norstedt G, Bader P, Ericson L. 2001. Polypores as indicators of conservation value in Corsican pine forests. *Biological Conservation* **99**: 347–354.

Normand P, Orso S, Cournoyer B, Jeannin P, Chapelon C, Dawson J, Evtushenko L, Misra AK. 1996. Molecular phylogeny of the genus *Frankia* and related genera and emendation of the family Frankiaceae. *International Journal of Systematic and Evolutionary Microbiology* 46: 1–9.

Peay KG, Bruns TD, Kennedy PG, Bergemann SE, Garbelotto M. 2007. A strong species–area relationship for eukaryotic soil microbes: island size matters for ectomycorrhizal fungi. *Ecology Letters* **10**: 470–480.

Peay KG, Garbelotto M, Bruns TD. 2010. Evidence of dispersal limitation in soil microorganisms: isolation reduces species richness on mycorrhizal tree islands. *Ecology* 91: 3631–3640.

Põlme S, Bahram M, Kõljalg U, Tedersoo L. 2014. Global biogeography of *Alnus*-associated *Frankia* actinobacteria. *New Phytologist* 204: 979–988.

Põlme S, Bahram M, Yamanaka T, Nara K, Dai YC, Grebenc T, Kraigher H, Toivonen M, Wang P-H, Matsuda Y. 2013. Biogeography of ectomycorrhizal fungi associated with alders (*Alnus* spp.) in relation to biotic and abiotic variables at the global scale. *New Phytologist* 198: 1239–1249.

Pozzi AC, Bautista-Guerrero HH, Nouioui I, Cotin-Galvan L, Pepin R, Fournier P, Menu F, Fernandez MP, Herrera-Belaroussi A. 2015. Inplanta sporulation phenotype: a major life history trait to understand the evolution of Alnus-infective Frankia strains. Environmental Microbiology 17: 3125–3138.

Richard F, Roy M, Shahin O, Sthultz C, Duchemin M, Joffre R, Selosse MA. 2011. Ectomycorrhizal communities in a Mediterranean forest ecosystem dominated by *Quercus ilex*: seasonal dynamics and response to drought in the surface organic horizon. *Annals of Forest Science* 68: 57–68.

Rochet J, Moreau PA, Manzi S, Gardes M. 2011. Comparative phylogenies and host specialization in the alder ectomycorrhizal fungi *Alnicola, Alpova* and *Lactarius* (Basidiomycota) in Europe. *BMC Evolutionary Biology* 11: 1–40.

Roy M, Pozzi AC, Gareil R, Nagati M, Manzi S, Nouioui I, Sharikadze N, Jargeat P, Gryta H, Moreau PA *et al.* 2017. Alder and the Golden Fleece: high diversity of *Frankia* and ectomycorrhizal fungi revealed from *Alnus glutinosa* subsp. *barbata* roots close to a Tertiary and glacial refugium. *PeerJ* 5: e3479.

Roy M, Rochet J, Manzi S, Jargeat P, Gryta H, Moreau PA, Gardes M. 2013. What determines *Alnus*-associated ectomycorrhizal community diversity and specificity? A comparison of host and habitat effects at a regional scale. *New Phytologist* **198**: 1228–1238.

Schwintzer CR. 1990. Spore-positive and spore-negative nodules. In: Schwintzer CR, Tjepkema JD, eds. *The biology of* Frankia *and actinorhizal plants*. San Diego, CA, USA: Academic Press, 177–193.

Sen A, Daubin V, Abrouk D, Gifford I, Berry AM, Normand P. 2014. Phylogeny of the class Actinobacteria revisited in the light of complete genomes. The orders 'Frankiales' and Micrococcales should be split into coherent entities: proposal of Frankiales ord. nov., Geodermatophilales ord. nov., Acidothermales ord. nov. and Nakamurellales ord. nov. International Journal of Systematic and Evolutionary Microbiology 64: 3821–3832.

Smouse PE, Peakall R. 1999. Spatial autocorrelation analysis of individual multiallele and multilocus genetic structure. *Heredity* 82: 561–573.

Stamatakis A, Hoover P, Rougemont J. 2008. A rapid bootstrap algorithm for the RAxML web servers. *Systematic Biology* 57: 758–771.

Taudiere A, Munoz F, Lesne A, Monnet A-C, Bellanger J-M, Selosse M-A, Moreau PA, Richard F. 2015. Beyond ectomycorrhizal bipartite networks: projected networks demonstrate contrasted patterns between early-and latesuccessional plants in Corsica. *Frontiers in Plant Science* **6**: 881.

Taylor JW, Jacobson DJ, Kroken S, Kasuga T, Geiser DM, Hibbett DS, Fisher MC. 2000. Phylogenetic species recognition and species concepts in fungi. *Fungal Genetics and Biology* 31: 21–32.

Tedersoo L, Pärtel K, Jairus T, Gates G, Põldmaa K, Tamm H. 2009. Ascomycetes associated with ectomycorrhizas: molecular diversity and ecology with particular reference to the Helotiales. *Environmental Microbiology* 11: 3166–3178.

Zambonelli A, Donnini D, Rana GL, Fascetti S, Benucci GMN, Iotti M, Morte A, Khabar L, Bawadekji A, Piattoni F. 2014. Hypogeous fungi in Mediterranean maquis, arid and semi-arid forests. *Plant Biosystems-An International Journal Dealing With all Aspects of Plant Biology* 148: 392–401.

Supporting Information

Additional Supporting Information may be found online in the Supporting Information tab for this article:

Fig. S1 Histogram of pairwise phylogenetic distances for *Frankia* species delineation.

Fig. S2 Ectomycorrhizal (EM) fungi and *Frankia* species accumulation curves.

Fig. S3 Frequency of ectomycorrhizal (EM) fungi reproducing below ground (hypogeous taxa, *Tylospora, Amphinema, Piloderma* and *Cenococcum geophilum*) and Ascomycota sequences.

Table S1 Sampled population information and new sites sampled in this study

Table S2 Soil variables for the most recent sampled sites

Table S3 Community matrix of ectomycorrhizal (EM) fungispecies and GenBank accession numbers

Table S4 Community matrix of Frankia species

Table S5 Community matrix of ectomycorrhizal (EM) fungi and*Frankia* specialization classes for the commonly sampled popula-tions

Table S6 Synthesis on environmental variables tested in NMDS

Please note: Wiley Blackwell are not responsible for the content or functionality of any Supporting Information supplied by the authors. Any queries (other than missing material) should be directed to the *New Phytologist* Central Office.