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# Chain stopper assisted characterization of supramolecular polymers

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**Abstract:** Supramolecular polymers are dynamic materials, consequently their molar mass is concentration dependent. However, the present experimental results show that an efficient chain stopper (*i.e.* a monofunctional monomer) can be used to block the concentration dependence of the molar mass of a hydrogen bonded supramolecular polymer, over a realistic concentration range. This fact was used to derive the molecular weight and radius of gyration of the stopped supramolecular chains (by light scattering), as well as the intrinsic viscosity. In a second step, the

molecular weight of the bis-urea based supramolecular polymer (EHUT) was determined in the absence of a chain stopper.

#### Introduction

Supramolecular polymers are chains of small molecules held together through reversible non-covalent interactions.<sup>13</sup> This reversibility is desirable because it is responsible for the appearance of new properties, as compared to those of usual covalent polymers. For instance, improved processability can be expected from the temperature dependence of the molar mass of supramolecular polymers.<sup>4</sup> Moreover, self-healing materials<sup>2,6</sup> may be obtained because of the dynamic nature of the chains.

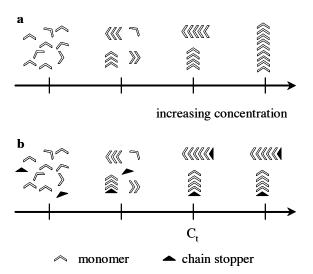
However, this reversibility is also a drawback, as far as characterization is concerned. In particular, molar mass measurement is impeded by the fact that molar mass changes with concentration. Thus, classical polymer characterization techniques such as size exclusion chromatography, static light scattering or osmometry cannot be used quantitatively. Up to now, only indirect methods have been used. These methods, such as fluorescence spectroscopy of labeled monomers, chain-end titration by NMR\*\*\* or FTIR\*\*\* spectroscopy, or calorimetry\*\*, rely on particular association models (isodesmic or cooperative association, ring-chain equilibrium, etc). The aim of this paper is to show (through a particular example) that the adequate use of a chain stopper (*i.e.* a monofunctional monomer) can help characterize supramolecular polymers.

Chain stoppers have previously been used to reduce the chain length of supramolecular polymers and thus the viscosity of their solutions, 5,16,22 but not to actually block the concentration dependence of the molar mass of supramolecular polymers.

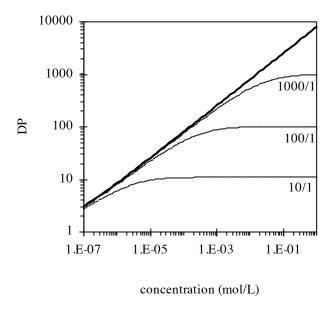
# Concept

The concept of the stopper-assisted mass control is illustrated in Figures 1 and 2 with the example of an AB type monomer, but can be generalized to any type of structure. As mentioned in the introduction, the chain length of a supramolecular polymer increases continuously as the concentration is increased (Figure 1a and 2). However, if a chain stopper with a strong association constant is present in a fixed stopper to monomer ratio, the chain length increases with the overall concentration only up to a certain concentration C<sub>i</sub>, where the vast majority of chains is terminated by a chain stopper (Figure 1b and 2). At concentrations above C<sub>1</sub>, the chain length cannot increase further because the stopped chains do not bear any complementary functional group. The difference of behavior between the two cases lies in the fact that for a chain stopped supramolecular polymer, there is a minimum number of chains present in solution (i.e. the number of chain stopper molecules) and thus a maximum chain length. On the contrary, for a stopper free supramolecular polymer, the number of chains present in solution can become vanishingly low when concentration is increased. Consequently, there is a range of concentrations  $(C \ge C)$  where the molar mass of a supramolecular polymer (with a fixed proportion of chain stopper) is constant. Over this range of concentrations, the physical properties of this system can be considered to vary in the same manner as those of covalent polymers. In particular, scattering or viscosity measurements can be extrapolated to zero concentration (over the range  $C \ge C_0$ ), in order to derive characteristic parameters such as the radius of gyration or the second virial coefficient of the chains.

Of course, for this general idea to be of any practical value, the threshold concentration  $C_1$  must be sufficiently low, which means that the self-association constant of the monomer (and thus the association constant of the chain stopper) must be large. The remaining of this paper shows through a particular example that it is indeed possible to experimentally apply this concept.



**Figure 1.** Schematic representation of the influence of concentration on the chain length of a supramolecular polymer without (a) or with (b) a chain stopper at a fixed stopper to monomer ratio.



**Figure 2.** Calculated number average degree of polymerization for a supramolecular polymer (bold curve) and for 1000/1, 100/1 and 10/1 monomer/chain stopper mixtures (plain curves), versus monomer concentration. The association constant is 6 10<sup>7</sup> L mol<sup>-1</sup> (see Supporting Information for details).

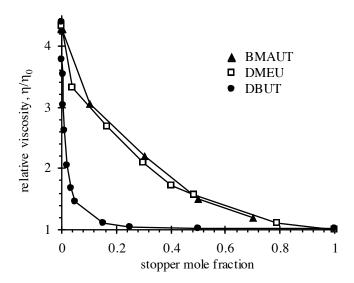
Chart 1. Structures of the monomer (EHUT) and of the chain stoppers (DBUT, BMAUT, DMEU).

### **Results and Discussion**

The experimental system chosen to test the present concept is bis-urea based supramolecular polymer **EHUT** (Chart 1) in carbon tetrachloride. This compound has been shown to reversibly self-assemble by hydrogen bonding in low polarity solvents, where it forms long wire-like structures.<sup>12,23</sup>

1. Selection of a suitable chain stopper. Potential chain stoppers have been designed by capping hydrogen bond donor groups on urea or bis-urea (Chart 1). They have been tested by measuring the drop in viscosity of EHUT solutions with an increasing proportion of chain stopper (Figure 3).

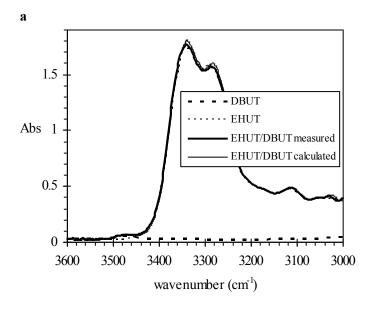
DBUT is a very efficient chain stopper because only a few percent is sufficient to dramatically reduce the viscosity of EHUT solutions.

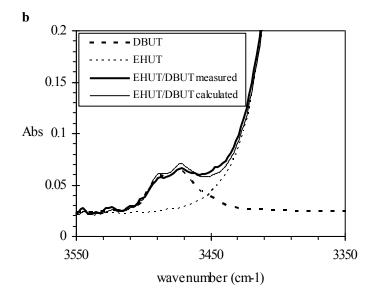


**Figure 3.** Relative viscosity of mixtures of **EHUT** and chain stoppers in CCl<sub>4</sub>, versus stopper mole fraction. The overall concentration is the same for all experiments ([**EHUT**] + [stopper] = 3.1 g L (7.2 mM for **EHUT/DBUT** mixtures)).

Mono-urea **DMEU** is much less efficient, showing that the two urea functions of **DBUT** interact cooperatively with **EHUT**, in the same way as **EHUT** was previously shown to self-assemble cooperatively. Remarkably, **BMAUT** does not seem to benefit from such a cooperative effect. This is attributed to the fact that the most stable conformations of the urea groups in **EHUT** and **BMAUT** are different. Indeed, it has been reported that in the most stable conformation, the carbonyl group of a mono-substituted urea is in the *cis* position relative to the substituent (as depicted in Chart 1). However, the carbonyl group of a N,N-disubstituted urea is more stable in the *trans* position relative to the aromatic substituent.

**DBUT** being the most efficient chain stopper, it was selected for the following experiments.



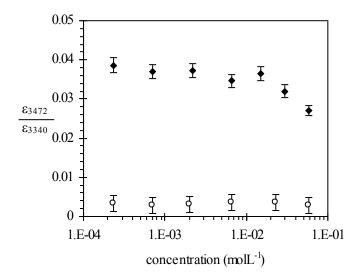


**Figure 4.** FTIR spectra of solutions in CCl<sub>4</sub> (complete NH region (a) and magnification of the free NH region (b)). "EHUT": **EHUT** at 2.0 x 10<sup>3</sup> mol L<sup>4</sup>; "DBUT": **DBUT** at 2.2 x 10<sup>4</sup> mol L<sup>4</sup>; "EHUT/DBUT experimental": **EHUT** at 2.0 x 10<sup>3</sup> mol L<sup>4</sup> with **DBUT** at 2.2 x 10<sup>4</sup> mol L<sup>4</sup>; "EHUT/DBUT calculated": summation of "EHUT" and "DBUT".

2. Determination of the relevant range of concentrations. By monitoring the presence of free NH groups, infrared spectroscopy is well suited to quantify the number of chain ends in the present system.<sup>12</sup> Figure 4a shows that a 90/10 EHUT/DBUT mixture at an overall concentration of 2.2mM, is characterized by a small free NH band (3472cm<sup>-1</sup>) and a large hydrogen bonded NH band (3340cm<sup>-1</sup>). Interestingly, this spectrum can be matched by adding the spectrum of self-associated **EHUT** to the spectrum of totally dissociated **DBUT** (Figure 4b). This fact yields no information about the interaction between **DBUT** and **EHUT**, because obviously, the same spectrum would be expected if no interactions were present. However, it means that for the 90/10 EHUT/DBUT mixture at a concentration of 2.2mM, the number of unstopped chains is negligible compared to the number of chain stopper (whether the latter are attached to a chain end or free). Moreover, if we assume that the association constant of DBUT with EHUT is at least of the same order of magnitude as that of the self-association of **EHUT** (as indicated by the viscosity measurements), then the concentration of free **DBUT** can be neglected. Consequently, the concentration at which the vast majority of chains are terminated by a chain stopper (C) is lower than 2.2mM. The results for the same experiment, repeated over a large range of concentrations, are summarized in Figure 5. At low concentrations (between 0.2 and 15mM), the ratio of the free NH band to the hydrogen bonded NH band is constant. It means first that  $C_1 < 0.2$ mM, because at concentrations lower than C, this ratio should increase due to the presence of unstopped chains. Additionally, it confirms that **DBUT** interacts with **EHUT** through the carbonyl groups and not through the NH groups, because the **DBUT** NH groups are not hydrogen bonded and we know from Figure 3 that **DBUT** is an efficient chain stopper in this concentration range. Furthermore, Figure 5 shows that at concentrations higher than 15mM, the free to hydrogen bonded NH ratio decreases, revealing that the NH functions of the chain stopper **DBUT** can also participate in hydrogen bonding. The butyl

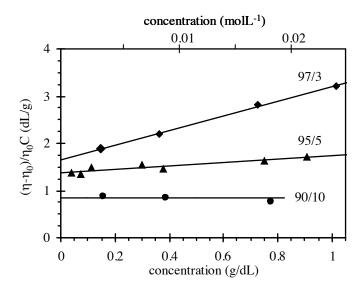
substituents are apparently not bulky enough to completely avoid this additional interaction, making **DBUT** unreliable as a chain stopper at concentrations higher than 15mM.

Consequently, the concentration dependence of the molar mass of 90/10 **EHUT/DBUT** mixtures can be considered to be effectively blocked over the range of concentrations from 0.2mM (or lower) to 15mM.<sup>26</sup>



**Figure 5.** Ratio of absorbances measured at 3472cm<sup>-1</sup> (free) and 3340cm<sup>-1</sup> (hydrogen bonded), versus concentration for **EHUT** solutions (o) and 90/10 **EHUT/DBUT** mixtures (♦), in CCl<sub>4</sub>.

**3.** Characterization of stopped chains. Figure 6 shows that EHUT/DBUT mixtures yield linear plots in the reduced specific viscosity representation classically used in polymer science. The intrinsic viscosity for these mixtures can be deduced by extrapolation to zero concentration and is reported in Table SI1 (Supporting Information). The intrinsic viscosity decreases when the chain stopper content increases, in agreement with the expected shortening of the supramolecular chains.

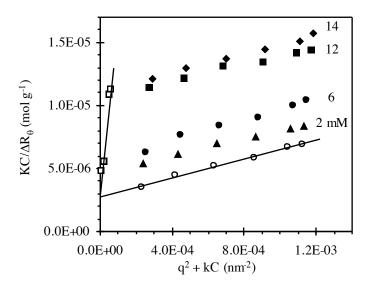


**Figure 6.** Reduced specific viscosity of 97/3, 95/5 and 90/10 **EHUT/DBUT** mixtures, versus overall concentration in CCl<sub>4</sub>.

Static light scattering is used in polymer science to measure the weight average molar mass by extrapolation to zero of both concentration and measuring angle. Figure 7 shows the classical Zimm representation used for this double extrapolation, in the case of 99.5/0.5 **EHUT/DBUT** mixtures. As in the case of non-aggregating covalent polymers, the plots are linear. Although the linearity of these data (light scattering and viscosity) does not prove in itself that the molar mass of the chains is constant, a non-linearity of the plot would be a clear sign of molar mass evolution with concentration. The characteristic information that can be derived from the Zimm plot (weight average molar mass  $(M_*)$ , radius of gyration  $(R_*)$  and second virial coefficient  $(A_*)$ ) are reported in Table 1. The values of  $M_*$  and  $R_*$  decrease significantly when the chain stopper content increases, again in agreement with the expected shortening of the supramolecular chains. The value of  $A_*$  seems to be unaffected by this shortening, and can thus be considered to be a characteristic of the

structure of **EHUT** chains in this solvent. More quantitatively, the measured values of M<sub>\*</sub> can be compared to the expected value for the number average molar mass (M<sub>\*</sub> in Table 1), calculated by assuming that each supramolecular chain contains one chain stopper. The ratio between measured M<sub>\*</sub> and expected M<sub>\*</sub> lies between 9 and 13. These large values can result from two non-exclusive effects. First, it is likely that the polydispersity of these systems is larger than 2, due to the non-isodesmic nature of the self-association of **EHUT**. Indeed, it has been shown that the polydispersity of **EHUT** supramolecular polymers in chloroform can be as high as 6. Second, we cannot exclude that several chain stopper molecules are necessary to effectively stop one supramolecular chain.

Dynamic light scattering performed on the same solutions reveals that the distributions of relaxation times derived from the correlation functions are characterized by a single relaxation process. In all cases, the characteristic average relaxation times ( $\tau$ ) are  $q^2$ -dependent (where q is the scattering wave vector), meaning that diffusive motions are probed. They can thus be used to calculate apparent diffusion coefficients  $D=(q^2\tau)^4$ . The plot of the apparent diffusion coefficient versus concentration is linear (Figure SII) and affords the diffusion coefficient ( $D_a$ ) by extrapolation to zero concentration. Finally, the hydrodynamic radius of the scattering species has been estimated using the Stokes-Einstein relation:  $R_a = kT/6\pi\eta D_a$ , where k is the Boltzmann constant, T the absolute temperature and  $\eta$  the viscosity of the solvent (Table 1). The high values of the ratio  $R_a/R_a$  are in agreement with the highly anisotropic shape previously deduced from SANS measurements.



**Figure 7.** Static light scattering data (Zimm plot) for 99.5/0.5 **EHUT/DBUT** mixtures, in CCl<sub>4</sub> (k = 0.01 cm<sup>3</sup> g<sup>-1</sup> nm<sup>2</sup>).

Table 1. Parameters deduced from light scattering data for mixtures of EHUT and DBUT in CCl<sub>+</sub>.

EHUT/DBUT <sup>a</sup>	$\mathbf{M}_{ ext{n, calc}}^{b}$	$\mathbf{M}_{\mathrm{w}}{}^{c}$	$R_{g}^{d}$	$A_{2}^{e}$	$\mathbf{R}_{\mathtt{h}}^{f}$
90/10	4.3	57	24	8.0	9
97/3	14.4	173	43	8.8	16
99.5/0.5	86.5	830	63	7.1	51

<sup>&</sup>lt;sup>4</sup> Molar ratio of the 2 compounds.

- <sup>6</sup> Measured weight average molar mass, in kg mol<sup>-1</sup>.
- <sup>4</sup> Radius of gyration, in nm.
- <sup>e</sup> Second virial coefficient, in 10<sup>4</sup> mol g<sup>2</sup> cm<sup>3</sup>.
- <sup>f</sup> Hydrodynamic radius, in nm.

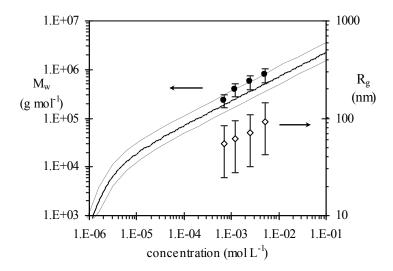
<sup>&</sup>lt;sup>b</sup> Calculated number average molar mass, assuming that each supramolecular chain contains one chain stopper, in kg mol<sup>1</sup>.

**4. Molar mass measurements in stopper free solutions.** The information deduced from these experiments with a chain stopper can then be used to characterize stopper free solutions. For example, the determination of the second virial coefficient ( $A_2$ ) makes it possible to correct finite concentration effects in pure **EHUT** solutions, if the concentration dependence of  $A_2$  is negligible, which seems to be the case here (Table 1). Accordingly,  $M_{w}$  and  $R_{z}$  have been determined for several **EHUT** concentrations (C), from the angle dependence of the Rayleigh ratio ( $\Delta R_{\theta}$ ), through eq 1.

$$\frac{KC}{\Delta R_{\theta}} - 2A_2C = \frac{1}{M_w} \left( 1 + \frac{q^2 R_g^2}{3} \right) \tag{1}$$

Figure 8 shows the results for concentrations below the overlap concentration, which can be estimated to be on the order of 0.01mol.L<sup>-1</sup> (see figure 1 from reference 12).

In order to test the validity of this approach, it is possible to compare the M<sub>\*</sub> data to results obtained independently from calorimetric measurements (ITC). The principle of the technique is to measure the heat evolved while aliquots of a relatively concentrated **EHUT** solution are injected into pure solvent (placed in a calorimetric cell at a fixed temperature). The recently described ITC procedure to determine the association constants of supramolecular polymers was applied to **EHUT** in CCl<sub>4</sub>. The detailed results are provided as Supporting Information. From the association constants, the evolution of M<sub>\*</sub> with concentration was calculated (Figure 8). The close agreement between light scattering and calorimetric data is particularly remarkable, as no fitting parameter is involved.



**Figure 8.** Determination by static light scattering (•) and calorimetry (full curve) of the weight average molar mass for **EHUT**, versus concentration in CCl<sub>4</sub>, at 25°C. The dotted curves represent the uncertainty of the calorimetric data.  $R_s$  ( $\diamond$ ) is determined by static light scattering.

#### Conclusion

The present experimental results show that an efficient chain stopper can actually block the concentration dependence of the molar mass of supramolecular polymers over a realistic concentration range. This property can be used to characterize the stopped supramolecular chains by classical polymer characterization techniques (viscosimetry, light scattering, etc). This in turn makes it possible to derive some information about the supramolecular polymer in the absence of a chain stopper.

### **Experimental Section**

Synthesis. The synthesis of EHUT and BMAUT was previously reported.236

**1,1-dibutyl-3-[3-(3,3-dibutylureido)-4-methylphenyl]urea (DBUT):** to a stirred solution of 2,4-toluene diisocyanate (Fluka, 99%, 3.4mL, 23mmol) in dichloromethane (125mL, distilled over phosphorous pentoxide), dibutylamine (Aldrich, 6.4g, 50mmol) in dichloromethane (75mL) was added under nitrogen. After 24h, the solvent was evaporated and purification was performed by silica gel column chromatography with dichloromethane / ethyl acetate (88/12) as the eluent. Recrystallization in heptane afforded 6.9g of a white solid (69%). m.p. 111°C. H NMR (300 MHz, D<sub>s</sub>-DMSO):  $\delta$  (ppm) = 8.02/7.61 (2s, 2H, Ar-N*H*),  $\delta$  = 7.33 (s, 1H, Ar-*H*),  $\delta$  = 7. 17/6.98 (2d, J = 9.4 Hz, 2H, Ar-H),  $\delta$  = 3.25 (m, 8H, N-C $H_2$ ),  $\delta$  = 2.06 (s, 3H, Ar-C $H_3$ ),  $\delta$  = 1.48/1.28 (m, 16H, C $H_2$ ),  $\delta$  = 0.90 (t, J = 7.1 Hz, 12H, C $H_3$ ). C NMR (75 MHz, D<sub>s</sub>-DMSO):  $\delta$  (ppm) = 155.6/155.2 (C=O),  $\delta$  = 138.4/137.9/129.2/126.3/118.1/116.4 (Ar),  $\delta$  = 46.0/45.9 (N-C $H_3$ ),  $\delta$  = 30.4/19.5/17.2 ( $CH_3$ ),  $\delta$  = 13.8/13.7 ( $CH_3$ ). Anal. Calcd for C<sub>8</sub>H<sub>4</sub>N,O<sub>5</sub>: C, 69.40; H, 10.25; N, 12.95; O, 7.40. Found: C, 68.86; H, 10.30; N, 12.85; O, 7.99.

**Characterizations.** Carbon tetrachloride was used as received. Solutions were prepared at least 1 day prior to the measurements.

**Viscometry.** Solutions were filtered through Millex filters (porosity 0.45  $\mu$ m). Measurements were performed at 25  $\pm$  0.1°C with a Cannon-Manning semi-micro capillary viscometer.

**IR spectroscopy.** Infrared spectra were recorded at room temperature on a Nicolet Avatar 320 spectrometer in KBr cells of 0.05 to 2.5 cm path length.

**Light scattering.** Static and dynamic light scattering measurements were performed with an experimental set up previously described,<sup>27</sup> at a wavelength ( $\lambda_0$ ) of 514.5 nm, and a temperature of 25°C. Solutions were filtered through Whatman Anotop filters (porosity 0.2  $\mu$ m). The specific refractive index increment for **EHUT** in CCl<sub>4</sub> has been measured by differential refractometry (dn/dC= 0.0756 mL/g).

Supporting Information Available: Viscosimetric, scattering and calorimetric data.

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### **Graphic Table of Content**

