



HAL
open science

Advancements on the molecular design of nanoantibiotics: current level of development and future challenges

Roxana Jijie, Alexandre Barras, Florina Teodorescu, Rabah Boukherroub, Sabine Szunerits

► To cite this version:

Roxana Jijie, Alexandre Barras, Florina Teodorescu, Rabah Boukherroub, Sabine Szunerits. Advancements on the molecular design of nanoantibiotics: current level of development and future challenges. *Molecular Systems Design & Engineering*, 2017, 2 (4), pp.349-369. 10.1039/C7ME00048K . hal-01686819

HAL Id: hal-01686819

<https://hal.science/hal-01686819v1>

Submitted on 17 Jan 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Advancements on the molecular design of nanoantibiotics: current level of development and future challenges

Authors: Roxana Jijie^a, Alexandre Barras^a, Florina Teodorescu^b, Rabah Boukherroub^{*a} and Sabine Szunerits^{*a}

^a Univ. Lille, CNRS, Centrale Lille, ISEN, Univ. Valenciennes, UMR 8520, IEMN, F-59000 Lille, France. E-mail: sabine.szunerits@univ-lille1.fr

^b Institute of Physical Chemistry “Ilie Murgulescu”, Romanian Academy, 060021 Bucharest, Romania

First published: 10/07/2017

DOI: [10.1039/C7ME00048K](https://doi.org/10.1039/C7ME00048K)

Abstract.

Numerous antimicrobial drugs have been developed and commercialized to kill and inhibit the growth of pathogenic microbes. The therapeutic efficiency of these drugs has however become often inadequate for the treatment of microbial infections, the reasons being multiple. Oral administration results often in only partial absorption and up to 50% of the active compound can be excreted in the urine. Furthermore, antibiotic therapy is also frequently accompanied by gastrointestinal complications, including vomiting, nausea and diarrhea. From a therapeutic point of view, the low solubility of antibiotics in lipid membranes presents a limitation for rapid and efficient treatment of infections. The preferred route for the delivery of antimicrobial drugs is the oral one; oral formulations capable of extending the duration of drug delivery and minimizing side effects have received much attention. The combination of antimicrobial agents with nanomaterials has been seen as a particularly promising strategy to overcome these limitations. Particulate formulations have proven their efficiency in achieving better pharmacokinetic profiles and improving the bio availability of several antibiotics. Antibiotic modified particles have shown their capability to protect some of the antimicrobial drugs from stomach acid and first-pass metabolisms in the gastrointestinal tract. Likewise, particle formulations can increase the circulation times of a drug and the local concentration gradient across absorptive cells. The present review describes the current status of different types of antibiotic loaded nano-systems. Key principles such as antibiotic loading, the release mechanism and the mode of action involved in pathogen killing or inhibition will be discussed in more detail. It is hoped that the general knowledge obtained here will help in the generation of advanced nanomaterials loaded with antimicrobials agents.

1. Introduction

Infectious microorganisms spread through the body via the circulatory system. They are normally removed from the blood stream by macrophages, which are present in all major organs such as the liver, spleen, bone marrow, etc. However, many microorganisms are able to escape the macrophage digestion process, making intracellular infections difficult to eradicate. Treatment with antimicrobial agents, in most cases antibiotics, is currently the only viable strategy to kill and inhibit further growth of pathogenic microorganisms. Typically, antimicrobials kill bacteria by binding to some vital compounds of bacterial metabolism or impeding normal cellular activities. For instance, β -lactams such as penicillin and cephalosporin inhibit bacterial cell wall synthesis; tetracyclines, macrolides, clindamycin, metronidazole and quinolones inhibit protein synthesis; sulphonamides and trimethoprim have an inhibitory effect on enzyme synthesis. Despite the great progress made in antimicrobial development, many infectious diseases remain difficult to treat. One major reason is that many antimicrobials are difficult to transport through cell membranes and have low activity inside the cells. Toxicity to healthy tissues and solubility issues are additional limitations for their use in large quantities. Another current issue when using antibiotics stems from acquired resistance from infectious microbes towards these drugs. To address these issues, alternative antimicrobial drug systems have been proposed, one being antibiotic loaded nanoparticles. Such “nanoantibiotics” allow altering and improving the pharmacokinetics and therapeutic levels of antimicrobial agents in contrast to the free counterparts. Many advantages of nanoparticle based drug delivery have been recognized, including improved solubility of drugs, prolonged circulation, release of drugs in a controlled and sustained manner, targeted delivery to tissues and cells of interest, as well as concurring delivery of multiple agents for combined therapy. This is also applicable to antimicrobial nanosystems. Moreover, antimicrobial loaded nanoparticles can enter host cells through endocytosis and then release the antibiotic load to treat intracellular infections. As a result, a number (<40) of nanoparticle-based systems are approved for clinical trials and are under different stages of clinical tests. Most of the clinically approved intravenous nanoparticle therapies are targeted for the treatment of various cancer types, including solid malignancies, ovarian, breast, leukemia, prostate, liver, lymphomas, bladder, pancreatic, etc. When it comes to the treatment of bacterial infections using nanoantibiotics, the only clinically approved system is AmBisome by Gildeas Science for the treatment of serious fungal infections and leishmaniasis ([Table 1](#)).¹ CAL02, with sphingomyelin and cholesterol loaded on liposomes for toxin neutralization, is under Ph I clinical trial against pneumonia infections. No antibiotic encapsulated nanosystem is under trial currently.

Table 1. Nanoparticle-based delivery systems clinically approved or under clinical trial against infection

Particles	Active agent	Name	Application
Liposomes	Sphingomyelin and cholesterol	CAL02 (Combioxin SA)	Ph I, pneumonia infections
Liposomes	Amphotericin B	AmBisome (Gildeas Science)	Clinically approved treatment of invasive fungal infections

While numerous studies for tumor-enhanced drug delivery have been performed, the corresponding research on the treatment of bacterial infections has just really started. One

sufficient challenge in nanomedicine translation is the contraction between prolonged circulation time achieved by the integration of PEG and other hydrophilic polymers, and targeted delivery to pathogens mostly requiring nanoparticles with a hydrophobic and positively charged surface. The negatively charged walls of many types of bacteria interact with the positively charged nanostructures through electrostatic interactions and results in permeability changes or even the complete destruction of the cell wall and thus the pathogen itself.

This review focuses on more recent developments of antibiotic-loaded nanostructures including liposomes, polymeric nanoparticles and dendrimers, metallic and magnetic nanoparticles, carbon-based nanostructures and mesoporous silica nanostructures (Fig. 1). Since liposomes were proposed as nanostructures for antibiotic delivery about forty years ago, the different nanotechnologies employed for increasing the bioavailability of antibiotics, thus allowing targeted antibiotic therapy, have been reviewed in several reports.^{2–10} The increase in public awareness of the emergence of antibiotic resistance and unsolved problems in treating bacterial infections makes a more timely review of importance. This feature article focuses in addition not only on the road map of antimicrobial lipid and polymer nanoparticles, which have been greatly covered by two recent review articles,^{4,9} but presents our vision of the progress in the use of metallic, magnetic and carbon-based nanostructures such as C-dots and graphene for antibiotic delivery and bacterial treatment. The characteristic features of different carbon-based nanostructures such as structural, optical, thermal and mechanical features, make them promising nanostructures for future antibacterial applications.

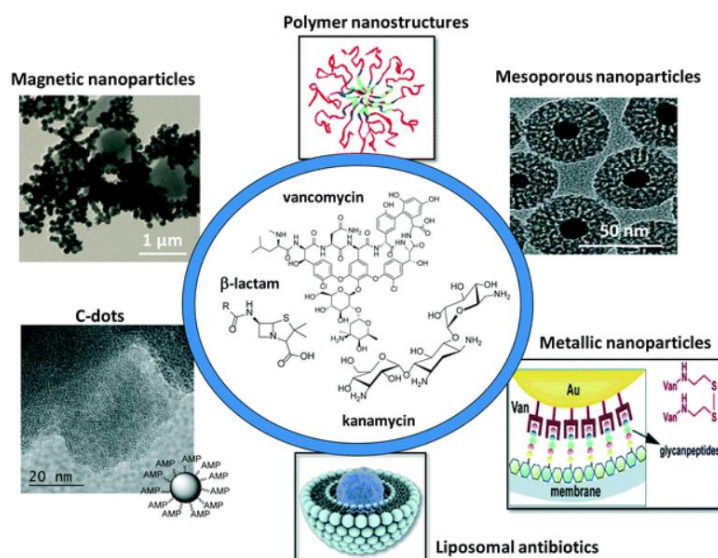


Figure 1.

Schematic illustration of the different nanoparticle platforms for antimicrobial drug loading and release.

2. Antibiotics: one of many antimicrobial agents

Antibiotics, a class of antimicrobial agents, used in the treatment and prevention of bacterial infections, have been considered as one of the wonder discoveries of the 20th century. Although there are well over 100 antibiotics on the market, the majority come

3

from only a few types of drugs (Table 2). Their mode of action (Fig. 2A) differs due to the nature of their structure and degree of affinity to certain target sites within bacterial cells.¹¹ Penicillin, cephalosporin, bacitracin and vancomycin are four antibiotics targeting the cell wall of bacteria by inhibiting cell wall synthesis. Cationic peptides such as colistin, on the other hand, are inhibitors of cell membrane functions. Membranes are important barriers that segregate and regulate the intra- and extracellular flow of substances. A disruption or damage to this structure results in leakage of important solutes essential for the cell's survival. Because this structure is found in both eukaryotic and prokaryotic cells, the action of this class of antibiotics is often poorly selective and can often be toxic for systemic use in the mammalian host. Most clinical usage is therefore limited to topical applications. Aminoglycosides, macrolides, and tetracyclines are inhibitors of protein synthesis, an essential process necessary for the multiplication and survival of all bacterial cells. These antibacterial agents target bacterial protein synthesis by binding to either the 30S or 50S subunits of the intracellular ribosomes. This activity then results in the disruption of the normal cellular metabolism of the bacteria, and consequently leads to the death of the organism or the inhibition of its growth and multiplication. Another mode of action is based on the inhibition of nucleic acid synthesis with quinolones, metronidazole and rifampin as examples. Other antibiotics act on selected cellular processes essential for the survival of the bacterial pathogens: sulfonamides and trimethoprim disrupt the folic acid pathway, which is a necessary step for bacteria to produce precursors important for DNA synthesis. Sulfonamides target and bind further to dihydropteroate synthase, while trimethoprim inhibit dihydrofolate reductase; both enzymes are essential for the production of folic acid, a vitamin synthesized by bacteria.

Table 2. Classification of antibiotics according to their mode of action

Antibiotic class	Examples	Target/mode of action
β -Lactams	Penicillin (ampicillin), cephalosporins (cephamycin), penemes (meropenem), monobactams (aztreonam)	Inhibiting cell wall synthesis
Aminoglycosides	Gentamicin, streptomycin, spectinomycin, and tobramycin	Inhibitors of protein synthesis
Glycopeptides	Vancomycin, teicoplanin	Inhibiting cell wall synthesis
Macrolides	Erythromycin, clarithromycin, azithromycin	Inhibitors of protein synthesis
Fluoroquinolones	Ciprofloxacin, levofloxacin, ofloxacin	Inhibition of nucleic acid synthesis
Sulfonamides	Co-trimoxazole and trimethoprim, sulfamethoxazole	Act on selected cellular processes and metabolism
Tetracyclines	Tetracycline and doxycycline	Inhibitors of protein synthesis
Cationic peptides	Colistin	Cell membrane functions

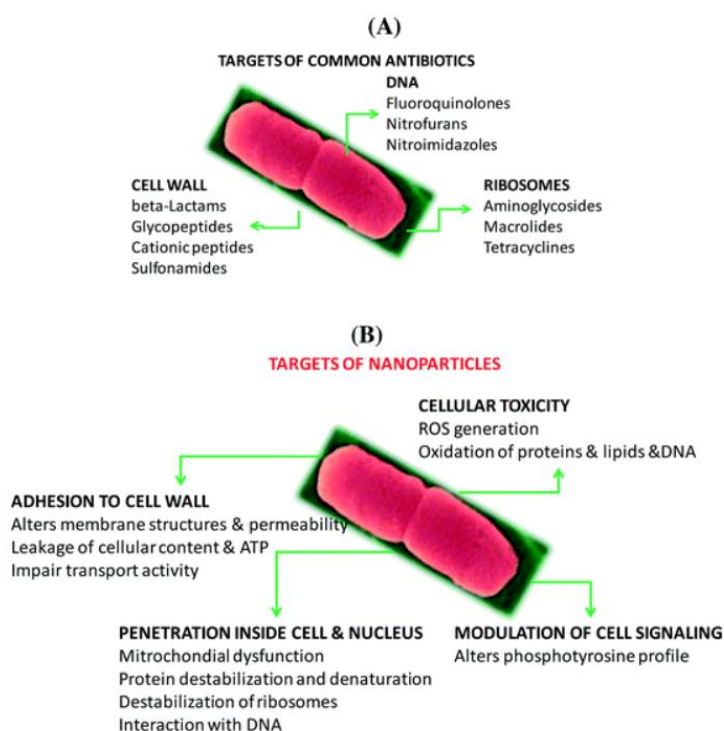


Figure 2 : Major targets of common antimicrobial agents (A) and nanoparticles (B).

Regrettably, the over and misuse of most of these drugs have been accompanied by the rapid appearance of resistant strains. A wide range of biochemical and physiological mechanisms may be responsible for resistance and are well described in some review articles; the reader is referred to them.^{12,13} Resistant strains of Gram-positive bacteria (such as methicillin-resistant *Staphylococcus aureus*, MRSA) pose a continuing threat. A range of antibiotics available to combat them remains comparatively robust. It is the emergence of resistant Gram-negative strains that currently presents the greater threat, in particular *Pseudomonas aeruginosa*, *Enterobacter cloacae* and *E. coli*. These bacteria can be deadly, causing infections from urinary tract infections (UTI) to life threatening pneumonias and bloodstream infections. As it is harder for antibiotics to penetrate Gram-negative bacterial cell walls, these pathogens are particularly difficult to eradicate and a very few antibiotics can currently combat them. This situation might be different when using nanostructures.

In the case of nanostructures, a wide range of pathogen inactivation mechanisms is possible depending greatly on the composition of the nanoparticles, their size, as well as surface charge and surface function (Fig. 2B). Next to cellular dysfunctions due to the interaction of the particles with the microbial cell membrane, changing permeability and transport activity, particles may also penetrate inside the cells and affect vital cellular functioning.¹⁴ Electrostatic interactions of metallic nanoparticles with sulfur-containing proteins, present in the cell wall, can cause irreversible changes in the cell wall structure, resulting in its disruption which in turn affects the integrity of the lipid bilayer and permeability of the cell membrane. An increase in membrane permeability affects a cell's ability to properly regulate transport activity through the plasma membrane and results in the leakage of cellular contents, including ions, proteins,

reducing sugars and sometimes the cellular energy reservoir, ATP. The thickness and composition of the cell wall of the microorganisms play an important role in the way nanoparticles interact (Fig. 2C). Gram-negative bacteria, such as *E. coli*, are more susceptible to cell membrane damage than Gram-positive bacteria, such as *S. aureus* due to the difference in the organization of a key component of the cell membrane, the peptidoglycans. In Gram-positive bacteria, the cell wall is composed of a negatively charged peptidoglycan layer of about 30 nm in thickness, while Gram-negative bacteria exhibit peptidoglycan layers of ~3–4 nm in thickness. Gram-negative bacteria contain on the other hand lipopolysaccharides (LPS) in the cell membrane, which contribute to the structural integrity of the membrane as well as membrane protection from chemical attacks. The negative charge of LPS together with the thinner peptidoglycan layer promotes adhesion of Ag NPs and makes bacteria more susceptible to antimicrobial therapy. Other modes of action of particles are based on blocking the sugar metabolism or increasing the cellular oxidative stress in the microbes. Indeed, oxidative stress is an indication of toxic effects caused by heavy metal ions, such as Ag⁺. Increased concentrations of metal ions are expected to cause an increase in cellular oxidative stress with the ability to produce reactive oxygen species (ROS) such as hydrogen peroxide (H₂O₂), the superoxide anion (O²⁻), hydroxyl radical (OH[·]), and singlet oxygen (¹O₂). Reactive oxygen species can also be generated intracellularly during mitochondrial oxidative phosphorylation. As the cycle of phosphorylation and dephosphorylation is essential for microbial growth and cellular activity, examining the phosphotyrosine profile of bacterial proteins from both Gram-positive and Gram-negative bacteria offers a useful way to study the effect of particles on their mode of action.

While a variety of nanostructures displaying intrinsic antibacterial properties have been identified (Ag NPs, Cu NPs, ZnO NPs, etc.) using one or several of the targeting mechanisms outlined in Fig. 2B, the focus of this review is directed only on the effect of antibiotic loaded nanostructures. While the development of bacterial resistance is common to all antibiotics, some previous studies showed that the activity of aminoglycosidic antibiotics (streptomycin, gentamycin, neomycin) is protected when immobilized on gold nanoparticles.¹⁵ One reason for this result might be due to the multivalent presentation of antibiotic ligands on the nanoparticles. As demonstrated for carbohydrate modified nanoparticles,^{16–21} a single particle with a large surface to volume ratio is ready for the attachment of multiple ligands, providing an easy and powerful possibility for enhancing the affinity and/or increasing locally the concentration of the ligand.

The formulation of different antibiotic modified nanostructures will thus be the focus of the rest of the review.

3. Different nanoantibiotic formulations

3.1. Liposomes

Liposomes are spherical vesicles made of one or more phospholipid bilayers with a diameter varying between 20 nm and 10 μm depending on the type of phospholipid used.²² Hydrophobic substances can be loaded inside the lipid bilayer, while hydrophilic compounds are entrapped in the aqueous core. For the entrapment of hydrophilic elements, large unilamellar vesicles (LUVs) are mainly used as the volume of encapsulated water is high. Multilamellar vesicles (MLVs) and in particular small unilamellar vesicles (SUVs) of ≈100 nm are more adapted for the integration of hydrophobic compounds such as antibiotics. The basic steps involved in the

preparation of antibiotic loaded liposomes consist of drying down the lipids using an organic solvent, dispersing the lipid in aqueous media, down-sizing the hydrated suspension by sonication or extrusion followed by purification.²³

Liposomes have become universal drug carrier systems due to the easy manner of manipulating their physico-chemical properties, enabling the design of nanocarriers with the desired pharmacokinetic properties.⁷ Numerous studies have focused on the entrapment of antibiotics in liposomes with the aim to enhance their antibacterial activity and to reduce toxic side effects during antibiotic therapy. The advantages of liposomal antibiotics include an improved plasma circulation time of the antibiotic when encapsulated in polyethylene glycol (PEG)-coated (STEALTH) formulations. The PEG layer protects the vesicles and allows a long liposome circulation in the blood system.²⁴ The possibility of adding targeting ligands to these nanocarriers allowed not only targeted treatment, but also improved the activity of antibiotics against extracellular pathogens and eradication of bacterial structures such as biofilms. In the area of antimicrobial resistance with a limited number of new antibiotics coming onto the market, improving currently available antibiotics upon the design of liposomal formulations is one way to overcome some of these resistance related issues. Promising results were for example reported for the eradication of drug-resistant *Pseudomonas aeruginosa* due to a direct interaction/fusion between liposomes and their bacterial cells.^{25,26}

Most studies regarding liposomal antibiotics deal with aminoglycosides, quinolones, polypeptides, and betalactams. Indeed, the first antibiotic to be entrapped was penicillin.²⁷ It was shown that entrapment extends indeed the rate of penicillin elimination from the plasma. Some years later, the capacity of dihydrostreptomycin (DHS) sulfate (a derivative of streptomycin, a semisynthetic aminoglycoside antibiotic) loaded liposomes formed from a mixture of egg phosphatidylcholine, cholesterol and phosphatidic acid in a 7:2:1 molar ratio to enhance the antimicrobial action towards *S. aureus* compared to the free drug was shown.²⁸ Multilamellar negatively charged liposomes (lipid bilayer vesicles) composed of phosphatidylcholine, cholesterol, and phosphatidylserine (molar ratio, 6:3:1) were produced by Desiderio and Campbell to entrap an aqueous solution of cephalothin.²⁹ This formulation was effective in the elimination of *Salmonella typhimurium* in infected macrophages. The possibility of entrapping more than one antibiotic was demonstrated in 1983 by Stevenson et al.,³⁰ who showed a 10-fold increase in intracellular antibacterial activity by loading streptomycin and chloramphenicol, within neutral or anionic unilamellar liposomes composed of egg phosphatidylcholine and cholesterol. The encapsulation of piperacillin, a β -lactam, into liposomes prepared with phosphatidylcholine and cholesterol (1:1) allowed the protection of the antibiotic from enzymatic hydrolysis by staphylococcal β -lactamase, resulting in enhanced activity against *Staphylococcus aureus*.³¹ Indeed, the literature is rich and some excellent recent reviews have been devoted to this topic.^{4,5,7} [Table 3](#) lists some more recent examples of liposomal antibiotics. The formulation of chloramphenicol in liposomes was recently proposed as an easy manner to overcome problems associated with the toxicity of the antibiotic.³² Li et al. showed that a liposomal co-delivery of daptomycin and clarithromycin is highly effective and less toxic for the treatment of methicillin-resistant *S. aureus* infections in the presence of only one-thirtieth of the concentration required in liposomes containing daptomycin only ([Fig. 3A](#)).³³ A liposomal complex containing the phospholipid cardiolipin co-entrapped with levofloxacin was formulated by Gaidukevich et al. to suppress the growth of *Mycobacterium tuberculosis* and reduce the minimum inhibitory concentration of cardiolipin (33.5 μM) and levofloxacin up to 2 $\mu\text{g ml}^{-1}$.³⁴ Antimicrobial activities of herbal extracts with lysozyme were evaluated against two Gram-positive (*B. subtilis* and *M. luteus*) and two Gram-negative (*E. coli*

and *S. marcescens*) bacteria by Matouskova et al.³⁵ The integration of selective antimicrobial peptides such as S-thanatin onto liposomes with encapsulated levofloxacin via a lipid PEG linker was recently reported by Fan et al.³⁶ (Fig. 3B). The positively charged nanostructures showed a strong interaction with bacterial cells with an increased uptake of levofloxacin. Indeed, the TS–liposome–levofloxacin (TS-LPs-LEV) complex exhibited excellent activity against *Klebsiella pneumoniae* ATCC 700603 and restored the susceptibility of multidrug-resistant *K. pneumoniae* clinical isolates to levofloxacin in vitro.

Table 3: Liposomal antibiotics used for bacteria eradication in the past 10 years

Active lipids	Antibiotics	Bacteria	Ref.
DC-Chol	Ciprofloxacin	<i>P. aeruginosa</i>	37 and 38
DC-Chol, Chol, DOPC, DOPE	Ampicillin, penicillin G	<i>E. coli</i> XL1 Blue strain	39
Soybean lecithin, Chol (chitosan)	Vancomycin hydrochloride	—	40
Chol, HSPC (mPEG ₂₀₀₀ -DSPE and chitosan)	Daptomycin, clarithromycin	MRSA	33
Commercial Lipoquin (ARD-3100)	Ciprofloxacin	<i>Francisella tularensis</i> strain Schu S4	41
Egg PC, Chol	Gentamicin	MRSA	42
DSPC, DSPG or Egg PC, DOPA	Doxycycline	<i>H. pylori</i>	43 and 44
DPPC, DOPC, DPPG	Tobramycin	<i>Burkholderia cepacia</i> complex	45
DPPC, DDAB, DCP, Chol	Clarithromycin	<i>P. aeruginosa</i> , <i>B. subtilis</i>	46
DPPC	Polymyxin B	<i>P. aeruginosa</i>	47
DPPC, POPC, Chol	Polymyxin B	<i>B. bronchiseptica</i> , <i>P. aeruginosa</i> , <i>E. coli</i> , <i>K. pneumoniae</i> , <i>A. lwoffii</i> , <i>A. baumannii</i>	48
DOPC, Chol	Colistin	—	49
DSPC, DCP, DMPG	Vancomycin	MRSA	50 and 51
PHEPC, Chol PEG-DSPE	Clarithromycin	<i>M. avium</i>	52
DPPC, Chol, cholesteryl tetraethyleneglycol fucose	Ampicillin, metronidazole	<i>H. pylori</i>	53
DPPC, Chol, (DSPE-PEG-Mal)	Gentamicin	<i>E. coli</i> , <i>P. aeruginosa</i> , <i>S. aureus</i>	54
PC/Chol + chitosan modified Au NPs	Vancomycin	MRSA	55

Chol: cholesterol; DC-Col: 3b-[N-(N',N'-dimethylaminoethane)-carbamoyl]cholesterol; DCP: dicetyl phosphate; DDAB: didecyldimethylammonium bromide; DMPG: 1,2-dimyristoyl-sn-glycero-3-phosphorylglycerol sodium salt; DOPC: dioleoylphosphatidylcholine; DPPC: dipalmitoylphosphatidylcholine; DPPG: 1,2-dipalmitoyl-sn-glycero-3-phosphorylglycerol sodium salt; DOPA: 1,2-di-(9Z-octadecenoyl)-sn-glycero-3-phosphate; DOPE: 1,2-dioleoyl-sn-glycero-3-phosphoethanolamine; DSPC: 1,2-distearoyl-sn-glycero-3-phosphocholine; DSPE: 1,2-distearoyl-sn-glycero-3-phosphoethanolamine; DSPG: 1,2-distearoyl-sn-glycero-3-phosphoglycerol; HSPC: hydrogenated soy phosphatidylcholine; Mal: maleimide; PC: phosphatidylcholine; PHEPC: partially hydrogenated egg phosphatidylcholine; POPC: 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine.

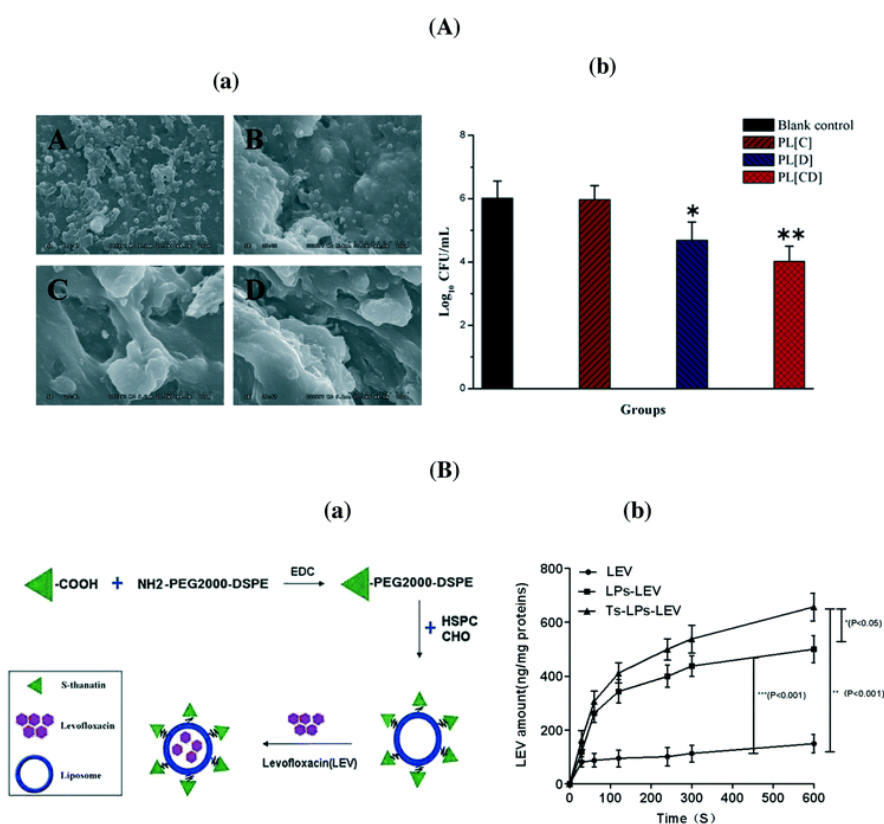


Fig. 3 (A) (a) MRSA 252 biofilm survival and morphology examined after seven days. (A) No treatment; (B) PEGylated liposome with clarithromycin, (C) PEGylated liposome with daptomycin; (D) PEGylated liposome with clarithromycin + daptomycin; (b) a comparison of bacterial survival in the presence of the indicated formulations; (B) (a) formulation of S-thanatins antibiotic liposomal formulation, (b) intra-bacterial levofloxacin accumulation of levofloxacin in *K. pneumoniae* ATCC 700603 (reprinted with permission from ref. 36).

Despite their common use, one of the main drawbacks of liposomal antibiotics is the short shelf-life of lipid vesicles which limits drug stability. This is mainly due to the hydrolysis of ester bonds or the oxidation of unsaturated acyl chains of the lipids used in their formulation. Passive

drug leakage is another issue to be controlled. Furthermore, the encapsulation process has an important influence on the therapeutic utility of the nanocarriers and liposomal antibiotics are only useful if a therapeutic amount of drug and a reasonable amount of lipids is used, as high doses of lipids can be toxic.⁵⁶

3.2. Polymer nanoparticles and hydrogels

To overcome some of the problems related to liposomal antibiotics, polymer particles loaded with antibiotics were developed. Polymer-based nanoparticles are submicron-sized colloidal particles in which the antibiotic can be embedded or encapsulated within their polymeric matrix or adsorbed or conjugated onto their surface.⁵⁷ Such particles are composed of biodegradable or biocompatible materials such as poly(lactic acid) (PLA), poly(lactic-co-glycolic acid) (PLGA), chitosan, gelatin, alginate, etc. and have been extensively explored as means for drug solubilization, stabilization and targeting.^{4,58} Compared to the free drug, polymeric particles have many advantages including improved drug bioavailability, higher carrier capacity, protection of the antibiotic against enzymatic and hydrolytic degradation, the ability to release the antibiotic in a controlled behavior and the capability to adapt to different routes of administration. By virtue of their enhanced permeability, they can concentrate in inflamed and infected locations. Furthermore, the advantages of polymeric particles are their high stability, controlled synthesis (size, surface charge, the presence of functional groups), and drug release profiles that can be accurately tuned by selecting the appropriate polymer. Some excellent review articles about polymeric nanoparticles have been published recently.^{4,9,59,60} Table 4 summarizes some of the recent developments in antibiotic loaded polymeric nanoparticles, including hydrogels to fight infections.

Table 4 Examples of polymeric nanoparticles and hydrogels conjugated with antibiotics from the past three years

Particles	Antibiotics	Bacterial strain	Ref.
PLGA	Cefazolin, ciprofloxacin, colistin, clindamycin, doxycycline, vancomycin	S. aureus, P. aeruginosa, E. coli	61
PLGA	Amikacin	P. aeruginosa	62
PLGA	Ciproflaxin	P. aeruginosa, S. aureus	63
PLGA/CAMP	Colistin	P. aeruginosa	64
PLGA/polypyrrole	Vancomycin	S. aureus	65 , 66
PLGA/PCL	Vancomycin, rifampin, linezolid, daptomycin	S. aureus	
POEGMA-b-PVA	Gentamicin	P. aeruginosa	67
PEG/PAA	Amikacin, colistin	S. aureus, E. coli	68
PEG-PCL-PAE	Vancomycin	S. aureus	69
p(DMAEMA-BMA-PAA)	Farnesol	Streptococcus mutans	70
Alginate/chitosan	Tobramycin	P. aeruginosa	71
Carboxymethyl-chitosan	Fentamycin sulfate (GS)	S. aureus	72

Core-shell supramolecular gelatin nanoparticles (SGNPs)	Vancomycin	S. epidermidis, S. aureus, E. coli, P. vulgaris, S. marcescens, P. aeruginosa	73
Polymeric triple-layered nanogel	Vancomycin	S. aureus	74
Mannose-modified hydrogel	Vancomycin	MRSA	75
Dendritic-based hydrogels	Amoxicillin	—	76
Chitosan-based hydrogel	Gentamycin sulfate	S. aureus	72
PEG/PAA	Amikacin, colistin	S. aureus, E. coli	68
PLGA-PLH-PEG	Vancomycin	S. aureus	77
PEG-b-PP, PEG-b-PC	Vancomycin, gentamicin	S. aureus	78
PAA/gelatin-based hydrogel	Gentamycin sulfate, vancomycin hydrochloride	—	79

BMA: butyl methacrylate; CAMP: cyclic adenosine monophosphate; DMAEMA = 2-(dimethylamino)ethyl methacrylate, PAA: 2-propylacrylic acid; PAE: poly(beta-amino ester); PCL: poly-caprolactone; PLGA: poly(lactic-co-glycolic acid); PLH: poly(L-histidine); PC: phenyl cephalosporin, PP: phenylacetic acid, PEG: poly(ethylene glycol); PEO: poly(ethylene oxide); POEGMA = poly(oligo(ethylene glycol) methacrylate); PQDMA: poly(2-dimethylamino)ethyl methacrylate; PVA: poly(vinyl alcohol).

PLGA particles containing colistin as well as cationic antimicrobial peptides were prepared by d'Angelo et al. using an emulsion/solvent diffusion technique.⁶⁴ Post-coating with chitosan and PVA allowed modulating surface properties and improving the transport of the nanosystem through artificial mucus; the particles displayed excellent efficacy in biofilm eradication compared to free colistin. Co-delivery of nitric oxide and gentamicin from polymeric nanoparticles was proposed recently by Nguyen et al. as efficient means for reducing the viability of *P. aeruginosa* biofilms by more than 90%.⁶⁷ The group of Scott developed tobramycin encapsulated polymer nanostructures, which showed the capability to penetrate the thick DNA-rich mucus on people with a *P. aeruginosa* pulmonary infection.⁷¹ Ciproflaxin loaded poly(lactic-co-glycolic acid) nanoparticles modified with DNase I were investigated by Torrents and co-workers for the eradication of established biofilms.⁶³ While the particles were not able to prevent biofilm formation, from planktonic bacteria, they successfully reduced biofilm mass and size (Fig. 4A). Repeated administration over 3 days reduced 95% and eradicated more than 99.8% of established biofilms.⁶³ Polymeric nanofibers, composed of PLGA and PCL, were developed by Ashbaugher et al. for local co-delivery of a combination of antibiotics.⁸⁰ The release can be adjusted by varying the PLGA/PCL polymer ratio and proved to be highly effective in preventing medical device infections in patients. Benoit and co-workers synthesized positively charged nanoparticles from diblock copolymers composed of 2-(dimethylamino)ethyl methacrylate (DMAEMA), butyl methacrylate (BMA) and 2-propylacrylic acid (PAA) for loading with farnesil, a hydrophobic antibacterial drug.⁷⁰ A pH dependent release was observed, resulting in the disruption of *Streptococcus mutans* biofilms 4-fold more efficiently than free farnesol (Fig. 4B). These particles also efficiently attenuated biofilm virulence in vivo using a clinically relevant topical treatment regime in a rodent dental carrier disease model.⁷⁰

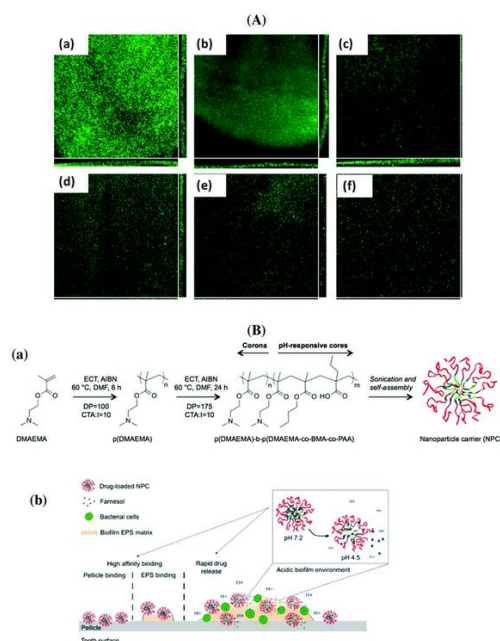


Fig. 4

(A) Flow cell analysis of *P. aeruginosa* biofilm formation in the absence (a) and when treated with free ciproflaxin (b), free ciproflaxin + DNase I (c), PLGA–ciproflaxin (d), PLGA–ciproflaxin + PLGA–DNase I (e) and PLGA–ciproflaxin–DNase I (f) (reprinted with permission from [ref. 63](#)); (B) (a) structure of diblock copolymers used in two-step RAFT polymerizations, (b) proposed mode of action of pH-responsive polymeric nanoparticles for prevention and/or treatment of biofilms (reprinted with permission from [ref. 70](#)).

In order to eliminate the adverse side effects due to premature and nonspecific antibiotic release, considerable efforts have been devoted to the development of ‘on-demand’ release strategies. In this context, hydrogels have attracted a great deal of attention as a matrix for the controlled delivery of antibiotics. Li et al. developed an efficient and specific antibiotic delivery system based on core–shell supramolecular gelatin nanoparticles (SGNPs).⁷³ The surface of the SGNPs was decorated with red blood cell (RBC) membranes and then vancomycin (Van) was encapsulated in the hybrid matrix (Van@SGNPs@RBC). After reaching the infection sites, the RBC membranes on the nanosystem reduce the clearance by the immune system and act as detoxifiers by absorbing the exotoxins produced by bacteria, while the gelatin core is degraded by the gelatinase and the encapsulated vancomycin is released and kills the local bacteria. In vitro studies proved that Van@SGNPs@RBC were internalized by macrophage cells and displayed negligible toxicity towards human embryonic kidney and human hepatocyte cell lines. The confocal microscopy images ([Fig. 5A](#)) revealed that the Van@SGNPs@RBC exhibited higher antibacterial activity against *S. aureus* than toward *S. epidermidis* with a MIC value of $3 \pm 0.8 \mu\text{g mL}^{-1}$ and no inhibition effect for Gram-negative bacteria.⁷³

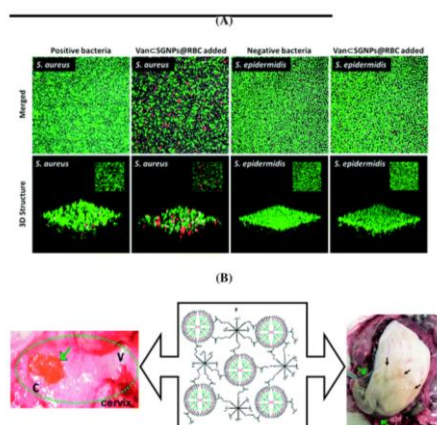


Fig. 5 (A) Antibacterial efficiency of VanC-SGNPs@RBC against *S. aureus* and *S. epidermidis* (reprinted with permission from [ref. 73](#)); (B) dendritic-based hydrogels (10 wt%) after intravaginal (left) and intracervical (right) application in pregnant guinea pigs (reprinted with permission from [ref. 76](#)).

A bacterial-lipase sensitive polymeric triple-layered nanogel (TLN) as a nanocarrier for vancomycin delivery to bacterial infection sites was proposed by Xiong et al.⁷⁴ They demonstrated that lipase-secreting *S. aureus* bacteria triggered rapid drug release and the released drug was able to kill the intracellular bacterial cells. The same group investigated mannosyl modified nanogels for targeted and activatable delivery of vancomycin to infection sites.⁷⁵ The phosphatase and phospholipase produced by the bacteria can degrade the polyphosphoester core of the nanogel, triggering antibiotic release. The mannosyl-modified gel exhibited excellent inhibition capacity against intracellular and in vivo bacterial infections, using Raw264.7 cells and zebrafish embryo infected with MRSA as models.⁷⁵

Recently, dendritic-based hydrogels showed great potential in the loading and release of therapeutic moieties. Navath and co-workers reported that amoxicillin entrapped into injectable and biodegradable hydrogels obtained by cross-linking of a thiopyridyl functionalized PAMAM dendrimer [(NH₂)₄₉-G₄-(NH-PDP)₁₅] with 8-arm PEG (MW ~ 20 kDa) is a viable therapeutic option to treat ascending genital infections during pregnancy.⁷⁶ The results confirmed that the dendritic-based hydrogels ensured a sustained intravaginal delivery of amoxicillin; additionally, the gel components were located on the epithelial surface and did not cross the fetal membrane ([Fig. 5B](#)). A gentamycin sulfate loaded genipin cross-linked carboxymethyl-chitosan hydrogel was proposed as a simple approach to achieve combined antibacterial efficiency and enhanced osteoblastic cell response, with significant inhibition of bacteria growth and biofilm formation of *S. aureus*. The antibacterial activity of the hydrogel was linked to the disruption of the bacterial wall and membrane.⁷² The design of materials that exhibit antibacterial properties and biocompatibility is of great impact in the biomedical field because biomaterial related infections remain a serious biomedical problem and are expensive and complex to treat.

Recently, dendrimeric macromolecules have been reported to be effective platforms for targeted drug delivery to treat bacterial infections ([Table 5](#)).

Table 5 Dendrimer nanoparticles loaded with antibiotics

Particles	Antibiotics	Bacteria	Ref.
Dendrimers	Ciprofloxacin	E. coli	81
Dendrimers	Vancomycin	S. aureus	82
Lipid-dendrimer NPs	Vancomycin	S. aureus, MRSA	83
Dendrimers	Tobramycin	P. aeruginosa	84
Dendrimers	Ciprofloxacin	S. aureus, E. coli	85
Dendrimers	Erythromycin, tobramycin	S. aureus, E. faecalis, E. coli, A. baumannii, K. pneumoniae, E. cloacae	86
Dendrimers	Norfloxacin	S. aureus, E. coli	87
Dendrimers	Nadifloxacin	E. coli, P. aeruginosa and P. hauseri	88
Dendrimers	Rifampicin	—	89
Dendrimers	Azythromycin	C. trachomatis	90

Choi et al. prepared ciprofloxacin attached to a lipopolysaccharide-binding poly(amidoamine) dendrimer nanoconjugate for bacterial cell wall-targeted delivery of the antibiotic under light irradiation ([Fig. 6A](#))⁸¹ and a vancomycin-conjugated poly(amidoamine) dendrimer for pathogen detection and isolation ([Fig. 6B](#)).⁸² A lipid-dendrimer hybrid nanoparticle (LDHN) system conjugated with vancomycin was synthesized by Sonawane et al.⁸³ to fight methicillin-resistant S. aureus (MRSA) infections. Due to the sustained release of vancomycin from LDHNs, the nanosystem showed prolonged and enhanced antibacterial activity. Reymond et al. reported the synergistic effect of glycopeptide dendrimers (FD2) with tobramycin against P. aeruginosa biofilm. The complex nanostructures displayed an effective biofilm inhibition and degradation properties at a sub-inhibitory concentration of both compounds, 4-fold lower than the FD2 minimal biofilm inhibitory concentration (MBIC) and 5-fold lower than the antibiotic MBIC. More recently, VanKoten et al. reported the enhanced antibacterial activity of fourth generation poly(amidoamine) dendrimers decorated with a quaternary ammonium compound against Gram-positive and Gram-negative bacteria.⁹¹

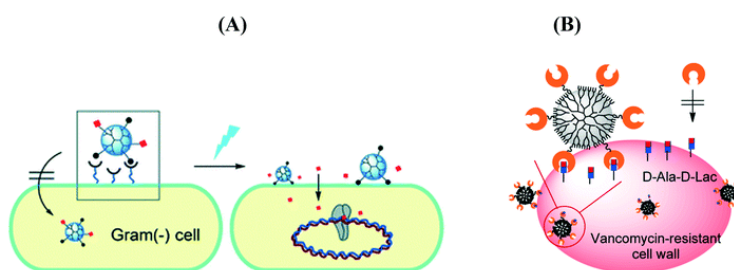


Fig. 6 (A) Light controlled release of ciprofloxacin by a cell wall-targeted dendrimer nanoconjugate (reprinted with permission from [ref. 81](#)); (B) vancomycin-conjugated poly(amidoamine) dendrimer as a nanoplatform for targeting Gram-positive bacteria (reprinted with permission from [ref. 82](#)).

3.3. Gold and silver nanostructures

Metallic nanoparticles (NPs) have generated huge interest in many biomedical fields due to their ease of synthesis and functionalization, and their plasmonic features, allowing the development of theranostic applications.^{92,93} Due to their small size, large surface area to volume ratio, stability over high temperature and translocation into the cells, inorganic nanoparticles are considered as a viable alternative for carrying a large amount of antibiotics and the efficacy of metallic NPs conjugated with antibiotics has thus been the subject of intense investigations. These studies showed that gold nanostructures^{94,95} used in combination with antibiotics exhibit enhanced bactericidal activity on a range of Gram-positive and Gram-negative bacteria and higher stability over an extended storage compared to the free antibiotics (Table 6). While silver nanoparticles (Ag NPs) are bactericidal per se and have proven to be the most effective against bacteria among the metallic nanoparticles,^{96,97} it has been reported that the combined use of Ag NPs with antibiotics results in enhanced and synergetic antimicrobial effects.⁹⁸

Table 6 Metallic nanoparticles conjugated with antibiotics

Particle	Antibiotics	Comments	Bacteria	Ref.
Au NPs	Gentamicin	d = 16 nm, citrate capped	E. coli K12	95
Au NPs	Vancomycin	d = 4–14 nm, biomass coated Au NPs of negative surface charge electrostatic interaction	E. coli, VRSA, VSSA	99
Au NPs	Vancomycin	d < 20 nm, interaction with bis(vancomycin) cystamide via Au–S bonds	E. coli, VRE	100
Au NPs	Cefaclor	d = 22–52 nm, primary amine group of cefaclor acted as both the reducing and the capping agent	S. aureus, E. coli	102
Au NPs	Gentamicin	d = 10–12 nm, electrostatic interaction	S. aureus	105
Au ³⁺ Au NPs	Penicillin G, amoxicillin, methicillin, cephalixin, cefixime, erythromycin, gentamicin, amikacin, tetracycline, ciprofloxacin, clindamycin, nitrofurantoin, nalidixic acid, vancomycin	d = 1.2–17.5 nm, tannin-free black tea reduction, Au ³⁺ coordination complex with available donor groups such as nitrogen, sulfur and phosphorus in antibiotics	P. aeruginosa, E. coli, S. aureus	108
Au NPs	Streptomycin, gentamycin, neomycin	d = 12–15 nm, citrate capped, electrostatic interactions	S. aureus, M. luteus, E. coli, P. aeruginosa	109
Au NPs	Ampicillin, streptomycin, kanamycin	d = 14 nm, NaBH ₄ reduction in the presence of antibiotics	E. coli DH5 α ; M. luteus, S. aureus	110

Au NPs	Ciprofloxacin	d = 4 and 20 nm, nitrogen atom of the NH moiety of piperazine group binds on Au NPs	—	111
Ag NPs	Ampicillin, penicillin G, isoniazid	d = 20–50 nm, Ag NPs embedded on silica shells functionalized with several antibiotics	<i>E. coli</i> , <i>P. aeruginosa</i> , <i>S. aureus</i> , <i>B. subtilis</i> , <i>C. albicans</i>	104
Ag NPs	Ampicillin, chloramphenicol, erythromycin, gentamicin, tetracycline, vancomycin	d = 3–20 nm, Allophylus cobbe leaf extract acts as a reducing as well as stabilizing/capping agent, mixed with antibiotics	<i>P. aeruginosa</i> , <i>S. flexneri</i> , <i>S. aureus</i> , <i>K. pneumoniae</i>	112
Ag NPs	Amoxicillin	d = 4 and 20 nm, mixed with antibiotics	<i>E. coli</i>	113
Ag NPs	Streptomycin, ampicillin, tetracycline	d = 38.3 (citrate), 19.3 (SDS), 16.0 nm (PVP), mixed with antibiotics	<i>E. coli</i> , <i>S. aureus</i>	114

PVP: polyvinylpyrrolidone.

Mainly two strategies are employed to conjugate antibiotics onto the surface of metallic NPs: (i) adsorption mediated by intermolecular forces such ionic interactions between the amino group of vancomycin and the negative surface charge of Au NPs⁹⁹ or via Au–S bonds^{100,101} and (ii) covalent binding.^{94,102–104} However, the exact mechanism governing the enhancement of the antibacterial properties of antibiotic modified metallic NPs is still highly speculative. In general, the enhanced antibacterial effects of metallic NP/antibiotic complexes have been attributed to the ability of the antibiotic to cause structural change and degradation together with the high affinity of metallic NPs to bind to the bacterial cell wall ensuring a local increase of the antibiotic concentration. Perry and co-workers¹⁰² reported a one-pot synthesis of spherical 52–22 nm Au NPs capped with cefaclor, a second-generation antibiotic where the primary amine groups of cefaclor acted as both the reducing and the capping agent for the synthesis of Au NPs leaving the β -lactam ring of cefaclor available for activity against microbes ([Fig. 7A](#)). The cefaclor reduced Au NPs showed a minimum inhibition concentration (MIC) for *S. aureus* and *E. coli* of 10 and 100 $\mu\text{g mL}^{-1}$, respectively. Coating the particles into a poly(ethylenimine) (PEI) modified glass surface resulted in an antimicrobial coating suitable for biomedical applications. AFM images suggested that the action of these particles takes place through the combination of cefaclor inhibiting the synthesis of peptidoglycan layers and the particles generating holes in the bacterial cell walls, thereby increasing the permeability of the cell wall, resulting in the leakage of cell contents and eventually cell death. In addition, the Au NPs increase the long-term activity of cefaclor, which maintains 100% antibacterial activity after 5 days, while cefaclor has a half-life of ca. 9 h with 11% activity in solution.

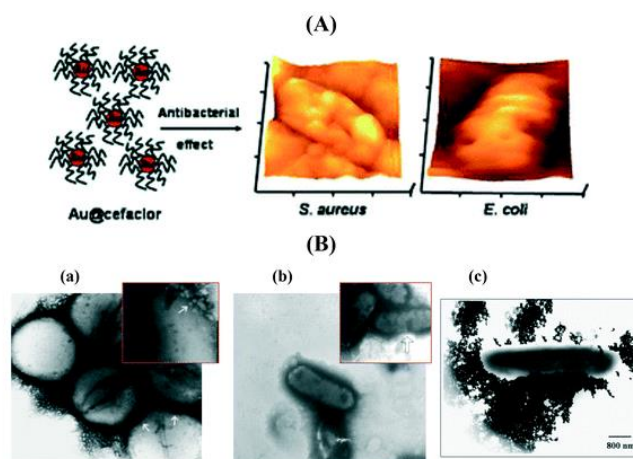


Fig. 7 (A) The antibacterial effect of cefaclor reduced spherical gold nanoparticles as shown by AFM images after incubation for 6 h at 37 °C (reprinted permission from [ref. 102](#)); (B) TEM micrographs of (a) vancomycin-resistant Staphylococcus aureus (VRSA) and (b) E. coli bacteria cells treated with vancomycin-modified Au NPs (reprinted with permission from [ref. 99](#)), (c) streptomycin-modified Au NPs interacting with E. coli (reprinted with permission from [ref. 94](#)).

Ahangari et al.¹⁰⁵ reported that gentamicin (Gm) can be successfully conjugated onto Au NPs via ionic interactions with 347 gentamicin molecules per gold nanosphere. Bio-distribution studies on BALB/c mice infected with Staphylococcus aureus illustrated a good retention of gentamicin conjugated Au NPs in the bacterial infection site. Similar results were reported by Gu et al.¹⁰⁰ and Fayaz et al.⁹⁹ using vancomycin coated Au NPs, exhibiting enhanced activities against vancomycin-resistant enterococci (VRE)¹⁰⁰ and vancomycin-resistant Staphylococcus aureus (VRSA), vancomycin-sensitive S. aureus (VSSA) and E. coli.⁹⁹ The vancomycin modified Au NPs interact with the building elements of the outer membrane causing structural changes, degradation and finally cell wall lysis. Indeed as seen from the TEM images,⁹⁹ the presence of vancomycin modified Au NPs is seen in abundance on the cell wall surface ([Fig. 7B](#)). Similar results were found by Saha et al.⁹⁴ using streptomycin-modified Au NPs ([Fig. 7B](#)). Importantly, the antibiotic-conjugated Au NPs are more stable than the free antibiotic withstanding even harsh storage conditions.

Shahverdi et al.¹⁰⁶ investigated the effect of trivalent gold ions (Au^{3+}) and gold nanoparticles (Au NPs) modified with 14 different antibiotics against P. aeruginosa, S. aureus and E. coli. The Au NPs showed no increase in the antibacterial effect of the antibiotics; the susceptibility of P. aeruginosa increased however in the presence of Au^{3+} and some antibiotics (methicillin, erythromycin, vancomycin, penicillin G, clindamycin, nalidixic acid) up to 147%. Independently, Burygin et al.⁹⁵ demonstrated that Au NPs had no enhancing effect on the antibacterial activity of gentamicin. They suggested that Au NPs enhanced the antibacterial activity of the antibiotic only when the drug was covalently attached onto the nanoparticles' surface, leading to a stable conjugate. In a different work, the same group demonstrated enhanced activity of ciprofloxacin in the presence of ZnO nanoparticles¹⁰⁷ and of different antibiotics in the presence of Ag NPs. The increased activity in the presence of ZnO nanoparticles against S. aureus was associated with the pumping activity of the NorA protein and enhanced adsorption of antibiotics into cells.

3.4. Carbon based materials

The use of carbon based materials for the removal of antibiotics by adsorption with high concentration has been well studied.^{115–119} Surprisingly the literature on the integration of antibiotics onto carbon based structures to ablate bacteria is rather limited ([Table 7](#)). In the case of fullerenes and in particular C60, photodynamic inactivation of bacteria was reported¹²⁰ and the effect proved to be more pronounced on Gram-positive than on Gram-negative bacteria, indicating that the bactericidal action was dependent on the fullerene insertion into the cell wall of the bacteria. Carbon nanotubes modified with amphotericin B, an antibiotic for the treatment of coronial fungal infections, were investigated by Bianco and co-workers for the treatment of *Candida parapsilosis* and *Candida albicans* based infections.¹²¹

Table 7 Carbon-based nanostructures conjugated with antibiotics

Particles	Antibiotics	Comments	Bacteria	Ref.
CDs	Ciprofloxacin	d = 2–5 nm, polyethylene glycol diamine capped CDs conjugated with glutamic acid ciprofloxacin	<i>E. coli</i>	128
CDs	Vancomycin	d = 2.2 nm, formation using citric acid and urea covalently linked with Van	<i>S. aureus</i> , <i>B. subtilis</i> , <i>B. monocutogenes</i>	126
CDs	Ferry ciprofloxacin hydrochloride	d = 2–5 nm, synthesis from gum arabic	<i>E. coli</i> , <i>P. aeruginosa</i> , <i>B. subtilis</i> , <i>S. aureus</i>	129
CDs	Tetracycline alone and associated with β -cyclodextrin	d = 2–5 nm, carbon dot coated alginate beads	—	130
CDs	Ampicillin	d = 40 nm, covalent linking to amine-functionalized C-dots formed from citric acid and ethylenediamine	<i>E. coli</i>	131
prGO	Ampicillin	Electrophoretically deposited prGO/PEI films modified covalently	<i>E. coli</i>	133
Graphene nanosheets	Gentamicin sulfate	Non-covalent loading of methanol derived graphene nanosheets	<i>E. coli</i>	132
Carbon nanotubes	Amphotericin B	Covalent linking via carboxylated sites along sidewalls of CNTs	<i>Candida parapsilosis</i> , <i>Candida albicans</i>	134

Functional carbon dots (CDs) have shown to open promising avenues for bacteria imaging, detection and inactivation.^{122–129} However, very few studies have used CDs as a carrier platform for antibiotics to control pathogenic infections. Krishna et al.¹²⁸ employed polyethylene glycol diamine capped CDs conjugated with glutamic acid and ciprofloxacin for simultaneous crack visualization and drug delivery to control bacterial infection ([Fig. 8A](#)). Thakur et al.¹²⁹ found that CDs can act as efficient nanosinks for delivery of ferry ciprofloxacin hydrochloride. The release of antibiotic was sustained and constant under physiological conditions over 24 h and

the nanocomposites displayed the highest antibacterial activity against *P. aeruginosa*. Gogoi et al.¹³⁰ found that the decoration of calcium alginate beads with carbon dots (CA-CDs) improved the stability of the CA beads, in addition to swelling and the antibiotic's loading capacity. Zhong et al.¹²⁶ synthesized CDs modified with vancomycin for fast and accurate sensing of Gram positive bacteria, including *S. aureus*, *B. subtilis* and *L. monocytogenes*. Our group reported that ampicillin loaded CDs are highly effective in inactivating the growth of *E. coli* (Fig. 8B).¹³¹ The antibacterial efficiency can be further increased upon visible light illumination where moderate quantities of reactive oxygen species were generated, resulting in additional photodynamic inactivation of the pathogens.

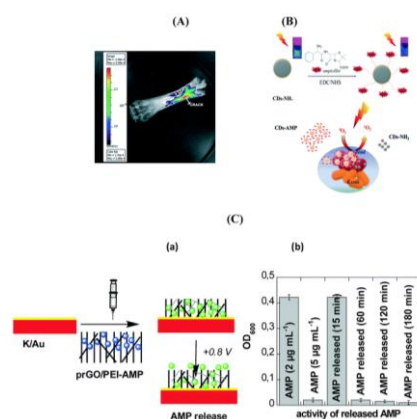


Fig. 8 (A) Bone crack detection by CDGAC imaged with the Xenogen (Caliper Life Sciences) IVIS system (reprinted with permission from [ref. 128](#)); (B) antibacterial activity of ampicillin (AMP) modified CDs functionalized with ampicillin together with the formation of single oxygen under visible light illumination;¹³¹ (C) (a) formation of an electrochemically activatable ampicillin (AMP) loaded interface based on the coating of flexible electrodes with prGO/PEI-AMP, (b) comparison of the action of free AMP ($2 \mu\text{g mL}^{-1}$; negative control), AMP ($5 \mu\text{g mL}^{-1}$; positive control) and electrochemically released AMP at different time intervals when incubated with *E. coli* ($1 \times 10^8 \text{ cfu mL}^{-1}$) for 2 h.¹³³

2D materials such as graphene based nanostructures also attracted huge interest in the loading and release of antibiotics. Pandey et al.¹³² investigated the loading and release behaviour of gentamicin sulphate on methanol derived graphene. An efficient drug release of 62.75% was obtained at pH 3. Moreover, the graphene nano-platform loaded with antibiotic inhibited bacterial growth in an enhanced manner. Porous reduced graphene oxide (prGO) modified with polyethyleneimine (PEI) was shown to be highly interesting for ampicillin loading and for on-demand drug release using electrochemical bias.¹³³ The interaction of ampicillin with prGO was believed to occur through π - π stacking and/or electrostatic interactions, resulting in the preservation of the amino group upon integration into prGO and its antibacterial function against Gram-negative bacteria. Application of +0.8 V resulted in a significant ampicillin release over time (Fig. 8C). The positively charged interface seems to be favorable for zwitterionic molecules such as ampicillin. Following the electrochemical release, the activity of the released ampicillin to kill pathogens was evaluated. The minimum inhibitory concentration of ampicillin against *E. coli* C43 isolates was $\text{MIC}_{50} = 4.9 \pm 0.5 \mu\text{g mL}^{-1}$. Application of +0.8 V for 15 min resulted in a release of $2 \mu\text{g mL}^{-1}$, below the MIC_{50} value, while after 1 h the critical $5 \mu\text{g mL}^{-1}$ was released from the matrix.

3.5. Magnetic nanoparticles

Different pathogen related issues such as the fast and efficient insulation and elimination of bacteria for water disinfection, food decontamination, infectious disease detection and identification, etc. can be addressed by using magnetic nanoparticles (MPs). MP based drug delivery systems retain also the ability to track the movement of the particles through the body¹³⁵ as they can interact with external magnetic fields, offering different opportunities in nanomedicine such as a contrast agent in MRI, magnetic hyperthermia or magnetically triggerable drug delivery systems.¹³⁶ While MPs are not bactericidal, loading of antibiotics on their surface makes them able to interact specifically with pathogens and might result in bacterial ablation (Table 8).

Table 8 Magnetic particles loaded with antibiotics

Particles	Antibiotics	Comments	Bacteria	Ref.
MPs	Vancomycin	d = 8–10 nm, surface thiol exchange with thiol of vancomycin	<i>S. aureus</i> , <i>S. saprophyticus</i>	137
MPs	Vancomycin	d = 50–2800 nm, covalent linkage	Gram-positive and Gram-negative	138
MPs	Methicillin	d = 83 nm, polymers on encapsulated MPs	<i>S. epidermidis</i> biofilm penetration	139

A series of vancomycin-modified MPs were prepared as the COOH groups of vancomycin can be easily activated and further integrated onto the nanostructures.^{137,138,140,141} Vancomycin, a glycopeptide antibiotic, interacts strongly with a broad range of Gram-positive bacteria through a simple five-hydrogen bond motif between the heptapeptide backbone of vancomycin and the D-alanyl-D-alanine-dipeptide extending from the cell wall. The interaction is strong with respect to small molecule–bimolecule interactions with a dissociation constant of 1–4 μM at pH 7.^{142,143} It was shown that the time required to efficiently magnetically confine bacteria decreases when the size of the MP and the vancomycin surface coverage increase.¹³⁸ Recently, a biocompatible multi-compartment nanocarrier containing hydrophobic MPs combined with mPEG-PDLLA (poly(D)-(L)-lactic acid) and the hydrophilic antibiotic methicillin was investigated for the treatment of medical-device associated infections (Fig. 9A).¹³⁹ The MP polymersomes penetrated 20 μm thick *S. epidermidis* biofilms with high efficiency following the application of an external magnetic field with complete biofilm eradication using an optimized MP formulation containing 40 $\mu\text{g mL}^{-1}$ MP and 20 $\mu\text{g mL}^{-1}$ methicillin. The evaluation of the capture capability of vancomycin loaded MPs was recently performed by Zhu et al. using the classical plate counting method.¹⁴¹ Excellent enrichment capabilities were observed once the MP/vancomycin particles were PEGylated (Fig. 9B) with an enrichment >60% even at ultralow bacteria concentrations of 1×10^1 cfu mL^{-1} . Our group demonstrated that magnetic PEGylated graphene oxide nanosheets loaded with 17 $\mu\text{g mL}^{-1}$ of ampicillin allows highly efficient magnetic separation and bacteria inactivation (Fig. 9C).

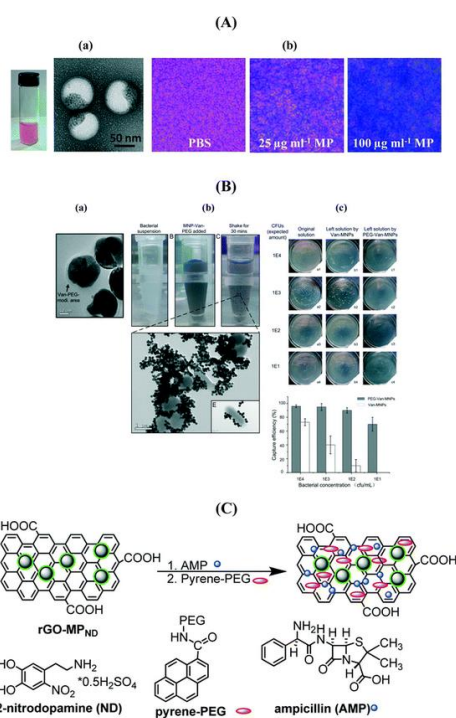


Fig. 9 (A) (a) Photograph of a magnetic/methicillin polymersome solution together with TEM images; (b) Prussian blue staining of biofilms treated with and without iron oxide-encapsulated polymersomes (reprinted with permission from [ref. 139](#)); (B) (a) TEM image of MP–vancomycin–PEG nanocarriers, (b) photographs of bacterial suspensions before and after shaking for 30 min with MP/vancomycin/PEG and TEM image showing the phenomenon of bacterial aggregation, (c) enrichment capability of MP/vancomycin and MP/van/PEG (reprinted with permission from [ref. 141](#)); (C) formulation of ampicillin (AMP) modified PEGylated magnetic reduced graphene oxide.

3.6. Other nanoparticles

The design of gated mesoporous supports able to deliver drugs is a fast-growing area that has attracted much attention of many researchers.¹⁴⁴ The high porosity of mesoporous silica nanoparticles (MSNs) ensuring a high drug-loading capacity and a sustained release of antibiotics at the target sites has been employed to maximize the antibacterial effects of antibiotics ([Table 9](#)). After the pioneering work by Vallet-Regi et al.¹⁴⁵ on ordered mesoporous silica as a matrix for drug delivery, loading with amoxicillin was performed.¹⁴⁶ The release of amoxicillin was observed to be dependent on the different physical states of the material with a faster release rate measured from powder than the disk form. This is in contrast to the gentamicin loaded highly ordered hexagonal mesoporous silica structure (SBA-15) where no significant difference was observed.¹⁴⁷ Nairi et al. postulated that ampicillin release under physiological conditions is strongly affected by the surface charge and not the pore size.¹⁴⁸ Similarly, Savimli and Yilmaz reported that the functionalization of MSNs with various groups, including amine, thiol and methyl can modulate the amount of amoxicillin loaded and released.¹⁴⁹ The introduction of ammonic groups greatly retarded the amoxicillin release from MCM-41.¹⁵⁰ Ciprofloxacin incorporated inside a silica shell (cip@SiO₂) was found to be active against both Gram-positive and Gram-negative bacteria, with a slight decrease in the MIC for *E. coli* DH5 α compared to the free antibiotic.¹⁵¹ The difference between the free drug and the

nanoconjugate was believed to be due to the difference in hydrophobicity. Most interestingly, silica-based porous materials were applied as a scaffold in bone repair (Fig. 10A)¹⁵² or as a coating material to incorporate antibiotics into metallic implantable materials to prevent bacterial infections (Fig. 10B).¹⁵³

Table 9 Mesoporous silica nanoparticles conjugated with antibiotics

Particles	Antibiotics	Ref.
SBA-15	Amoxicillin	146
SBA-15	Gentamicin	147
mSiNTs	Ampicillin	154
MSNs	Ampicillin	148
MSNs	Amoxicillin	149
SBA-15	Erythromycin	155
MSNs	Amoxicillin	150
SiO ₂	Ciprofloxacin	151
MSNs	Gentamicin sulfate	152
Silica xerogel loaded into a porous TiO ₂	Tetracycline-hydrochloride	153
MCM-41 capped with ε-polylysine	Vancomycin	156

SBA-15: highly ordered hexagonal mesoporous silica structure; MSNs: mesoporous silica nanostructures; mSiNTs: mesoporous silica nanotubes.

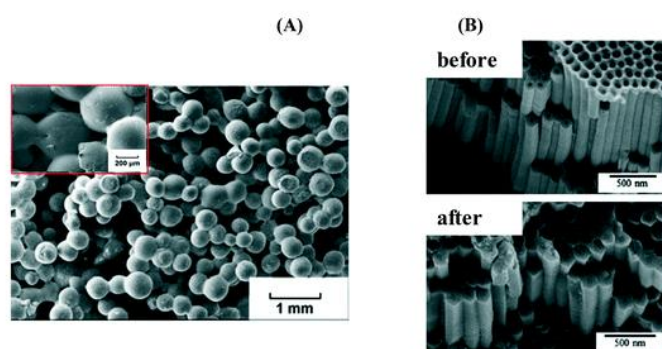


Fig. 10 (A) SEM images of (a) gentamycin sulfate (50 wt%) loaded mesoporous silica–hydroxyapatite particles (HMS–HA) embedded in poly(lactide-co-glycolide) (PLGA) proposed as a biodegradable drug delivery composite for bone replacement (reprinted with permission from [ref. 152](#)); (B) SEM images of porous TiO₂ before (a) and after (b) silica xerogel containing tetracycline-hydrochloride filtration (reprinted with permission from [ref. 153](#)).

4. Conclusion and perspectives

The use of nanoparticles as delivery vehicles for antimicrobial agents represents a new and promising approach in the design of effective therapies against pathogenic and antibiotic resistant bacteria. Such structures possess several clinical advantages due to the intrinsic physico-chemical properties of the nanostructures. Due to the large surface-to-volume ratio, several antibiotics together with targeting molecules can be loaded on the nanostructures. Co-delivery of two or more antibiotics allows the development of a therapeutic modality for combination therapy. This might help in targeting specific pathogens but might be also a long-term strategy to overcome antibiotic resistance. The use of antibiotic loaded nanocarriers overcomes in addition solubility and stability issues of current antibiotics and minimizes drug-induced side effects. The administration of PEG-modified antibiotic-loaded particles increases in addition the drug circulation time and has a strong impact on the overall pharmacokinetics. While, currently, most research is focused on the use of these nanostructures via intravenous administration, nanoparticles' administration can be extended to oral, nasal, pulmonary, parenteral, intra-ocular, or transdermal routes. The numerous benefits of antibiotic-loaded nanostructures over free antimicrobial agents are thus opening a wide range of opportunities. Especially, the possibility of stimulus-controlled delivery is a promising strategy for treating localized infections, assuring a high local concentration of therapeutic molecules and the conjugation of nanoparticles with specific ligands allows a targeted delivery.

The choice of the nanoparticle based platform might depend on the looked for application. While all of the different nanostructures showed improved antibacterial properties when compared to the free antibiotic in solution, differences in synthetic effort, toxicity and mechanical and chemical properties make these nanoantibiotics rather different from each other. [Table 10](#) tries to point out the advantages and disadvantages of each nanosystem.

Table 10 Comparison of the different features of the presented nanoantibiotics

Particles	Advantages	Disadvantages
Liposomes	Improved pharmacokinetics	Short shelf-life which limits drug stability
	Better biodistribution	Drug leakage over time
	Decreased toxicity	Strong influence of the encapsulation process on therapeutic utility
	Target selective	Complex preparation on laboratory scale Expensive
Polymers	Good stability	Presence of a reactive group can influence the rate and yield of the conjugation reaction and stability of the polymer
	Good encapsulation efficiency	
	Controllable release rate/duration	Nondegradable backbones must have a molecular weight below the renal threshold, which is typically around 50 kDa to be safe
	Wide availability of polymers	
	Access to antimicrobial polymers (chitosan)	
Target selective		
Hydrogels	High drug loading	

Particles	Advantages	Disadvantages
	Humidity controlled environment favorable to drug stability	Low elastic force of many hydrogels limits their use in load-bearing applications as these can decompose and release drug
	Responsive structures for drug release	
	Target selective	
Metallic particles	Intrinsic antibacterial activity	Aggregation issues resulting in decreased antibacterial effects
	Easy to synthesise with different shapes, forms	Release of metal ions into the medium which can be potentially toxic
	Improved stability of the drug	Accumulation in the body after administration
	Combination with other antibacterial approaches (PTT, PDT)	
	Target selective	
Magnetic structures	Ability to track their movement through the body	Corrosion related issues
	Useful for imaging applications (MRI)	Magnetic forces might not be strong enough to overcome the force of blood to help in the accumulation to the target site
Carbon-based materials	Easy and cheap fabrication	Diversity in C-based materials making commercialization impossible
	Covalently and non-covalently linked depending on carbon structures	Presence of heavy metal ion impurities from the synthetic procedure
	Tuneable chemical and physical properties allowing imaging	
Mesoporous silica	Sustained and controlled release of drug if protected	Burst release
		Inhibition of release due to blocking of pores
	Non-toxic	Difficulties in controlling the pore size on large scale synthesis

One key obstacle in treating infectious diseases with these nanoantibiotics is in translating this exciting technology into clinical applications. Currently, only one particle formulation for the treatment of fungal diseases is clinically approved and another for the treatment of *Pseudomonas* is undergoing Ph1 clinical trials. Profound knowledge about the potential toxicity of these novel nanoantibiotics is greatly needed to guarantee successful clinical translation. The toxic effects of antimicrobial nanosystems on the central nervous system are still not known and the interaction of particles with cells and tissues remains poorly understood. Clinical translation and integration rely on a consistent and reproducible product. The size-specific properties of these nanosystems demand, in addition, the development and use of synthetic methods with high reproducibility on a large scale. With few exceptions, nanoparticles used in

pre-clinical studies are almost exclusively synthesized in small batches. One of the main challenges facing clinical translation of nanoparticles is controlling their biological fate. Rapid clearance by immune cells can be overcome using PEG, sugar, serum protein modified particles, or concepts which have been successfully used in clinically approved products like Doxil and PEGylated doxorubicin loaded liposomes. Overall, the outlook for nanoparticle based drug delivery is promising and will hopefully in the future lead to promising clinical products for antibiotic-modified carriers. Unfortunately, the results presented to date seem to represent evolutionary improvements over existing options rather than major breakthroughs; further work will clearly be required to reach the goal of more general and powerful treatments and cures.

Acknowledgements

Financial support from the Centre National de la Recherche Scientifique (CNRS), the University Lille 1, the Hauts-de-France region, the CPER “Photonics for Society”, the Agence National de la Recherche (ANR) and the EU through FLAG-ERA JTC 2015-Graphivity, and the Marie Skłodowska-Curie Action (H2020-MSCA-RISE-2015, PANG-690836) is acknowledged.

References

1. A. C. Anselmo and S. Mitragotri, *Bioeng. Transl. Med.*, 2016, 1, 10–29 [Search](#) .
2. H.-J. Choi, D. Wendell and C. D. Montemagno, *NanoBiotechnology*, 2007, 3, 66–75.
3. R. P. Rennie, *Handb. Exp. Pharmacol.*, 2012, 211, 45–65
4. K. Forier, K. Raemdonck, S. C. De Smedt, J. Demeester, T. Coenye and K. Braeckmans, *J. Controlled Release*, 2014, 190, 607–623
5. H. Pinto-Alphandary, A. Andreumont and P. Couvreur, *Int. J. Antimicrob. Agents*, 2000, 13, 155–168
6. M. Veerapandian and K. Yun, *Appl. Microbiol. Biotechnol.*, 2011, 90, 1655–1667
7. Z. Drulis-Kawa and A. Dorotkiewicz-Jach, *Int. J. Pharm.*, 2010, 387, 187–198
8. L. Zhang, D. Pornpattananangkul, C.-M. J. Hu and C.-M. Huang, *Curr. Med. Chem.*, 2010, 17, 585–594
9. A. Muñoz-Bonilla and M. Fernández-García, *Eur. Polym. J.*, 2015, 65, 46–62
10. Y. Zhao and X. Jiang, *Nanoscale*, 2013, 5, 8340–8350
11. M. Morar and G. D. Wright, *Annu. Rev. Genet.*, 2010, 44, 25–51
12. S. B. Levy and B. Marshall, *Nat. Med.*, 2004, 10, S122–S129
13. J. Davies and D. Davies, *Microbiol. Mol. Biol. Rev.*, 2010, 74, 417–433
14. M. B. Habash, A. J. Park, E. C. Vis, R. J. Harris and C. M. Khursigara, *Antimicrob. Agents Chemother.*, 2014, 58, 5818–5830
15. A. Nirmala Grace and K. Pandian, *Colloids Surf., A*, 2007, 297, 63–70
16. Y. C. Lee and R. T. Lee, *Acc. Chem. Res.*, 1995, 28, 321–327
17. M. Mammen, S.-K. Choi and G. M. Whitesides, *Angew. Chem., Int. Ed.*, 1998, 37, 2754–2794
18. C.-C. Lin, Y.-C. Yeh, C.-Y. Yang, C.-L. Chen, G.-F. Chen, C.-C. Chen and Y.-C. Wu, *J. Am. Chem. Soc.*, 2002, 124, 3508–3509
19. J. J. Lundquist and E. J. Toone, *Chem. Rev.*, 2002, 102, 555–578
20. L. L. Kiessling, J. E. Gestwicki and L. E. Strong, *Curr. Opin. Chem. Biol.*, 2000, 4, 696–703

21. M. Durka, K. Buffet, J. Iehl, M. Holler, J.-F. Nierengarten, J. Taganna, J. Bouckaert and S. P. Vincent, *Chem. Commun.*, 2011, 47, 1321–1323
22. A. Bangham, M. M. Standish and G. Weissmann, *J. Mol. Biol.*, 1965, 13, 253–259
23. A. Akbarzadeh, R. Rezaei-Sadabady, S. Davaran, S. W. Joo, N. Zarghami, Y. Hanifehpour, M. Samiei, M. Kouhi and K. Nejati-Koshki, *Nanoscale Res. Lett.*, 2013, 8, 102
24. S. Labana, R. Pandey, S. Sharma and G. Khuller, *Int. J. Antimicrob. Agents*, 2002, 20, 301–304
25. Z. Drulis-Kawa, A. Dorotkiewicz-Jach, J. Gubernator, G. Gula, T. Bocser and W. Doroszkiewicz, *Int. J. Pharm.*, 2009, 367, 211–219
26. R. E. Hancock and F. S. Brinkman, *Annu. Rev. Microbiol.*, 2002, 56, 17–38
27. G. Gregoriadis, *FEBS Lett.*, 1973, 36, 292–296
28. P. F. Bonventre and G. Gregoriadis, *Antimicrob. Agents Chemother.*, 1978, 13, 1049
29. J. V. Desiderio and S. G. Campbell, *J. Infect. Dis.*, 1983, 148, 563–570
30. M. Stevenson, A. Baillie and R. Richards, *Antimicrob. Agents Chemother.*, 1983, 24, 742–749
31. M. Nacucchio, M. Bellora, D. Sordelli and M. D'aquino, *Antimicrob. Agents Chemother.*, 1985, 27, 137–139
32. S. G. Ingebrigtsen, A. Didriksen, M. Johannessen, N. Škalko-Basnet and A. M. Holsæter, *Int. J. Pharm.*, 2017, 526, 538–546
33. Y. Li, T. Su, Y. Zhang, X. Huang, J. Li and C. Li, *Drug Delivery*, 2015, 22, 627–637
34. S. Gaidukevich, Y. L. Mikulovich, T. Smirnova, S. Andreevskaya, G. Sorokoumova, L. Chernousova, A. Selishcheva and V. Shvets, *Bull. Exp. Biol. Med.*, 2016, 160, 675
35. P. Matouskova, I. Marova, J. Bokrova and P. Benesova, *Food Technol. Biotechnol.*, 2016, 54, 304
36. X. Fan, J. Fan, X. Wang, P. Wu and G. Wu, *Frontiers in pharmacology*, 2015, 6, 249
37. C. Liu, J. Shi, Q. Dai, X. Yin, X. Zhang and A. Zheng, *Drug Dev. Ind. Pharm.*, 2015, 41, 272–278
38. S. Antonela Antoniu, *Expert Rev. Anti-Infect. Ther.*, 2012, 10, 1439–1446
39. I. Colzi, A. N. Troyan, B. Perito, E. Casalone, R. Romoli, G. Pieraccini, N. Škalko-Basnet, A. Adessi, F. Rossi and C. Gonnelli, *Eur. J. Pharm. Biopharm.*, 2015, 94, 411–418
40. Z. Yang, J. Liu, J. Gao, S. Chen and G. Huang, *Int. J. Pharm.*, 2015, 495, 508–515
41. K. A. Hamblin, S. J. Armstrong, K. B. Barnes, C. Davies, J. P. Wong, J. D. Blanchard, S. V. Harding, A. J. Simpson and H. S. Atkins, *Antimicrob. Agents Chemother.*, 2014, 58, 3053–3059
42. D. G. Atashbeyk, B. Khameneh, M. Tafaghodi and B. S. Fazly Bazzaz, *Pharm. Biol.*, 2014, 52, 1423–1428
43. S. Thamphiwatana, W. Gao, D. Pornpattananangkul, Q. Zhang, V. Fu, J. Li, J. Li, M. Obonyo and L. Zhang, *J. Mater. Chem. B*, 2014, 2, 8201–8207
44. S. Thamphiwatana, V. Fu, J. Zhu, D. Lu, W. Gao and L. Zhang, *Langmuir*, 2013, 29, 12228–12233
45. A.-S. Messiaen, K. Forier, H. Nelis, K. Braeckmans and T. Coenye, *PLoS One*, 2013, 8, e79220
46. M. Alhajlan, M. Alhariri and A. Omri, *Antimicrob. Agents Chemother.*, 2013, 57, 2694–2704

47. J. He, K. Abdelraouf, K. R. Ledesma, D. S.-L. Chow and V. H. Tam, *Int. J. Antimicrob. Agents*, 2013, 42, 559–564
48. M. Alipour, M. Halwani, A. Omri and Z. E. Suntres, *Int. J. Pharm.*, 2008, 355, 293–298
49. S. J. Wallace, J. Li, R. L. Nation, R. J. Prankerd and B. J. Boyd, *J. Pharm. Sci.*, 2012, 101, 3347–3359
50. L. Sande, M. Sanchez, J. Montes, A. J. Wolf, M. A. Morgan, A. Omri and G. Y. Liu, *J. Antimicrob. Chemother.*, 2012, 67, 2191–2194
51. A. Pumerantz, K. Muppidi, S. Agnihotri, C. Guerra, V. Venketaraman, J. Wang and G. Betageri, *Int. J. Antimicrob. Agents*, 2011, 37, 140–144
52. J. E. de Steenwinkel, W. van Vianen, T. Marian, H. A. Verbrugh, M. A. van Agtmael, R. M. Schiffelers and I. A. Bakker-Woudenberg, *J. Antimicrob. Chemother.*, 2007, 60, 1064–1073
53. P.-L. Bardonnet, V. Faivre, P. Boullanger, J.-C. Piffaretti and F. Falson, *Eur. J. Pharm. Biopharm.*, 2008, 69, 908–922
54. N. Monteiro, M. Martins, A. Martins, N. A. Fonseca, J. N. Moreira, R. L. Reis and N. M. Neves, *Acta Biomater.*, 2015, 18, 196–205
55. D. Pornpattananangkul, L. Zhang, S. Olson, S. Aryal, M. Obonyo, K. Vecchio, C.-M. Huang and L. Zhang, *J. Am. Chem. Soc.*, 2011, 133, 4132–4139
56. A. Sharma and U. S. Sharma, *Int. J. Pharm.*, 1997, 154, 123–140
57. K. M. El-Say and H. S. El-Sawy, *Int. J. Pharm.*, 2017, 528, 675–691
58. B. Bharatwaj, L. Wu, J. A. Whittum-Hudson and S. R. da Rocha, *Biomaterials*, 2010, 31, 7376–7385
59. W. S. Cheow and K. Hadinoto, *Microb. Biofilms*, 2014, 227–238
60. W. Gao, R. H. Fang, S. Thamphiwatana, B. T. Luk, J. Li, P. Angsantikul, Q. Zhang, C.-M. J. Hu and L. Zhang, *Nano Lett.*, 2015, 15, 1403–1409
61. S. R. Shah, A. M. Henslee, P. P. Spicer, S. Yokota, S. Petrichenko, S. Allahabadi, G. N. Bennett, M. E. Wong, F. K. Kasper and A. G. Mikos, *Pharm. Res.*, 2014, 31, 3379–3389
62. P. Sabaeifard, A. Abdi-Ali, M. R. Soudi, C. Gamazo and J. M. Irache, *Eur. J. Pharm. Sci.*, 2016, 93, 392–398
63. A. Baelo, R. Levato, E. Julián, A. Crespo, J. Astola, J. Gavaldà, E. Engel, M. A. Mateos-Timoneda and E. Torrents, *J. Controlled Release*, 2015, 209, 150–158
64. I. d'Angelo, B. Casciaro, A. Miro, F. Quaglia, M. L. Mangoni and F. Ungaro, *Colloids Surf., B*, 2015, 135, 717–725
65. W.-L. Chiang, T.-T. Lin, R. Sureshbabu, W.-T. Chia, H.-C. Hsiao, H.-Y. Liu, C.-M. Yang and H.-W. Sung, *J. Controlled Release*, 2015, 199, 53–62
66. D. Das and S. Pal, *Adv.*, 2015, 5, 25014–25050
67. T.-K. Nguyen, R. Selvanayagam, K. K. Ho, R. Chen, S. K. Kutty, S. A. Rice, N. Kumar, N. Barraud, H. T. Duong and C. Boyer, *Chem. Sci.*, 2016, 7, 1016–1027
68. Y. Wu, J. Liang, F. Horkay and M. Libera, *J. Polym. Sci., Part B: Polym. Phys.*, 2016, 54, 64–72
69. L. Chu, H. Gao, T. Cheng, Y. Zhang, J. Liu, F. Huang, C. Yang, L. Shi and J. Liu, *Chem. Commun.*, 2016, 52, 6265–6268
70. B. Horev, M. I. Klein, G. Hwang, Y. Li, D. Kim, H. Koo and D. S. Benoit, *ACS Nano*, 2015, 9, 2390–2404
71. J. Deacon, S. M. Abdelghany, D. J. Quinn, D. Schmid, J. Megaw, R. F. Donnelly, D. S. Jones, A. Kissenpfennig, J. S. Elborn and B. F. Gilmore, *J. Controlled Release*, 2015, 198, 55–61

72. F. Wu, G. Meng, J. He, Y. Wu, F. Wu and Z. Gu, *ACS Appl. Mater. Interfaces*, 2014, 6, 10005–10013
73. L.-L. Li, J.-H. Xu, G.-B. Qi, X. Zhao, F. Yu and H. Wang, *ACS Nano*, 2014, 8, 4975–4983
74. M.-H. Xiong, Y. Bao, X.-Z. Yang, Y.-C. Wang, B. Sun and J. Wang, *J. Am. Chem. Soc.*, 2012, 134, 4355–4362
75. M. H. Xiong, Y. J. Li, Y. Bao, X. Z. Yang, B. Hu and J. Wang, *Adv. Mater.*, 2012, 24, 6175–6180
76. R. S. Navath, A. R. Menjoge, H. Dai, R. Romero, S. Kannan and R. M. Kannan, *Mol. Pharmaceutics*, 2011, 8, 1209–1223
77. A. F. Radovic-Moreno, T. K. Lu, V. A. Puscasu, C. J. Yoon, R. Langer and O. C. Farokhzad, *ACS Nano*, 2012, 6, 279–4287
78. Y. Li, G. Liu, X. Wang, J. Hu and S. Liu, *Angew. Chem., Int. Ed.*, 2016, 55, 1760–1764
79. M. Changez, V. Koul and A. K. Dinda, *Biomaterials*, 2005, 26, 2095–2104
80. A. G. Ashbaugh, X. Jiang, J. Zheng, A. S. Tsai, W.-S. Kim, J. M. Thompson, R. J. Miller, J. H. Shahbazian, Y. Wang and C. A. Dillen, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, 201613722
81. P. T. Wong, S. Tang, J. Mukherjee, K. Tang, K. Gam, D. Isham, C. Murat, R. Sun, J. R. Baker and S. K. Choi, *Chem. Commun.*, 2016, 52, 10357–10360
82. S. K. Choi, A. Myc, J. E. Silpe, M. Sumit, P. T. Wong, K. McCarthy, A. M. Desai, T. P. Thomas, A. Kotlyar, M. M. B. Holl, B. G. Orr and J. R. Baker, *ACS Nano*, 2013, 7, 214–228
83. S. J. Sonawane, R. S. Kalhapure, S. Rambharose, C. Mocktar, S. B. Vepuri, M. Soliman and T. Govender, *Int. J. Pharm.*, 2016, 504, 1–10
84. G. Michaud, R. Visini, M. Bergmann, G. Salerno, R. Bosco, E. Gillon, B. Richichi, C. Nativi, A. Imberty and A. Stocker, *Chem. Sci.*, 2016, 7, 166–182
85. N. Wrońska, A. Felczak, K. Zawadzka, A. Janaszewska, B. Klajnert, M. Bryszewska and K. Lisowska, *New J. Chem.*, 2014, 38, 2987–2992
86. K. Winnicka, M. Wroblewska, P. Wiczorek, P. T. Sacha and E. A. Trynieszewska, *Molecules*, 2013, 18, 8607–8617
87. Y.-J. Zhou, M.-X. Zhang, R. C. Hider and T. Zhou, *FEMS Microbiol. Lett.*, 2014, 355, 124–130
88. A. Felczak, K. Zawadzka, N. Wrońska, A. Janaszewska, B. Klajnert, M. Bryszewska, D. Appelhans, B. Voit and K. Lisowska, *New J. Chem.*, 2013, 37, 4156–4162
89. P. V. Kumar, A. Asthana, T. Dutta and N. K. Jain, *J. Drug Targeting*, 2006, 14, 546–556
90. M. K. Mishra, K. Kotta, M. Hali, S. Wykes, H. C. Gerard, A. P. Hudson, J. A. Whittum-Hudson and R. M. Kannan, *Nanomedicine*, 2011, 7, 935–944
91. H. W. VanKoten, W. M. Dlakic, R. Engel and M. J. Cloninger, *Mol. Pharmaceutics*, 2016, 13, 3827–3834
92. S. Szunerits and R. Boukherroub, *Chem. Commun.*, 2012, 48, 8999–9010
93. N. Khlebtsov, V. Bogatyrev, L. Dykman, B. Khlebtsov, S. Staroverov, A. Shirokov, L. Matora, V. Khanadeev, T. Pylaev and N. Tsyganova, *Theranostics*, 2013, 3, 167–180
94. B. Saha, J. Bhattacharya, A. Mukherjee, A. Ghosh, C. Santra, A. K. Dasgupta and P. Karmakar, *Nanoscale Res. Lett.*, 2007, 2, 614
95. G. Burygin, B. Khlebtsov, A. Shantrokha, L. Dykman, V. Bogatyrev and N. Khlebtsov, *Nanoscale Res. Lett.*, 2009, 4, 794

96. Y.-H. Hsueh, K.-S. Lin, W.-J. Ke, C.-T. Hsieh, C.-L. Chiang, D.-Y. Tzou and S.-T. Liu, *PLoS One*, 2015, 10, e0144306
97. M. A. Raza, Z. Kanwal, A. Rauf, A. N. Sabri, S. Riaz and S. Naseem, *Nanomaterials*, 2016, 6, 74
98. M. Rai, A. Yadav and A. Gade, *Biotechnol. Adv.*, 2009, 27, 76–83
99. A. M. Fayaz, M. Girilal, S. A. Mahdy, S. Somsundar, R. Venkatesan and P. Kalaichelvan, *Process Biochem.*, 2011, 46, 636–641
100. H. Gu, P. Ho, E. Tong, L. Wang and B. Xu, *Nano Lett.*, 2003, 3, 1261–1263
101. A. N. Brown, K. Smith, T. A. Samuels, J. Lu, S. O. Obare and M. E. Scott, *Appl. Environ. Microbiol.*, 2012, 78, 2768–2774
102. A. Rai, A. Prabhune and C. C. Perry, *J. Mater. Chem.*, 2010, 20, 6789–6798
103. Y. E. Hur, S. Kim, J.-H. Kim, S.-H. Cha, M.-J. Choi, S. Cho and Y. Park, *Mater. Lett.*, 2014, 129, 185–190
104. M. Tudose, D. C. Culita, C. Munteanu, J. Pandeale, E. Hristea, P. Ionita, I. Zarafu and M. C. Chifiriuc, *J. Inorg. Organomet. Polym. Mater.*, 2015, 25, 869–878
105. A. Ahangari, M. Salouti, Z. Heidari, A. R. Kazemizadeh and A. A. Safari, *Drug Delivery*, 2013, 20, 34–39
106. A. R. Shahverdi, A. Fakhimi, H. R. Shahverdi and S. Minaian, *Nanomedicine*, 2007, 3, 168–171
107. M. Banoee, S. Seif, Z. E. Nazari, P. Jafari-Fesharaki, H. R. Shahverdi, A. Moballeg, K. M. Moghaddam and A. R. Shahverdi, *J. Biomed. Mater. Res., Part B*, 2010, 93, 557–561
108. Z. E. Nazari, M. Banoee, A. A. Sepahi, F. Rafii and A. R. Shahverdi, *Gold Bull.*, 2012, 45, 53–59
109. A. N. Grace and K. Pandian, *Colloids Surf., A*, 2007, 297, 63–70
110. D. Bhattacharya, B. Saha, A. Mukherjee, C. R. Santra and P. Karmakar, *Nanosci. Nanotechnol.*, 2012, 2, 14–21
111. R. T. Tom, V. Suryanarayanan, P. G. Reddy, S. Baskaran and T. Pradeep, *Langmuir*, 2004, 20, 1909–1914
112. S. Gurunathan, J. W. Han, D.-N. Kwon and J.-H. Kim, *Nanoscale Res. Lett.*, 2014, 9, 373
113. P. Li, J. Li, C. Wu, Q. Wu and J. Li, *Nanotechnology*, 2005, 16, 1912
114. A. J. Kora and L. Rastogi, *Bioinorg. Chem. Appl.*, 2013, 2013, 871097
115. O. Akhavan, E. Ghaderi and A. Esfandiari, *J. Phys. Chem. B*, 2011, 115, 6279–6288
116. M. Bottini, N. Rosato and N. Bottini, *Biomacromolecules*, 2011, 12, 3381–3393
117. B. Peng, L. Chen, C. Que, K. Yang, F. Deng, X. Deng, G. Shi, G. Xu and M. Wu, *Sci. Rep.*, 2016, 6, 31920
118. X. Zhang, J. Shen, N. Zhuo, Z. Tian, P. Xu, Z. Yang and W. Yang, *ACS Appl. Mater. Interfaces*, 2016, 8, 24273–24280
119. Y. Gao, J. Wu, X. Ren, X. Tan, T. Hayat, A. Alsaedi, C. Cheng and C. Chen, *Environ. Sci.: Nano*, 2017, 4, 1016–1024
120. P. Mroz, G. P. Tegos, H. Gali, T. Wharton, T. Sarna and M. R. Hamblin, *Photochem. Photobiol. Sci.*, 2007, 6, 1139–1149
121. W. Wu, S. Wieckowski, G. Pastorin, M. Benincasa, C. Klumpp, J. P. Briand, R. Gennaro, M. Prato and A. Bianco, *Angew. Chem., Int. Ed.*, 2005, 44, 6358–6362

122. Y.-P. Sun, B. Zhou, Y. Lin, W. Wang, K. S. Fernando, P. Pathak, M. J. Mezziani, B. A. Harruff, X. Wang and H. Wang, *J. Am. Chem. Soc.*, 2006, 128, 7756–7757
123. L. Cao, S.-T. Yang, X. Wang, P. G. Luo, J.-H. Liu, S. Sahu, Y. Liu and Y.-P. Sun, *Theranostics*, 2012, 2, 295–301
124. Q. Wang, X. Huang, Y. Long, X. Wang, H. Zhang, R. Zhu, L. Liang, P. Teng and H. Zheng, *Carbon*, 2013, 59, 192–199
125. C.-I. Weng, H.-T. Chang, C.-H. Lin, Y.-W. Shen, B. Unnikrishnan, Y.-J. Li and C.-C. Huang, *Biosens. Bioelectron.*, 2015, 68, 1–6
126. D. Zhong, Y. Zhuo, Y. Feng and X. Yang, *Biosens. Bioelectron.*, 2015, 74, 546–553
127. M. J. Mezziani, X. Dong, L. Zhu, L. P. Jones, G. E. LeCroy, F. Yang, S. Wang, P. Wang, Y. Zhao and L. Yang, *ACS Appl. Mater. Interfaces*, 2016, 8, 10761–10766
128. A. S. Krishna, C. Radhakumary, M. Antony and K. Sreenivasan, *J. Mater. Chem. B*, 2014, 2, 8626–8632
129. M. Thakur, S. Pandey, A. Mewada, V. Patil, M. Khade, E. Goshi and M. Sharon, *J. Drug Delivery*, 2014, 2014, 282193
130. N. Gogoi and D. Chowdhury, *J. Mater. Chem. B*, 2014, 2, 4089–4099
131. R. Jijie, A. Barras, J. Bouckaert, N. Dumitrascu, S. Szunerits and R. Boukherroub, *J. Mater. Chem.*, 2017, submitted.
132. H. Pandey, V. Parashar, R. Parashar, R. Prakash, P. W. Ramteke and A. C. Pandey, *Nanoscale*, 2011, 3, 4104–4108
133. S. Boulahneche, R. Jijie, A. Barras, F. Chekin, S. K. Singh, J. Bouckaert, M. S. Medjram, S. Kurungot, R. Boukherroub and S. Szunerits, *J. Mater. Chem. B*, 2017 [10.1039/C7TB00687J](https://doi.org/10.1039/C7TB00687J)
134. W. Wu, S. Wieckowski, G. Pastorin, M. Benincasa, C. Klumpp, J.-P. Briand, R. Gennaro, M. Prato and A. Bianoc, *Angew. Chem., Int. Ed.*, 2005, 44, 6358–6362
135. O. Veisoh, J. W. Gunn and M. Zhang, *Adv. Drug Delivery Rev.*, 2010, 62, 284–304
136. H. Mok and M. Zhang, *Expert Opin. Drug Delivery*, 2013, 10, 73–87
137. H. Gu, K. Xu, C. Xu and B. Xu, *Chem. Commun.*, 2006, 941–949.
138. A. J. Kell, G. Stewart, S. Ryan, R. Peytavi, M. Boissinot, A. Huletsky, M. G. Bergeron and B. Simard, *ACS Nano*, 2008, 2, 1777–1788.
139. B. M. Geilich, I. Gelfat, S. Sridhar, A. L. van de Ven and T. J. Webster, *Biomaterials*, 2017, 119, 78–85.
140. . Gu, P.-L. Ho, K. W. Tsang, L. Wang and B. Xu, *J. Am. Chem. Soc.*, 2003, 125, 15702–15703.
141. M. Zhu, W. Liu, H. Liu, Y. Liao, J. Wei, X. Zhou and D. Xing, *ACS Appl. Mater. Interfaces*, 2015, 7, 12873–12881.
142. J. Rao, I. J. Colton and G. M. Whitesides, *J. Am. Chem. Soc.*, 1997, 119, 9336–9340.
143. B. K. Hubbard and C. T. Walsh, *Angew. Chem., Int. Ed.*, 2003, 42, 730–765.
144. A. P. A. P. Wight and M. E. Davis, *Chem. Rev.*, 2002, 102, 3589–3614.
145. M. Vallet-Regi, A. Ramila, R. Del Real and J. Pérez-Pariente, *Chem. Mater.*, 2001, 13, 308–311.
146. M. Vallet-Regi, J. Doadrio, A. Doadrio, I. Izquierdo-Barba and J. Pérez-Pariente, *Solid State Ionics*, 2004, 172, 435–439.

147. A. Doadrio, E. Sousa, J. Doadrio, J. P. Pariente, I. Izquierdo-Barba and M. Vallet-Regí, *J. Controlled Release*, 2004, 97, 125–132.
148. V. Nairi, L. Medda, M. Monduzzi and A. Salis, *J. Colloid Interface Sci.*, 2017, 497, 217–225.
149. Sevimli and A. Yilmaz, *Microporous Mesoporous Mater.*, 2012, 158, 281–291.
150. Z. Li, K. Su, B. Cheng and Y. Deng, *J. Colloid Interface Sci.*, 2010, 342, 607–613.
151. M. Rosemary, I. MacLaren and T. Pradeep, *Langmuir*, 2006, 22, 10125–10129.
152. X. Shi, Y. Wang, L. Ren, N. Zhao, Y. Gong and D.-A. Wang, *Acta Biomaterial.*, 2009, 5, 1697–1707.
153. C.-M. Han, E.-J. Lee, H.-E. Kim, Y.-H. Koh and J.-H. Jang, *Thin Solid Films*, 2011, 519, 8074–8076.
154. R. K. Singh, T.-H. Kim, J.-J. Kim, E.-J. Lee, J. C. Knowles and H.-W. Kim, *Adv.*, 2013, 3, 8692–8704.
155. J. C. Doadrio, E. M. Sousa, I. Izquierdo-Barba, A. L. Doadrio, J. Perez-Pariente and M. Vallet-Regí, *J. Mater. Chem.*, 2006, 16, 462–466.
156. N. Mas, I. Galiana, L. Mondragon, E. Aznar, E. Climent, N. Cabedo, F. Sancenon, J. Ramano Murguia, R. Martinez-Manez, M. D. Marcos and P. Amoros, *Chem. – Eur. J.*, 2013, 19, 11167–11171.