

Natural polymeric 3-alkylpyridinium salt affects vertebrate skeletal muscle contractility by preferentially blocking neuromuscular transmission

Monika Cecilija Žužek, Marjana Grandič, Evelyne Benoit, Robert Frangež

▶ To cite this version:

Monika Cecilija Žužek, Marjana Grandič, Evelyne Benoit, Robert Frangež. Natural polymeric 3-alkylpyridinium salt affects vertebrate skeletal muscle contractility by preferentially blocking neuromuscular transmission. Toxicology Letters, 2017, 281, pp.95-101. 10.1016/j.toxlet.2017.09.007. hal-01661797

HAL Id: hal-01661797

https://hal.science/hal-01661797

Submitted on 21 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Natural polymeric 3-alkylpyridinium salt affects vertebrate skeletal muscle

2 contractility by preferentially blocking neuromuscular transmission

3

- 4 Monika Cecilija Žužek^a, Marjana Grandič^b, Evelyne Benoit^c and Robert Frangež^a
- ^aInstitute of Preclinical Sciences, Veterinary Faculty, University of Ljubljana, Gerbičeva 60,
- 6 1000 Ljubljana, Slovenia
- 7 bInstitute of food safety, feed and environment, Department of environment, animal nutrition,
- 8 welfare and hygiene, Veterinary Faculty, University of Ljubljana, Cesta v Mestni log 47,
- 9 1000 Ljubljana, Slovenia
- 10 °DRF/Institut des Sciences de la Vie Frédéric Joliot/SIMOPRO, CEA de Saclay, and Institut
- des Neurosciences Paris-Saclay (Neuro-PSI), UMR 9197 CNRS/Université Paris-Sud, 91190
- 12 Gif-sur-Yvette, France
- *corresponding author: Robert Frangež; E-mail address: robert.frangez@vf.uni-lj.si; tel.:
- 14 00386 1 477 91 31

15

16

Highlights

- nPoly-3-APS preferentially blocks the skeletal neuromuscular transmission
- nPoly-3-APS acts by a non-depolarizing mechanism
- In vivo toxicity of nPoly-3-APS mainly occurs through antagonist action on nAChRs

20

21

Abstract

- 22 The effects of natural polymeric alkylpyridinium salt (nPoly-3-APS), a potent
- 23 acetylcholinesterase inhibitor isolated from the marine sponge *Reniera sarai*, were studied on
- 24 isolated mouse phrenic nerve-hemidiaphragm muscle preparations using electrophysiological
- 25 approaches. nPoly-3-APS inhibited nerve-evoked isometric muscle twitch and tetanic
- 26 contraction in a concentration-dependent manner (IC₅₀ = 29.4 μ M and 18.5 μ M, respectively)
- and produced a 30-44% decrease of directly muscle-elicited twitch and tetanus amplitudes at
- 28 54.4 μM . Additionally, nPoly-3-APS (9.1-27.2 μM) markedly decreased the amplitude of
- 29 miniature endplate potentials, while their frequency was only affected at the highest
- 30 concentration used. Endplate potentials were also inhibited by nPoly-3-APS in a
- 31 concentration-dependent manner (IC $_{50} = 20.1 \mu M$), without significant change in the resting
- membrane potential of muscle fibers (up to $54.4 \mu M$). In conclusion, our results show, for the
- first time, that nPoly-3-APS preferentially blocks the neuromuscular transmission, *in vitro*, by
- a non-depolarizing mechanism. This strongly suggests that the *in vivo* toxicity of nPoly-3-
- 35 APS mainly occurs through an antagonist action of the compound on nicotinic acetylcholine
- 36 receptors of skeletal muscles.

- 38 Key words: Natural Poly-APS; Neuromuscular junction; AChE inhibitors; Motor endplate
- 39 potential; Skeletal muscle contraction

1 Introduction

40

41

42 43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67 68

69

70

71

72 73

74

75

Natural polymeric 3-alkylpyridinium salt (nPoly-3-APS), isolated from the marine sponge Reniera sarai (Sepcic et al., 1997), shows a broad spectrum of biological activities, including antibacterial and antifouling activities, acetylcholinesterase (AChE) inhibition, cytotoxicity, as well as hemolysis and selective toxicity against non-small cell lung cancer (NSCLC) tumor cells in vitro and in vivo (Sepcic et al., 1997; Sepcic et al., 1998; Malovrh et al., 1999; Faimali et al., 2003; Tucker et al., 2003; Chelossi et al., 2006; Paleari et al., 2006; Catassi et al., 2008; Elersek et al., 2008). nPoly-3-APS can also be used for the stable transfection of nucleated mammalian cells (Tucker et al., 2003). Its likely application in medicine led to the synthesis of a series of analogs with different degree of polymerization and length of alkyl chains (Houssen et al., 2010). Synthetic analogs of Poly-3-APS, such as APS12-2, APS3 (Grandic et al., 2011; Grandic et al., 2013), APS8 (unpublished data) and structurally related compounds, are promising new chemotherapeutic agents that exhibit very low in vivo toxicity in experimental animals. Recently, the effects of some of these synthetic analogs possessing potent AChE-inhibitory properties were studied on neuromuscular transmission in skeletal muscle (reviewed in Grandic and Frangez, 2014) to avoid unwanted peripheral side effects that could appear in patients treated with some AChE inhibitors. APS12-2 acts as a potent non-competitive AChE inhibitor with an inhibitory constant (K_i) of 34×10^{-3} nM (Houssen et al., 2010). It exerts hemolytic, antibacterial and antifungal actions (Houssen et al., 2010). Some nicotinic cholinergic antagonists, such as APS3 and APS8, have shown promise in lung cancer treatment because of their selective cytotoxicity against NSCLC, the most common form of lung cancer (Zovko et al., 2013). NSCLC cells express cholinergic signaling mediators, including neuronal-type α7-nicotinic acetylcholine receptors (Paleari et al., 2006; Catassi et al., 2008). The inhibitory effects of the synthetic analogs APS12-2 and APS-3 on neuromuscular transmission and nerve-evoked skeletal muscle contraction were previously reported (Grandic et al., 2012; Grandic et al., 2013). It was demonstrated that the neuromuscular blockade produced by these two compounds is due to their direct inhibitory effect on muscle-type nicotinic acetylcholine receptors (nAChRs). Therefore, it was of interest to determine whether the natural compound nPoly-3-APS shares the same biological effects on neuromuscular transmission and skeletal muscle functioning. In particular, due to the structural similarity between nPoly-3-APS and quaternary ammonium compounds that have been reported to block muscle-type nAChRs, the aim of this study was to investigate the effects and underlying mechanisms of the natural compound on neuromuscular transmission and skeletal muscle contraction. We show, for the first time, that nPoly-3-APS affects the skeletal muscle functioning, in vitro, by preferentially blocking the neuromuscular transmission probably through an antagonist action on muscle-type nAChRs.

76

77

2 Materials and methods

78 2.1 Materials

79 2.1.1 Drugs

80 nPoly-3-APS was isolated from the marine sponge Reniera sarai according to Sepčić et al.

81 (1997). Before use, the toxin was dissolved in a sterile 0.9% saline solution at a stock

- 82 concentration of 10 mg/mL and was stored at 4°C. Neostigmine methyl sulfate (Tokyo
- Chemical Industry CO./LTD, Japan), 3,4-diaminopyridine (3,4-DAP; Sigma-Aldrich, USA)
- 84 and μ-conotoxin GIIIB (Bachem, Switzerland) were of the highest grade available.

85 2.1.2 Experimental animals and neuromuscular preparations

- 86 Adult male Balb/C mice were obtained from the animal breeding facility at the Veterinary
- 87 Faculty (University of Ljubljana, Slovenia). All the experiments followed ethical standards
- and were approved by the administration of the Republic of Slovenia for food safety,
- veterinary and plant protection with permit no. 34401-20/2009/30.
- 90 Mice were sacrificed by cervical dislocation followed by immediate exsanguination. The
- 91 diaphragm muscle along with both phrenic nerves was dissected and split into two
- 92 neuromuscular preparations. Each hemidiaphragm was maintained in oxygenated standard
- Krebs-Ringer solution composed of the following: 154 mM NaCl, 5 mM KCl, 2 mM CaCl₂, 1
- 94 mM MgCl₂, 5 mM HEPES and 11 mM D-glucose (pH 7.4). All experiments were performed
- 95 at 22-24°C.

97

96 **2.2 Methods**

2.2.1 Muscle twitch and tetanus recordings

- 98 The hemidiaphragm was pinned to the organ bath on its lateral side. It was connected to an
- 99 isometric mechano-electrical transducer (Grass Instruments, West Warwick, RI, USA) by
- silky thread via a stainless-steel hook, attached to its tendinous part. The motor nerve of the
- isolated neuromuscular preparation was stimulated using a square pulse S-48 stimulator
- 102 (Grass Instruments, West Warwick, RI, USA) via a suction electrode with pulses of 0.1 ms
- and supramaximal voltage (typically 5-10 V) applied at a 0.1-Hz frequency. Direct muscle
- stimulation was evoked by electric field stimulation via a platinum electrode assembly in the
- organ bath connected to the S-48 Grass stimulator and producing pulses of 0.1 ms and 50-120
- V applied at a 0.1-Hz frequency. Directly- (via muscle stimulation) or indirectly- (via nerve
- stimulation) evoked tetanic contraction was obtained with a pulse train duration of 1000 ms
- and a stimulation rate of 80 Hz. To achieve the maximal contractile response upon nerve-
- evoked stimulation, the stable resting tension for each neuromuscular preparation (typically
- 1.5-2.5 g) was adjusted approximately 20 min before starting the experiment. Muscle twitch
- or tetanic tension was measured using a Grass FT03 force transducer (Grass Instruments,
- West Warwick, RI, USA). Electrical signals were amplified using a P122 strain gauge
- amplifier (Grass Instruments, West Warwick, RI, USA) and digitized using a data acquisition
- system (Digidata 1440A; Molecular Devices, Sunnyvale, CA, USA) at a sampling rate of 1
- kHz. The inhibitory response of nPoly-3-APS was continuously measured during 60-90 min
- after compound application. The muscle twitch or tetanic tension blockade produced by
- 117 nPoly-3-APS was expressed as the percentage of the maximal response recorded before
- compound application. The concentrations of nPoly-3-APS used were 9.1, 18.1, 27.2, 36.2,
- 119 45.3 and 54.4 μM.

120

2.2.2 Membrane potential recordings

- Hemidiaphragm preparations were equilibrated for 30 min in standard Krebs-Ringer solution
- added with 1.6 μM μ-conotoxin GIIIB, which blocks Na_V1.4 muscle sodium channel subtypes

(Cruz et al., 1985; Hong and Chang, 1989) and allows the recording of the full-sized amplitude of the endplate potentials (EPPs). Miniature endplate potentials (MEPPs), EPPs and the resting membrane potential (rV_m) were recorded from superficial hemidiaphragm muscle fibers using intracellular borosilicate glass microelectrodes and Axoclamp 900A microelectrode amplifier (Molecular Devices, Sunnyvale, CA, USA). Microelectrodes were pulled using a P-97 Flaming/Brown micropipette puller (Sutter Instruments, Novato, CA, USA) and were filled with 3 M KCl solution. Only microelectrodes with resistance of 10-20 $M\Omega$ were used. Endplate regions, where EPPs were recorded, were indicated by the presence of MEPPs. EPPs were evoked by stimulating the phrenic nerve via the bipolar suction electrode with supramaximal square pulses of 0.1 ms at 0.5 Hz using the S-48 Grass stimulator. MEPPs and EPPs were digitized at 25 kHz using Digidata 1440A and pCLAMP 10 (Molecular Devices, Sunnyvale, CA, USA) and were stored for later analysis using pCLAMP-Clampfit 10 software (Molecular Devices, Union City, CA, USA). The recordings were performed before treatment, 45 and 90 min after the application of nPoly-3-APS and 15 min after the compound wash-out. MEPP and EPP amplitudes were normalized to a rV_m of -70 mV using the formula $V_c = V_o x (-70)/rV_m$, where V_c is the normalized amplitude of MEPPs and EPPs, and V_o is their recorded amplitude (Pardo et al., 2006). The concentrations of nPoly-3-APS used were 1.8, 9.1, 18.1, 27.2, 36.2 and 54.4 μM.

141 2.2.3 Data analysis and statistics

The results are presented as the means \pm S.E. The nPoly-3-APS concentrations producing 50% inhibition of initial responses (IC₅₀) were determined by fitting the concentration-response relationships using the four-parameter nonlinear regression model (GraphPad Prism version 6.00). The data were statistically analyzed using Sigma Plot for Windows version 12.5 (Systat Software Inc., USA). Student's two-tailed *t*-test was used for statistical analysis of the data. Statistical significance was set at a *P* value \leq 0.05.

3 Results

3.1 nPoly-3-APS blocks muscle contraction in vitro

The effects of natural nPoly-3-APS (9.1-54.4 μM) were first determined on nerve-evoked and directly muscle-elicited isometric twitch and tetanic contraction in mouse hemidiaphragm preparations. A representative example of the time-course of nPoly-3-APS effects on muscle contraction is shown in Fig. 1A. The compound (36.2 μM) blocked nerve-evoked twitch and tetanic contraction within 90 min (Fig. 1A and B). This effect was in part reversible since, after about 30 min of thorough washing of the preparation with standard physiological solution, the nerve-evoked muscle twitch and tetanic contraction were restored partially (Fig. 1A). In contrast, directly muscle-elicited twitch and tetanic contraction were much less affected by a similar compound concentration (Fig. 1A and C). The nPoly-3-APS-induced block of muscle contraction occurred in a concentration-dependent manner (Fig. 1D and E). The IC₅₀ values, calculated from the concentration-response curves, were 18.5-29.4 μM for nerve-evoked muscle contraction, and 56-90 μM for directly muscle-elicited contraction (Fig. 1E). Therefore, nPoly-3-APS was more than 3 times more efficient to block nerve-evoked than directly muscle-elicited twitch and tetanic contraction. It is worth noting that no decrease

of muscle contraction amplitude occurred during 90-min control experiments during which neuromuscular preparations were only bathed with Krebs-Ringer solution (data not shown).

Figure 1 near here

- During the development of the contraction block, obtained 30 min after the application of
- 36.2 μM nPoly-3-APS (i.e. $33 \pm 4.4\%$, n = 4), 3 μM neostigmine, which is known to inhibit
- 170 AChE (Alderdice, 1982), did not modify muscle twitch and tetanic contraction (data not
- shown). Additionally, 300 µM 3,4-diaminopyridine, a well-known potassium channel blocker
- in nerve terminals (Molgo et al., 1980), significantly increased the twitch and tetanic
- 173 contraction amplitudes to approximately $120 \pm 11\%$ (n = 4) of their initial value (data not
- 174 shown).

167

3.2 Effects of nPoly-3-APS on rV_m, MEPPs and EPPs

- In a second set of experiments, we tested whether nPoly-3-APS has some effects on rV_m,
- 177 MEPPs and EPPs since the compound was shown to block preferentially nerve-evoked
- muscle twitch and tetanic contraction.

179 3.2.1 nPoly-3-APS does not affect rV_m

- nPoly-3-APS (1.8-54.4 μM), i.e. at concentrations that partially or completely block nerve-
- evoked muscle twitch and tetanic contraction in mouse hemidiaphragms (see Fig. 1D and E),
- did not significantly affect the rV_m of muscle fibers recorded in the endplate regions of the
- surface fibers after 45 and 90 min of compound application (Fig. 2). In particular, the mean
- 184 rV_m was -65.5 \pm 1.7 mV under control conditions (i.e. at time zero, before compound
- application), and -64.3 \pm 1.5 mV and -61.2 \pm 0.6 mV, 45 and 90 min after exposure to 54.4
- 186 μM nPoly-3-APS, respectively.

Figure 2 near here

3.2.2 nPoly-3-APS reduces MEPP and EPP amplitudes

- The effects of nPoly-3-APS on the MEPP amplitude and frequency are shown in Fig. 3.
- 190 nPoly-3-APS (9.1-27.2 μM), in a concentration-dependent manner, significantly reduced the
- amplitude of MEPPs recorded in superficial muscle fibers of the mouse hemidiaphragm (Fig.
- 192 3A and B), without significant decrease of their frequency except at the highest compound
- concentration (Fig. 3C). At higher concentrations (36.2-54.4 µM), nPoly-3-APS completely
- abolished MEPPs (Fig. 3A). No significant change in the EPP half-decay time was observed
- among the concentrations studied (Fig. 3A).

Figure 3 near here

- 197 Electrophysiological recordings of EPPs from the endplates of the mouse hemidiaphragm
- exposed to nPoly-3-APS (1.8-54.4 μM) for 90 min revealed that the compound reduced their
- amplitude in a concentration-dependent manner (Fig. 4). The IC₅₀ calculated value for the
- 200 nPoly-3-APS-induced reduction of the EPP amplitude was 20.1 μM (Fig. 4B). No significant
- 201 change in the EPP half-decay time was observed among the concentrations studied (Fig. 4A).

Figure 4 near here

203

4. Discussion

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219220

221

222

223224

225

226

227

228

229

230231

232

233

234

235

236

237

238

239

240

241242

243

244

245

246

247

This study shows, for the first time, the effects of natural alkylpyridinium polymer (nPoly-3-APS), produced by the marine sponge Reniera sarai, on neuromuscular transmission and skeletal muscle functioning. In a previous study, it was shown that nPoly-3-APS after i.v. administration at a sublethal dose to male Wistar rats increased the residual respiratory volume, prolonged expirium, produced bradypnea; at a lethal dose, nPoly-3-APS causes respiratory arrest (Bunc et al., 2000). However, nPoly-3-APS showed many interesting biological activities (reviewed in Turk et al., 2007). Its application in medicine led to the synthesis of a series of synthetic analogs with different degrees of polymerization and different lengths of alkyl chains (Houssen et al., 2010). Recently, the effects of some of these synthetic analogs due to their AChE inhibitory potency were studied on the skeletal neuromuscular transmission (reviewed in Grandic and Frangez, 2014). Experiments on skeletal neuromuscular preparations revealed that APS12-2, a non-competitive AChE inhibitor which was previously shown to be selectively cytotoxic toward NSLC cells in vitro, is a potent antagonist of skeletal muscle nAChRs (Grandic et al., 2012). Later, it was shown that the synthetic polymeric 3-alkylpyridinium salt (APS3) in vitro inhibits nerve-evoked muscle contraction of the mouse hemidiaphragm due to specific block of muscle-type nAChRs with an IC₅₀ of 20.3 µM (Grandic et al., 2012). Therefore, it was of interest to examine the natural molecule nPoly-3-APS for similar biological effects on skeletal neuromuscular transmission and to propose different mechanistic hypotheses to explain its underlying molecular mechanism of action.

Our results obtained from phrenic nerve-hemidiaphragm preparations are summarized in Table 1. They show that nPoly-3-APS, at concentrations ranging from 9.1 to 54.4 μM, progressively blocked the maximal amplitude of nerve-evoked isometric muscle twitch, sustained (tetanic) contraction and EPPs, without any noticeable change in their decay phase. Additionally, relatively small decrease of maximal amplitude was observed on directly muscle-elicited twitch and tetanic contraction. The natural compound, without notably affecting the frequency (except a reduction observed in the presence of the highest compound concentration) and the decay phase, decreased the MEPP amplitude measured in hemidiaphragm superficial muscle fibers. Taking into account that the muscle fiber rV_m remained unchanged even after the longest exposure to the highest concentration of nPoly-3-APS used, these results indicate a preferential postsynaptic activity of the natural compound. Indeed, they are exactly what is expected if nPoly-3-APS produces muscle-type nAChR blockage (Table 1). This mechanistic hypothesis was previously proposed to explain the effects of APS12-2 and APS3, two synthetic analogs of polymeric alkylpyridinium salts isolated from Reniera sarai, which were similar to those of the natural compound (Grandic et al., 2012; Grandic et al., 2013). Additionally, available information in the literature indicates that 3-alkylpyridinium polymers from the marine sponge *Reniera sarai*, at approximatively 200-2000 times lower concentrations than those used in the present study, possess strong AChE activity, and a detailed mechanism of kinetics of AChE inhibition has been proposed (Sepcic et al., 1997; Sepcic et al., 1998; reviewed in Turk et al., 2007). The lack of AChE activity at the neuromuscular junction level allows acetylcholine (ACh) to accumulate in the synaptic cleft and thus to activate repetitively nAChRs (Van der Kloot et al., 1994). It is characterized, in particular, by muscle inability to sustain a tetanic contraction under repetitive high frequency stimulation of the motor nerve, due to both ACh accumulation and decreased rV_m, by enhancement of nerve-evoked muscle twitch, EPP and MEPP amplitudes, as well as MEPP frequency, and by prolongation of their decay phase (Table 1; Chang et al., 1986; Katz and Miledi, 1973). However, no physiological evidence was obtained in the present study of an eventual anti-AChE effect of nPoly-3-APS since we did not observe any of these AChE inhibition characteristics (Table 1). Furthermore, the nerve-evoked muscle twitch block was not antagonized/augmented by 3 µM neostigmine, which almost completely inhibits AChE in the neuromuscular junctions of the mouse hemidiaphragm (Minic et al., 2003). Possible involvement of anti-AChE activity of the natural compound is therefore excluded in the present results. Different hypotheses can be proposed to explain the absence of an observation of nPoly-3-APS-induced AChE inhibition at the level of mouse skeletal neuromuscular junction. The first is that such an inhibition is masked by the nPoly-3-APSproduced blockage of muscle-type nAChRs. However, none of the AChE inhibition characteristics were observed at low concentrations (1.8-9.1 µM) which produced no marked nAChR blockage even if they were 100-1000 times higher than those reported previously to inhibit AChE (Sepcic et al., 1997). The second hypothesis is the polycationic nature of nPoly-3-APS, leading to limited ability of the natural compound to reach its binding site since its AChE activity was previously shown to occur only in aqueous solutions (Sepcic et al., 1997), which may also have an impact on its activity in biological systems. Finally, it is worth noting that these authors reported only inhibition of recombinant insect, electric eel and human erythrocyte AChE, as well as equine BuChe, without any information on mammal neuronal AChE. Knowing that there is low percentage of similarity between mammal (human and mouse), insect and fish AChE amino-acid sequences (Wiesner et al., 2007), one cannot exclude that the affinity of nPoly-3-APS for mouse AChE is very low compared to that for mouse muscle-type AChRs.

248

249

250

251

252253

254

255

256

257258

259

260

261262

263

264

265

266

267

268

269270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285286

287

288

289

290

Table 1 near here

The partial block of nerve-evoked muscle twitch produced by nPoly-3-APS was completely reversed by the potassium channel blocker 3,4-diaminopyridine, known to prolong the duration of action potential, to increase both Ca²⁺ entry into nerve terminals and ACh release from motor terminals (Molgo et al., 1980; Hong and Chang, 1990). Antagonist effects of 3,4diaminopyridine on mouse hemidiaphragm muscle, partially paralyzed by the action of nPoly-3-APS, further suggest postsynaptic effects. In the range of the concentrations studied, nPoly-3-APS also showed a concentration-dependent effect on muscle excitability and excitationcontraction coupling processes, as determined by the observed small blockade of contractile responses by the compound (18.1-54.4 µM) upon single and tetanic direct muscle stimulation. nPoly-3-APS has been previously reported to be hemolytic and cytotoxic against different mammalian cell lines (Malovrh et al., 1999) and to produce concentration-dependent reversible/irreversible plasma membrane depolarization in neuronal cells (McClelland et al., 2003). Persistent depolarization may affect muscle excitability and directly-elicited muscle contractility (for review see Ulbricht, 2005). However, our results clearly show that the compound-induced small decrease of directly muscle-elicited twitch and tetanic contraction cannot be explained by a reduction of rV_m.

In conclusion, our results reveal, for the first time, that micromolar concentrations of nPoly-3-

APS completely inhibit nerve-evoked twitch and tetanic contraction as well as, although 291 292 partly, directly muscle-elicited twitch and tetanic contraction. No evidence was obtained in the present study for an eventual anticholinesterase effect of nPoly-3-APS. The in vivo 293 toxicity of the natural compound is thus likely to occur through a preferential inhibition of 294 295 neuromuscular transmission. The most likely mechanistic hypothesis to explain such an 296 inhibition is a slowly reversible antagonist effect on muscle-type nAChRs since nPoly-3-APS reduced the number of open nAChRs by altering neither muscle fiber passive membrane 297 properties nor neurotransmitter ACh release. This blocking nAChR activity remains however 298 to be confirmed by studying the natural compound effects on muscle-type nAChRs expressed 299 300 or incorporated in *Xenopus* oocytes.

301

302

Funding information

- 303 This work was supported by Slovenian research grant [P4-0053] and French/Slovenian
- 304 Proteus grant [BI-FR-PROTEUS/17-18-001, 37446QC].

305

306

Compliance with Ethical Standards

- 307 All experiments on isolated tissue followed ethical standards and were approved by The
- 308 Administration of the Republic of Slovenia for food safety, veterinary and plant protection (34401-
- 309 20/2009/30). The manuscript does not contain clinical studies or patient data.

310

311

Conflicts of interest

312 The authors declare that there are no conflicts of interest.

313

314

Acknowledgments

- 315 The authors gratefully acknowledge American Journal Experts for the English proofreading
- of the manuscript, and thank Boštjan Drolc and Katarina Babnik for excellent technical
- 317 support.

318

319

References

- Alderdice, M.T., 1982. Further comparison of the effects of physostigmine and neostigmine
- on frog neuromuscular transmission. Clin. Exp. Pharmacol. Physiol. 9, 35-43. doi:
- 322 10.1111/j.1440-1681.1982.tb00776.x
- Bunc, M., Sepcic, K., Turk, T., and Suput, D., 2000. In vivo effects of head-to-tail 3-
- 324 alkylpiridinium polymers isolated from the marine sponge *Raniera sarai*. Pflugers Arch.
- 325 440(Suppl 1), R173–R174. doi:10.1007/s004240000052
- 326 Catassi, A., Servent, D., Paleari, L., Cesario, A., and Russo, P., 2008. Multiple roles of
- nicotine on cell proliferation and inhibition of apoptosis: Implications on lung carcinogenesis.
- 328 Mutat. Res. 659, 221–231. doi: 10.1016/j.mrrev.2008.04.002

- Chang, C.C., Hong, S.J., and Ko, J.L., 1986. Mechanisms of the inhibition by neostigmine of
- 330 tetanic contraction in the mouse diaphragm. Br. J. Pharmacol. 87, 757-762. doi:
- 331 10.1111/j.1476-5381.1986.tb14594.x
- Chelossi, E., Mancini, I., Sepcic, K., Turk, T., and Faimali, M., 2006. Comparative
- antibacterial activity of polymeric 3-alkylpyridinium salts isolated from the Mediterranean
- sponge Reniera sarai and their synthetic analogues. Biomol. Eng. 23, 317–323. doi:
- 335 10.1016/j.bioeng.2006.09.005
- 336 Cruz, L.J., Gray, W.R., Olivera, B.M., Zeikus, R.D., Kerr, L., Yoshikami, D., and
- 337 Moczydlowski, E., 1985. Conus geographus toxins that discriminate between neuronal and
- 338 muscle sodium channels. J. Biol. Chem. 260, 9280–9288.
- Elersek, T., Kosi, G., Turk, T., Pohleven, F., and Sepcic, K., 2008. Influence of polymeric 3-
- alkylpyridinium salts from the marine sponge *Reniera sarai* on the growth of algae and wood
- decay fungi. Biofouling 24, 137–143. doi: 10.1080/08927010801935745
- Faimali, M., Sepcic, K., Turk, T., and Geraci, S., 2003. Non-toxic antifouling activity of
- 343 polymeric 3-alkylpyridinium salts from the Mediterranean sponge Reniera sarai (Pulitzer-
- 344 Finali). Biofouling 19, 47–56. doi: 10.1080/0892701021000036966
- Grandic, M., and Frangez, R., 2014. Pathophysiological effects of synthetic derivatives of
- polymeric alkylpyridinium salts from the marine sponge, Reniera sarai. Mar. Drugs 12,
- 347 2408–2421. doi: 10.3390/md12052408
- Grandic, M., Araoz, R., Molgò, J., Turk, T., Sepcic, K., Benoit, E., and Frangez, R., 2013.
- Toxicity of the synthetic polymeric 3-alkylpyridinium salt (APS3) is due to specific block of
- nicotinic acetylcholine receptors. Toxicology 303, 25–33. doi: 10.1016/j.tox.2012.10.013
- 351 Grandic, M., Araoz, R., Molgò, J., Turk, T., Sepcic, K., Benoit, E., and Frangez, R., 2012.
- 352 The non-competitive acetylcholinesterase inhibitor APS12-2 is a potent antagonist of skeletal
- muscle nicotinic acetylcholine receptors. Toxicol. Appl. Pharmacol. 265, 221–228. doi:
- 354 10.1016/j.taap.2012.09.024
- 355 Grandic, M., Sepcic, K., Turk, T., Juntes, P., and Frangez, R., 2011. In vivo toxic and lethal
- 356 cardiovascular effects of a synthetic polymeric 1,3-dodecylpyridinium salt in rodents.
- 357 Toxicol. Appl. Pharmacol. 255, 86–93. doi: 10.1016/j.taap.2011.06.003
- Hong, S.J., and Chang, C.C., 1989. Use of geographutoxin II (mu-conotoxin) for the study of
- neuromuscular transmission in mouse. Br. J. Pharmacol. 97, 934–940. doi: 10.1111/j.1476-
- 360 5381.1989.tb12034.x
- Hong, S.J., and Chang, C.C., 1990. Facilitation by 3,4-diaminopyridine of regenerative
- acetylcholine release from mouse motor nerve. Br. J. Pharmacol. 101, 793-798. doi:
- 363 10.1111/j.1476-5381
- Houssen, W.E., Lu, Z., Edrada-Ebel, R., Chatzi, C., Tucker, S.J., Sepcic, K., Turk, T., Zovko,
- 365 A., Shen, S., Mancini, I., Scott, R.H., and Jaspars, M., 2010. Chemical synthesis and
- 366 biological activities of 3-alkyl pyridinium polymeric analogues of marine toxins. J. Chem.
- 367 Biol. 3, 113–125. doi: 10.1007/s12154-010-0036-4
- Katz, B., and Miledi, R., 1973. The binding of acetylcholine to receptors and its removal from

- 369 the synaptic cleft. .J Physiol. 231, 549–574.
- Malovrh, P., Sepcic, K., Turk, T., and Macek, P., 1999. Characterization of hemolytic activity
- of 3-alkylpyridinium polymers from the marine sponge Reniera sarai. Comp. Biochem.
- 372 Physiol. C Pharmacol. Toxicol. Endocrinol. 124, 221-226. doi: 10.1016/S0742-
- 373 8413(99)00067-5
- McClelland, D., Evans, R.M., Abidin, I., Sharma, S., Choudhry, F.Z., Jaspars, M., Sepcic, K.,
- and Scott, R.H., 2003. Irreversible and reversible pore formation by polymeric
- alkylpyridinium salts (poly-APS) from the sponge Reniera sarai. Br. J. Pharmacol. 139,
- 377 1399–1408. doi: 10.1038/sj.bjp.0705374
- 378 Minic, J., Chatonnet, A., Krejci, E., and Molgó, J., 2003. Butyrylcholinesterase and
- 379 acetylcholinesterase activity and quantal transmitter release at normal and
- acetylcholinesterase knockout mouse neuromuscular junctions. Br. J. Pharmacol. 138, 177-
- 381 187. doi: 10.1038/sj.bjp.0705010
- Molgo, J., Lundh, H., and Thesleff, S., 1980. Potency of 3,4-diaminopyridine and 4-
- aminopyridine on mammalian neuromuscular transmission and the effect of pH changes. Eur.
- J. Pharmacol. 61, 25–34. doi: 10.1016/0014-2999(80)90378-7
- Paleari, L., Trombino, S., Falugi, C., Gallus, L., Carlone, S., Angelini, C., Sepcic, K., Turk,
- 386 T., Faimali, M., Noonan, D.M., and Albini, A., 2006. Marine sponge-derived polymeric
- 387 alkylpyridinium salts as a novel tumor chemotherapeutic targeting the cholinergic system in
- 388 lung tumors. Int. J. Oncol. 29, 1381–1388. doi: 10.3892/ijo.29.6.1381
- Pardo, N.E., Hajela, R.K., and Atchison, W.D., 2006. Acetylcholine release at neuromuscular
- junctions of adult tottering mice is controlled by N-(cav2.2) and R-type (cav2.3) but not L-
- 391 type (cav1.2) Ca2+ channels. J. Pharmacol. Exp. Ther. 319, 1009–1020. doi:
- 392 10.1124/jpet.106.108670
- Sepcic, K., Guella, G., Mancini, I., Pietra, F., Serra, M.D., Menestrina, G., Tubbs, K., Macek,
- P., and Turk, T., 1997. Characterization of anticholinesterase-active 3-alkylpyridinium
- polymers from the marine sponge *Reniera sarai* in aqueous solutions. J. Nat. Prod. 60, 991–
- 396 996. doi: 10.1021/np970292q
- Sepcic, K., Marcel, V., Klaebe, A., Turk, T., Suput, D., and Fournier, D., 1998. Inhibition of
- 398 acetylcholinesterase by an alkylpyridinium polymer from the marine sponge, *Reniera sarai*.
- 399 Biochim. Biophys. Acta. 1387, 217–225. doi: 10.1016/S0167-4838(98)00125-3
- Tucker, S.J., McClelland, D., Jaspars, M., Sepcic, K., MacEwan, D.J., and Scott, R.H., 2003.
- 401 The influence of alkyl pyridinium sponge toxins on membrane properties, cytotoxicity,
- transfection and protein expression in mammalian cells. Biochim. Biophys. Acta 1614, 171-
- 403 181. doi: 10.1016/S0005-2736(03)00175-5
- 404 Turk, T., Frangez, R., and Sepcic, K., 2007. Mechanisms of toxicity of 3-alkylpyridinium
- 405 polymers from marine sponge Reniera sarai. Mar. Drugs 5, 157–167. doi:
- 406 10.3390/MD504157
- 407 Ulbricht, W., 2005. Sodium channel inactivation: Molecular determinants and modulation.
- 408 Physiol. Rev. 85, 1271–1301. doi: 10.1152/physrev.00024.2004

- Van der Kloot, W., Balezina, O.P., Molgò, J., and Naves, L.A., 1994. The timing of channel
- opening during miniature endplate currents at the frog and mouse neuromuscular junctions:
- effects of fasciculin-2, other anti-cholinesterases and vesamicol. Pflugers Arch. 428, 114–126.
- 412 doi: 10.1007/BF00374848
- Wiesner J., Kriz Z., Kuca K., Jun D., Koca J., 2007. Acetylcholinesterases-the structural
- 414 similarities and differences. J Enzyme Inhib Med Chem. 22, 417-424. doi:
- 415 10.1080/14756360701421294
- Zovko, A., Viktorsson, K., Lewensohn, R., Kolosa, K., Filipic, M., Xing, H., Kem, W.R.,
- Paleari, L., and Turk, T., 2013. APS8, a polymeric alkylpyridinium salt blocks alpha7 nAChR
- and induces apoptosis in non-small cell lung carcinoma. Mar. Drugs. 11, 2574–2594. doi:
- 419 10.3390/md11072574

Table 1 Investigated parameters of mouse neuromuscular system in function of protocol stimulation, present results obtained from nPoly-3-APS effects on these parameters, and expected results assuming that the natural compound either inhibits AChE activity or blocks nAChR functioning.

Protocol stimulation	Investigated parameters ¹	Present results ¹	Expected results with nAChR blockage 1	Expected results with AChE inhibition
Nerve	Twitch amplitude	⊻ (important)	Я	7
	Twitch decay phase	→	\rightarrow	7
	Tetanic contraction amplitude	(important) لا	И	И
	Tetanic contraction decay phase	→	→	7
	EPP amplitude	نا (important)	Я	7
	EPP decay phase	→	→	7
Muscle	Twitch amplitude	الا (small)	\rightarrow	\rightarrow
	Twitch decay phase	→	\rightarrow	\rightarrow
	Tetanic contraction amplitude	ע (small)	→	\rightarrow
	Tetanic contraction decay phase	→	\rightarrow	\rightarrow
None	MEPP amplitude	ע (important)	Я	7
	MEPP frequency	∠ (high concentrations)	Я	7
	MEPP decay phase	→	→	7
	Resting membrane potential	→	→	Я

⁴²⁴ Specific effects due to nAChR blockage are underline in grey.

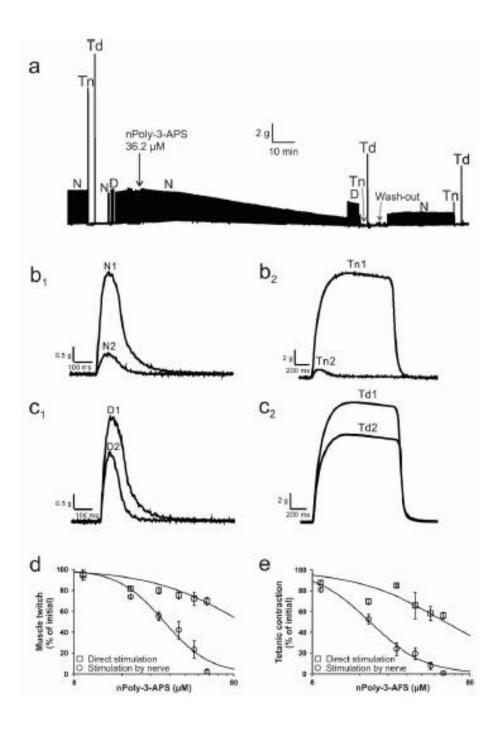


Fig 1 Effects of nPoly-3-APS on nerve-evoked and directly muscle-elicited twitch and tetanic contraction in isolated mouse hemidiaphragm neuromuscular preparations. (a) Representative time-course of nPoly-3-APS (36.2 μM) effects on muscle contraction showing the partial and reversible block of nerve-evoked and directly muscle-elicited twitch and tetanic contraction induced by the compound. N denotes nerve-evoked muscle twitch; D denotes directly muscle-elicited twitch; Tn denotes nerve-evoked tetanic contraction; Td denotes directly muscle-elicited tetanic contraction. (b1) Traces of nerve-evoked twitch recorded before (N1) and 90 min after (N2) the application of 36.2 μM nPoly-3-APS. (b2) Traces of tetanic contraction recorded before (Tn1) and 90 min after (Tn2) the application of 36.2 μM nPoly-3-APS. (c1) Representative recordings of twitch elicited by direct muscle stimulation, before (D1) and 90 min after (D2) the application of 36.2 μM nPoly-3-APS. (c2) Representative recordings of

tetanic contraction elicited by direct muscle stimulation, before (Td1) and 90 min after (Td2) the application of 36.2 μ M nPoly-3-APS. (d) Concentration-response curves for the inhibition of directly muscle-elicited and nerve-evoked twitch by nPoly-3-APS. Muscle twitch was expressed as percentage of its maximal initial value. Each value represents the mean \pm S.E. obtained from five different muscles. IC₅₀ values, determined by fitting the relationships with the four-parameter nonlinear regression model, were 29.4 and 90 μ M for nerve-evoked and directly muscle-elicited twitches, respectively ($r^2 \geq 0.993$). (e) Concentration-response curves for the inhibition of directly muscle-elicited and nerve-evoked tetanic contraction by nPoly-3-APS. Tetanic contraction was expressed as percentage of its maximal initial value. Each value represents the mean \pm S.E. obtained from five different muscles. IC₅₀ values, determined by fitting the relationships with the four-parameter nonlinear regression model, were 18.5 and 56 μ M for nerve-evoked and directly muscle-elicited tetanic contractions, respectively ($r^2 \geq 0.978$)

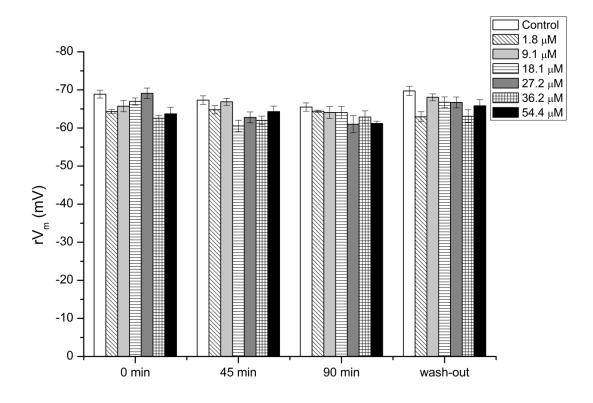


Fig 2 Concentration- and time-dependent effect of nPoly-3-APS on the muscle fiber resting membrane potential (rV_m) of the mouse hemidiaphragm. Neuromuscular preparations were exposed to 1.8-54.4 μ M nPoly-3-APS. Measurements were performed before (time 0), 45 and 90 min after nPoly-3-APS application, and after about 30 min of thorough washing of the preparation with standard physiological solution. Each value represents the mean \pm S.E. of rV_m recorded from 3 different hemidiaphragms (n = 7-10 fibers from each)

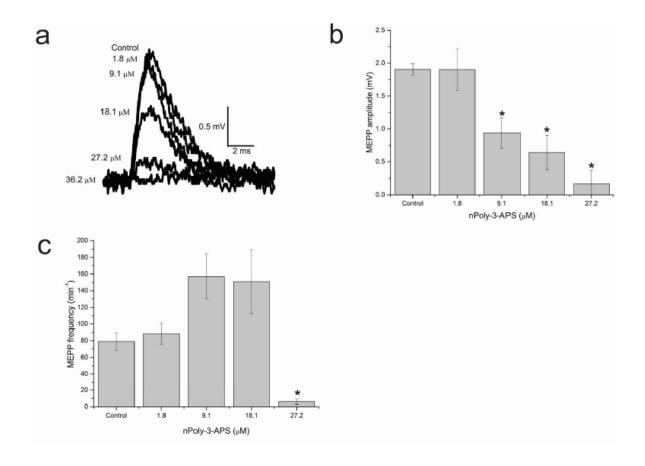


Fig 3 Effects of nPoly-3-APS on the MEPP amplitude and frequency in the muscle fibers of isolated mouse hemidiaphragm. (a) MEPP traces recorded under control conditions and after the indicated concentrations of nPoly-3-APS applied for 90 min. At 36.2 μ M, the compound completely blocked MEPPs. (b) MEPP amplitude and (c) MEPP frequency in muscle fibers exposed to different nPoly-3-APS concentrations. Each bar represents the mean value \pm S.E. from 3 different hemidiaphragms (n = 7-10 fibers from each). * indicates statistically significant difference ($P \le 0.05$)

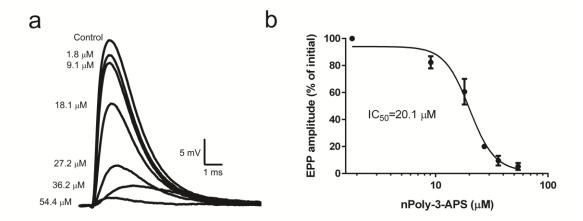


Fig 4 Effects of nPoly-3-APS on EPP amplitude in mouse hemidiaphragm skeletal muscle fibers. The compound blocked EPPs in a concentration-dependent manner. (a) EPP traces recorded under control conditions and 90 min after exposure of muscle fibers to various nPoly-3-APS concentrations. Note that the decrease of EPP amplitudes occurred without significant change in rV_m . (b) Concentration-response curve for the inhibition of EPP amplitude, expressed as a percentage of its maximal initial value, by nPoly-3-APS3. Each value represents the mean \pm S.E. of EPPs recorded from at least 3 different hemidiaphragms (n = 7-10 muscle fibers from each). The IC₅₀ value, determined by fitting the relationship with the four-parameter nonlinear regression model, was 20.1 μ M (r^2 =0.985)