Dairy food structures influence the rates of nutrient digestion

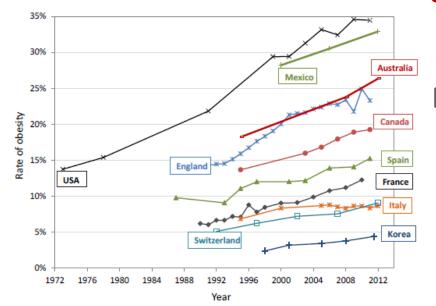


Dr Didier DUPONT

INRA, Rennes, France



Why are we interested in understanding food digestion?



Diet-related diseases ↑

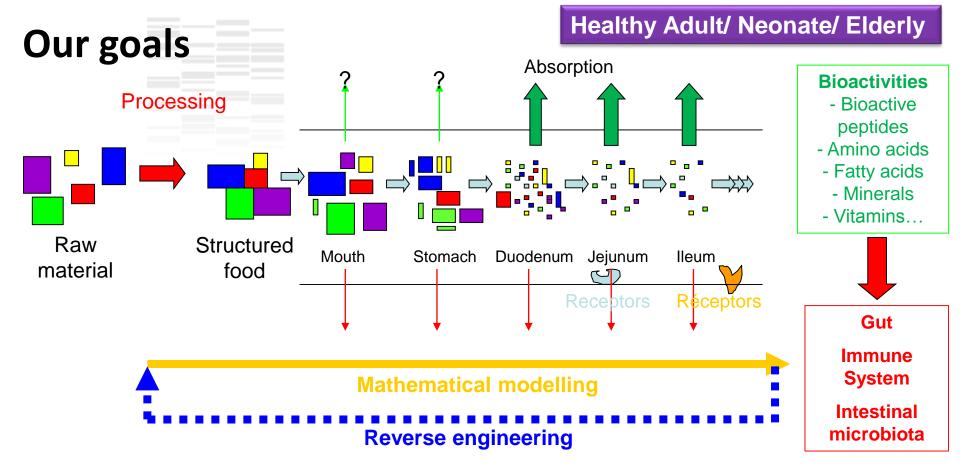
Prevent these pathologies rather than cure them



Gut = interface between food and human body
Digestion releases food components that can have a beneficial or a
deleterious effect on human health

... but the mechanisms of food disintegration in the gastrointestinal tract remain unclear and the digestive process has been considered as a black box so far

By increasing our knowledge on food digestion, we will increase our knowledge on the effect of food on human health

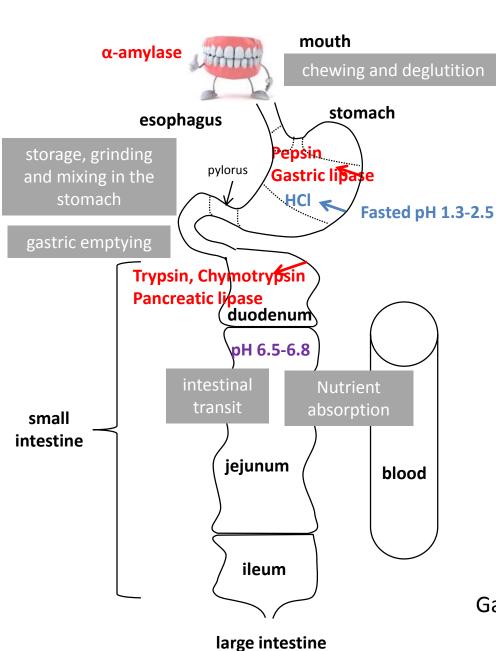


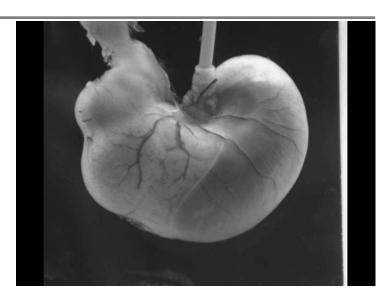
- To understand the mechanisms of breakdown of food matrices and their constituents in the gut and identify the beneficial/deleterious food components released during digestion
- To determine the impact of the structure of food matrices on nutrient bioavailability
- To model these phenomena in order to develop a reverse engineering approach



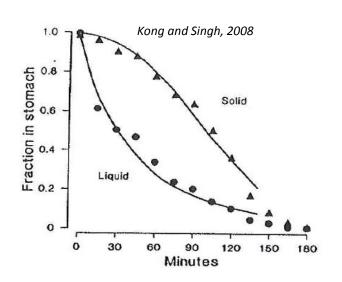


The digestive process



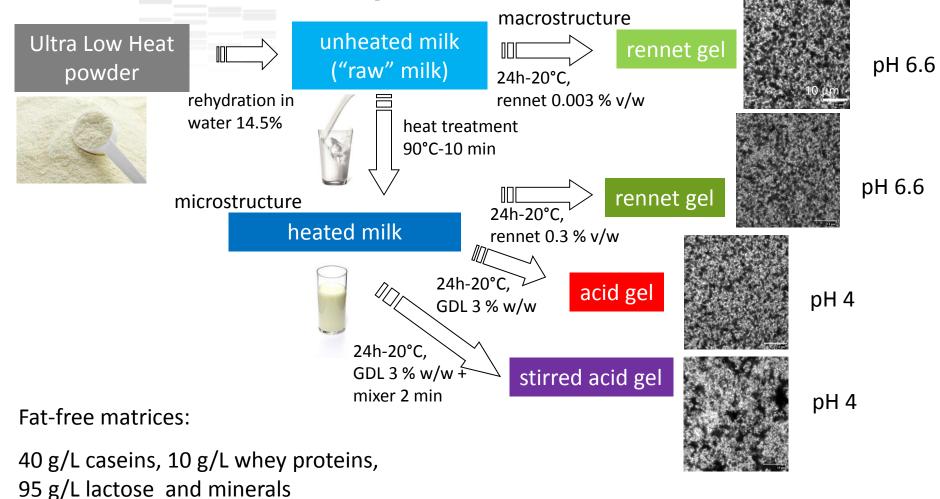


From Roger Lentle, Massey Univ. NZ



Gastric phase = a very complex but crucial step for the whole digestion process

Do dairy products of identical composition but different structure exhibit similar kinetics of digestion?

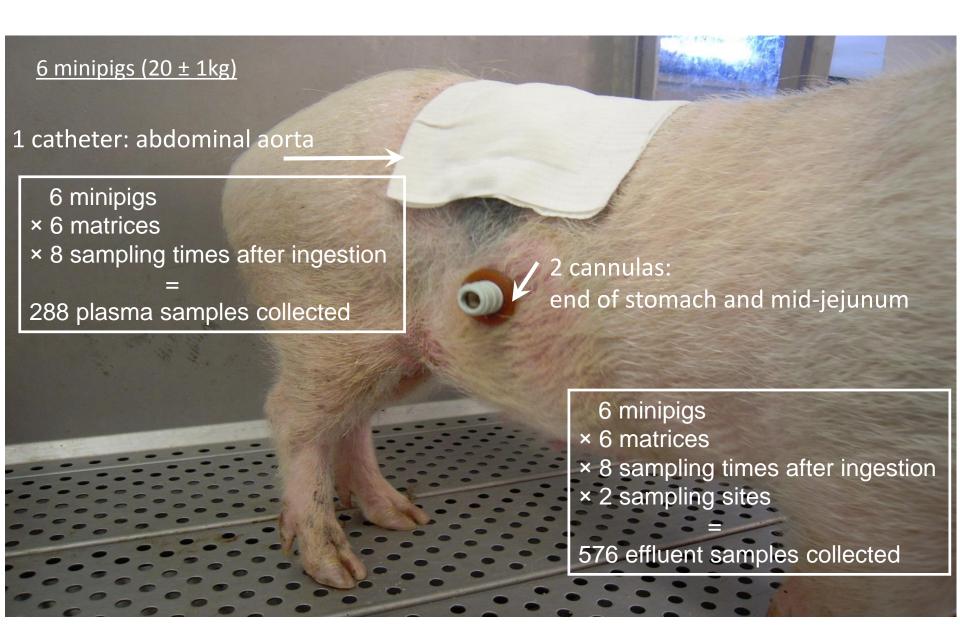


+ marker of the meal transit (Cr^{2+} -EDTA) \rightarrow Mean Retention Time in the stomach

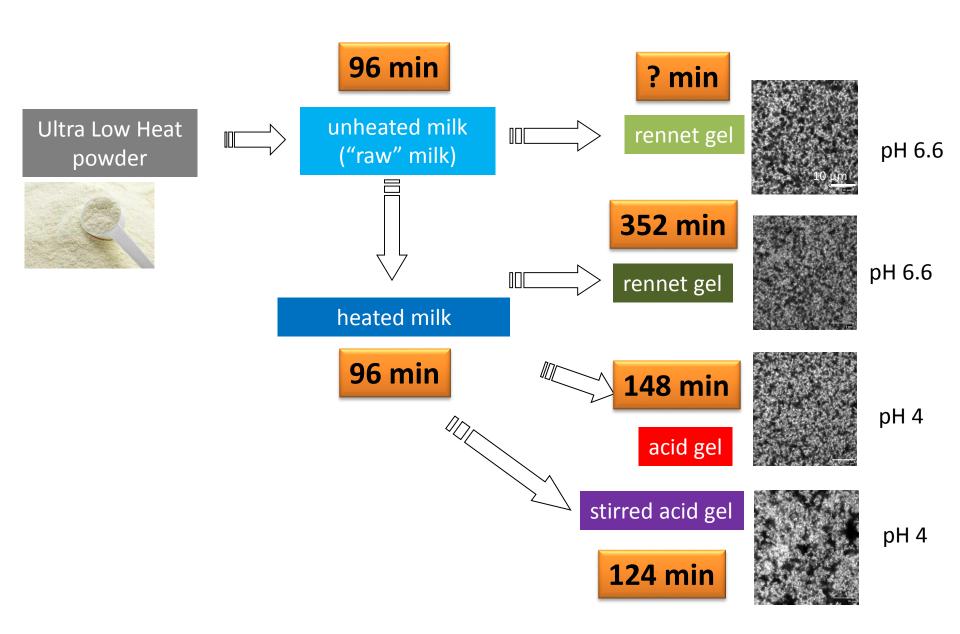




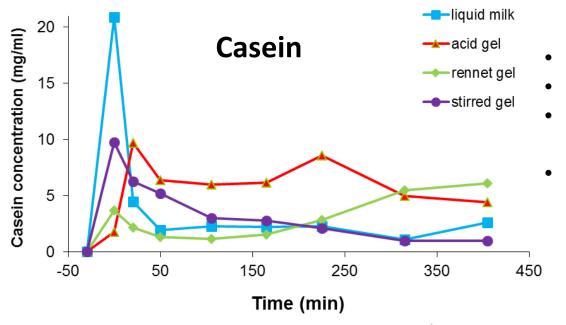
The multi-canulated mini-pigs



Time of residence in the stomach

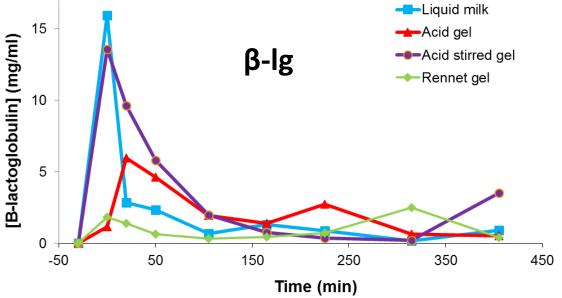


Milk proteins in the duodenum (ELISA)



- Intense and early peak with milk
- Lower and delayed with gels
- Intermediate behaviour with stirred gel
- Low concentrations with rennet gel but casein release tends to increase over time

- Only traces of milk proteins found in the jejunum
- Dairy products remain highly digestible



Bioactive peptides released during digestion differ from one matrix to another

More than 16000 peptides were unambiguously identified!

D	6	A	Defenses		20		405	465	225	245	1
	Sequence	Activity	Reference	4	20	50	105	165	225	315	ı
αs1	1-23	EMUL	Shimizu et al. (1984)								ı
αs1	23-34	HYP	Maruyama & Suzuki (1982)								ı
αs1	30-45	MB	Meisel et al. (1991)								ı
αs1	40-52	MB	Adamson & Reynolds (1996)								ı
as1	43-58	MB	Meisel et al. (1991)								ı
as1	91-100	STRE	Miclo et al. (2001)							ı	ı
as1	99-109	MIC	McCann et al. (2006)								ı
as1	167-180	MIC	Hayes et al. (2006)								ı
αs1	180-193	MIC	Hayes et al. (2006)								ı
αs2	1-24	MB	Miquel et al. (2005)								ı
αs2	124-146	MB	Miquel et al. (2005)								ı
as2	183-206	TRAN	Kizawa et al. (1996)								ı
as2	183-207	MIC	Recio & Visser (1999)								ı
as2	189-197	HYP	Maeno et al. (1996)								ı
as2	190-197	HYP	Maeno et al. (1996)								ı
β	1-24	MB	Bouhallab et al. (1999)								ı
β	33-52	MB	Miquel et al. (2005)								ı
β	60-80	OPI	Jinsmaa & Yoshikawa (1999)								ı
β	98-105	OXI	Rival et al. (2001)								ı
β	114-119	OPI	Jinsmaa & Yoshikawa (1999)								ı
β	132-140	HYP	Robert et al. (2004)								ı
β	192-209	IMM	Coste et al. (1992)								ı
β	193-202	IMM	Kayser & Meisel (1996)								ı
β	193-209	IMM	Coste et al. (1992)								ı
κ	18-24	HYP	Lopez-Exposito et al. (2007)								ı
κ	106-116	THR	Jolles et al. (1986)								ı
β –lg	32-40	HYP	Pihlanto-Leppala et al. (2000)								ı
β –lg	92-100	MIC	Pellegrini et al. (2001)				_				ı
β –lg	142-148	HYP	Mullally et al. (1997)								ı
											ı

Protein	Sequence	Activity	Reference Adamson & Reynolds	4	20	50	105	165	225	315
α s1	40-52	MB	(1996)							
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αs1	167-180	MIC	Hayes et al. (2006)							
as1	180-193	MIC	Hayes et al. (2006)							
α s2	1-24	MB	Miquel et al. (2005)							
α s2	189-197	HYP	Maeno et al. (1996)				<u> </u>			
β	33-52	MB	Miquel et al. (2005)				i			
β	166-175	HYP	Hayes et al. (2007)							
β	193-202	IMM	Kayser & Meisel (1996)							
β –lg	92-100	MIC	(8))							
βlg	142-148	HYP	(9))							

Rennet Gel

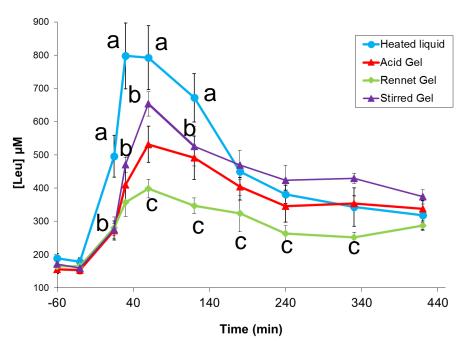
- More bioactive peptides identified during digestion of acid gel than rennet gel
- Nature of peptides is identical (clearly defined by the digestive enzyme specificity)
- Kinetics of release are different

Acid Gel

Barbé et al. 2014 Food Res Int

The liquid-gel transition

2) effect on absorption

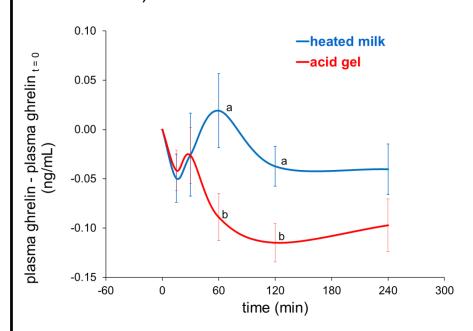


milk gelation:

→ delayed proteins transit → delayed AA absorption

maximal AA concentration in the plasma

3) <u>potential effect on satiety</u> ghrelin (gastrointestinal hormone → appetite stimulation)



milk gelation:

postprandial ghrelin concentration = satiety?





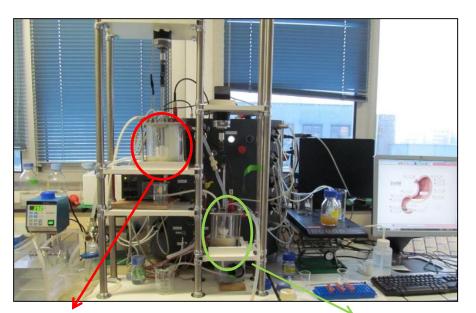




Differential behaviour of acid/rennet gels in gastric conditions

- Acid/Rennet gel: identical composition, similar rheological properties and pore size
- - How can we explain this difference? Dynamic in vitro digestion of the 2 gels

Ménard *et al*. Food Chem 2014



Emptying:

Elashoff's model

DIDGI®

StoRM® software

Stomach

- Pepsine
- Gastric lipase
- Simulated gastric fluid
- HCI



Small intestine



- Pancreatin
- Bile
- Simulated intestinal fluid
- NaHCO₃

Emptying: Elashoff's model

Behaviour of acid and rennet gels in the stomach during in vitro dynamic digestion

Barbé et al. Food Chem. 2014





Formation of a strong coagulum with rennet gel \rightarrow slow down the gastric emptying of caseins

The structure that a food adopts in the stomach is essential to understand its digestion











Understanding the mechanisms of dairy gel particles degradation in the stomach

Soleil is a particle (electron) accelerator that produces the synchrotron radiation, an extremely powerful source of light that permits exploration of inert or living matter



DISCO is a VUV to visible beamline dedicated to biochemistry, chemistry and cell biology. The spectral region is optimized between 60 and 700 nm with conservation of the natural polarization of the light

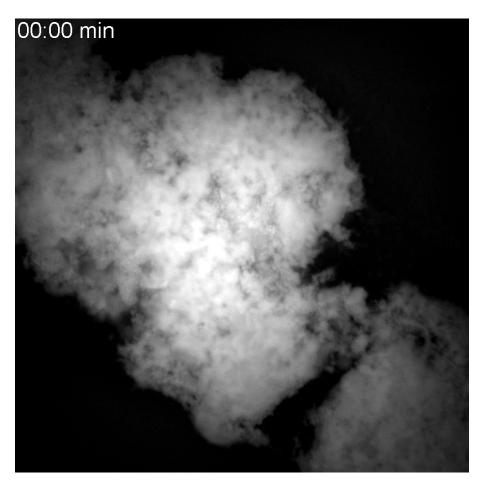
Allow the imaging of protein intrinsic fluorescence with a UV microscope

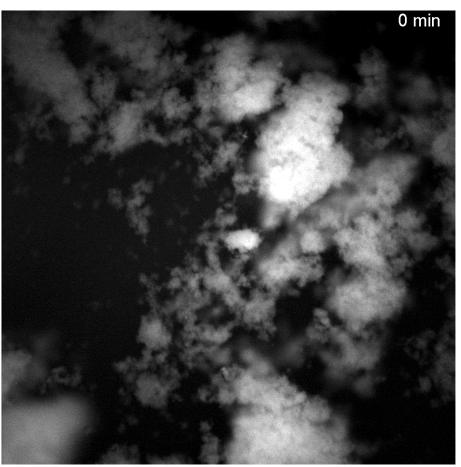




Kinetics of gel particles disintegration

Floury et al. Food Chem. 2017





Rennet Gel

Acid Gel





Understanding how structures of human milk are disintegrated during digestion to create new biomimetic infant formula



CHU Rennes Preterm

In vivo study in the preterm infant

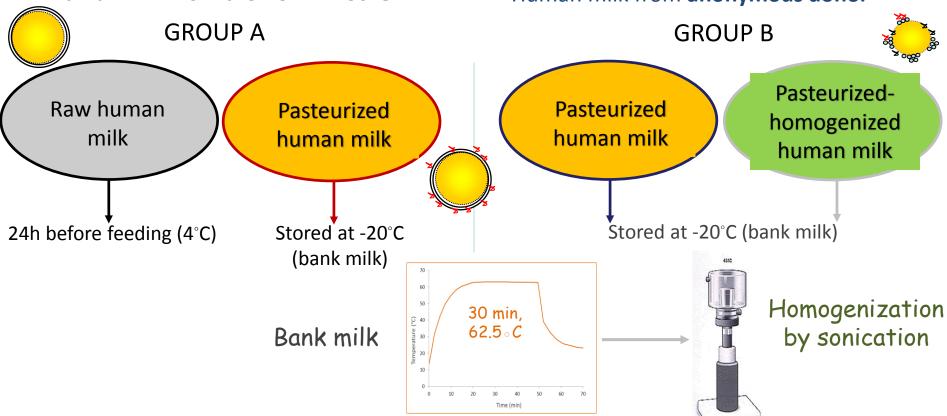
ClinincalTrials.gov Identifier: NCT02112331

ARCHILACT

Preterm hospitalized infants Feeding nasogastric tube Fed every 3 hours

Aim: to compare meals with similar composition but different structure

Human milk from their **own mother**Human milk from **anonymous donor**

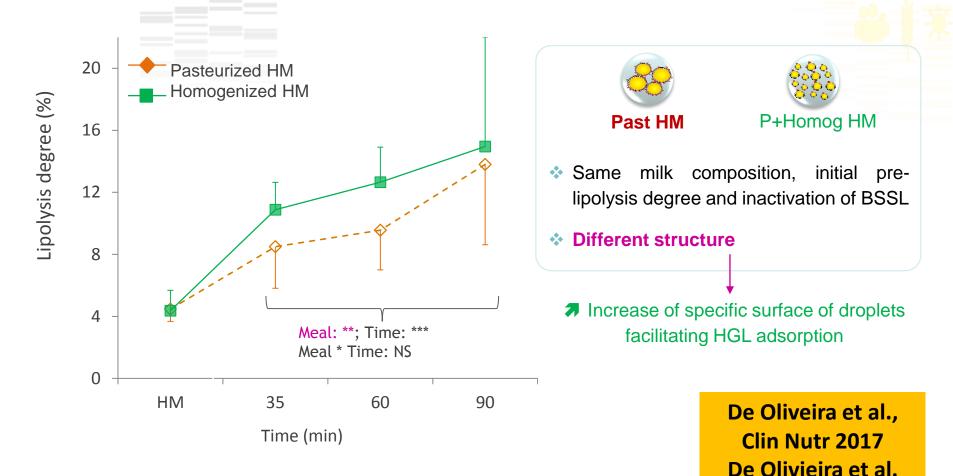








Homogenization impacted gastric lipolysis



Homogenization of human milk: a new way of stimulating the growth and development of pre-term neonate?









Am J Clin, Nutr 2017



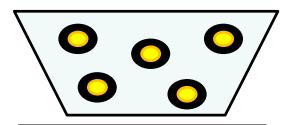


Infant formulas: Can we create lipid structures biomimetic on the native fat globule?



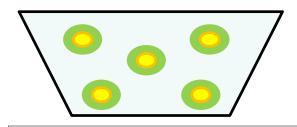
Infant formulas: can we create lipid structures biomimetic on the native fat globule?

Formula T1



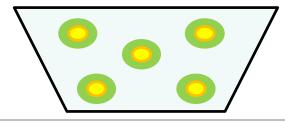
Interface 100 % Proteins 100% vegetable oil

Formula T2

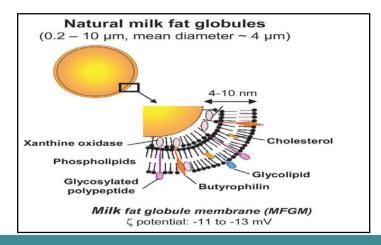


Interface 100 % phospholipids 100% vegetable oil

Formula T3



Interface 100 % phospholipides 40% vegetable oil + 60% milk fat





Lopez, (2007)

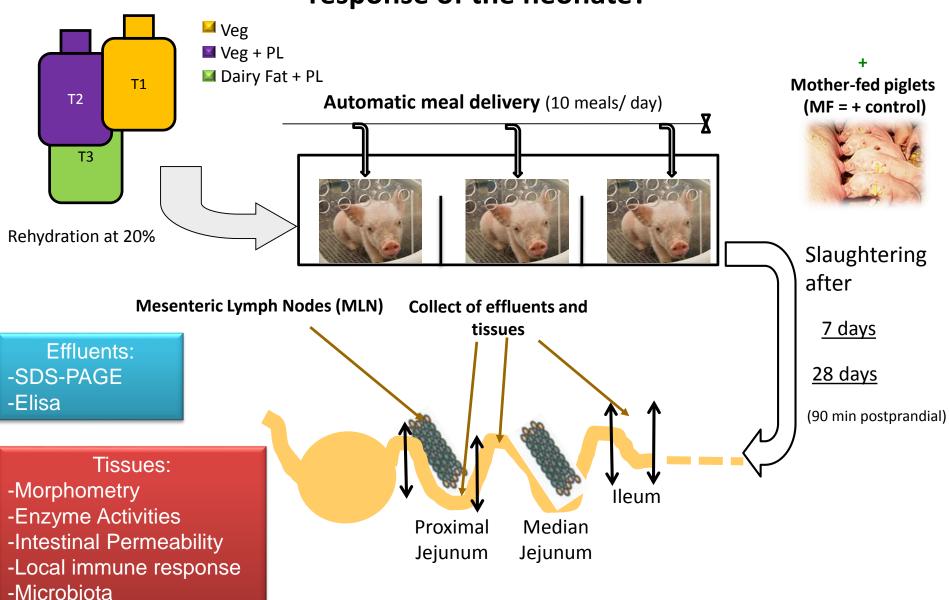




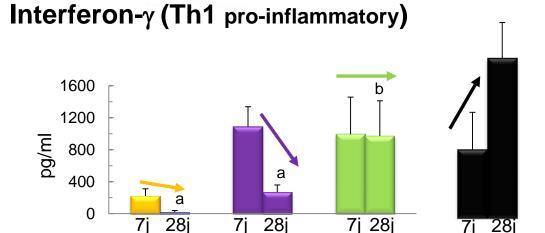


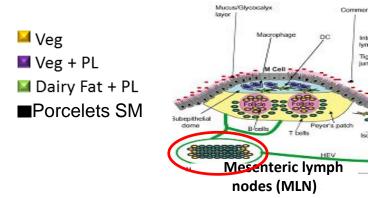


Can the composition of infant formula modulate the physiological response of the neonate?

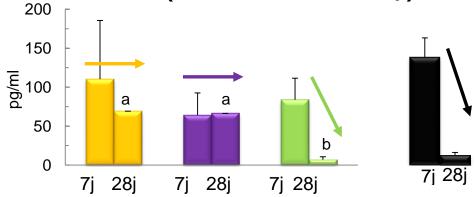


Secretory activity of MLN





Interleukine-10 (Th2 anti-inflammatory)



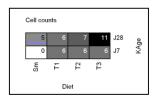
Milk lipids → maturation of the piglet's immune system more similar than with sow's milk

Le Huerou et al. Eur J Nutr 2017







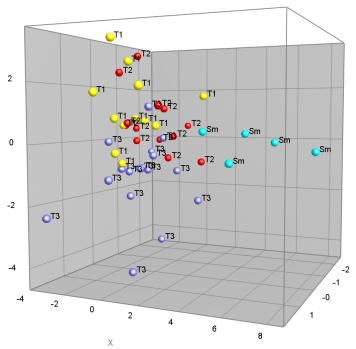


Microbiota by DHPLC



D7 & D28

D28

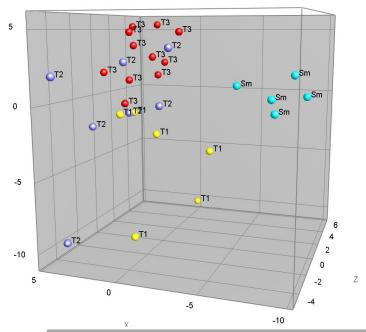


The composition/structure of the infant

formula « orientates » the microbiota

More Proteobacteria with milk fat / More Firmicutes with plant oil

Bourlieu et al. Eur J Lipid Sci Technol 2016





Conclusion

The structure/composition of food regulate the kinetics of protein and lipid digestion and the release of amino acids in the bloodstream

Gastric emptying rate will highly depend on the structure that the product will adopt in the stomach cavity.

Understanding the mechanisms of food particle breakdown in the stomach is critical to control the structure a food will adopt in gastric conditions

Being able to design food structures for controlling the kinetics of hydrolysis of macronutrients will allow to obtain food particularly adapted to specific population











The Bioactivity & Nutrition team

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Amélie DEGLAIRE — Lecturer
Juliane FLOURY — Lecturer
Catherine GUERIN - Lecturer
Joëlle LEONIL — Senior Scientist
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Masters students









Improving health properties of food by sharing our knowledge on the digestive process

International Network

Dr. Didier DUPONT, Senior Scientist, INRA, France









Industry involvement

42 private companies are following INFOGEST























Heinz







triskelion by







Chair Didier Dupont - France

INFOGEST







Vice-chair Alan Mackie - UK

www.cost-infogest.eu

In vitro/in vivo correlations WG1

In vitro semidynamic model of digestion WG2

Models for specific populations WG3

Digestive lipases and lipid digestion WG4

Digestive amylases and starch digestion WG5

In silico models of digestion WG6



Didier Dupont



Alan Mackie



Guy Vergeres



Uri Lesmes

The "Mindthe-Gap" group



Myriam Grundy



Frederic Carriere



Nadja Siegert





Fred Warren



Steven Le Feunteun





Granada, Spain April 2019

We are pleased to announce the next 6th International Conference on Food Digestion



in Granada, Spain, 2-4 April 2019