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**Remobilization of polycyclic aromatic hydrocarbons and organic matter in seawater during sediment resuspension experiments from a polluted coastal environment: insights from Toulon Bay (France)**

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## Abstract

Polycyclic aromatic hydrocarbons (PAHs) and organic matter contents were measured in seawater during resuspension experiments using sediments collected from Toulon Bay (Northwestern Mediterranean Sea, France). The studied sediments were very highly contaminated in PAHs, especially in 4-ring compounds emitted from combustion processes. The sediments used for resuspension experiments were collected at 0-2 cm (diagenetically new organic matter, OM) and 30-32 cm depths (diagenetically transformed OM). They were both mostly composed of fine particles ( $< 63 \mu\text{m}$ ), enriched in organic carbon (8.2 and 6.3%, respectively) and in PAHs (concentration of  $\Sigma 34$  PAHs:  $38.2$  and  $35.2 \times 10^3 \text{ ng g}^{-1}$ , respectively). The resuspension of these sediments led to an increase in concentrations of dissolved  $\Sigma 34$  PAHs, dissolved organic carbon (DOC) and dissolved humic- and tryptophan-like fluorophores in seawater up to 10-, 1.3-, 4.4- and 5.7-fold, respectively. The remobilization in seawater was higher for 4-6 ring PAHs, especially benzo(g,h,i) perylene, whose concentration exceeded the threshold values of the European Water Framework Directive. This noted the potential harmful effects of sediment resuspension on marine biota. From these sediment resuspension experiments, we determined OC-normalized partition coefficients of PAHs between sediment and water ( $K_{oc}$ ) and found that during such events, the transfer of PAHs from sediment particles to seawater was lower than that predicted from octanol-water partition coefficients ( $K_{ow}$ ) (i.e., measured  $K_{oc} > K_{oc}$  predicted from  $K_{ow}$ ). The results confirmed the sequestration role of sedimentary OC quality and grain size on PAHs; the OM diagenetic state seemed to impact the partition process but in a relatively minor way. Furthermore, differences were observed between 2-4 ring and 5-6 ring PAHs, with the latter displaying a relatively higher mobility towards seawater. These differences may be explained by the distribution of these two PAH pools within different OM moieties, such as humic substances and black carbon.

**Keywords:** Sediment resuspension, remobilization, polycyclic aromatic hydrocarbons, fluorescent dissolved organic matter, sediment-water partition.

## 1. Introduction

Polycyclic aromatic hydrocarbons (PAHs) are among the most widespread organic pollutants in marine environments. Due to their hydrophobicity and low water solubility, PAHs entering marine waters tend to sorb onto particulate organic matter (OM) (Gustafsson et al., 2001; Kennish, 1992; May, 1978) that is then deposited to the sediments *via* vertical sinking. This mechanism is recognized as a major pathway for the removal and global cycling of hydrocarbons in the ocean (Adhikari et al., 2015; Berrojalbiz et al., 2011; Gustafsson et al., 1997). However, due to natural (waves, currents, storms) and anthropogenic (dredging, trawling, ship traffic) forcing, coastal sediments are frequently resuspended, refocused and transported along the continental shelf (Durrieu de Madron et al., 2008; Schoellhamer, 1996) and thus may turn into a potential source of organic pollutants, including PAHs, for the water column (Dong et al., 2015; Eggleton and Thomas, 2004). While natural forcing generally induces the resuspension of surface sediments (the first centimeters), human activities, particularly dredging or harbor improvements, may lead to the resuspension over sediment depths of several dozens of centimeters. Dissolved PAHs originating from sediment resuspension may then have harmful effects for organisms living in the water column (Gewurtz et al., 2007; Varanasi et al., 1985; Woodhead et al., 1999).

The partitioning of PAHs between water and sediment evaluated through the determination of the sediment-water or sediment OM-water partition coefficients ( $K_d$  or  $K_{oc}$ , respectively) is largely driven by grain size and OM content of sediments and may vary during resuspension

events (Cornelissen et al., 2006; Karickhoff et al., 1979; Rockne et al., 2002; Tremblay et al., 2005). However, little is known about the association between PAHs and sedimentary OM subjected to transformations during burial and diagenesis (Berner, 1980; Henrichs, 1993); that is, the change in  $K_d$  and  $K_{oc}$  values with sediment depth or OM aging. Further, studies dealing with PAH partitioning between water and sediment should better consider the interactions between dissolved PAHs and dissolved OM (DOM) (Akkanen et al., 2005; McCarthy and Jimenez, 1985; Yang et al., 2016). Over the last decade, the dynamics of DOM in coastal waters has been evaluated through the study of its fluorescent fraction (FDOM). Recently, FDOM associated with marine or river sediments has been characterized, i.e., the FDOM from sediment pore waters (Burdige et al., 2004; Dang et al., 2014; Sierra et al., 2001) and from sediment particles (Brym et al., 2014; Dang et al., 2014; He et al., 2016), as well as FDOM released by coastal sediment resuspension (Komada et al., 2002). Hence, due to the influence of OM in PAH partitioning processes, it is generally observed that theoretical  $K_{oc}$ , i.e.,  $K_{oc}$  predicted from their octanol-water partition coefficients ( $K_{ow}$ ), are lower than measured  $K_{oc}$  (Accardi-Dey and Gschwend, 2002; Feng et al., 2007; Fernandes et al., 1997). As a corollary, concentrations in water that are determined from sediment concentrations and theoretical  $K_{oc}$  are frequently overestimated. This also leads to an over-estimation of uptake processes and/or effects of those chemicals on exposed organisms (Arp et al., 2009; Hawthorne et al., 2007). Measuring  $K_{oc}$  is thus essential for more accurate predictions of PAH toxicity in the water column.

In the Mediterranean Sea, Toulon harbor (Southern France, Northwestern Mediterranean Sea) hosts the main French Navy structure. It is located deep inside Toulon Bay and enclosed by a sea wall. Because of this separation, and also the irregular freshwater inputs and the low tide, water circulation in this part of the Bay is limited, leading to low water regeneration and potentially strong accumulation of chemical contaminants such as PAHs in the sediments

(Benlahcen et al., 1997; Misson et al., 2016; Wafo et al., 2016). To maintain a navigable water depth, harbor dredging is regularly required in accordance with the current legislation. Such dredging may induce a sediment resuspension over 100 cm sedimentary depths, thus allowing surface and deep sediments to be resuspended. Since surface and deep sediments have different OM contents (from potentially different origins along with diagenetic transformations with time/depth), their resuspension could generate various remobilization kinetics of pollutants in seawater. Several studies have reported high concentrations of trace metals, metalloids and organometallics during Toulon Bay seabed disturbances (Cossa et al., 2014; Dang et al., 2015a, Pougnet et al., 2014; Tessier et al., 2011). In addition, Dang et al. (2015b) demonstrated that Pb was significantly released in seawater and further transferred to organisms such as mussels during resuspension experiments on surface and deep sediments from Toulon Bay. Nevertheless, to our knowledge, no data are available describing PAH remobilization during sediment resuspension events in such a strongly dredged and multi-contaminated area.

Therefore, in the present study, we carried out supplementary experiments simulating the resuspension of surface and deep sediments in Toulon Bay, as proposed by Dang et al. (2015b). Our objectives were (i) to assess the contamination level and origin of PAHs in a sediment core from Toulon Bay, (ii) to evaluate the effect of surface and deep sediment resuspension on the remobilization of PAH and OM, as well as on the water quality, and (iii) to compare theoretical and measured  $K_{oc}$  to better understand the factors controlling PAH remobilization during sediment resuspension experiments (SRE). To our knowledge, this study represents the first assessment of remobilization kinetics of both dissolved PAHs and DOM during SRE involving surface and deep sediments.

## **2. Materials and Methods**

## **2.1 Study area**

Located on the French Northwestern Mediterranean coast, Toulon city is a part of a large urban area of approximately  $500 \times 10^3$  inhabitants. Toulon Bay is divided into two unequal basins, a small basin (9.8 km<sup>2</sup>, semi-enclosed) submitted to various anthropogenic inputs (the French Navy, commercial traffic, raw sewage from the urban area, industries) and a large basin (42.2 km<sup>2</sup>) that is less impacted and open to the sea (Fig. 1). Toulon harbor is situated in the small bay, which is the discharge area for the watershed. Low tides in the Mediterranean Sea associated with weak currents in this area of Toulon Bay have significant implications for the accumulation of contaminants in sediments (Dufresne et al., 2014; Tessier et al., 2011).

## **2.2. Sampling and sample treatment**

Sediments and seawater were collected at the Missiessy (MIS) site within the nuclear submarine harbor of the French Navy, on May 5<sup>th</sup> 2014, with the support of the French Navy (ship, material, divers) (Fig. 1).

### **2.2.1. Sediment core**

A sediment core of ca. 50 cm was collected through the use of an interface corer (Plexiglas tube, 10 cm diameter and 1 m long), by Navy divers keeping undisturbed the bottom water column and the upper sediment column, and so preserving the water-sediment interface as described in Dang et al. (2014, 2015a), Tessier et al. (2011). The collected sediment core was maintained vertically and was carefully transferred (by boat and then van) to the laboratory before being installed on a home-made slicing table under glove box. The core was sliced with a 2-cm resolution under inert atmosphere (N<sub>2</sub>) to preserve oxidation-reduction (redox) conditions.

Each slice was then homogenized in a 150 mL high-density-polyethylene bottle and split into 3 × 50 mL polypropylene centrifuge tubes under N<sub>2</sub> atmosphere. Then, porewater was extracted by centrifugation (15 min, 20 °C, 400 rpm, Sigma 3-18 K), recovered under N<sub>2</sub> atmosphere by filtration (0.2-µm on-line syringe filters, cellulose nitrate, Sartorius) and stored in required vessels depending on further chemical analyses. Such methodology was successfully applied to study depth profile variation of main diagenesis tracers and OM characteristics (Chen and Hur, 2015). In accordance with previous studies on trace element sedimentary dynamics in the same area (Dang et al., 2015b), two slice samples were selected from this sediment core to perform SRE: the 0-2 cm sediment layer (denoted hereafter “0-2 cm sediment”) and the 30-32 cm sediment layer (denoted hereafter “30-32 cm sediment”) (see § 2.3). The PAH concentrations were determined in each layer of the core (freeze-drying and 2 mm sieving) to assess the contamination level and potential toxicity of PAHs at this specific site. In addition to PAHs, particle grain size, organic carbon and nitrogen (OC and ON) contents, and aliphatic hydrocarbons (AHs) were determined in the 0-2 and 30-32 cm sediment layers dedicated to SRE. Additionally, porewater measurements of pH, redox potential (Eh), dissolved organic carbon (DOC), dissolved inorganic carbon (DIC), total nitrogen (N<sub>T</sub>), ammonium (NH<sub>4</sub><sup>+</sup>), soluble reactive phosphorus (SRP), silicate (Si(OH)<sub>4</sub>), manganese (Mn<sub>T</sub>), iron (Fe<sub>T</sub>), sulfate (SO<sub>4</sub><sup>2-</sup>) and total sulfide (ΣHS<sup>-</sup>) were taken to verify the redox status of these two sediments according to previously described procedures (Dang et al., 2014, 2015a, 2015b).

### ***2.2.2. Seawater***

Seawater was collected to characterize the water column in the small basin of the bay in terms of PAH and DOM concentrations and to conduct SRE. Samples were taken in duplicates at 0.5, 2.5, 5 and 13 m (overlying water ~1 m above the sediments) depths using a 2.2-L Wildco

Van-Dorn horizontal water sampler. Before use, the latter was rinsed with  $3 \times 1$  L of ultrapure (Milli-Q) water acidified to 0.2% HCl (TraceSelect, Fluka) and then several times with sampling water. The water samples were poured into polycarbonate bottles (Nalgene). Before use, the bottles were washed with 1 M HCl followed by a wash with ultrapure water and three washes with the respective sample.

At the laboratory, the water samples were immediately filtered under a low vacuum ( $< 50$  mm Hg) through precombusted ( $450$  °C, 6 h) glass fiber filters (GF/F,  $\sim 0.7$   $\mu\text{m}$ , 47 mm diameter, Whatman) using all-glassware systems for dissolved PAHs, DOC and FDOM. The filtered samples for dissolved PAH analyses were stored in 2-L SCHOTT glass bottles with 50 mL dichloromethane ( $\text{CH}_2\text{Cl}_2$ ) at  $4$  °C in the dark before solvent extraction (within 24 h) while the samples for DOC and FDOM were stored frozen until analysis.

### **2.3. Sediment resuspension experiments**

The solid/liquid (S/L) ratio was set at  $\sim 1$   $\text{g L}^{-1}$  (expressed in dry weight), a value close to *in situ* levels of suspended particulate matter measured during surface sediment reworking as previously published (Dang et al., 2015b). To apply this ratio for each experiment,  $\sim 11$  g of wet sediment from the 0-2 cm slice (62% moisture) and  $\sim 8$  g of wet sediment from the 30-32 cm slice (52% moisture) were introduced into 4 L Nalgene bottles filled with 0.7- $\mu\text{m}$  filtered surface seawater (collected at 0.5 m depth). The bottles were then placed on roller agitation devices (Wheaton, 8 rpm) at room temperature ( $21$ - $23$  °C). Experiments lasted 14 days and were run in duplicate (i.e., four 4 L bottles in total). The water subsamples were collected at 1, 3, 7 h and 1, 3, 7, 10, 14 days and were analyzed for dissolved PAHs, DOC and FDOM. For each subsample, agitation was stopped for approx. 30 min allowing most of the sediment particles to settle. Then, 400 mL of water was collected from the top of the bottle and immediately filtered for analysis.

To minimize S/L ratio variation, the same volume (400 mL) of filtered initial seawater was added to the bottle after each subsampling. Because subsamples were taken several minutes after stopping agitation, the loss of sediment particles from this successive subsampling was estimated to be negligible over the course of the SRE (Dang et al., 2015b). Also, according to Dong et al. (2016), such short stops cannot be considered as successive sediment resuspension-deposition events. It should be noted that the biological activity was not stopped in these experiments since (i) any added poison could have affected OM measurements and (ii) our aim was to mimic the PAH and OM sorption/desorption as close as possible to the natural conditions.

## **2.4. Hydrocarbon analysis**

### ***2.4.1. Hydrocarbon extraction and purification***

PAHs were measured in each layer of the whole sediment core and in seawater samples (water column profile and SRE), while AHs were measured only in the 0-2 and 30-32 cm sediments used for SRE. Hydrocarbons (PAHs and AHs) were extracted from sediment samples in 5 mL cells with CH<sub>2</sub>Cl<sub>2</sub> and copper powder to remove elemental sulfur, using pressurized liquid extraction (trade name ASE for accelerated solvent extraction) with a Dionex ASE 350 system (150 °C, 110 bars, 3 cycles of 10 min, 100% of rinsing, purging for 60 s; ThermoElectron). Dissolved PAHs present in the fraction < 0.7 μm (water column profile and SRE) were extracted from water by liquid-liquid extraction with CH<sub>2</sub>Cl<sub>2</sub> (2 × 80 mL per liter). The protocol was then the same for sediment and water extracts. The solvent volume was reduced by rotary evaporation and the solvent was changed to *n*-hexane prior to purification. The hexane-solubilized extracts were purified to separate hydrocarbons from more polar compounds. These extracts were fractionated on a 500-mg silica column. Silica gel (extra pure, Merck) was

activated at 450 °C for 6 h followed by partial deactivation with 4% water by weight. The extracts were deposited using 2-mL *n*-hexane and hydrocarbons were eluted with 3-mL *n*-hexane/CH<sub>2</sub>Cl<sub>2</sub> (3:1 v/v) (Guigue et al., 2011, 2014, 2015). All solvents were of organic trace-analysis quality (Rathburn Chemicals Ltd.).

#### ***2.4.2. Analysis of hydrocarbons***

The purified extracts were analyzed by gas chromatography-mass spectrometry (GC-MS) (TraceISQ, ThermoElectron) operating at an ionization energy of 70 eV for a m/z range of 50-400 (full scan and selected ion monitoring (SIM) modes processed simultaneously), using helium as carrier gas at a flow rate of 1.2 mL min<sup>-1</sup>. The GC-MS was equipped with a HP-5 MS ultra-inert column (30 m × 0.25 mm × 0.25 μm, J&W Scientific, Agilent Technologies). The injector (used in splitless mode) and detector temperatures were 270 and 320 °C, respectively. The initial column temperature was held for 3 min at 70 °C, then ramped at 15 °C min<sup>-1</sup> (ramp 1) to 150 °C and then at 7 °C min<sup>-1</sup> (ramp 2) to a final temperature of 320 °C, which was held for 60 min. AHs and PAHs were identified and quantified in scan and SIM modes simultaneously using two distinct methods (Guigue et al., 2011, 2014).

#### ***2.4.3. Quality assurance and quality control***

All glassware was washed with 1 M HCl and ultrapure water and combusted at 450 °C for 6 h. All the materials that could not be baked were washed with 1M HCl and ultrapure water and dried at room temperature.

Standard mixtures (04071, Fluka and 47543-U, Supelco among others) and natural samples (for alkylated homologue PAHs) were used to validate the identification methods. Further, deuterated standard mixtures (C<sub>16</sub>-d<sub>34</sub> and Naph-d<sub>8</sub> for lower MW compounds, i.e., <*n*-C<sub>20</sub> alkanes

and 2-ring PAHs,  $C_{24}$ - $d_{50}$  and Phe- $d_{10}$  for medium MW compounds, i.e.,  $n$ - $C_{20}$ - $n$ - $C_{29}$  alkanes and 3-4 ring PAHs,  $C_{36}$ - $d_{74}$  and Per- $d_{12}$  for higher MW compounds, i.e.,  $>n$ - $C_{30}$  alkanes and 5-6 ring PAHs) were introduced as surrogates before extraction, as well as a supplementary deuterated standard (Chr- $d_{12}$ ) before injection, to assess the procedural recoveries and to refine the quantification of hydrocarbons in the samples. The recoveries of Naph- $d_8$ , Phe- $d_{10}$  and Per- $d_{12}$  were on average 44, 61 and 81% in water and 54, 73 and 93% in sediment samples, respectively. Caution was taken during the evaporation because dryness could lead to the complete loss of the more volatile compounds. In addition, blanks were run for the whole procedure including the use of the Nalgene polycarbonate bottles, extraction, solvent concentration and purification. All concentration values were blank- and recovery-corrected (procedure fully described in Guigue et al., 2015). Instrumental detection limits for individual compounds varied from 1 to 30 pg of injected chemical. No certified reference standard is currently available for PAHs in water but the protocol for sediment was validated based on the agreement of individual compound quantification compared to NIST, 1941b (organics in sediment) (agreement of  $97 \pm 10\%$ ).

#### ***2.4.4. Determination of individual hydrocarbons***

For PAHs, we determined the concentrations of 19 parent PAHs and alkylated homologues of the target compounds Naph, Flu, Phe/Ant, Flt/Pyr and Chr, leading to a total of 34 PAHs (see full names of PAHs in Text S1). Naph and its alkylated homologues are 2-ring compounds. Acy, Ace, Flu, DBT, Phe, Ant and alkylated homologues of Flu, Phe and Ant are 3-ring compounds. Flt, Pyr, BaA, Chr and alkylated homologues of Flt, Pyr and Chr are 4-ring compounds. BbF, BkF, BaP, Per and DBA are 5-ring, while BP and IndP are 6-ring, compounds. Concerning AHs, we determined the concentrations of the resolved  $n$ -alkane series ( $R$ ) from  $n$ - $C_{15}$  to  $n$ - $C_{36}$  with two isoprenoids, pristane (Pr) and phytane (Phy), as well as the unresolved complex mixture

(UCM) concentrations. All ratios and indices used to determine PAH and AH potential origins are given in the supplementary material (see Text S1).

## **2.5. Sediment grain size and contents in total organic carbon and nitrogen**

Grain size and total OC and ON contents were determined for the 0-2 and 30-32 cm sediment layers. Grain size was determined with a Beckman Coulter LS 13 320 laser granulometer after OM removal (Ghilardi et al., 2012). The relative abundance of sand (2000 to 63  $\mu\text{m}$ ), silt (63 to 2  $\mu\text{m}$ ) and clay (< 2  $\mu\text{m}$ ) was then measured. Contents in OC and ON were determined simultaneously, after acidification, with an AutoAnalyser II Technicon using the wet oxidation procedure (Raimbault et al., 1999). The contents are expressed as a percentage (%) of sed. dw.

## **2.6. Dissolved organic matter measurements**

DOC was determined by high-temperature catalytic oxidation using a Shimadzu TOC 5000 Total Carbon Analyzer (Kyoto, Japan) according to Sohrin and Sempéré (2005). Two replicates were analyzed for each sample. The concentrations are the mean of the two replicates with a coefficient of variance (CV) < 2%.

FDOM analyses were performed with a Hitachi F-7000 spectrofluorometer (Tokyo, Japan). The method is fully described in Tedetti et al. (2012, 2016) and Ferretto et al. (2014). Briefly, excitation-emission matrices (EEMs) were generated for excitation wavelengths ( $\lambda_{\text{Ex}}$ ) between 230 and 500 nm and for emission wavelengths ( $\lambda_{\text{Em}}$ ) between 280 and 550 nm. Two replicates were run for each sample. To correct EEMs for potential inner filtering effects, dilution method was used. Fluorescence intensities were blank-corrected and converted to quinine sulfate units (QSU), where 1 QSU corresponds to the fluorescence of 1  $\mu\text{g L}^{-1}$  quinine sulfate in 0.05 M

sulfuric acid at  $\lambda_{\text{Ex}}/\lambda_{\text{Em}}$  of 350/450 nm (5nm slit widths). The fluorescence intensities in QSU provided for each sample are the mean of the two replicates with a CV < 8%. The data were processed using parallel factor analysis (PARAFAC) operated using the DOMFluor toolbox v1.6. running under MATLAB 7.10.0 (R2010a). The PARAFAC model was created and validated for 81 samples according to the method of Stedmon and Bro (2008).

## 2.7. Determination of sediment-water partition coefficients of PAHs

The partition coefficients of PAHs between sediment and water ( $K_d$ ) were determined using the following equation:

$$K_d = C_p / C_d$$

where  $C_p$  is the concentration of individual PAHs in sediment ( $\text{ng kg}^{-1}$ ) and  $C_d$  is the concentration in the water-dissolved phase ( $\text{ng L}^{-1}$ ). The OC-normalized partition coefficients of PAHs between sediment and water ( $K_{oc}$ ) were then calculated using the following equation:

$$K_{oc} = K_d / f_{oc}$$

where  $f_{oc}$  is the fraction of organic carbon in resuspended sediment particles (% OC in Table 2). The values for  $\log K_d$  and  $\log K_{oc}$  were computed for the 16 priority PAHs.

## 2.8. Statistics

Student's *t*-test, used to compare the mean of two independent data groups, and Pearson's linear correlation matrices, as well as non-linear regression analyses were carried out using XLSTAT 2013.5.01 (Microsoft Excel add-in program). For the different analyses and tests, the significance threshold was set at  $p < 0.05$ .

### 3. Results and discussion

#### 3.1. Characterization of sediment and water from the MIS site

##### 3.1.1 Contamination levels and origin of PAHs in the sediment core

In the first 40 cm of the sediment core,  $\Sigma 34$  PAH concentrations varied between  $30.7 \times 10^3$  at the 10-12 cm depth and  $123 \times 10^3$  ng g<sup>-1</sup> sed. dw at the 36-38 cm depth, with a high value of  $231 \times 10^3$  ng g<sup>-1</sup> sed. dw recorded at the 24-26 cm depth (Fig. S1). The PAH concentrations in the surface layer of the MIS sediment core were in agreement with previous recordings from the MIS site ( $34.0 \times 10^3$  ng g<sup>-1</sup>; Misson et al., 2016) and in the Toulon coastal area ( $48.1 \times 10^3$  ng g<sup>-1</sup>; Benlahcen et al., 1997). The PAH concentrations we recorded in the whole core were in the upper-range of values reported for sediments from other regions of the Mediterranean Sea (see Table 6 in Zaghden et al., 2017). For instance, they were close to the highest values measured in the surface sediments from the Taranto Gulf, Italy ( $28.9$ - $262 \times 10^3$  ng g<sup>-1</sup> sed. dw; Annicchiarico et al., 2011). The environmental quality guidelines are detailed in the supplementary material (Text S2). The 16 priority PAHs displayed concentrations above L1 and ERL levels at almost all sediment depths and above L2 and ERM levels at many depths (Table 1), which mirrors high to extreme pollution levels according to both JORF and SQG, in agreement with previous results for trace elements from the small basin of Toulon Bay (Tessier et al., 2011).

The profile shape of  $\Sigma 34$  PAH concentrations with depth at MIS was in good agreement with previous studies showing disturbed sedimentation during the past (raising of a scuttled warship) and the present (harbor extension) activities (Fig. S1; Dang et al., 2015b; Misson et al., 2016). Despite episodic events disturbing sedimentation, the surface layers are considered to be more recent than deeper ones (Dang et al., 2015b; Tessier et al., 2011). The surprising value of  $231 \times$

$10^3 \text{ ng g}^{-1}$  recorded at 24-26 cm, which had a similar molecular distribution to the other layers (data not shown), might not be attributed to a special historic period/contamination but more probably to the heterogeneity in the vertical distribution of sediment particles or to the presence of a tiny piece of coal block.

The PAH molecular distribution in the sediment core barely varied with depth and was dominated by 4-ring compounds, as shown for the 0-2 and 30-32 cm sediments in Fig. 2. This distribution reflected the dominance of Flt and Pyr, which has been observed in other coastal areas of the Mediterranean Sea (Ben Othman et al., 2017; Pérez-Fernández et al., 2015). The isomeric ratios were quite close from one depth to another, and showed that PAHs in the MIS sediment more probably originated from combustion processes, especially of grass/wood/coal, rather than from burned/unburned petroleum residues (Fig. S2). The dominance of pyrogenic PAHs in this sediment is in agreement with our knowledge of current and past activities at Toulon harbor (loading dock for coal and coal-fired Navy vessels) and also reflects a highly anthropized coastal environment subjected to important watershed and atmospheric particulate inputs (Fig. S2; Vane et al., 2014; Yunker et al., 2002). Nevertheless, it is worth recalling that isomeric ratios have to be used with caution for the determination of PAH origin because they may evolve with time or with the distance from emission sources (Katsoyiannis and Breivik, 2014).

### ***3.1.2. Comparison of the 0-2 and 30-32 cm sediments***

The 0-2 and 30-32 cm sediments used for SRE were exclusively composed of fine particles <  $63 \mu\text{m}$ : 69 and 82% of silt, 31 and 18% of clay, respectively. Their OC content was 8.2 and 6.3%, respectively (Table 2). The predominance of silts, which promote the accumulation of contaminants, and the OC content in these sediments are in good agreement with previous studies

from Toulon Bay (Dang et al., 2014; Tessier et al., 2011). Additionally, the OC content was in the upper range of that reported for surface sediments of the Northern Mediterranean Sea (from 0.38 to 6.2%) (Benlahcen et al., 1997; Lipiatou and Saliot, 1991).

The C/N ratios in the 0-2 and 30-32 cm sediments (13.5 and 22, respectively) (Table 2) were higher than typical values reported for most coastal sediments (C/N = 6-10; Wang et al., 2001) but lower than that of the alkaline-extracted OM previously recorded in this area (C/N =  $36 \pm 9$ ; Dang et al., 2014). The C/N ratio may provide information about the OM origin, as well as the diagenetic degradation state. A high C/N ratio ( $> 20$ ) reveals a terrestrial origin of OM due to the low N percentage in the higher vegetation (Emerson and Hedges, 1988; Muller, 1977). On the other hand, a low C/N ratio (5-7) implies a marine origin (plankton or seaweeds) (Meyers, 1994). In addition, it has previously been demonstrated that the buried particulate OM in Toulon Bay (at another location than MIS) is N-, P- and Si-depleted, due to the sedimentary OM mineralization and transformation processes (Dang et al., 2014). This depletion could result in increases in the C/N ratio. In the present case, the C/N ratios suggest a mixed origin, with both terrestrial and autochthonous marine sources at both depths along with higher diagenetic processes at the 30-32 cm depth (N-depletion; Table 2) according to Dang et al. (2014).

The porewaters of the two studied sediment layers were slightly acidic and enriched in DOC, DIC,  $\text{NH}_4^+$ , SRP,  $\text{Si}(\text{OH})_4$  and  $\text{Fe}_T$  compared to seawater, showing important diagenesis processes (Table S1), and differed in their redox status. The 0-2 cm sediment porewaters had positive Eh (109 mV/ENH) and high  $\text{Fe}_T$  concentrations (70.2  $\mu\text{M}$ ) typical of oxidizing conditions, while the 30-32 cm sediment porewaters displayed negative Eh (-145 mV/ENH) and much lower  $\text{Fe}_T$  concentrations (7.7  $\mu\text{M}$ ), reflecting reducing conditions. Additionally, they had differences in DOC (367 and 717  $\mu\text{M}$ , respectively),  $\text{N}_T$  (104 and 173  $\mu\text{M}$ ),  $\text{NH}_4^+$  (72.6 and 54.8  $\mu\text{M}$ , respectively) and  $\text{Mn}_T$  (2.3 and 0.7 nM, respectively) concentrations (Table S1).

The  $\Sigma 34$  PAH concentrations in these samples were  $38.2$  and  $35.7 \times 10^3$  ng g<sup>-1</sup> sed. dw, respectively (Fig. S1; Table 2) and all 16 priority compounds showed concentrations above L1 and ERL levels. Additionally, Ace and BaA in the 0-2 cm sediment and Phe, Pyr, BaP and DBA in the 0-2 and 30-32 cm sediments had concentrations above the L2 and ERM levels (Table 1). For both the sediment layers, as in the whole core, 2-ring compounds accounted on average for 8% of total PAHs, 3-rings for 17%, 4-rings for 41% (with a high contribution of Flt and Pyr), 5-rings for 22% and 6-rings for 11% (Fig. 2).

The  $R$  concentrations ( $n$ -C<sub>15</sub> to  $n$ -C<sub>36</sub> with Pr and Phy) were  $2.4$  and  $10.0 \times 10^3$  ng g<sup>-1</sup> sed. dw with UCM/ $R$  values of 30 and 118, respectively (Table 2). The  $R$  values were relatively low compared to the PAH content of these sediments and were situated in the middle-lower range of  $n$ -alkane concentrations reported from other Mediterranean environments (see Table 4 in Zaghdien et al., 2017). For example, they were of the same order of magnitude as those determined in the Gulf of Tunis ( $1.8$  and  $10.0 \times 10^3$  ng g<sup>-1</sup> sed. dw; Mzoughi and Chouba, 2011). The  $R$  and UCM/ $R$  values were four times higher at the 30-32 cm than at the 0-2 cm depth. The  $R$  molecular profiles were characterized by bi-modal distributions. Odd-carbon numbered dominated the  $n$ -alkane patterns:  $n$ -C<sub>17</sub> for the LMW compound profiles ( $\leq n$ -C<sub>24</sub>) and  $n$ -C<sub>29</sub>,  $n$ -C<sub>31</sub> and  $n$ -C<sub>33</sub> for the HMW compound profiles ( $>n$ -C<sub>24</sub>) (Fig. S3). This led to CPI values slightly above 1. CPI<sub>15-24</sub> was 1.3 in both sediments, showing that autochthonous algal material and anthropogenic  $n$ -alkanes accounted for 13 and 87% of total  $n$ -alkanes, respectively. CPI<sub>25-36</sub> was 2.9 and 2.2 in the 0-2 and 30-32 cm sediments, respectively. This underscores that continental higher plant debris  $n$ -alkanes accounted for 49 and 37%, and anthropogenic  $n$ -alkanes for 51 and 63% of total  $n$ -alkanes, respectively (Table 2). In both sediments, these results ( $R$ , UCM/ $R$ , CPI values) reflected the superposition of compounds originating from biogenic and anthropogenic inputs, as well as their incomplete degradation (Blumer et al., 1971; Bouloubassi and Saliot,

1993; Douglas and Eglinton, 1966). However, compared to the 0-2 cm sediment, the 30-32 cm sediment presented a higher anthropogenic fingerprint along with a higher diagenetic state. This greater anthropization level was very likely due to the Second World War and activities during and after this period (Tessier et al., 2011). The higher contribution of Phy at this depth might also be attributed to the diagenetic state of sediments, even though diagnostics using isoprenoid compounds must be taken with caution (Grossi et al., 1998; Rontani and Bonin, 2011).

In conclusion, C/N ratios, diagenetic tracers in porewaters and AHs showed that the 0-2 cm sediment was globally less contaminated, presenting more biogenic inputs, especially terrigenous, and was more recent/less degraded or diagenetic transformed than the 30-32 cm sediment. This finding confirmed the differences already reported in previous studies combining redox potential and particulate OM aging (Dang et al., 2014, 2015b). The OM aging of sediment particles might play a role in PAH-OM interactions and therefore on the PAH mobility during seabed disturbances, as suspected for other organic contaminants (Calvo et al., 1991). In contrast, PAH distribution did not make it possible to highlight such differences between the two sediments.

### ***3.1.3. Characterization of the water column***

The concentrations of dissolved  $\Sigma 34$  PAHs in the water column were  $2.8 \pm 0.7$ ,  $2.1 \pm 0.5$ ,  $3.8 \pm 1.5$  and  $4.6 \pm 0.9$  ng L<sup>-1</sup> at 0.5, 2.5, 5 and 13 m depths, respectively (Fig. S4a). Dissolved PAHs were dominated by 2- and 3-ring compounds ( $38 \pm 9$  and  $49 \pm 10\%$ , respectively), whereas 4- and 5-rings represented only  $10 \pm 2$  and  $3 \pm 7\%$ , respectively, with 6-ring compounds being under the detection limits (Fig. 2). These concentrations are in the lower range of those previously observed in some Northwestern Mediterranean coastal areas, whereas the molecular distribution (dominance of 2-3 ring compounds) was in agreement with these studies (Guigue et al., 2011, 2014; Guitart et al., 2004).

DOC concentrations were  $79.4 \pm 1.3$ ,  $75.6 \pm 1.0$ ,  $74.8 \pm 1.0$  and  $76.0 \pm 0.2$   $\mu\text{M}$  at 0.5, 2.5, 5 and 13 m depths, respectively (Fig. S4b). These concentrations were in the same range as those previously measured in this area (Dang et al., 2014). PARAFAC applied to EEMs revealed the presence of two main FDOM fluorophores in seawater (for both depth profiles and SRE). One fluorophore displayed two fluorescence maxima at  $\lambda_{\text{Ex1}}$ ,  $\lambda_{\text{Ex2}}/\lambda_{\text{Em}}$  of 230, 340/452 nm. This humic-like fluorophore corresponds to peaks A + C in Coble's (1996) classification and to component 2 in Ishii and Boyer's (2012) classification. The other, with two fluorescence maxima located at  $\lambda_{\text{Ex1}}$ ,  $\lambda_{\text{Ex2}}/\lambda_{\text{Em}}$  of 230, 280/348 nm, corresponds to a tryptophan-like fluorophore, i.e., peaks T1 + T2 according to Coble (1996) and Hudson et al. (2007) (contour plots of PARAFAC components not shown). It should be noticed that this fluorophore could also indicate the presence of PAHs and specifically naphthalene, whose fluorescence spectral domain is superimposed on that of tryptophan (Tedetti et al., 2010; Ferretto et al., 2014). However, the sum of Naph and alkylated Naph homologue concentrations measured here ( $< 5$   $\text{ng L}^{-1}$ ) in water were not high enough to generate such fluorescence signatures (detection limit of Naph by EEM method;  $\sim 1$   $\mu\text{g L}^{-1}$ ; Ferretto et al., 2014). The fluorescence intensities of humic- and tryptophan-like fluorophores at 0.5, 2.5, 5 and 13 m depths were  $2.7 \pm 1.2$ ,  $3.5 \pm 0.9$ ,  $2.9 \pm 0.3$  and  $4.0 \pm 0.3$  QSU (Fig. S4c) and  $4.8 \pm 2.4$ ,  $6.2 \pm 1.4$ ,  $7.4 \pm 0.7$  and  $8.4 \pm 1.6$  QSU, respectively (Fig. S4d).

These two fluorophores have been reported in several coastal environments, including the Northwestern Mediterranean Sea (Ferretto et al., 2017; Para et al., 2010; Tedetti et al., 2012). The tryptophan-like fluorophore represents free amino acids or amino acids bound as peptides or proteins. It is known to serve as a fresh and labile bioavailable product for heterotrophic bacteria (Romera-Castillo et al., 2010; Yamashita and Tanoue, 2004). The humic-like fluorophore has maximal absorption in the UVC and UVA wavelengths. It is more hydrophobic, more condensed and more photodegradable compared to the tryptophan material, as indicated by its higher  $\lambda_{\text{Ex}}$  and

$\lambda_{Em}$ . Dang et al. (2014) and Hur et al. (2014) detected these two fluorophores (humic and tryptophan) in sediment particles after an alkaline extraction, showing that they could also originate from sediment, in addition to the water column. According to Dang et al. (2014), the tryptophan-like fluorophore would be derived from fresh POC in surface sediments, while humic-like fluorophore would be either produced by the hydrolysis of the buried HMW-POM or by the geo-polymerization of LMW-DOM. Overall, these data show that water from 0.5 m depth was not subjected to an important recent remobilization and was well adapted to forthcoming experiments.

### **3.2. Remobilization of PAHs and OM during sediment resuspension experiments (SRE)**

#### ***3.2.1. Dissolved phase enrichment***

Concentrations of dissolved  $\Sigma 34$  PAHs, DOC and FDOM fluorophores in water over the course of the 0-2 and 30-32 cm SRE are presented in Fig. 3. The standard deviation (to estimate the variance) is given for the two-bottle replicates ( $n = 2$ ). For the four parameters, both SRE led first to an immediate (within one hour after the sediment addition) release in water. This fast PAH release in the dissolved phase (within the first hour) was also observed by Yang et al. (2008). Then, from the first hour to the end (14 days) of the SRE, concentrations in water tended to increase: from  $8.6 \pm 1.6$  to  $24 \pm 9.9$  ng L<sup>-1</sup> (Fig. 3a) and from  $9.5 \pm 3.9$  to  $12 \pm 5.4$  ng L<sup>-1</sup> (Fig. 3b) for dissolved  $\Sigma 34$  PAHs, from  $87 \pm 0.2$  to  $91 \pm 0.3$   $\mu$ M (Fig. 3c) and from  $89 \pm 1.4$  to  $98 \pm 2.9$   $\mu$ M (Fig. 3d) for DOC, from  $5.4 \pm 0.1$  to  $11 \pm 1.8$  QSU (Fig. 3e) and from  $4.8 \pm 0.9$  to  $11 \pm 0.2$  QSU (Fig. 3f) for the humic-like fluorophore, and from  $9.1 \pm 0.3$  to  $28 \pm 3.2$  QSU (Fig. 3g) and from  $8.0 \pm 1.4$  to  $15 \pm 0.9$  QSU (Fig. 3h) for the tryptophan-like fluorophore. These results seem to be in agreement with desorption biphasic models of organic compounds from

soils/sediments, i.e., a rapid desorption phase of PAHs adsorbed onto particles followed by a more slowly desorbing phase of PAHs strongly bound into particles (Karickhoff and Morris, 1985; Mitra and Dickhut, 1999; Van Noort, et al., 2003).

Enrichment factors (EFs, computed as the ratios of the concentration in water at time  $t$  over that at the initial time; that is, just before sediment addition) for dissolved PAHs, DOC and the humic-like fluorophore were quite similar between the 0-2 and 30-32 cm SRE, ranging from 2.6-8.5 and 2.5-10 (PAHs), 1.1-1.2 and 1.1-1.3 (DOC), and 2.0-4.2 and 1.8-4.4 (humic), respectively (Fig. 3a-f). For the tryptophan-like fluorophore, they were clearly higher in the 0-2 cm SRE (1.7-5.7) than in the 30-32 cm SRE (1.7-3.2) (Fig. 3g,h). EFs recorded here for PAHs were of the same order of magnitude as those reported in previous studies under natural conditions (Dong et al., 2015).

### ***3.2.2. Change in dissolved PAH distribution profile***

During SRE, all the individual PAHs investigated were, to some extent, released into the dissolved phase. For both SRE, dissolved PAH distribution profiles were dominated by 3-ring compounds ( $33 \pm 8$  and  $38 \pm 8\%$  of total PAHs) (Fig. 2). Moreover, in the range of 2-4-ring compounds, SRE PAH profiles were clearly intermediate between (and significantly different from) that of the water column (0.5-13 m depths; dominated by 3-ring compounds, accounting for  $49 \pm 10\%$  of total PAHs) and those of sediments (dominated by 4-ring compounds, accounting for 39 and 43% of total PAHs) (Fig. 2). However, for 5- and 6-ring compounds, the relative abundances were much closer between SRE (16-19 and 6-7%, respectively) and sediments (21-24 and 11-12%, respectively). Hence, compared to the water column, the remobilization of PAHs led to a shift in the molecular structure profiles towards HMW compounds, with a decrease in the relative abundances of 2- and 3-ring compounds (by 49 and

29% on average, respectively), an increase in the relative abundances of 4- and 5-ring compounds (by 115 and 414% on average, respectively) and the appearance of 6-ring compounds (Fig. 2).

### ***3.2.3. Comparison of the remobilization kinetics between surface and deep sediments***

Remobilization kinetics were studied by following dissolved PAH and DOM concentration evolution, as well as correlations between these parameters within each SRE. First, to help in comparing the behavior of parameters between each other and between the two experiments, we sought to model the kinetic patterns. Several models obtained by least square fits were tested (linear, exponential, power, logarithmic, polynomial of degrees 2-3). The polynomial function of degree 3 (cubic functions) best fitted our data. All these cubic relationships were significant ( $r = 0.78-1.00$ ,  $p \leq 0.0001-0.02$ ,  $n = 8$ ), except for DOC in the 30-32 cm SRE ( $r = 0.72$ ,  $p = 0.06$ ,  $n = 7$ ; Fig. 3d). Nevertheless, we observed some differences in the shape of these cubic functions between the 0-2 cm and the 30-32 cm SRE.

In the 0-2 cm SRE, DOC and the tryptophan-like fluorophore presented a cubic curve with a negative leading coefficient ( $-0.045$  and  $-0.005$ , respectively; Fig. 3c,g). Dissolved PAHs and humic-like fluorophores presented a different pattern due to the positive leading coefficient of their equation ( $0.031$  and  $0.002$ , respectively; Fig. 3a,e). In addition, maximal values for PAHs and the humic- and tryptophan-like fluorophores were recorded at the end of the experiment ( $t = 14$  days) of  $24 \pm 9.9 \text{ ng L}^{-1}$ ,  $11 \pm 1.8$  and  $28 \pm 3.2 \text{ QSU}$ , respectively (Fig. 3a,e,g), whereas that of DOC was found at  $t = 10$  days ( $99 \pm 2.9 \text{ } \mu\text{M}$ ) (Fig. 3c). Only humic- and tryptophan-like fluorophore intensities were significantly linearly correlated for the 0-2 cm SRE ( $r = 0.98$ ,  $p = 0.0003$ ,  $n = 8$ ) (Table 3).

In contrast, for the 30-32 cm SRE, leading coefficients were all positive (Fig. 3b,f,h). For dissolved PAHs and the humic-like fluorophore, maximal values ( $29 \pm 20 \text{ ng L}^{-1}$  and  $12 \pm 1.5$

QSU, respectively) were found at  $t = 7$  days (Fig. 3b,f). Those for DOC concentration ( $100 \pm 8.5$   $\mu\text{M}$ ) and the tryptophan-like fluorophore ( $15 \pm 0.9$  QSU) were observed at  $t = 3$  days (Fig. 3d) and  $t = 14$  days (Fig. 3h), respectively. For the 30-32 cm SRE, the correlation between humic- and tryptophan-like fluorophores was still observed ( $r = 0.92$ ,  $p = 0.001$ ,  $n = 8$ ), but in addition, significant linear correlations were found between dissolved PAHs and the humic-like fluorophore ( $r = 0.75$ ,  $p = 0.025$ ,  $n = 8$ ) and between DOC and the tryptophan-like fluorophore ( $r = 0.78$ ,  $p = 0.04$ ,  $n = 7$ ) (Table 3). These correlations are further investigated in § 3.3.3.

In the present study, the adjustment of dissolved concentrations to functions of degree 3 is in agreement with Delle Site (2001) and Karickhoff et al. (1979), who presented the sorption processes of a chemical from one phase to another as the result of a reversible reaction (sorption-desorption) reaching a final equilibrium condition between the concentrations of the chemical in the two phases. One can even suspect redistribution of the chemical between and within phases. Furthermore, the significant leading coefficient being mainly positive suggested that the dissolved  $\Sigma 34$  PAH and DOM patterns for the two SRE may evidence such successive processes: first, desorption from particles related to dissolution or colloid formation from particle collisions that can be followed by some sorption onto/into particles or new aggregations (because of change in polarity or oxygenation conditions for example, especially for the 30-32 cm sediment) and, possibly, some bacterial degradation and volatilization occurring at different time scales between parameters and the two sediments. Even though sorption cubic models globally fit for the two SRE, the remobilization patterns of dissolved PAHs and DOM during SRE appeared to be sediment-dependent. Accordingly, Dang et al. (2015b) also observed different remobilization patterns for Pb between the 0-2 cm and 30-32 cm sediment that were attributed to differences in the oxidation degree, which is further linked to differences in the OM diagenetic state (see § 3.1.2.) of these two sediments.

### 3.3. Factors influencing the PAH sediment-water partitioning

Partition coefficients ( $K_d$  and  $K_{oc}$ ) are key parameters to understand and describe the distribution of PAHs between sediment and water. The  $\log K_d$  and  $\log K_{oc}$  values for PAHs found in the literature display great variability. Even though it is clear that  $K_d$  and  $K_{oc}$  are related to physico-chemical characteristics of the compounds, the variability between studies would come from the strong heterogeneity in the nature/composition of sedimentary particles and OM, which influence the whole sediment sorption capacity.

#### 3.3.1. Hydrophobicity of PAHs

The values for  $\log K_{oc}$  we determined here were plotted *versus* the corresponding  $\log K_{ow}$  in Fig. 4. A strong positive significant correlation was observed between  $\log K_{ow}$  and the measured  $\log K_{oc}$  for the 2-4 ring PAHs for both SRE:

$$\text{For 0-2 cm SRE: } \log K_{oc} = 0.59 \times \log K_{ow} + 4.9 \quad (r = 0.89, p = 0.001, n = 10) \quad (\text{a})$$

$$\text{For 30-32 cm SRE: } \log K_{oc} = 0.64 \times \log K_{ow} + 4.7 \quad (r = 0.91, p < 0.0001, n = 10) \quad (\text{b})$$

The equations (a and b,  $r = 0.89-0.91$ ) revealed that the linear free energy relationship was applicable for 2-4 ring PAHs. The  $\log K_{oc}$  values for the 5-6 ring compounds diverged from the 2-4 ring compound linearity area (Fig. 4). They were lower than expected from equation (a) or (b), and very close to each other, depicting a plateau. They were also closer to  $\log K_{ow}$  values and to  $\log K_{oc}$  predicted from Bouloubassi and Saliot (1992) and Means et al. (1980). These results confirmed that the hydrophobicity of the concerned PAHs remains an essential factor for their partitioning between water and suspended particles. This is in agreement with most of the previous studies (Accardi-Dey and Gschwend, 2003; Bouloubassi and Saliot, 1992; Dong et al.,

2015). However, it is not the only factor, because (i) the log  $K_{oc}$  measured here were generally above log  $K_{ow}$  (Fig. 4), as observed in several studies (Bouloubassi and Saliot, 1992; Feng et al., 2007; Rockne et al., 2002), and above log  $K_{oc}$  measured in previous studies dealing with the resuspension of sediment or porewater contents (Table S2), (ii) the slopes of the linear regressions for the 2-4 ring PAH were  $< 1$  (0.59 and 0.64), which is in accordance with numerous studies, showing that sediment retention capacity was higher for lower MW than for higher MW PAHs (2->3->4-rings) (Bouloubassi and Saliot, 1992; Cao et al., 2015; Fernandes et al., 1997), (iii) the mobility of 5-6 ring PAH diverged from that of 2-4 rings, suggesting that the transport processes differed between these two groups (Feng et al., 2007), and (iv) the remobilization kinetics for the surface and deep sediment were shown to be sediment-dependent. For all of these reasons, other factors, such as sedimentary grain size and OM quality, were suspected to play an important role in PAH partitioning.

### ***3.3.2. Sedimentary grain size and OM quality***

In this work, sediments were characterized by fine-grained particles that promote the accumulation of hydrophobic compounds such as PAHs (Yang et al., 2008). Concerning OM sorption capacities, Cornelissen and Gustafsson (2004 and references therein) support the dual-mode sorption concept. The latter suggests that sedimentary OM retention is due to absorption into amorphous OM, including biopolymers and humic substances (HS), and adsorption onto carbonaceous geosorbents, including unburned coal, kerogen and black carbon (BC). HS and BC are as two major effective sorbents for PAHs in sediment particles (Delle Site, 2001; Khan and Shnitzer, 1972; Cornelissen and Gustafsson., 2004). Cornelissen et al. (2005) supported that important content in carbonaceous geosorbents might explain 1-2 order of magnitude higher  $K_{oc}$  values. HS and BC were not measured in the sediment in the present work. Nonetheless, humic-

like material was evidenced in these Toulon sediments by Dang et al. (2014) and we can reasonably make the assumption of a high presence of sedimentary soot BC because of PAHs originating mainly from combustion-residue-sources (Fig. S2, Readman et al., 1987; Zhou et al., 1999) that can be both released in seawater during our SRE (Dong et al., 2016; Kaal et al., 2016). Therefore, the 2-4 ring and 5-6 ring PAHs could have been partitioned differently between HS and BC, which both present peculiar binding capacities.

The decoupling between 2-4 and 5-6 ring compounds could also be linked to the association of these two groups of PAHs with different sedimentary particle sizes. Indeed, Karickhoff et al. (1979) and Latimer et al. (1999) showed that HMW PAHs are preferentially associated with larger size particles and will therefore have different transport characteristics and fate. In the present case, coarse silts would display lower active surface area, leading to lower retention capacities than the finer ones. This may also explain the relatively higher mobility of 5-6 ring PAHs towards the dissolved phase compared to LMW PAHs.

### **3.3.3. DOM**

Due to the complexity of interactions driving the PAH partitioning between water and sediment, it is very likely that such contaminants were distributed into three phases: the first phase (solid phase, sediment), the second phase (solution phase, water) and the third phase (colloidal or DOM-associated phase) (Delle Site, 2001). Within the third phase, the formation of PAH-DOM complexes may increase the transfer of PAHs towards the dissolved phase (Dong et al., 2016). The effect of the third phase is known to be more pronounced for compounds exhibiting higher hydrophobicity ( $\log K_{ow} > 5$ ) (Delle Site, 2001; Dong et al., 2016).

The remobilization of dissolved PAHs, especially during the 30-32 cm SRE, seemed to be associated with the release of dissolved/colloidal diagenetic HS (correlation between PAHs and

humic-like fluorophore; Table 3, Dang et al., 2015b). This is consistent with the fact that this humic-like fluorophore in the Toulon sediment would arise from the OM diagenetic transformation (Dang et al., 2014). We may assume that these dissolved PAHs released from sediment concomitantly to humic-like material were present in water both as free compounds and as complexes with DOM, which in turn might substantially modify their fate in the water column (Hur et al., 2014; Kim and Kwon, 2010; Sabbah et al., 2004). The coupling between PAHs and HS could be favored by the anoxic/diagenetic state of this deep sediment, since previous studies have shown that the binding capacity between PAHs and HS was negatively related to the oxidation degree and positively related to the OM humification degree (Chen et al. 2010; He and Wang, 2011; Perminova et al., 1999).

Consequently, the relative higher mobility of 5-6 ring compounds towards the dissolved phase we observed here, already highlighted by Mitra et al. (1999), could be explained by their interactions with HS (third phase effect), which would be in accordance with observations from Delle Site (2001). However, a detailed correlation analysis revealed that only 2-4 ring compounds displayed a significant correlation with HS at the 30-32 cm depth ( $r = 0.81$ ,  $p = 0.013$ ,  $n = 8$ ), while 5-6 rings did not correlate ( $r = 0.51$ ,  $p = 0.21$ ,  $n = 8$ ) (not shown in Table 3). This does not necessarily exclude the hypothesis of the third phase distribution for 5-6 ring PAHs, but rather suggests that the latter would occur from their interaction with other OM moieties. Since PAHs in sediment most likely originated from combustion processes, the mobility of 5-6 ring PAHs could be related preferentially to the release of dissolved black carbon (Kaal et al., 2016).

#### ***3.3.4. Equilibration time***

In parallel to previous conclusions, the present high  $\log K_{oc}$  values may be explained by the non-equilibrium partition status. Many authors indicated that partition coefficients measured in

non-equilibrium situations can lead to errors in  $\log K_{oc}$  (Delle Site, 2001). Moreover, if the suspected bi-phase desorption model is effective in the present case, the fast release might have taken a short time (1 h) but the total equilibrium may need months to years to be achieved (Delle Site, 2001 and references therein). Thus, considering the remobilization kinetics, it is very likely that the equilibrium between phases has not been reached and concentrations in the dissolved phase were expected to continue to increase. In spite of this, we aimed to focus on real conditions of few-to-many-day dredging resuspension sediments, and we thought this work might provide useful insight into processes that control PAH fate and transport.

### **3.4. Impact of the MIS sediment resuspension on water quality**

Although the sediment from the MIS site was very highly contaminated and in spite of the PAH dissolved enrichment during the presented experiments, the concentrations of individual PAHs in water during both SRE remained far below the EU WFD MAC-EQS, except for one compound, BP, which exceeded by 3.8 (0-2 cm SRE)- to 5.1 (30-32 cm SRE)-fold the threshold value of  $0.82 \text{ ng L}^{-1}$  (Table 4). These BP concentrations showed that sediment resuspension in Toulon Bay may lead to potential harmful effects on the biota (Feng et al., 2007). However, BP (6-ring compound) may be most likely associated with colloids or DOM, which might in turn reduce its bioavailability and toxicity.

## **4. Conclusion**

Sediment at the MIS site (Toulon Bay) appears to be one of the most contaminated sediments of the Mediterranean coasts in terms of PAHs and trace elements. Sediment resuspension is a

geochemically significant process for PAHs and OM in coastal environments. The two simulated SRE showed 2- to 6-ring PAH remobilization from the sediment to the dissolved phase, increasing seawater concentrations, up to 10-fold. These concentrations remained, however, below the toxicity thresholds, limiting potential adverse effects, except for BP. The PAH transfer from the Toulon sediment to water was lower than predicted from  $\log K_{ow}$ , confirming the sequestration role of such a fine-grained and OC-enriched sediment for hydrophobic contaminants. The remobilization patterns of dissolved PAHs and DOM were different for the two sediments very likely due to their respective diagenetic state and redox status. Additionally, the difference in the relationships between  $\log K_{ow}$  and  $\log K_{oc}$  for 2-4 ring PAHs and 5-6 ring PAHs may be explained by their partition between different OM moieties, i.e., HS and BC, respectively, but also different particle grain sizes, i.e., finer and larger, respectively. Additional experiments would be necessary to discriminate the roles of the particle size and associated OM, as well as weathering processes (degradation, volatilization), particularly for LMW PAHs, on the sediment-water PAH partitioning at this site. The combination of sedimentary OM and insufficient equilibration time may easily explain 2-3 order of magnitude higher  $K_{oc}$  values compared to other studies. Resuspension of sediments remains a complex issue to interlink particle concentrations with the degradation of water quality and the living organism exposure. These results should be considered for the future management operations and environmental policy in polluted coastal areas such as Toulon harbor.

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## Figure captions

**Figure 1.** Location of the Missiessy (MIS) site in Toulon harbor (Southern France, Northwestern Mediterranean Sea).

**Figure 2.** Distribution profile of 1) PAHs in the 0-2 and 30-32 cm sediments used for sediment resuspension experiments (SRE) (“0-2 cm sediment” and “30-32 cm sediment”), 2) dissolved PAHs in water samples of SRE performed on the 0-2 cm and 30-32 cm sediments (for each SRE, mean of the eight times: 1, 3, 7 h, 1, 3, 7, 10 and 14 days, with two bottle replicates for each time;  $n = 16$ ) (“0-2 cm SRE” and “30-32 cm SRE”), and 3) dissolved PAHs in the water column (mean of the four depths: 0.5, 2.5, 5 and 13 m with two bottle replicates for each depth;  $n = 8$ ) (“Water column”). Different letters (*a*, *b*, *c*, *d* or *e*) are used for significantly different means (*t*-test,  $p < 0.05$ ).

**Figure 3.** Concentrations of dissolved  $\Sigma 34$  PAHs ( $\text{ng L}^{-1}$ ) (a,b), concentrations in DOC ( $\mu\text{M}$ ) (c, d), and fluorescence intensities of humic-like (e, f) and tryptophan-like (g, h) fluorophores (QSU) in water during the 0-2 cm (white dots) and 30-32 cm (gray dots) sediment resuspension experiments (SRE). Polynomial functions of degree 3 (cubic functions) obtained by least square fits are shown as black lines along with their respective equation and correlation coefficient. The dotted line represents the concentration in the water before sediment addition for each parameter. The standard deviation is given for the two bottle replicates ( $n = 2$ ). For DOC, the sample at day 7 for 30-32 cm SRE, considered as an outlier, was removed.

**Figure 4.** Relationship between  $\log K_{ow}$  and  $\log K_{oc}$  determined during the sediment resuspension experiments performed on the 0-2 cm (white dots for 2-4 ring compounds, white square for 5-6 ring ones) and 30-32 cm (gray dots for 2-4 ring compounds, gray square for 5-6 ring ones) sediments for the 16 priority PAHs. The equations for 2-4 ring and 5-6 ring compounds in the 0-2 cm SRE were  $y = 0.59x + 4.9$  ( $r = 0.89$ ,  $n = 10$ ,  $p < 0.05$ ) and  $y = 0.07x + 7.3$  ( $r = 0.12$ ,  $n = 6$ ,  $p > 0.05$ ), respectively. The equations related to 2-4 ring and 5-6 ring compounds in the 30-32 cm SRE were  $y = 0.64x + 4.7$  ( $r = 0.91$ ,  $n = 10$ ,  $p < 0.05$ ) and  $y = -0.17x + 8.7$  ( $r = 0.24$ ,  $n = 6$ ,  $p > 0.05$ ), respectively. Crosses represent the theoretical relationship  $\log K_{oc} = \log K_{ow} - 0.32$  according to Means et al. (1980). Black diamonds represent values from Bouloubassi and Saliot(1992). The standard deviation of  $\log K_{oc}$  for each compound is given for the two bottle replicates ( $n = 2$ ). The  $\log K_{ow}$  are the means of different values reported in Accardi-Dey and Gschwend (2003); Dong et al. (2015); Feng et al. (2007); Socha and Carpenter (1987).

**Table 1.** Concentration range of the 16 priority PAHs ( $10^3 \text{ ng g}^{-1} \text{ sed. dw}$ ) in the whole sediment core and in the 0-2 and 30-32 cm sediments used for sediment resuspension experiments (SRE). Comparison with the toxicity critical levels published in JORF for the French legislation and with the international SQG references proposed by Long et al. (1995).

| Compound                 | Whole core | 0-2 cm deep | 30-32 cm deep | JORF (fraction < 2 mm) |       | SQG (Long et al., 1995) |       |
|--------------------------|------------|-------------|---------------|------------------------|-------|-------------------------|-------|
|                          |            |             |               | L1                     | L2    | ERL                     | ERM   |
| Naph                     | 0.161–1.22 | 0.526       | 0.585         | 0.160                  | 1.13  | 0.160                   | 2.10  |
| Acy                      | 0.102–1.01 | 0.179       | 0.247         | 0.040                  | 0.340 | 0.044                   | 0.640 |
| Ace                      | 0.233–2.38 | 0.769       | 0.468         | 0.015                  | 0.260 | 0.016                   | 0.500 |
| Flu                      | 0.287–2.45 | 0.484       | 0.383         | 0.020                  | 0.280 | 0.019                   | 0.540 |
| Phe                      | 2.37–24.6  | 2.98        | 2.96          | 0.240                  | 0.870 | 0.240                   | 1.50  |
| Ant                      | 0.526–3.75 | 0.588       | 0.634         | 0.085                  | 0.590 | 0.085                   | 1.10  |
| Flt                      | 3.22–46.6  | 4.38        | 4.41          | 0.600                  | 2.85  | 0.600                   | 5.10  |
| Pyr                      | 2.78–36.1  | 3.41        | 3.70          | 0.500                  | 1.50  | 0.665                   | 2.60  |
| BaA                      | 0.988–11.3 | 1.83        | 1.39          | 0.260                  | 0.930 | 0.261                   | 1.60  |
| Chr                      | 0.988–10.1 | 1.78        | 1.43          | 0.380                  | 1.59  | 0.384                   | 2.80  |
| BbF                      | 2.020–13.8 | 3.42        | 2.84          | 0.400                  | 0.900 |                         |       |
| BkF                      | 0.601–4.30 | 0.912       | 0.878         | 0.200                  | 0.400 |                         |       |
| BaP                      | 1.15–9.12  | 2.13        | 1.71          | 0.430                  | 1.02  | 0.430                   | 1.60  |
| IndP                     | 1.50–7.48  | 2.71        | 1.99          | 1.70                   | 5.65  |                         |       |
| DBA                      | 0.331–2.09 | 0.553       | 0.452         | 0.060                  | 0.160 | 0.063                   | 0.260 |
| BP                       | 1.05–6.47  | 1.89        | 1.77          | 1.70                   | 5.65  |                         |       |
| $\Sigma 16 \text{ PAHs}$ | 22.9-182   | 28.5        | 25.8          |                        |       |                         |       |
| $\Sigma 34 \text{ PAHs}$ | 30.7-231   | 38.2        | 35.7          |                        |       |                         |       |

JORF: French Republic Official Journal; SQG: sediment quality guidelines. L1 and L2: critical levels 1 and 2; ERL and ERM: effects range-low and effects range-median.

**Table 2.** Physical and chemical characteristics of the 0-2 and 30-32 cm sediments used for sediment resuspension experiments (SRE).

|  | 0-2 cm sediment | 30-32 cm sediment |
|--|-----------------|-------------------|
| Moisture (%)                                   | 62              | 52                |
| Sand (%)                                       | 0               | 0                 |
| Silt (%)                                       | 69              | 82                |
| Clay (%)                                       | 31              | 18                |
| OC (%)   | 8.2             | 6.3               |
| ON (%)   | 0.61            | 0.29              |
| C/N  | 13.5            | 22.0              |
| $\Sigma 34$ PAHs ( $10^3$ ng g <sup>-1</sup> ) | 38.2            | 35.7              |
| <i>R</i> ( $10^3$ ng g <sup>-1</sup> )         | 2.41            | 10.0              |
| UCM/ <i>R</i>                                  | 30              | 118               |
| CPI <sub>15-24</sub>                           | 1.3             | 1.3               |
| CPI <sub>25-36</sub>                           | 2.9             | 2.2               |

OC and ON: organic carbon and nitrogen (% for 1 g sed. dw);  $\Sigma 34$  PAHs: concentrations in total polycyclic aromatic hydrocarbons ( $10^3$  ng g<sup>-1</sup> sed. dw); *R*: concentration in resolved *n*-alkanes from *n*-C<sub>15</sub> to *n*-C<sub>36</sub> with two isoprenoids, pristane (Pr) and phytane (Phy) ( $10^3$  ng g<sup>-1</sup> sed. dw); UCM: unresolved complex mixture. CPI<sub>15-24</sub> and CPI<sub>25-36</sub>: carbon preference indices in the ranges *n*-C<sub>15</sub>-*n*-C<sub>24</sub> and *n*-C<sub>25</sub>-*n*-C<sub>36</sub>.

**Table 3.** Pearson's correlation matrices for the concentrations of dissolved  $\Sigma 34$  PAHs and dissolved organic matter (DOM) during sediment resuspension experiments (SRE) conducted on the 0-2 and 30-32 cm sediments. Significant linear correlations ( $p < 0.05$ ) are in bold.

a) 0-2 cm SRE (n = 8)

| Variables  | $\Sigma 34$ PAHs | DOC  | Humic       |
|------------|------------------|------|-------------|
| DOC        | -0.03            |      |             |
| Humic      | 0.69             | 0.39 |             |
| Tryptophan | 0.56             | 0.43 | <b>0.98</b> |

b) 30-32 cm SRE (n = 7 or 8)

| Variables  | $\Sigma 34$ PAHs | DOC         | Humic       |
|------------|------------------|-------------|-------------|
| DOC        | 0.50             |             |             |
| Humic      | <b>0.75</b>      | 0.66        |             |
| Tryptophan | 0.54             | <b>0.78</b> | <b>0.92</b> |

PAHs: polycyclic aromatic hydrocarbons; DOC: dissolved organic carbon; Humic and tryptophan: Humic- and tryptophan-like fluorophores of fluorescent dissolved organic matter (FDOM).

**Table 4.** Concentration ranges of the 16 priority PAHs in water ( $\text{ng L}^{-1}$ ) during sediment resuspension experiments (SRE) performed on the

0-2 and 30-32 cm sediments. Comparison with the MAC-EQS of the EU Water Framework Directive (WFD, 2013/39/EU).

| Compound | 0-2 cm SRE      | 30-32 cm SRE    | MAC-EQS |
|----------|-----------------|-----------------|---------|
| Naph     | 0.3–1.7         | 0.6–2.7         | 130 000 |
| Acy      | 0.1–0.2         | 0.1–1.9         |         |
| Ace      | 0.3–0.8         | 0.5–3.1         |         |
| Flu      | 0.3–1.6         | 0.4–2.9         |         |
| Phe      | 0.5–2.2         | 0.6–4.5         |         |
| Ant      | <i>bdl</i> –0.7 | <i>bdl</i> –1.7 | 100     |
| Flt      | 0.2–3.9         | 0.3–3.3         | 120     |
| Pyr      | 0.8–4.8         | 0.2–2.9         |         |
| BaA      | <i>bdl</i> –0.2 | <i>bdl</i> –0.6 |         |
| Chr      | 0.1–3.4         | <i>bdl</i> –2.2 |         |
| BbF      | 0.6–2.6         | 0.7–3.7         | 17      |
| BkF      | 0.2–1.3         | <i>bdl</i> –1.9 | 17      |
| BaP      | 0.1–1.6         | <i>bdl</i> –2.5 | 27      |
| IndP     | <i>bdl</i> –2.0 | <i>bdl</i> –2.1 |         |
| DBA      | <i>bdl</i> –0.4 | <i>bdl</i> –1.0 |         |
| BP       | 0.2–3.1         | <i>bdl</i> –4.2 | 0.82    |

MAC-EQS: Maximum allowable concentration-environmental quality standards in water for a particular event/forcing; *bdl*: below detection limit.

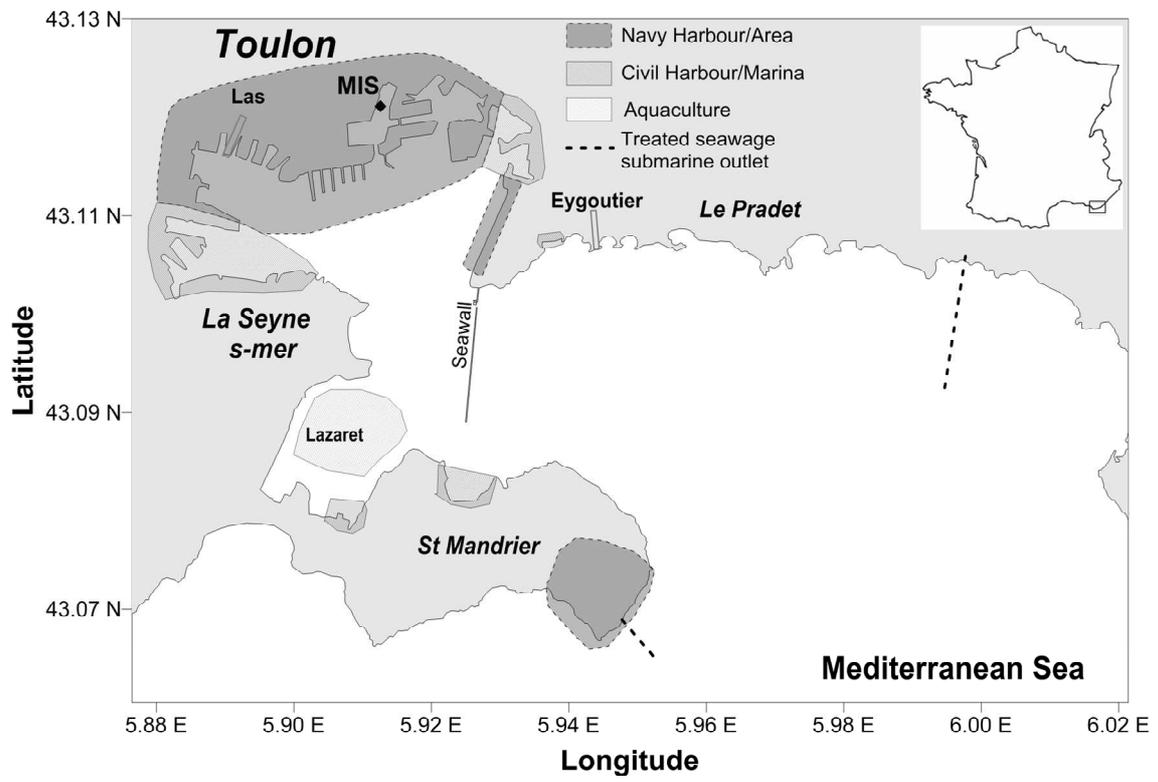


Figure 1

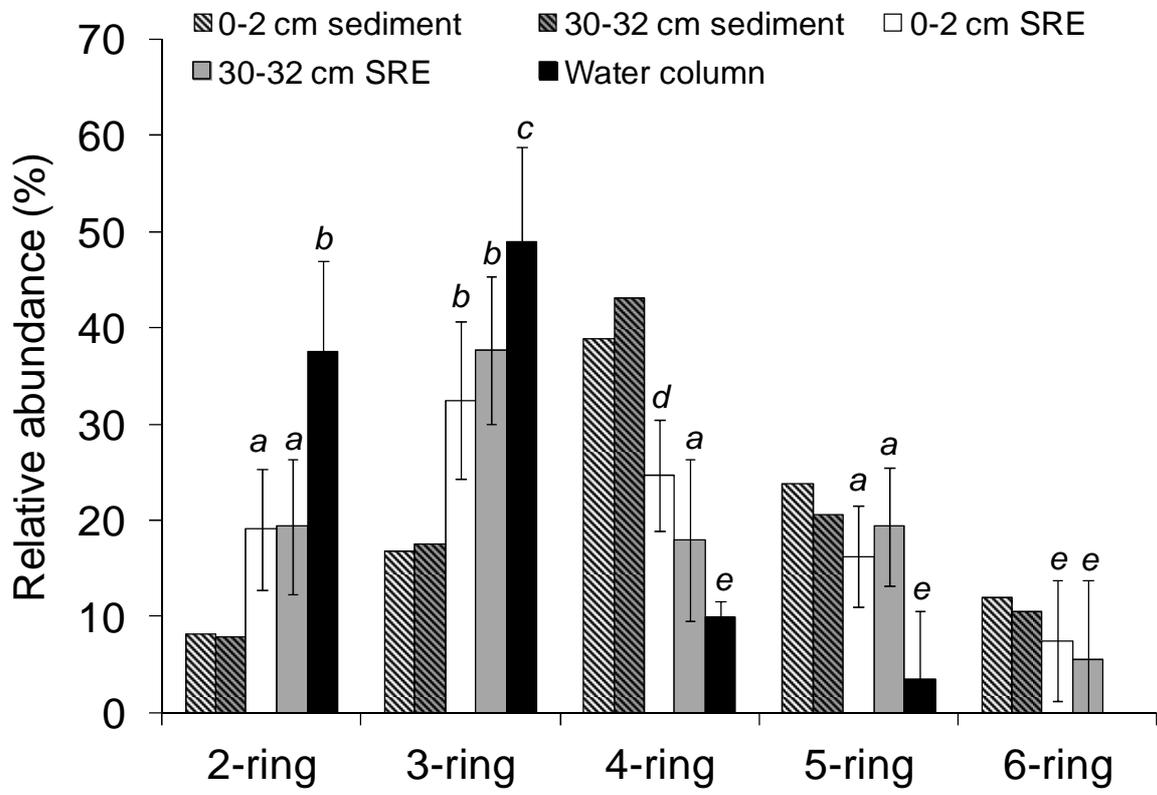


Figure 2

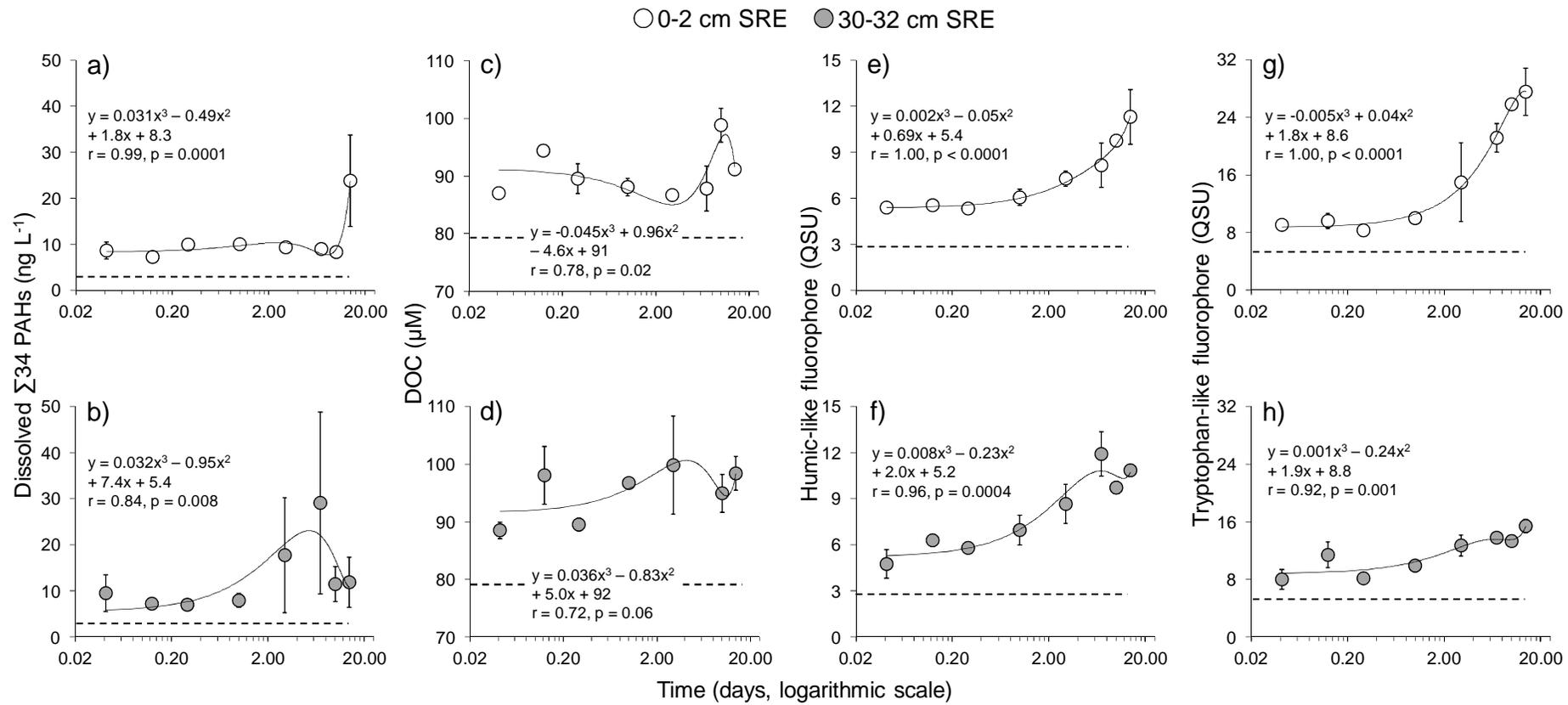


Figure 3

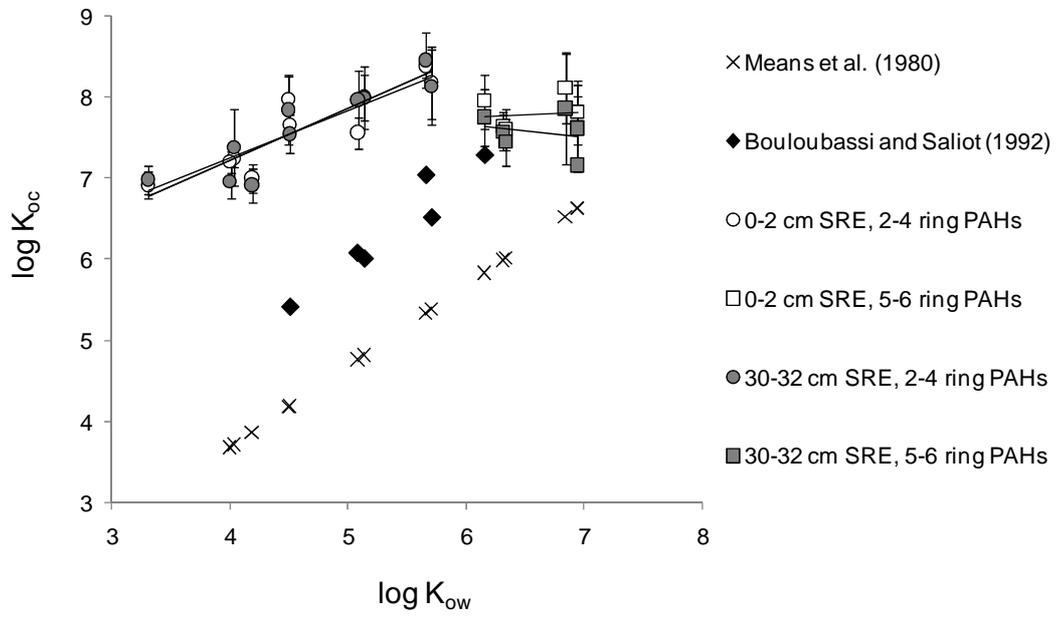


Figure 4

**Remobilization of polycyclic aromatic hydrocarbons and organic matter in seawater during sediment resuspension experiments from a polluted coastal environment: insights from Toulon Bay (France)**

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**Supplementary material**

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### **Text S1. Determination of hydrocarbons**

Concerning PAHs, measured compounds are namely naphthalene (Naph), acenaphthylene (Acy), acenaphthene (Ace), fluorene (Flu), dibenzothiophene (DBT), phenanthrene (Phe), anthracene (Ant), fluoranthene (Flt), pyrene (Pyr), benz[*a*]anthracene (BaA), chrysene (Chr), benzo[*b*]fluoranthene (BbF), benzo[*k*]fluoranthene (BkF), benzo[*e*]pyrene (BeP), benzo[*a*]pyrene (BaP), perylene (Per), dibenz[*a,h*]anthracene (DBA), benzo[*g,h,i*]perylene (BP), indeno[1,2,3-*cd*]pyrene (IndP). The isomeric ratios (Ant/Ant+Phe, BaA/BaA+Chr, IndP/IndP+BP and Flt/Flt+Pyr) were used to assess the origin of the sediment particles (Tobiszewski and Namieśnik, 2012; Yunker et al., 2002).

Concerning AHs, the UCM hump corresponds to a mixture of many structurally complex isomers and homologues of branched and cyclic hydrocarbons that cannot be resolved by capillary GC columns (Bouloubassi and Saliot, 1993). Its relative importance, expressed as the ratio of unresolved to resolved compounds (UCM/*R*), is commonly used as a diagnostic criterion of petroleum inputs (Mazurek and Simoneit, 1984). We also determined the carbon preference indices in the ranges *n*-C<sub>15</sub>-*n*-C<sub>24</sub> and *n*-C<sub>25</sub>-*n*-C<sub>36</sub> (CPI<sub>15-24</sub> and CPI<sub>25-36</sub>, respectively), expressed as the ratio of odd to even carbon numbered *n*-alkanes (Blumer et al., 1971; Bouloubassi and Saliot, 1993; Douglas and Eglinton, 1966). Finally, we also assessed the relative proportions of biogenic (marine and terrigenous) and petroleum *n*-alkanes using the formula: % anthropogenic *n*-alkanes =  $2 / (CPI + 1) \times 100$

### **Text S2. Environmental quality guidelines**

For the present study, the PAH concentrations were compared either to the (i) “Official Journal of the French Republic” (JORF), number 0046, February 23<sup>th</sup> 2013 as current local legislation and to (ii) the “sediment quality guidelines” (SQG) proposed by Long et al. (1995) as international reference (Barhoumi et al., 2014; Guo et al., 2007; Zaghden et al., 2017). They both proposed two critical levels: the levels 1 and 2 for JORF (L1 and L2, respectively)

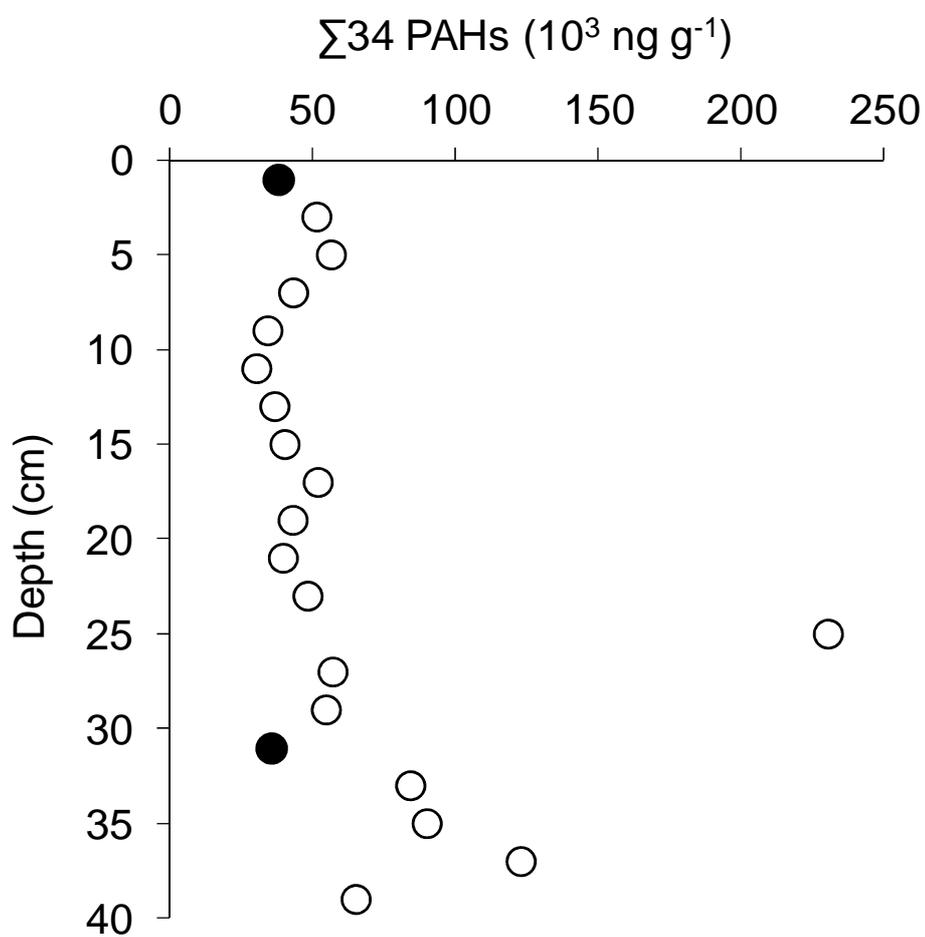
and the effect range-low (ERL) and effect range-median (ERM) for SQG. The two low levels (L1 and ERL), when exist, are very close to each other, whereas ERM are almost the double of L2 (Table 1). Briefly, the concentrations below L1 and ERL values reflect no particular contamination or conditions in which biologically adverse effects would be rarely observed. The concentrations between L1 and L2 or ERL and ERM values represent a potential contamination with adverse effects occasionally occurring and imply complementary investigations. The concentrations equal or above L2 or ERM values represent a high contamination with adverse effects frequently occurring. For waters, the PAH concentrations were compared to the maximum allowable concentration for environmental quality standards (MAC-EQS) defined by the European Union Water Framework Directive (WFD, 2013/39/EU).

**Table S1.** Chemical characteristics of porewaters of the 0-2 and 30-32 cm sediments used for sediment resuspension experiments.

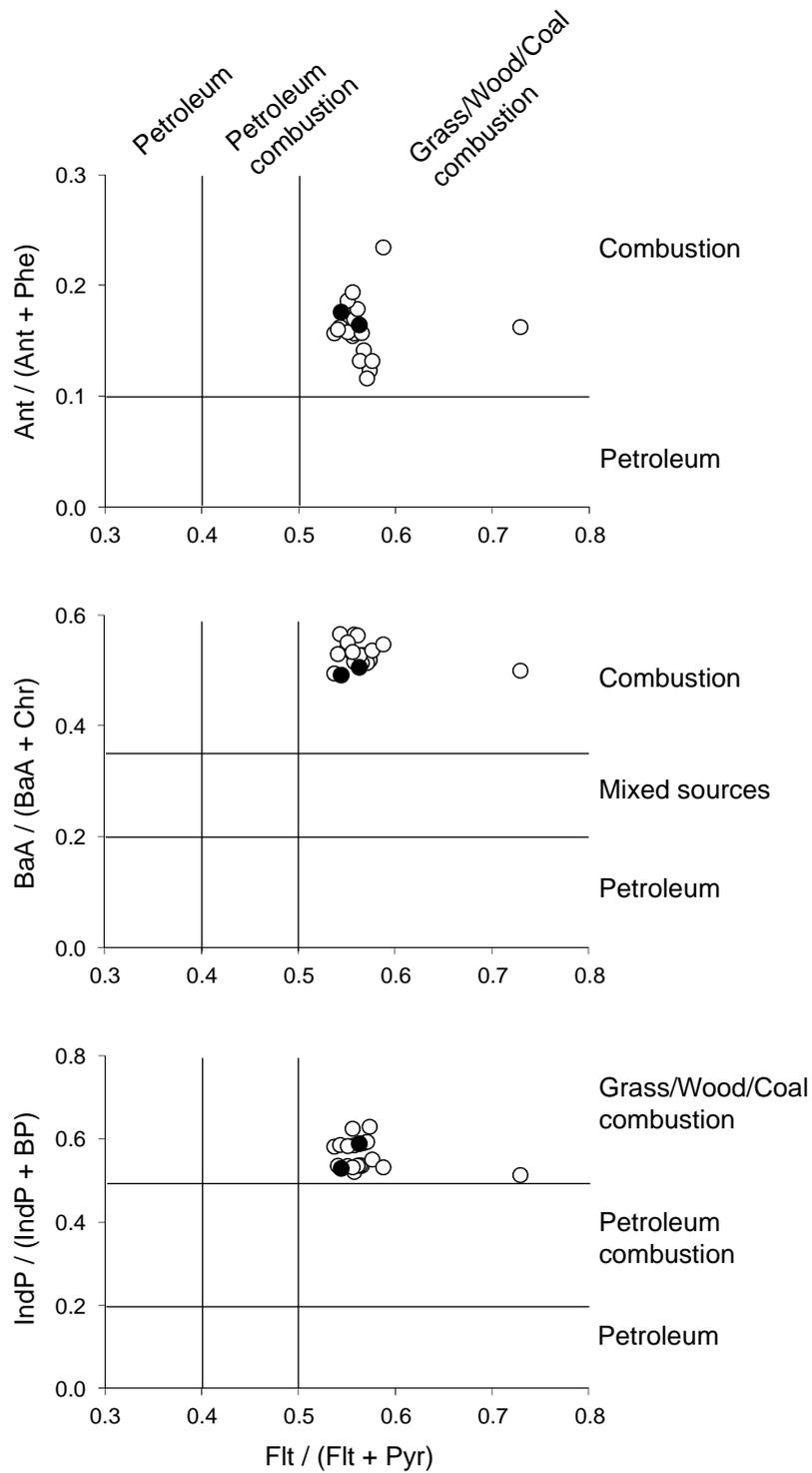
|   | 0-2 cm sediment | 30-32 cm sediment |
|---|-----------------|-------------------|
| pH  | 7.49            | 7.51              |
| Eh (mV/ENH)   | 109             | -145              |
| DOC ( $\mu\text{M}$ )   | $367 \pm 8$     | $717 \pm 8$       |
| DIC (mM)  | $2.81 \pm 0.02$ | $2.78 \pm 0.03$   |
| $\text{N}_\text{T}$ ( $\mu\text{M}$ )                         | $104 \pm 2$     | $173 \pm 8$       |
| $\text{NH}_4^+$ ( $\mu\text{M}$ )                             | 72.6            | 54.8              |
| SRP ( $\mu\text{M}$ )   | 7.1             | 5.1               |
| $\text{Si}(\text{OH})_4$ ( $\mu\text{M}$ )                    | 95.4            | 110.2             |
| $\text{Mn}_\text{T}$ (nM)                                     | 2.3             | 0.7               |
| $\text{Fe}(\text{II}) = \text{Fe}_\text{T}$ ( $\mu\text{M}$ ) | 70.2            | 7.7               |
| $\text{SO}_4^{2-}$ (mM)                                       | 30.7            | 28.8              |
| $\Sigma\text{HS}^-$ ( $\mu\text{M}$ )                         | <0.3            | <0.3              |

**Table S2.** Partition coefficients of the individual PAHs between sediment and water ( $\log K_d$ ) and between sediment organic matter and water ( $\log K_{oc}$ ) determined in this study during sediment resuspension experiments (SRE). Comparison with the  $\log K_d$ ,  $\log K_{oc}$  and partition coefficients between octanol and water ( $\log K_{ow}$ ) from the literature. The  $\log K_{ow}$  are the means of different values reported in Accardi-Dey and Gschwend (2003); Dong et al. (2015); Feng et al. (2007); Socha and Carpenter (1987).

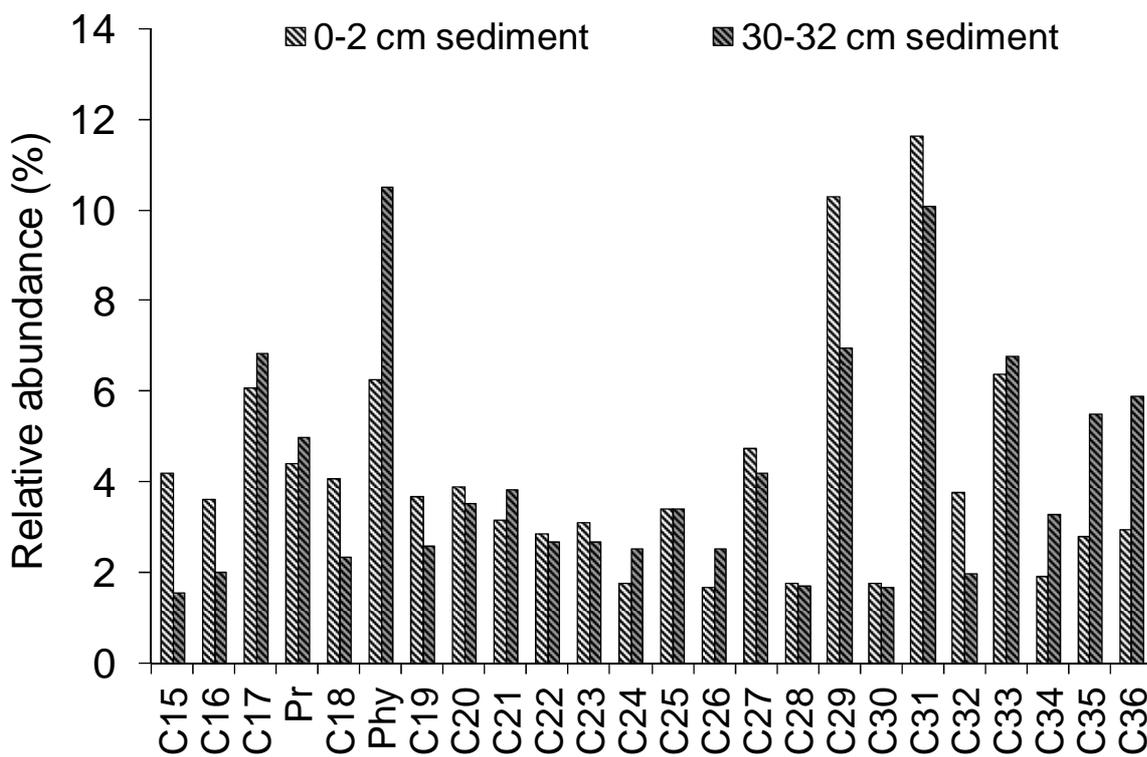
|                                 |                | <b>Naph</b>           |                        | <b>Acy</b>            |                        | <b>Ace</b>            |                        | <b>Flu</b>            |                        | <b>Phe</b>            |                        | <b>Ant</b>            |                        | <b>Flt</b>            |                        | <b>Pyr</b>            |                        |
|---------------------------------|----------------|-----------------------|------------------------|-----------------------|------------------------|-----------------------|------------------------|-----------------------|------------------------|-----------------------|------------------------|-----------------------|------------------------|-----------------------|------------------------|-----------------------|------------------------|
| <i>log K<sub>ow</sub></i>       |                | 3.31                  |                        | 4.04                  |                        | 4.00                  |                        | 4.19                  |                        | 4.51                  |                        | 4.50                  |                        | 5.14                  |                        | 5.09                  |                        |
|                                 | %<br>OC        | log<br>K <sub>d</sub> | log<br>K <sub>oc</sub> |
| <b>This study</b>               | <b>6.3–8.2</b> | <b>5.8</b>            | <b>6.9–7.0</b>         | <b>6.2</b>            | <b>7.3–7.4</b>         | <b>5.8–6.1</b>        | <b>7.0–7.2</b>         | <b>5.7–5.9</b>        | <b>6.9–7.0</b>         | <b>6.3–6.6</b>        | <b>7.5–7.7</b>         | <b>6.6–6.9</b>        | <b>7.8–8.0</b>         | <b>6.8–6.9</b>        | <b>8.0</b>             | <b>6.5–6.8</b>        | <b>7.6–8.0</b>         |
| Accardi-Dey and Gschwend (2002) | 1.2–3.1        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        | 3.5–3.7               | 5.3                    |
| Accardi-Dey and Gschwend (2003) |                |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       | 3.9–5.6                |
| Cornelissen et al. (2006)       | 1.7–7.4        |                       |                        |                       |                        |                       |                        |                       |                        |                       | 6.0–6.2                |                       |                        |                       | 5.8–6.4                |                       | 5.9–6.3                |
| Dong et al. (2015)              | 0.9–4.8        |                       | 4.0–4.9                |                       | 5.2–5.7                |                       | 4.8–5.4                |                       | 5.0–5.7                |                       | 5.3–6.1                |                       | 4.7–5.7                |                       | 5.2–6.2                |                       | 4.5–6.4                |
| Feng et al. (2007)              | 1.1–6.3        |                       |                        |                       |                        | 2.7–3.2               |                        | 2.2–2.9               |                        | 2.5–3.1               |                        | 2.4                   |                        | 3.1–3.3               |                        | 3.2–3.7               |                        |
| Jonker and Koelmans (2001)      |                |                       |                        |                       |                        |                       |                        |                       |                        |                       | 5.7                    |                       |                        |                       |                        |                       | 6.1                    |
| Latimer et al. (1999)           | 2.5–5.6        | 2.0–3.4               |                        | 2.3–4.6               |                        | 2.6–4.4               |                        |                       |                        | 3.9–6.2               |                        | 3.9–4.1               |                        | 4.3–5.3               |                        | 4.3–5.0               |                        |
| Mitra et al. (1999a)            |                |                       |                        |                       |                        |                       | 3.6–6.1                |                       |                        |                       |                        |                       |                        |                       |                        |                       | 3.1–6.5                |
| Mitra et al. (1999b)            | 1.5–4.5        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       | 5.1–5.4                |
| Socha and Carpenter (1987)      | 0.4–2.7        |                       |                        |                       |                        |                       |                        |                       |                        |                       | 5.0                    |                       |                        |                       | 4.5                    |                       | 5.1                    |
|                                 |                | <b>BaA</b>            |                        | <b>Chr</b>            |                        | <b>BbF</b>            |                        | <b>BkF</b>            |                        | <b>BaP</b>            |                        | <b>IndP</b>           |                        | <b>DBA</b>            |                        | <b>BP</b>             |                        |
| <i>log K<sub>ow</sub></i>       |                | 5.66                  |                        | 5.71                  |                        | 6.31                  |                        | 6.34                  |                        | 6.15                  |                        | 6.84                  |                        | 6.94                  |                        | 6.94                  |                        |
|                                 | %<br>OC        | log<br>K <sub>d</sub> | log<br>K <sub>oc</sub> |
| <b>This study</b>               | <b>6.3–8.2</b> | <b>7.3</b>            | <b>8.4–8.5</b>         | <b>6.9–7.1</b>        | <b>8.1–8.2</b>         | <b>6.4–6.6</b>        | <b>7.6</b>             | <b>6.2–6.5</b>        | <b>7.4–7.6</b>         | <b>6.6–6.9</b>        | <b>7.8–8.0</b>         | <b>6.6–7.0</b>        | <b>7.9–8.1</b>         | <b>6.0–6.5</b>        | <b>7.2–7.6</b>         | <b>6.4–6.7</b>        | <b>7.6–7.8</b>         |
| Cornelissen et al. (2006)       | 1.7–7.4        |                       |                        |                       | 6.9–7.5                |                       | 6.9–7.3                |                       |                        |                       | 6.8–7.5                |                       | 7.3–8.1                |                       |                        |                       | 7.6–8.1                |
| Dong et al. (2015)              | 0.9–4.8        |                       | 5.4–6.4                |                       | 5.1–6.0                |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |                       |                        |
| Latimer et al. (1999)           | 2.5–5.6        | 4.2–4.9               |                        | 4.3–4.9               |                        | 4.1–4.6               |                        | 4.0–5.5               |                        | 4.6–5.2               |                        | 3.6–4.8               |                        | 2.3–4.0               |                        | 3.7–4.9               |                        |
| Jonker and Koelmans (2001)      |                |                       |                        |                       |                        |                       |                        |                       |                        |                       | 7.96                   |                       |                        |                       |                        |                       | 7.84                   |
| Lohman et al. (2005)            |                |                       |                        |                       |                        |                       |                        |                       |                        |                       | 7.0–7.5                |                       |                        |                       |                        |                       |                        |
| Mitra et al. (1999a)            |                |                       |                        |                       |                        |                       | 4.1–7.1                |                       |                        |                       | 2.7–7.0                |                       | 4.0–7.0                |                       | 2.5–6.6                |                       |                        |
| Mitra et al. (1999b)            | 1.5–4.5        |                       |                        |                       |                        |                       | 5.1–6.0                |                       | 5.1–6.0                |                       | 5.4–6.2                |                       |                        |                       |                        |                       | 5.1–6.0                |
| Socha and Carpenter (1987)      | 0.4–2.7        |                       | 5.5                    |                       | 5.5                    |                       | 5.8                    |                       | 5.8                    |                       | 5.9                    |                       |                        |                       |                        |                       | 6.4                    |



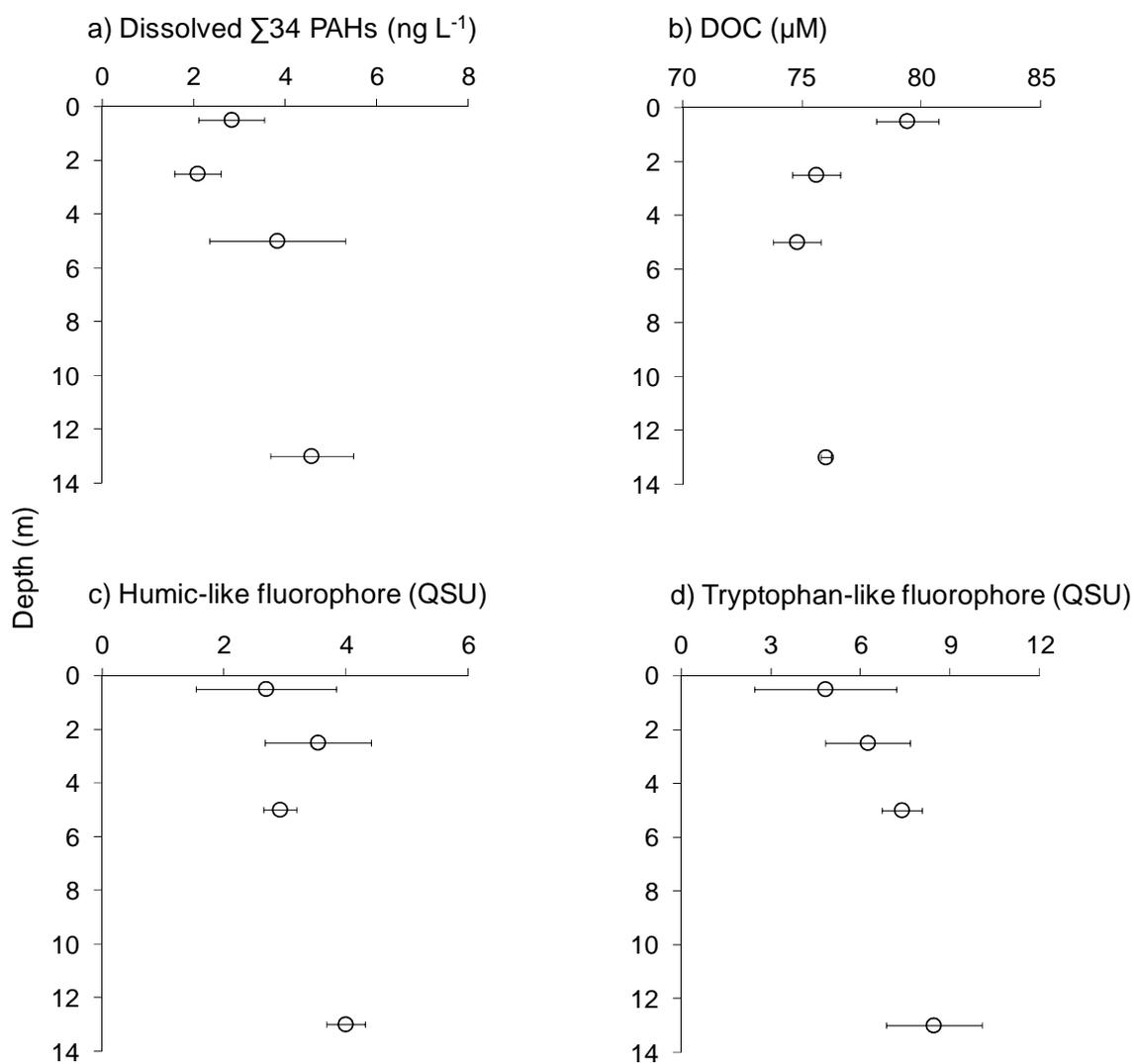
**Figure S1.** Concentrations of  $\Sigma 34$  PAHs ( $10^3 \text{ ng g}^{-1}$  sed. dw) with depth in the whole sediment core from the MIS site. The black circles represent the 0-2 and 30-32 cm sediment samples used for sediment resuspension experiments (SRE).



**Figure S2.** Cross plots of PAH isomeric ratios for the whole sediment core from the MIS site. The black circles represent the 0-2 and 30-32 cm sediment samples used for sediment resuspension experiments (SRE).



**Figure S3.** Molecular distribution profile (%) of resolved *n*-alkanes with pristane (Pr) and phytane (Phy) for the 0-2 and 30-32 cm sediments used for sediment resuspension experiments (SRE).



**Figure S4.** Concentrations in a) dissolved  $\Sigma 34$  PAHs ( $\text{ng L}^{-1}$ ), b) DOC ( $\mu\text{M}$ ) and fluorescence intensities of c) humic-like and d) tryptophan-like fluorophores (QSU) with depth in the water column from the MIS site.

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