

# Fate of sulfamethoxazole, its main metabolite N-ac-sulfamethoxazole and ciprofloxacin in agricultural soils amended or not by organic waste products

Aurore Andriamalala, Laure Vieublé, Valerie Dumeny, Philippe Cambier

# ▶ To cite this version:

Aurore Andriamalala, Laure Vieublé, Valerie Dumeny, Philippe Cambier. Fate of sulfamethoxazole, its main metabolite N-ac-sulfamethoxazole and ciprofloxacin in agricultural soils amended or not by organic waste products. Chemosphere, 2018, 191, pp.607-615. 10.1016/j.chemosphere.2017.10.093 . hal-01629852

# HAL Id: hal-01629852 https://hal.science/hal-01629852

Submitted on 5 Feb 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Fate of Sulfamethoxazole, its main metabolite N-ac-Sulfamethoxazole and Ciprofloxacin
2	in agricultural soils amended or not by organic waste products
3	Aurore Andriamalala <sup>a</sup> , Laure Vieublé-Gonod <sup>b</sup> , Valérie Dumeny <sup>a</sup> . Philippe Cambier <sup>a</sup>
4	<sup>a</sup> UMR Ecosys, INRA, AgroParisTech, Université Paris-Saclay, 78850, Thiverval Grignon,
5	France
6	<sup>b</sup> UMR Ecosys, AgroParisTech, INRA, Université Paris-Saclay, 78850, Thiverval Grignon,
7	France
8	E-mail: laure.vieuble@agroparistech.fr
9	Corresponding author:
10	Philippe Cambier (Tel. 330130815988; E-mail:philippe.cambier@inra.fr)

## 12 Abstract:

13 Spreading organic waste products (OWP) issued from sewage sludge or manures into soil may disseminate antibiotics with unknown risks for human health and environment. Our 14 15 objectives were to compare the fate of two sulfonamides, sulfamethoxazole (SMX) and its metabolite N-acetyl-sulfamethoxazole (N-ac-SMX), and one fluoroquinolone, ciprofloxacin 16 (CIP), in an unamended soil, and two soils regularly amended since 1998 with a sewage 17 sludge and green waste compost and with farmyard manure respectively. Incubations of soil 18 spiked with <sup>14</sup>C labelled SMX or N-ac-SMX (0.02 mg.kg<sup>-1</sup>) or CIP (0.15 mg.kg<sup>-1</sup>) allowed a 19 quantification of radiolabeled molecules in the mineralized, easily, hardly and non-extractable 20 fractions after 3 and 156 days. Nature of <sup>14</sup>C molecules was also analyzed by HPLC in 21 extractable fractions after 3 and 156 days. 22

SMX and N-ac-SMX dissipation was fast and due to i) mineralization (~10% of recovered <sup>14</sup>C
 after 156 days) or incomplete degradation (production of metabolites), ii) adsorption, even if

25	both sulfonamides present low Kd (<3 L.kg <sup>-1</sup> ) and iii) formation of non-extractable residues
26	(NER), representing more than 50% of recovered radioactivity.
27	N-ac-SMX was more mineralized than SMX, and formed more progressively NER, after a
28	step of deacetylation. Adsorption of CIP was fast and formed mainly NER (> 72%) whereas
29	its mineralization was negligible. Repeated applications of OWP tend to enhance adsorption
30	of antibiotics and lower their degradation, through the quantity and quality of the built up soil
31	organic matter. If applications of sewage sludge compost favor adsorption and inhibit
32	mineralization, applications of manure boost the formation of non-extractable residues.
33	
34	Keywords: Soil; Antibiotic; Metabolite; Organic waste products; Biodegradation; Adsorption
35	
36	Highlights:
37	N-ac-SMX slightly more mobile and more mineralized than SMX during soil incubations
38	Differences in fates of N-ac-SMX and SMX partly due to a step of deacetylation
39	SMX more available in soil than CIP but both mainly non-extractable after 156 days
40	Regular organic amendments enhance sorption and lower mineralization of antibiotics
41	

#### 43 1. Introduction

The increasing occurrence of antibiotics (ATB) in soils, surface water and ground water becomes of more and more concern because of environmental and human health threat (Boxall et al., 2012).

Once in the soil, ATB may be degraded by biotic or abiotic path, retained on soil constituents and/or transferred to surface and groundwater through runoff and infiltration (Jechalke et al., 2014). Volatilization and photodegradation of antibiotics are generally negligible processes in soils (Ozaki et al., 2011; Jechalke et al., 2014). Indeed, the fate of antibiotics depends on antibiotics and soil properties. Generally detected in the µg-mg per kg range (Golet et al., 2002; Boxall et al., 2004), ATB may persist for years in soils (Du and Liu, 2012) while maintaining an activity even at a low dose (Boxall et al., 2004).

One of the entry routes of veterinary and human antibiotics into agricultural soils is the 54 application of organic waste products (OWP) (Kümmerer, 2001), which is a worldwide 55 increasingly common practice of recycling valuable nutrients and organic matter (Chang et 56 al., 1995). However OWP may contain antibiotics. When administered to livestock, 10 to 57 90% of ATB are excreted as parent compounds or metabolites (Sarmah et al., 2006). 58 Consequently ATB and metabolites can be present directly in livestock effluents (Boxall et 59 60 al., 2004). ATB in human's urine and feces are not completely eliminated in waste water plants and can persist in effluents and/or sludge (Nieto et al., 2010). 61

OWP addition may also impact the fate of antibiotics in soils, through changing soil physical,
chemical and biological properties (Li et al., 2015). Contradictory results were obtained under
laboratory conditions. Some works highlighted a stimulation of ATB biodegradation after
OWP addition (Accinelli et al., 2007; Zhang et al., 2017) while others showed an increase of
ATB adsorption and inhibition of mineralization (Li et al., 2015).

However studies based on addition of OWP into soils in the lab miss the long-term effects of 67 68 OWP applications on soil and their indirect effect on the fate of ATB. In this study, we compared the fate of ATB in three soils from the same field experiment. Two have been 69 amended every two years since 1998, with a farmyard manure (FYM) or a co-compost of 70 sewage sludge and green waste (SGW), and the third one has never been amended 71 (CONT).We focused on two ATB widely used in human and veterinary medicine, 72 73 sulfamethoxazole (SMX) and ciprofloxacin (CIP), belonging respectively to sulfonamides and fluoroquinolones families. These molecules are among the most frequently detected ATB in 74 the environment (Barnes et al., 2004; Kolpin et al., 2002), and present contrasting behavior in 75 76 soils. SMX is relatively mobile and rapidly degraded with half-lives ranging from a few hours 77 to a few days (Salvia et al., 2014; Srinivasan and Sarmah, 2014), while CIP is known to have a high affinity to soils (Nowara et al., 1997; Vasudevan et al., 2009) and may persist for 78 79 several months to years (Hamscher et al., 2005; Rosendahl et al., 2012). However some studies suggested that sulfonamides may be more persistent than would be predicted from 80 laboratory controlled studies (Bialk et al., 2005). The different characteristics of CIP and 81 SMX make them good candidates to represent a range of antibiotics in the environment (Chen 82 et al., 2011). 83

We also considered the main metabolite of SMX, the N-acetyl-sulfamethoxazole (N-ac-SMX). When excreted, this metabolite accounts for approximately 50% of the administered SMX. But N-ac-SMX may undergo deacetylation in the environment and SMX can be regenerated (Göbel et al., 2005).

Thus the objectives of this study were to determine i) the fate of antibiotics with contrasted characteristics in soils and ii) how long-term applications of different OWP to soils impacted the fate of these antibiotics.

We monitored <sup>14</sup>C-labelled antibiotics during 6 months incubations by quantifying
radioactivity in the mineralized, extractable and non-extractable fractions, and by qualifying
radioactivity (parent molecules or metabolites) in the extractable fractions.

94

### 95 **2. Materials and methods**

## 96 2.1. Antibiotics

<sup>14</sup>C-ring-labeled sulfamethoxazole (chemical purity 98%, specific activity 77 mCi.mmol<sup>-1</sup>)
and <sup>14</sup>C-ring-labeled N-acetyl-sulfamethoxazole (chemical purity 93%, specific activity 77
mCi.mmol<sup>-1</sup>) were purchased from Isobio (ARC). <sup>14</sup>C-ring-labeled ciprofloxacin (chemical purity 98.4%, specific activity 15 mCi.mmol<sup>-1</sup>), was purchased from Moravek Biochemical.
The structures, pKa, and labelling of molecules are presented in Fig. S1.

102

### 103 2.2. Soil sampling

Soils were sampled in 2014 from the ploughed layers (0-25 cm) of the experimental field 104 QualiAgro (48°53' N, 1°58' E) located at Feucherolles (Yvelines, France): a soil control that 105 has never received OWP and 2 soils amended every 2 years since 1998 with a co-compost of 106 green waste and sewage sludge (SGW) or farmyard manure (FYM) (Houot et al., 2002). At 107 108 sampling, amended soils (SGW or FYM) have received 9 applications of OWP. The soil is a silt loam Glossic Luvisol and contains on average 19% clay, 75% silt and 6% sand in the 109 ploughed layer. Clay minerals identified are kaolinite, illite and smectite, with traces of 110 111 chlorite (Cornu, personal communication). The field is cropped with a biannual rotation winter wheat-maize. Selected properties of the soil samples and of the OWP applied in 2013 112 are listed in Table 1. The soils were homogenized, sieved through a 4-mm mesh and stored 3 113 weeks at 4°C in the dark before using. 114

## 116 **Table 1**

117 Physicochemical properties of soils (CONT, SGW, FYM) and OWP used as amendment in

118	2013
118	2013

		Soils		OWP			
	CONT	SGW	FYM	Compost sewage sludge-green waste	Farmyard manure		
Organic carbon (g.kg <sup>-1</sup> dw)	10.2	17.6	15.5	257.3	354.7		
Total nitrogen (g.kg <sup>-1</sup> dw)	1.0	1.7	1.4	25.5	20.1		
pH	6.7	6.8	7.3	7.7	8.8		
CEC (cmol+.kg <sup>-1</sup> )	7.6	9.8	9.9	-	-		
C/N	10.2	10.4	11.1	9.9	17.6		
$I_{ROC}^{a}$	-	-	-	78	67		
[SMX] (µg.kg <sup>-1</sup> soil dw) <sup>b</sup>	nd	nd	nd	nd	nd		
[CIP] (µg.kg <sup>-1</sup> soil dw) <sup>b</sup>	nd	$7.3 \pm 1.9$	nd	$309 \pm 22$	nd		

<sup>a</sup> Proportion of total organic carbon potentially incorporated into soil organic matter

120 (Lashermes et al., 2009).

<sup>b</sup> Concentrations of ATB were analysed in OWP of 2013 and soil sampled in November 2104

- 122 (Bourdat-Deschamps et al., 2017).
- 123 nd: not detected
- 124

## 125 2.3. Sorption Coefficient of SMX and N-ac-SMX

Sorption coefficients (Kd) of SMX and N-ac-SMX were determined on gamma-ray sterilized (40–70 kGy) samples of the 3 studied soils as follows: three replicates of 3 g equivalent dry soil were mixed with 10 ml of CaCl<sub>2</sub> 0.01 M solution sterilized by filtration at 0.2  $\mu$ m and 7.96  $\mu$ l of <sup>14</sup>C labelled SMX or 10.14  $\mu$ l of <sup>14</sup>C labelled N-ac-SMX (0.02  $\mu$ g.g<sup>-1</sup> soil), before shaking at 15 rotations.min<sup>-1</sup> for 24 h in the dark at 20°C. Samples were then centrifuged 10 min at 34155 g at 20°C and the <sup>14</sup>C remaining in the supernatants was quantified by liquid scintillation counting.

133 Then Kd (L.kg<sup>-1</sup>) was calculated as follows:

134 Kd = Cads/Ce

Cads was the concentration of ATB adsorbed on soil (mg.kg<sup>-1</sup>) and Ce, the concentration of
ATB in soil solution after equilibrium (mg.L<sup>-1</sup>).

137

138 2.4. Incubations

Ten grams (equivalent dry soil) of CONT, SGW and FYM soils were amended with 1 ml of solutions of <sup>14</sup>C-SMX, <sup>14</sup>C-N-ac-SMX or <sup>14</sup>C-CIP to provide final concentrations of 0.02, 0.02 or 0.15 µg.g<sup>-1</sup> soil respectively (1543 MBq.mM<sup>-1</sup> per sample) and a moisture content corresponding to 80% of maximum water holding capacity (0.22 g of water per g of soil). Six replicates per treatment were prepared. Spiked soils were incubated in 1 L airtight jars in presence of 2 vials containing respectively 10 ml of deionized water and 6 ml of sodium hydroxide 1N, at 28 °C for 156 days.

At 3 days and then weekly, <sup>14</sup>C-activity in the NaOH solutions was measured with a TriCarb liquid scintillation counter after mixing with 10 mL of UltimaGold liquid scintillation cocktail. Samples were kept for at least 24 h in the dark before counting to avoid potential chemiluminescence. The counting time was 10 min.

The opening of the jars to replace the NaOH traps ensured aerobic conditions during all incubation time. The soil moisture was checked over time and readjusted when necessary to 80% of WHC by adding deionized water.

153

## 154 2.5. Quantification of $^{14}C$ residues

At days 3 and 156, three successive extractions were performed to quantify the distribution of radioactivity from <sup>14</sup>C-SMX, <sup>14</sup>C-N-ac-SMX and <sup>14</sup>C-CIP in the easily and hardly extractable and the non-extractable fractions. The extraction protocol was adapted from Ferhi et al. (2016).

To measure the easily extractable fractions, 30 ml of cyclodextrin HPCD (0.05 mol.L<sup>-1</sup>) were 159 160 added to soil samples in 250 ml PPCO tubes. After shaking during 16 hours at 15 rotations.min<sup>-1</sup>, samples were centrifuged (20 min, 20°C, 34155 g). Supernatants were 161 weighted and mixed with 10 ml of Ultima Gold XR scintillation liquid (Perkin Elmer) for 162 counting. The next extraction was performed by adding acetonitrile MeCN (99.9% 163 Acetonitrile Carlo Erba Reagents Dasit Group, 27.5 ml) combined with a Mc Ilvaine pH9 164 165 buffer solution (9.3 mL of 1/29 (v/v) citric acid/phosphate). pH9 was reached with addition of NaOH 10 N. Samples were ultra-sonicated twice 10 min before centrifugation (20 min, 20°C, 166 34155 g). Radioactivity was quantified in the supernatants by liquid scintillation. The last 167 168 extraction was performed by addition of MeCN (99.9% Acetonitrile Carlo Erba Reagents Dasit Group, 27.5 ml) combined with a Mc Ilvaine pH2 buffer solution (11 mL of 27/3 (v/v) 169 citric acid/phosphate). pH2 was reached by adding HCl 37%. Samples were ultra-sonicated 170 171 twice for 10 min, before centrifugation (20 min, 20°C, 34155 g). Because of a salting out effect due to the high quantity of EDTA salt, two phases were obtained. Radioactivity was 172 quantified in the lower and upper phases by liquid scintillation. The hardly extractable 173 fraction was defined as the sum of pH9 and pH2 extracts. 174

After extractions, soil samples containing non-extractable <sup>14</sup>C-residues (NER) were air dried
at 35°C. Radioactivity from NER was measured by combustion (Biological Oxidizer OX 700,
Zinsser Analytic, Frankfurt, Germany) and trapping of the <sup>14</sup>CO<sub>2</sub> in Carbosorb and liquid
scintillation counting (Packard Instruments, Meriden, CT, USA).

179

## 180 2.6. Qualitative analysis of $^{14}C$ residues in extractable fractions

181 A qualitative analysis of <sup>14</sup>C in HPCD and MeCN extracts was carried out by high-182 performance liquid chromatography (HPLC) with a Waters (Milford,MA, USA) 183 chromatography appliance (Systemcontroller 610, Autosampler 717) coupled with a

radioactive flow detector (Flo-one A-500, Packard-Radiomatic) when radioactivity in 184 samples was greater than 250 Bq.mL<sup>-1</sup>. In certain cases, the 3 repetitions per extracts were 185 pooled to obtain sufficient radioactivity for analysis (Table S1). HPCD extracts were 186 concentrated on SPE cartridges (Strata X-A 33 µm Polymeric Stron Anion, Phenomenex) and 187 then with nitrogen before HPLC analyses. Radioactivity in HPCD extracts was too low after 188 156 days of incubation to be analyzed. MeCN-McIlvaine extracts were evaporated before 189 190 addition of 2 mL methanol. After centrifugation, samples were analyzed by HPLC. Radioactivity in the lower phase after MeCN-McIlvaine extraction was insufficient to be 191 analyzed. Samples analyzed by HPLC (Perkin Elmer: Flexar System Quaternary Pump, 192 Automatic Sample, PDA detector, Chromera® CDS piloting and acquisition Software; 193 Column: Waters X-Bridge C18 5µ / 4.6×250 mm; detector: Radio-HPLC FLO 194 ONE/Beta/Gamma C150TR; elution method: ACN + 0.01% TFA and H<sub>2</sub>O + 0.01% TFA) are 195 196 synthetized in Table S1.

197

## 198 2.7. Statistical analyses

Statistical analyses, using R software, were performed *via* ANOVA and Tukey tests when values were parametric, Kruskal and Dunn tests when values were non-parametric, to compare the fate of <sup>14</sup>C SMX, N-ac-SMX and CIP in the 3 studied soils at d3 and d156. Pvalues less than 0.05 were considered as statistically significant.

203

205 **3. Results** 

206 3.1. Fate of SMX and N-ac-SMX into soils amended or not with OWP

207 3.1.1. Kd of SMX and N-ac-SMX

208 Kd of SMX was lower in CONT than in SGW and intermediate in FYM (Table 2). Kd of N-

ac-SMX was significantly different in the three soils, in the following order:
SGW>CONT>FYM (Table 2).

211

### 212 **Table 2**

Sorption coefficients (Kd) of SMX and N-ac-SMX (L.kg<sup>-1</sup>) added at a concentration of 0.02  $\mu$ g.g<sup>-1</sup> soil in sterilized CONT, SGW and FYM soils (mean ± standard-deviation from 3 replicates). Different letters indicate statistical differences (P<0.05) between the three soils.

Kd (L.kg <sup>-1</sup> )	SMX		N-ac-SMX		SMX/N-ac-SMX
CONT	$2.58\pm0.11$	b	$1.21\pm0.03$	b	2.1
SGW	$3.20\pm0.10$	а	$1.33\pm0.00$	а	2.4
FYM	$2.95\pm0.07$	ab	$1.02\pm0.02$	c	2.9

216

Kd of SMX appeared highly correlated to the C content of soils ( $R^2=0.94$ ) in contrast to N-ac-SMX ( $R^2=0.008$ ).

In the three soils, Kd of SMX was two to three folds higher than Kd of N-ac-SMX. In all cases, at a concentration of  $0.02 \ \mu g.g^{-1}$ , Kd values were low, whatever the molecules and whatever the soils.

222

223 3.1.2. Fate of SMX

SMX mineralization started without latency in the 3 soils and reached 6.6, 8.9 and 10.5% of
added <sup>14</sup>C-SMX in the SGW, FYM and CONT soils respectively after 156 days (Fig. 1).
Although low in all cases, SMX mineralization was significantly greater in CONT than in
FYM and SGW.



**Fig. 1.** Cumulative <sup>14</sup>C-SMX mineralization in the control soil and in the soils that have received 9 applications of a co-compost of sewage sludge and green waste (SGW) or farmyard manure (FYM). Error bars indicate standard deviations of 3 replicates.

The total recovery of <sup>14</sup>C in the mineralized, extractable (HPCD and MeCN) and nonextractable <sup>14</sup>C fractions reached 87.4-88.8% of added <sup>14</sup>C-SMX after 3 days and 81.7-91% after 156 days. For interpretation, the results were expressed as percentages of recovered <sup>14</sup>C. The sum of all fractions corresponds therefore to 100% (Fig. 2).





Fig. 2. Distribution of <sup>14</sup>C-SMX in the different soil fractions (mineralized, HPCD and MeCN
extractable fractions, non-extractable residues), at (A) day 3, and (B) day 156, in the control
soil (CONT) and the soils amended with a sewage sludge-green waste compost (SGW) or
farmyard manure (FYM).

243

At d3 (Fig. 2A), <sup>14</sup>C was mainly distributed in the extractable and non-extractable fractions. SMX mineralization was negligible and varied between 1.1% (FYM) and 1.7% (CONT and SGW). Radioactivity in easily extractable (HPCD) fraction ranged between 17.9% (FYM) and 20.9% (SGW) of recovered <sup>14</sup>C. In hardly extractable (MeCN) fraction, <sup>14</sup>C ranged between 21.3% (FYM) and 27.3% (SGW). HPCD and MeCN extractable fractions were significantly lower in FYM. NER represented more than half of the recovered radioactivity as early as 3 days (Fig. 2A). They were significantly higher in FYM than in CONT and SGW.

Between 3 and 156 days, we observed mainly a decrease of the HPCD fraction (from about 20% to 6.1-7.7%) and an increase of the mineralized fraction. During the same period, mineralization was multiplied by a factor of about 6.5 and reached 7.4% (SGW), 11.0% (FYM) and 11.5% (CONT) of recovered <sup>14</sup>C (Fig. 2B).

The MeCN fraction slightly but significantly increased to reach 24.1-28.7%, whereas NER did not significantly vary between d3 and d156.

After 156 days, CONT presented the highest SMX mineralization and SGW the lowest mineralization and the highest HPCD extractable fraction. At last, FYM presented the lowest MeCN <sup>14</sup>C fraction.

260

261 3.1.3. Fate of N-ac-SMX

As for SMX, N-ac-SMX was immediately mineralized and cumulative <sup>14</sup>C-CO<sub>2</sub> ranged between 8.5 and 11.6% at d156 (Fig. 3). As for SMX, N-ac-SMX mineralization was higher in CONT than in amended soils and higher in FYM compared to SGW.



265

Fig. 3. Cumulative <sup>14</sup>C N-ac-SMX mineralization in the control soil (CONT) and the soils
amended with a sewage sludge-green waste compost (SGW) or farmyard manure (FYM).
Error bars indicate standard deviations of 3 replicates.

269

The total recovery of <sup>14</sup>C in the different fractions ranged from 71.7 to 82.2% of added <sup>14</sup>C Nac-SMX after 3 days and from 82.1 to 86.8% after 156 days. For interpretation, the results were expressed as percentages of recovered <sup>14</sup>C (Fig. 4).



Fig. 4. Distribution of <sup>14</sup>C-N-ac-SMX in the different soil fractions (mineralized, HPCD and MeCN extractable fractions, non-extractable residues), at (A) day 3, and (B) day 156, in the control soil (CONT) and the soils amended with a sewage sludge-green waste compost (SGW) or farmyard manure (FYM).

278

After 3 days, N-ac-SMX mineralization was low (2.1-3.0%) and followed the order
CONT≥FYM>SGW. The HPCD fractions represented 22.9-25.6% of recovered <sup>14</sup>C and were
significantly higher in SGW than in CONT, FYM presenting intermediate values. Hardly
(MeCN) extractable fractions were not significantly different in the 3 soils (34.4-37.3%).
NER varied between 35% (SGW) and 37.7% (FYM).

Between 3 and 156 days, mineralization increased by a 4.4-4.8 factor. NER increased to reach 49.7% (CONT), 55.9% (SGW) and 60.1% (FYM) of recovered <sup>14</sup>C. On the contrary, the HPCD and MeCN <sup>14</sup>C fractions decreased and ranged between 6.3-7.3% and 21.2-30.6% respectively. At d156, the HPCD fractions were not significantly different between the 3 soils. CONT presented the highest mineralized and MeCN fractions. FYM was characterized by a lower MeCN fraction and higher NER. The majority of <sup>14</sup>C was concentrated in the NER for the 3 soils.

291

292 3.1.4. Comparison between SMX and N-ac-SMX at d3 and d156

At d3, radioactivity in the mineralized, HPCD and MeCN extractable fractions was significantly lower for SMX than for N-ac-SMX (Table S2). The mineralization from <sup>14</sup>C-SMX was about half of that of N-ac-SMX. So, at d3, N-Ac-SMX was more mineralized and remained more extractable than SMX. At the opposite, addition of SMX produced more NER than N-ac-SMX. After 156 days, the HPCD fraction was significantly higher for SMX than N-ac-SMX except in CONT where it was similar. The MeCN fraction was similar for SMX

and N-ac-SMX in the three soils. NER were significantly higher in CONT after addition of
SMX while they were similar for SMX and N-ac-SMX in SGW and FYM. When comparing
fraction by fraction, differences between SMX and N-ac-SMX were similar for the 3 soils.

303 3.1.5. Qualitative analysis of  $^{14}$ C in extractable fractions

Three days after SMX addition, HPCD fractions that represented around 20% of recovered  $^{14}C$ , were mainly composed of  $^{14}C$ -SMX (71.3-80.5% of extractable  $^{14}C$ ). Four other metabolites including N-ac-SMX were detected but in smaller quantities. Metabolites differed among the 3 soils (Table 3).

In the MeCN pH9 fraction, we quantified no SMX but polar metabolites (M1, M2, M3) and an apolar metabolite (M9) at d3. Results were similar for the 3 soils. Between 3 and 156 days, metabolites evolved differently in the MeCN pH9 extracts. M2 and M3 disappeared while M9 and especially M1 increased. M1 (66.7-83.4% of extractable <sup>14</sup>C and 10.1-11.9% of total <sup>14</sup>C), the main metabolite and M9 were present in the 3 soils. M5 seemed specifically appear in the amended soils after 156 days.

SMX was present in the upper phase of MeCN pH2 extracts and represented 46.1 to 100% of extractable  ${}^{14}C$  (3.9 and 7.7% of total  ${}^{14}C$ ) in CONT and SGW respectively. SMX was predominant in this fraction for the 3 soils but metabolites (M6, M8, M9) were detected at smaller doses in one or another soil. CONT contained a greater diversity of metabolites than amended soils. In the MeCN pH2 extracts, SMX decreased over time but was still present at d156 in CONT and SGW. M8 and M9 disappeared and M4 and M6 remained the only quantifiable metabolites, representing 22.8-66% of extractable  ${}^{14}C$  (2.4-7% of total  ${}^{14}C$ ).

Three days after its addition, N-ac-SMX has almost completely disappeared in the HPCD fraction, subsisting only in CONT at about 12% of recovered <sup>14</sup>C, and was converted into SMX and other metabolites (M6 et M7). SMX was present in the 3 soils but in larger

quantities in the amended soils compared to CONT. CONT also presented a higher diversityof metabolites.

The metabolites present in the MeCN pH9 fraction 3 days after N-ac-SMX addition were the same in the 3 soils (M1, M2, M3, M9) with similar distributions to those obtained after SMX addition.

In the MeCN pH2 extracts at d3, N-ac-SMX subsisted around 12% of extractable <sup>14</sup>C in the 3
soils and was mostly transformed into SMX (70%-87%).

At d156, only the MeCN pH9 extracts contained a sufficient radioactivity and could be

analyzed by HPLC. Between d3 and d156, the polar metabolite M1 increased by a factor of

2.3 to 3.5 and represented 59-71% of extractable <sup>14</sup>C in the 3 soils. M2 and M3 decreased or

disappeared and M9 increased to reach around 20% of extractable <sup>14</sup>C in CONT and SGW. As

for SMX, M5 appeared only in amended soils. The evolution of the MeCN pH9 metabolites

336 was the same for SMX and N-ac-SMX (Table 3).

**338 Table 3** 

- 339 Qualitative analysis of <sup>14</sup>C by HPLC after addition of SMX or N-ac-SMX in the easily extractable fraction (HPCD), hardly extractable fraction at
- 340 pH9 (MeCN pH9) and hardly extractable fraction (upper phase) at pH2 (MeCN pH2) of CONT, SGW and FYM respectively after 3 (d3) and 156
- days (d156) of incubation: detection of SMX, N-ac-SMX and 9 other metabolites (M1-M9)
- 342 Results are expressed as % of extractable  ${}^{14}C$  or % of total  ${}^{14}C$ .
- 343 Extracts non analyzed by HPLC because of a too low radioactivity

			SMX						N-ac-SMX					
			CON	Т	SGW	Ι	FYN	1	CON	NT SGW		/ FYN		М
			Extractable <sup>14</sup> C (%)	Total <sup>14</sup> C (%)	Extractable <sup>14</sup> C (%)	Total <sup>14</sup> C (%)	Extractable <sup>14</sup> C (%)	Total <sup>14</sup> C (%)	Extractable <sup>14</sup> C (%)	Total <sup>14</sup> C (%)	Extractable <sup>14</sup> C (%)	Total <sup>14</sup> C (%)	Extractable <sup>14</sup> C (%)	r.
		M6 (24.4')	0	0	8.7	1.8	10.3	1.8	27.8	6.4	0	0	0	
		SMX (25.4')	74.7	15.6	71.3	14.9	80.5	14.4	36.1	8.3	67.9	17.3	100	
	UDCD	M7 (26.3')	18.6	3.9	13.2	2.8	0	0	24.2	5.5	32.1	8.2	0	
	прср	N-ac-SMX (27.8')	0	0	6.9	1.4	0	0	11.9	2.7	0	0	0	
		M9 (37.3')	6.7	1.4	0	0	9.2	1.7	0	0	0	0	0	
		Total	100	20.9	100	20.9	100	17.9	100	22.9	100	25.5	100	
	MeCN	M1 (3.7')	23.6	4.1	24.0	4.5	27.3	3.4	30.6	7.3	21.0	5.6	28.7	
		M2 (6.5')	37.8	6.6	27.1	5.0	20.9	2.6	20.1	4.8	20.0	5.4	17.6	
d3		M3 (9.8')	31.0	5.4	42.0	7.8	44.7	5.6	43.1	10.3	50.0	13.5	42.8	
	p119	M9 (37.3')	7.7	1.3	7.0	1.3	7.2	0.9	6.2	1.5	8.8	2.4	11.0	
		Total	100	17.4	100	18.6	100	12.5	100	23.8	100	26.9	100	
		M6 (24.4')	10.3	0.9	0	0	0	0	2.8	0.3	0	0	0	
		SMX (25.4')	46.1	3.9	100	7.7	76.7	5.2	74.7	6.9	87.4	8.2	69.9	
	MeCN	N-ac-SMX (27.8')	0	0	0	0	0	0	12.5	1.2	12.6	1.2	10.1	
	pH2	M8 (35.0)	16.2	1.4	0	0	23.3	1.6	0	0	0	0	0	
		M9 (37.3')	27.4	2.3	0	0	0	0	10.1	0.9	0	0	20.0	
		Total	100	8.4	100	7.7	100	6.8	100	9.2	100	9.4	100	
d156	MeCN	M1 (3.7')	66.7	10.1	67.1	11.9	69.6	11.6	71.2	13.1	61.0	9.4	58.8	

pH9	M2 (6.5')	12.4	1.9	0	0	0	0	7.8	1.4	0	0	12.9
	M5 (21.6')	0	0	16.2	2.9	17.0	2.8	0	0	19.5	3.0	22.3
	M9 (37.3')	20.9	3.2	16.7	3.0	13.5	2.3	21.0	3.9	19.6	3.0	6.0
	Total	100	15.2	100	17.8	100	16.7	100	18.4	100	15.4	100
	M4 (16.7')	22.8	2.4	0	0							
MeCN	M6 (24.4')	66.0	7.0	43.8	3.5	-	-	-	-	-	-	-
pH2	SMX (25.4')	11.2	1.2	56.2	4.4	-	-	-	-	-	-	-
	Total	100	10.6	100	7.9	-	-	-	-	-	-	-

### 346 *3.2. Fate of CIP into soils amended or not with OWP*

The <sup>14</sup>C balances ranged from 76 to 91.7% of added <sup>14</sup>C-CIP after 3 days and from 73.8 to 83.2% after 156 days. Presented results are expressed as percentages of recovered <sup>14</sup>C (Fig. 5).

350



**Fig. 5.** Distribution of <sup>14</sup>C-CIP in the different soil fractions (mineralized, HPCD and MeCN extractable fractions, non-extractable residues), at (A) day 3, and (B) day 156, in the control soil (CONT) and the soils amended with a sewage sludge-green waste compost (SGW) or farmyard manure (FYM).

356

We observed few evolutions between d3 and d156 in all of the <sup>14</sup>C fractions. CIP 357 mineralization was considered as negligible since % of mineralization (0.5-0.6%) were below 358 the impurity threshold (1.6%) in the 3 soils after 156 days. The radioactivity present in the 359 HPCD fraction represented only a few percent of recovered <sup>14</sup>C. At d156, radioactivity was 360 concentrated in MeCN fractions (20.4-25.9%) and mainly in NER (72.2-77.6%). CIP became 361 rapidly strongly adsorbed or non-extractable. When comparing the 3 soils, NER were 362 significantly higher in FYM while MeCN residues were higher in CONT. SGW presented 363 intermediate values. 364

### 366 **4. Discussion**

### 367 4.1. Fate of SMX and N-ac-SMX

After addition of <sup>14</sup>C-SMX and <sup>14</sup>C N-ac-SMX as a solution, the fraction of easily extractable 368 <sup>14</sup>C dropped in all soils till d3 and then decreased more slowly between 3 and 156 days. The 369 decrease with time of SMX and N-ac-SMX in the HPCD fraction can be due to i) sorption on 370 371 soil constituents, ii) physical sequestration in soil and/or iii) degradation. Then, radioactivity was quickly transferred to the hardly extractable fraction and NER. SMX and N-ac-SMX 372 sorption affinity to soils was relatively low as shown by the Kd values in the 3 soils (1-3.2 373 L.kg<sup>-1</sup>) that are in the same range of other studies (Drillia et al., 2005; Höltge and Kreuzig, 374 2007; Kodešová et al., 2015). Different interactions may be involved in sorption of 375 sulfonamides in soils, including surface complexation, ionic and hydrophobic interactions, 376 377 some of them being pH-dependent (Leal et al., 2013; Tolls, 2001; Gao et al., 2010). In this study, soil pH ranged between 6.7 and 7.3, above the pKas of SMX and N-ac-SMX (Fig. S1), 378 so that these molecules mainly existed in their anionic form, limiting ionic interactions with 379 clay minerals (Gao et al, 2005). But sorption also depends on hydrophobicity, and SMX and 380 381 its metabolites may thus interact with soil OM (Gao et al., 2010; Leal et al., 2013; Li et al., 382 2015). Thiele-Bruhn et al. (2004) highlighted the strong affinity between soil organic carbon and the sulfonamides. Furthermore, according to Martinez-Hernandez et al. (2016), sorption is 383 fast and can play a key role during the first 48h of soil incubations. Gulkowska et al. (2014) 384 385 attributed the fast formation of NER to chemical bonds between humic substances and the sulfonamide sulfamethazine. Thus sorption and chemical reactions of SMX and its 386 metabolites could partially explain the high increase of <sup>14</sup>C in the MeCN and non-extractable 387 fractions at d3, leading to unavailability of these compounds to soil microorganisms. This 388 hypothesis was also confirmed by HPLC analyses showing that after 3 days, SMX, N-ac-389

SMX and other metabolites were partially transferred to the MeCN fraction. Then 390 391 sulfonamides may desorb with time from this fraction to be transferred to readily extractable or non-extractable fractions (Drillia et al., 2005; Sukul et al., 2008). At the end of incubation, 392 NER represented more than 50% of recovered <sup>14</sup>C for SMX and N-ac-SMX. NER can have a 393 chemical origin with binding of ATB and their metabolites to soil constituents (Gulkowska et 394 al., 2014) but can also result from a physical sequestration of compounds or a microbial 395 396 transformation of ATB and subsequent incorporation into microbial biomass (Kästner et al., 397 2014).

Previous work showed that SMX and N-ac-SMX are mainly degraded by soil microorganisms 398 399 and that abiotic degradation is negligible (Acinnelli et al., 2007; Alvarino et al., 2016). Our 400 results showed that SMX and N-ac-SMX biodegradation could be complete (~ 10% of mineralization at d156) or incomplete with metabolite production as previously showed by 401 402 Liu et al. (2009) and Martinez-Hernandez et al. (2016). The production of metabolites, mainly more polar than parent compounds, started quickly after spiking soils with SMX and N-ac-403 SMX and confirmed results previously observed by Richardson and Ternes (2011) and Koba 404 et al. (2017). Few metabolites were identified but the degradation pathway of SMX and N-ac-405 406 SMX path remained unclear. The main metabolite of sulfonamides is their acetylated form 407 (Baran et al., 2011), whose formation can be reversible (Höltge & Kreuzig, 2007). Metabolites may also vary according to microbial species (Larcher and Yargeau, 2012) and 408 environmental conditions (Müller et al., 2013). 409

410 Mineralization was significantly greater for N-ac-SMX than for SMX in the first days of 411 incubation. These results are consistent with the lower adsorption and the higher extractability 412 of N-ac-SMX in soil, leading to a higher availability of N-ac-SMX to soil microorganisms. At 413 the opposite, the higher Kd of SMX can partially explain the higher production of abiotic 414 NER. Höltge and Kreuzig (2007) previously observed higher Kd for SMX compared to N-ac-

SMX in most of soils and OWP they studied. Considering HPLC analyses, N-ac-SMX has almost completely disappeared from the HPCD extracts after 3 days and was present only in the MeCN pH2 fractions. The quantification of SMX as the most abundant molecule in the HPCD and MeCN pH2 extracts after N-ac-SMX addition in the 3 soils confirmed the reversible transformation of N-Ac-SMX to SMX (Höltge and Kreuzig, 2007; Zarfl et al., 2009).

421 The fate of SMX and N-ac-SMX also differs in the second part of incubation. For SMX, the production of MeCN and non-extractable residues was very fast up to 3 days and then evolved 422 little between d3 and d156. For N-ac-SMX, hardly extracted residues increased till 3 days 423 424 before decreasing. The production of NER was more progressive with N-ac-SMX than for SMX and was explained by a decrease of easily and hardly extractable fractions. Zarfl et al. 425 (2009) suggested by simulation that NER were formed directly from easily extractable 426 427 residues after sulfonamide addition. This would mean that some of the hardly extractable residues would go back to soil solution before contributing to the formation of NER. 428

Even if evolution with time was different for the 2 compounds, the HPCD, MeCN and non-429 extractable fractions were very similar at d156. For SMX and N-ac-SMX, radioactivity was 430 mainly concentrated in NER and represented more than 50% of recovered <sup>14</sup>C. Besides, Zarfl 431 432 et al. (2009) also showed that the formation of NER from an acetylated metabolite of sulfadiazine was negligible. Therefore, deacetylation of N-ac-SMX to reform the parent 433 compound should be necessary before its transfer to NER. Thus the transformation of N-SMX 434 435 into SMX leads at long term to a similar distribution of radioactivity in extractable and nonextractable fractions for N-ac-SMX and SMX. On the other hand, the mineralization of the 2 436 molecules started from the first days of incubation and explains why in this case, differences 437 persisted over time. 438

440 *4.2. Fate of CIP* 

The fate of the fluoroquinolone CIP is very different from sulfonamides. No significant mineralization (less than 1%) was detected after 156 days. Except in artificial conditions, e.g., pure cultures with optimal growth conditions, liquid media, bioreactor (Dorival-Garcia et al., 2013) or specific conditions with microorganisms from drinking water biofilter and adapted to high concentrations in CIP (Liao et al., 2016), most of the studies concluded that ciprofloxacin was recalcitrant to biodegradation (Girardi et al., 2011, Jechalke et al., 2014; Goulas et al., 2016).

The low CIP mineralization is due to its strong sorption and its resulting low bioavailability. 448 449 Kd of CIP was not measured in this study, but the literature reports very high values of Kd, > 500 kg.L<sup>-1</sup> (Leal et al., 2013; Vasudevan et al., 2009). Different mechanisms such as cation 450 exchange, cation bridging, complexation, and H-binding may be responsible for sorption of 451 452 fluoroquinolones in soils (Tolls, 2001; Vasudevan et al., 2009). At soil pH, zwitterionic species predominate in the soil solution. Hence a strong attraction between positive charges of 453 CIP and negative charges of soil constituents can explain fast and strong CIP adsorption in 454 soil. Drillia et al. (2005) measured very high desorption coefficients of fluoroquinolones, 455 meaning that they mainly remained sorbed in the soil. Furthermore CIP addition in soil 456 457 induces a high production of NER (Girardi et al., 2011). The decline of extractable CIP is thus mainly governed by sorption and formation of NER. In our study, from 3 days and throughout 458 the incubation time, the radioactivity was concentrated in the hardly extractable fraction 459 460 (20.4-25.9% at d156) and even more in NER (72.2-77.6% at d156). Girardi et al. (2011) comparing CIP degradation in biotic and abiotic conditions concluded that NER were mainly 461 from abiotic origin. 462

But no mineralization does not mean no biodegradation. Due to the low radioactivity in the extracts, HPLC analyses could not be performed in this work. But other studies highlighted a

possible incomplete degradation and the appearance of metabolites (Girardi et al., 2011),
especially in presence of fungi (Parshikov et al., 1999; Wetzstein et al., 1999).

467

## 468 4.3. Influence of soil properties on the fate of antibiotics

The use of soils amended or not with OWP had for objective to study how long term applications of different OWP to soils impacted the fate of ATB. Two hypotheses were tested: - OWP repeated application may increase ATB biodegradation (Accinelli et al., 2007) stimulating the soil indigenous microorganisms, introducing exogenous degrading microorganisms in soils or promoting the development of an adapted microflora due to the presence of antibiotics in OWP (Topp et al., 2013)

- OWP repeated application may enhance antibiotics sorption limiting their bioavailability and
decreasing their mineralization (Li et al., 2015).

We found SMX and N-ac-SMX mineralization lower in OWP amended soils compared to
CONT. Li et al (2015) previously observed that the addition of biosolids and compost to soil
decreased SMX mineralization but the level of inhibition depended on the quantity of OM.

When comparing results of HPLC analyses between amended and unamended soils after 480 SMX addition, the differences mainly appeared in the MeCN pH2 fraction, with less SMX 481 482 and more metabolites present in CONT, as if the biodegradation was more advanced while remaining incomplete. These effects appeared at d3 and persisted over time. In soils incubated 483 with N-ac-SMX, the main effect of amendments appeared in HPCD fractions, where organic 484 485 matter seemed to have accelerated the de-acetylation into SMX and inhibited the formation of one metabolite (M6). The M5 metabolite present at d156 after addition of SMX and N-ac-486 SMX seemed specifically produced in amended soils. 487

On another side, differences were observed between SGW and FYM, i.e., SMX and N-acSMX mineralization was more inhibited in SGW than in FYM. HPLC analyses also showed

that the proportion of easily extractable SMX was higher in FYM while strongly adsorbed SMX was higher in SGW. These results are consistent with soil organic carbon contents higher in SGW than in FYM (17.6 vs 15.5 g.kg<sup>-1</sup>) and values of Kd higher in SGW than in FYM for both SMX and N-ac-SMX. Furthermore, Racke and Lichtenstein (1985) reported that the quality of OM and its degree of decomposition were also key factors. OM is more stable and less degradable in compost of sewage sludge and green waste than in manure.

496 Finally organic amendments stimulated the production of NER whatever the studied antibiotic (SMX, N-ac-SMX, CIP). The increase of NER occurred very quickly with SMX and CIP and 497 later with N-ac-SMX. In addition, this effect was more pronounced in FYM than in SGW. Li 498 499 et al. (2015) also showed that OWP amendment increased the formation and accumulation of NER in a soil spiked with SMX from the first days of incubation. They suggested that OWP 500 provide additional sites facilitating binding or conjugation interactions between contaminants 501 502 and soil constituents and/or promoted microbial transformation leading to the formation of intermediates that were readily bound to the matrix. 503

504

## 505 5. Conclusion

The fate of two sulfonamides, SMX and N-ac-SMX, and one fluoroquinolone, CIP, in soils amended or not with manure or a co-compost of sludge and green waste depends on molecules and soil characteristics. Antibiotic mineralization was nil or low after 156 days (CIP << SMX < N-ac-SMX) while the production of NER accounted for more than 50% of recovered  $^{14}C$ .

511 Organic amendments decrease antibiotics degradation and mineralization (SMX and N-ac-512 SMX) and increase the production of NER (SMX, N-ac-SMX and CIP). The effects also 513 depend on the quantity and quality of added OM. The SGW compost further inhibits 514 mineralization while FYM enhances the production of NER. Land application of OWP is an

increasing practice in agriculture and urban gardening and so the impact of OWP on the fate 515 516 of ATB and its associated health and environmental risks are of great interest to be studied. But OWP often contain multiple contaminants (other xenobiotics, metals ...), as well as a 517 complex organic matrix that interacts with the contaminants during storage or treatment 518 (composting, methanisation, liming, drying...), and then after soil spreading. Consequently it 519 would be interesting to evaluate whether the concomitant addition of contaminants (cocktail 520 521 effects) in soils and/or the evolution of OM-ATB interactions over time have an impact on the fate of antibiotics. 522

523

### 524 6. Acknowledgements

Acknowledgements are due to ADEME and INRA, Department of Environment and Agronomy, for the PhD grant of the first author, and to the French National Research Agency who financed the project CEMABS (ANR-13-CESA-0016-01). Soil samples came from the QualiAgro field experiment supported by Veolia Research and Innovation and by AnaEE-France. We also thank the staff of ECOSYS who helped us during sampling, experiments and data treatments.

531

### 532 7. Bibliographie

533 Accinelli, C., Koskinen, W.C., Becker, J.M., Sadowsky, M.J. (2007). Environmental Fate of

Two Sulfonamide Antimicrobial Agents in Soil. J. Agric. Food. Chem., 55, 2677–2682.
http://doi.org/10.1021/jf063709j

Alvarino, T., Nastold, P., Suarez, S., Omil, F., Corvini, P.F.X., Bouju, H. (2016). Role of

537 biotransformation , sorption and mineralization of 14 C-labelled sulfamethoxazole under

different redox conditions. Science of the Total Environment, 542, 706–715.

539 http://doi.org/10.1016/j.scitotenv.2015.10.140

- 540 Baran, W., Adamek, E., Ziemiańska, J., Sobczak, A. (2011). Effects of the presence of
- sulfonamides in the environment and their influence on human health. J. Hazard. Mater.,196, 1-15.
- Barnes, K.K., Christenson, S.C., Kolpin D.W., Focazio, M., Furlong, E.T., Zaugg, S.D.,
  Meyer, M.T., Barber, L.B. (2004). Pharmaceuticals and other organic waste water
  contaminants within a leachate plume downgradient of a municipal landfill. Ground
  Water Monit. Rem., 24 (2), 119–126.
- 547 Bialk, H.M., Simpson A.J., Pedersen, J.A. (2005). Cross-Coupling of Sulfonamide
  548 Antimicrobial Agents with Model Humic Constituents. Env. Sci. Technol., 39(12),
  549 4463–4473. doi 10.1021/es0500916
- Bourdat-Deschamps, M., Ferhi, S., Bernet, N., Feder, F., Crouzet, O., Patureau, P.,
  Montenach, D., Moussard, G.D., Mercier, V., Benoit, P., Houot, S. (2017). Fate and
  impacts of pharmaceuticals and personal care products after repeated applications of
  organic waste products in long-term field experiments. Science of the Total Environment
  607-608, 271-280.
- Boxall, A.B.A., Kay, P., Blackwell, P.A., Fogg, L.A., (2004). Fate of veterinary medicines
  applied to soils. In: Kümmerer, K. (Ed.), Pharmaceuticals in the Environment: Sources,
  Fate, Effects and Risks. Springer-Verlag, Berlin, pp. 165–180.
- 558 Boxall, A.B.A., Rudd, M.A., Brooks, B.W., Caldwell, D.J., Choi, K., Hickmann, S., Innes, E.,
- Ostapyk, K., Staveley, J., P., Beazley, K.F. (2012). Review Pharmaceuticals and
  Personal Care Products in the Environment: What Are the Big Questions?. Environ.
  Health Perspect., 120(9), 1221–1229. doi 10.1289/ehp.1104477
- Chang, A.C., Pan, G., Page, A.L., Asano, T. (1995). Developing Human Health-related
  Chemical Guidelines for Reclaimed Waster and Sewage Sludge Applications in
  Agriculture. World Health Organization, Geneva, 1995., WHO/EOS/95.20.

565	Chen, H., Gao, B., Li, H., Ma, L.Q. (2011). Effects of pH and ionic strength on
566	sulfamethoxazole and ciprofloxacin transport in saturated porous media. J. Contam.
567	Hydrol., 126(1–2), 29–36. http://doi.org/10.1016/j.jconhyd.2011.06.002

- 568 Dorival-García, N., Zafra-Gómez, A., Navalón, A., González, J., Hontoria, E., Vílchez, J.L.
- 569 (2013). Removal of quinolone antibiotics from wastewaters by sorption and biological
- 570 degradation in laboratory-scale membrane bioreactors. Sci. Total Environ., 442, 317–

571 328. http://doi.org/10.1016/j.scitotenv.2012.10.026

- 572 Drillia, P., Stamatelatou, K., Lyberatos, G. (2005). Fate and mobility of pharmaceuticals in
  573 solid matrices. Chemosphere, 60, 1034–1044.
  574 http://doi.org/10.1016/j.chemosphere.2005.01.032
- Du, L., Liu, W. (2012). Occurrence, fate, and ecotoxicity of antibiotics in agro-ecosystems. A
  review. Agron. Sustain. Dev., 32, 309–327. http://doi.org/10.1007/s13593-011-0062-9
- Ferhi, S., Bourdat-Deschamps, M., Daudin, J.J., Houot, S., Nélieu, S. (2016). Factors
  influencing the extraction of pharmaceuticals from sewage sludge and soil: an
  experimental design approach. Anal Bioanal Chem., 408, 6153–6168.
- Gao, J., Pedersen, J. (2005). Adsorption of Sulfonamide Antimicrobial Agents to Clay
  Minerals. Environ. Sci. Technol., 39, 9509-9516.
- Gao, J., Pedersen, J. (2010). Sorption of Sulfonamide Antimicrobial Agents to Humic Acid –
  Clay Complexes. J. Environ. Qual., 39, 228–235. doi:10.2134/jeq2008.0274

Girardi, C., Greve, J., Lamshöft, M., Fetzer, I., Miltner, A., Schäffer, A., Kästner, M. (2011).
Biodegradation of ciprofloxacin in water and soil and its effects on the microbial
communities. J. Hazard. Mater., 198, 22–30.
http://doi.org/10.1016/j.jhazmat.2011.10.004

Göbel, A., Thomsen, A., McArdell, C.S., Joss, A., Giger, W. (2005). Occurrence and sorption
behavior of sulfonamides, macrolides, and trimethoprim in activated sludge treatment.

- 590 Env. Sci. Technol., 39, 3981–3989. http://doi.org/10.1021/es048550a
- Golet, E.M., Strehler, A., Alder, A.C., Giger, W. (2002). Determination of fluoroquinolone
  antibacterial agents in sewage sludge and sludge-treated soil using accelerated solvent
  extraction followed by solid-phase extraction. Anal. Chem., 74(21), 5455–5462.
- Goulas, A., Haudin, C., Bourdat-Deschamps, M., Benoit, P., Ferhi, S. (2016). A new
  extraction method to assess the environmental availability of ciprofloxacin in agricultural
  soils amended with exogenous organic matter. Chemosphere, 165, 460–469.
  doi.org/10.1016/j.chemosphere.2016.09.040
- Gulkowska, A., Thalmann, B., Hollender, J., Krauss, M. (2014). Nonextractable residue
  formation of sulfonamide antimicrobials: New insights from soil incubation experiments.
  Chemosphere 107, 366–372.
- Hamscher, G., Pawelzick, H.T., Höper, H., Nau, H. (2005). Different behaviour of
  tetracyclines and sulfonamides in sandy soils after repeated fertilization with liquid
  manure. Environ. Toxicol. Chem., 24, 861–8.
- Höltge, S., Kreuzig, R. (2007). Laboratory testing of sulfamethoxazole and its metabolite
  acetyl-sulfamethoxazole in soil. Clean Soil Air Water, 35(1), 104–110.
  doi.org/10.1002/clen.200600019
- Houot, S., Clergeot, D., Michelin, J., Francou, C., Bourgeois, S., Caria, G., Ciesielski, H.
  (2002). Agronomic value and environmental impacts of urban composts used in
  agriculture. In: Insam, H., Klammer, S. (Eds.), Microbiology of Composting. SpringerVerlag, Berlin Heidelberg, pp. 457-472
- Jechalke, S., Heuer, H., Siemens, J., Amelung, W., Smalla, K. (2014). Fate and effects of
  veterinary antibiotics in soil. Trends Microbiol., 22(9), 536–545.
  http://doi.org/10.1016/j.tim.2014.05.005.
- Kästner, M., Nowak, K.M.; Miltner, A., Trapp, S., Schaffer, A. (2014) Classification and

- Modelling of Non extractable Residue (NER) Formation of Xenobiotics in Soil A
  Synthesis. Crit. Rev. Env. Sci. Technol., 44(19), 2107-2171.
- Koba, O., Golovko, O., Kodešová, R., Fér, M., Grabic, R. (2017). Antibiotics degradation in
  soil: A case of clindamycin, trimethoprim, sulfamethoxazole and their transformation
  products. Environ. Pollut., 220, 1251-1263.
- Kodešová, R., Grabic, R., Kočárek, M., Klement, A., Golovko, O., Fér, M., Nikodem, A.,
  Jakšík. O. (2015). Pharmaceuticals' sorptions relative to properties of thirteen different
- 622 soils. Sci. Total Environ., 511, 435-443. doi: 10.1016/j.scitotenv.2014.12.088
- 623 Kolpin, D.W., Furlong, E.T., Meyer, M.T., Thurman, E.M., Zaugg., S.D., Barber, L.B.,
- Buxton, H.T. (2002). Pharmaceuticals, Hormones, and Other Organic Wastewater
  Contaminants in U.S. Streams, 1999 2000: A National Reconnaissance. Environ. Sci.
  Technol., 36(6), 1202–1211.
- Kümmerer, K. (2001). Drugs in the environment: emission of drugs, diagnostic aids and
  disinfectants into wastewater by hospitals in relation to other sources a review.
  Chemosphere, 45,957–969.
- Larcher, S and Yargeau, V. (2012). Biodegradation of sulfamethoxazole: current knowledge
  and perspectives. Appl Microbiol Biotechnol., 96:309-318.
- 632 Lashermes, G., Nicolardot, B., Parnaudeau, V., Thuriès, L., Chaussod, R., Guillotin, M.L.,
- 633 Linères, M., Mary, B., Metzger, L., Morvan, T., Tricaud, A., Vilette, C., Houot, S.
- 634 (2009). Indicator of Potential Residual Carbon in Soils after Exogenous Organic Matter
- Application. European Journal of Soil Science., 60 (2), 297–310.
- Leal, R.M.P., Alleoni, L.R.F., Tornisielo, V.L., Regitano, J.B. (2013). Sorption of
  fluoroquinolones and sulfonamides in 13 Brazilian soils. Chemosphere, 92(8), 979–985.
- 638 http://doi.org/10.1016/j.chemosphere.2013.03.018
- 639 Li, J., Ye, Q., Gan, J. (2015). Influence of organic amendment on fate of acetaminophen and

640 sulfamethoxazole in soil. Environ. Pollut., 206, 543–550.
641 http://doi.org/10.1016/j.envpol.2015.08.019

- Liao, X., Li, B., Zou, R., Dai, Y., Xie, S., Yuan, B. (2016). Biodegradation of antibiotic
  ciprofloxacin: pathways, influential factors, and bacterial community structure. Environ.
  Sci. Pollut. Res. Int., 23(8), 7911–7918. http://doi.org/10.1007/s11356-016-6054-1
- Liu, F., Ying, G.G., Tao, R., Zhao, J.L., Yang, J.F., Zhao, L.F. (2009). Effects of six selected
- antibiotics on plant growth and soil microbial and enzymatic activities. Environ. Pollut.,
  157(5), 1636–1642. http://doi.org/10.1016/j.envpol.2008.12.021
- 648 Martínez-Hernández, V., Meffe, R., Herrera, S., Bustamante, I.D. (2016). The role of sorption
- and biodegradation in the removal of acetaminophen, carbamazepine, caffeine, naproxen
- and sulfamethoxazole during soil contact: A kinetics study. Sci. Total Envir., 559, 232–

651 241. http://doi.org/10.1016/j.scitotenv.2016.03.131

- Müller, M., Schüssler, W., Horn H., Lemmer, H. (2013). Aerobic biodegradation of the
  sulfonamide antibiotic sulfamethoxazole by activated sludge applied as co-substrate and
  sole carbon and nitrogen source. Chemosphere 92, 969–978.
- Nieto A., Borruli F., Pocurull E., Marce R.M., (2010). Ocurence of pharmaceuticals and
  hormones in sewage sludge. Environ Toxicol Chem., 29,1484-1489.
- Nowara, A., Burhenne, J., Spiteller, M. (1997). Binding of Fluoroquinolone Carboxylic Acid
  Derivatives to Clay Minerals. J. Agric. Food. Chem., 45, 1459–1463.
  http://doi.org/10.1021/jf9602151
- 660 Ozaki, N., Bester, K., Moldrup, P., Henriksen, K., Komatsu, T. (2011). Photodegradation of
- the synthetic fragrance OTNE and the bactericide triclosan adsorbed on dried loamy sand
- Results from models and experiments. Chemosphere, 83(11), 1475–1479.
- 663 http://doi.org/10.1016/j.chemosphere.2011.03.006
- Parshikov, I.A., Freeman, J.P., Lay, J.O., Beger, R.D., Williams, A.J., Sutherland, J.B. (1999).

- Regioselective transformation of ciprofloxacin to N -acetylciprofloxacin by the fungus *Mucor ramannianus*. FEMS Microbiol. Lett., 177, 131–135.
- Racke, K.D., Lichtenstein, E.P. (1985). Effects of soil microorganisms on the release of
  bound 14C residues from soils previously treated with [14C] parathion. J. Agric. Food
  Chem., 33, 938-943.
- Richardson, S.D., Ternes, T.A. (2011). Water analysis: emerging contaminants and current
  issues. Anal. Chem., 83, 4614-4648.
- Rosendahl, I., Siemens, J., Kindler, R., Groeneweg, J., Zimmermann, J., Czerwinski, S.,
  Lamshöft, M., Wilke, B.M., Vereeken, H., Amelung, W. (2012). Persistence of the
  Fluoroquinolone Antibiotic Difloxacin in Soil and Lacking Effects on Nitrogen
  Turnover. J. Environ. Qual., 41, 1275-1283. http://doi.org/10.2134/jeq2011.0459
- Salvia, M.-V., Experton, J., Geandel, C., Cren-Olivé, C., Vulliet, E. (2014). Fate of
  pharmaceutical compounds and steroid hormones in soil: study of transfer and
  degradation in soil columns, Environ. Sci. Pollut. Res., 21, 10525–10535.
  http://doi.org/10.1007/s11356-014-3038-x
- Sarmah, A. K., Meyer, M. T., Boxall, A.B. (2006). A global perspective on the use, sales,
  exposure pathways, occurrence, fate and effects of veterinary antibiotics (VAs ) in the
  environment. Chemosphere, 65, 725–759.
- 683 http://doi.org/10.1016/j.chemosphere.2006.03.026
- Srinivasan, P., Sarmah, A.K. (2014). Assessing the sorption and leaching behaviour of three
  sulfonamides in pasture soils through batch and column studies. Sci. Total Environ., 493,
  535–543. http://doi.org/10.1016/j.scitotenv.2014.06.034
- Sukul, P., Lamshöft, M., Zühlke, S., Spiteller, M. (2008). Sorption and desorption of
  sulfadiazine in soil and soil-manure systems. Chemosphere, 73(8), 1344–1350.
  http://doi.org/10.1016/j.chemosphere.2008.06.066

- Thiele-Bruhn, S., Seibicke, T., Schulten, H.R., Leinweber P. (2004). Sorption of sulfonamide
  pharmaceutical antibiotics on whole soils and particle-size fractions. J. Environ. Qual.,
  33, 1331-1342.
- Tolls, J. (2001). Sorption of Veterinary Pharmaceuticals in Soils: A Review. Envir. Sci.
  Technol., 35(17), 3397–3406.
- Topp, E., Chapman, R., Devers-Lamrani, M., Hartmann, A., Marti, R., Martin-Laurent, F., 695 696 Sabourin, L., Scott, A., Sumarah, M. (2013). Accelerated Biodegradation of Veterinary Antibiotics in Agricultural Soil following Long-Term Exposure, and Isolation of a 697 J. Sulfamethazine-degrading Environ. Qual., 42(1), 173–178. 698 sp. 699 http://doi.org/10.2134/jeq2012.0162
- Vasudevan, D., Bruland, G.L., Torrance, B.S., Upchurch, V.G., MacKay, A.A. (2009). pHdependent ciprofloxacin sorption to soils: Interaction mechanisms and soil factors
  influencing sorption. Geoderma, 151, 68–76.
  http://doi.org/10.1016/j.geoderma.2009.03.007
- Wetzstein, H.G., Stadler, M., Tichy, H.V., Dalhoff, A., Karl, W. (1999). Degradation of
  Ciprofloxacin by Basidiomycetes and Identification of Metabolites Generated by the
  Brown Rot Fungus *Gloeophyllum striatum*. Appl. Environ. Microbiol., 65(4), 1556–
  1563.
- Zarfl, C., Klasmeier, J., Matthies, M. (2009). A conceptual model describing the fate of
   sulfadiazine and its metabolites observed in manure-amended soils. Chemosphere, 77(6),
- 710 720–726. http://doi.org/10.1016/j.chemosphere.2009.08.035
- Zhang, Y., Hu, S., Zhang, H., Shen, G., Yuan, Z., & Zhang, W. (2017). Degradation kinetics
  and mechanism of sulfadiazine and sulfamethoxazole in an agricultural soil system with
  manure application. Science of the Total Environment., 607–608, 1348–1356.
  http://doi.org/10.1016/j.scitotenv.2017.07.08

# 718 Appendix A. Supplementary data

719

720 Fig. S1. Molecular structure, pKa and labelling of sulfamethoxazole, N-acetyl-

sulfamethoxazole and ciprofloxacin (\*  $^{14}$ C labelling)



# **Table S1**

## 727 Selected extracts for qualitative analyses of radioactivity by HPLC

HPCD extractionsMeCN pH9 extractionsMeCN pH2 uper phase extractionsMoleculeSoilDayConcentrationPoolConcentrationPoolConcentrationPoolN-ac-SMX $\begin{array}{c} CONT \\ FYM \end{array}$ 3 $\begin{array}{c} SPE \ cartridge strata \\ XA \ type \end{array}$ noEvaporationnoEvaporationpoolN-ac-SMX $\begin{array}{c} SGW \\ FYM \end{array}$ 156 $\begin{array}{c} FVE \ cartridge strata \\ XA \ type \end{array}$ noEvaporationnoEvaporationyesN-ac-SMX $\begin{array}{c} SGW \\ FYM \end{array}$ 156 $\begin{array}{c} FVE \ cartridge strata \\ XA \ type \end{array}$ noEvaporationnoEvaporationyesSMX $\begin{array}{c} CONT \\ SGW \\ FYM \end{array}$ 3 $\begin{array}{c} SPE \ cartridge strata \\ XA \ type \end{array}$ noEvaporationnoEvaporationyesSMX $\begin{array}{c} SGW \\ SGW \\ FYM \end{array}$ 156 $\begin{array}{c} FVE \ cartridge strata \\ XA \ type \end{array}$ noEvaporationnoEvaporationyesSMX $\begin{array}{c} SGW \\ SGW \\ FYM \end{array}$ 156 $\begin{array}{c} FVE \ cartridge strata \\ XA \ type \end{array}$ noEvaporationnoEvaporationyesSMX $\begin{array}{c} SGW \\ SGW \\ FYM \end{array}$ 156 $\begin{array}{c} FVE \ cartridge strata \\ XA \ type \end{array}$ noEvaporationnoEvaporationyesSMX $\begin{array}{c} SGW \\ SGW \\ FYM \end{array}$ 156 $\begin{array}{c} FVE \ cartridge strata \\ XA \ type \end{array}$ noEvaporationnoEvaporationyes									
MoleculeSoilDayConcentrationPoolConcentrationPoolConcentrationPoolN-ac-SMXCONT FYM3SPE cartridge strata XA typenoEvaporationnoEvaporationnoN-ac-SMXCONT FYM156SPE cartridge strata XA typenoEvaporationnoEvaporationyesN-ac-SMXSGW FYM156SPE cartridge strata XA typenoEvaporationnoEvaporationyesSMXSGW FYM3SPE cartridge strata XA typenoEvaporationnoEvaporationyesSMXSGW FYM156SPE cartridge strata XA typenoEvaporationnoEvaporationyesSMXSGW FYM156SPE cartridge strata XA typenoEvaporationnoEvaporationyes				HPCD extractio	ns	MeCN pH9 ext	ractions	MeCN pH2 upper phase extractions	
N-ac-SMXCONT SGW FYM3SPE cartridge strata XA typenoEvaporationnoEvaporationyesN-ac-SMXCONT FYM156EvaporationnoEvaporationnoEvaporationyesSMXSGW FYM156SPE cartridge strata XA typenoEvaporationnoEvaporationyesSMXCONT FYM3SPE cartridge strata XA typenoEvaporationnoEvaporationyesSMXSGW FYM156SPE cartridge strata XA typenoEvaporationnoEvaporationyes	Molecule	Soil	Day	Concentration	Pool	Concentration	Pool	Concentration	Pool
N-ac-SMXCONT SGW FYM156EvaporationnoEvaporationyesSMXCONT SGW FYM3SPE cartridge strata XA typenoEvaporationnoEvaporationyesSMXSGW FYM3SPE cartridge strata XA typenoEvaporationnoEvaporationyesSMXSGW FYM156EvaporationnoEvaporationyes	N-ac-SMX	CONT SGW FYM	3	SPE cartridge strata XA type	no	Evaporation	no	Evaporation	yes
SMXCONT SGW FYM3SPE cartridge strata XA typenoEvaporationnoEvaporationyesSMXCONT SGW FYMSGW156EvaporationnoEvaporationyes	N-ac-SMX	CONT SGW FYM	156			Evaporation	no	Evaporation	yes
CONT SMX SGW 156 Evaporation no Evaporation yes FYM	SMX	CONT SGW FYM	3	SPE cartridge strata XA type	no	Evaporation	no	Evaporation	yes
	SMX	CONT SGW FYM	156			Evaporation	no	Evaporation	yes

**Table S2.** Ratio between percentages of <sup>14</sup>C-SMX and <sup>14</sup>C- N-ac-SMX in the mineralized,

HPCD, MeCN and non-extractable fractions at day 3 and day 156.

\* means that the percentages of radioactivity in the considered fraction are significantly

733 different for SMX and N-ac-SMX.

	SMX/N-ac-SMX											
	Miner frac	alized tion	HPCD e frac	xtractable	MeCN of fra	extractable action	Non extractable fraction					
	d3	d156	d3	d156	d3	d156	d3	d156				
CONT	0.6 *	0.8 *	0.9 *	0.9	0.7 *	0.9	1.4 *	1.1 *				
SGW	0.5 *	0.7 *	0.8 *	1.1 *	0.7 *	1.0	1.4 *	1.0				
FYM	0.6 *	0.9	0.7 *	1.1 *	0.6 *	1.1	1.7 *	1.0				