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▶ To cite this version:

Fares Gouzi, Christian Prefaut, Aldjia Abdellaoui, Emilie Roudier, Philippe de Rigal, et al.. Blunted muscle angiogenic training-response in COPD patients versus sedentary controls. European Respiratory Journal, 2013, 41 (4), pp.806-814. 10.1183/09031936.00053512. hal-01624893

HAL Id: hal-01624893

https://hal.science/hal-01624893

Submitted on 15 Aug 2018

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TITLE:

BLUNTED MUSCLE ANGIOGENIC TRAINING-RESPONSE IN COPD PATIENTS

VERSUS SEDENTARY CONTROLS

AUTHORS

Fares Gouzi^{1,2}; Christian Préfaut¹; Aldjia Abdellaoui^{1,3}; Emilie Roudier⁴; Philippe de Rigal²;

Nicolas Molinari⁵; Dalila Laoudi-Chenivesse¹; Jacques Mercier¹; Olivier Birot⁴; Maurice

Havot¹

¹ INSERM U-1046, CHRU Montpellier, Department of Clinical Physiology, University of

Montpellier I, University of Montpellier II, F-34295 Montpellier, France

² Pulmonary Rehabilitation Center "La Solane", Fontalvie Group, F-66340 Osséja, France

³ Pulmonary Rehabilitation Center "La Vallonie", Fontalvie Group, F-34700 Lodève, France

⁴ York University, Faculty of Health, Angiogenesis Research Group, Toronto, ON, M3J 1P3,

Canada

⁵ CHRU Montpellier, Department of Medical Information, UMR 729 MISTEA, University of

Montpellier I, F-34295 Montpellier, France

RUNNING HEAD: BLUNTED MUSCLE ANGIOGENIC RESPONSE IN COPD

Corresponding author

Fares Gouzi, MD, PhD

INSERM U-1046, University of Montpellier I, University of Montpellier II

Department of Clinical Physiology, CHRU Montpellier

34295 Montpellier cedex 5, France

0

Phone: +33 467 335 908

Fax: +33 467 335 923

e-mail: $\underline{f\text{-}gouzi@chu\text{-}montpellier.fr}$

Word count: 2958

ABSTRACT

The impaired skeletal muscle of chronic obstructive pulmonary disease (COPD) patients reduces exercise capacity. Similar to the oxidative muscle fibers, the angio-adaptation of muscle to training may be blunted in these patients, as in other chronic conditions. We therefore compared muscle functional responses and angio-adaptations after training in COPD patients and sedentary healthy subjects (SHS).

Twenty-four COPD patients [FEV₁ (in %pred.): 45.6±17.5] and 23 SHS (<150 min/week of moderate-to-vigorous exercise) completed a 6-week rehabilitation program based on individualized moderate-intensity endurance training. Histomorphological muscle analysis and measurements of pro-angiogenic vascular endothelial factor-A (VEGF-A) and antiangiogenic thrombospondin-1 (TSP-1) were conducted before and after training.

COPD patients and SHS showed improved symptom-limited oxygen consumption and muscle endurance, although improvements were lower in COPD patients (+0.96±2.4 vs. +2.9±2.6 mL/kg/min, p<0.05 and +65% vs. +108%, p=0.06, respectively). The capillary-to-fiber (C/F) ratio increased less in COPD patients than SHS (16±10% vs. 37±20%, p<0.05) and no fiber type switch occurred in patients. The VEGF-A/TSP-1 ratio increased in COPD patients and SHS (+65% vs. +35%, p<0.05). Changes in C/F and VO2_{SL} were correlated (r=0.51, p<0.05). In addition to a lack of fiber switch, COPD patients displayed a blunted angiogenic response to training.

Abstract word count: 194

KEY-WORDS: exercise testing, chronic diseases, muscle dysfunction in COPD (myopathy), vascular remodeling, rehabilitation programs, remodeling

INTRODUCTION

Skeletal muscle dysfunction has substantial implications in many chronic diseases [chronic heart failure (CHF), diabetes, peripheral arterial disease [1], obesity, etc.]. In chronic obstructive pulmonary disease (COPD), the skeletal muscle dysfunction affects the patient's prognosis, exercise tolerance, and health-related quality of life [2]. This muscle impairment is characterized by cellular changes, with decreases in the proportion of type I oxidative fibers, oxidative enzymes [3], and capillaries [4]. In particular, the alteration in the capillary supply [4] is well-correlated with the impaired muscle function and aerobic exercise capacity of these patients.

The impaired skeletal muscle of COPD patients has been associated with physical inactivity and, although exercise training is the most efficient therapy for the skeletal muscle dysfunction in these patients [5], it has never been able to fully reverse this dysfunction [6]. Systemic factors may explain this incomplete recovery [2], but a lack of muscle adaptation at the cellular level has also been incriminated. For example, several studies have shown no increase in the type I fiber proportion after exercise training in the quadriceps of COPD patients [7-10], in contrast to the observations in healthy subjects [11].

Consistent with the role of capillarization in muscle function, it is possible that impaired training-induced angiogenesis would also explain the incomplete muscle function recovery in COPD patients. Because exercise training significantly increases muscle capillarization in these patients [7, 8, 12], it could be hypothesized that the angiogenic response of muscle to training is present but blunted in COPD, as in other chronic conditions with similar skeletal muscle impairment (diabetes [13] and CHF [14]). Unfortunately, no study has ever compared the training-induced capillarization in COPD patients versus sedentary healthy subjects

(SHS). Moreover, the impact of COPD on the training-induced expression of the main molecular factors controlling skeletal muscle angio-adaptation, such as pro-angiogenic vascular endothelial growth factor-A (VEGF-A) or anti-angiogenic thrombospondin-1 (TSP-1), remains unknown.

Therefore, the aim of our study was to determine whether COPD patients experience a specific alteration in their muscle angio-adaptive capacity. To do so, we compared functional responses, myofiber remodeling, capillarization, and the main angiogenic factors after a moderate-intensity training program in COPD patients and sedentary healthy subjects with the same pattern of physical inactivity.

MATERIALS AND METHODS

See additional information and references in the online supplement.

Study population

From March 2008 to February 2009, we recruited 24 SHS (age from 50 to 75 years) with no disease and less than 150 minutes of moderate-to-vigorous physical activity per week (corresponding to the recommended threshold for health improvement [15]) and 24 patients with COPD defined by dyspnea, chronic cough or sputum production and/or a history of exposure to risk factors for the disease, and post-bronchodilatator FEV1/FVC <70% [16]. Exclusion criteria included other respiratory diagnosis, decompensated comorbidity, exacerbation in the last 3 months, and previous participation in an exercise training program. All subjects and patients performed the tests at INSERM U-1046, CHRU Montpellier, France, or "La Solane" and "La Vallonie" Pulmonary Rehabilitation Centers in Osseja and Lodève, France, respectively. Informed written consent was obtained from all subjects, and the

research protocol was approved by the institutional ethics committee of the Montpellier University Hospitals (n° 2008-EESSS-V2 and n°2009-04-BPCO-V2) and conducted in accordance with the Helsinki declaration and the European guidelines for "good clinical practice.

Physical activity

In order to assess the physical activity (PA) of our sedentary-selected healthy population, we used the Voorrips questionnaire (modified Baecke's questionnaire), a PA questionnaire validated and used in this indication [17]. "Objective" PA level was assessed in 20 COPD patients and 20 SHS who wore a triaxial accelerometer (Tritrac RT3 Research Tracker, Stayhealthy, Monrovia, CA, USA), validated in COPD [18], for 7 consecutive days.

The QUANTAP interview-administered survey is a computer-assisted tool designed to determine PA over a lifetime in 4 dimensions (sports at school, leisure sports, occupation, and daily activities). This questionnaire is reliable to assess lifetime PA and has been validated for use in elderly French subjects and in the context of various pathologies, recently in COPD [19].

Pulmonary function tests

All subjects underwent whole body plethysmography (Transmural Bodybox 2800; Sensomedics, Yorba Linda, CA, USA). The values were compared with normal values [20] (see online supplement).

Exercise testing and muscle function assessment

The 6-minute walking test (6MWT) was performed in a 30-meter corridor. The distance walked during the test (6MWD) was compared with the reference values [21]. Participants performed an incremental test on an electrically-braked cycle ergometer (Ergoselect 200P,

Ergolyne, Bitz, Germany) to determine symptom-limited oxygen consumption (VO_{2SL}) and the ventilatory threshold [22] (see online supplement). If the ventilatory threshold could not be detected, we assessed the dyspnea threshold by rating breathlessness every minute of the test, using the patient's own vocabulary (see online supplement) [23] (see online supplement). The quadriceps strength and endurance were measured by isometric maximal voluntary contraction (qMVC) and endurance time (T.lim). Fat-free and muscle mass were estimated with multi-frequency bioelectrical impedancemetry (QuadScan 4000, Bodystat, Isle of Man, UK). Additional information is provided in the online supplement.

Exercise training

The exercise training sessions were part of a multi-component and comprehensive pulmonary rehabilitation course, including an education program [5]. Twenty sessions of endurance exercise were conducted 3 or 4 times per week for 6 weeks. The exercise intensity was set as the heart rate at the ventilatory or dyspnea threshold [5, 22, 23]. Each session lasted 45 minutes, and in 1 out of 2 sessions, was followed by 30 minutes of strength-building exercise performed at 40% of the one-repetition maximum (1-RM). All sessions were supervised by an experienced clinician, and the training intensity was increased during the training protocol.

Muscle biopsy analysis

Muscle biopsies were performed in the vastus lateralis of the quadriceps before the exercise training program and 48 hours after the last training session [24].

Muscle immunohistochemistry

Muscle fiber type and mean cross-sectional area (CSA) were assessed after immunohistochemistry on frozen sections (10-µm thickness) from the muscle biopsies, using

antibodies specific to each myosin heavy chain (MHC) isotype (University of Iowa, IO, USA) [4, 25]. Capillaries were visualized by immunochemistry with the monoclonal antibody against CD31 (BD Bioscience, #550389, Franklin Lakes, NJ, USA). CSA, capillary density and the capillary-to-fiber ratio were determined after counting capillaries and myofibers on 1 to 2 cryosections from each muscle biopsy (see online supplement).

Western blotting

Blots were probed for vascular endothelial growth factor-A (VEGF-A, clone VG-1, Millipore #05-1117, Etobicoke, ON, Canada) and thrombospondin-1 (TSP-1, clone A6.1, Invitrogen #399300, Burlington, ON, Canada). β -actin protein was also measured as our loading control (see online supplement).

Statistical analysis

Data are presented as mean \pm SD. Distribution normality was tested by the Kolmogorov-Smirnov test. Pearson coefficients describe the correlations. We used a multiple linear regression model. Comparisons of the training effect between groups were performed by mixed linear regression modeling [26]. Each patient/subject was evaluated twice (repeated measures). Thus, measured variables were adjusted using a linear mixed-effects model for repeated measures to take into account the repeated measures as random variables. Patient group and time of measure were used as fixed effects in the model. The group effect was then tested with the mixed-linear model. A p-value of <0.05 was considered statistically significant. Data were analyzed using R.2.13.0 software.

RESULTS

Baseline characteristics of the COPD patients and SHS

All the SHS had a Voorrips score below 9.4, indicating that they were all sedentary. The COPD patients and SHS were well-matched according to age and physical activity (PA) level (Table 1). Moreover, there was no significant difference between the COPD patients and SHS for their PA level over the past 15 years (in metabolic equivalents: 14949±9470 vs. 12341±5201, p=0.26). At the morphological level (Figure E1, online supplement), COPD muscle presented a significantly lower proportion of type I fibers compared with that of SHS (Table 2). Additional details are provided in the online supplement.

Exercise training-induced functional and histomorphological adaptations

Exercise training was performed at the intensity of the ventilatory or dyspnea (n=6/24 COPD patients) threshold (mL/kg/min and % of predicted VO_{2SL}) in COPD patients and SHS: 12.7 ± 1.9 , $48.7\pm11.4\%$ and 15.4 ± 3.5 , $63.5\pm10.1\%$, respectively; p<0.001. Expressed in % of maximal heart rate, the relative intensity was not significantly different in COPD patients and SHS ($66\pm6.8\%$ and $69\pm6.3\%$, respectively, p=0.27). Expressed in % of VO_{2SL}, the relative intensity was slightly higher in COPD patients vs. SHS ($66\pm6\%$ vs. $60\pm5\%$, respectively; p=0.05). Relative improvements for all functional parameters are shown in Figure 1. The training significantly improved 6MWD (m) in both groups (COPD: $+37\pm35$ and SHS: $+31\pm35$, p<0.001; Figure 1). The VO_{2SL} showed significant interaction between group and time (p<0.05). Last, a significant increase in T.lim (s) occurred in both groups (COPD: $+121\pm169$ and SHS $+334\pm274$, p<0.001), with an interaction between group and time that almost reached significance (p=0.06).

Representative changes in type I fiber proportions in COPD patients and SHS are depicted in Figure 2A. The type I fiber proportion was significantly increased in the SHS group

(+8 \pm 11.6%, p<0.01; Figure 2C) and was mirrored by a decrease in the percentage of type IIx fibers (-8 \pm 10.0%, p=0.001). Conversely, no significant change occurred in the COPD group (Figure 2B). We observed a reduction in fiber CSA in COPD patients (5107 \pm 1375 to 4565 \pm 1477 μ m², p<0.05) and no significant change in the SHS group (4409 \pm 1679 to 4629 \pm 1670, p=0.51). Both type I and type IIa fiber CSA showed this reduction in COPD patients (5905 \pm 1588 to 5260 \pm 1493, p<0.05, and 5400 \pm 2012 to 4657 \pm 1460, p<0.05).

Exercise training-induced muscle angio-adaptations

Capillaries were visualized after immunodetection of the endothelial marker CD31 (Figure 3A). In both groups, the exercise training stimulated angiogenesis, as reflected by the increase in the capillary-to-fiber ratio (COPD: 16±10% and SHS: 37±20%, p<0.001; Figure 3B.). Yet there was a significant interaction between group and time (p<0.01), indicating lower improvement in COPD patients. An improvement in capillary density was observed in both groups (COPD: 29±28% and SHS: 9±17%, p<0.01), with no group-time interaction (p=0.11).

Exercise training had no significant effect on the VEGF-A protein level in either healthy subjects or COPD subjects. TSP-1 protein expression decreased in response to exercise training in both groups (p<0.05) with a 44% decrease observed in the COPD population (0.134±0.022 in pre-training vs. 0.075±0.013 in post-training, p<0.05; Figure 4D). Interestingly, the VEGF-A/TSP-1 ratio was significantly increased by exercise training in both the COPD and SHS groups (+65% and +35%, respectively, p<0.05; Figure 4E), in favor of an angiogenic response.

Angiogenesis-function relationships

At baseline, the capillary-to-fiber ratio was significantly and positively correlated with VO_{2SL} and qMVC in COPD patients and SHS (r=0.60, p<0.01 and r=0.67, p<0.01, Figures 5A and 5B). The capillary density was not significantly correlated with the type I fiber proportion (p=0.07). The absolute training-induced changes in the capillary-to-fiber ratio was significantly correlated with the changes in VO_{2SL} in both groups (r=0.51, p<0.05; Figure 5C).

DISCUSSION

The major finding of our study was that the skeletal muscle in COPD patients showed a blunted response to training in terms of muscle capillarization response, which was linked with the lower functional improvements. We also observed an increase in the VEGF-A/TSP-1 ratio in the two groups, in favor of an angiogenic process. Last, a lack of fiber type switch response to training was observed in the COPD patients compared with the age- and PA-matched healthy subjects.

Effect of the training stimulus

Our program of exercise training at moderate intensity significantly increased the exercise capacity in both COPD patients and SHS. Additional details are provided in the online supplement. Interestingly, there was a significant interaction between group and time effects for VO_{2SL} and T.lim, indicating that the functional changes were lower in the COPD patients for these parameters. These lower functional improvements in COPD patients compared with SHS are compatible with the lack of exercise-induced muscle fiber type changes [27].

Regarding the exercise training intensity, all participants were trained at their own ventilatory (or dyspnea) threshold, which is a method for targeting the training stimulus to the individual muscle aerobic capacity [28]. The relative intensity was similar in terms of the percentage of

maximal heart rate, which is another classical method to target relative exercise intensity [5, 22]. Studying trained and untrained healthy subjects, it has appeared that the relative but not the absolute intensity was the determinant of the level of transcriptional activation of muscle genes in response to exercise training [29]. Greater muscle responses after continuous exercise performed at similar relative intensity (50 and 65% of the VO_{2SL}) have even been observed in COPD patients compared with healthy subjects, especially for PGC-1α, which is a master regulator of the fiber type switch [30]. Thus, in our study, it is very likely that at the muscle level, the training stimulus was at least similar in COPD patients and SHS.

Muscle histomorphological and functional responses to training in COPD patients

In agreement with previous works, the baseline level of muscle capillarization was lower in COPD patients [4, 7] and well-correlated with the muscle dysfunction and O₂ consumption before training [4]. We observed an increase in the capillary-to-fiber ratio, which may indicate a training-induced improvement in the capacity to transfer O₂ to myofibers in both COPD patients and SHS. Indeed, recent work has shown that the O₂ transfer to the cell is mostly a function of the capillary-to-fiber interface [31], as reflected by the capillary-to-fiber ratio [32]. This was highlighted by many of our cryosections, which showed an increase in the length of the capillary contact and thus the length of the interface (Figure 3A).

Moreover, the increase in the muscle capillary-to-fiber ratio (i.e., the capillary-to-fiber interface, and thus the O_2 transfer capacity) may have functioned as a master-driver of muscle O_2 uptake and thus improved the endurance after moderate-intensity exercise training in the COPD patients. This would be consistent with evidence showing the critical role of O_2 transfer to cells across capillaries in the maximal O_2 uptake of muscle from healthy subjects [33]. Indeed, in our study, we found no change in the capacity of myofibers to consume O_2 . In

contrast to SHS [34], COPD patients showed no shift toward more oxidative type I fibers [7, 12], and the preliminary data in our COPD patients (n=10) indicated no improvement in the maximal rates of oxygen consumption of permeabilized fibers (see Methods and Results in the online supplement). Last, the significant and positive correlation between the capillary-to-fiber ratio and the VO_{2SL} changes, as previously observed in healthy subjects [35], strengthens the credibility of a role for capillarization in the improvement of maximal O₂ uptake in our COPD patients.

Training-induced response of the angiogenic factors in COPD patients

Angiogenesis is a complex and multi-step biological process tightly orchestrated by a balance between pro- and anti-angiogenic factors. Therefore, we chose to target key pro- and antiangiogenic factors with a well-characterized clinical impact, i.e., VEGF-A [36] and TSP-1. The deletion of VEGF-A in the myofibers from transgenic mice dramatically reduced muscle capillarization and decreased exercise endurance time by approximately 80% [37]. Gene therapy targeting VEGF gene expression improved muscle capillarization [38]. TSP-1 is a large matrix glycoprotein whose action principally serves to inhibit angiogenesis [36]. Its clinical impact has been highlighted by increased muscle capillarity and 67% greater endurance in mice knocked out for the TSP-1 gene compared with wild-type [39]. Altogether, the VEGF-A/TSP-1 ratio has emerged as the most relevant marker of the muscle angioadaptive balance [36]. After hindlimb unloading [40], an increase of the VEGF-A/TSP-1 ratio was observed, consistent with the capillarization increase. Thus, assessing the VEGF-A/TSP-1 ratio appears to be a more accurate approach than analyzing the expression of molecular factors individually [36]. In our study, we also observed a significant increase in this ratio in both groups, which indicates that the angiogenic balance in our patients shifted in favor of angiogenesis, consistent with the capillarization increase.

Which mechanisms for the impaired muscle cellular responses to training in COPD patients? The defective muscle adaptation to exercise training in COPD patients was characterized by a lack of fiber type shift and a blunted capillarization response. Thus, we can hypothesize a pleiotropic patho-biological mechanism. In COPD, systemic factors (hypoxemia, oxidative stress or systemic low-grade inflammation – all enhanced during exercise) have already been incriminated in the peripheral muscle dysfunction [2]. Nonetheless, a molecular link between the cellular pathways regulating muscle angiogenesis and the fiber type cannot be eliminated.

Clinical implications

A key observation in our study was the link between reduced function (quadriceps T.lim and VO_{2SL}) and capillarization. Moreover, our study indicates a probable role of the blunted muscle capillarization response to training in the reduced VO_{2SL} improvement in the COPD patients, as angiogenesis has also shown an impact on the improvement in VO_{2SL} in two other chronic diseases: CHF [41] and peripheral arterial disease [42]. Our finding of lower improvement in the capillary-to-fiber ratio thus appears to be clinically significant. The blunted angiogenic capacity in COPD muscle is consistent with observations in other chronic diseases with similar peripheral muscle dysfunction (diabetes [13] and CHF [14]). In diseases associated with lower limb ischemia, muscle angiogenesis may thus be a relevant target for therapeutic interventions designed to improve muscle capillarization [43]. However, although O₂ transfer to muscle appears to be critical, there is no evidence of chronic lower limb ischemia in COPD patients [44]. Nonetheless, patients with hypoxemia and/or co-morbidities (vascular and/or diabetic) can experience chronic ischemia and in this case therapeutic interventions in association with exercise training might be an adequate approach.

In conclusion, exercise training increased the muscle capillarization in COPD patients, which might have driven the functional improvements. Although the most relevant parameter of the angio-adaptative balance (i.e., the VEGF-A/TSP-1 ratio) increased in favor of an angiogenic process in the COPD patients and SHS, the increase in capillarization was nevertheless significantly blunted in the patients. In addition to the lack of fiber type switch, reduced angiogenesis constitutes another feature of the altered muscle response to training in COPD patients.

ACKNOWLEDGMENT

The authors gratefully acknowledge A. Bourret and G. Hugon for their technical support in the laboratory and all the team in the pulmonary rehabilitation centers "La Solane" and "La Vallonie" for their contribution to this work. G. Caris and A. Suc are acknowledged for their help with the muscle biopsies. The authors would like to thank G. Carnac for managing the first meeting between the Montpellier and Toronto teams, C. Notarnicola for her expertise on the immunohistochemistry experiments, and C. Stott for his critical reading of the manuscript.

REFERENCES

- 1. Eisner MD, Anthonisen N, Coultas D, Kuenzli N, Perez-Padilla R, Postma D, Romieu I, Silverman EK, Balmes JR. An official American Thoracic Society public policy statement: Novel risk factors and the global burden of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2010; 182: 693-718.
- 2. Couillard A, Prefaut C. From muscle disuse to myopathy in COPD: potential contribution of oxidative stress. *Eur Respir J* 2005; 26: 703-719.
- 3. Gosker HR, van Mameren H, van Dijk PJ, Engelen MP, van der Vusse GJ, Wouters EF, Schols AM. Skeletal muscle fibre-type shifting and metabolic profile in patients with chronic obstructive pulmonary disease. *Eur Respir J* 2002; 19: 617-625.
- 4. Eliason G, Abdel-Halim SM, Piehl-Aulin K, Kadi F. Alterations in the muscle-to-capillary interface in patients with different degrees of chronic obstructive pulmonary disease. *Respir Res* 2010; 11: 97.
- 5. Nici L, Donner C, Wouters E, Zuwallack R, Ambrosino N, Bourbeau J, Carone M, Celli B, Engelen M, Fahy B, Garvey C, Goldstein R, Gosselink R, Lareau S, MacIntyre N, Maltais F, Morgan M, O'Donnell D, Prefault C, Reardon J, Rochester C, Schols A, Singh S,

- Troosters T. American Thoracic Society/European Respiratory Society statement on pulmonary rehabilitation. *Am J Respir Crit Care Med* 2006; 173: 1390-1413.
- 6. Bernard S, Whittom F, Leblanc P, Jobin J, Belleau R, Berube C, Carrier G, Maltais F. Aerobic and strength training in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1999; 159: 896-901.
- 7. Whittom F, Jobin J, Simard PM, Leblanc P, Simard C, Bernard S, Belleau R, Maltais F. Histochemical and morphological characteristics of the vastus lateralis muscle in patients with chronic obstructive pulmonary disease. *Med Sci Sports Exerc* 1998; 30: 1467-1474.
- 8. Vogiatzis I, Stratakos G, Simoes DC, Terzis G, Georgiadou O, Roussos C, Zakynthinos S. Effects of rehabilitative exercise on peripheral muscle TNFalpha, IL-6, IGF-I and MyoD expression in patients with COPD. *Thorax* 2007; 62: 950-956.
- 9. Gosker HR, Schrauwen P, Broekhuizen R, Hesselink MK, Moonen-Kornips E, Ward KA, Franssen FM, Wouters EF, Schols AM. Exercise training restores uncoupling protein-3 content in limb muscles of patients with chronic obstructive pulmonary disease. *Am J Physiol Endocrinol Metab* 2006; 290: E976-981.
- 10. Lewis MI, Fournier M, Storer TW, Bhasin S, Porszasz J, Ren SG, Da X, Casaburi R. Skeletal muscle adaptations to testosterone and resistance training in men with COPD. *J Appl Physiol* 2007; 103: 1299-1310.
- 11. Russell AP, Feilchenfeldt J, Schreiber S, Praz M, Crettenand A, Gobelet C, Meier CA, Bell DR, Kralli A, Giacobino JP, Deriaz O. Endurance training in humans leads to fiber type-specific increases in levels of peroxisome proliferator-activated receptor-gamma coactivator-1 and peroxisome proliferator-activated receptor-alpha in skeletal muscle. *Diabetes* 2003; 52: 2874-2881.

- 12. Vogiatzis I, Simoes DC, Stratakos G, Kourepini E, Terzis G, Manta P, Athanasopoulos D, Roussos C, Wagner PD, Zakynthinos S. Effect of pulmonary rehabilitation on muscle remodelling in cachectic patients with COPD. *Eur Respir J* 2010; 36: 301-310.
- 13. Wallberg-Henriksson H, Gunnarsson R, Henriksson J, Ostman J, Wahren J. Influence of physical training on formation of muscle capillaries in type I diabetes. *Diabetes* 1984; 33: 851-857.
- 14. Kiilavuori K, Naveri H, Salmi T, Harkonen M. The effect of physical training on skeletal muscle in patients with chronic heart failure. *Eur J Heart Fail* 2000; 2: 53-63.
- 15. Nelson ME, Rejeski WJ, Blair SN, Duncan PW, Judge JO, King AC, Macera CA, Castaneda-Sceppa C. Physical activity and public health in older adults: recommendation from the American College of Sports Medicine and the American Heart Association. *Med Sci Sports Exerc* 2007; 39: 1435-1445.
- 16. Rabe KF, Hurd S, Anzueto A, Barnes PJ, Buist SA, Calverley P, Fukuchi Y, Jenkins C, Rodriguez-Roisin R, van Weel C, Zielinski J. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. *Am J Respir Crit Care Med* 2007; 176: 532-555.
- 17. Voorrips LE, Ravelli AC, Dongelmans PC, Deurenberg P, Van Staveren WA. A physical activity questionnaire for the elderly. *Med Sci Sports Exerc* 1991; 23: 974-979.
- 18. Steele BG, Holt L, Belza B, Ferris S, Lakshminaryan S, Buchner DM. Quantitating physical activity in COPD using a triaxial accelerometer. *Chest* 2000; 117: 1359-1367.

- 19. Gouzi F, Prefaut C, Abdellaoui A, Vuillemin A, Molinari N, Ninot G, Caris G, Hayot M. Evidence of an Early Physical Activity Reduction in Chronic Obstructive Pulmonary Disease Patients. *Arch Phys Med Rehabil* 2011.
- 20. Quanjer PH, Tammeling GJ, Cotes JE, Pedersen OF, Peslin R, Yernault JC. Lung volumes and forced ventilatory flows. Report Working Party Standardization of Lung Function Tests, European Community for Steel and Coal. Official Statement of the European Respiratory Society. *Eur Respir J Suppl* 1993; 16: 5-40.
- 21. ATS statement: guidelines for the six-minute walk test. *Am J Respir Crit Care Med* 2002; 166: 111-117.
- 22. Vallet G, Ahmaidi S, Serres I, Fabre C, Bourgouin D, Desplan J, Varray A, Prefaut C. Comparison of two training programmes in chronic airway limitation patients: standardized versus individualized protocols. *Eur Respir J* 1997; 10: 114-122.
- 23. Préfaut C, Gautier-Dechaud V, Fuchs-Climent D, Poulain M. Quality of life in patients with chronic obstructive pulmonary disease on long-term oxygen therapy: role of exercise training. *Human Kinetics* 2000; 15: 177-186.
- 24. Hayot M, Michaud A, Koechlin C, Caron MA, Leblanc P, Prefaut C, Maltais F. Skeletal muscle microbiopsy: a validation study of a minimally invasive technique. *Eur Respir J* 2005; 25: 431-440.
- 25. Abdellaoui A, Prefaut C, Gouzi F, Couillard A, Coisy-Quivy M, Hugon G, Molinari N, Lafontaine T, Jonquet O, Laoudj-Chenivesse D, Hayot M. Skeletal muscle effects of electrostimulation after COPD exacerbation: a pilot study. *Eur Respir J* 2011; 38: 781-788.

- 26. Verbeke G, Molenberghs G. Linear Mixed Models for Longitudinal Data. *In*: Verbeke G, Molenberghs G Linear Mixed Models for Longitudinal Data 2nd ed. Springer-Verlag New York Inc, New York, 2009; pp: 570.
- 27. Horowitz JF, Sidossis LS, Coyle EF. High efficiency of type I muscle fibers improves performance. *Int J Sports Med* 1994; 15: 152-157.
- 28. Vago P, Mercier J, Ramonatxo M, Prefaut C. Is ventilatory anaerobic threshold a good index of endurance capacity? *Int J Sports Med* 1987; 8: 190-195.
- 29. Nordsborg NB, Lundby C, Leick L, Pilegaard H. Relative workload determines exercise-induced increases in PGC-1alpha mRNA. *Med Sci Sports Exerc* 2010; 42: 1477-1484.
- 30. Puente-Maestu L, Lazaro A, Tejedor A, Camano S, Fuentes M, Cuervo M, Navarro BO, Agusti A. Effects of exercise on mitochondrial DNA content in skeletal muscle of patients with COPD. *Thorax* 2011; 66: 121-127.
- 31. Hepple RT, Hogan MC, Stary C, Bebout DE, Mathieu-Costello O, Wagner PD. Structural basis of muscle O(2) diffusing capacity: evidence from muscle function in situ. *J Appl Physiol* 2000; 88: 560-566.
- 32. Hepple RT, Agey PJ, Hazelwood L, Szewczak JM, MacMillen RE, Mathieu-Costello O. Increased capillarity in leg muscle of finches living at altitude. *J Appl Physiol* 1998; 85: 1871-1876.
- 33. Richardson RS, Grassi B, Gavin TP, Haseler LJ, Tagore K, Roca J, Wagner PD. Evidence of O2 supply-dependent VO2 max in the exercise-trained human quadriceps. *J Appl Physiol* 1999; 86: 1048-1053.

- 34. Simoneau JA, Lortie G, Boulay MR, Marcotte M, Thibault MC, Bouchard C. Human skeletal muscle fiber type alteration with high-intensity intermittent training. *Eur J Appl Physiol Occup Physiol* 1985; 54: 250-253.
- 35. Hepple RT, Mackinnon SL, Thomas SG, Goodman JM, Plyley MJ. Quantitating the capillary supply and the response to resistance training in older men. *Pflugers Arch* 1997; 433: 238-244.
- 36. Olfert IM, Birot O. Importance of anti-angiogenic factors in the regulation of skeletal muscle angiogenesis. *Microcirculation* 2011; 18: 316-330.
- 37. Olfert IM, Howlett RA, Tang K, Dalton ND, Gu Y, Peterson KL, Wagner PD, Breen EC. Muscle-specific VEGF deficiency greatly reduces exercise endurance in mice. *J Physiol* 2009; 587: 1755-1767.
- 38. Korpisalo P, Hytonen JP, Laitinen JT, Laidinen S, Parviainen H, Karvinen H, Siponen J, Marjomaki V, Vajanto I, Rissanen TT, Yla-Herttuala S. Capillary enlargement, not sprouting angiogenesis, determines beneficial therapeutic effects and side effects of angiogenic gene therapy. *Eur Heart J* 2011; 32: 1664-1672.
- 39. Malek MH, Olfert IM. Global deletion of thrombospondin-1 increases cardiac and skeletal muscle capillarity and exercise capacity in mice. *Exp Physiol* 2009; 94: 749-760.
- 40. Roudier E, Gineste C, Wazna A, Dehghan K, Desplanches D, Birot O. Angio-adaptation in unloaded skeletal muscle: new insights into an early and muscle type-specific dynamic process. *J Physiol* 2010; 588: 4579-4591.

- 41. Esposito F, Reese V, Shabetai R, Wagner PD, Richardson RS. Isolated quadriceps training increases maximal exercise capacity in chronic heart failure: the role of skeletal muscle convective and diffusive oxygen transport. *J Am Coll Cardiol* 2011; 58: 1353-1362.
- 42. Wang J, Zhou S, Bronks R, Graham J, Myers S. Effects of supervised treadmill walking training on calf muscle capillarization in patients with intermittent claudication. *Angiology* 2009; 60: 36-41.
- 43. Moazzami K, Majdzadeh R, Nedjat S. Local intramuscular transplantation of autologous mononuclear cells for critical lower limb ischaemia. *Cochrane Database Syst Rev* 2011; CD008347.
- 44. Vogiatzis I, Athanasopoulos D, Stratakos G, Garagouni C, Koutsoukou A, Boushel R, Roussos C, Zakynthinos S. Exercise-induced skeletal muscle deoxygenation in Osupplemented COPD patients. *Scand J Med Sci Sports* 2009; 19: 364-372.

FIGURE LEGENDS

Figure 1. Relative changes in functional parameters (%) in chronic obstructive pulmonary disease (COPD) patients (dark) and sedentary healthy subjects (SHS) (white) following exercise training. 6MWD (m): 6-minute walking distance in meters; 6MWD (%pred.): 6-minute walking distance in percentage of the predicted value; VO_{2SL} (mL/kg/min): peak oxygen uptake in milliliters per kilogram and per minute; T.lim (s): endurance time in

seconds; qMVC: quadriceps maximal voluntary contraction in kilograms; BMI: body mass index (kg/m²); FFMI: fat-free mass index in kilograms. Data are presented as mean \pm standard error of the mean. Statistical differences: * time effect, p<0.001; # group-time interaction, p<0.05.

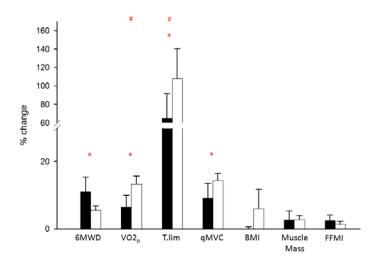


Figure 2. A) Representative transverse cryosections of the *vastus lateralis* stained with the type I myosin heavy chain isotype antibody at a magnification of * 10, in a COPD patient and an SHS before and after exercise. Note the increase in the proportion of type I MHC fibers in the SHS. B) Proportion of the muscle fiber type (%) before (dark) and after (white) the exercise training program in COPD patients (n=21) and C) SHS (n=23). Data are presented as mean \pm standard error of the mean. * p<0.05, after exercise training.

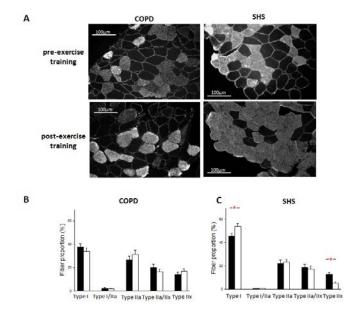


Figure 3. A) Representative pictures of *vastus lateralis* capillarization after staining for the endothelial marker CD31 at a magnification of * 10, in a COPD patient and an SHS before and after exercise. Note the transverse capillaries surrounding the myofiber (black arrows) in pre-exercise training and the increased size of the capillary-to-fiber interface post-exercise training (dotted arrows). B) Mean values and standard error of the mean in COPD patients and SHS before (dark) and after (white) training for the capillary-to-fiber ratio. * p<0.05, after exercise training. # p<0.05, between groups.

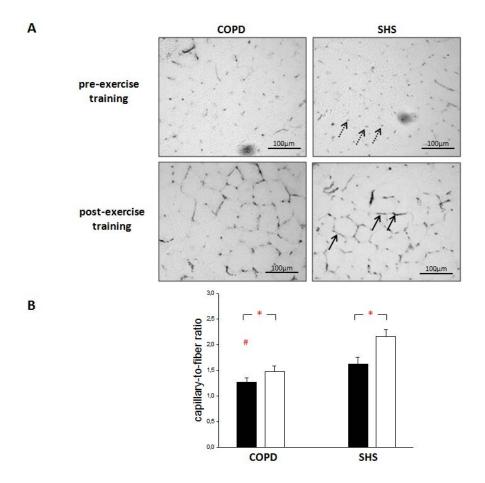


Figure 4. Representative Western blot of vascular endothelial growth factor-A (VEGF-A) and thrombospondin-1 (TSP-1) (respectively, panels A and B) protein expression in the *vastus lateralis* of COPD patients and SHS before (dark) and after (white) training. β -actin is used as a loading control. Densitometry analysis of VEGF-A (C) and TSP-1 (D) protein and the VEGF-A/TSP-1 ratio (E) in the vastus lateralis of COPD patients and SHS before (dark) and after (white) training. Mean \pm standard error of the mean. Statistical difference after training: * p<0.05; between groups (post-training): ** p<0.05.

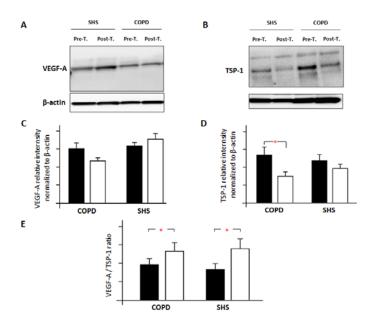
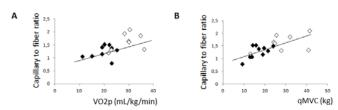


Figure 5. A) The capillary-to-fiber ratio was positively correlated with the peak oxygen uptake (VO_{2SL}), expressed in milliliters per kilogram and per minute (r=0.60, p<0.01), in COPD patients (dark) and SHS (white), and B) with the quadriceps maximal voluntary contraction (qMVC) expressed in kilograms (r=0.67, p<0.01). C) The capillary-to-fiber ratio changes were positively correlated with the changes in peak oxygen uptake (VO_{2SL}), expressed in milliliters per kilogram and per minute (r=0.51, p<0.05), in COPD patients (dark) and SHS (white).

BASELINE



TRAINING-INDUCED CHANGES

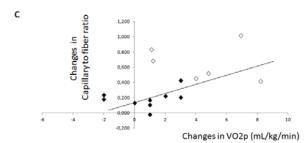


Table 1. Baseline clinical and functional characteristics of the chronic obstructive pulmonary disease (COPD) patients and the sedentary healthy subjects (SHS)

TABLES

	COPD patients	SHS	p
n	24	23	1.00
Age (y)	60.8 ± 8.0	61.5 ± 5.7	0.756
Gender (M/F)	18/6	11/12	
FEV ₁ /VC (%)	42.3 ± 8.9	75.8 ± 4.3	< 0.001
FEV ₁ (% pred.)	45.6 ± 17.5	106.0 ± 12.9	< 0.001
VC (%pred)	90.0 ± 23.7	111.5 ± 14.3	< 0.01
RV (%pred.)	183.8 ± 26.3	114.1 ± 34.6	< 0.001
RV/TLC (%)	52.6 ± 12.2	37.9 ± 6.0	< 0.01
FRC (%pred)	156.7 ± 22.2	109.7 ± 22.3	< 0.001
PaO2 (mmHg)	72.1 ± 10.1	-	
PA level	6.3 [2.5-7.9]	4.3 [3.2-5.6]	0.474
(Voorrips score)			
PA level	123 ± 44	137 ± 51	0.490
(activity counts, AU)	(n=9)	(n=19)	
6MWD (m)	480 [426-568]	615 [580-652]	< 0.001
6MWD (%pred.)	70.4 ± 16.2	93.4 ± 10.5	< 0.001
VO _{2SL} (mL/kg/min)	16.8 ± 5.0	25.7 ± 6.0	< 0.001
VO _{2SL} (%pred.)	64.1 ± 20.2	107.7 ± 13.8	< 0.001
BMI (kg/m^2)	24.7 ± 4.1	25.9 ± 2.8	0.285
FFMI (kg/m²)	18.1 ± 2.0	18.9 ± 2.9	0.287
qMVC (kg)	17.6 ± 5.6	21.7 ± 9.0	0.105
Quadriceps T.lim (s)	272 [159-368]	340 [221-656]	0.05

Definition of abbreviations: M: male/F: female; FEV_1 (% pred.): forced expiratory volume in 1 second in percentage of the predicted value; VC (% pred.): slow vital capacity, in percentage of the predicted value; RV (%pred.): residual volume, in percentage of the predicted value; RV/TLC (%): ratio of the residual volume and total lung capacity, in percentage; FRC (% pred.): functional residual capacity, in percentage of the predicted value; PA: physical activity; 6MWD (m): 6-minute walking distance, in meters; 6MWD (%pred.): 6-minute walking distance, in percentage of the predicted value; VO_{2SL} (mL/kg/min): peak oxygen uptake, in milliliters per kilogram and per minute; BMI: body mass index (kg/m²); FFMI: fat-free mass index (kg/m²); qMVC (kg): quadriceps maximal voluntary contraction, in

kilograms; T.lim (s): quadriceps endurance time, in seconds. Results are expressed in mean \pm SD or median [interquartile range (IQR)].

Table 2. Muscle characteristics of the chronic obstructive pulmonary disease (COPD) patients and the sedentary healthy subjects (SHS)

	COPD patients	SHS	p
n	24	23	
Muscle fiber %			
Type I	37.5 ± 12.6	44.5 ± 11.1	< 0.05
Type I/IIa	2.1 ± 4.1	0.8 ± 0.8	0.118
Type IIa	26.4 ± 15.9	22.2 ± 14.0	0.356
Type IIa/IIx	20.0 ± 13.5	19.5 ± 12.1	0.886
Type IIx	13.8 ± 10.8	13.0 ± 8.6	0.784
All fiber cross-sectional	5107 ± 1376	4409 ± 1679	0.141
area (µm²)			
Capillary-to-fiber ratio	1.273 ± 0.247	1.625 ± 0.327	< 0.05
	(n=10)	(n=8)	
Capillary density (/mm²)	256 ± 539	309 ± 557	0.057
-	(n=10)	(n=8)	

Results are expressed in mean \pm SD