

# Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure

Davy Guignard, Véronique V. Gayrard-Troy, Marlène Z. Lacroix, Sylvie Puel, Nicole Picard-Hagen, Catherine Viguié

# ▶ To cite this version:

Davy Guignard, Véronique V. Gayrard-Troy, Marlène Z. Lacroix, Sylvie Puel, Nicole Picard-Hagen, et al.. Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure. Chemosphere, 2017, 182, pp.458-467. 10.1016/j.chemosphere.2017.05.028. hal-01607369

# HAL Id: hal-01607369 https://hal.science/hal-01607369

Submitted on 26 May 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Accepted Manuscript

Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure

Davy Guignard, Véronique Gayrard, Marlène Lacroix, Sylvie Puel, Nicole Picard-Hagen, Catherine Viguié

PII:	S0045-6535(17)30723-3
DOI:	10.1016/j.chemosphere.2017.05.028
Reference:	CHEM 19243
To appear in:	Chemosphere
Received Date:	10 January 2017
Revised Date:	28 April 2017
Accepted Date:	04 May 2017

Please cite this article as: Davy Guignard, Véronique Gayrard, Marlène Lacroix, Sylvie Puel, Nicole Picard-Hagen, Catherine Viguié, Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure, *Chemosphere* (2017), doi: 10.1016/j.chemosphere.2017.05.028

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the 1 pregnant ewe at low level representative of human exposure 2 Davy Guignard, Véronique Gayrard, Marlène Lacroix, Sylvie Puel, Nicole Picard-Hagen, 3 Catherine Viguié 4 Toxalim (Research Centre in Food Toxicology), Université de Toulouse, INRA, ENVT, INP-5 Purpan, UPS, Toulouse, France 6 7 Corresponding author: Catherine Viguié 8 TOXALIM UMR 1331, INRA UPS 9 10 Ecole Nationale Vétérinaire de Toulouse 23 chemin des Capelles, BP 87614 11 31076 Toulouse cedex3, France 12 Phone: +33 561 19 39 13 13 Email: catherine.viguie@inra.fr 14 15 16

#### 17 <u>Abstract</u>

Many uncertainties remain regarding the potential of bisphenol A (BPA) as a thyroid disruptor 18 in mammals and the relevance of experimental data to humans. The relevance of the exposure 19 schemes used in experimental in vivo studies is also a major source of uncertainty when 20 analysing the risk of BPA exposure for human health. In this context, the goals of our study, 21 conducted in an ovine model relevant to human gestation and thyroid physiologies, were to: 1) 22 determine the equivalence of subcutaneous and dietary exposures and 2) determine if 23 environmentally relevant doses of BPA can alter gestational and newborn thyroid functions. 24 The difference between the two routes of exposure was mainly related to the overall BPA 25 exposure and much less to the peak serum concentrations. Interestingly, BPA-GLUC (the main 26 metabolite of BPA) internal exposure via both routes was almost identical. The decrease in 27 thyroid hormones concentration overtime was more accentuated in ewes treated with BPA, 28 particularly with the medium dose (50  $\mu$ g/(kg.d); SC) for which the maximum BPA 29 concentrations were predicted to be within the 1-10 ng/mL range i.e. very similar to the highest 30 blood concentrations reported in humans. The balance between TT4 and rT3 varied differently 31 between the vehicle and the medium dose group. The mechanisms underlying those 32 modifications of maternal thyroid homeostasis remain to be determined. Our study did not 33 evidence significant modification of TSH secretion or binding to serum proteins but might 34 suggest an effect at the level of deiodinases. 35

- 36
- 37 Highlights
- Maximum BPA serum concentration is similar between subcutaneous and dietary routes
- 39 -Bisphenol A alters the maternal thyroid status in pregnant ewe
- 40 -Thyroid disruption is observed at BPA serum concentration relevant to human exposure

41 -BPA might disrupt the deiodinase activity in pregnant ewe

#### 42 Abbreviations:

43 AUC, area under the curve; BPA, bisphenol A; BPA-GLUC, bisphenol A glucuronide; BPA-

44 SULF, bisphenol A sulfate; Cmax, maximum concentration; GD, gestational day; LOQ, limit

45 of quantification; LOD, limit of detection; PK, pharmacokinetic; PND, post-natal day; rT3,

46 reverse triiothyronine; S.C., subcutaneous; TH, thyroid hormones; TSH, thyroid stimulating

47 hormone; TT3, total triiothyronine; TT4, total thyroxine; fT4, free thyroxine.

48

49 Keywords: bisphenol A; toxicokinetic; exposure; thyroid disruption; pregnancy; ovine model

#### 1. <u>Introduction</u>

50

Bisphenol A (BPA), a major molecule used in the plastics industry, is now recognized to be an endocrine disruptor that exerts deleterious effects on human health (Chapin et al., 2008; vom Saal et al., 2007). Most investigations have focused on reproductive functions based on the estrogenomimetic properties of this compound. However, accumulated evidence suggests that BPA might have negative effects on other endocrine systems (Richter et al., 2007; Rubin, 2011) including thyroid function (Zoeller, 2010).

Although sometimes controversial, the results of numerous epidemiological studies in humans 57 indicate that BPA exposure might be associated with diverse degrees of thyroid disruption in 58 adults, children and new born. Epidemiological data obtained in a cohort of men from infertile 59 couples (Meeker et al., 2010) and in a larger cohort of healthy individuals (NHANES cohort; 60 Meeker and Ferguson, 2011) suggest that BPA exposure might be associated with some degrees 61 of thyroid disruption characterized by an inverse relationship between urinary BPA 62 concentrations and total thyroxine (TT4) concentrations. In another cohort of adults, urinary 63 BPA concentration was, respectively, positively and negatively correlated with free T3 and 64 TSH serum concentrations (Wang et al., 2013). In a large cohort of Thai adults, BPA serum 65 concentrations were shown to be negatively correlated with free T4 (fT4) concentrations in men 66 but not in women (Sriphrapradang et al., 2013). In a cohort of 9 to 11 year old Chinese children, 67 the BPA urinary concentration in morning spot sampling was negatively correlated with thyroid 68 gland volume (Wang et al., 2015) and this correlation did not seem to depend upon the iodine 69 supply and/or iodine status of the children. 70

As far as gestational exposure is concerned, data are very scarce in human. However, there is
some evidence that infant thyroid function might be altered in relation to maternal exposure to
BPA. In a cohort of pregnant women and their infants (CHAMACOS study), urinary BPA in

the mother was shown to be negatively correlated with maternal TT4 blood concentration when the urine BPA and TT4 blood samples were collected close together in time (Chevrier et al., 2013). As a result, the average maternal urinary BPA was related to increased odds of low TT4 in the mother. An inverse relationship between maternal urinary BPA and neonatal blood TSH was evidenced in newborn boys. In the HOME study, 26-week maternal urinary BPA concentrations were inversely correlated with serum TSH concentrations in newborn girls but not in boys (Romano et al., 2015).

Very few experimental studies have been done on the effect of BPA on thyroid function in 81 mammals. In one study, the physiological increase in circulating TT4 that generally occurs 82 around post-natal day 15 (PND15) was amplified in rats born to and fed by mothers treated with 83 an oral BPA dose of 1 mg/(kg.d) from gestational day (GD) 6 until weaning (Zoeller et al., 84 2005). In another study in pregnant rats (Xu et al., 2007), BPA oral treatment of 0.1 mg/L in 85 drinking water (approximately 10 µg/(kg.d)) led to a transitory decrease in fT4 concentration 86 in mothers during the early postpartum period (PND0 to PND7), while their male offspring 87 showed an increase in fT4 at PND7 followed by a decrease at PND21. 88

The relevance to humans of thyroid regulation in rodents is nevertheless controversial due to 89 90 the different timing of thyroid function ontogenesis and the lack of expression of thyroxine binding globulin (TBG), the high affinity specific binding protein in adult rats (Fisher et al., 91 92 1976). This serum protein is considered a key player in the maintenance of thyroid homeostasis 93 (Hard, 1998; Wu and Farrelly, 2006). Ovine model is considered as a more relevant model to human than rodents from the thyroid regulation standpoint. Indeed, in this species as in human, 94 TBG is the major serum carrier protein of thyroid hormones. Furthermore, ovine and human 95 species are both considered as precocious species for the ontogenesis of the thyroid system 96 because fetal thyroid axis is almost mature at birth. In sheep, thyroid gland can accumulate 97 iodine and synthesize T4 around the 70<sup>th</sup> day of gestation, the neuroendocrine control of the 98

99 thyroid axis starts to develop as soon as the 50<sup>th</sup> day of gestation (Fisher et al., 1976) and TH 100 receptor expression is observable around 80 days of gestation in the brain and the liver (Polk et 101 al., 1989). T3 levels in blood are much lower in the fetus than in the mother and start to rise 102 significantly only on the very late stages of pregnancy in both human (Fisher et al., 2002) and 103 sheep (Fisher et al., 1994).

In one of our studies in pregnant ewes, BPA exposure was associated with hypothyroxinemia
(about 30% decrease in TT4) in both the mothers and newborn lambs after subcutaneous
exposure of the ewes to 5 mg/(kg.d) throughout gestation (Viguié et al., 2013).

Evidence that BPA can be a potential thyroid disruptor has also arisen from studies in 107 mammalian cells and in amphibian or fish larvae models. The results of these studies converge 108 to suggest that BPA can exert an antagonistic effect on TH-dependent mechanisms (Fini et al., 109 110 2007; Goto et al., 2006; Heimeier et al., 2009; Iwamuro et al., 2003; Kaneko et al., 2008; Sun et al., 2009; Terrien et al., 2011; Yang and Chan, 2015) and that several pathways involved in 111 TH economy can be affected by BPA. Hepatic regulation of thyroid hormone homeostasis can 112 also be affected by BPA as shown in a zebrafish liver cell line in which BPA (45-180 µM) 113 decreased the expression of several genes involved in thyroid homeostasis i.e., deiodinases I 114 115 and III, sulfotransferases 1,2 and 3, UGT2A1 and thyroid receptor  $\beta$  (Yang and Chan, 2015). In addition, the antagonistic effect of BPA could be mediated, at least in cell models, by the 116 117 increased recruitment of co-repressors of TH nuclear receptors from the N-CoR family (Moriyama et al., 2002; Sheng et al., 2012). 118

Overall, it is apparent that uncertainties remain regarding the potential of BPA as a thyroid disruptor in mammals and the relevance of experimental data to humans. Another major source of uncertainty on BPA risk analysis for human health based on experimental data obtained in animals is related to the level of internal exposure, particularly with regard to the route of

administration. Indeed, in many good quality experimental studies, animals were exposed via
parenteral or gastric gavage routes to ensure better control of the dose administered although
none of these routes is totally representative of the acknowledged main route of human exposure
i.e. via food contamination (Guignard et al., 2016; Taylor et al., 2011; Vom Saal et al., 2014).
Thus, the relevance of the route of administration is a critical issue that every scientist needs to
bear in mind when performing toxicological evaluations of small lipophilic molecules which,
alike BPA, are subjected to a high hepatic first pass effect.

In this context, the goals of our study conducted in an ovine model relevant to human gestation and thyroid physiologies were to 1) determine the equivalence of subcutaneous and dietary exposures and 2) determine if environmentally relevant doses of BPA can alter gestational and newborn thyroid functions.

134

135

#### 2. Materials and Methods

#### 136 2.1. <u>Chemicals</u>

Bisphenol A 99% purity (Sigma-Aldrich, Saint-Quentin Fallavier, France) was dissolved in 137 corn oil / absolute ethanol (6/1: V/V). For experiment 1, a single BPA solution (100 mg/mL) 138 was prepared and used for both routes of administration. The volume administered to the 139 animals was adjusted to the bodyweight recorded the day before the administrations. For the 140 dietary route, BPA solution was spilled onto the pellet ration (on average 6 mL of solution on 141 142 75g of pellets). The subcutaneous (S.C.) injection of BPA was performed on the distal part of the neck (on average 3 mL/ewe). For experiment 2, a vehicle and three different BPA solutions 143 (0.01 - 1 and 100 mg/mL) were prepared so that the injected volume adjusted to individual 144 bodyweight was the same in all groups. The solutions were renewed monthly and were stored 145 at ambient temperature in sealed amber glass bottles for the entire duration of their use. The 146

vehicle solution was prepared and stored in a different room. All material used for the solution
preparation, sample processing and assays was either made of glass or of BPA-free plastics.

149

## 150 2.2. <u>Animal husbandry</u>

All animal procedures were carried out in accordance with the accepted standards of humane 151 animal care under agreement 311155515 for animal experimentation from the French Ministry 152 of Agriculture and validated by the local ethical committee for animal experimentation (project 153 number: 2016 0323 10542323) and by the French Ministry of Higher Education and Research 154 (project number: 00890.01) for experiments 2 and 1, respectively. The study was performed on 155 adult Lacaune ewes in the sheep research facility at the National Veterinary School of Toulouse. 156 France. The ewes were kept under natural photoperiodic and temperature conditions, with free 157 access to water, hay ad libitum and plant based pellets (Brebilac® RAGT, Rodez, France). The 158 absence of BPA contamination of water, hay and pellets was checked by assaying BPA in 100 159 mg samples of each matrix homogenized in 1 mL of acetonitrile. BPA was not detected in any 160 of the samples (LOD 10 ng/g of food). 161

In experiment 1, animals were penned individually the day before the administrations and for 24h following administrations. For experiment 2, all animals in a same group were kept together and no direct contact was allowed between animals from different groups, with separate food and water supplies provided for each group.

166

#### 167 2.3. Experimental design

168 Experiment 1: relative bioavailability via subcutaneous and dietary routes of administration

This experiment was designed to evaluate the equivalence, in terms of internal exposure to 169 170 BPA, of the S.C. route of administration used for convenience in this experiment and the dietary one thought to be the main route of exposure in human populations. It was conducted on 4 ewes 171 (3-5 year old; 50-73 kg) and was divided into 2 periods conducted at least 13 days apart. The 172 first period was devoted to the S.C. route of administration (5 mg/kg of body weight). During 173 the second period, the same ewes received BPA via the diet (10 mg/kg of body weight). Before 174 the dietary period, ewes were fasted overnight to facilitate intake of the BPA-treated pellets. 175 The ewes ingested the pellets within 2-9 minutes. Free access to hav and water was maintained 176 after the administrations. The different doses for the different routes were based on previous 177 178 studies and were chosen to produce BPA concentrations above our assay limit of quantification, for at least 8-10 hours after the administrations. Use of different doses for the two routes was 179 possible because there is evidence that the parameters of BPA pharmacokinetics in several 180 species including sheep are linear functions of the dose (Collet et al., 2010; Taylor et al., 2011; 181 Tominaga et al., 2006) over a dose range of 5 to 10 mg/kg. Results regarding the dietary route 182 have been described in detail in a previous publication (Guignard et al., 2016). 183

Serial jugular blood samples were collected before all administrations and 0.5 - 1 - 2 - 4 - 6 - 88 - 10 - 24 h after the S.C. administration or 0.03 - 0.08 - 0.16 - 0.25 - 0.5 - 1 - 2 - 4 - 6 - 8- 10 - 26 h after the dietary exposure. For the dietary administration, time 0 was the time when the ewes had finished ingesting the pellets. The blood samples were centrifuged at 3000*g* at 4°C for 20min, then the serum was decanted and stored in propylene tubes at -20°C until assayed.

190 Experiment 2: Effect of environmentally relevant doses of BPA on maternal and fetal thyroid
191 status.

This experiment was conducted on 22 pregnant ewes (1-3 year old; 55-103 kg) artificially 192 inseminated on the same day after hormonal synchronization of estrus. A vitamin and 193 iodine/selenium-enriched mineral supplement (Alimal gestante® Alliance Elevage, Souvigny, 194 France) was provided to cover the specific requirements of gestation. The ewes were weighed 195 twice a month. The amount of pellets was regularly adjusted to pregnancy requirements and 196 changes in body weight. The ewes were randomly allocated to 4 treatment groups balanced for 197 TT4 concentration, body weight and age. From GD28 to GD132-134, each group received 198 different daily doses of BPA by S.C. administration i.e., low dose group 0.5 µg/(kg.d) (n=6); 199 medium dose group 50  $\mu$ g/(kg.d) (n=5); high dose 5000  $\mu$ g/(kg.d) (n=5) and vehicle group 200 201 (n=6). The administered volume was regularly adjusted to the most recently recorded body 202 weight.

203 Maternal blood samples were collected twice a week during the treatment period, just before 204 the next BPA administration. The first maternal blood sample was taken on GD28 before the 205 first BPA or vehicle administration. On GD62, serial blood samples were collected from 206 animals in the high dose treatment group (0 - 1.5 - 4 - 8 and 10 h post administration).

At GD133-135, i.e. about two weeks before the expected term, the fetuses were delivered by cesarean section under rachianesthesia and local anesthesia of the muscle wall (5 mL of lidocaine in the sacrolombar space and 10 mL along the incision line on the abdominal wall; Lurocaine®, Vetoquinol SA, Lure, France) to avoid spontaneous deliveries. Although not totally mature at this age, thyroid function is already well established at this stage in the sheep as in human.

Maternal and fetal jugular blood, cord blood and amniotic fluid were collected at delivery after
which the ewes and fetuses were immediately euthanized by intravenous overdose of sodium
pentobarbital (Doléthal®, Vetoquinol France, 60 mg/kg of pentobarbital i.v.).

217

#### 2.4. <u>BPA</u>, <u>BPA-GLUC</u> and <u>BPA-SULF</u> assay

BPA, BPA-GLUC and BPA-SULF (BPA sulfate) concentrations were simultaneously
measured in the serum samples collected during experiment 1, in the amniotic fluid, cord blood
and maternal and fetal blood samples collected at delivery, and in the serial samples collected
from the high dose group in experiment 2, using a previously described UPLC/MS/MS method
(Lacroix et al., 2011). The validated LOQs were 1 ng/mL for BPA and 5 ng/mL for BPAGLUC/S. The mean intra- and inter-day coefficients of variation for three concentration levels
of BPA and BPA-GLUC were lower than 15%.

- 225
- 226

227

## 2.5. Pharmacokinetic analyses

All pharmacokinetic analyses were performed with the Phoenix® software, version 6.4(Pharsight Corporation, Cary, NC, USA).

#### 230 Experiment 1:

Serum concentration-time profiles of BPA and BPA-GLUC were first analyzed using a non-231 232 compartmental approach. The areas under the BPA and BPA-GLUC serum concentrations curves (BPA and BPA-GLUC AUC<sub>last</sub>) were calculated using the linear trapezoidal rule from 233 dosing time to the last quantifiable (i.e. >LOQ) serum concentration sampling time. The AUC 234 from t=0 to infinity was obtained by adding to AUC<sub>last</sub> the area extrapolated from the last 235 observation to infinity obtained by dividing the last observed quantifiable serum concentration 236 by the slope of the terminal phase estimated by linear regression, using the best fit option of 237 Phoenix®. As previously explained (Gayrard et al., 2013; Guignard et al., 2016), to avoid any 238 confusion due to high concentration resulting from direct drainage of the mouth by the jugular 239

Comment citer ce document : Guignard, D., Gayard, V., Lacroix, M., Puel, S., Picard-Hagen, N., Viguié, C. (Auteur de correspondance) (2017). Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure. Chemosphere, 182, 458-467.

veins, the data obtained during the first 10 minutes after final pellet ingestion were not taken into account in the calculation of BPA AUC<sub>inf</sub>. The digestive absorption of BPA being close to 100% (Doerge et al., 2010), the molar ratio AUC<sub>inf</sub> BPA-GLUC / AUC<sub>inf</sub> BPA could be used as an index of BPA metabolism. The relative bioavailability of BPA by dietary and subcutaneous routes was compared using the following equation:

245 
$$Relative \ bioavailability = \frac{BPA\ AUCinf\ diet}{BPA\ AUCinf\ S.C.} X \frac{BPA\ dose\ S.C.}{BPA\ dose\ diet}$$

The time courses of serum BPA concentrations obtained in the 4 ewes after the S.C. administration were fitted to a triexponential equation, corresponding to an extravascular administration, to generate the pharmacokinetic (PK) parameters required to predict the time course of BPA blood concentrations for the lowest doses in experiment 2:

250

$$C(t) = A.e^{-\alpha.t} + B.e^{-\beta.t} + C.e^{-K01.t}$$

251

## 252 Experiment 2:

Given the known BPA PK parameters and our assay LOQ, the maternal serum concentrations 253 of BPA were likely to be non-quantifiable after repeated S.C. administrations of BPA at 0.5 or 254 50  $\mu$ g/(kg.d). Time-concentration profiles of BPA were therefore simulated, using the PK 255 parameters and the model of experiment 1, to estimate the animal internal exposure. BPA 256 disposition was assumed to be linear overtime, as no accumulation or increase in the 257 metabolism of BPA or BPA-GLUC had been observed with repeated administrations (Collet et 258 al., 2010; Viguié et al., 2013) and because gestation does not affect BPA metabolic rate in the 259 ewe (Corbel et al., 2013). The goodness-of-fit was determined using data obtained from the 260 serial samples collected from ewes receiving the highest dose at GD62. 261

262

#### 263 2.6. <u>TH and TSH assays</u>

Total and free T4and total T3 concentrations were measured in one every two samples 264 collected twice a week in ewes from GD28 to GD128 and from fetal jugular samples collected 265 at delivery using a RIA kit (T4 MAb RIA kit, MP Biomedicals, Illkirch, France). The LOQ of 266 the RIA assays (lowest value with an intra-assay CV lower than 20% and an accuracy above 267 90%) was validated at 10 ng/mL and 0.4 ng/mL for total T4 and T3, respectively. For free T4, 268 a limit of sensitivity was set at the lowest concentration of the standard curve providing that 269 270 the intra-assay CV was below 20% (3.2 pg/mL). The mean intra- and inter-assay CVs for three QC pools were systematically below 10%. 271

272

273 The balance between T4 activation into T3 and its inactivation to rT3, through different deiodinase activities, particularly within the placenta, has a key role in ensuring adequate TH 274 supply to the fetus to meet its needs. We therefore used a previously described UPLC/MS/MS 275 assay (Roques et al., 2012) to simultaneously determine TT4 and rT3 concentrations in the 276 cord blood samples and in maternal samples collected after TT4 modification had been 277 established, at GD 41 - 44 - 48 - 51 - 55 - 58 - 72 - 86 - 100 - 114 and 128. The LOD for 278 each assay run was defined as the calculated concentration corresponding to a peak area at a 279 given elution time, equal to three fold the area obtained by solvent injection. These LOD were 280 2.8, 0.5 and 0.15 ng/mL for TT4, rT3 and TT3, respectively. Mean intra- and inter-assay CVs 281 for three QC pools were systematically below 15% and accuracy was between 88 and 108%. 282 To determine the consistency between RIA and UPLC/MS assays, we assayed TT4 in a pool 283 284 of 43 fetal sheep blood samples from another experiment using both methods. We found a mean inter-method coefficient of variation of 5.2% for concentrations ranging from 53 to 138 285 ng/ml. 286

Comment citer ce document : Guignard, D., Gayard, V., Lacroix, M., Puel, S., Picard-Hagen, N., Viguié, C. (Auteur de correspondance) (2017). Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure. Chemosphere, 182, 458-467.

10.1016/i.chem

287	
288	TSH serum concentration was measured in maternal samples collected at GD $34 - 65 - 97$ and
289	132 using an ELISA kit (Sheep Ultrasensitive TSH ELISA kit®, Blue Gene, Shanghai, China)
290	following the manufacturer's instructions. The lowest measurable concentration was 1 $\mu$ UI/ml.
291	
292	2.7. <u>Statistical analyses</u>
293	Statistical analyses were performed with Systat® (Systat Software Inc., San Jose
294	California,USA).
295	For experiment 1, mean AUC and Cmax for both BPA and BPA-GLUC concentrations were
296	compared using paired T-tests.
297	For experiment 2, the effect of the treatment on the time course of maternal TT4, fT4, TT3 and
298	TSH was analyzed by using a two-way repeated-measures ANOVA, with time, treatment and
299	their interaction as fixed factors and animals nested in the treatment as a random effect factor.
300	A Bonferroni test was performed to identify the dose that individually would give a different
301	serum TH profile to the one observed in the control group.
302	The effect of the treatment on hormone concentrations measured in fetal and maternal blood at
303	delivery was analyzed with a one-way ANOVA.
304	
305	<u>3. Results</u>
306	3.1. Relative bioavailability by subcutaneous and dietary routes of administration
307	(Experiment 1)
308	Fig. 1 (A-B) shows the mean (± S.E.M.) serum BPA and BPA-GLUC concentration-time
309	profiles following S.C. (5 mg/kg) or dietary (10 mg/kg) administration of BPA. The

concentrations were not normalized by the dose. The concentration-time profiles varied greatly 310 between the two routes. With the dietary route, the maximum serum concentration  $(C_{max})$  was 311 attained very quickly (at 0.12 h for 2 ewes and 0.20 h for 2 others) and BPA concentrations 312 decreased quickly to reach the assay LOQ within 8 h post-administration. For the S.C. route, 313 C<sub>max</sub> was obtained later (after2 h for 3 ewes and 1 h for one ewe) and the BPA concentrations 314 decreased more slowly. The mean normalized by the dose BPA  $C_{max}$  for the S.C. route was only 315  $4.6 \pm 1.5$  fold higher than for the dietary route (p=0.07; Fig. 1 C). By contrast to BPA, the 316 BPA-GLUC concentration time course for the two routes of exposure was very similar and the 317 maximum concentration attained with both routes, when normalized by the administered dose, 318 319 was about 2.4 nmol/mL (Fig. 1 D).

320





322 Fig. 1: A-B) mean (± S.E.M.) serum BPA (A) and BPA-GLUC (B) concentration-time profiles

323 obtained for the same ewes (n=4) after subcutaneous (5 mg/kg) and dietary (10 mg/kg)

administrations. Blood samples were collected 0.5 - 1 - 2 - 4 - 6 - 8 - 10 and 24 h after subcutaneous administration and 0.03 - 0.08 - 0.16 - 0.25 - 0.5 - 1 - 2 - 4 - 6 - 8 - 10 and 24 h after dietary administration. The serum concentrations were not normalized by the dose. C-D) Mean ( $\pm$  S.E.M.) maximum concentration of BPA (C) and BPA-GLUC (D) normalized by the BPA dose obtained after subcutaneous and dietary administrations.

329

Administration by S.C. route led to a higher overall internal exposure to BPA characterized by a much higher normalized by the dose AUC<sub>inf</sub> (p=0.002; Fig.2 A). The calculated relative bioavailability of BPA for the dietary as compared to S.C. route was  $3.3 \pm 0.3$  %. The AUC<sub>inf</sub> of BPA-GLUC was very similar for both routes of administration (p=0.11; Fig. 2 B). The ratio of BPA-GLUC to BPA AUC<sub>inf</sub> was about 25 fold higher for the dietary route than for the S.C. route (p=0.002; Fig. 2 C). This can be explained by the extensive hepatic first-pass effect that immediately transforms BPA into BPA-GLUC after BPA intestinal absorption.



337

Comment citer ce document : Guignard, D., Gayard, V., Lacroix, M., Puel, S., Picard-Hagen, N., Viguié, C. (Auteur de correspondance) (2017). Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure. Chemosphere, 182, 458-467.

1016/i.chemosphere.2017.05.028

Fig. 2: A-B) mean (± S.E.M.) AUC<sub>inf</sub> of BPA (A) and BPA-GLUC (B) concentrations
normalized by the BPA dose obtained after subcutaneous (5 mg/kg) and dietary (10 mg/kg)
administrations in the same ewes (n=4). The BPA concentrations obtained during the first 10
minutes after the end of food intake were not taken into account in the AUC estimation. C)
Mean (± S.E.M.) ratio of the BPA to the BPA-GLUC molar dose-normalized AUC<sub>inf</sub> after
subcutaneous and dietary administrations.

344

Overall, the maximum serum concentration of BPA obtained after subcutaneous administration was not dramatically higher than after dietary administration. However, BPA high level exposure was more prolonged after administration by S.C. route than by dietary route. Interestingly, the internal exposure to BPA-GLUC after both routes of administration was almost identical.

350

351 3.2. Maternal and fetal monitoring of BPA and BPA-GLUC internal exposure (Experiment
 352 2)

Table 1 describes the mean BPA, BPA-GLUC and BPA-SULF concentrations measured in the 353 maternal and fetal jugular blood, cord blood and amniotic fluid collected at delivery for the 354 medium and high BPA dose groups. Concentrations were below the assay LOQ in all samples 355 from the control group. BPA concentrations were quantifiable in all collected blood samples 356 357 only for animals in the highest dose group and in the amniotic fluid from the medium and high dose groups. It was quantifiable in cord blood samples from 4/10 and 5/10 lambs from the low 358 and medium dose groups, respectively. BPA-GLUC concentrations could be systematically 359 measured in fetal jugular blood, cord blood and amniotic fluid samples from the medium and 360 high dose groups and in a few fetal jugular (5/10), cord blood (3/10) and amniotic fluid (3/7)361

samples from the low dose group. Overall, BPA-GLUC concentrations in the fetal compartment 362 were much higher than the BPA fetal concentrations or the BPA-GLUC maternal 363 concentrations. BPA was detectable in the maternal blood only for the highest dose group. BPA-364 GLUC in the maternal blood was detectable in a few samples of the low and medium doses 365 groups and was systematically quantified in all the ewes from the high dose group. When 366 quantifiable, BPA maternal concentrations were very similar to the amniotic ones, while for 367 BPA-GLUC amniotic fluid concentrations were much higher than the maternal ones and within 368 the same range of the fetal ones. 369

BPA-SULF could be measured in samples from the fetal compartment (jugular fetal and cord blood) in all samples from the medium and high dose groups. For the amniotic fluid and maternal samples, it was rarely measurable in samples from the medium dose group. In the fetal and cord blood samples from the high dose group, BPA-SULF was detected at levels about 50 to 80 fold higher than BPA and about 20 fold lower than BPA-GLUC. BPA-SULF concentration in amniotic fluid were about 10 fold lower than that in the fetus while they were about 10 fold higher than that of the maternal compartment.

Table 1: mean (± SEM) BPA and BPA-GLUC/S concentrations (nmol/mL) measured in the 377 maternal and fetal jugular blood, cord blood and amniotic fluid after treatment of pregnant ewes 378 with BPA from GD 28 to 132-134. LOQ were validated at 1 ng/mL and 5 ng/ml for BPA and 379 BPA-GLUC/S, respectively (4.4, 12.4 and 16.2nmol/L for BPA, BPA-GLUC and BPA-SULF, 380 respectively). Mean concentration were noted as "<LOQ" when the concentrations in more than 381 half the samples were below LOQ. When BPA and/or its metabolites were measurable in at 382 least half the samples, the non-measurable values were replaced by the LOQ/2 for the mean 383 calculation. 384

		Cord blood			Fetal blood			Amniotic fluid			Maternal blood		
		BPA	BPA- GLUC	BPA- SULF	BPA	BPA- GLUC	BPA- SULF	BPA	BPA- GLUC	BPA- SULF	BPA	BPA- GLUC	BPA- SULF
Medium	Mean	0.007	1.180	0.046	<loq< td=""><td>1.09</td><td>0.040</td><td>0.012</td><td>0.284</td><td><loq< td=""><td><loq< td=""><td>0.022</td><td><loq< td=""></loq<></td></loq<></td></loq<></td></loq<>	1.09	0.040	0.012	0.284	<loq< td=""><td><loq< td=""><td>0.022</td><td><loq< td=""></loq<></td></loq<></td></loq<>	<loq< td=""><td>0.022</td><td><loq< td=""></loq<></td></loq<>	0.022	<loq< td=""></loq<>
$\begin{array}{c} \text{dose} \\ 50 \ \mu\text{g}/(\text{kg.d}) \\ 0 \ 2 \end{array}$	S.E.M.	0.002	0.077	0.01		0.058	0.006	0.005	0.034			0.008	
μmol/(kg.d)	n/tot	5/10	10/10	10/10	2/10	10/10	10/10	7/8	8/8	3/8	0/5	3/5	2/5
High dose	Mean	0.106	95.873	5.277	0.058	95.698	4.555	0.259	28.041	0.502	0.236	1.624	0.056
5000 μg/(kg.d) 20	S.E.M.	0.012	11.540	0.766	0.005	14.422	0.454	0.055	2.582	0.206	0.030	0.153	0.005
μmol/(kg.d)	n/tot	8/8	8/8	8/8	8.8	8/8	8/8	8/8	8/8	8/8	5/5	5/5	5/5

385

Comment citer ce document : Guignard, D., Gayard, V., Lacroix, M., Puel, S., Picard-Hagen, N., Viguié, C. (Auteur de correspondance) (2017). Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure. Chemosphere, 182, 458-467. DOI : 10.1016/i.chemosphere.2017.05.028

Fig. 3 shows the predicted BPA serum concentrations in ewes treated with BPA (0.5 - 50 or)386 5000 µg/(kg.d); S.C.) using a bi-compartmental model based on the PK parameters generated 387 during experiment 1. The values for BPA serum concentrations measured in a representative 388 ewe from the 5000 µg/(kg.d) group at GD 62 were superimposed on the prediction curve and 389 showed a good fitting of the data by our model. BPA serum concentrations were predicted to 390 fluctuate between 2 – 32 pg/mL (8.8 pmol/L-0.14 nmol/L), between 0.2- 3.2ng/mL (0.88-14 391 nmol/L) and between 20 - 320 ng/mL (88-1400 nmol/L) for the BPA doses of 0.5, 50 and 5000 392 393  $\mu g/(kg.d)$ , respectively.



394

**Fig. 3:** predicted serum concentrations of BPA in pregnant ewes treated with repeated subcutaneous administrations of BPA (0.5 - 50 and  $5000 \ \mu g/(kg.d)$ ). Concentrations were predicted using a bi-compartmental model and appropriate pharmacokinetic parameters previously established in the ewe for a dose of 5 mg/(kg.d). Observed concentrations are the BPA serum concentrations measured in blood samples collected from one representative ewe treated with BPA ( $5000 \ \mu g/(kg.d)$ ; S.C.) from GD 28 to 132-134. Blood samples were collected at GD 62 at time 0 - 1.5 - 4 - 8 and 10 h after the S.C. administration.

402

#### 403 3.3. Maternal and fetal thyroid status (Experiment 2)

Fig. 4 shows the time course of mean totalT4, free T4and total T3 concentrations measured in the maternal serum samples throughout the treatment period for each BPA group compared individually to the control group. For total T4, no significant interaction between time and treatment could be evidenced (p=0.37).

The time course of maternal free T4 serum concentrations appeared to be affected by the treatments (interaction time x treatment, p<0.001). A significant difference between the control group and the medium (p=0.003) and high (p=0.015) dose groups was observed which was characterized by a decrease in fT4 concentrations. Visual inspection of the graph indicates that this decrease was initiated around GD 40 (i.e. after about 12 days of treatment) in BPA-treated animals. The time course of the ratio of fT4 to TT4 concentrations in maternal blood was not modified by the treatment (interaction time\* treatment, p=0.7).

Throughout pregnancy, the time course of TT3 concentrations in the mothers was significantly affected by the treatment (interaction time x treatment, p=0.04). The profile of the medium dose group differed from that of the control group (p=0.03). Overall, TT3 concentrations were lower in BPA-treated animals from GD 60-70 (visual inspection). This interaction was not significant for the high dose group (p=0.44).

420 No effect of BPA at any dose on serum TSH concentrations (Fig. 5) could be evidenced in
421 maternal blood samples collected at GD 34, 65,97 and 132 (interaction time \* treatment, p=0.23
422 and treatment effect p=0.19).



.

Version postprint

**Fig. 4:** time course of mean ( $\pm$  SEM) total T4, free T4 and total T3 serum concentrations in pregnant ewes treated subcutaneously with vehicle (n=6) or BPA (0.5 (µg/(kg.d), n=6). 50 µg/(kg.d), n=5 or 5000 µg/(kg.d), n=5) from GD 28 to 132-134. Blood samples were collected on GD 28 – 30 – 37 – 44 – 51 – 58 – 65 – 72 – 79 – 86 – 93 – 100 – 107 – 114 – 121 and 128. Time \* treatment significant interactions (P<0.05) were evidenced in several instances signifying that the time course of the hormones differed between the BPA and vehicle-treated animals.

431

- 432
- 433



435 **Fig. 5:** mean ( $\pm$  SEM) serum TSH concentrations in pregnant ewes treated subcutaneously with 436 vehicle (n=6) or BPA S.C. (0.5 µg/(kg.d), n=6; 50 µg/(kg.d), n=5 or 5000 µg/(kg.d), n=5) from 437 GD 28 to 132-134. Blood samples were collected on GD 34 – 65 – 97 and 132. No significant 438 effect of BPA treatments could be evidenced.

439

434

Disruption of the maternal thyroid status was further characterized by measuring rT3 jointly to 440 TT4 using HPLC/MS in samples collected from the 41<sup>st</sup> day of gestation, i.e. after fT4 441 concentrations have started to decrease in the medium dose group. A treatment effect could be 442 evidenced on TT4 concentrations with no interaction with the time. On rT3 profiles, a trend 443 444 (p=0.09) toward an interaction between time and treatment was evidenced suggesting that modifications of the rT3 patterns might occur later than for T4. The time course of the rT3/TT4 445 ratio (Fig. 6) was significantly affected by BPA exposure (interaction time x treatment p=0.04) 446 over the period considered. These modifications were due to differences between the control 447 and medium dose groups with a time-dependent effect on the rT3 to TT4 ratio (p=0.024) 448 characterized by a higher rT3 to TT4 ratio in the BPA group from GD50 to 90, approximately. 449



450

**Fig. 6:** time course of mean ( $\pm$  S.E.M.) ratio (%) of rT3 to total T4 serum concentrations measured by HPLC/MS in pregnant ewes treated subcutaneously with control or BPA 0.5 – 50 and 5000 µg/(kg.d) from GD 40 to 132-134. Blood samples were collected on GD 41 – 44 – 48 – 51 – 55 – 58 – 72 – 86 – 100 – 114 and 128. A time \* treatment significant interaction was evidenced only in the groupe treated with the middle dose (p=0.024) signifying that the time course of the rT3/T4 ratio in this group differed from vehicle animals.

458 Neither TT4 nor fT4 in fetal jugular blood (Fig. 7) were affected by BPA treatments at any of 459 the doses tested (individual comparison with the control group, p>0.05). TT3 was never 460 detectable in fetal jugular blood.

461



463 <u>Fig. 7:</u> mean ( $\pm$  S.E.M.) free (A) and total (B) T4 serum concentrations measured by a RIA 464 method in fetal jugular blood collected at GD132-134. Fetuses came from mothers treated with 465 vehicle or BPA 0.5 – 50 and 5000 µg/(kg.d) from GD 28 to 132-134 S.C. Data were analyzed 466 using a one factor ANOVA. No significant effect of the treatments could be evidenced (p>0.05).

467

462

The thyroid hormone concentrations in cord blood were measured by HPLC/MS. TT3 in the fetuses was almost always below our assay LOD. No effect of the treatment on rT3, TT4 concentrations and rT3/TT4 ratio could be evidenced in cord blood.

471

#### 472 **4.** <u>Discussion</u>

Comment citer ce document : Guignard, D., Gayard, V., Lacroix, M., Puel, S., Picard-Hagen, N., Viguié, C. (Auteur de correspondance) (2017). Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis in the pregnant ewe at low level representative of human exposure. Chemosphere, 182, 458-467. DOI : 10.1016/i.chemosphere.2017.05.028

Our study is one of the rare studies to directly assess the equivalence between a route of 473 exposure commonly used in experimental studies of BPA toxicity and the dietary route 474 acknowledged as the main route of exposure for human populations. Our findings suggest that 475 the bioavailability of BPA might be 30 fold lower when administered by dietary route than by 476 subcutaneous route. Our current results indicate that the difference between the two routes of 477 exposure is mainly related to the overall BPA exposure and much less to the peak serum 478 concentrations. Our results also show that BPA-GLUC exposure, unlike that of BPA, is very 479 similar for both routes of administration. BPA-GLUC is usually assumed to be devoid of 480 biological activity. However, in some sensitive tissues, the balance between BPA detoxification 481 by conjugation into BPA-GLUC and BPA-GLUC reactivation by deconjugation into BPA is in 482 favor of deconjugation (Corbel et al., 2014). Although the mechanism enabling BPA-GLUC (a 483 hydrophilic compound) to reach the intracellular space remains unknown, these observations 484 485 indicate that internal exposure to BPA metabolite might also be of biological significance and should be more thoroughly considered in toxicological studies. Furthermore, it is noteworthy 486 that BPA-GLUC blood profiles differ markedly after gavage administration as compared to the 487 dietary one (Guignard et al., 2016). Thus when considering potential effects of BPA-GLUC, 488 gavage might not be better than the subcutaneous route for evaluating BPA oral toxicity. 489

Our TK model enabled us to estimate the internal exposure of ewes treated by chronic exposure 490 via the subcutaneous route. Interestingly, both the medium and low doses selected were 491 predicted to reproduce BPA serum concentrations in ewes within the range of concentrations 492 commonly reported in the most recent studies in pregnant women i.e., from 100 pg/mL 493 (Teeguarden et al., 2016) to 4 ng/mL (Shekhar et al., 2016). However, in both those studies, 494 free BPA would account for about 60-80% of total BPA while in our study the ratio of BPA-495 GLUC to BPA was markedly in favor of BPA-GLUC in both the mother and the fetus at 496 delivery, i.e. at least 24 h after the last BPA administration. 497

Concerning fetal exposure, our exposure schemes made it possible to encompass the range of 498 499 aglycone BPA concentrations reported in fetal compartments in humans, whether in amniotic fluid or cord blood. Indeed, for BPA, the geometric mean of about 4.8 ng/mL in the amniotic 500 fluid of full-term human newborns (Shekhar et al., 2016) is similar to the concentrations 501 measured in the amniotic fluid of lambs from the medium dose group in our study. Similarly, 502 cord blood concentrations in human newborns (about 1.2 ng/mL; Chevalier et al., 2015) are 503 very close to the few measurable values obtained in the cord blood of lambs from our medium 504 dose group. Our study evidenced a high accumulation rate of BPA-GLUC in the fetal 505 compartment in agreement with other study with short term exposure. Indeed, in our current 506 507 study, as in the short term fetal exposure study performed at GD120 (Corbel et al., 2013), amniotic fluid concentrations were about 4 fold lower than concentration in fetal blood (jugular 508 or cord blood). However, in a first study with chronic gestational exposure (Viguié et al., 2013), 509 510 BPA-GLUC concentrations in full term lambs were 2 fold higher in amniotic fluid than in fetal blood. The most likely explanation for such fetal age –dependent differences in fetal exposure 511 to BPA-GLUC could rely on the ontogeny of UGT activity in the fetal sheep with a late rise 512 toward the end of pregnancy (Pretheeban et al., 2011) similarly to human (Divakaran et al., 513 2014). The biological significance of the very high fetal exposure to BPA-GLUC produced in 514 515 our sheep model and its relevance to humans remains to be elucidated. From this standpoint, it is noteworthy that some data in monkeys suggest that, similarly to what is observed in the ovine 516 fetus, BPA-GLUC accumulates in the fetal compartment (Patterson et al., 2013). 517

518 BPA-SULF appeared to accumulate as well in the fetal compartment. However, it is noteworthy 519 that it does not follow the same pattern in the fetal compartment than BPA-GLUC. BPA-SULF 520 concentrations in the amniotic fluid are indeed about 10 fold lower than in the fetal 521 compartment. This suggests that BPA-SULF might not be eliminated from the fetal blood

through the kidney and thus the amniotic fluid or that BPA-SULF is very rapidly and efficientlycleared from the amniotic fluid.

Our results concerning the thyroid status of the mother reveal a certain degree of thyroid 524 disruption oriented toward a decrease in concentrations of the circulating hormones, fT4 and 525 TT3. The medium dose (50  $\mu$ g/(kg.d)) seemed to be pivotal as this level of BPA treatment 526 significantly affected the time courses of both hormones. Interestingly, the time course of TT3, 527 the biologically active hormone, tended to be significantly modified by our lowest dose (0.5 528  $\mu g/(kg.d)$ ). This suggests that pregnant females might be sensitive to BPA-induced thyroid 529 disruption even at environmentally-relevant exposure. Our study did not provide an assessment 530 of the minimal active BPA dose. It is noteworthy however that the effect observed at our lowest 531 dose are limited to a trend in TT3 decrease while both fT4 and TT3 were modified with the 532 middle dose. Although our highest dose (5000 µg/(kg.d)) enabled us to reproduce the 533 hypothyroxinemia described in a previous study, surprisingly, modifications of the time course 534 were only observed for fT4 and not for TT4. The fact that BPA induced a disrupting effect on 535 some thyroid parameters at the medium and/or low doses but not at the high dose suggests a 536 non-monotonic effect of BPA on thyroid function in pregnant animals. Nevertheless, this 537 interpretation needs to be treated with caution and more doses and a larger number of animals 538 would be required to evidence and characterize such a complicated dose-response relationship. 539

No effect of BPA treatments on fetal thyroid status could be evidenced in cord and jugular blood samples. This contrasts with our previous results in sheep (Viguié et al., 2013). One major difference between this study and the previous study in sheep is related to the gestational stage and fetal age and/or the different fetal exposure to BPA/BPA-GLUC. In our earlier experiment, the lambs were extracted at term, whereas in the current experiment they were extracted 10 to 14 days before term. Thus, the final stage of thyroid maturation might not yet have occurred in these animals. This explanation is fully consistent with the fact that TT3 could not be detected

in the fetal blood samples. Indeed, TT3 concentration in the blood of sheep fetuses, as well as
human fetuses (Fisher, 2002), is only detectable in the later stages of pregnancy (the last week
for sheep; (Fisher et al., 1994). It is noteworthy that, in rodents, BPA-induced thyroid disruption
due to maternal exposure was mainly evidenced during the postnatal period, which corresponds
in this species to the final stage of maturation of thyroid function (Xu et al., 2007; Zoeller et
al., 2005), i.e., equivalent to the two missing weeks required, in the current experiment, for the
ewes to attain full term pregnancy.

Another explanation for such a discrepancy might arise from the difference in mineral supply 554 provided to the animal. Indeed, in the current experiment, ewes received a pellet ration 555 containing a mineral supply adapted to pregnancy requirements and thus enriched in mineral 556 iodine (calcium iodate about 1.5 mg/day) and selenium (sodium selenite about 0.62 mg/day). 557 Iodine supply might have somehow compensated at least in part the BPA-induced thyroid 558 disruptive process or made the fetal thyroid function less sensitive to thyroid disruption. This 559 kind of relationship between sensitivity to thyroid disruption and iodine status is supported by 560 epidemiological observations in humans. The inverse maternal urinary BPA /newborn girl TSH 561 relationship was shown to depend, at least in part, on the maternal iodine status in the human 562 biomonitoring "HOME" survey (Romano et al., 2015), suggesting that the maternal iodine 563 supply might somehow affect the sensitivity of the developing fetus to chemical thyroid 564 disruption. Other examples of interaction between the iodine status and diverse thyroid stressors 565 have been found in humans such as mercury in pregnant women (Llop et al., 2015) or 566 perfluoroalkyl acids in anti-TPO antibody positive humans (Webster et al., 2016). 567

The mechanisms by which BPA exposure induces maternal thyroid disruption remain unclear. From our results, we could not get any evidence that either TSH secretion or TH binding to carrier serum proteins might be substantially altered in BPA-treated ewes. The absence of significant modifications of TSH in our BPA-treated pregnant ewes should not be considered

as an ultimate proof that the thyroid axis alteration were of no potential functional 572 573 consequences. Disruption of thyroid homeostasis indeed occurs in the absence of modification of blood TSH in so called states of hypothyroxinemia (decreased free-T4 concentration with or 574 without modifications of TSH). There are some experimental evidences that such maternal 575 hypothyroxinemia might be of detrimental consequences on fetal neural development and as so 576 should not be regarded as trivial (Morreale de Escobar et al., 2000; Lavado-Autric et al., 2003; 577 Gilbert et al., 2012). Accordingly, some epidemiological surveys highlight a link between 578 maternal hypothyroxinemia during pregnancy and increased risk of autism impaired 579 psychomotor development, Attention Deficit Hyperactivity Disorder, language delay, lower 580 581 mental scores (Zoeller et al., 2002; Henrichs et al., 2013; Román et al., 2013).

The balance between deiodinase activities converting T4 into either the bioactive T3 or 582 inactiverT3 is a key factor in the maintenance of fetal thyroid homeostasis. This is why we 583 584 measured rT3 and T4 concentrations in cord blood and in some bi-weekly maternal samples, using MS detection, in an attempt to gain new insights into the balance between T4 and rT3 and 585 the potential mechanisms of BPA action. The results obtained with MS in maternal samples, 586 collected from GD 41 to 128, suggest that a modification of the deiodination balance, 587 characterized by a time-dependent modulation of the rT3/TT4 ratio, occurred in mothers 588 589 receiving the medium BPA dose compared to those in the vehicle group. The placenta is a key actor in fetal thyroid balance. It not only expresses deiodinases, in particular type 3, to produce 590 a balance between the active and inactive thyroid hormone supply to the fetus, it also 591 592 synthesizes transthyretin which regulates the transplacental transfer of maternal thyroxine to the fetal compartments (Landers et al., 2009) and regulates iodide exchanges through NIS (Na/I 593 symporter) expression. In vitro data suggest that the placenta is a sensitive target tissue for BPA, 594 including at environmentally relevant doses (Mannelli et al., 2015; Sieppi et al., 2016; Tait et 595 al., 2015). However, no evaluation of the impact of BPA on thyroid-relevant placental functions 596

are currently available and there is no evidence that BPA can alter deiodinase expression and/or activity in any tissues. In the light of our current results indicating that in pregnant ewes BPA exposure modifies the balance between T4 and rT3 at a late stage of pregnancy, it could be very interesting to assess the effect of BPA on placental deiodinase activities.

601

#### 602 **5.** <u>Conclusion</u>

603 Our study carried out in an animal model relevant to humans in terms of thyroid and gestation physiologies shows that BPA can affect maternal thyroid function and equilibrium during 604 605 pregnancy. This effect was expressed at levels of internal exposure (as assessed by serum 606 aglycone BPA concentrations) similar to those described in humans i.e. for BPA blood concentrations below the ng/ml range. Although within the framework of this study, these 607 maternal modifications could not be tied with alterations in some patterns of fetal TH serum 608 concentration, the possibility remains that TH action and concentrations in target tissues, 609 particularly in the brain, might have been affected. Interestingly, our data suggest that BPA 610 exposure might somehow modify the deiodination balance, thus pinpointing a possible effect 611 of BPA on the thyroid-related endocrine function of the placenta. 612

613

614

# 615 Acknowledgements

We thank the INRA experimental unit of Domaine de Lafage for providing the animals and thestaff from the ENVT sheep experimental unit for their involvement in animal care.

This research did not receive any specific grant from funding agencies in the public,
commercial, or not-for-profit sectors. D Guignard beneficiated from PhD Grant of the doctoral
School SEVAB from the Institut National Polytechnique of Toulouse, France.

621

#### 622 **References**

- Chapin, R.E., Adams, J., Boekelheide, K., Gray, L.E., Hayward, S.W., Lees, P.S.J., McIntyre,
  B.S., Portier, K.M., Schnorr, T.M., Selevan, S.G., Vandenbergh, J.G., Woskie, S.R.,
  2008. NTP-CERHR expert panel report on the reproductive and developmental toxicity
  of bisphenol A. Birth Defects Res. B. Dev. Reprod. Toxicol. 83, 157–395.
  doi:10.1002/bdrb.20147
- Chevalier, N., Brucker-Davis, F., Lahlou, N., Coquillard, P., Pugeat, M., Pacini, P., PanaïaFerrari, P., Wagner-Mahler, K., Fénichel, P., 2015. A negative correlation between
  insulin-like peptide 3 and bisphenol A in human cord blood suggests an effect of
  endocrine disruptors on testicular descent during fetal development. Hum. Reprod. 30,
  447–453. doi:10.1093/humrep/deu340
- Chevrier, J., Gunier, R.B., Bradman, A., Holland, N.T., Calafat, A.M., Eskenazi, B., Harley,
  K.G., 2013. Maternal urinary bisphenol a during pregnancy and maternal and neonatal
  thyroid function in the CHAMACOS study. Environ. Health Perspect. 121, 138–144.
  doi:10.1289/ehp.1205092
- Collet, S.H., Picard-Hagen, N., Viguié, C., Lacroix, M.Z., Toutain, P.-L., Gayrard, V., 2010.
  Estrogenicity of Bisphenol A: A Concentration-Effect Relationship on Luteinizing
  Hormone Secretion in a Sensitive Model of Prepubertal Lamb. Toxicol. Sci. 117, 54–
  62. doi:10.1093/toxsci/kfq186
- 641 Corbel, T., Gayrard, V., Puel, S., Lacroix, M.Z., Berrebi, A., Gil, S., Viguié, C., Toutain, P.-L.,
- 642 Picard-Hagen, N., 2014. Bidirectional placental transfer of Bisphenol A and its main

- 643 metabolite, Bisphenol A-Glucuronide, in the isolated perfused human placenta. Reprod.
- 644 Toxicol. 47, 51–58. doi:10.1016/j.reprotox.2014.06.001
- 645 Corbel, T., Gayrard, V., Viguié, C., Puel, S., Lacroix, M.Z., Toutain, P.-L., Picard-Hagen, N.,
- 646 2013. Bisphenol A Disposition in the Sheep Maternal-Placental-Fetal Unit:
  647 Mechanisms Determining Fetal Internal Exposure. Biol. Reprod. 89, 11.
  648 doi:10.1095/biolreprod.112.106369
- 649 Doerge, D.R., Twaddle, N.C., Vanlandingham, M., Fisher, J.W., 2010. Pharmacokinetics of
  650 bisphenol A in neonatal and adult Sprague-Dawley rats. Toxicol. Appl. Pharmacol. 247,
- 651 158–165. doi:10.1016/j.taap.2010.06.008
- Fini, J.-B., Le Mevel, S., Turque, N., Palmier, K., Zalko, D., Cravedi, J.-P., Demeneix, B.A.,
  2007. An in vivo multiwell-based fluorescent screen for monitoring vertebrate thyroid
  hormone disruption. Environ. Sci. Technol. 41, 5908–5914.
- Fisher, D.A., 2002. Endocrinology of fetal development, in: Williams Textbook of
  Endocrinology. Saunders, pp. 811–842.
- Fisher, D.A., Dussault, J.H., Sack, J., Chopra, I.J., 1976. Ontogenesis of hypothalamic-pituitary--thyroid function and metabolism in man, sheep, and rat. Recent Prog. Horm.
  Res. 33, 59–116.
- Fisher, D.A., Polk, D.H., Wu, S.Y., 1994. Fetal thyroid metabolism: a pluralistic system.
  Thyroid 4, 367–371.
- Gayrard, V., Lacroix, M.Z., Collet, S.H., Viguié, C., Bousquet-Melou, A., Toutain, P.-L.,
  Picard-Hagen, N., 2013. High bioavailability of bisphenol A from sublingual exposure.
  Environ. Health Perspect. 121, 951–956. doi:10.1289/ehp.1206339
- Gilbert, M.E., Rovet, J., Chen, Z., Koibuchi, N., 2012. Developmental thyroid hormone
  disruption: Prevalence, environmental contaminants and neurodevelopmental
  consequences. Neurotoxicology 33, 842–852.

- Goto, Y., Kitamura, S., Kashiwagi, K., Oofusa, K., Tooi, O., Yoshizato, K., Sato, J., Ohta, S.,
  Kashiwagi, A., 2006. Suppression of amphibian metamorphosis by bisphenol A and
  related chemical substances. J. Health Sci. 52, 160–168. doi:10.1248/jhs.52.160
- 671 Guignard, D., Gauderat, G., Gayrard, V., Lacroix, M.Z., Picard-Hagen, N., Puel, S., Toutain,
- P.-L., Viguié, C., 2016. Characterization of the contribution of buccal absorption to
  internal exposure to bisphenol A through the diet. Food Chem. Toxicol.
  doi:10.1016/j.fct.2016.04.004
- Hard, G.C., 1998. Recent developments in the investigation of thyroid regulation and thyroid
  carcinogenesis. Environ. Health Perspect. 106, 427–436.
- Heimeier, R.A., Das, B., Buchholz, D.R., Shi, Y.-B., 2009. The Xenoestrogen Bisphenol A
  Inhibits Postembryonic Vertebrate Development by Antagonizing Gene Regulation by
  Thyroid Hormone. Endocrinology 150, 2964–2973. doi:10.1210/en.2008-1503
- Henrichs, J., Ghassabian, A., Peeters, R.P., Tiemeier, H., 2013. Maternal hypothyroxinemia
  and effects on cognitive functioning in childhood: how and why? Clin. Endocrinol.
  (Oxf.) 79, 152–162. doi:10.1111/cen.12227
- Iwamuro, S., Sakakibara, M., Terao, M., Ozawa, A., Kurobe, C., Shigeura, T., Kato, M.,
  Kikuyama, S., 2003. Teratogenic and anti-metamorphic effects of bisphenol A on
  embryonic and larval Xenopus laevis. Gen. Comp. Endocrinol. 133, 189–198.
  doi:10.1016/S0016-6480(03)00188-6
- 687 Kaneko, M., Okada, R., Yamamoto, K., Nakamura, M., Mosconi, G., Polzonetti-Magni, A.M.,
- Kikuyama, S., 2008. Bisphenol A acts differently from and independently of thyroid
  hormone in suppressing thyrotropin release from the bullfrog pituitary. Gen. Comp.
  Endocrinol. 155, 574–580. doi:10.1016/j.ygcen.2007.09.009
- 691 Lacroix, M.Z., Puel, S., Collet, S.H., Corbel, T., Picard-Hagen, N., Toutain, P.L., Viguié, C.,
- 692 Gayrard, V., 2011. Simultaneous quantification of bisphenol A and its glucuronide

metabolite (BPA-G) in plasma and urine: applicability to toxicokinetic investigations.

694	Talanta 85, 2053–2059. doi:10.1016/j.talanta.2011.07.040
695	Landers, K.A., McKinnon, B.D., Li, H., Subramaniam, V.N., Mortimer, R.H., Richard, K.,
696	2009. Carrier-Mediated Thyroid Hormone Transport into Placenta by Placental
697	Transthyretin. J. Clin. Endocrinol. Metab. 94, 2610-2616. doi:10.1210/jc.2009-0048
698	Lavado-Autric, R., Ausó, E., García-Velasco, J.V., del Carmen Arufe, M., Escobar del Rey, F.,
699	Berbel, P., Morreale de Escobar, G., 2003. Early maternal hypothyroxinemia alters
700	histogenesis and cerebral cortex cytoarchitecture of the progeny. J. Clin. Invest. 111,
701	1073–1082. doi:10.1172/JCI16262
702	Llop, S., Lopez-Espinosa, MJ., Murcia, M., Alvarez-Pedrerol, M., Vioque, J., Aguinagalde,
703	X., Julvez, J., Aurrekoetxea, J.J., Espada, M., Santa-Marina, L., Rebagliato, M.,
704	Ballester, F., 2015. Synergism between exposure to mercury and use of iodine
705	supplements on thyroid hormones in pregnant women. Environ. Res. 138, 298-305.
706	doi:10.1016/j.envres.2015.02.026
707	Mannelli, C., Ietta, F., Avanzati, A.M., Skarzynski, D., Paulesu, L., 2015. Biological Tools to
708	Study the Effects of Environmental Contaminants at the Feto-Maternal Interface. Dose-
709	Response 13. doi:10.1177/1559325815611902
710	Meeker, J.D., Calafat, A.M., Hauser, R., 2010. Urinary Bisphenol A Concentrations in Relation
711	to Serum Thyroid and Reproductive Hormone Levels in Men from an Infertility Clinic.
712	Environ. Sci. Technol. 44, 1458–1463. doi:10.1021/es9028292
713	Meeker, J.D., Ferguson, K.K., 2011. Relationship between Urinary Phthalate and Bisphenol A
714	Concentrations and Serum Thyroid Measures in U.S. Adults and Adolescents from the

- 715 National Health and Nutrition Examination Survey (NHANES) 2007–2008. Environ.
- 716 Health Perspect. 119, 1396–1402. doi:10.1289/ehp.1103582

693

717	Moriyama, K., Tagami, T., Akamizu, T., Usui, T., Saijo, M., Kanamoto, N., Hataya, Y.,
718	Shimatsu, A., Kuzuya, H., Nakao, K., 2002. Thyroid hormone action is disrupted by
719	bisphenol A as an antagonist. J. Clin. Endocrinol. Metab. 87, 5185-5190.
720	doi:10.1210/jc.2002-020209
721	Morreale de Escobar, G., Obregon, M.J., Escobar del Rey, F., 2000. Is Neuropsychological
722	Development Related to Maternal Hypothyroidism or to Maternal Hypothyroxinemia?
723	J. Clin. Endocrinol. Metab. 85, 3975–3987.
724	Patterson, T.A., Twaddle, N.C., Roegge, C.S., Callicott, R.J., Fisher, J.W., Doerge, D.R., 2013.
725	Concurrent determination of bisphenol A pharmacokinetics in maternal and fetal rhesus
726	monkeys. Toxicol. Appl. Pharmacol. 267, 41-48. doi:10.1016/j.taap.2012.12.006
727	Polk, D., Cheromcha, D., Reviczky, A., Fisher, D.A., 1989. Nuclear thyroid hormone receptors:
728	ontogeny and thyroid hormone effects in sheep. Am. J. Physiol. 256, E543-549.
729	Richter, C.A., Birnbaum, L.S., Farabollini, F., Newbold, R.R., Rubin, B.S., Talsness, C.E.,
730	Vandenbergh, J.G., Walser-Kuntz, D.R., vom Saal, F.S., 2007. In vivo effects of
731	bisphenol A in laboratory rodent studies. Reprod. Toxicol. Elmsford NY 24, 199.
732	Román, G.C., Ghassabian, A., Bongers-Schokking, J.J., Jaddoe, V.W.V., Hofman, A., de Rijke,
733	Y.B., Verhulst, F.C., Tiemeier, H., 2013. Association of gestational maternal
734	hypothyroxinemia and increased autism risk. Ann. Neurol. 74, 733-742.
735	doi:10.1002/ana.23976
736	Romano, M.E., Webster, G.M., Vuong, A.M., Thomas Zoeller, R., Chen, A., Hoofnagle, A.N.,
737	Calafat, A.M., Karagas, M.R., Yolton, K., Lanphear, B.P., Braun, J.M., 2015.
738	Gestational urinary bisphenol A and maternal and newborn thyroid hormone
739	concentrations: The HOME Study. Environ. Res. 138, 453-460.

doi:10.1016/j.envres.2015.03.003 740

- 741 Roques, B.B., Lacroix, M.Z., Puel, S., Gayrard, V., Picard-Hagen, N., Jouanin, I., Perdu, E.,
- Martin, P.G., Viguié, C., 2012. CYP450-dependent biotransformation of the insecticide
  fipronil into fipronil sulfone can mediate fipronil-induced thyroid disruption in rats.
  Toxicol. Sci. Off. J. Soc. Toxicol. 127, 29–41. doi:10.1093/toxsci/kfs094
- Rubin, B.S., 2011. Bisphenol A: An endocrine disruptor with widespread exposure and multiple
  effects. J. Steroid Biochem. Mol. Biol. 127, 27–34. doi:10.1016/j.jsbmb.2011.05.002
- Shekhar, S., Sood, S., Showkat, S., Lite, C., Chandrasekhar, A., Vairamani, M., Barathi, S.,
  Santosh, W., 2016. Detection of phenolic endocrine disrupting chemicals (EDCs) from
  maternal blood plasma and amniotic fluid in Indian population. Gen. Comp. Endocrinol.
  doi:10.1016/j.ygcen.2016.05.025
- Sheng, Z.-G., Tang, Y., Liu, Y.-X., Yuan, Y., Zhao, B.-Q., Chao, X.-J., Zhu, B.-Z., 2012. Low
  concentrations of bisphenol a suppress thyroid hormone receptor transcription through
  a nongenomic mechanism. Toxicol. Appl. Pharmacol. 259, 133–142.
  doi:10.1016/j.taap.2011.12.018
- Sieppi, E., Vähäkangas, K., Rautio, A., Ietta, F., Paulesu, L., Myllynen, P., 2016. The
  xenoestrogens, bisphenol A and para-nonylphenol, decrease the expression of the
  ABCG2 transporter protein in human term placental explant cultures. Mol. Cell.
  Endocrinol. 429, 41–49. doi:10.1016/j.mce.2016.03.034
- Sriphrapradang, C., Chailurkit, L., Aekplakorn, W., Ongphiphadhanakul, B., 2013. Association
   between bisphenol A and abnormal free thyroxine level in men. Endocrine 44, 441–447.
   doi:10.1007/s12020-013-9889-y
- Sun, H., Shen, O.-X., Wang, X.-R., Zhou, L., Zhen, S., Chen, X., 2009. Anti-thyroid hormone 762 activity of bisphenol A, tetrabromobisphenol A and tetrachlorobisphenol A in an 763 improved Toxicol. Vitro 950-954. 764 reporter gene assay. In 23, doi:10.1016/j.tiv.2009.05.004 765

- Tait, S., Tassinari, R., Maranghi, F., Mantovani, A., 2015. Bisphenol A affects placental layers
  morphology and angiogenesis during early pregnancy phase in mice. J. Appl. Toxicol.
  JAT 35, 1278–1291. doi:10.1002/jat.3176
- 769 Taylor, J.A., vom Saal, F.S., Welshons, W.V., Drury, B., Rottinghaus, G., Hunt, P.A., Toutain,
- P.-L., Laffont, C.M., VandeVoort, C.A., 2011. Similarity of Bisphenol A
  Pharmacokinetics in Rhesus Monkeys and Mice: Relevance for Human Exposure.
  Environ. Health Perspect. 119, 422–430. doi:10.1289/ehp.1002514
- Teeguarden, J.G., Twaddle, N.C., Churchwell, M.I., Doerge, D.R., 2016. Urine and serum
  biomonitoring of exposure to environmental estrogens I: Bisphenol A in pregnant
  women. Food Chem. Toxicol. 92, 129–142. doi:10.1016/j.fct.2016.03.023
- Terrien, X., Fini, J.-B., Demeneix, B.A., Schramm, K.-W., Prunet, P., 2011. Generation of
  fluorescent zebrafish to study endocrine disruption and potential crosstalk between
  thyroid hormone and corticosteroids. Aquat. Toxicol. 105, 13–20.
  doi:10.1016/j.aquatox.2011.04.007
- Tominaga, T., Negishi, T., Hirooka, H., Miyachi, A., Inoue, A., Hayasaka, I., Yoshikawa, Y.,
  2006. Toxicokinetics of bisphenol A in rats, monkeys and chimpanzees by the LC–
  MS/MS method. Toxicology 226, 208–217. doi:10.1016/j.tox.2006.07.004
- Viguié, C., Collet, S.H., Gayrard, V., Picard-Hagen, N., Puel, S., Roques, B.B., Toutain, P.-L.,
  Lacroix, M.Z., 2013. Maternal and fetal exposure to bisphenol a is associated with
  alterations of thyroid function in pregnant ewes and their newborn lambs.
  Endocrinology 154, 521–528. doi:10.1210/en.2012-1401
- vom Saal, F.S., Akingbemi, B.T., Belcher, S.M., Birnbaum, L.S., Crain, D.A., Eriksen, M.,
  Farabollini, F., Guillette, L.J., Hauser, R., Heindel, J.J., Ho, S.-M., Hunt, P.A., Iguchi,
  T., Jobling, S., Kanno, J., Keri, R.A., Knudsen, K.E., Laufer, H., LeBlanc, G.A.,
  Marcus, M., McLachlan, J.A., Myers, J.P., Nadal, A., Newbold, R.R., Olea, N., Prins,

Comment citer ce document :

Guignard, D., Gayard, V., Lacroix, M., Puel, S., Picard-Hagen, N., Viguié, C. (Auteur de correspondance) (2017). Evidence for bisphenol A-induced disruption of maternal thyroid homeostasis

in the pregnant ewe at low level representative of human exposure. Chemosphere, 182, 458-467.

<sup>10.1016/</sup>i.chemosphere.2017.05.028

G.S., Richter, C.A., Rubin, B.S., Sonnenschein, C., Soto, A.M., Talsness, C.E.,
Vandenbergh, J.G., Vandenberg, L.N., Walser-Kuntz, D.R., Watson, C.S., Welshons,
W.V., Wetherill, Y., Zoeller, R.T., 2007. Chapel Hill bisphenol A expert panel
consensus statement: integration of mechanisms, effects in animals and potential to
impact human health at current levels of exposure. Reprod. Toxicol. Elmsford N 24,
131–138. doi:10.1016/j.reprotox.2007.07.005

- Vom Saal, F.S., VandeVoort, C.A., Taylor, J.A., Welshons, W.V., Toutain, P.-L., Hunt, P.A., 797 2014. Bisphenol A (BPA) pharmacokinetics with daily oral bolus or continuous 798 exposure via silastic capsules in pregnant rhesus monkeys: Relevance for human 799 800 exposures. Reprod. Toxicol. Elmsford Ν 45, 105–116. doi:10.1016/j.reprotox.2014.01.007 801
- Wang, N., Zhou, Y., Fu, C., Wang, H., Huang, P., Wang, B., Su, M., Jiang, F., Fang, H., Zhao,
  Q., Chen, Y., Jiang, Q., 2015. Influence of Bisphenol A on Thyroid Volume and
  Structure Independent of Iodine in School Children. PLOS ONE 10, e0141248.
  doi:10.1371/journal.pone.0141248
- Wang, T., Lu, J., Xu, M., Xu, Y., Li, M., Liu, Y., Tian, X., Chen, Y., Dai, M., Wang, W., Lai,
  S., Bi, Y., Ning, G., 2013. Urinary bisphenol a concentration and thyroid function in
  Chinese adults. Epidemiol. Camb. Mass 24, 295–302.
  doi:10.1097/EDE.0b013e318280e02f
- 810 Webster, G.M., Rauch, S.A., Marie, N.S., Mattman, A., Lanphear, B.P., Venners, S.A., 2016.
- 811 Cross-Sectional Associations of Serum Perfluoroalkyl Acids and Thyroid Hormones in
- 812 U.S. Adults: Variation According to TPOAb and Iodine Status (NHANES 2007-2008).
- Environ. Health Perspect. 124, 935–942. doi:10.1289/ehp.1409589
- 814 Wu, K.-M., Farrelly, J.G., 2006. Preclinical development of new drugs that enhance thyroid
- 815 hormone metabolism and clearance: inadequacy of using rats as an animal model for

epresentative of numan exposure. Che 10.1016/i.chemosphere 2017 05 028

- predicting human risks in an IND and NDA. Am. J. Ther. 13, 141–144.
  doi:10.1097/01.mjt.0000209673.01885.b0
- Xu, X., Liu, Y., Sadamatsu, M., Tsutsumi, S., Akaike, M., Ushijima, H., Kato, N., 2007.
  Perinatal bisphenol A affects the behavior and SRC-1 expression of male pups but does
  not influence on the thyroid hormone receptors and its responsive gene. Neurosci. Res.
- 821 58, 149–155. doi:10.1016/j.neures.2007.02.011
- 822 Yang, J., Chan, K.M., 2015. Evaluation of the toxic effects of brominated compounds (BDE-
- 47, 99, 209, TBBPA) and bisphenol A (BPA) using a zebrafish liver cell line, ZFL.
- Aquat. Toxicol. 159, 138–147. doi:10.1016/j.aquatox.2014.12.011
- Zoeller, R.T., Bansal, R., Parris, C., 2005. Bisphenol-A, an environmental contaminant that acts
   as a thyroid hormone receptor antagonist in vitro, increases serum thyroxine, and alters
   RC3/neurogranin expression in the developing rat brain. Endocrinology 146, 607–612.
   doi:10.1210/en.2004-1018
- Zoeller, T.R., 2010. Environmental chemicals targeting thyroid. Horm. Athens 9, 28–40.
- Zoeller, R.T., Dowling, A.L.S., Herzig, C., Iannacone, E.A., Gauger, K.J., Bansal, R., 2002.
- 831 Thyroid Hormone, Brain Development, and the Environment. Environmental health832 perspectives 110.

833



route