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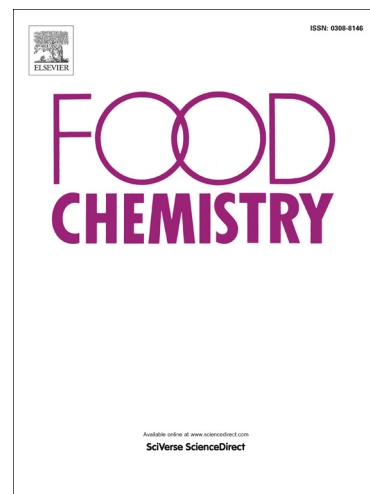
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Impact of canning and storage on apricot carotenoids and polyphenols

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Abstract

Apricot polyphenols and carotenoids were monitored after industrial and domestic cooking, and after 2 months of storage for industrial processing.

The main apricot polyphenols were flavan-3-ols, flavan-3-ol monomers and oligomers, with an average degree of polymerization between 4.7 and 10.7 and caffeoylquinic acids. Flavonols and anthocyanins were minor phenolic compounds. Upon processing procyanidins were retained in apricot tissue. Hydroxycinnamic acids, flavan-3-ol monomers, flavonols and anthocyanins leached in the syrup. Flavonol concentrations on per-can basis were significantly increased after processing. Industrial processing effects were higher than domestic cooking probably due to higher temperature and longer duration. After 2 months of storage, among polyphenols only hydroxycinnamic acids, flavan-3-ol monomers and anthocyanins were reduced.

Whichever the processing method, no significant reductions of total carotenoids were observed after processing. The *cis*- β -carotene isomer was significantly increased after processing but with a lower extent in domestic cooking. Significant decreased in total carotenoid compounds occurred during storage.

Keywords: polyphenol, carotenoid, sugar, acid, processing

Highlights

Apricot carotenoids are more stable than phenolics upon thermal treatments

Polyphenol losses are mainly due to thermal degradation and leaching

Apricot phenolic are more stable than carotenoids upon storage

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Introduction

Apricot (*Prunus armeniaca* L.) is a fruit of high economic and nutritional relevance. Apricots are a rich source of phenolics and carotenoids that may contribute to reduced risks of several degenerative diseases, such as cancer, cardiovascular diseases, caused by oxidative stress (Terry et al., 2001).

Because of its climacteric characteristic, apricots present a challenge in postharvest storage as they have a very short storage life due to high respiration rate and rapid ripening process linked to ethylene behavior after picking (Gouble et al., 2006). In order to extend their consumption and because of their short seasonal availability, apricots are commonly preserved and consumed after thermal processing methods including canning (halves in syrup for example) and pureeing.

Heat treatments are known to induce significant changes in chemical composition of foods, affecting the bioaccessibility and contents of compounds such as vitamins, carotenoids and polyphenols. Thermal processing of peaches results in a loss of 21% of total procyanidins (Hong, Barrett, & Mitchell, 2004) and some of these losses could be attributed to a leaching into the syrup of the low molecular-weight procyanidins. Cooking of pear results also in a preferential retention of the larger procyanidins in the pear sections whereas procyanidins of low degree of polymerisation and caffeoylquinic acid leach into the cooking water (Renard, 2005). The same mechanism has been observed in apple puree (Le Bourvellec, Bouzerzour, Ginies, Regis, Plé, & Renard, 2011). For cherry products, there is little loss of total phenolics with canning, but approximately 50% of total phenolics are redistributed to the syrup (Chaovanalikit & Wolstrad, 2004a). These results contrast with those of Brownmiller, Howard, & Prior (2009) who report that after canning berries 65% of the total procyanidins are retained. The thermal degradative processes depend also on phytomicronutrient structures. Hydroxycinnamic acids and (-)-epicatechin are more labile than flavonol glycosides during

processing of cherry (Chaovanalikit & Wolstrad, 2004b). Pasteurization of peach significantly reduces the carotenoid concentrations. Concentration of zeaxanthin and β -cryptoxanthin are reduced whereas levels of lutein and β -carotene are unaffected by the heat treatment (Oliveira, Pintado, & Almeida, 2012). Furthermore, thermal processing also induces a change in isomeric composition of canned tomato, due to the trans to cis isomerization (Lessin, Catigani, & Schwartz, 1997). Lessin et al. (1997) also found an increase in total β -carotene, α -carotene and β -cryptoxanthin following canning of carrots, collard greens, spinach and sweet potatoes, but losses in peaches and tomatoes. Overall, the discrepancy observed between the different food matrices might be attributable to the great variability of the shape of the food matrices (Bureau et al., 2015). Canning may improve extractability of carotenoids from cellular matrix, therefore resulting in higher levels in thermally processed products. However, excess heat may also lead to degradation.

Storage may also affect retention of phytonutrients. Oliveira et al. (2012) report a reduction of total phenolic in canned peach during the initial 18 days of storage, followed by a plateau. However, the authors did not assay the syrup in this study. Decreases of procyanidin concentration has been reported during storage of canned peaches, however these losses are due to the migration from fruit matrix to syrup (Hong et al., 2004). Anthocyanin degradation occurs in canned cherries and their syrup stored for 5 months at room temperature (Chaovanalikit & Wolstrad, 2004a, b), and cyanidin-3-O-rutinoside and pelargonidin-3-O-rutinoside undergo partial hydrolysis during storage (Chaovanalikit & Wolstrad, 2004b). Oliveira et al. (2012) show that changes in carotenoids occur during storage of peach with increased levels of zeaxanthin and reduction, by decreasing order, of β -cryptoxanthin > β -carotene > lutein. Campbell & Padilla-Zakour (2013) also reported that β -cryptoxanthin and β -carotene decrease over the storage period (in canned apricots and peaches) while lutein (in peaches) and zeaxanthin (in both fruits) are lost. Based on HPLC

data apricot phenolic and carotenoid compounds are more stable under storage than those of peaches (Campbell & Padilla-Zakour, 2013). However, tomato paste β -carotene and lycopene remained constant through 12 months of storage (Koh, Charoenprasert, Mitchell, 2011). The contents of carotenoid and phenolic compounds during food processing depend on the chemical structure of the phytonutrients, on the fruit matrix, on the intensity of the heat treatment and also on storage.

With the increased recognition of the role of plant-based phenolics and carotenoids in human health and nutrition, it is increasingly important to understand how various processing conditions and storage impact these phytonutrients. Nevertheless, information is lacking on how different thermal processing methods and storage of processed products influence the apricot qualities. This study evaluated changes in sugar, acid, polyphenol and carotenoid compositions and contents in response to thermal processing (industrial processing and domestic cooking, halves in syrup) and storage of canned apricots.

Material and methods

Chemical

Acetonitrile of HPLC grade and acetic acid were from Fischer Scientific (Pittsburgh, PA, USA). Tert-Butyl methyl ether (MTBE) of HPLC grade was from Merck (Darmstadt, Germany). Methanol of HPLC grade was from VWR international (Fontenay-sous-bois, France). 5-O-caffeoylquinic acid, (+)-catechin, (–)-epicatechin, sucrose, glucose, fructose, citric acid, β -carotene, β -apo8'-carotenal, and toluene- α -thiol were from Sigma Aldrich (Darmstadt, Germany). Quercetin, cyanidin-3-O-galactoside, and 3-O-caffeoylquinic acid were obtained from Extrasynthèse (Lyon, France). Malic acid was obtained from R-Biopharm AG (Darmstadt, Germany). Phytoene was obtained from CaroteNature (Lupsingen, Switzerland).

Plant material and sample preparation

Four apricot cultivars were collected in June 2015 in INRA experimental orchards, Gotheron at St Marcel-les-Valence (Drôme, France) and Amarine at Bellegarde (Gard, France) and in SERFEL (Station d'Expérimentation Régionale pour les Fruits et Légumes) orchard at Saint Gilles (Gard, France): 'Orangered® Bhart', 'Iranien', 'Goldrich' and 'Hargrand'. These various orchards, located in different regions, allowed to have all 4 cultivars at the same two dates (23 and 25/06), imposed by canning schedule. These cultivars presented different colors: 'Orangered® Bhart', a dark orange apricot with red blush, 'Iranien' a green-white cultivar with a little red blush on sunny fruits, 'Goldrich' a dark orange one without blush and 'Hargrand' a pale orange apricot without blush. They were chosen according to their texture evolutions after processing (Ribas-Agusti, Gouble, Bureau, Maingonnat, Audergon, & Renard, 2017). For each cultivar, about 250 fruits were harvested at a commercial maturity stage. Firmness of entire fruits was assessed by compression test

(see section physical characterization) in order to obtain samples of homogeneous representative texture hence maturity level for each cultivar and between cultivars. So, for each cultivar, 86 fruits were sorted and divided in three batches: the first batch of 24 fruits was used for analysis of fresh apricots, the second batch of 20 fruits was used for domestic cooking, and the third batch of 42 fruits was used for canning. For each process, part of the fruits was for biochemical analysis and the other part for texture analysis.

Fresh fruit

For biochemical analysis, three replicates of four fruits each were constituted randomly. In each replicate, fruits were cut into small pieces and immediately frozen using liquid nitrogen. Apricots were then divided in two bags, A and B. Bags A were freeze-dried, stored at -20 °C, and used for phenolic characterization. Bags B were stored at -80 °C and used for infrared, total soluble solids (TSS), titratable acidity (TA), carotenoids, sugars and organic acids.

Canning treatment

Canning was carried out by the Centre Technique de la Conservation des Produits Agricoles (CTCPA, Avignon, France). The fruits were cleaned, manually cut through the middle and pitted. Six apricot halves were put into each can (425 mL), to which hot syrup (70°C) was added (16° Brix, sucrose). The fruit proportion in the final product varied from 42% ('Iranien') to 67% ('Hargrand'). Cans were sealed and then heated at 95 °C. The temperature of the syrup in the can was monitored by placing thermocouples into two cans; it reached 95 °C after 14 min. and remained at this temperature for 2 min thereafter. Finally cans were sprayed with cold water to reach 30 °C.

For each cultivar, 14 cans of 6 halves each were obtained and distributed as follows:

- 3 cans for biochemical analysis one day after canning;

- 4 cans for texture analysis one day after canning
- 3 cans for biochemical analysis after 60 days storage at room temperature (22 °C);
- 4 cans for texture analysis after 60 days storage at room temperature (22 °C).

For biochemical analysis, syrup and fruits, of each can, were separated and weighted. Fruits were analyzed as described for fresh ones. Syrup was homogenized and 50 mL were frozen and stored at -20 °C until freeze-drying and polyphenol analysis.

Domestic cooking

The 20 fruits were cut through the middle and pitted. Apricot halves (40) were plunged in cane sugar syrup (16° Brix, 900 mL) at 70 °C and cooked for about 20 min, until a final temperature of 85 °C was reached inside the apricot halves as monitored with a Fluke 51 K/J thermometer (John Fluke, Everett, Washington, USA). Then apricot halves were immediately drained and cooled on ice bed. The fruit proportion in the containers varied from 22% ('Iranien') to 29% ('Hargrand').

After cooking, syrup and fruits were separated, 18 halves, distributed in three samples of six halves, were used for biochemical characterization. Syrup and fruits were weighted. Syrup was homogenized and 50 mL were frozen and stored at -20 °C until freeze-drying and polyphenol analysis.

Physical characterization

In order to compare the fruits and the cultivars on the same bases, a standardization and an harmonization of fruit batches was performed by using fruits at the same global firmness. Firmness of each fruit was measured by uniaxial compression test with a 50 mm flat probe using a texture analyzer (Pénélaup, Sérissud, Montpellier, France). The fruit compression strain was kept low (3% of apricot height) to ensure that no irreversible damage or stress to the fruit occurred. Test operating conditions were conducted with test-speed 10 cm

min⁻¹. Force corresponding to 3% strain was measured and used to compute the related compression pressure, expressed in Pascal (Grotte et al., 2001). For each cultivar, fruits were sorted by decreasing firmness and 86 fruits were selected with the more homogeneous firmness and dispatched homogeneously into the 3 conditions. The global firmness for ‘Orangered® Bhart’, ‘Goldrich’ and ‘Hargrand’ was 115 kPa (standard deviation 11) (respectively 114, 117 and 115 kPa). For ‘Iranien’, due to the imperative date for canning, fruits were less mature, with firmness of 293 kPa (standard deviation 27).

The fruit texture, before and after thermal treatments, was determined by penetrometry test. For each half (10 to 12 by condition), a slice of about 1.5 cm thick was cut longitudinally and four measurements were made on each slice (Ribas-Agusti et al., 2017). Texture measurements were performed using a multi-purpose texturometer TaPlus (Lloyd, Ametek, Elancourt, France). The system measured the maximum force (N) required to penetrate 9 mm of pulp, at constant penetration rate (100 mm min⁻¹), using a flat cylinder tip (2.0 mm diameter).

Mid-infrared spectroscopy

As described in Bureau et al. (2013), mid-infrared spectra were collected at 23 °C with a Bruker Tensor 27 FTIR spectrometer (Wissembourg, France) equipped with a horizontal attenuated total reflectance (ATR) sampling accessory and deuterated triglycine sulphate (DTGS) detector. The homogenized (only halves were ground) samples were placed at the surface of the zinc selenide crystal providing six internal reflections into the samples. The sample consistency, a thick liquid, allowed a good contact between sample and crystal and did not require pressing. The spectrum of each sample was obtained by taking the average of 32 scans at a resolution of 4 cm⁻¹, with scanner velocity of 10 KHz. The crystal was cleaned between measurements with deionized water and dried with lint-free tissue.

Analysis methods

Total Soluble Solids (TSS) were determined with a digital refractometer (PR-101 ATAGO, Norfolk, VA, USA) and expressed in °Brix at 20 °C. Titratable acidity (TA) was determined by titration up to pH 8.1 with 0.1 mol L⁻¹ NaOH and expressed in mmol H⁺ kg⁻¹ fresh weight (FW) using an autotitrator (Methrom, Herisau, Switzerland).

Sugars (glucose, fructose, and sucrose) and organic acids (malic acid and citric acid) were quantified using colorimetric-enzymatic methods (R-Biopharm AG, Darmstadt, Germany) and expressed in g kg⁻¹ FW. These measurements were performed with a SAFAS flx-Xenius XM spectrofluorimeter (SAFAS, Monaco).

TSS, TA, sugars and organic acids were quantified in halve fruit only.

Phenolics were measured by high-performance liquid chromatography (HPLC)-diode array detection (DAD) after thioacidolysis using a method adapted from Le Bourvellec et al. (2011) and Guyot, Marnet, & Drilleau, (2001) on fruit and syrup. Procyanidins were characterized by thioacidolysis to determine subunit composition, average molecular mass and the average degree of polymerization (mDP). The mDP of procyanidins was calculated as the molar ratio of all the flavan-3-ol units (thioether adducts plus terminal units) to (-)-epicatechin and (+)-catechin corresponding to terminal units. Thioacidolysis was performed on the freeze-dried samples according to the procedure established by Le Bourvellec et al. (2011) and Guyot, Marnet, & Drilleau, (2001) except that the amount of added concentrated HCl was increased to take into account the buffering capacity of the fruit. Precisely weighted powder (c.a. 100 mg for fruit and c.a 1200 mg for syrup) was suspended in 1.5 mL Eppendorf vial with 400 µL of dried methanol acidified by concentrated HCl (250 mL/L) and 800 µL of toluene α thiol solution (50 mL/L) in dried methanol. The reaction was carried out at 40°C for 30 min with agitation every 10 min. Then, the vials were cooled in an ice bath for at least 5 min. After filtration (PTFE, 0.45 µm), the reaction medium was directly injected (20 µL) into

the HPLC-DAD system. In order to distinguish between native flavan-3-ol monomers and flavan-3-ol monomers coming from the terminal units of procyanidins a 'crude' seed analysis without thiolysis was performed. Precisely weighted powders (c.a. 100 mg for fruit and c.a. 1200 mg for syrup) were suspended in 1.5 mL Eppendorf vial with 1200 μ L of methanol acidified by acetic acid (10 mL/L). The reaction was carried out in an ultrasonic bath during 15 min. After filtration (PTFE, 0.45 μ m), the reaction mixture was directly injected (20 μ L) into the HPLC-DAD system. Analyses were performed using an Ultra Fast Liquid Chromatography Shimadzu Prominence system (Kyoto, Japan) including two pumps LC-20AD Prominence liquid chromatograph UFLC, a DGU-20A5 Prominence degasser, a SIL-20AHT Prominence autosampler, a CTO-20AC Prominence column oven, a SPD-M20A Prominence diode array detector, a CBM-20A Prominence communication bus module and controlled by a LC Solution software (Shimadzu, Kyoto, Japan). Separations were achieved in Le Bourvellec et al. (2011) conditions. Individual compounds were quantified in mg kg^{-1} FW by comparisons with external standards:

-at 280 nm for (+)-catechin, (-)-epicatechin, (-)-epicatechin benzyl thioether (quantified as (-)-epicatechin);

-at 320 nm for 3-O-caffeoylquinic acid 5-O-caffeoylquinic acid, and their methylated derivatives obtained during thioacidolysis reaction quantified as their respective non-methylated equivalents;

-at 350 nm for quercetin-3-O-glycosides (quantified as quercetin);

-at 540 nm for cyanidin glycosides (quantified as cyanidin-3-O-glucoside).

Fruit carotenoid extraction was carried out using the micromethod described by Page, Van Stratum, Degrou, & Renard, (2012), including the addition of β -apo8'-carotenal used as internal standard. HPLC-DAD analyses were performed using an Ultra Fast Liquid chromatography Shimadzu Prominence system (Kyoto, Japan) as described above.

Separations were achieved on a 250 mm × 4.6 mm i.d. 3 μm Develosil C30 (Nomura Chemical Co. Ltd., Seto, Aichi, Japan) protected with a precolumn containing the same stationary phase, in an oven maintained at 30°C. Elution was performed at a solvent flow rate 1.4 mL min⁻¹. The mobile phase was methanol and the polarity gradient was achieved with MTBE, starting with 100% methanol, increasing to 40% MTBE in 10 min, then increase up to 60% MTBE in 2 min, maintaining that percentage of MTBE for 7 min, return to 100% Methanol in 2 min and hold for 10 min. Carotenoids were detected at wavelength from 280 nm to 600 nm, and quantified at 450 nm (for β-apo8'-carotenal, β-carotene), 350 nm (for phytofluene) and 290 nm (for phytoene). β-carotene and β-apo8'-carotenal were identified by co-elution of standards. The others (phytoene, and phytofluene), and especially β-carotene isomers, were tentatively identified according to their UV spectrum and quantified as equivalent of β-carotene and corrected according to their extinction coefficient. Quantification was performed relative to the peak area of the internal standard (β-apo8'-carotenal). Response factors were calculated for β-carotene from standard solutions. The results are expressed equivalent β-carotene using its response factor. Carotenoid levels were expressed in mg kg⁻¹ of fresh weight.

Statistical analysis

Results are presented as mean values. Two-way analysis of variance (ANOVA) by Fisher's test (F) was used to compare the means and performed using the XI Stat package of Microsoft Excel. Difference was considered significant at $P \leq 0.05$.

Results and discussion

Global characterization of apricot after canning and storage by mid-infrared spectroscopy (MIRS)

For each variety and each processing condition, samples (fruits without syrup) were analysed by MIRS reflecting information on changes of biochemical composition especially for major metabolites such as sugars and organic acids (Bureau et al., 2009a). ANOVA performed on spectral data allowed to classify factors, here cultivar and cooking conditions, according to their effect on apricot spectral data. The Fisher values (F) was higher for cultivar (F=180) than for cooking conditions (F=60) (results not shown). ANOVA allowed also identifying the most informative spectral regions according to the studied effects, which were in this study between 1500 and 1000 cm^{-1} for the cultivar and between 1800 and 1500 cm^{-1} for the cooking conditions. In order to highlight the effect of the cooking conditions, a Principal Component Analysis (PCA) was performed on the second region ranging from 1800 and 1500 cm^{-1} (Figure 1A). PC1 represented 84.3% of the total variability whereas PC2 only 7.1%. The main variations were then observed on the PC1. 'Goldrich' and 'Iranien' were clearly separated on the PC1. 'Hargrand' and 'Orangered® Bhart' appeared to be overlapped on PC1 (84.3% variation) but not on PC2 (7.1% variation). Inside each cultivar group, cooking conditions always respected the same trend: on the left, fresh and domestic cooking samples (F and D) and on the right, canned apricot with 1 day and 2 months of storage (C0 and C2). The first and second eigenvectors (Figure 1, B and C) showed negative peaks at 1717 and 1636 cm^{-1} for the first eigenvectors and positive peaks at 1724 and 1589 cm^{-1} for the second one. The sample discrimination seemed to be in relation with organic acid and water content. Indeed, peaks at 1717, 1724 and 1589 cm^{-1} could be attributed to the C=O fonction of carboxylic acid with absorption in the range 1740-1660 cm^{-1} and when ionisated characterized by 2 bands with one between 1630-1550 cm^{-1} (Coates, 2000; Bertrand & Dufour, 2000).

Moreover the deformation of O-H (bending scissoring) provides a band with a maximum at 1636 cm^{-1} in our conditions.

Sugar and acid

Sugar and acid determinations

Sucrose was the main sugar in fresh apricots accounting for 37 to 77% of total sugars, followed by glucose (13-33%) and fructose (9-39%) except in 'Iranien' cultivar where fructose was the second main sugar (Table 1). Sucrose concentrations were in accordance with results from other studies (Schmitzer, Slatnar, Mikulic-Petkovsek, Veberic, Krska, & Stampar, 2010; Caliskan, Bayazit, & Sumbul, 2012; Aubert & Chanforan, 2007; Bureau et al., 2009b) with reported concentration ranging from 12 to 110 g kg^{-1} FW. Glucose and fructose concentrations ranges measured by Aubert & Chanforan (2007), Bureau et al. (2009b), Schmitzer et al. (2010), and Caliskan et al. (2012), were from 6.0 to 54.6 g kg^{-1} FW and from 1.0 to 44.2 g kg^{-1} FW, respectively in agreement with the present data.

The mean levels of malic and citric acid concentrations in fresh apricots were quite similar, ranging respectively from 3.8 to 10.7 g kg^{-1} FW and to 5.5 to 13 g kg^{-1} FW, in agreement with those measured by Aubert & Chanforan (2007), Schmitzer et al. (2010) and Bureau et al. (2009b) with 'Orangered® Bhart' and 'Iranien' the less acid and 'Hargrand' and 'Goldrich' the more acidic cultivars. It was to note that usually the 'Iranien' acid contents are lower than those of 'Orangered® Bhart' (Ribas-Agusti et al., 2017). This difference was due to the fact that 'Iranian' was less mature.

Canning effect

Except for 'Hargrand', a significant increased of sucrose concentration was observed for industrial canning. In contrast, for domestic cooking, except for 'Goldrich', sucrose

concentration decreased. The increase observed could be due to an equilibration of the fruit and syrup sucrose concentration consecutive to a softening of the fruit tissue. For glucose and fructose all cases were observed, their concentrations in the processed fruits may be close to, lower, or higher than in the fresh fruit, again probably due to an equilibration of the concentrations between the fruit and the syrup.

Pasteurization of fresh apricots significantly affected the organic acid concentrations (Table 1), but with different trends. Industrial processing caused a significantly higher loss of organic acids when compared to domestic cooking, for which organic acid concentrations remained close to those observed in fresh fruit, in accordance with MIRS. Loss by leaching from fruit to syrup was more favored by the industrial treatment that induced a stronger softening of the flesh (Supplementary data S1). Moreover, the observed trends may also result from a kinetic effect. After domestic cooking the separation between the fruit halves and the liquid took place very quickly after the cooking, whereas in the case of industrial treatment the separation between the solid and the liquid was done one day after the cooking of the cans.

Storage effect

A significant increase of glucose and fructose concentrations was observed with storage except for 'Iranien'. This could be due to a syrup sucrose hydrolysis during storage due to low syrup pH (below 4.0, data not shown). In fact samples showed a slight decrease in sucrose with storage but it was not significant.

The concentration of malic acid, but not citric acid, decreased significantly after two months of storage. The fact that malic acid decreased during storage while citric acid was stable could be due to their tissue distribution. While malic acid is located preferentially in the pulp, citric acid has a subepidermal distribution (Lichou & Jay, 2012). The localization of the

malic acid would thus promote its leaching in the syrup as opposed to the citric hence the differences observed during storage for both acids.

Phenolic compounds

Phenolic profile

Four major phenolic groups with a total of 9 identified individual compounds were quantified (Table 2). Phenolic compounds included flavan-3-ols ((+)-catechin and (-)-epicatechin (monomer) and procyanidins (polymers)), two hydroxycinnamic acids (3-O-caffeoylquinic and 5-O-caffeoylquinic acids), two flavonols (quercetin-3-O-rutinoside, and -3-O-glucoside) and two anthocyanins (cyanidin-3-O-rutinoside and cyanidin-3-O-glucoside). These groups coincide with those found previously in other stone fruits (peach and nectarine) (Tomas-Barberan, Gil, Cremin, Waterhouse, Hess-Pierce, & Kader, 2001; Tavarini et al., 2011) and apricots (Ruiz, Egea, Gil, & Tomas-Barberan, F.A. 2005a).

For fresh fruits, phenolic levels ranged between 406 and 2013 mg kg⁻¹ FW. Overall, these quantities are noticeably higher than previously published data (Ruiz et al., 2005a). Ruiz et al., (2005a) found that total phenolic content, quantified as the sum of the individual compounds identified by HPLC, between 326 and 1600 mg kg⁻¹ FW of edible tissue. Differences observed may be due to the cultivar analyzed and to the thiolysis-HPLC method that allows a better estimation of polymerized proanthocyanidins. These compounds are usually underestimated when alcoholic or hydro-alcoholic extractions are used because most of them are not extracted (Ruiz et al., 2005a). Moreover, after extraction, their estimation by direct HPLC remains incomplete because polymeric forms do not give well-resolved peaks on chromatograms. Therefore, at best, only oligomers have been quantified by direct HPLC analyses (Ruiz et al., 2005a).

Among the four major groups, procyanidins were the predominant class. The structure of apricot procyanidin showed a great homogeneity for all cultivar on the basis of the nature of the constitutive flavan-3-ol units. (-)-Epicatechin was always the predominant constitutive unit, accounting for more than 85% of total units for all cultivars with a small proportion of (+)-catechin detected only as terminal units. The proportions of (-)-epicatechin and (+)-catechin as terminal units were noticeably variables from one cultivar to another. Except 'Goldrich' cultivar (8% terminal (+)-catechin and 14% terminal (-)-epicatechin), terminal (+)-catechin was always predominant i.e. 9% terminal (+)-catechin and 1% terminal (-)-epicatechin for 'Hargrand', 11% terminal (+)-catechin and 8% terminal (-)-epicatechin for 'Orangered® Bhart' and 13% terminal (+)-catechin and 3% terminal (-)-epicatechin for 'Iranien'. These percentages may be slightly overestimated because of the epimerization reaction occurring under the conditions of thiolysis. Procyanidins' average degrees of polymerisation (DP_n) ranged between 4.7 and 10.7 and were close to those of apple procyanidins (Guyot et al., 2001; Le Bourvellec et al., 2011). The procyanidins' DP_n determined in this study were higher than those calculated from normal-phase HPLC-DAD by Ruiz et al. (2005a). The only flavan-3-ol monomers detected were (+)-catechin and (-)-epicatechin. Hydroxycinnamic acids represented the second phenolic class. Except in 'Goldrich' cultivar, the amount of 5-O-caffeoylquinic acid was higher than that of 3-O-caffeoylquinic acid. The occurrence of quercetin-3-O-rutinoside and quercetin-3-O-glucoside in apricots was reported in (Ruiz et al., 2005a ; Campbell, Merwin, & Padilla-Zakour, 2013) as well as other flavonols derivatives. Cyanidin-3-O-glucoside and cyanidin-3-O-rutinoside were the only two anthocyanin detected in 'Orangered® Bhart' cultivar (Ruiz et al., 2005a). However, Bureau, Renard, Reich, Ginies, & Audergon, (2009c) detected three anthocyanins in red apricot fruits, the major compound being cyanidin-3-O-rutinoside, and two minor compounds cyanidin-3-O-glucoside and peonidin-3-O-rutinoside. The anthocyanins, despite

being in low concentrations (32 mg kg^{-1} FW) had a clear impact in apricot skin color, providing a red blush. Among the analyzed cultivars, 3 have no supracolor while only 'Orangered® Bhart' has one, so the results are consistent with the visual characterization.

All the concentrations of monomeric phenolic compound (flavan-3-ol monomers, hydroxycinnamic acids, flavonols and anthocyanins) are consistent with previous works (Campbell et al., 2013) which further showed a wide variation in the phenolic content of different cultivars (Campbell et al., 2013; Ruiz et al., 2005a).

Canning effect

Pasteurization of fresh apricots significantly affected the phenolic compound concentrations (Table 2), as demonstrated on various fruits (Asami, Hong, Barrett, & Mitchell A.E. 2003; Chaovanalikit & Wrolstad, 2004a; Brownmiller et al., 2009). Industrial thermal processing caused a significant higher loss of total phenolic compounds from 13% (in 'Hargrand') to 47% (in 'Iranien') when compared to domestic cooking from 2% (in 'Hargrand') to 33% (in 'Orangered® Bhart'), which may be related to the different temperature levels. Moreover, higher temperatures may increase membranes permeability facilitating leaching (Jimenez, Martinez-Tomé, Romojaro, Murcia, 2008; Chaovanalikit & Wrolstad, 2004a). Asami et al. (2003) report a 20% reduction in total phenolics after pasteurization of peach samples at 110°C . Decreases in total phenolic content after canning of blueberry or cherry products have also been reported (Chaovanalikit & Wrolstad, 2004a; Brownmiller et al., 2009).

Retention of polyphenol in canned apricots was higher for flavan-3-ols. Processing, industrial and domestic, resulted in 2.4% (in 'Hargrand') to 44% (in 'Iranien') loss in procyanidins, but the type of processing had no significant effect on procyanidin losses. Procyanidin's DPn significantly increased from fresh fruit to canned products. Procyanidins are known to form complexes with food macromolecules (proteins and carbohydrates) (Le

Bourvellec & Renard, 2012). Especially, procyanidins with high degree of polymerization are adsorbed onto cell walls by physico-chemical interactions. So the increase observed could be due to leaching of procyanidins of low molecular weight into the syrup during cooking and retention of procyanidins of high molecular weight within fruit tissues. Renard (2005) also observed this phenomenon during pear cooking. Flavan-3-ol monomers were significantly decreased after processing, and notably after industrial cooking.

Hydroxycinnamic acids, 3-O-caffeoylquinic acid and 5-O-caffeoylquinic acid, were significantly lost from apricot halves after cooking (Table 2) and this loss significantly differed between the two processes. Lower losses were observed after domestic cooking. Except in 'Iranien' cultivar, an increased was observed for flavonols after domestic cooking. This might be due to a matrix softening effect, which increased the extraction efficiency of flavonols in the softened peel, as in most fruit flavonols are highly concentrated in the peel, resulting in a higher concentration with respect to the raw material. However, after industrial cooking a decrease in flavonol concentrations was observed, except for 'Hargrand'. The flavonol effects observed with industrial cooking may be due to a negative balance between extractability in the softened fruits (Supplementary material S1) and a loss by degradation and/or leaching.

'Orangered® Bhart' anthocyanins were significantly altered by the heat treatment (Table 2), but there was no difference between processes. Heating also decreases anthocyanin concentrations in nectarines but not in cherries, peaches and plums (Chaovanalikit & Wolstrad, 2004b; Leong & Oey, 2012).

Storage effect

The 2 months storage did not provide a significant loss of total phenolic compounds; this was mainly due to the fact that procyanidins were stable during storage (Table 2). This agree with the study of Jimenez et al., (2008) who found no changes in the different

antioxidant activities assays from 1 to 150 day of storage of canned apricots. It should be pointed out, however, that 'Iranien' and 'Orangered® Bhart', the less concentrated cultivars, were those which had the most losses, even if it was not significant.

While flavan-3-ol monomer and hydroxycinnamic acid concentrations decreased significantly after storage, flavonol glycosides differed in their responses: quercetin-3-O-glucoside concentrations being lower, close to, or higher than in the fresh fruit, in contrast to quercetin-3-O-rutinoside. Moreover, the flavonol glycosides response to storage was cultivar dependent. For anthocyanins, significant losses after two months of storage were observed as in the case of canned cherries (Chaovanalikit & Wolstrad, 2004b).

Balance between degradation and leaching

In order to estimate the proportion of losses due to thermal degradation or migration from the food matrix to the syrup, the levels of phenolic compounds were calculated on a per-can basis (weights of fruit and syrup) and compared to the content calculated for fresh apricots (fruit weight, 100%) (Figure 2). (+)-Catechin losses observed were mainly due to degradation induced by industrial processing and storage in contrast to (-)-epicatechin which was both degraded and leached into the syrup (Figure 2A and B). Industrial processing which increased the matrix softening effect compared to domestic cooking (Supplementary material S1) favored the leaching into the fruit syrup. The apparent susceptibility of (-)-epicatechin to leaching into the syrup compared to (+)-catechin could be attributed to a mechanism of epimerization of (-) epicatechin to (+)-catechin which might occurred in these heated conditions (Vidal, Cartalade, Souquet, Fulcrand & Cheynier, 2002 ; Guyot, Doco, Souquet, Moutounet & Drilleau, 1997). Procyanidins were mainly degraded by processing (Figure 2C), except in the case of 'Hargrand' and 'Orangered® Bhart' cultivars for which a leaching into the syrup was also observed. Their susceptibility to degradation was also cultivar dependent and followed the same trend as the monomeric flavan-3-ols. Changes in 3- and 5-

caffeoylquinic acids (Figure 2D and E) contents resulted from thermal degradation and leaching favored by industrial processing except in the case of 'Iranien' cultivar. Anthocyanins were also prone to thermal degradation and leaching (Figure 2F). Changes in flavonols upon cooking resulted in higher content with respect with to the raw material due to a food matrix destructuring effect. However, the softening of the food matrix structure was not correlated to the extractability increase. For example 'Goldrich' was the cultivar for which the softening was the strongest (Supplementary material S1) but it was not the one for which there was the highest increase of the extractability.

In view of these results, for total polyphenol contents, the cultivar susceptibility to leaching and degradation was in the following order: 'Iranien' = 'Orangered® Bhart' > 'Goldrich' > 'Hargrand'. It appeared that this result was associated with levels of TPP that were Orangered® Bhart' < 'Iranien' < 'Goldrich' < 'Hargrand'. The less the cultivars were concentrated in polyphenols the more susceptible they were to degradation and leaching.

Carotenoids

Four different carotenoids were identified in fresh and cooked apricots (Table 4). Carotenoid analysis was not performed on the syrups because Campbell & Padilla-Zakour (2013) have shown that leaching of apricot carotenoid after canning was minimal (<1%). Because of the general insolubility of carotenoids in water, the presence of this compound in syrup was thought to be largely due to dispersed fruit fragment.

The total carotenoid contents varied from 7.8 mg kg⁻¹ FW in 'Iranien' to 108.9 mg kg⁻¹ FW in 'Hargrand'. On the whole, mean values of total carotenoids were similar to those reported by Ruiz, Egea, Tomas-Barberan, & Gil, (2005b) (15-165 mg kg⁻¹ FW) for 37 apricot cultivars and hybrids, except for 'Iranien' which showed very low carotenoid contents (in accordance with low color values, data not shown).

In agreement with other published results (Leccese et al., 2010; Ruiz et al., 2005b; Kurz, Carle, & Schieber, 2008; Campbell et al., 2013), the most abundant carotenoid in apricot fruit depends on the cultivar, trans- β -carotene in Goldrich and Orangered[®] Bhart, phytoene on 'Hargrand', and phytofluene on 'Iranien'.

Microscopic investigations revealed that within the chromoplast, β -carotene can be either in a crystalline form or partially solubilized in lipid droplets as a function of fruits and vegetables (Pénicaud, Achir, Dhuique-Mayer, Dornier & Bohuon, 2011). It seemed that, in apricot, β -carotene was partially solubilized in lipid droplets, and a portion of cis- β -carotene naturally occurs (Kurz et al., 2008). A cis- β -carotene isomer was detected in fresh apricots (Table 4). According to our chromatographic conditions, UV/vis spectral data and literature data (Carle, & Schieber, 2005 ; Kurz et al. 2008), therefore, we assumed that the cis isomer of β -carotene detected in apricots was the 13-cis-isomer. The physical state of the carotenoids present in apricots could be an important factor for the stability of β -carotene both during the process and during the storage.

Canning effect

Concentrations of total carotenoids exhibited no significant change after heat processing (Table 3). In fact, samples showed a slight increase in total carotenoids with canning. This might be due to an increased of extraction efficiency in the softened fruits, disruption of caroteno-protein complexes, and/or inactivation of oxidizing enzymes (Lessin et al., 1997). Among the four carotenoids analyzed, only cis- β -carotene was significantly increased after heat processing. Several studies have reported an increase in cis-isomers in vegetables subjected to thermal processes (Schieber & Carle, 2005; Lessin et al., 1997). In particular, thermal treatment is known to cause the formation of the 13-cis-isomer (Schieber & Carle, 2005). The assumption that the cis isomer of β -carotene detected in our samples was the 13-cis-isomer was corroborated by the fact that the cis- β -carotene detected in fresh apricot

was significantly increased after heat processing and that no new peaks were visible on the HPLC chromatograms. The physical state of carotenes can be considered as the most critical factor leading to isomerization (Marx, Stuparic, Schieber & Carle, 2003). Hence, The increased of *cis*- β -carotene could also be attributed to the partial dissolution of crystalline carotene present in the intact apricot in lipid droplets (Kurz et al., 2008) as *trans-cis*-isomerization is facilitated when carotenoids are dissolved, whereas crystalline carotenoids tend to be more stable (Schieber & Carle, 2005 ; Pénicaud et al., 2011).

Significant lower contents of *cis*- β -carotene were obtained after domestic cooking than canning, probably due to lower temperature of processing.

Storage effect

Total carotenoid compounds significantly decrease over the storage period. Campbell & Padilla-Zakour (2013) have also shown a decrease of carotenoids after 6 months of storage. Carotenoids from kiwifruit puree tend also to be significantly reduced over time (Benlloch-Tinoco, Kaulmann, Corte-Real, Rodrigo, Martínez-Navarrete, & Bohn, 2015). Carotenoid degradation during storage might be due to oxidation reactions leading to the formation of low molecular weight epoxycarotenoids and apocarotenoids (Oliveira et al., 2012), however no new peaks were visible on the HPLC chromatograms after storage. Among carotenoids, *trans*- β -carotene concentrations were not modified by storage. Tomato paste β -carotene and lycopene remained also constant through 12 months of storage (Koh et al., 2011). Overall, the discrepancy with data in the literature might be attributable to the great variability of carotenoid stability in different food matrices (Lee & Coates, 1999). Moreover, the fact that *trans*- β -carotene concentrations were not modified by storage could also be attributed to their physical state as crystalline carotenoids tend to be more stable than dissolved carotenoids (Schieber & Carle, 2005). Moreover, it seemed also that isomerization of β -carotene was a prerequisite for oxidation degradation (Pénicaud et al., 2011).

Conclusions

The effect of cooking on polyphenol and carotenoid concentrations depends on the processing parameters, the structure of the food matrix, the chemical nature and the physical state of the phytonutrients. Carotenoids were more stable than phenolic compounds upon thermal treatments. Thus, domestic cooking ensured better preservation of polyphenols than industrial processing: due to lower temperature used, thermal degradation was limited. Finally, it appeared that the polyphenol losses observed were largely due to initial thermal degradation and diffusion in the syrup, but they were relatively stable during storage. However, carotenoids showed a different behavior, thermal treatment induced increased extraction efficiency leading to an apparent increase upon processing but they were degraded during storage.

Moreover, it seemed that the stability of carotenoids could mainly be attributed to their physical state, i.e. the crystalline form. The physical appearance of carotenoids appears to be an important factor in their stability during the processes. Thus, it appeared that in apricot, within the chromoplast, β -carotene could be in a crystalline form, partially solubilized in lipid droplets and a portion of cis- β -carotene naturally occurred. During processing, the crystalline form remained stable and the form partially solubilized in lipid droplets isomerized resulting in a stabilization of trans- β -carotene and an increased in cis- β -carotene. During storage, it appeared that the crystalline form remained stable as evidenced by trans- β -carotene contents after 2 months of storage whereas the cis-isomer forms were reduced probably due to degradation by oxidation.

The content of polyphenols and carotenoids after processing and storage is a net result of a combined increase in phytonutrient extractability, and a loss by degradation and leaching. It should also be noted that processing preserved a good part of the potential of the fruit, which is of interest to the consumer.

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Figure captions

Figure 1: A. Principal Component Analysis (PCA) on MIR spectral data acquired on apricot purees. Fig. A. Plot of samples distribution according to PC1 and PC2 with the distinction of cultivars ('Hargrand', 'Iranien', 'Goldrich' and 'Orangered® Bhart') and cooking conditions: F for fresh before processing, D: domestic cooking, C0: canning after 1 day of storage and C2: canning after 2 months. Fig. B and C eigenvectors 1 and 2 respectively.

Figure 2: Levels of phenolic content in a per-can basis compared to the initial entire fruit content expressed in percentage.

GD: 'Goldrich', HG: 'Hargrand', IR: 'Iranien', OR: 'Orangered® Bhart', D: domestic cooking, C0: canning after 1 day of storage and C2: canning after 2 months of storage.

Black line: initial entire fruit content in percentage. ■: fruit content in percentage, ■: syrup content in percentage.

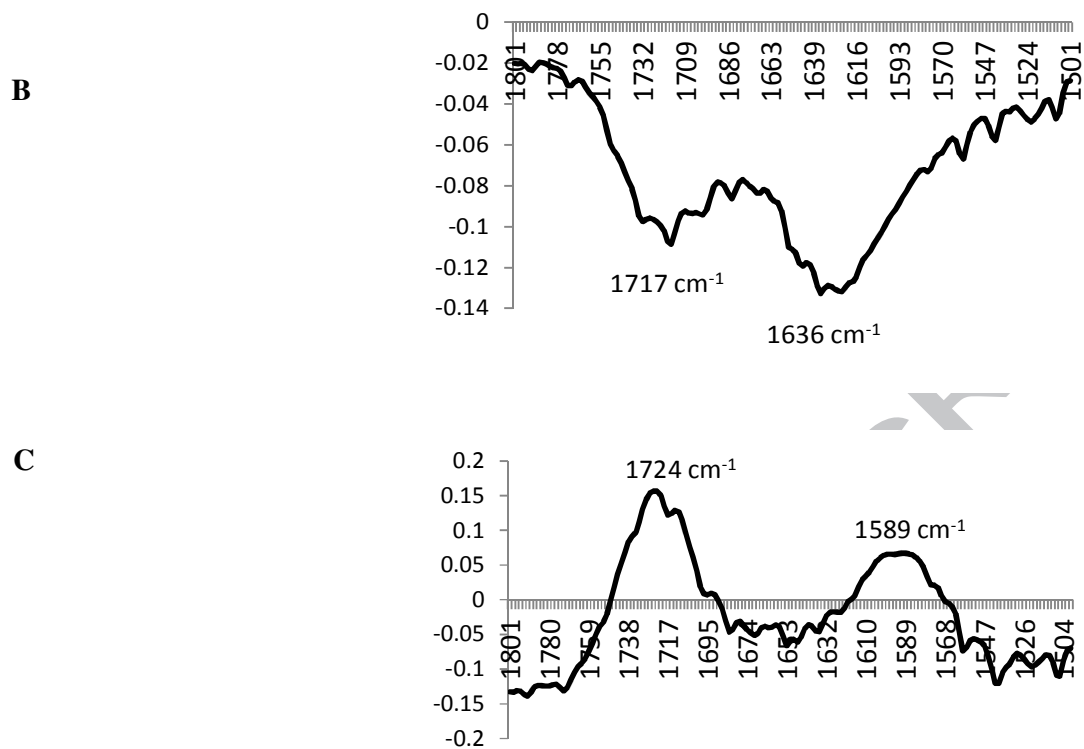
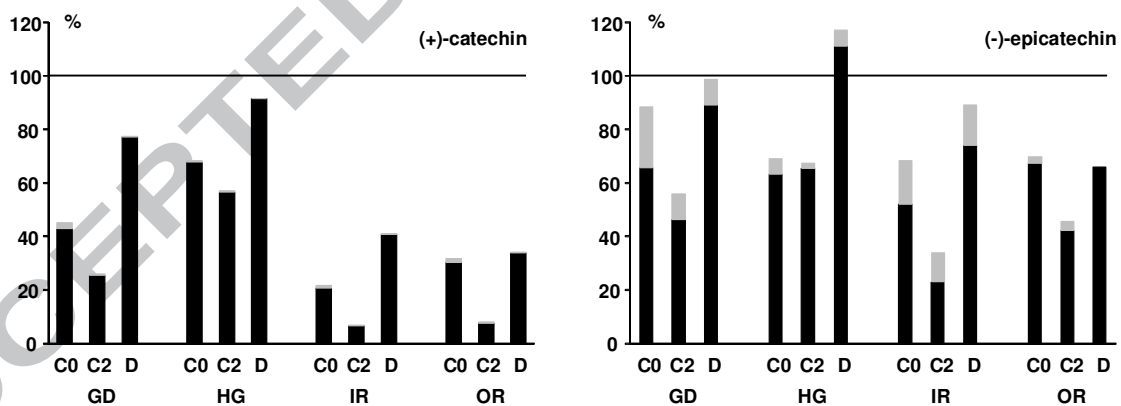


Fig 2 :



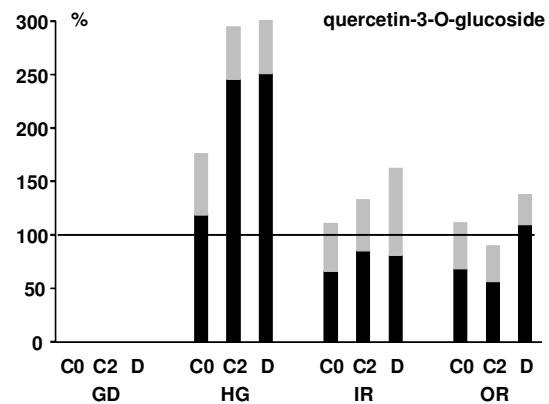
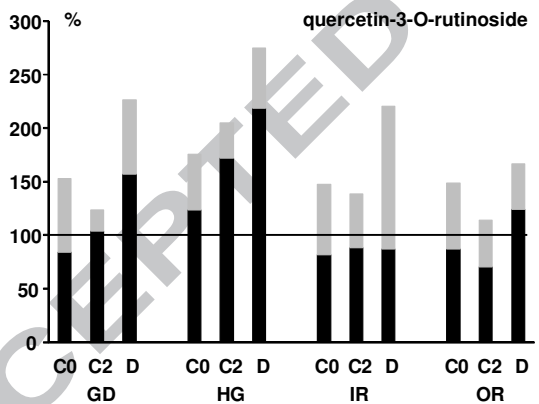
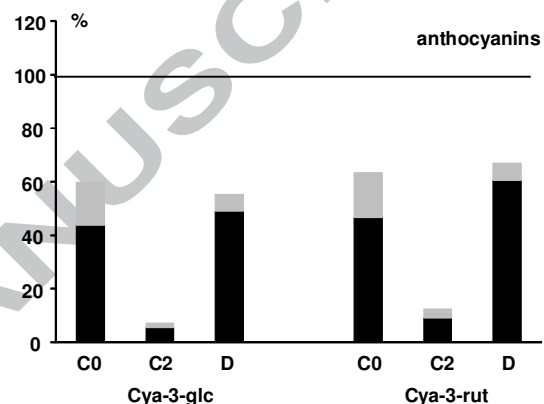
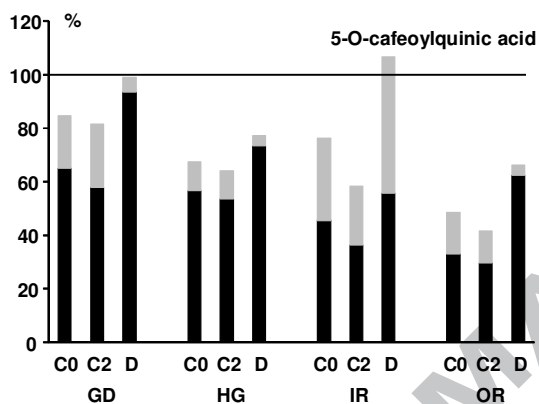
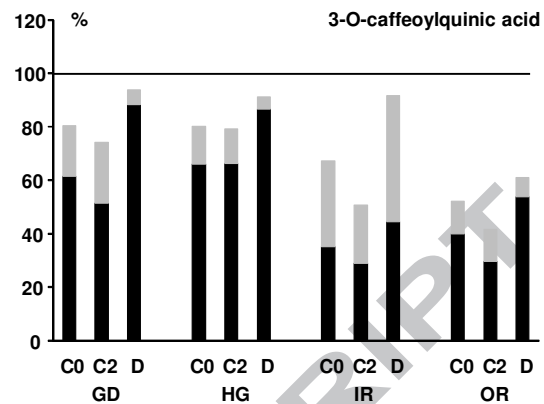
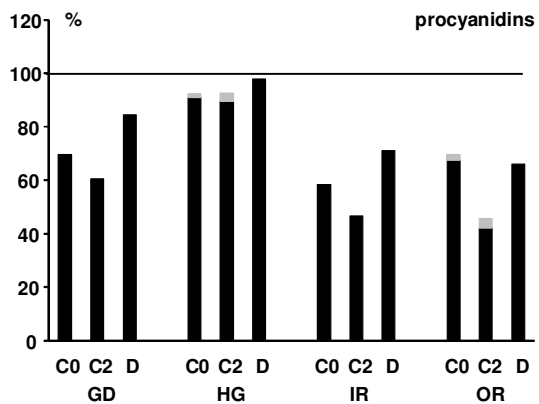


Table 1: Sugar and acid compositions (g kg^{-1} FW) in apricots as function of cultivars, processing and storage.

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Cultivars	Process ing	St orage	Gl ucose	Fr uctose	S ucose	Ma lic acid	Citr ic acid
Iranien	fresh		1 9.8	5 8.2	7 1.1	3.8	6.2
Iranien	canned	1 day	1 8.9	1 5.3	8 0.7	1.5	3.0
Iranien	canned	6 0 days	1 8.5	1 3.0	8 3.9	1.5	3.4
Iranien	domest ic cooking	0 day	2 0.4	5 3.3	6 6.0	3.3	5.6
Goldrich	fresh		1 4.8	8. 5	4 5.1	10. 7	13. 0
Goldrich	canned	1 day	1 5.5	1 1.4	7 0.5	6.8	6.7
Goldrich	canned	6 0 days	2 1.2	3 9.2	6 6.0	5.7	7.4
Goldrich	domest ic cooking	0 day	1 2.7	8. 0	4 9.9	10. 3	10. 0
Orangered® Bhart	fresh		2 0.5	9. 0	4 5.5	4.0	5.5
Orangered®	canned	1	1	7.	6	2.2	2.2

Bhart		day	1.3	2	0.6		
Orangered®	canned	6	1	1	5	1.9	2.8
Bhart		0 days	6.4	3.0	2.5		
Orangered®	domest	0	1	9.	3	4.2	4.6
Bhart	ic cooking	day	7.3	2	1.6		
Hargrand	fresh		2	1	7	9.0	9.7
			0.2	1.9	2.0		
Hargrand	canned	1	1	1	4	6.8	6.6
		day	7.6	1.2	6.4		
Hargrand	canned	6	3	2	3	6.5	6.3
		0 days	2.4	9.8	7.0		
Hargrand	domest	0	1	1	4	9.0	10.
	ic cooking	day	8.1	1.8	2.1		6
Statistics		P	1.	1.	5.	0.3	0.4
		ooled SD	0	9	0		
F value and significance							
Cultivar			5.	1	1	74.	40.
			84	85.65	1.20	55	12
			*	*	*	**	**
				**	*	*	*
Process			7.	5	2	10	79.

			96	5.17	1.44	0.87	82
			*	*	*	**	**
			*	**	**	*	*
Type of process			3. 28	8 4.56	5 4.01	25 8.00	10 7.89
			n s	* **	* **	** *	** *
Storage			7 8.30	6 0.90	1. 29	7.2 6	1.5 8
			* **	* **	n s	* *	ns

Pooled SD : pool standard deviation ddl = 32, ns (non significant), * ($P \leq 0.05$), ** ($P \leq 0.01$), *** ($P \leq 0.001$).

Values are means of 3 replicates.

Table 2: Phenolic composition (mg kg⁻¹ FW) in apricots as function of cultivars, processing and storage.

Cultivars	Processing	Storage	Form fruit/syrup	Flavan-3-ols				Phenolic acids		Flavonols		Anthocyanins		Total PP
				C	AT	CA	DP	-CQA	-CQA	Quercetin-3-rut	Quercetin-3-glc	Yanidin-3-glc	Yanidin-3-rut	
Iranien	fresh			7	8	88	1.2	7	3	2			77	
Iranien	canned	1 day	fruit	4	4	66	1.9	0	9	0			55	
Iranien	canned	1 day	syrup			d	d		0				7	
Iranien	canned	60 days	fruit	1	1	32	1.2		5	0			88	

Iranien	canned	60	sy									n		2
		days	rup			d	d					d	d	3
Iranien	domestic	0	fr									n		3
	cooking	day	uit	7	4	03	.9	2	4	1		d	d	22
Iranien	domestic	0	sy									n		2
	cooking	day	rup			d	d					d	d	1
Goldrich	fresh											n		1
				94	4	82	.7	8	8	4		d	d	249
Goldrich	canned	1	fr									n		8
		day	uit	27	3	43	.4	0	1	2		d	d	45
Goldrich	canned	1	sy									n		1
		day	rup	2	8	d	d	2	8	1		d	d	21
Goldrich	canned	60	fr									n		7
		days	uit	0	7	74	.9	0	5	4		d	d	12
Goldrich	canned	60	sy									n		1

		days	rup	0		3	.5	4	0			d	d	17
Goldrich	domestic	0	fr									n		1
	cooking	day	uit	73	3	58	.9	6	2	2		d	d	086
Goldrich	domestic	0	sy									n		1
	cooking	day	rup			d	d				d	d	d	8
Orangered®	fresh											1		4
Bhart				7	4	33	.7	6	14	9		0.0	9.8	06
Orangered®	canned	1	fr									4		2
Bhart		day	uit	4	3	0	.7	8	1	6		.4	.2	29
Orangered®	canned	1	sy									1		4
Bhart		day	rup				.3		3	6		.4	.0	9
Orangered®	canned	60	fr									0		1
Bhart		days	uit			6	.0	1	8	1		.6	.8	45
Orangered®	canned	60	sy									0		4
Bhart		days	rup				.3		6	1		.1	.6	3

Orangered®	domestic	0	fr									4		2
Bhart	cooking	day	uit	3	2	8	.2	5	1	7		.9	1.9	72
Orangered®	domestic	0	sy									0		1
Bhart	cooking	day	rup			d					d	.2	.4	0
Hargrand	fresh											n		2
				3	20	681	0.7	1	4	3		d	d	013
Hargrand	canned	1	fr									n		1
		day	uit	3	4	517	1.0	0	8	6		d	d	749
Hargrand	canned	1	sy									n		1
		day	rup	0	5	7	.2		8	4		d	d	14
Hargrand	canned	60	fr									n		1
		days	uit	5	9	516	1.6	1	5	2		d	d	740
Hargrand	canned	60	sy									n		1
		days	rup			21	.4		1	0		d	d	73
Hargrand	domestic	0	fr									n		1

	cooking	day	uit	2	13	641	.3	7	2	8		d	d	966
Hargrand	domestic	0	sy									n		1
	cooking	day	rup			d	d				d	d	d	0
Statistics		Pool										0		5
		ed SD		.2	.2	6.2	.4	.2	.4	.8	.4	.5	.0	6.1
F value and significance														
Cultivar				07.32	6.61	6.21	1.99	6.6	6.81	5.43	7.31	2.35	1.91	6.83
												*		*
				**	*	**	*	**	**	*	**	*	**	**
Process				3.28	6.92	.87	.21	3.48	8.20	6.10	0.57	6.87	4.11	0.81
												*		*

				**	*			**	**	*	*	*	*	*
Type of process				1.14	2.38	.59	.89	0.82	0.80	5.82	9.32	.37	.58	0.51
				**	*	s	s	**	*	*	*	s	s	*
Storage				4.40	9.50	.79	.90	3.57	0.3	.64	8.88	17.19	82.01	.45
				**	**	s		*	*	s	*	**	**	s

EC: (-)-epicatechin, CAT: (+)-catechin, PCA: procyanidins, mDP: average degree of polymerization of procyanidins, 3-CQA: 3-O-caffeoylquinic acid, 5-CQA : 5-O-caffeoylquinic acid, Quer-3-rut: quercetin-3-O-rutinoside, Quer-3-glc: quercetin-3-O-glucoside, Cya-3-glc: cyanidine-3-O-glucoside, Cya-3-rut: cyanidine-3-O-rutinoside, TPP: total polyphenols, Nd : not detected, pooled SD : pooled standard deviation ddl = 56, ns : non significant, * (P≤0.05), ** (P≤0.01), *** (P≤0.001). Values are means of 3 replicates.

Table 3: Carotenoids concentrations (mg kg⁻¹ FW) in apricots as function of cultivars, processing and storage.

Cultivars	Pr ocessing	Stor age	P hytoene*	Ph ytofluene*	Trans-β- carotene	C is-β- caroten e	T C
Iranien	fre sh		2. 4	3.0	1 .4	1 .0	7 .8
Iranien	ca nned	1 day	3. 2	4.4	1 .9	1 .1	1 0.5
Iranien	ca nned	60 days	2. 2	3.3	1 .6	1 .0	8 .2
Iranien	DC	0 day	2. 3	3.4	1 .4	0 .9	8 .0
		Poo led SD ^a	0. 13	0.2 0	0 .10	0 .07	0 .4
Goldrich	fre sh		1 7.3	12. 7	2 1.2	3 .7	5 4.9
Goldrich	ca nned	1 day	1 5.5	11. 9	2 2.9	6 .0	5 6.4
Goldrich	ca nned	60 days	1 3.9	10. 8	1 9.8	4 .2	4 46.6

Goldrich	DC	0	1	12.	2	3	5
		day	6.2	2	2.9	.6	5.0
Orangered®	fre		7.	6.0	2	4	4
Bhart	sh		0		7.2	.5	4.7
Orangered®	ca	1	6.	6.0	2	7	4
Bhart	nned	day	6		5.0	.5	5.0
Orangered®	ca	60	6.	5.0	2	3	3
Bhart	nned	days	2		3.6	.8	8.6
Orangered®	DC	0	7.	6.3	2	4	4
Bhart		day	3		6.8	.5	4.9
Hargrand	fre		5	37.	1	3	1
	sh		2.0	6	5.9	.4	08.9
Hargrand	ca	1	5	39.	1	4	1
	nned	day	3.0	5	8.0	.2	14.6
Hargrand	ca	60	4	33.	1	4	9
	nned	days	6.4	3	5.3	.0	9.1
Hargrand	DC	0	4	34.	1	3	1
		day	8.6	8	5.3	.8	02.5
Statistics		Poo	1.	1.1	0	0	2
		led SD ^b	61	2	.97	.28	.73

F value and significance							
Cultivar			5	48	2	9	5
			66.23	1.30	93.54	7.39	45.81
			*	**	*	*	*
			**	*	**	**	**
Process			0.	1.5	0	8	3
			67	7	.97	3.46	.20
			n	ns	n	*	n
			s		s	**	s
Type of process			0.	2.5	0	1	4
			71	3	.85	02.05	.94
			n	ns	n	*	*
			s		s	**	
Storage			6.	9.8	4	4	2
			71	1	.74	8.29	0.32
			*	*	n	*	*
					s	**	**

DC: domestic cooking, TC: total carotenoids. *quantified as equivalent of β -Carotene, pooled SD: pooled standard deviation, a: ddl =8, b: ddl = 24, ns (non significant), * ($P \leq 0.05$), ** ($P \leq 0.01$), *** ($P \leq 0.001$). Values are means of 3 replicates.

Highlights

Apricot carotenoids are more stable than phenolics upon thermal treatments

Polyphenol losses are mainly due to thermal degradation and leaching

Apricot phenolic are more stable than carotenoids upon storage

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