



HAL
open science

A User's View of the Parameters Derived from the Induction Curves of Maximal Chlorophyll a Fluorescence: Perspectives for Analyzing Stress

Julie Ripoll, Nadia Bertin, Luc Bidel, Laurent Urban

► **To cite this version:**

Julie Ripoll, Nadia Bertin, Luc Bidel, Laurent Urban. A User's View of the Parameters Derived from the Induction Curves of Maximal Chlorophyll a Fluorescence: Perspectives for Analyzing Stress. *Frontiers in Plant Science*, 2016, 7, 10.3389/fpls.2016.01679 . hal-01605733

HAL Id: hal-01605733

<https://hal.science/hal-01605733>

Submitted on 28 Jun 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



A User's View of the Parameters Derived from the Induction Curves of Maximal Chlorophyll *a* Fluorescence: Perspectives for Analyzing Stress

Julie Ripoll^{1,2}, Nadia Bertin¹, Luc P. R. Bidel³ and Laurent Urban^{2*}

¹ INRA – Centre d'Avignon, UR 1115 Plantes et Systèmes de Culture Horticoles, Avignon, France, ² UMR QualiSud, Université d'Avignon et des Pays du Vaucluse, Avignon, France, ³ INRA, UMR AGAP, Montpellier, France

OPEN ACCESS

Edited by:

Vasileios Fotopoulos,
Cyprus University of Technology,
Cyprus

Reviewed by:

Amarendra Narayan Misra,
Central University of Jharkhand, India
Adeyemi Oladapo Aremu,
University of KwaZulu-Natal,
South Africa

*Correspondence:

Laurent Urban
laurent.urban@univ-avignon.fr

Specialty section:

This article was submitted to
Plant Physiology,
a section of the journal
Frontiers in Plant Science

Received: 21 September 2016

Accepted: 25 October 2016

Published: 11 November 2016

Citation:

Ripoll J, Bertin N, Bidel LPR and
Urban L (2016) A User's View of the
Parameters Derived from the
Induction Curves of Maximal
Chlorophyll *a* Fluorescence:
Perspectives for Analyzing Stress.
Front. Plant Sci. 7:1679.
doi: 10.3389/fpls.2016.01679

Analysis of the fast kinetics of the induction curve of maximal fluorescence represents a relatively recent development for chlorophyll *a* fluorescence measurements. The parameters of the so-called JIP-test are exploited by an increasingly large community of users to assess plant stress and its consequences. We provide here evidence that these parameters are capable to distinguish between stresses of different natures or intensities, and between stressed plants of different genetic background or at different developmental stages at the time of stress. It is, however, important to keep in mind that the JIP-test is inherently limited in scope, that it is based on assumptions which are not fully validated and that precautions must be taken to ensure that measurements are meaningful. Recent advances suggest that some improvements could be implemented to increase the reliability of measurements and the pertinence of the parameters calculated. We moreover advocate for using the JIP-test in combination with other techniques to build comprehensive pictures of plant responses to stress.

Keywords: chlorophyll *a* fluorescence, JIP-test, potential and limits, *Solanum lycopersicum* L., stress response, water deficit

INTRODUCTION

There is a growing interest for ChlF since the last 15 years (e.g., 14400 articles in 2015 vs. 4870 in 2000; research made on google scholar using “chlorophyll *a* fluorescence” as a keyword). When excitation energy arrives in a RC where the donor side cannot evacuate energy toward an acceptor, energy is essentially lost under the form of heat and ChlF. Measurements of ChlF therefore gives insight into efficiencies of energy transfer and heat dissipation. Stress may impact all the steps from light energy absorption to electron transfer to the final acceptors. So, ChlF can be used to characterize the effects of stress on adaptive mechanisms (Misra et al., 2012; Kalaji et al., 2014, 2016).

There are several types of instruments for analyzing ChlF. Steady-state instruments are designed for quenching analysis and for coupled measurements of ChlF and gas exchanges which give insight into downstream processes. The focus here is on the analysis of OJIP fluorescence transients (JIP-test) which become increasingly popular among users, thanks to the development of relatively cheap and users-friendly devices (Kalaji et al., 2014). In

Abbreviations: CEF, cyclic electron flux; ChlF, chlorophyll *a* fluorescence; KMC, Kinetic Monte Carlo; NPQ, non-photochemical quenching; PC, plastocyanin; PQ, plastoquinone; PS, photosystem; RC, reaction center; WD, water deficit.

this paper, we shall exploit the experience we have gained using the JIP-test for evaluating the effects of different intensities of WD on tomato plants according to genetic diversity and to plant developmental stage at the time of stress, to discuss its potential. We shall also stress some theoretical and practical limitations of the JIP-test and evoke its potential when combined to other techniques.

THE OJIP/OLKJIP MODEL: PRINCIPLE AND SHORT DESCRIPTION

The OJIP model allows to analyze the ChlF induction curve when a leaf acclimated to dark conditions is suddenly exposed to a saturating pulse of light (Kautsky effect). The induction curve appears as a fast wave (ca. 0.3 s) with characteristic steps named O, J, I, and P, plotted on a logarithmic time scale, starting from initial fluorescence F_0 (dark adapted) to maximal fluorescence F_M (light-saturated; Strasser and Strasser, 1995). Level O corresponds to the initial fluorescence emitted, whereas levels J and I correspond to the fluorescence emitted after, respectively, 2 and 30 ms. The level P corresponds to F_M (Strasser et al., 2000). Under specific conditions, like heat stress, another inflection in the induction curve can appear around 300 μ s, called K. Eventually a shift of the induction curve between 50 and 300 μ s, influenced by the excitation energy transfer between PS II units, may appear, the so-called L band (Strasser and Stirbet, 1998).

The JIP-test is based on several assumptions (Stirbet and Govindjee, 2011). The most important assumption is that the fluorescence increase from F_0 to F_M reflects mainly the redox state of Q_A protein (Kalaji et al., 2014) in PSII RC. This basic assumption is a matter of debate (Schansker et al., 2014). For some authors, the alterations induced at the acceptor side of PSII RC during the rapid turnover of oxidation of the Q_B pool at the Q_B site may be essential in the triggering of photoinactivation and D1 protein damage (Gong and Ohad, 1991). Within this view it is the Q_B state occupancy which has the highest influence on fluorescence yield (Zivčák et al., 2015). The JIP-test is not only based on the assumption that the F_0 to F_M rise reflects the Q_A redox state, but also on the assumption that NPQ processes do not hinder the rise to F_M .

The JIP-test can be used as a signature of (1) diverse events translating into changes in the redox state of the components of the linear electron transport flow, (2) the involvement of alternative electron routes, (3) the build-up of a transmembrane pH gradient (and membrane potential), (4) the activation of different NPQ processes, (5) the activation of the Calvin-Benson cycle (Stirbet et al., 2014). The OJ section reflects the reduction of the acceptor side of PSII, the JI section the partial reduction of the PQ pool and finally the IP section the reduction of the acceptor side of PSI. The reader will find an excellent introduction to the parameters derived from the JIP-test mathematical model in Strasser et al. (2004) and Stirbet and Govindjee (2011) for instance. The key idea underpinning stress characterization and analysis using the JIP-test, is that stress necessarily impacts the efficiencies and fluxes of electrons and of energy in and around

PSI and PSII, and that their variations can be assessed and analyzed using the parameters derived from the OJIP/OLKJIP transients (Maxwell and Johnson, 2000).

WHAT CAN WE LEARN FROM THE PARAMETERS OF THE JIP-TEST?

Plants have to adapt to the risk of photooxidative damage which results from the imbalance between the incoming energy under the form of photon flux and the energy quenched by photosynthetic processes (Gururani et al., 2015). Stress, for instance by limiting stomatal conductance and CO_2 supply to the Calvin-Benson cycle, exacerbates the risk of photooxidative damage. Therefore, stress triggers adaptive responses aiming at reducing the quantity of energy entering the leaf, reducing the amount of absorbed energy converted into electron flux, and rerouting electron fluxes. Each step of OJIP/OLKJIP curves can be associated to the efficiency of energy or electron transfers between the components of PSII and PSI (Figure 1).

The JIP-test provides invaluable parameters to analyze upstream adaptive mechanisms to different types of stress (Kalaji et al., 2016). It is, however, important to keep in mind that no single parameter derived from the JIP-test can be considered as specific of a given type of stress. It is rather a combination of parameters that may be considered as relevant. The rate of energy dissipation by processes other than trapping expressed either per RCs or on an absorbed energy basis J_0^{DI}/RC or J_0^{DI}/J^{ABS} is used to evaluate heat dissipation processes. The ratio $V_K/V_J = [(F_{0.3\text{ms}} - F_0)/(F_{2\text{ms}} - F_0)]$ is associated to limitation/inactivation and possibly damage of the oxygen-evolving complex. The I-P phase, consequently the rate of electron transport from Q_B to PSI acceptors J_0^{RE1}/RC or J_0^{RE1}/J^{ABS} is considered to give insight into the CEF (Harbinson and Foyer, 1991; Schansker et al., 2014; Zivčák et al., 2015). The CEF contributes to the balance of the ATP/NADPH output ratio and can provide protection against photooxidative stress (Martin et al., 2004; Huang et al., 2016), offsetting the decline of the linear electron flow under WD (Mladenov et al., 2015).

There is an increasing amount of evidence suggesting that the JIP-test is a discriminating one. The JIP-test is capable not only to assess different kind of stresses but also to distinguish between specific responses for a given type of stress, due, for instance, to genotypic differences, to differences in stress intensity, or to differences in the developmental stage of plants at the time stress is applied. JIP parameters were found to be capable to distinguish among tree species (Pollastrini et al., 2016). The above-mentioned parameters, J_0^{DI}/J^{ABS} , V_K/V_J and J_0^{RE1}/J^{ABS} , were found to be relevant parameters to evaluate responses to WD as a function of genotype diversity for a given plant species, in barley (Oukarroum et al., 2007, 2009) and in tomato (Table 1). Considering differences according to stress intensity, we observed a difference in J_0^{RE1}/J^{ABS} between tomato plants at the reproductive stage submitted to severe WD and tomato plants at the same stage of development submitted to repeated cycles of WD and recovery (Table 1). We also found differences due to developmental stage at the time of stress since we observed

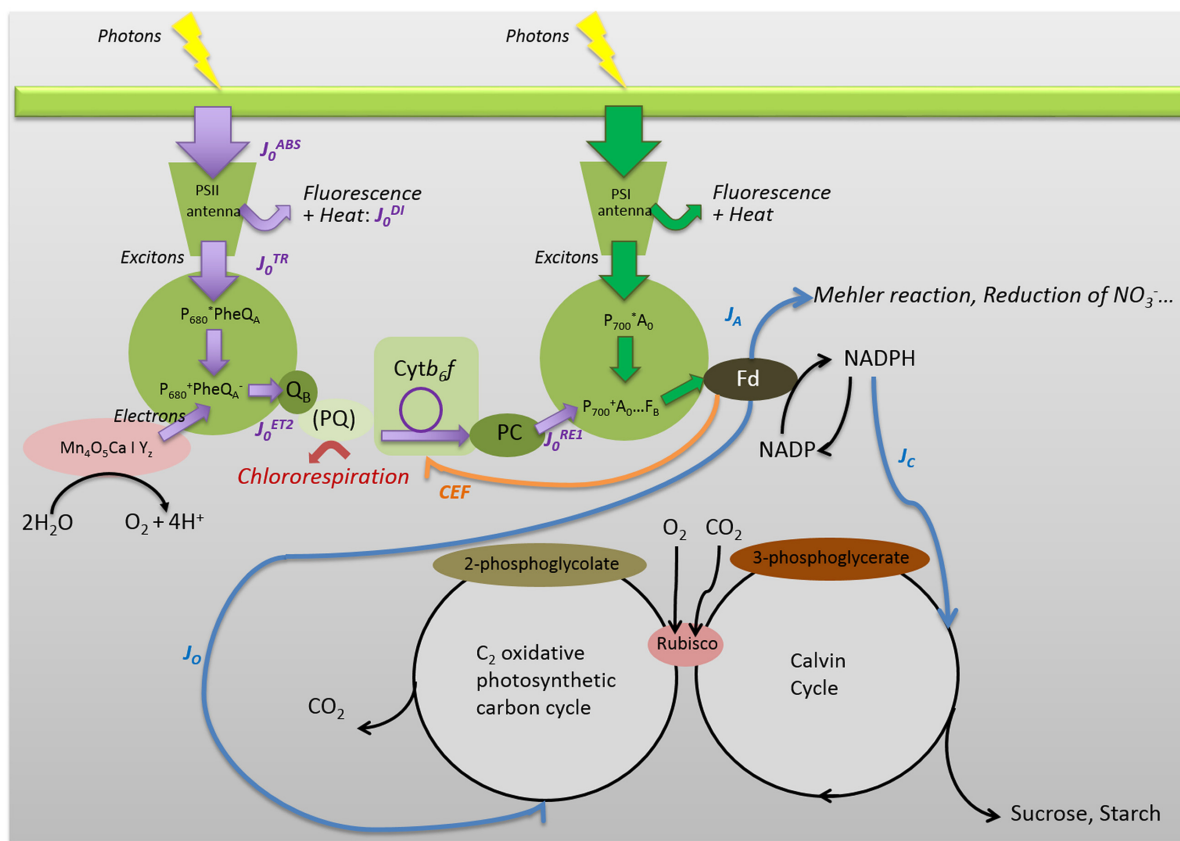


FIGURE 1 | Simplified representation of the main energy pathways in and around PSI and PSII, down to ferredoxin (Fd) and downstream. Some of the JIP-test parameters were indicated. Since most ChlF originates in PSII antenna, J_0^{ABS} represents the rate of photon absorption by all PSII antenna pigments. The dissipated energy flux J_0^{DI} represents the part of the absorbed photon flux dissipated through direct fluorescence and other non-radiative processes (as heat), and the trapped exciton flux J_0^{TR} represents the rate of exciton trapping by the PSII RC P_{680} . The trapped energy is used for charge separation using the components of the PSII RC, i.e., pheophytin molecules (Phe) and Q_A and Q_B quinones [linked to D1 and D2 proteins not represented, Q_A (bound to D2) and Q_B (bound to D1)]. The complex $Mn_4O_5CaIY_2$ corresponds to the oxygen-evolving complex. The flux J_0^{ET2} represents the electron transport flux from Q_A to Q_B . J_0^{RE1} represents the rate of electrons from Q_B to PSI acceptors. J_C , J_O , and J_A represent the electron flows for carboxylation, oxygenation, and alternative sinks, respectively. PQ and PC represent plastoquinones and plastocyanins. Violet arrows are associated to the JIP-test. Blue arrows correspond to the fluxes evaluated from combined measurements of modulated ChlF and gas exchanges (Valentini et al., 1995). The orange arrow corresponds to the CEF (Kotakis et al., 2006) and the red to the chlororespiration (Rumeau et al., 2007).

an increase in J_0^{DI}/RC and J_0^{DI}/J_0^{ABS} in tomato plants at the reproductive stage submitted to severe WD (with the exception of LA1420), whereas there was an increase in F_0 and a decrease in S_m (the normalized area of the maximal fluorescence induction curve) in plants at the vegetative stage submitted to a similar stress (Table 1). The latter shifts are suspected to be indicators of damage (Christen et al., 2007; Yordanov et al., 2008).

It is near to impossible not to evoke the popular Performance Index (PI) of Strasser when discussing the parameters derived from the JIP-test (Silvestre et al., 2014; Zivčák et al., 2014). Recently Kalaji et al. (2016) recommended the non-specialist to resort to the PI in the absence of a serious capacity to understand and exploit the other parameters. We experienced that the PI is not always as easy to interpret as usually believed. For instance, the 19.8% decrease in PI_{ABS} of LA1420 plants at the reproductive stage submitted to repeated cycles of WD and recovery (generally believed to favor acclimation) withstands interpretation since

PI_{ABS} did not decrease in similar plants submitted to severe WD (Table 1). One would have expected the reverse. Our opinion is that it is generally more rewarding to make use of the full set of parameters that can be derived from the JIP-test.

LIMITATIONS ORIGINATING FROM THE THEORETICAL BACKGROUND, RECENT ADVANCES AND PROSPECTS OFFERED BY COMBINING APPROACHES

There are several limitations associated with the JIP-test, some arising from the physiological assumptions behind the theory and others concerning good practices (Murchie and Lawson, 2013; Kalaji et al., 2014). As said before, it is essential to keep in mind that the JIP-test model is based on a sum of

TABLE 1 | Relative differences for the parameters derived from JIP-test, performed on 30 min dark-adapted leaves with a Plant Efficiency Analyzer (Hansatech Instrument, King's Lynn, UK) on vegetative and reproductive plants of Cervil, LA1420, PlovdivXXIVa and Levovil tomato accessions exposed to two WD treatments.

	Severe water deficit Vegetative plants				Severe water deficit Reproductive plants				Repeated cycles of water deficit Reproductive plants			
	Cervil	LA1420	Plovdiv	Levovil	Cervil	LA1420	Plovdiv	Levovil	Cervil	LA1420	Plovdiv	Levovil
	F_0	15.0	21.6	-1.6	10.3	-4.8	-0.5	2.0	3.2	5.3	-3.1	2.4
F_m	8.9	-6.9	-3.3	7.5	-9.8	2.3	-6.8	-6.1	-1.3	-12.5	-9.8	-12.4
Technical fluorescence parameters												
Sm	-71.6	-32.9	-34.2	-28.2	-1.6	17.9	-9.3	0.4	10.0	10.1	6.1	1.7
N	-74.2	-34.9	-38.9	-39.4	2.3	11.9	-9.0	1.4	12.2	10.3	6.8	2.9
V_k/V_j	-7.6	1.6	-7.2	-15.5	4.9	-4.4	1.0	1.3	-4.8	-3.0	-2.2	-4.3
Specific energy fluxes per reaction centers (RC)												
J_0^{ABS}/RC	-6.5	10.3	-6.8	-15.0	6.3	-5.1	2.0	2.7	-3.1	-0.7	0.4	-1.8
J_0^{PI}/RC	-1.7	56.8	-5.8	-13.1	12.0	-7.3	11.7	12.9	4.6	11.1	15.0	11.5
J_0^{TR}/RC	-7.8	1.7	-7.2	-15.5	5.1	-4.6	0.4	0.8	-4.7	-2.9	-2.3	-4.4
J_0^{ET2}/RC	-13.3	-17.1	-16.2	-17.0	-10.0	-8.6	-10.1	-12.2	-8.6	-5.0	-8.5	-13.2
J_0^{REI}/RC	-46.4	-40.7	-47.4	-37.3	-19.1	-10.6	-29.2	-23.7	6.3	18.0	8.7	9.1
Efficiencies and probabilities												
J_0^{PI}/J_0^{ABS}	5.9	34.0	1.5	2.5	7.0	-3.2	9.9	9.3	8.0	11.7	14.3	13.7
J_0^{TR}/J_0^{ABS}	-1.2	-6.9	-0.2	-0.7	-1.3	0.6	-1.7	-1.7	-1.7	-2.2	-2.7	-2.7
J_0^{ET2}/J_0^{ABS}	-6.9	-18.3	-9.7	-1.8	-14.4	-3.1	-11.6	-14.3	-5.2	-4.5	-8.7	-12.0
J_0^{REI}/J_0^{ABS}	-42.6	-38.5	-43.3	-26.7	-22.3	-7.0	-30.2	-23.1	9.1	17.6	7.9	11.1
J_0^{REI}/J_0^{ET2}	-38.8	-27.7	-37.9	-24.5	-11.7	-3.4	-21.5	-11.5	18.7	24.3	20.5	29.3
Performance indexes												
PI_{ABS}	-14.2	-50.6	-14.6	10.6	-37.3	-0.5	-34.1	-39.7	-12.6	-19.8	-19.4	-32.0
PI_{ABS}^{TOT}	-66.6	-45.9	-65.2	-30.8	-40.0	-5.1	-53.0	-50.1	-3.6	-14.2	-20.7	-25.1
Color scale	-80	-45	0	30	60							

The first treatment consisted in a severe WD (10 days without water) applied both at the vegetative stage (30 days after sowing) and at the reproductive stage (90 days after sowing). The second treatment consisted in three repeated cycles of WD of increasing intensity followed by recovery periods (Ripoll et al., 2016), during plant development (from 40 to 100 days after sowing). Relative differences were calculated based on means of initial values vs. stressed values for the severe WD and on total area under the curve of the stressed versus means of control values for the repeated cycles of WD treatment (n ≥ 5). The percentages were scaled by color (green for high and red for low values). Significant differences are indicated by using bold, italic, and underlined fonts (P < 0.05). The Tukey test was used when conditions of ANOVA are respected, i.e., for F_0 and F_m for vegetative plants; and for J_0^{ABS}/RC , J_0^{PI}/J_0^{ABS} and PI_{ABS} for reproductive plants. Alternatively, the Kruskal–Wallis test was used.

assumptions (Stirbet and Govindjee, 2011). For instance, during the measurement of an OJIP transient, all PSII units are considered to be homogeneous and active, which is probably not true (Vredenberg, 2011). Recent mathematical models using KMC simulation can help to deal with this limitation (Guo and Tan, 2011, 2014). KMC simulation should help to take into account the variability in the number of RCs, in PQ pool size, in the number of active Q_B sites and in Q_A reduction rate events (Guo and Tan, 2011, 2014).

However, the information supplied by JIP parameters does not allow for comprehensive interpretation of the adaptive strategies adopted by stressed plants. This is a shortcoming inherent to the fact that all the information derived from the JIP-test is about energy and electron fluxes and transfer efficiencies upstream PSI, whereas it is quite clear that downstream allocation of electron fluxes among the Calvin-Benson cycle, photorespiration and alternative electron sinks play a key-role along with antioxidant mechanisms in the strategy of plants facing photooxidative damage (Figure 1). The fluorescence steps beyond F_M so-called PSMT phase (Kalaji et al., 2016) could be used for analyses in relation to the activation of the ferredoxin-NADP⁺ reductase and the Calvin-Benson cycle through the ferredoxin-thioredoxin system (Stirbet et al., 2014). So far, unfortunately, the PSMT phase appears less reproducible than the OJIP phase (Stirbet et al., 2014; Vredenberg, 2015).

Recent studies bridged the gap between the scientific sub-communities by associating analysis of the OJIP transients, measurements of gas exchanges and simultaneous measurements of PSI and PSII activities, with the objective to characterize PSI functioning (Brestič et al., 2014; Zivčák et al., 2015). Such approaches should be more developed in the future to build broader pictures of the mechanisms of plant acclimation to stress at play both before and beyond PSI. The information obtained could possibly be used to improve the PSMT model and to gain new insight in the functioning of the components of the photosynthetic machinery (Belyaeva et al., 2016). Of course there is also ample room for progress by studying jointly parameters

derived from ChlF measurements and molecular and biochemical markers (Hao et al., 2012; Mladenov et al., 2015; Yin et al., 2015).

CONCLUDING REMARKS

JIP parameters are gaining recognition among plant biologists besides other indicators of physiological status (Chen et al., 2014; Wituszyńska et al., 2015). There is little doubt that improvements and novel techniques like JIP-test imaging (Jedrowski and Brüggemann, 2015) will go on fueling the interest of the scientific community for these parameters in the future, possibly in phenotyping platforms. The potential of JIP parameters to distinguish between plant stress responses and to assess genetic diversity is more and more well recognized. However, for interesting they are, the parameters derived from the JIP-test have inherent limitations. We therefore recommend to associate to JIP parameters to parameters derived from combined measurements of gas-exchanges and steady-state ChlF, and even to other molecular or biochemical markers, to get the most comprehensive pictures possible of the plant adaptive mechanisms involved in stress responses.

AUTHOR CONTRIBUTIONS

JR, NB, LB, and LU compiled data, developed theory and wrote the paper. JR performed the experiments and analyses.

ACKNOWLEDGMENTS

JR was supported by a Ph.D. fellowship of the *Federative Research Structure Tersys*. The authors thank the team of the UR1052 Genetics and Improvement of Fruit and Vegetables (INRA, Montfavet) for providing plant material and the team of the UR1115 Plants and cropping Systems in Horticulture (INRA, Avignon) for technical assistance.

REFERENCES

- Belyaeva, N. E., Bulychev, A. A., Riznichenko, G. Y., and Rubin, A. B. (2016). Thylakoid membrane model of the Chl a fluorescence transient and P700 induction kinetics in plant leaves. *Photosynth. Res.* 130, 491–515. doi: 10.1007/s11120-016-0289-z
- Brestič, M., Zivčák, M., Olšovská, K., Shao, H. B., Kalaji, H. M., and Allakhverdiev, S. I. (2014). Reduced glutamine synthetase activity plays a key role in control of photosynthetic responses to high light in barley leaves. *Plant Physiol. Biochem.* 81, 74–83. doi: 10.1016/j.plaphy.2014.01.002
- Chen, S., Kim, C., Lee, J. M., Lee, H. A., Fei, Z., Wang, L., et al. (2014). Blocking the Q_B -binding site of photosystem II by tenuazonic acid, a non-host specific toxin of *Alternaria alternata*, activates singlet oxygen-mediated and EXECUTER-dependent signalling in *Arabidopsis*. *Plant Cell Environ.* 38, 1069–1080. doi: 10.1111/pce.12462
- Christen, D., Schönmann, S., Jermini, M., Strasser, R. J., and Défago, G. (2007). Characterization and early detection of grapevine (*Vitis vinifera*) stress responses to esca disease by in situ chlorophyll fluorescence and comparison with drought stress. *Environ. Exp. Bot.* 60, 504–514. doi: 10.1016/j.envexpbot.2007.02.003
- Gong, H., and Ohad, I. (1991). The PQ/PQH2 ratio and occupancy of photosystem II- Q_B site by plastoquinone control the degradation of D1 protein during photoinhibition in vivo. *J. Biol. Chem.* 266, 21293–21299.
- Guo, Y., and Tan, J. (2011). Modeling and simulation of the initial phases of chlorophyll fluorescence from photosystem II. *Biosystems* 103, 152–157. doi: 10.1016/j.biosystems.2010.10.008
- Guo, Y., and Tan, J. (2014). Kinetic Monte Carlo simulation of the initial phases of chlorophyll fluorescence from photosystem II. *Biosystems* 115, 1–4. doi: 10.1016/j.biosystems.2013.10.004
- Gururani, M. A., Venkatesh, J., and Tran, L. S. P. (2015). Regulation of photosynthesis during abiotic stress-induced photoinhibition. *Mol. Plant.* 8, 1304–1320. doi: 10.1016/j.molp.2015.05.005
- Hao, D., Chao, M., Yin, Z., and Yu, D. (2012). Genome-wide association analysis detecting significant single nucleotide polymorphisms for chlorophyll and chlorophyll fluorescence parameters in soybean (*Glycine max*) landraces. *Euphytica* 186, 919–931. doi: 10.1007/s10681-012-0697-x
- Harbison, J., and Foyer, C. H. (1991). Relationships between the efficiencies of photosystems I and II and stromal redox state in CO₂-free air. Evidence for cyclic electron flow in vivo. *Plant Physiol.* 97, 41–49. doi: 10.1104/pp.97.1.41

- Huang, W., Yang, Y.-J., Hu, H., Zhang, S.-B., and Cao, K.-F. (2016). Evidence for the role of cyclic electron flow in photoprotection for oxygen-evolving complex. *J. Plant Physiol.* 194, 54–60. doi: 10.1016/j.jplph.2016.02.016
- Jedemski, C., and Brüggemann, W. (2015). Imaging of fast chlorophyll fluorescence induction curve (OJIP) parameters, applied in a screening study with wild barley (*Hordeum spontaneum*) genotypes under heat stress. *J. Photochem. Photobiol. B* 151, 153–160. doi: 10.1016/j.jphotobiol.2015.07.020
- Kalaji, H. M., Jajoo, A., Oukarroum, A., Brestic, M., Zivcak, M., Samborska, I. A., et al. (2016). Chlorophyll a fluorescence as a tool to monitor physiological status of plants under abiotic stress conditions. *Acta Physiol. Plant.* 38, 102. doi: 10.1007/s11738-016-2113-y
- Kalaji, H. M., Schansker, G., Ladle, R. J., Goltsev, V., Bosa, K., Allakhverdiev, S. I., et al. (2014). Frequently asked questions about in vivo chlorophyll fluorescence: practical issues. *Photosynth. Res.* 122, 121–158. doi: 10.1007/s11120-014-0024-6
- Kotakis, C., Petropoulou, Y., Stamatakis, K., Yirotis, C., and Manetas, Y. (2006). Evidence for active cyclic electron flow in twig chlorenchyma in the presence of an extremely deficient linear electron transport activity. *Planta* 225, 245–253. doi: 10.1007/s00425-006-0327-8
- Martin, M., Casano, L. M., Zapata, J. M., Guéra, A., Del Campo, E. M., Schmitz-Linneweber, C., et al. (2004). Role of thylakoid Ndh complex and peroxidase in the protection against photo-oxidative stress: fluorescence and enzyme activities in wild-type and ndhF-deficient tobacco. *Physiol. Plant.* 122, 443–452. doi: 10.1111/j.1399-3054.2004.00417.x
- Maxwell, K., and Johnson, G. N. (2000). Chlorophyll fluorescence, a practical guide. *J. Exp. Bot.* 51, 659–668. doi: 10.1093/jexbot/51.345.659
- Misra, A. N., Misra, M., and Singh, R. (2012). “Chlorophyll fluorescence in plant biology,” in *Biophysics*, ed. A. N. Misra (Shanghai: Intech), 171–192.
- Mladenov, P., Finazzi, G., Bligny, R., Moyankova, D., Zaslava, D., Boisson, A. M., et al. (2015). In vivo spectroscopy and NMR metabolite fingerprinting approaches to connect the dynamics of photosynthetic and metabolic phenotypes in resurrection plant *Haberlea rhodopensis* during desiccation and recovery. *Front. Plant Sci.* 6:564. doi: 10.3389/fpls.2015.00564
- Murchie, E. H., and Lawson, T. (2013). Chlorophyll fluorescence analysis: a guide to good practice and understanding some new applications. *J. Exp. Bot.* 64, 3983–3998. doi: 10.1093/jxb/ert208
- Oukarroum, A., Madidi, S. O. E., Schansker, G., and Strasser, R. J. (2007). Probing the responses of barley cultivars (*Hordeum vulgare* L.) by chlorophyll a fluorescence OLKJIP under drought stress and re-watering. *Environ. Exp. Bot.* 60, 438–446. doi: 10.1016/j.envexpbot.2007.01.002
- Oukarroum, A., Schansker, G., and Strasser, R. J. (2009). Drought stress effects on photosystem I content and photosystem II thermotolerance analysed using Chl a fluorescence kinetics in barley varieties differing in their drought tolerance. *Physiol. Plant.* 137, 188–199. doi: 10.1111/j.1399-3054.2009.01273.x
- Pollastrini, M., Holland, V., Brüggemann, W., Bruelheide, H., Danila, I., Jaroszewicz, B., et al. (2016). Taxonomic and ecological relevance of the chlorophyll a fluorescence signature of tree species in mixed European forests. *New Phytol.* 212, 51–65. doi: 10.1111/nph.14026
- Ripoll, J., Urban, L., and Bertin, N. (2016). The Potential of the MAGIC TOM parental accessions to explore the genetic variability in tomato acclimation to repeated cycles of water deficit and recovery. *Front. Plant Sci.* 6:1172. doi: 10.3389/fpls.2015.01172
- Rumeau, D., Peltier, G., and Cournac, L. (2007). Chlororespiration and cyclic electron flow around PSI during photosynthesis and plant stress response. *Plant Cell Environ.* 30, 1041–1051. doi: 10.1111/j.1365-3040.2007.01675.x
- Schansker, G., Toth, S. Z., Holzwarth, A. R., and Garab, G. (2014). Chlorophyll a fluorescence: beyond the limits of the QA model. *Photosynth. Res.* 120, 43–58. doi: 10.1007/s11120-013-9806-5
- Silvestre, S., de Sousa Araújo, S., Carlota Vaz Patto, M., and Marques da Silva, J. (2014). Performance index: an expeditious tool to screen for improved drought resistance in the *Lathyrus* genus. *J. Int. Plant Biol.* 56, 610–621. doi: 10.1111/jipb.12186
- Stirbet, A., and Govindjee. (2011). On the relation between the Kautsky effect (chlorophyll a fluorescence induction) and Photosystem II: basics and applications of the OJIP fluorescence transient. *J. Photochem. Photobiol. B* 104, 236–257. doi: 10.1016/j.jphotobiol.2010.12.010
- Stirbet, A., Riznichenko, G. Y., Rubin, A. B., and Govindjee. (2014). Modeling chlorophyll a fluorescence transient: relation to photosynthesis. *Biochemistry (Mosc.)* 79, 291–323. doi: 10.1134/S0006297914040014
- Strasser, B. J., and Strasser, R. J. (1995). *Measuring Fast Fluorescence Transients to Address Environmental Questions: the JIP-test*. Dordrech: Kluwer Academic Publishers.
- Strasser, R. J., Srivastava, A., and Tsimilli-Michael, M. (2000). “The fluorescence transient as a tool to characterize and screen photosynthetic samples,” in *Probing Photosynthesis: Mechanism, Regulation and Adaptation*, eds M. Yunus, U. Pathre, and P. Mohanty (London: Taylor and Francis), 445–483.
- Strasser, R. J., and Stirbet, D. A. (1998). Heterogeneity of photosystem II probed by the numerically simulated chlorophyll a fluorescence rise (O-J-I-P). *Math. Comput. Simulat.* 48, 3–9. doi: 10.1016/j.jtbi.2008.11.018
- Strasser, R. J., Tsimilli-Michael, M., and Srivastava, A. (2004). “Analysis of the chlorophyll a fluorescence transient,” in *Chlorophyll a Fluorescence*, eds G. Papageorgiou and Govindjee (Dordrecht: Springer), 321–362.
- Valentini, R., Epron, D., De Angelis, P., Matteucci, G., and Dreyer, E. (1995). In situ estimation of net CO₂ assimilation, photosynthetic electron flow and photorespiration in Turkey oak (*Q. cerris* L.) leaves: diurnal cycles under different levels of water supply. *Plant Cell Environ.* 18, 631–640. doi: 10.1111/j.1365-3040.1995.tb00564.x
- Vredenberg, W. (2011). Kinetic analyses and mathematical modeling of primary photochemical and photoelectrochemical processes in plant photosystems. *Biosystems* 103, 138–151. doi: 10.1016/j.biosystems.2010.10.016
- Vredenberg, W. (2015). A simple routine for quantitative analysis of light and dark kinetics of photochemical and non-photochemical quenching of chlorophyll fluorescence in intact leaves. *Photosynth. Res.* 124, 87–106. doi: 10.1007/s11120-015-0097-x
- Wituszyńska, W., Szechyńska-Hebda, M., Sobczak, M., Rusaczonok, A., Kozłowska-Makulska, A., Witoń, D., et al. (2015). Lesion simulating disease 1 and enhanced disease susceptibility 1 differentially regulate UV-C-induced photooxidative stress signalling and programmed cell death in *Arabidopsis thaliana*. *Plant Cell Environ.* 38, 315–330. doi: 10.1111/pce.12288
- Yin, Z., Qin, Q., Wu, F., Zhang, J., Chen, T., Sun, Q., et al. (2015). Quantitative trait locus mapping of chlorophyll a fluorescence parameters using a recombinant inbred line population in maize. *Euphytica* 205, 25–35. doi: 10.1007/s10681-015-1380-9
- Yordanov, I., Goltsev, V., Stefanov, D., Chernev, P., Zaharieva, I., Kirova, M., et al. (2008). Preservation of photosynthetic electron transport from senescence-induced inactivation in primary leaves after decapitation and defoliation of bean plants. *J. Plant Physiol.* 165, 1954–1963. doi: 10.1016/j.jplph.2008.05.003
- Zivčák, M., Brestič, M., Kunderlikova, K., Olšovská, K., and Allakhverdiev, S. I. (2015). Effect of photosystem I inactivation on chlorophyll a fluorescence induction in wheat leaves: does activity of photosystem I play any role in OJIP rise? *J. Photochem. Photobiol. B* 152, 318–324. doi: 10.1016/j.jphotobiol.2015.08.024
- Zivčák, M., Olšovská, K., Slamka, P., Galambošová, J., Rataj, V., Shao, H. B., et al. (2014). Application of chlorophyll fluorescence performance indices to assess the wheat photosynthetic functions influenced by nitrogen deficiency. *Plant Soil Environ.* 60, 210–215.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Ripoll, Bertin, Bidel and Urban. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.