

# Corpus luteum vascularization and progesterone production in Autumn and Winter cycles of the mare: relationship between ultrasonographic characteristics of corpora lutea and plasma progesterone concentration in the last cycles before anestrus

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Corpus luteum vascularization and progesterone production in Autumn and Winter cycles of
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- 8
- 9 Abstract:

In 20 estrus cycles of 15 mares, Color Doppler ultrasound of corpora lutea and plasma progesterone 10 concentration ([P4]) were analyzed on days 6, 10, 14, 16 and 18 after ovulation. [P4] was positively 11 12 correlated with corpora lutea cross sectional area (CSA), vascularized area (VA) and index of 13 vascularization (IV=VA/CSA) (P<0.0001). CSA, VA and IV in corpora lutea of mares with [P4] < 1 ng/ml were significantly lower than in corpora lutea of mares with [P4] > 1 ng/ml. Mares with 14 15 CSA <3473 pixels, VA<25.5 pixels and a IV <7.6% were prone to express [P4] <1 ng/ml 25.4, 7.9 16 and 7.6 times more than mares with higher values, respectively. CLs analyzed parameters differed 17 significantly between last cycles of the breeding season and previous cycles until day 14 after ovulation (P<0.05). No significant differences were found in [P4] between last cycles and previous 18 19 ones.

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- 20
- 21 Keywords:
- 22 Mare; Corpus Luteum; Color Doppler Ultrasound; Progesterone; Fall Transition
- 23
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#### 27 **1. Introduction:**

Mares' autumn transition into anestrus is a process still not completely explored. In the northern hemisphere temperate regions, although most mares stop cycling between November and January [1] anestrus can begin between August and February [1-3]. Factors analyzed to explain seasonality in the mare have been: latitude [4], age [4,5], recent reproductive history (foaled or barren) [5] and nutrition and fat reserves (leptin blood concentration) [6,7] In non-pregnant mares, corpus luteum (CL) progesterone production seems to decline significantly from one cycle to the subsequent at the end of the breeding season [1,3,8-10].

Also luteolysis appears to be negatively affected by fall transition, with an increase of the incidence
of a persistent corpus luteum (25%) compared to the breeding season (8-10%) [11].

Color Doppler ultrasound has been applied for the evaluation of CL vascularization and perfusion
in cattle and horses [12-19]. Corpora lutea maximum diameter [18,19] and vascularization
[12,16,18,19] has been significantly correlated with plasma progesterone concentration during
diestrus and pregnancy in the mare.

Diagnostic accuracy studies address how well a test identifies the target condition. Sensitivity, specificity and Likelihood Ratios (LRs) are all different ways of expressing test performances [20]. In the bovine species, to define CLs functionality, CL diameter [21], area, and vascularization area [22] cutoffs have been defined when plasma [P4] was above 1 ng/ml. Sensitivity and specificity (expressed in %) were defined as the probability to have a CL above a definite diameter [21] or area and vascularization [22] cutoffs when [P4] was above 1 or below 1 ng/ml, respectively.

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Aims of this study were: to describe a cutoff to predict the CL functionality based on CL diameter
and vascularization and to verify the existence of differences on CLs diameter, vascularization and
P4 production between the last cycle before anestrus and previous cycles.

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#### 52 **2.** Materials and methods:

53 Animals, estrus cycles monitoring and blood sampling - Between October 2013 and January 2014, 54 15 barren, cyclic Standardbred mares aged 7 to 14 years, maintained in paddocks and fed with a 55 balanced ratio of hay and commercial horse fodder, were studied for a total of 20 estrous cycles. 56 Body condition score of the mares at the beginning of the study was 5-6 (where 1 is thin, 5 is 57 moderate and 9 is obese) [23]. Ovarian activity was monitored twice a week for the whole period and daily during estrus until ovulation, by B-mode trans-rectal ultrasound (MyLab<sup>TM</sup> 30 Gold 58 machine equipped with a 5.0-7.5 MHz linear probe and a Color Doppler function; Esaote S.p.A, 59 60 Florence, Italy). Mares were considered fallen into anestrus when no ovulations or CLs were 61 evidenced by ultrasound for at least 4 weeks. On days 6, 10, 14, 16 and 18 after ovulation, CLs were scanned by Color Doppler (n= 20) and blood was collected by jugular venipuncture (n= 11). 62 63 Blood was centrifuged, and plasma separated and stored at -20 °C until [P4] determination.

Color Doppler corpus luteum scanning -. For every CL, three images were taken in color flow 64 65 mode at a standard depth of 10 cm, frequency of 5.0 MHz, 70% gain, and with a pulse repetition 66 frequency (PRF) of 2.8, at maximum cross sectional diameter. Images taken were analyzed using 67 ImageJ software (National Institutes of Health, Bethesda, USA). Cross sectional areas (CSA) of the 68 CL were measured and any lacuna present was excluded. Images were cropped and colored zones 69 selected (vascularized area, VA). The CSA and VA areas of the CL were measured and evaluated 70 by counting the number of grey and color pixels, for each image taken respectively. The mean CSA 71 and VA areas were calculated using the computer-assisted image analysis software, ImageJ 72 (National Health Institutes, Bethesda, USA) and the mean ratio of VA to CSA was taken as an 73 index of vascularization (IV) [18,19].

*Plasma progesterone evaluation method* – Plasma progesterone was analyzed with an ELISA assay adapted from Canepa et al 2008 [24]. To summarize, the monoclonal antibody anti-P4 was obtained from the mouse against P4 conjugated to bovine serum albumin. This antibody diluted at 1/160000 was aliquoted on 94 wells of a 96 wells plate (Immuno Maxisorp C96, Nunc) previously coated with an anti-mouse immunoglobulins G obtained in goat. After that, 10µl of each sample or of each

79 standard were added. After incubation, a solution of progesterone- $11\alpha$ -hemi-succinate-phosphatase 80 alkaline conjugate was distributed in all the wells. After one hour, the plate was washed and a solution with p-nitrophenylphosphate and 100mM Diethanolamine 5mM MgCl2 was added. The 81 82 plates were read at 405 nm in a photometer. For the validation of this method for equine plasma, a 83 test of dilution was done. On this first test, the correlation coefficient was 0.992 between expected and observed data with a slope of 1.04. A test with an addition of P4 was done, and the correlation 84 coefficient between expected and observed data was 0.914 with a slope of 0.97. The resulting 85 86 coefficient of inter-assay variation was 10.3 and the intra-assay variation was 10.6. These coefficients were calculated on a reference which had an average of 1.0 ng/ml. 87

Progesterone concentration < 1 ng/ml was taken as the limit for a non-functional CL: mares</li>
showing P4 concentration < 1ng/ml were assumed not to have a functional CL at the time of</li>
sampling [25,26].

Statistical analysis - Statistical analysis was performed using GraphPad Prism version 6.00 for Mac
Os X (GraphPad Software, La Jolla California USA, <u>www.graphpad.com</u>).

93 The normality of the distribution of populations studied was analyzed by Shapiro-Wilk test.

Pearson's correlation test was run to assess the relationship between [P4] and CSA, VA and IV,
respectively.

96 Corpora lutea CSA and VA taken when [P4] <1 ng/ml and [P4] ≥1 ng/ml were compared using</li>
97 Unpaired Student's t-tests.

Sensitivity (denoted in %) was defined as the number of mares correctly identified, by trans-rectal ultrasound (CSA, VA, and IV) as having a non-functional CL divided by the total number of mares with [P4] <1 ng/ml. Specificity was the number of mares identified by CSA, VA and IV as having a functional CL, divided by the total number of mares with  $[P4] \ge 1$  ng/ml. Positive Likelihood ratio was defined as the probability to have [P4] < 1 ng/ml if CSA, VA and IV were below the cutoff (sensitivity/1-specificity) [20].

The CLs CSA, VA and IV cutoffs sensitivity and specificity were defined at the highest Positive
Likelihood ratio for the detection of a non-functional CL (P4 <1 ng/mL) and were evaluated using</li>
receiver operating characteristic (ROC) analysis. [20-22].

<mark>1</mark>07

Differences between corpora lutea CSA, VA, IV (CL diameter and vascularization) and [P4]
 production between last cycles (=LC) before anestrus and previous cycles (=PC), on days 6, 10, 14,

**1**10 16 and 18 after mares' ovulations were analyzed using unpaired Student's t-tests.

111 An area under the curve (AUC) has been calculated for each mare CSA, VA and [P4] between day

**1**12 6 and 18 and data from PC and LC were compared using unpaired Student's t-tests.

**1**13 Results were considered as statistically different with P<0.05.

<mark>1</mark>14

### **1**15 **3. Results:**

Seven of the 15 mares went into anestrus during the period of the study: in 5 cases, corpora lutea progressively disappeared, at US examination, around day 18 after ovulation and no follicular growth and ovulation occurred anymore. These cycles were considered as last cycles (LC, N=5) before anestrus. In 2 cases, corpora lutea images persisted for 4 weeks after day 18 before progressively disappearing; no follicular growth and ovulation occurred thereafter. These cycles were considered as having a corpus luteum persistence before anestrus.

122 Eight of 15 mares did not enter into anestrus during the period of the study.

123 Consequently, the distribution of the 20 cycles studied and the data analyzed resulted as follows:

- 5 LC before anestrus that provided data on CSA, VA, IV (N=5) and [P4] levels (N=5)
   analyzed for relationship between CL diameter and vascularization and [P4] production and
   for differences between LC and PC on the same parameters;
- 2 cycles with persistent corpora lutea before anestrus that provided data on CSA, VA, IV
  (N=2) and [P4] levels (N= 2) analyzed for relationship between CL diameter and
  vascularization and [P4] production but not for differences between LC and PC,

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-	130	- 13 PC that prov	vided data on CSA, VA, IV	(N=13) and [P4] levels (	N= 4) analyzed for				
-	<mark>1</mark> 31	relationship bet	ween CL diameter and vas	scularization and [P4] [	production and for				
-	<mark>1</mark> 32	differences betw	een LC and PC on the same pa	arameters.					
-	<mark>1</mark> 33								
-	<mark>1</mark> 34	Consequently, relati	onship between CSA, VA, IV	and [P4] were studied or	n 11 cycles (5 LC, 2				
-	<mark>1</mark> 35	corpus luteum persi	stence and 4 PC) while differ	rences on CSA, VA, IV b	between LC and PC				
-	<mark>1</mark> 36	were studied on 18	cycles (5 LC and 13 PC) and c	lifferences on [P4] levels	between LC and PC				
-	<mark>1</mark> 37	were studied on 9 cy	cles (5 LC and 4 PC).						
-	<mark>1</mark> 38			6					
-	<mark>1</mark> 39	The data analyzed into	this study (CSA, VA, IV, [P4]	) were normally distribute	ed as determined by				
-	<mark>1</mark> 40	Shapiro-Wilk test (P>0.05).							
-	<mark>1</mark> 41	A strong positive correlation (P<0.0001) resulted for plasma [P4] between days 6 and 18 after							
tprint	<mark>1</mark> 42	ovulation and CSA (r=0.527), VA (r=0.695) and IV (r=0.712). CSA, VA and IV statistically							
<u> </u>	<mark>1</mark> 43	explained 28%, 48% an	d 51% of the variability in [P4	], respectively.					
ersio	143 144	The differences between CLs diameter and vascularization taken when [P4] <1 ng/ml and [P4] $\geq$ 1							
$\geq$	<mark>1</mark> 45	ng/ml are described in Table 1.							
-	<mark>1</mark> 46								
-	<mark>1</mark> 47	Table 1: Differences be	tween corpora lutea CSA (pixe	els), VA (pixels) and IV (9	%), when [P4] <				
-	148	48 $1ng/ml \text{ or } \ge 1ng/ml \text{ (N= 11 cycles)}.$							
		[P4] ng/ml (range)	CSA (pixels)	VA (pixels)	IV (%)				
			mean $\pm$ sd (range)	$mean \pm sd \ (range)$	mean $\pm$ sd (range)				
		[P4] <1 ng/ml (0-0.8)	2949.6 ± 1567.1 (1024-6044) <sup>a</sup>	$19.9 \pm 69.9 \ (0-287)^{a}$	0.4 ± 1.3 (0-11) <sup>a</sup>				
		[P4] ≥1 ng/ml (1.2-16.7)	9547.2 ± 4870.2 (3470-25592) <sup>b</sup>	$1139.4 \pm 796.4 (0-2384)^{b}$	$11.7 \pm 8.3 (0-38)^{b}$				

- 149 Within columns  $^{a\neq b}$ : P<0.05
- 150 Area under the ROC curves for CSA, VA and IV were 0.95, 0.93 and 0.93 respectively with a
- 151 P<0.0001. The CSA, VA and IV cutoffs (at the highest Positive Likelihood ratio) of a non-

- **1**52 functional ([P4] <1 ng/mL) corpus luteum, evaluated using a receiver operating characteristic curve
- **1**53 analysis were described in Table 2.
- <mark>1</mark>54
- **1**55 **Table 2:** Highest CSA (pixels), VA (pixels) and IV (%) cutoffs (at the highest Positive Likelihood
- **1**56 ratio) of a non-functional ([P4] <1 ng/mL) CL, evaluated using a receiver operating characteristic
- **1**57 (ROC) curve analysis (N=11 cycles).

Parameter	Value	Sensitivity	Specificity	Highest Positive Likelihood ratio
CSA (pixels)	<3473	70.6%	97.2%	25.4
VA (pixels)	<25.5	88.2%	88.9%	7.9
IV (%)	<5.5	88.9%	88.2%	7.6

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Considering the areas under the curve (AUCs) between diestrus days 6 and 18, PC and LC differed
significantly for CSA (109438 ± 24488 vs 83045 ± 16893 pixels; P<0.05) and VA (18514 ± 8330 vs</li>
8989 ± 4746 pixels; P<0.05), but not [P4] (80.7 ± 43.7 vs 73.7 ± 32.3 ng/ml; P>0.05).

162 The same occurred evaluating data of the single days: there were statistically significant differences

163 for CSA (days 6 and 10) VA (days 6, 10 and 14), and IV (day 14) (P<0.05) but not for [P4] (Table

**1**64 3).

<mark>1</mark>65

**Table 3:** Mean ± sd CSA (pixels), VA (pixels), IV (%) and [P4] (ng/ml) in days 6, 10, 14, 16 and
18 post-ovulation of PC and LC.

<b>F</b>	Cycle	Day 6	Day 10	Day 14	Day 16	Day 18
Factor		$(\text{mean} \pm \text{sd})$	(mean $\pm$ sd)	$(\text{mean} \pm \text{sd})$	$(\text{mean} \pm \text{sd})$	(mean $\pm$ sd)
CSA (pixels)	PC (n=13)	$16713 \pm 6245^{a}$	$12127\pm5817^a$	6277 ± 2690	4151 ± 1862	$2819\pm2082$
	LC	$10595 \pm 2514^{b}$	$8712\pm636^b$	$5935 \pm 3700$	$3382\pm2004$	2981 ± 1230

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	(n=5)					
VA (pixels)	PC (n=13)	$2419\pm880^a$	$2325\pm976^a$	$1312\pm968^a$	$365\pm650$	103 ± 168
	LC (n=5)	$1596 \pm 668^{b}$	$1144 \pm 817^{b}$	$544 \pm 464^{b}$	64 ± 120	0 ± 0
IV (%)	PC (n=13)	$17.6 \pm 8.4$	22.0 ± 10.0	16.6 ± 10.9 <sup>a</sup>	6.1 ± 10.6	2.6 ± 4.4
	LC (n=5)	16.9 ± 10.1	12.8 ± 8.6	$6.9 \pm 4.8^{b}$	1.2 ± 2.3	0 ± 0
<b>P4</b> (ng/ml)	PC (n=4)	8.3 ± 5.0	9.8 ± 4.7	5.7 ± 3.2	2.3 ± 2.7	0.1 ± 0.2
Within supersor	LC (n=5)	9.1 ± 3.8	8.7 ± 2.9	4.0 ± 5.0	0.2 ± 0.4	0.0 ± 0.0

Within superscript <sup> $a\neq b$ </sup>: P<0.05

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170 **Discussion and Conclusions:** It has been shown how luteal functionality, in terms of [P4] 171 production, slowly decreases from summer cycles to the last cycle before anestrus [1,3,8,9]. 172 A correlation between [P4] and CL vascularization was already found in mares [12,16] cattle [17,22] and woman [27,28]. In our study, although limited by the number of cycles **1**73 174 analyzed, CSA, VA and especially IV of mares CLs had a strong correlation with [P4]. 175 Crabtree and Wilsher also found a strong correlation of [P4] with CSA and VA but not with 176 IV in pregnant mares [18]. IV is a parameter that changes when CSA and VA growth is 177 divergent, and in that study, in the physiologic pregnant CLs, probably their growth was **1**78 parallel making IV a constant. The areas under the ROC curves higher than 0.9 and their 179 high significance (P<0.0001) for CSA, VA and IV showed that these parameters were able

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180 to predict when [P4] was lower than 1 ng/ml. These results are consistent with the bovine **1**81 species for CL dimensions and vascularization [21,22]. Corpora lutea producing **1**82 progesterone, in the present study, had 3-4 folds larger CSAs and 10 folds larger VAs and 183 IVs than not functional CLs ([P4] <1 ng/ml; P<0.05). Areas under the curves of CSA and **1**84 VA showed to be significantly higher in previous cycles than during last cycles of the year, 185 but [P4] did not, even though AUC for this hormone was numerically lower. CLs ultrasonographic differences between PC and LC were shown clearly by the AUCs, but also 186 evident between single examination days, up to the 14<sup>th</sup> day after ovulation. This trend was 187 **1**88 not found in [P4], may be also due to the smaller number of cycles evaluated. In this study 2/7 (28.6%) of the mares' last cycles developed in a persistent CL compared to 0/13 (0%) of 189 190 the previous cycle. This is consistent with the 25% incidence of persistence of CL reported **1**91 previously for the last cycle before vernal anestrus versus the 10% of normal cycles of the **1**92 season [11]. In several livestock species has been reported that low level or absence of 193 plasma [P4] resulted in compromised PGF2 $\alpha$  response [11,29-31]. Although we were not **1**94 able to show significant differences, previous cycles had a numerically higher [P4] on each **1**95 sampling day, and this might have been the cause for such persistent CLs. In conclusion, 196 CLs ultrasonographic characteristics seem to reflect their functionality, confirming previous **1**97 studies performed in different species [14,16-19,21,22], and, on the group of animals **1**98 studied, CLs of the last cycles of the year were smaller and less vascularized than those of 199 the previous cycles.

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- Highlights
- Mares corpora lutea functionality was analysed by plasma progesterone concentration and Color Doppler ultrasound
- Last cycle corpora lutea functionality was compared with previous mares' cycles
- A positive correlation was found between plasma progesterone concentration and corpora lutea cross-sectional area and vascularization.
- ROC curve identified B-Mode and Color Doppler non-functional corpora lutea ultrasound characteristics.
- Color Doppler evidenced significant differences between last cycle and normal cycle's corpora lutea but not between the plasma progesterone concentrations, respectively.