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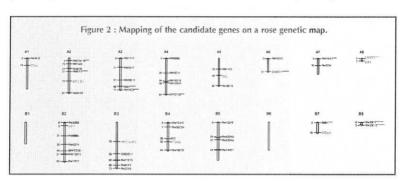
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# A gene candidate approach to study the control of recurrent blooming in rose.

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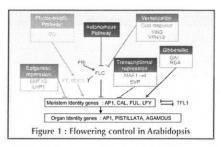
In rose, flowering can occur once a year (non-recurrent blooming) or continuously during the favourable season (recurrent blooming). Recurrent blooming has been shown to be under the control of a major recessive gene, *RB*, which was localized on genetic map (Crespel et al, 2002, fig. 2). In order to understand the molecular basis of recurrent blooming, we have undertaken a gene-candidate approach. Genetic and physiological data suggest that gibberelic acid or floral repressor may be involved in this control. Genes implicated in these pathways have been well characterized in Arabidopsis (fig. 1).



■ To look for co-localization with the *RB* gene, we mapped the genes on a genetic map developed in the laboratory with AFLP and SSR markers (fig. 2). A *RGA* homologue, *RGAa*, was mapped in the vicinity of *RB*. To better understand the involvement of these genes in the floral process, we have followed their expression by RT-PCR during the floral process and in different tissues (fig. 3). The genes showed different patterns of expression and almost all the genes are expressed during the floral initiation (between stage lb and II), suggesting that they may be implicated in this process.

Table 1 : List of the flowering genes isolated in rose.

	R	IN	N	YL	L	A	FB	P	I	Ib	11	Ш		
RGAa														
RGAb														
LHP1				nd										
EMF2		nd	nd						nd					
CEN														
AP1a														No expression
AP1b				Ī										Low expression High expression
AP3 B3													nd	
AP3 euB3													R: re IN:	internode,
PI (BP)													YL:	iode, youg leaf, xpanded leaf,
AG													A: v	youg leaf, xpanded leaf, egetative apex; flower bud,
E1F													P: p	etal. e I, Ib, II and III defined in figure 4



Cloning and characterization of flowering genes in rose

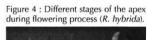
■ Using sequences available in the databases or degenerate primers, we were able to isolate genes homologue to genes involved in GA perception (*RGA/GAI*), floral repression (*LHP1*, *EMF2* and *TFL1*), floral control (*AP1*, *SOC1* and *LFY*) and flower development (*AP1*, *PI* and *AG*) (tab.1).

	Genes		Source	% similarity	Molecular marker	Genetic position
GA perception	RGA	a	EST	72	Acryl	AT
	RUA	b	Primer	84	Acryl	85
Floral repressors	TFL2/LH	P1	Primer	56	no	-
	EMF2		Primer	56	no	
	CEN/TF	L1	Primer	77	Acryl	A1
Floral Centrol	SOC1		Primer	73	no	
	LEAFY		Primer	90	no	-
	ADETALAS	а	Primer	81	Acryl	B2/A8
	APETALA1	b	Primer	75	no	
Flower development	4 DET 4 1 4 2	B3	cDNA	67	Acryl	A2
	APETALA3	euB3	cDNA	70	Acryl	В3
	PISTILLA	ATA	cDNA	68	no	-
	AGAMO	US	cDNA	81	no	

## Construction of a cDNA libraries and EST production Figure 4: Different

Two cDNA libraries have been constructed from:

(1) vegetative apices (Rosa wichuraiana)
(2) apices during the floral transition (R. hybrida Black Baccara \*, stage I, Ib and II).





Floral initiation

Histological analysis revealed that the floral initiation takes place between stage Ib and II (data not shown). 5000 EST have been produced from the two libraries and are presently analysed.

### Conclusions and perspectives

We were able to isolate in a perennial plant (rose) homologues of genes involved in floral control in an annual one (Arabidopsis). Expression analysis seems indicate that these genes are implicated in the floral process. Production of EST may allow us to develop differential approach to isolate new genes in rose involved in the control of flowering.

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