



**HAL**  
open science

## Seasonal effects on plasma cortisol concentrations in the Bedouin buck: circadian studies and response to ACTH

N. Chergui, Pierre Mormède, Aline Foury, F. Khammar, Z. Amirat

### ► To cite this version:

N. Chergui, Pierre Mormède, Aline Foury, F. Khammar, Z. Amirat. Seasonal effects on plasma cortisol concentrations in the Bedouin buck: circadian studies and response to ACTH. *Animal*, 2017, 11 (3), pp.445-451. 10.1017/S1751731116001671 . hal-01602754

**HAL Id: hal-01602754**

**<https://hal.science/hal-01602754v1>**

Submitted on 11 Nov 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - ShareAlike 4.0 International License

# Seasonal effects on plasma cortisol concentrations in the Bedouin buck: circadian studies and response to ACTH

N. Chergui<sup>1†</sup>, P. Mormede<sup>2a</sup>, A. Foury<sup>2</sup>, F. Khammar<sup>1</sup> and Z. Amirat<sup>1</sup>

<sup>1</sup>Laboratoire de Recherches sur les Zones Arides (LRZA), Faculty of Biology, Houari Boumediene University of Sciences and Technology (USTHB), PB 39, El Alia, Algiers, 16110, Algeria; <sup>2</sup>Nutrition et Neurobiologie Intégrée (NutriNeuro), Institut National de la Recherche Agronomique (INRA), F-33076 Bordeaux, France

(Received 18 December 2015; Accepted 11 July 2016; First published online 11 August 2016)

*Our work aims at the exploration of cortisol secretion in the Bedouin goat, native to the Algerian Sahara desert, to understand the mechanisms of adaptation to extreme hot climates. In the present study, diurnal and seasonal variations of cortisol concentrations were measured in basal conditions, as well as the response to ACTH stimulation tests across seasons in bucks. The plasma concentrations of cortisol showed no diurnal cycle but a large variation across seasons. The highest levels occurred in summer and winter when the environmental conditions are at their extreme levels. The rectal temperature showed nycthemeral and seasonal variations, and BW was also different across seasons with highest values in summer and lowest in winter. The results obtained after administration of two doses (2 or 10 µg/kg BW) of synthetic ACTH to three different age groups (kids, adults and elderly animals) showed a strong increase in plasma cortisol concentrations under all conditions with maximum levels achieved between 15 and 120 min. The analysis of the area under the cortisol curve showed no significant difference between the responses to the two doses of ACTH and between age groups, but showed seasonal variations with the lowest response in autumn than in other seasons. We conclude that season significantly affects secretion of cortisol in both basal state and under ACTH stimulation. However, the variation of adrenal reactivity to ACTH is not sufficient to explain seasonal differences, and in particular the summer peak in basal circulating cortisol concentrations. Further research should focus on the respective contribution of environmental factors (such as day length, temperature, humidity) and the mechanisms involved in cortisol regulation.*

**Keywords:** Bedouin goat, cortisol, ACTH stimulation test, diurnal cycle, season

## Implications

Understanding the biological mechanisms of adaptation to hot environments is a challenge to select the most efficient animals in these conditions. We studied these processes in the Bedouin goat, usually considered as adapted to the desert climate. Our results show a dissociation between circulating cortisol levels and the adrenal reactivity to ACTH. Further research should focus on the respective contribution of environmental factors (such as day length, temperature, humidity) and the mechanisms involved in cortisol regulation. The Bedouin goat in its natural desert environment appears as good model for these studies.

## Introduction

In order to improve animal production in harsh environments, it is important to understand adaptation mechanisms

to extreme environmental conditions (temperature, food availability). Goats are considered to be well adapted to hot and dry climates (Silanikove, 2000; Aboul-Naga *et al.*, 2014), but the mechanisms of this adaptation, beyond specific digestive characteristics, are not well understood (Silanikove *et al.*, 1993; Choshniak *et al.*, 1995; Shamay *et al.*, 2000; Maltz *et al.*, 2009). The hypothalamic–pituitary–adrenal (HPA) axis is a critical neuroendocrine system involved in metabolism and adaptation, and circulating glucocorticoid hormone (GC) concentrations or the reactivity of the adrenal cortex to ACTH are often used as stress indicators (Mormede *et al.*, 2007).

In most animal species, a circadian timing system has evolved as a strategy to cope with 24-h rhythms in the environment (Dickmeis *et al.*, 2013). The circadian rhythm of cortisol is paralleled by other physiological parameters such as body temperature and melatonin, demonstrating the power of the circadian pacemaker in the suprachiasmatic nucleus to regulate the timing of physiological rhythms (Pevet and Challet, 2011; Dickmeis *et al.*, 2013). Seasonal changes have also been described (Romero, 2002;

<sup>a</sup> Present address: Génétique, Physiologie et Systèmes d'Élevage (GenPhySE), INRA, F-31326 Castanet-Tolosan, France.

<sup>†</sup> E-mail: nadiachergui@yahoo.fr

Dickmeis *et al.*, 2013) but the amplitude and timing of the cycle largely differ across studies, possibly reflecting the variety of environmental conditions of these studies (e.g. Rhind and McMillen, 1995; Alila-Johansson *et al.*, 2003; Meza-Herrera *et al.*, 2007; Al-Busaidi *et al.*, 2008; Al-Samavi *et al.*, 2014 in goats). A characteristic feature of the desert climate is the large amplitude of variation in temperature and hygrometry, during the nycthemeron and across seasons. As the HPA axis is extremely sensitive to these environmental conditions (e.g. Marple *et al.*, 1972), its activity may be a good index of the intensity of adaptive adjustments. Indeed, we previously described large seasonal changes in HPA axis activity in a wild desert species, the sand rat (Amirat *et al.*, 1980).

In order to evaluate the intensity of adaptation processes in the Bedouin buck exposed to extreme environmental conditions around the year in the Sahara desert, we investigated the seasonal changes in circadian plasma cortisol concentrations, rectal temperature, BW and in the reactivity of the adrenal cortex to exogenous ACTH.

## Material and methods

### Animals

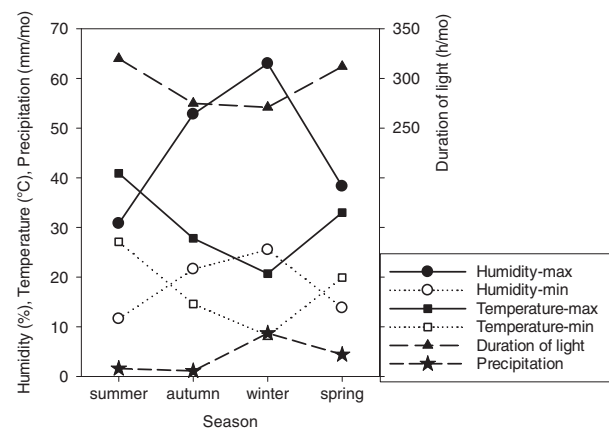
The experimental animals were male goats from the Bedouin breed. They exhibited similar phenotypic characteristics, and were clinically healthy. They were raised in the Béni Abbès research station (30°07' N, 2°10' W, Bechar, South Algeria) located in the Algerian Sahara desert. The lifespan of this domestic species is about 10 years. The males were kept separated from females, under open shelter undergoing the influence of natural variations of climatic factors. Animals were fed forage and a mixture of oats and barley (about 1 kg) once daily (1630 h); water was provided *ad libitum*.

The experiments were conducted over 2 consecutive calendar years under the largely differing environmental conditions, which occur in the Sahara desert during summer (July), autumn (October), winter (February and March) and spring (May) (Figure 1). Each animal was studied only once per season; 1 h before the start of experiments, animals were weighed and housed individually in order to reduce the stress induced by handling. The meteorological data were obtained from the Algerian meteorological center.

Blood samples (3 ml) were collected from the jugular vein by trained personnel using heparinized Vacutainer® tubes (lithium heparinate, Venoject Terumo; Leuven, Belgium) and kept on ice until centrifugation at  $3000 \times g$  for 10 min at 4°C. Plasma samples were stored at -25°C until assayed.

The rectal temperature of each buck was measured at 0800 and 2000 h using a digital clinical thermometer (Eco Temp Smart, MC-341-E; Omron Healthcare, Hoofddorp, Netherlands) inserted about 3 cm into the rectum in contact with the mucosal wall of the rectum. The value was taken after the thermometer gave an alarm sound (60 to 90 s), indicating that the reading had stabilized.

All experimental protocols were accepted by the ethical committee and the project commission of the institution.



**Figure 1** Climatic data during the periods of experimentation. Means of the monthly measures obtained from the Béni Abbès meteorological station (30.13°N, 2.17°O, elevation: 500 m) for the 2 experimental years.

### Basal adrenocortical activity

For the study of basal adrenocortical activity, we used seven adult males (2 to 3 years,  $31.9 \pm 1.9$  kg BW). A few animals were culled and eventually replaced, so that six were available in spring, summer and autumn, and three in winter. Blood samples were taken every 2 h for 22 h, from 1000 h on the 1<sup>st</sup> day to 0800 h on the next day in order to catch the circadian changes.

### Adrenal response to ACTH

For the study of the reactivity to ACTH, we used seven young (<18 months,  $19.5 \pm 0.8$  kg BW), three adult (2 to 2.5 years,  $24.7 \pm 2.1$  kg BW) and three old males (5 to 8 years,  $40.0 \pm 1.6$  kg BW). Due to the shift of several animals from the 'young' to the 'adult' group, and to the culling of several animals, the number of trials available is variable and mentioned in the caption of figures. In order to reduce the possible effect of the circadian rhythm on adrenal reactivity to ACTH (Reinberg *et al.*, 1983), all stimulations were performed between 0800 and 1200 h. At each season, the animals received two doses (2 then 10  $\mu\text{g}/\text{kg}$  BW, 72 h apart) of synthetic ACTH i.v. in the jugular vein (Cortrosyn; Organon, Oss, Netherlands; 1 mg equivalent to 100 IU), diluted in 1 ml of apyrogenic physiological serum. Blood samples were collected in refrigerated tubes at -15, 0, 3, 8, 15, 30, 45, 60, 90 and 120 min, and then at 24 and 48 h after each ACTH injection. The goal was to obtain a maximal response of the adrenal glands, a phenotype that we have shown to be highly heritable in pigs (Larzul *et al.*, 2015).

### Cortisol assay

In order to define the main GC hormone secreted by the adrenal cortex of the Bedouin goat, the relative proportions of cortisol and corticosterone were measured in 17 samples, four before and 13 after ACTH injection. Plasma (100  $\mu\text{l}$ ) was extracted with 3 ml cyclohexane/ethyl acetate (50/50) at a recovery rate of  $71.0 \pm 1.5\%$ . Hormones were separated by paper chromatography (Bush B5 solvent system, no. 2

Whatman paper), as previously described (Amirat *et al.*, 1980). Next, hormones were measured by radioimmunoassay (RIA) using specific antibodies raised in rabbit, kindly provided by Prof. Claustrat (Lyon, France) and by Prof. Oliver (Marseille, France). Plasma cortisol was then routinely measured by RIA after ethanol extraction without previous separation with Prof. Oliver's antibody, as described in the study by Désautés *et al.* (1997). Intra and inter-assay CV were 6.7% and 7.4%, respectively, for corticosterone, and 13.6% and 7.8%, respectively, for cortisol, with a detection limit of 0.1 ng/ml.

#### Statistical analysis

Data are presented as means  $\pm$  standard error of the mean. The areas under the curve (AUC) after ACTH injection were calculated with respect to ground by the trapezoidal approach (Pruessner *et al.*, 2003). The area was limited to the 0 to 90 min period after ACTH injection because samples taken at 120 min were lost for one experimental group. Cortisol concentrations or AUC after ACTH injection were analyzed using linear mixed effects models (package 'nlme' of R) with the animal as a random factor to account for repeated measures and time of the day and season (basal cortisol concentrations), or ACTH dosage, season and age (AUC) as fixed factors. Statistical significance was set at  $P < 0.05$ . Diurnal variations were also analyzed by cosinor with the R package 'season' (Tong, 1976).

## Results

In the Bedouin buck, basal plasma cortisol concentration ranged between 1 and 18 ng/ml in the analyzed samples, whereas plasma corticosterone concentration was only about 0.1 ng/ml, with a 60:1 ratio. This large difference was also found after ACTH injection (1:30 ratio). Therefore, immunoreactive cortisol was measured in further experiments.

#### Seasonal variation in the base levels of plasma cortisol concentrations

The analysis of basal cortisol concentrations showed a tendency only for the effect of time of the day ( $P = 0.054$ ) and a highly significant influence of season ( $P < 0.0001$ ) with a season  $\times$  time interaction ( $P = 0.011$ ) (Figure 2). Mean cortisol concentrations were lowest in autumn and spring and highest in summer and winter ( $P = 0.0003$  and  $0.0001$  v. autumn, respectively). Subsequent analysis per season showed that the time of the day was significant in summer ( $P = 0.03$ ), autumn ( $P = 0.04$ ) and winter ( $P = 0.01$ ), but not in spring ( $P = 0.17$ ). However, these variations did not fit the classical sigmoidal curve as analyzed by the cosinor algorithm (all  $P > 0.05$ ).

#### Seasonal variation of rectal temperature and BW

The rectal temperature showed circadian fluctuations, with higher values at 2000 than 0800 h ( $P = 0.01$ ) and no significant time  $\times$  season interaction (Figure 3). It was also

different across seasons ( $P < 0.0001$ ), following the changes of external temperature and day length, with highest values occurring in summer and spring and lowest in autumn and winter. BW varied significantly across seasons ( $P = 0.0001$ ), the mean values being highest in summer and lowest in winter.

#### Seasonal variation of adrenal response to ACTH

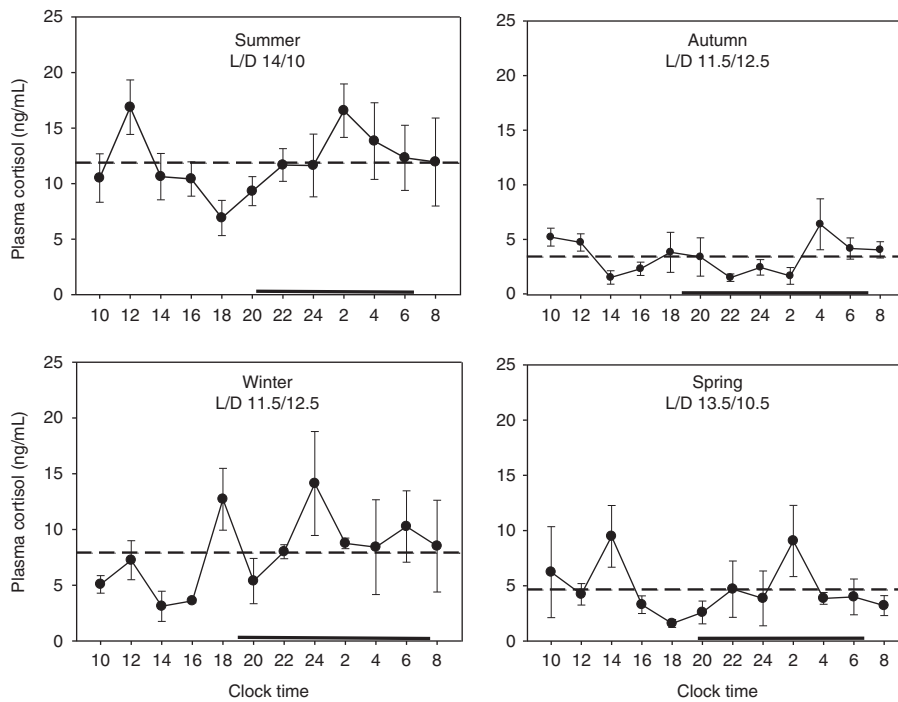
The responses of plasma cortisol concentrations to 2 and 10  $\mu\text{g}/\text{kg}$  BW of ACTH are shown in Figure 4. Both doses of ACTH induced a strong increase in cortisol concentrations in all conditions with maximum levels achieved between 15 and 60 min (2  $\mu\text{g}/\text{kg}$ ) or 90 min (10  $\mu\text{g}/\text{kg}$ ). The response to ACTH (AUC) did not differ between ACTH doses ( $P = 0.12$ ) but the effect of season was highly significant ( $P < 0.0001$ ), the AUC being lowest in autumn and summer and highest in spring (autumn  $P < 0.05$ , winter  $P < 0.01$ , spring  $P < 0.001$  v. summer). The global influence of age did not reach significance ( $P = 0.11$ ), although the youngest animals had a lower AUC as compared with adults ( $P < 0.05$ ).

## Discussion

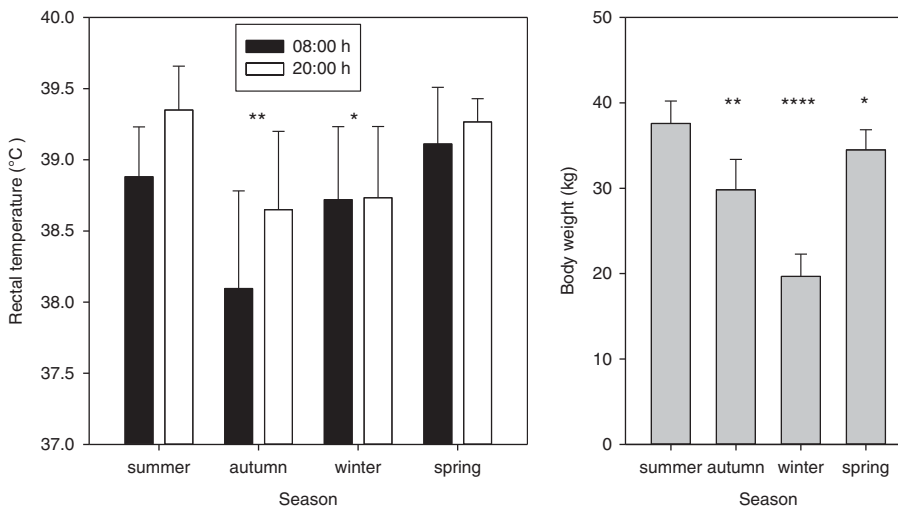
These data show that the Bedouin goat produces both cortisol and corticosterone as adrenal GC hormones, with a clear predominance of cortisol. Basal plasma cortisol concentrations measured here are in the range of previously published values obtained under similar conditions (Rhind and McMillen, 1995; Alila-Johansson *et al.*, 2003; Meza-Herrera *et al.*, 2007). The adrenal steroid hormones play a crucial role in physiological adjustments to various types of environmental and stress factors (Mormede *et al.*, 2007).

The daily rhythm of HPA axis activity is well described in most mammalian species with a peak at the beginning of the behaviorally active period, morning in diurnal species. In ruminants, it was described in sheep (Fulkerson and Tang, 1979; Mesbah and Brudieux, 1982) but it is not very pronounced in cattle and some studies failed to show any diurnal rhythm (see Mormede *et al.*, 2007 for discussion). In the present study with goats, there was no clear-cut diurnal cycle, as previously shown by others (Kokkonen *et al.*, 2001; Alila-Johansson *et al.*, 2003). Although the diurnal cycle is principally driven by light, nutritional factors are also involved (see Blum *et al.*, 2012 for review). The specific feature of a dampened cycle of nutrients absorption in ruminants may be operative in this reduced diurnal cycle of HPA axis activity, although to the best of our knowledge, this hypothesis has never been examined experimentally.

Circulating concentrations of cortisol were primarily influenced by season, with 3.5-fold higher mean peak levels in summer than at the trough in autumn. Variable effects of season on HPA axis activity have been published, both for the range of change and the circannual shape of the concentration curve (Rhind and McMillen, 1995; Alila-Johansson *et al.*, 2003; Meza-Herrera *et al.*, 2007; Al-Busaidi *et al.*, 2008; Al-Samawi *et al.*, 2014 in goats). Part of these



**Figure 2** 22-h Plasma cortisol concentration profiles in adult bucks measured in summer, autumn, winter and spring (means  $\pm$  SEM,  $n = 6$  except in winter  $n = 3$ ). Dashed lines show the mean value. Black bars on the x-axis denote hours of darkness. LD = light/dark cycle.



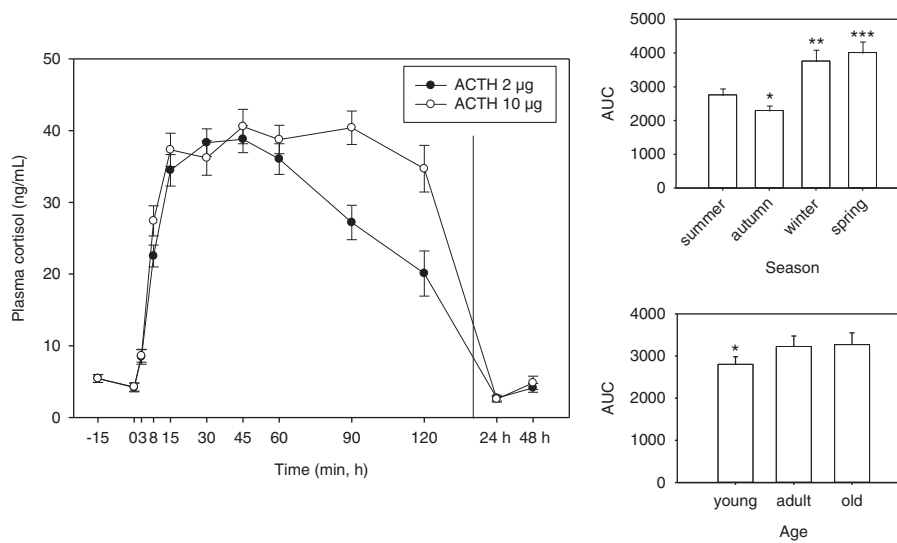
**Figure 3** Left panel: seasonal variations of rectal temperature measured at 0800 and 2000 h in adult bucks; right panel: seasonal variations of BW (means  $\pm$  SEM,  $n = 6$  except in winter  $n = 3$ ). \*  $P < 0.05$ , \*\*  $P < 0.01$  and \*\*\*\*  $P < 0.0001$  v. summer.

variations across studies most probably arises from the large differences in the local experimental conditions (such as the duration and intensity of light, temperature and humidity). It raises the question of the nature of environmental factors driving the changes in HPA axis activity.

A first candidate is day length and the pineal hormone melatonin secreted during the night and in larger amounts in winter (see Malpaux *et al.*, 2001 for review and Alila-Johansson *et al.*, 2006; Todini *et al.*, 2011; Carcangiu *et al.*, 2015 in goats). Melatonin takes part in the regulation of circadian rhythms (Pevet and Challet, 2011) and exerts direct inhibitory actions on adrenal responses to ACTH in sheep (Torres-Farfan *et al.*, 2008). In goats, melatonin was shown

to inhibit heat stress responses, including plasma cortisol increase and expression of heat-shock proteins (HSP) in peripheral blood mononuclear cells (PBMC) (Sharma *et al.*, 2013), as well as the cortisol response to weaning stress (Redondo *et al.*, 2010). More studies will be necessary to know the range of fluctuation of melatonin levels in our experimental conditions and explore its possible influence on HPA axis activity.

Temperature and humidity are well-documented environmental factors influencing HPA axis activity, although their influence is rather complex. As shown in cows (Johnson and Vanjonack, 1976; Lee *et al.*, 1976) and in pigs (Marple *et al.*, 1972; Campos *et al.*, 2014) adaptation to heat reduces



**Figure 4** Left panel: kinetics of the plasma cortisol response to two ACTH doses (2 and 10 µg/kg, means ± SEM,  $n = 37$ ). Right panels: integrated responses to ACTH (areas under the curve (AUC)) from 0 to 90 min as a function of season (upper right, means ± SEM,  $n = 30, 16, 14, 14$  in summer, autumn, winter and spring, respectively) and age (lower right, means ± SEM,  $n = 26, 24$  and  $24$  in young, adults and old bucks, respectively). Each bar represents the grand mean of all tests available for the specific group. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  v. summer (season panel) and \* $P < 0.05$  v. adult (age panel). The vertical line in the left panel indicates the rupture of the time scale between 120 min and 24 h.

plasma cortisol levels, but acute exposure to high temperatures (in the range of 40°C as observed during summer in the present experiments) increases plasma cortisol levels and triggers other stress responses such as the expression of HSP in PBMC (Dangi *et al.*, 2012; Sharma *et al.*, 2013). These non-linear effects of increased temperature on HPA axis activity can explain apparently divergent experimental results of the effects of seasons, depending on the range of temperatures encountered at each experimental location. High ambient temperatures and high solar radiation may be responsible for the highest cortisol levels observed during summer in our experiments, similar to findings of Al-Samawi *et al.* (2014) in Saudi Arabia (38.5°C), as compared with the low levels measured in summer by Al-Busaidi *et al.* (2008) in Oman (34°C) and by Meza-Herrera *et al.* (2007) in Northern Mexico (26.1°C). Indeed, the increase in rectal temperature during the hot season indicates that animals were subjected to heat stress (Minka and Ayo, 2012; Al-Samawi *et al.*, 2014).

The seasonal rhythm in BW coincides with reproductive activity; indeed, the gain was observed when bucks were reproductively inactive, between March and mid-August, and the loss during the breeding season, which occurs between late summer and late autumn (Amirat *et al.*, 2001; Kumar *et al.*, 2014). Furthermore, energy mobilization is necessary in winter to maintain body temperature, and could be partly responsible for the decrease in BW observed in this period. This same seasonal rhythm in BW has been reported by other authors, which commonly correlates with the influence of the season (photoperiod) on physiological state and reproductive performance of bucks, independently of the feeding level (Zarazaga *et al.*, 2009). For example, Barenton *et al.* (1988) demonstrated that variation in photoperiod has a direct effect on BW: long days stimulate and short days inhibit weight gain. The HPA axis and other neuroendocrine systems

may be operative in these effects in rams (Cahill *et al.*, 2013; Ebling, 2015).

Different timings of the seasonal rhythm in HPA axis activity can be shown at different levels of the axis. In rams, Ssewanyana *et al.* (1990) showed the highest levels of circulating hormones such as ACTH and cortisol from May to July, and  $\beta$ -endorphin in August. The greatest pituitary response (ACTH,  $\beta$ -endorphin) to arginine vasopressin (AVP) and/or corticotropin-releasing hormone (CRH) was obtained in summer and autumn and the maximal adrenal response (cortisol) to dexamethasone/ACTH in spring at the time of the peak in cortisol secretion. Another study reported no significant seasonal changes in plasma cortisol concentrations but a seasonal cycle in the levels of  $\beta$ -endorphin occurring in parallel with the cycle in plasma FSH and BW (Ebling and Lincoln, 1987).

In the present experiments, we investigated the adrenal response to ACTH as the major source of variability in cortisol levels (Mormede and Terenina, 2012), including the effect of season (Reinberg *et al.*, 1983). No significant effect of age or dose was found, the adrenal being already maximally stimulated at the lowest dose (2 µg/kg). Although a marked effect of season was observed, it did not match the variation in plasma cortisol levels, as the largest response to ACTH was measured in spring, when cortisol levels were low. However, the smallest response to ACTH was measured in autumn when cortisol levels were also at their lowest. Therefore, the adrenal reactivity to ACTH may contribute to the seasonal cycle of cortisol but other levels of the HPA axis could be predominant in the transduction of the rhythm. Arginine vasopressin is a potent secretagogue of ACTH, particularly in goats where it is more potent than CRH (Katoh *et al.*, 2005). This peptide, together with the adrenal cortex hormones, plays an important role in water homeostasis, a critical

function in desert animals exposed to high temperatures and dehydration (El-Husseini and Haggag, 1974; Haggag and El-Husseini, 1974; Yagil and Etzion, 1979), but its possible role in seasonal cortisol secretion rhythms has not been investigated. More studies are necessary to identify the contribution of the various season-related environmental factors and to monitor simultaneously the different levels of the HPA axis potentially involved in their transduction.

## Conclusion

The present data show that the Bedouin buck produces both cortisol and corticosterone as adrenal GC hormones, with a clear predominance of cortisol, both at basal conditions and after ACTH stimulation. No clear circadian variation could be seen in circulating cortisol concentrations but large variations across seasons were found with higher levels in summer and winter. However, the adrenal response to ACTH does not follow this pattern, the response being the highest in spring and the lowest in autumn, both seasons with low basal plasma cortisol concentrations; it was low in summer when plasma cortisol concentrations are the highest. The rectal temperature also shows nycthemeral and seasonal variations, with higher values at 2000 than 0800 h in all seasons, particularly in summer and spring. The BW was also different across seasons, with higher values in summer (period of energy restoration) and it was lowest in winter (period of energy deficit). Further experiments will allow to explore the nature of the most important environmental factors influencing HPA axis activity and their mechanisms of action. This will help to better understand the physiological basis of adaptation of goats to harsh environments.

## Acknowledgments

The authors thank all the staff of the experimental station of Beni-Abbes (W. Bechar). Special thanks to Zahreddine Djazouli (University Saad Dahlab, W. Blida) and Yann Labrune (INRA, Castanet-Tolosan) for help with statistical analyses, and Nouria Boukenaoui of the Research Laboratory LRZA/USTHB for technical support with cortisol immunoassays. The authors dedicate this work to the memory of their colleague Yahia Lakhdari, LRZA/USTHB. Special thanks to Prof. Byron C. Jones (UTHSC Memphis, TN, USA) for editing the manuscript.

## References

Aboul-Naga A, Osman MA, Alary V, Hassan F, Daoud I and Tourrand JF 2014. Raising goats as adaptation process to long drought incidence at the Coastal Zone of Western Desert in Egypt. *Small Ruminant Research* 121, 106–110.

Al-Busaidi R, Johnson EH and Mahgoub O 2008. Seasonal variations of phagocytic response, immunoglobulin G (IgG) and plasma cortisol levels in Dhofari goats. *Small Ruminant Research* 79, 118–123.

Alila-Johansson A, Eriksson L, Soveri T and Laakso M-L 2003. Serum cortisol levels in goats exhibit seasonal but not daily rhythmicity. *Chronobiology International* 20, 65–79.

Alila-Johansson A, Eriksson L, Soveri T and Laakso M-L 2006. The daily rhythms of melatonin and free fatty acids in goats under varying photoperiods and constant darkness. *Chronobiology International* 23, 565–581.

Al-Samawi KA, Al-Hassan MJ and Swelum AA 2014. Thermoregulation of female Aardi goats exposed to environmental heat stress in Saudi Arabia. *Indian Journal of Animal Research* 48, 344–349.

Amirat Z, Khammar F and Brudieux R 1980. Seasonal changes in plasma and adrenal concentrations of cortisol, corticosterone, aldosterone, and electrolytes in the adult male sand rat (*Psammomys obesus*). *General and Comparative Endocrinology* 40, 36–43.

Amirat Z, Lakdari Y and Khammar F 2001. Cycle saisonnier de l'activité sexuelle chez le bouc de race Bédouine (Seasonal cycle of sexual activity in male Bedouin goat). *Rencontres Autour des Recherches sur les Ruminants* 8, 373.

Barenton B, Ravault J-P, Chabanet C, Daveau A, Pelletier J and Ortavant R 1988. Photoperiodic control of growth hormone secretion and body weight in rams. *Domestic Animal Endocrinology* 5, 247–255.

Blum ID, Lamont EW and Abizaid A 2012. Competing clocks: metabolic status moderates signals from the master circadian pacemaker. *Neuroscience and Biobehavioral Reviews* 36, 254–270.

Cahill S, Tuplin E and Holahan MR 2013. Circannual changes in stress and feeding hormones and their effect on food-seeking behaviors. *Frontiers in Neuroscience* 7, 140.

Campos PHRF, Noblet J, Jaguelin-Peyraud Y, Gilbert H, Mormede P, Donzele RFM de O, Donzele JL and Renaudeau D 2014. Thermoregulatory responses during thermal acclimation in pigs divergently selected for residual feed intake. *International Journal of Biometeorology* 58, 1545–1557.

Carcangiu V, Giannetto C, Luridiana S, Fazio F, Mura MC, Parmeggiani A and Piccione G 2015. Seasons induce changes in the daily rhythm of plasma melatonin in goats (*Capra hircus*). *Animal Biology* 65, 13–20.

Choshniak I, Ben-Kohav N, Taylor CR, Robertshaw D, Barnes RJ, Dobson A, Belkin V and Shkolnik A 1995. Metabolic adaptations for desert survival in the Bedouin goat. *American Journal of Physiology* 268, R1101–R1110.

Dangi SS, Gupta M, Maurya D, Yadav VP, Panda RP, Singh G, Mohan NH, Nital H, Bhure SK, Das BC, Bag S, Mahapatra R, Taru Sharma G and Sarkar M 2012. Expression profile of HSP genes during different seasons in goats (*Capra hircus*). *Tropical Animal Health and Production* 44, 1905–1912.

Desautels C, Bidanel JP and Mormede P 1997. Genetic study of behavioral and pituitary-adrenocortical reactivity in response to an environmental challenge in pigs. *Physiology and Behavior* 62, 337–345.

Dickmeis T, Weger BD and Weger M 2013. The circadian clock and glucocorticoids – interactions across many time scales. *Molecular and Cellular Endocrinology* 380, 2–15.

Ebling FJP 2015. Hypothalamic control of seasonal changes in food intake and body weight. *Frontiers in Neuroendocrinology* 37, 97–107.

Ebling FJP and Lincoln GA 1987. Beta-endorphin secretion in rams related to season and photoperiod. *Endocrinology* 120, 809–818.

El-Husseini M and Haggag G 1974. Antidiuretic hormone and water conservation in desert rodents. *Comparative Biochemistry and Physiology Part A: Physiology* 47, 347–350.

Fulkerson WJ and Tang BY 1979. Ultradian and circadian rhythms in the plasma concentration of cortisol in sheep. *Journal of Endocrinology* 81, 135–141.

Haggag G and El-Husseini M 1974. The adrenal cortex and water conservation in desert rodents. *Comparative Biochemistry and Physiology Part A: Physiology* 47, 351–359.

Johnson HD and Vanjonack WJ 1976. Effects of environmental and other stressors on blood hormone patterns in lactating animals. *Journal of Dairy Science* 59, 1603–1617.

Katoh K, Yoshida M, Kobayashi Y, Onodera M, Kogusa K and Obara Y 2005. Responses induced by arginine-vasopressin injection in the plasma concentrations of adrenocorticotrophic hormone, cortisol, growth hormone and metabolites around weaning time in goats. *Journal of Endocrinology* 187, 249–256.

Kokkonen U, Riskila P, Roihankorpi M and Soveri T 2001. Circadian variation of plasma atrial natriuretic peptide, cortisol and fluid balance in the goat. *Acta Physiologica Scandinavica* 171, 1–8.

Kumar R, Pramod RK, Kumar R, Negi M, Singh SP, Singh R and Mitra A 2014. Testicular biometry and seasonal variations in semen parameters of Black Bengal goats. *Indian Journal of Animal Science* 84, 635–639.

- Larzul C, Terenina E, Foury A, Billon Y, Louveau I, Merlot E and Mormede P 2015. The cortisol response to ACTH in pigs, heritability and influence of corticosteroid-binding globulin. *Animal* 9, 1929–1934.
- Lee JA, Rousset JD and Beatty JF 1976. Effect of temperature-season on bovine adrenal cortical function, blood cell profile, and milk production. *Journal of Dairy Science* 59, 104–108.
- Malpaux B, Migaud M, Tricoire H and Chemineau P 2001. Biology of mammalian photoperiodism and the critical role of the pineal gland and melatonin. *Journal of Biological Rhythms* 16, 336–347.
- Maltz E, Silanikove N and Shkolnik A 2009. Energy cost and water requirement of black Bedouin goats at different levels of production. *Journal of Agricultural Science* 98, 499–504.
- Marple DN, Aberle ED, Forrest JC, Blake WH and Judge MD 1972. Effects of humidity and temperature on porcine plasma adrenal corticoids, ACTH and growth hormone levels. *Journal of Animal Science* 34, 809–812.
- Mesbah S and Brudieux R 1982. Diurnal variation of plasma concentrations of cortisol, aldosterone and electrolytes in the ram. *Hormone and Metabolic Research* 14, 320–323.
- Meza-Herrera CA, Bocanegra JA, Banuelos R, Arechiga CF, Rincon RM, Ochoa-Cordero MA, Juarez-Reyes AS, Cerrillo-Soto MA and Salinas H 2007. Circannual fluctuations in serum cortisol and glucose concentrations and hair coat growth in goats. *Journal of Applied Animal Research* 31, 79–82.
- Minka NS and Ayo JO 2012. Assessment of thermal load on transported goats administered with ascorbic acid during the hot-dry conditions. *International Journal of Biometeorology* 56, 333–341.
- Mormede P, Andanson S, Auperin B, Beerda B, Guemene D, Malmkvist J, Manteca X, Manteuffel G, Prunet P, van Reenen CG, Richard S and Veissier I 2007. Exploration of the hypothalamic-pituitary-adrenal function as a tool to evaluate animal welfare. *Physiology and Behavior* 92, 317–339.
- Mormede P and Terenina E 2012. Molecular genetics of the adrenocortical axis and breeding for robustness. *Domestic Animal Endocrinology* 43, 116–131.
- Pevet P and Challet E 2011. Melatonin: both master clock output and internal time-giver in the circadian clocks network. *Journal of Physiology, Paris* 105, 170–182.
- Pruessner JC, Kirschbaum C, Meinlschmid G and Hellhammer DH 2003. Two formulae for the computation of the area under the curve represent measures of total hormone concentration versus time-dependent change. *Psychoneuroendocrinology* 28, 916–931.
- Redondo E, Franco A, García A and Masot AJ 2010. Changes in concentrations of cortisol and melatonin in plasma, expression of synaptophysin, and ultra-structural properties of pinealocytes in goat kids in situations of stress due to early weaning: the effect of melatonin. *New Zealand Veterinary Journal* 58, 160–167.
- Reinberg A, Touitou Y, Levi F and Nicolai A 1983. Circadian and seasonal changes in ACTH-induced effects in healthy young men. *European Journal of Clinical Pharmacology* 25, 657–665.
- Rhind SM and McMillen SR 1995. Seasonal changes in systemic hormone profiles and their relationship to patterns of fibre growth and moulting in goats of contrasting genotypes. *Australian Journal of Agricultural Research* 46, 1273–1283.
- Romero M 2002. Seasonal changes in plasma glucocorticoid concentrations in free-living vertebrates. *General and Comparative Endocrinology* 128, 1–24.
- Shamay A, Mabjeesh S, Shapiro F and Silanikove N 2000. Adrenocorticotrophic hormone and dexamethasone failed to affect milk yield in dairy goats: comparative aspects. *Small Ruminant Research* 38, 255–259.
- Sharma S, Ramesh K, Hyder I, Uniyal S, Yadav VP, Panda RP, Maurya VP, Singh G, Kumar P, Mitra A and Sarkar M 2013. Effect of melatonin administration on thyroid hormones, cortisol and expression profile of heat shock proteins in goats (*Capra hircus*) exposed to heat stress. *Small Ruminant Research* 112, 216–223.
- Silanikove N 2000. The physiological basis of adaptation in goats to harsh environments. *Small Ruminant Research* 35, 181–193.
- Silanikove N, Tagari H and Shkolnik A 1993. Comparison of rate of passage, fermentation rate and efficiency of digestion of high fiber diet in desert Bedouin goats compared to Swiss Saanen goats. *Small Ruminant Research* 12, 45–60.
- Ssewanyana E, Lincoln GA, Linton EA and Lowry PJ 1990. Regulation of the seasonal cycle of  $\beta$ -endorphin and ACTH secretion into the peripheral blood of rams. *Journal of Endocrinology* 124, 443–454.
- Todini L, Terzano GM, Borghese A, Debenedetti A and Malfatti A 2011. Plasma melatonin in domestic female Mediterranean sheep (Comisana breed) and goats (Maltese and Red Syrian). *Research in Veterinary Science* 90, 35–39.
- Tong YL 1976. Parameter estimation in studying circadian rhythms. *Biometrics* 32, 85–94.
- Torres-Farfan C, Valenzuela FJ, Mondaca M, Valenzuela GJ, Krause B, Herrera EA, Riquelme R, Llanos AJ and Seron-Ferre M 2008. Evidence of a role for melatonin in fetal sheep physiology: direct actions of melatonin on fetal cerebral artery, brown adipose tissue and adrenal gland. *Journal of Physiology* 586, 4017–4027.
- Yagil R and Etzion Z 1979. The role of antidiuretic hormone and aldosterone in the dehydrated and rehydrated camel. *Comparative Biochemistry and Physiology Part A: Physiology* 63, 275–278.
- Zarazaga LA, Guzmán JL, Domínguez C, Pérez MC and Prieto R 2009. Effects of season and feeding level on reproductive activity and semen quality in Payoya buck goats. *Theriogenology* 71, 1316–1325.