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## Development and Characterization of an In Vivo Pathogenic Molecular Clone of Equine Infectious Anemia Virus

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An infectious nonpathogenic molecular clone (19-2-6A) of equine infectious anemia virus (EIAV) was modified by substitution of a 3.3-kbp fragment amplified by PCR techniques from a pathogenic variant (EIAV<sub>PV</sub>) of the cell culture-adapted strain of EIAV (EIAV<sub>PR</sub>). This substitution consisted of coding sequences for 77 amino acids at the carboxyl terminus of the integrase, the S1 (encoding the second exon of *tat*), S2, and S3 (encoding the second exon of *rev*) open reading frames, the complete *env* gene (including the first exon of *rev*), and the 3' long terminal repeat (LTR). Modified 19-2-6A molecular clones were designated EIAV<sub>PV3.3</sub>, and infection of a single pony (678) with viruses derived from a mixture of five of these molecular clones induced clinical signs of acute equine infectious anemia (EIA) at 23 days postinfection (dpi). As a consequence of this initial study, a single molecular clone, EIAV<sub>PV3.3#3</sub> (redesignated EIAV<sub>UK</sub>), was selected for further study and inoculated into two ponies (613 and 614) and two horses (700 and 764). Pony 614 and the two horses developed febrile responses by 12 dpi, which was accompanied by a 48 to 64% reduction in platelet number, whereas pony 613 did not develop fever (40.6°C) until 76 dpi. EIAV could be isolated from the plasma of these animals by 5 to 7 dpi, and all became seropositive for antibodies to this virus by 21 dpi. Analysis of the complete nucleotide sequence demonstrated that the 3.3-kbp 3' fragment of EIAV<sub>UK</sub> differed from the consensus sequence of EIAV<sub>PV</sub> by just a single amino acid residue in the second exon of the *rev* gene. Complete homology with the EIAV<sub>PV</sub> consensus sequence was observed in the hypervariable region of the LTR. However, EIAV<sub>UK</sub> was found to contain an unusual 68-bp nucleotide insertion/duplication in a normally conserved region of the LTR sequence. These results demonstrate that substitution of a 3.3-kbp fragment from the EIAV<sub>PV</sub> strain into the infectious nonpathogenic molecular clone 19-2-6A leads to the production of progeny virus particles with the ability to induce clinical signs of EIA. Therefore, EIAV<sub>UK</sub>, which is the first pathogenic, cell culture-adapted molecular clone of EIAV to be described, should be of value in identifying viral determinants of pathogenicity.

Infection of horses and ponies (*Equus caballus*) with the equine lentivirus equine infectious anemia virus (EIAV) results in a persistent, life-long infection in which evidence of extensive active viral replication can be detected in blood samples as much as 20 years after the initial diagnosis (30). However, the course and severity of the disease following infection vary considerably, with some animals suffering debilitating cycles of illness and occasionally even death, while others show no obvious clinical signs. In experimental infections where disease is apparent, the first clinical episode usually occurs within 42 days after administration of the virus (reviewed in references 6, 10, 40, and 54). By convention, this first fever episode is referred to as the acute stage of equine infectious anemia (EIA) and is characterized by pyrexia, thrombocytopenia, anorexia, depression, and high plasma viremia levels (reviewed in references 6, 10, 40, and 54). Anemia is not usually detected at this stage (reviewed in references 6, 10, 40, and 54). Resolution of this first febrile episode is normally observed after 1 to 5 days and occurs concomitantly with a dramatic drop in the amount of plasma-associated virus (reviewed in references 6, 10, 40, and 54). Following the acute stage, some

animals may remain clinically normal (20), while others go on to experience multiple bouts of illness in which severe anemia may accompany pyrexia, thrombocytopenia, edema, and dramatic weight loss (reviewed in references 6, 10, 40, and 54).

The mechanisms that control the broad range of clinical responses are unknown but almost certainly involve both host and viral factors, including the probable existence of EIAV strains with widely different pathogenic potentials. In the laboratory, it is possible to attenuate virulent wild-type isolates of EIAV by adaptation to growth in fibroblastic cell types (3, 19). These wild-type isolates of EIAV are normally tropic for cells of the monocyte/macrophage lineage (24–27, 39, 53, 59), and so virulence determinants must either be lost or altered when selection pressure in the form of growth in nonnatural target cells is applied.

In this laboratory, the avirulent, cell-adapted version of the macrophage tropic Wyoming isolate (36) is designated the prototype (EIAV<sub>PR</sub>) strain of EIAV. Infection of horses and ponies with this virus produces no discernible signs of disease as monitored by rectal temperature, platelet numbers, or behavioral changes, within a normal observation period of 100 to 150 days (21, 22). However, pathogenic properties were restored when the EIAV<sub>PR</sub> strain was subjected to three serial passages in ponies (44), followed by selection and biological cloning of a neutralization escape mutant (50). This antigenic variant is designated EIAV<sub>PV</sub>. Infection of horses and ponies

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with this strain routinely induces symptoms of acute EIA within 42 days (11, 20, 22, 60), although variability between individual animals in the severity of disease and the number of febrile episodes is observed. Interestingly, EIAV<sub>PV</sub> retained the in vitro cell tropism characteristics of the EIAV<sub>PR</sub> strain, indicating that virulence and ability to grow in fibroblastic cell cultures are not mutually exclusive (50). The existence of related strains of EIAV with clearly defined virulence characteristics provides the basis of an excellent system to study viral factors associated with pathogenicity. However, to exploit this system fully, the development of pathogenic infectious molecular clones of EIAV with properties similar to those of the EIAV<sub>PV</sub> strain is required. This would allow the identification of pathogenic determinants within the EIAV genome by the production of recombinants between pathogenic and the nonpathogenic molecular clones that have been described previously (45).

A number of infectious molecular clones have been produced from fetal equine kidney (FEK) cells apparently infected with the EIAV<sub>PV</sub> strain (45). However, viruses derived from these molecular clones were found to be nonpathogenic when inoculated into ponies (45). Furthermore, a comparison of the nucleotide sequence of the surface unit (SU) glycoprotein demonstrated a closer homology to the EIAV<sub>PR</sub> strain (49) than to a consensus sequence for the EIAV<sub>PV</sub> strain (32, 34). This conclusion was reinforced by neutralization studies using two of the infectious molecular clones (44-1 and 19-2), where greater reactivity was observed with serum samples taken from ponies infected with the EIAV<sub>PR</sub> strain than the EIAV<sub>PV</sub> strain (7, 9).

We have confirmed that the infectious clone 19-2 is not pathogenic in ponies and that pathogenicity is not restored (33) by the introduction of a second copy of the putative transcription regulatory element, CAAT, into the long terminal repeat (LTR). The rationale for this modification was that while a double CAAT-box motif is present in the LTR of the virulent EIAV<sub>WY</sub> strain, only a single copy is found in the nonpathogenic EIAV<sub>PR</sub> strain (29, 35, 46, 49). Furthermore, pathogenicity was not restored to 19-2 by substitution of a EIAV<sub>PV</sub> strain-like 3' LTR or by a double substitution involving the EIAV<sub>PV</sub> 3' LTR and modification of the variable regions of the SU glycoprotein gene to make them identical to those of the consensus sequence for the EIAV<sub>PV</sub> strain (34).

In an attempt to produce an infectious molecular clone of EIAV with the ability to induce disease in horses and ponies, we modified the version of molecular clone 19-2 containing a double CAAT motif (19-2-6A) in the 3' LTR by substituting a 3.3-kbp fragment obtained by PCR amplification of EIAV<sub>PV</sub>-infected FEK cells. This fragment was replaced using a unique *NcoI* site situated in the *pol* gene of EIAV. Therefore, it encompassed sequences encoding 77 amino acids from the carboxyl terminus of the integrase protein, the S1 (encoding the second exon of *tat* [14, 42, 56]), S2 (encoding a 7-kDa polypeptide of unknown function [52]), and S3 (encoding the second exon of *rev* [17, 42, 48, 56]) open reading frames (ORFs), the complete *env* gene (including the first exon of *rev* [42, 48, 56]), and the 3' LTR. These modified molecular clones were tested for the ability to induce disease in horses and ponies.

#### MATERIALS AND METHODS

**Virus strains and cell culture.** The pathogenic EIAV<sub>PV</sub> strain (44, 50), derived from the avirulent cell-adapted EIAV<sub>PR</sub> strain (36), was grown in primary FEK cell cultures as described previously (22).

**Preparation of DNA from EIAV<sub>PV</sub>-infected cell cultures.** EIAV<sub>PV</sub>-infected cell cultures were trypsinized and washed in Hanks buffered saline solution to re-

move contaminating proteins from the maintenance medium. The cells were resuspended and lysed in 0.6% sodium dodecyl sulfate (SDS)-50 mM Tris hydrochloride (pH 7.5)-10 mM EDTA. This lysate was digested with RNase (100 µg/ml) at 50°C for 30 min and with proteinase K (0.1 mg/ml) at 50°C for 15 h and then subjected to phenol-chloroform-isoamyl alcohol extractions (25:24:1) to remove protein, followed by two chloroform-isoamyl alcohol (24:1) extractions and ethanol precipitation.

**PCR amplification and molecular cloning of the 3.3-kbp 3'-terminal fragments from the EIAV<sub>PV</sub> strain.** The 3.3-kbp 3'-terminal fragment of the EIAV<sub>PV</sub> strain was amplified from FEK-infected cell DNA by using primers 4876 and 8196 (Fig. 1). To increase fidelity, this amplification was conducted with KlenTaq 1 (Ab Peptides, Inc., St. Louis, Mo.) and *Pfu* (Stratagene, La Jolla, Calif.) thermostable polymerases as described by Barnes (2). Briefly, each 100-µl reaction mixture contained 20 mM Tris-HCl (pH 9.1), 1.6 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 3.5 mM MgCl<sub>2</sub>, 15 µg of bovine serum albumin, 200 µM each deoxynucleoside triphosphate, 20 pmol of each primer, 25 U of KlenTaq 1, and 0.156 U of *Pfu* polymerase. Following the addition of 0.1 µg of EIAV<sub>PV</sub>-infected FEK DNA, each reaction was subjected to 30 cycles of melting at 94°C for 20 s, annealing at 66°C for 30 s, and extension at 68°C for 8 min. An aliquot of each reaction mixture was analyzed for the presence of a 3.3-kbp product on an 0.8% agarose-Tris borate gel (51), while additional aliquots (approximately 500 ng) were blunt ended by using a PCR polishing kit (Stratagene) according to the manufacturer's instructions. Blunt-ended PCR products were extracted once with phenol-chloroform-isoamyl alcohol (25:24:1) and twice with chloroform-isoamyl alcohol (24:1) before being ethanol precipitated. This precipitated material was resuspended in sterile distilled H<sub>2</sub>O, digested with *NcoI*, and separated in a 1% low-melting-point agarose gel in TAE (51) buffer. The 3.3-kbp fragment was purified from agarose gel by digestion with Agar ACE (Promega, Madison, Wis.) according to the manufacturer's recommendations. This purified fragment was ligated into *NcoI*-*SmaI*-digested pLG338/30 plasmid DNA (Fig. 1), and the products of this ligation were used to transform *Escherichia coli* Sure cells (Stratagene). Plasmid pLG338/30 is a low-copy-number vector designed to stabilize the EIAV *env* gene during molecular cloning in *E. coli* (12). The resultant molecular clones were designated pLG338/PV3.3.

**Construction of EIAV<sub>PV3.3</sub> chimeric molecular clones.** A modified 19-2 infectious molecular clone of EIAV (45) containing a double CAAT-box motif in the 3' LTR, designated 19-2-6A (33), was digested with *NcoI* and *EcoRI*, and the fragments (9.7 and 3.7 kbp) were separated on a low-melting-point agarose gel. The 9.7-kbp fragments, containing the plasmid vector, 5' LTR, *gag*, and *pol* sequences, was recovered from the agar by digestion with Agar ACE. Five pLG338/PV3.3 molecular clones were selected after partial sequencing of the *env* gene, their plasmid DNAs were digested with *NcoI* and *EcoRI*, and the 3.3-kbp EIAV<sub>PV</sub> fragment was recovered by Agar ACE digestion from a 1% low-melting-point agarose gel. These EIAV<sub>PV</sub> 3.3-kbp *NcoI*-*EcoRI* fragments were ligated to the 9.7-kbp purified fragment from 19-2-6A to generate chimeric proviral molecular clones of EIAV designated EIAV<sub>PV3.3</sub> (Fig. 1).

**Transfection of cell cultures.** Plasmid DNA preparations from five EIAV<sub>PV3.3</sub> molecular clones of EIAV were purified with minor modifications by a polyethylene glycol precipitation procedure described previously (28). These modifications included a phenol-chloroform-isoamyl alcohol (25:24:1) extraction of the supernatant obtained after centrifugation of the bacterial cell lysate and digestion of RNase A-treated nucleic acids with proteinase K (250 µg/ml) in the presence of 0.25% SDS for 30 min at 37°C. Purified EIAV<sub>PV3.3</sub> plasmid DNAs (4 µg) were transfected into FEK cell cultures (at  $2 \times 10^5$  cells per 35-mm-diameter well) by using a polycationic lipid transfection reagent (Lipofectamine; Gibco Life Technologies, Gaithersburg, Md.) according to the manufacturer's instructions. After a 21- to 24-day incubation period (which included passage and amplification of the cell numbers), tissue culture fluids from transfected cells cultures were screened for released reverse transcriptase activity (8), aliquoted, and stored at -70°C until further use. The titers of the chimeric virus stocks were determined by performing serial log<sub>10</sub> dilutions in modified Eagle's medium containing 3% fetal bovine serum, 25 mM HEPES, 10 mM NaHCO<sub>3</sub>, 100 IU of penicillin/ml, and 100 µg of streptomycin/ml and inoculating each dilution onto fresh FEK cell cultures. These FEK cell cultures were monitored for 21 days, after which the culture fluids were assayed for reverse transcriptase activity (8) and the titration endpoint was determined by the Karber method (15).

**Experimental subjects.** The ponies (613, 614, and 678) used in these studies were of mixed breed origin, while the horses (700 and 764) were thoroughbreds. Prior to the experiment, serum samples from all animals were tested by the agar gel immunodiffusion test (IDEXX Laboratories Inc., Westbrook, Mass.) to ensure that they were seronegative for EIAV. The animals were housed in screened box stalls to exclude hematophagous insects, and all animal handling protocols were approved by the University of Kentucky Institutional Animal Care and Use Committee. The horses and ponies were treated with Ivermectin for internal parasites and vaccinated against tetanus toxoid (Merck, Sharp and Dohme, Rahway, N.J.). Animals were inoculated intravenously, using clarified (3,000 × g for 30 min at 4°C) cell culture fluids from EIAV<sub>PV3.3</sub>-transfected cell cultures at  $4 \times 10^5$  or  $1 \times 10^5$  50% tissue culture infective doses (TCID<sub>50</sub>) and monitored on a daily basis for rectal temperature and overall clinical status. Blood samples were collected at regular intervals by jugular venipuncture into sterile glass evacuated tubes (Becton Dickinson, Rutherford, N.J.) for serum or with EDTA for plasma (Sherwood Medical, St. Louis, Mo.). Serum and plasma were stored

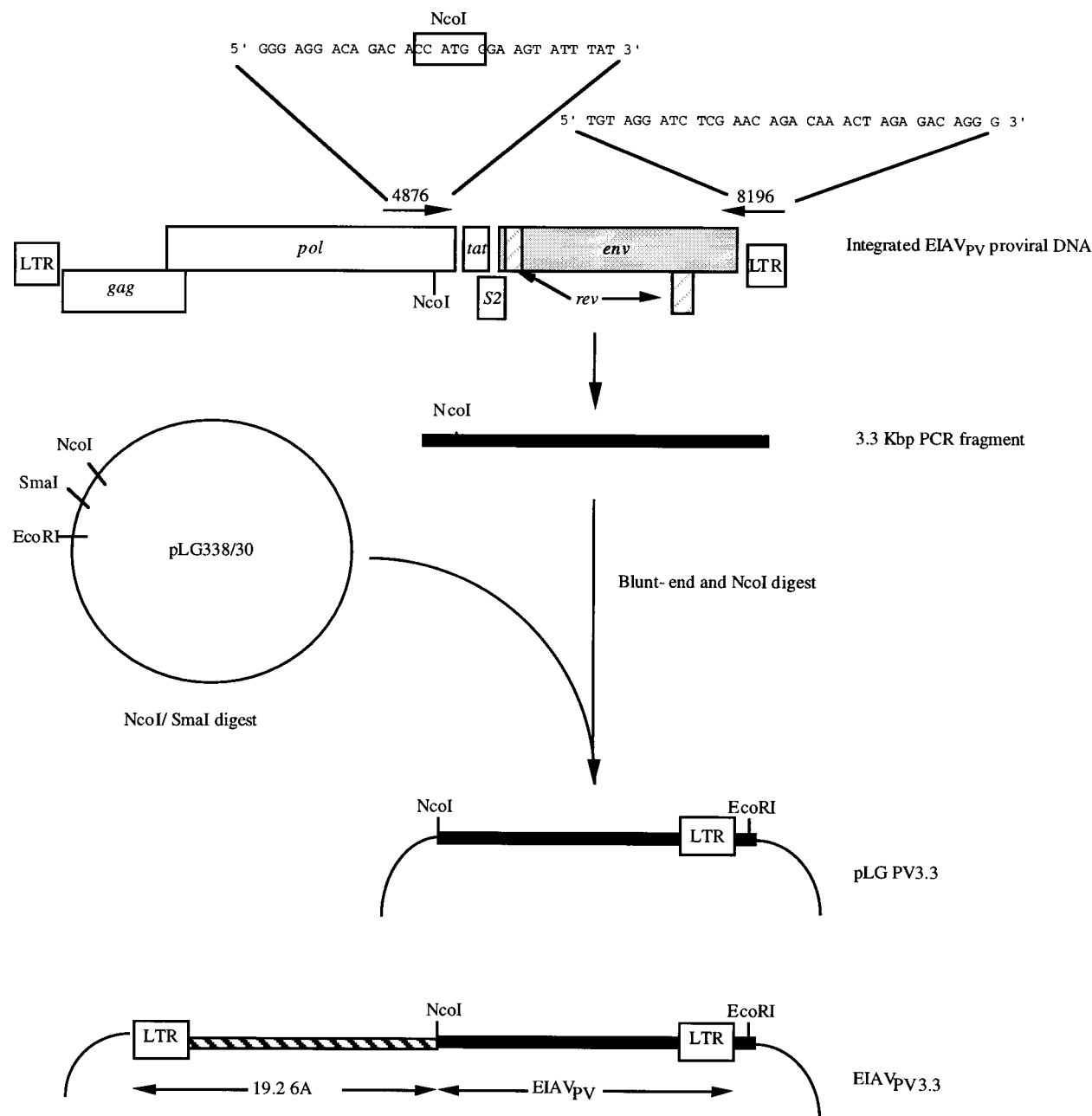


FIG. 1. Amplification and molecular cloning of the 3.3-kbp 3'-terminal fragment of EIAV<sub>PV</sub>. A 3.3-kbp fragment from EIAV<sub>PV</sub>-infected cell cultures was amplified by PCR, blunt ended, and digested with *NcoI* prior to ligation with *NcoI*/*SmaI*-digested pLG338/30 vector DNA. The resultant pLG338/PV3.3 molecular clones were digested with *NcoI* and *EcoRI*, and the 3.3-kbp EIAV<sub>PV</sub> fragment was inserted into the infectious molecular clone 19-2-6A. This produced chimeric proviral molecular clones designated EIAV<sub>PV3.3</sub>.

at  $-20$  and  $-70^{\circ}\text{C}$ , respectively. In addition, fresh blood samples collected in the presence of EDTA were analyzed to determine platelet numbers, using the Unopette system (Becton Dickinson). Platelet numbers were determined at least weekly and during each febrile episode ( $>39.4^{\circ}\text{C}$ ). Virus isolation from plasma samples on FEK cells cultures was described previously (22, 60), and determination of viral titers in these samples was conducted by inoculation of serial  $\log_{10}$  dilutions onto FEK cells (22, 60). All cultures were maintained for at least 21 days, after which they were assayed for reverse transcriptase activity (8). Serum samples were tested for antibodies to the p26 antigen of EIAV by equine infectious anemia-competitive enzyme-linked immunosorbent assay (EIA-CELISA; IDEXX Laboratories).

**Nucleotide sequencing of EIAV<sub>UK</sub> (EIAV<sub>PV3.3#3</sub>).** Plasmid DNA was extracted and purified with a Midiprep kit (Qiagen, Chatsworth, Calif.). This purified DNA was sequenced automatically with a Taq Dye Deoxy Terminator Cycle Sequencer kit (Perkin-Elmer, Applied Biosystems Foster City, Calif.), using overlapping

EIAV primers. DNA sequences were resolved with an ABI Prism 373 DNA sequencer (Applied Biosystems). The sequences were analyzed by using the Genetics Computer Group (Madison, Wis.) package analysis software (18). Oligonucleotide primers for these sequencing reactions were derived from published sequences of the EIAV genome (32, 45, 49, 55 [GenBank accession no. U01866]).

**Nucleotide sequence accession number.** The sequence of EIAV<sub>UK</sub> was submitted to GenBank and has been assigned accession no. AF016316.

## RESULTS

**Amplification of the 3.3-kbp 3'-terminal fragment from EIAV<sub>PV</sub>-infected cell cultures and production of EIAV<sub>PV3.3</sub> chimeric proviral DNA molecular clones.** A modified version



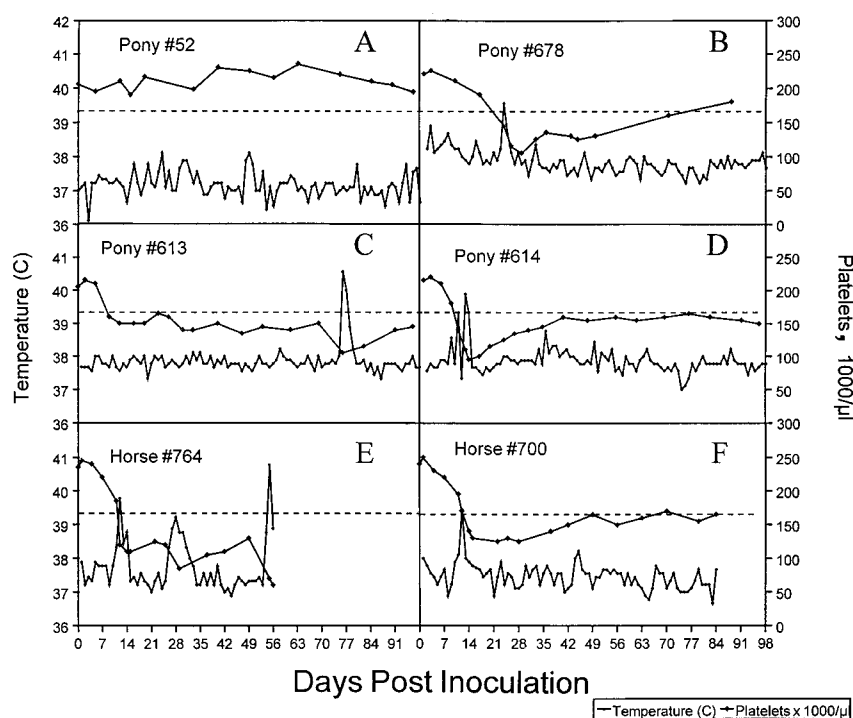


FIG. 2. Clinical responses in animals infected with EIAV derived from infectious molecular clones. Body temperature and platelet count are shown for animals infected with viruses from molecular clone 19-2-6A (A), viruses from a mixture of EIAV<sub>PV3.3</sub> molecular clones (B), and viruses from molecular clone EIAV<sub>UK</sub> (C to F).

of the PCR (2) was used with the oligonucleotide primer pair 4876-8196 (Fig. 1) against a template DNA prepared from EIAV<sub>PV</sub>-infected FEK cells cultures to produce 3'-terminal fragments of the proviral genome. This primer pair was designed to produce a 3,340-bp fragment encompassing coding sequences for 77 amino acids at the carboxyl terminus of integrase, the S1, S2, and S3 ORFs, the complete *env* gene, and the 3' LTR. Analysis of the PCR products by agarose gel electrophoresis, followed by ethidium bromide staining, demonstrated the presence of a single band with the predicted size of approximately 3.3 kbp. These PCR products were incubated with *Pfu* polymerase to remove possible 3' single nucleotide overhangs that may have been added during PCR and then digested with *NcoI*. A unique *NcoI* site is situated at position 4889 within the genomes of all strains of EIAV which have been examined in this laboratory, and this was included in the oligonucleotide primer 4876 (Fig. 1) to simplify subsequent molecular cloning steps. The 3.3-kbp fragments containing a cleaved *NcoI* site at the 5' terminus and a blunt 3' terminus were ligated into the low-copy-number plasmid vector pLG338/30 that had been digested with *NcoI* and *SmaI* (Fig. 1). Five of the resultant molecular clones, designated pLG338/PV3.3, were selected at random and partially characterized by determining the nucleotide sequence of a 459-bp segment encompassing four of the eight variable regions (32) within the coding domain for the SU glycoprotein. At both nucleotide and amino acid levels, two of the pLG338/PV3.3 molecular clones showed complete homology with the consensus sequence for the EIAV<sub>PV</sub> strain (32, 34), while two differed by just a single amino acid (99.95% homology) and the remaining clone differed by three amino acids (98.04% homology). These results suggested that the pLG338/PV3.3 molecular clones were a family of closely related sequences, representative of those found in viral stocks of the EIAV<sub>PV</sub> strain (31, 32, 34).

Therefore, the 3.3-kbp EIAV<sub>PV</sub> sequences were excised from the plasmid by digestion with *NcoI*/*EcoRI* and substituted into the infectious molecular clone 19-2-6A (Fig. 1) to create chimeric proviral molecular clones designated EIAV<sub>PV3.3</sub>. These chimeric molecular clones were transfected into FEK cell cultures, either individually or as a mixture. In the case of the mixture, each of the five plasmid DNA preparations was added in equal proportions (0.8  $\mu$ g) to produce a total of 4.0  $\mu$ g of DNA per transfection. Following an incubation period of 3 weeks, reverse transcriptase activity was detected in cell culture fluids from both the mixed and individual molecular clone transfections. The amount of virus produced by the transfected cell cultures was determined by titration of cell culture fluids in FEK cells. The mixture and each of the individual EIAV<sub>PV3.3</sub> molecular clones produced infectious progeny virus titers of  $10^{6.0}$  TCID<sub>50</sub>/ml, which is equivalent to the yields obtained following transfection with the parental 19-2-6A molecular clone.

**Clinical responses following infection with viruses derived from a mixture of EIAV<sub>PV3.3</sub> molecular clones.** Infection of ponies with viruses from molecular clone 19-2-6A produced none of the clinical responses (6, 10, 40, 54), such as fever or thrombocytopenia, which are normally associated with acute EIA (Fig. 2A). Therefore, a preliminary experiment was conducted to investigate the effects in vivo of replacing the 3.3-kbp 3'-terminal fragment from 19-2-6A with equivalent sequences derived from the EIAV<sub>PV</sub> strain. For this preliminary experiment, a single pony (678) was inoculated intravenously with  $4 \times 10^{6.0}$  TCID<sub>50</sub> of the virus population derived from the mixed transfection of five EIAV<sub>PV3.3</sub> molecular clones. A mixture rather than a single source was used because it was considered potentially more economical in terms of animal resources for the initial screening of chimeric molecular clones.

Pony 678 experienced a febrile episode (39.6°C) 23 days

TABLE 1. Serological and virological responses following infection with EIAV<sub>PV3.3</sub> strains

Animal no.	Virus strain	Time (dpi) of viremia <sup>a</sup>	Viremia titer <sup>b</sup> during 1st febrile episode	Time (dpi) of seroconversion <sup>c</sup>
678	EIAV <sub>PV3.3</sub> mix <sup>d</sup>	2–21	10 <sup>4.0</sup>	18
613	EIAV <sub>UK</sub>	5–14, 16, 21, 28, 35	10 <sup>3.5</sup>	19
614	EIAV <sub>UK</sub>	5–16, 22	10 <sup>4.5</sup>	19
700	EIAV <sub>UK</sub>	7–18	10 <sup>4.0</sup>	21
764	EIAV <sub>UK</sub>	7–18	10 <sup>5.0</sup>	21

<sup>a</sup> Virus isolation in FEK cells was carried out on plasma samples collected until 21 dpi for animal 678, up to 40 dpi for animals 613 and 614, and until 18 dpi for animals 700 and 764.

<sup>b</sup> Expressed as TCID<sub>50</sub>/ml of plasma sample.

<sup>c</sup> Seroconversion is defined as the time taken following infection to produce a positive reaction in the EIA-CELISA.

<sup>d</sup> Mixture of five EIAV<sub>PV3.3</sub> infectious molecular clones.

postinfection (dpi) with a concomitant decline in platelet levels from 220,000/μl before infection to 105,000/μl at 28 dpi (Fig. 2B). This pony developed antibodies to the major core protein (p26) of EIAV as measured by the commercially available EIA-CELISA by 18 dpi (Table 1), and EIAV could be isolated on FEK cell cultures from plasma samples as early as 2 dpi (Table 1). In addition, the plasma viremia titer during the febrile episode (23 dpi) was 10<sup>4.0</sup> TCID<sub>50</sub>/ml, which is similar to the levels found in many ponies infected with the EIAV<sub>PV</sub> (11, 20, 22, 60).

**Clinical responses following infection of horses and ponies with viruses derived from a single EIAV<sub>PV3.3</sub> molecular clone.** The results obtained with pony 678 suggested that a pathogenic phenotype typical of acute EIA in terms of involving pyrexia and thrombocytopenia (reviewed references 6, 10, 40, and 54) could be conferred on virions from the 19-2-6A molecular clone by replacement of the 3.3-kbp 3'-terminal fragment with equivalent sequences derived from the EIAV<sub>PV</sub> strain. However, it was important to determine if the clinical responses in pony 678 could be duplicated in other animals by using viruses derived from a single EIAV<sub>PV3.3</sub> molecular clone. As a starting point for these experiments, the chimeric molecular clone EIAV<sub>PV3.3#3</sub> (redesignated EIAV<sub>UK</sub>) was selected because it had complete homology to the consensus sequence of the EIAV<sub>PV</sub> strain in the 459-bp region of the EIAV SU glycoprotein that was used in preliminary characterization of the 3.3-kb EIAV<sub>PV</sub> fragments.

Two ponies (613 and 614) and two horses (700 and 764) were inoculated intravenously with viruses derived from molecular clone EIAV<sub>UK</sub> at doses of  $4 \times 10^{6.0}$  and  $1 \times 10^{5.0}$  TCID<sub>50</sub>, respectively. The rectal temperature of pony 614 reached 39.3°C at 10 dpi, declined to normal levels (37 to 38°C) at 11 dpi, and then rose again to 39.9°C at 12 dpi (Fig. 2D). Platelet numbers in this animal declined 56% from 215,000/μl before infection to 95,000/μl at 13 dpi (Fig. 2D). Febrile episodes ( $\geq 39.4^\circ\text{C}$ ) were also observed in the two horses at 12 dpi, with maximal declines in platelet numbers of 48% for horse 700 and 64% for horse 764 during the first 28 dpi (Fig. 2E and F). In contrast, pony 613 did not exhibit a febrile response (40.6°C) until 76 dpi and showed only a slight decrease (27%) in platelet levels during the first 28 dpi. However, during the febrile episode, platelet levels in pony 613 did decline to 105,000/μl, which is a decrease of 49% from the preinfection level (Fig. 2C). Ponies 613 and 614 and horse 700 experienced only a single bout of disease during the postinfection observation periods of 98 and 84 dpi, respectively. However, horse 764

had a second febrile episode (39.2°C) at 28 dpi, with a decrease in platelets from 120,000/μl at 25 dpi to 85,000/μl at 28 dpi. In addition, at 55 dpi this horse became severely depressed and anorexic, with a body temperature of 40.8°C. Platelet levels in 764 decreased to 60,000/μl, and the animal was euthanized at 56 dpi. At necropsy, no gross macroscopic lesions were detected, although histochemical examination revealed Kupffer cell hyperplasia in sections of the liver. These cells contained hemosiderin aggregates, which is a pathological feature associated with EIA (6, 10, 40, 54). Bacteriologic examination of tissues (lung, liver, small intestine, and colon) from 764 produced no evidence of pathogenic bacteria.

**Serological and virological responses following infection with EIAV<sub>UK</sub>.** Serological responses to the infection with viruses from EIAV<sub>UK</sub> plasmid were examined by using a commercially available EIA-CELISA. In this test, both ponies (613 and 614) became seropositive to p26 at 19 dpi, with the horses (700 and 764) becoming positive at 21 dpi (Table 1). All animals remained seropositive throughout the duration of the experiment.

Plasma samples from all animals (613, 614, 700, and 764) were assayed at various time points postinfection for the presence of EIAV by inoculation onto susceptible FEK cell cultures, followed by measurement of reverse transcriptase activity in cell culture fluids. Using this technique, we detected virus in pony 613 plasma samples from 5 to 14 dpi, with additional positive samples at 16, 21, 28, and 35 dpi (Table 1). Similarly, virus was detected in samples from pony 614 at each time point from 5 to 16 dpi and then again at 22 dpi (Table 1). In the case of the horses, virus was detected at each time point from 7 to 18 dpi.

This analysis was extended to determine the amounts of virus present during the first febrile episode. Titrations of plasma samples revealed peak viremia of 10<sup>4.5</sup> TCID<sub>50</sub>/ml for pony 614, 10<sup>3.5</sup> TCID<sub>50</sub>/ml for pony 613, 10<sup>4.0</sup> TCID<sub>50</sub>/ml for horse 700, and 10<sup>5.0</sup> TCID<sub>50</sub>/ml for horse 764 (Table 1). These titers are very similar to those observed during febrile episodes in ponies infected with the EIAV<sub>PV</sub> strain (11, 20, 22, 60).

**Complete nucleotide sequence of EIAV<sub>UK</sub>.** The complete nucleotide sequence of molecular clone EIAV<sub>UK</sub> was determined as a prerequisite to the identification of viral determinants of pathogenicity. Comparison of these sequences revealed complete homology with those of molecular clone 19-2-6A from nucleotide position 1 in the 5' LTR to the *NcoI* site at position 4889 (data not shown). Although this result might be expected, it demonstrates that no mutations had been introduced into the sequence upstream of the *NcoI* site during molecular cloning steps. The nucleotide sequences downstream of the *NcoI* site were compared with the consensus sequence for the EIAV<sub>PV</sub> strain (32, 34), a cloned representative of EIAV<sub>PR</sub> and molecular clone 19-2 (Fig. 3). The nucleotide sequence of ORF S1, which encodes the second exon of *tat* (14, 42, 56), was identical in all viruses examined (data not shown). In the case of ORF S2, 19-2 was found to differ from the other viruses at amino acid positions 4 and 19 (Fig. 3A; Table 2). In the second exon of *rev* (ORF S3), the avirulent 19-2 and EIAV<sub>PR</sub> strains contained valine and tyrosine at amino acid positions 14 and 91, whereas the same positions in EIAV<sub>PV</sub> and EIAV<sub>UK</sub> were occupied by isoleucine and histidine (Fig. 3B; Table 2). Furthermore, 19-2 was found to contain a glycine rather than a serine residue at position 8 and EIAV<sub>UK</sub> was found to contain an arginine rather than a tryptophan at position 103 (Fig. 3B; Table 2). In the sequences encoding polypeptides downstream of the *NcoI*, there were just three differences at the nucleotide level between EIAV<sub>UK</sub> and the consensus sequence for the EIAV<sub>PV</sub> strain. Of these dif-

A	EIAV <sub>UK</sub>	1	66
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	YMGLEFGKGV <sup>*</sup> T WSASHSMGGS QGESQPLLPN SQKNLSVRRT QCFNLIVIM TVRTAWQNRR KQETKK	
Nucleoprotein motif			
B	EIAV <sub>UK</sub>	1	100
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	consensus	DPQGPLESDQ WCR-LRQSLP EEKIPSTQCI ARRHLGPGPT QHTPSRRDRW IRGQILQAEV LQERLEWRIR GVQQAARELG EVNRGIWREL -FREDQRGDF	
	EIAV <sub>UK</sub>	101	135
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	consensus	SAWGGYQRAQ ERLWGEQSSP RVLPRGDSKR RRKHL	
C	EIAV <sub>UK</sub>	1	100
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	MVSIAPFYGGI PGGISTPITQ QSEKSKCEEN TMFPQPYCYNN <u>DSKNSMAESK</u> <u>EARDQEMNLK</u> <u>ESKBEKRRN</u> DWWKIGMFLI CLAGTTGGIL WWYEGLPQQH	
	first exon rev		
	EIAV <sub>UK</sub>	101	200
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	YIGLVAIGCR <u>LNGSGQSNAI</u> ECWGSFPGCR PFQNYFSYET <u>NRSMHMDNNT</u> ATLEAYHRE ITFIYKSSCT DSDHCQBYQC KKVNLSSDS SN-VRVEDVM	
	EIAV <sub>UK</sub>	201	300
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	NT-EYWGPKW <u>LECNQYENFK</u> TILVPENEMV <u>NINDTDTWIP</u> KGCNETWARV KRCPIDILYG IHPRLCVQP PFFLVQEKGI A-TSRIGNCG PTIFLGVLED	
	EIAV <sub>UK</sub>	301	400
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	NKGVRG-YT <u>ACNV-RL-IN</u> RKDYTGIIYQV PIFYTC-PTN <u>ITSCNNEPII</u> SVMYETNQV QYLLCENNNS NNYNQVVQSF GVIGQAHLEL PRPNKRIRNQ	
	EIAV <sub>UK</sub>	401	500
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	SFNQYNCSIN <u>NKTELETWKL</u> VKTSGITPLP ISSEANTGLI RHKRD <sup>*</sup> FGISA IVAATVAATA TAASATMSYV ALTEVNKIME VQNTTFEVEN STLNGMDLIE	
	gp90 gp45		
	EIAV <sub>UK</sub>	501	600
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	RQKILYAMI LQTHADVQLL KERQQVEETF NLIGCIERTH VFCHTGPWN MSWHLNEST QWDDWVSKME DLNQEILT-L HGARNNLAQS MITFNTFDSI	
	EIAV <sub>UK</sub>	601	700
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	AQFGKDLWSH IGNWIPGLGA SIISKYIVMPL LIYLLLTSSP KILRALWKVT SGAGSSGSRY LKKKPHHKHA SREDTWDQAQ HNIHLAGVTG GSGDKYYKQK	
	EIAV <sub>UK</sub>	701	800
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	-SRNDWNGES EBYNRRPKSW VKSIEAFGES YISEKTKGEI SQPGAAINEH KNGSGGNNPH QGSLDLLEIRS EGGNIYDCCI KAQEGTLAIP CCGFPLWLFW	
	EIAV <sub>UK</sub>	801	859
	EIAV <sub>PV</sub>		
	EIAV <sub>PR</sub>		
	19.2		
	Consensus	GLVIIVGRIA GYGLRGLAVI IRICTIRGLNL IFBIIRKMLD YIGRALNPGT SHVSMPPQYV	

FIG. 3. Comparison of the deduced amino acid sequences encoded by EIAV<sub>UK</sub> with the 19-2 sequence, the EIAV<sub>PV</sub> consensus sequence, and the sequence of a cloned representative of the EIAV<sub>PR</sub> strain. Comparisons shown are for ORF S2 (A), ORF S3, which encodes the second exon of *rev* (B), and the envelope glycoprotein (C). Only the amino acid residues that differ from the total consensus sequence are indicated. A dash indicates that no consensus sequence was found. Potential N-linked glycosylation sites are underlined. Potential N-linked glycosylation sites that differ between (EIAV<sub>UK</sub> and EIAV<sub>PV</sub>) and EIAV<sub>PR</sub> and 19-2) are marked by asterisks.

TABLE 2. Differences in amino acid content between EIAV<sub>UK</sub> 19-2, EIAV<sub>PV</sub> (consensus), and EIAV<sub>PR</sub>

Coding sequence	Amino acid position	Amino acid <sup>a</sup>			
		EIAV <sub>UK</sub>	19-2	EIAV <sub>PV</sub>	EIAV <sub>PR</sub>
ORF S2	4	L	V	L	L
	19	G	E	G	G
ORF S3	8	S	G	S	S
	14	I	V	I	V
	91	H	Y	H	Y
	103	R	W	W	W
Env SU	193	P	S	P	S
	200	M	M	M	T
	203	T <sup>b</sup>	A	T <sup>b</sup>	A
	282	N <sup>b</sup>	D	N <sup>b</sup>	D
	308	N <sup>b</sup>	D	N <sup>b</sup>	D
	315	S <sup>b</sup>	R	S <sup>b</sup>	R
	318	K	N	K	N
	337	N <sup>b</sup>	T	N <sup>b</sup>	T
TM	426	I	I	I	V
	589	I	T	I	T
	701	C	Y	C	Y

<sup>a</sup> Indicated in single-letter code.<sup>b</sup> Change creates a potential N-linked glycosylation site.

ferences, only one, resulting in the arginine residue at position 103 in ORF S3, was nonsynonymous. In sequences encoding the SU glycoprotein, EIAV<sub>UK</sub> differed from 19-2 and EIAV<sub>PR</sub> by seven and nine amino acids, respectively. Furthermore, five of these differences resulted in the creation of potential N-linked glycosylation sites (Table 2). The transmembrane (TM) glycoprotein contained just two amino acid differences in which isoleucine at position 589 and cysteine at 701 in EIAV<sub>UK</sub> were represented by threonine and tyrosine in 19-2 and EIAV<sub>PR</sub> (Fig. 3C; Table 2). It is interesting that the differences between EIAV<sub>UK</sub> and 19-2 amounted to only 1.1% of the total amino acid content. However, a similar low level of variation has been observed previously between pathogenic and non-pathogenic (or weakly pathogenic) molecular clones of simian immunodeficiency virus (SIV) and visna virus (1, 38, 58). With one exception, extensive homology between EIAV<sub>UK</sub> and the EIAV<sub>PV</sub> consensus sequence was observed in the 3' LTR, which contains the *cis*-acting transcriptional control elements

(Fig. 4). The major difference between the two sequences was the presence of an unusual 68-bp duplication/insertion located in a normally conserved U3 region, 85 nucleotides from the 5' end of the 3' LTR (Fig. 5). With the exception of this duplication, the sequences downstream of the *Nco*I site in molecular clone EIAV<sub>UK</sub> are typical of the predominant forms found in the quasispecies comprising the EIAV<sub>PV</sub> strain.

## DISCUSSION

Previously it was established that viruses produced by transfection of the 19-2 and 19-2-6A proviral molecular clones of EIAV in equine cell cultures can establish a persistent infection in an equine host but are unable to induce clinical responses (33, 45). However, in this report, it is demonstrated that a virulent phenotype is conferred on viruses produced from the 19-2-6A molecular clone by replacement of the 3'-terminal 3,340-bp fragment with corresponding sequences derived from the pathogenic biological clone EIAV<sub>PV</sub>. The *in vivo* pathogenic properties of these chimeric molecular clones were evaluated by using viruses derived by transfection of FEK cell cultures at dose levels equivalent to those used for the 19-2, 19-2-6A, and EIAV<sub>PR</sub> strains (11, 22, 33, 34, 60). In the case of the infection with the single clone, EIAV<sub>UK</sub>, disease was observed by 12 dpi in three of the four recipients, which is well within the time frame expected for the onset of clinical signs following infection with similar amounts of the EIAV<sub>PV</sub> strain (21). The remaining recipient (613) did not experience a febrile episode until 76 dpi; however, the earliest detectable plasma viremia, the duration of the first viremia, and the approximate time of seroconversion were similar to those for the other EIAV<sub>UK</sub>-infected animals. A delayed onset of disease (which in this laboratory is defined as clinical signs occurring after 42 dpi) despite apparently normal kinetics of initial viral replication (as defined by initial detection of plasma viremia, duration of initial viremia, and time of seroconversion) has been previously observed in 3 of 23 EIAV<sub>PV</sub>-infected ponies (21). This finding suggests that although all ponies are susceptible to infection by EIAV<sub>PV</sub>, some are partially resistant to disease. In addition to a delayed onset of clinical signs, there is also variation between animals in the severity of disease following infection with EIAV. In a recent study investigating maturation of immune responses in four EIAV<sub>PV</sub>-infected po-

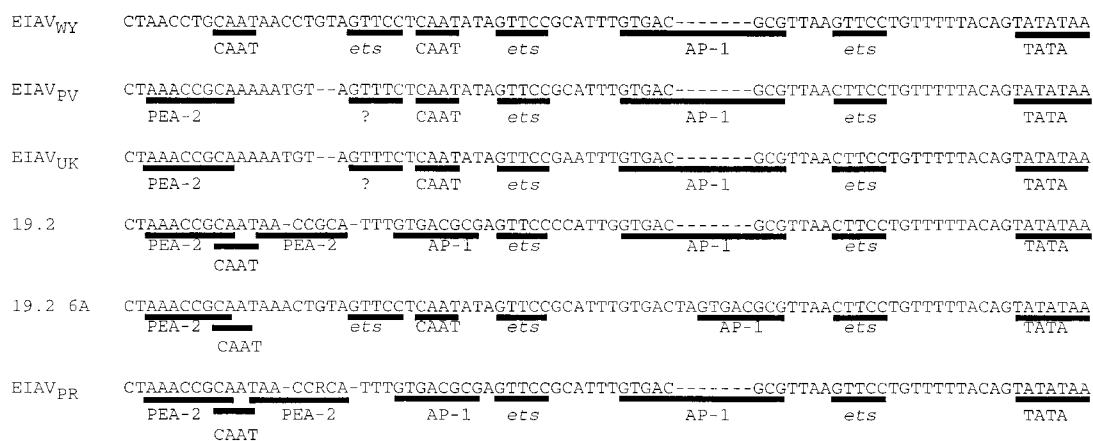


FIG. 4. Comparison of the U3-LTR hypervariable regions of the EIAV<sub>WY</sub> strain with the EIAV<sub>PV</sub> consensus sequence EIAV<sub>UK</sub>, the 19-2-6A sequence, and a consensus sequence for the EIAV<sub>PR</sub> strain. Potential *cis*-acting sequences (PEA-2, CAAT, *ets*, AP-1, and TATA) are underlined. The presence of a possible new *cis*-acting element is indicated by ?. Gaps (indicated by dashes) were introduced to align the sequence. In EIAV<sub>PR</sub>, R indicates either G or A, as at this location 54% of the population is represented by G and 46% is represented by A (29).



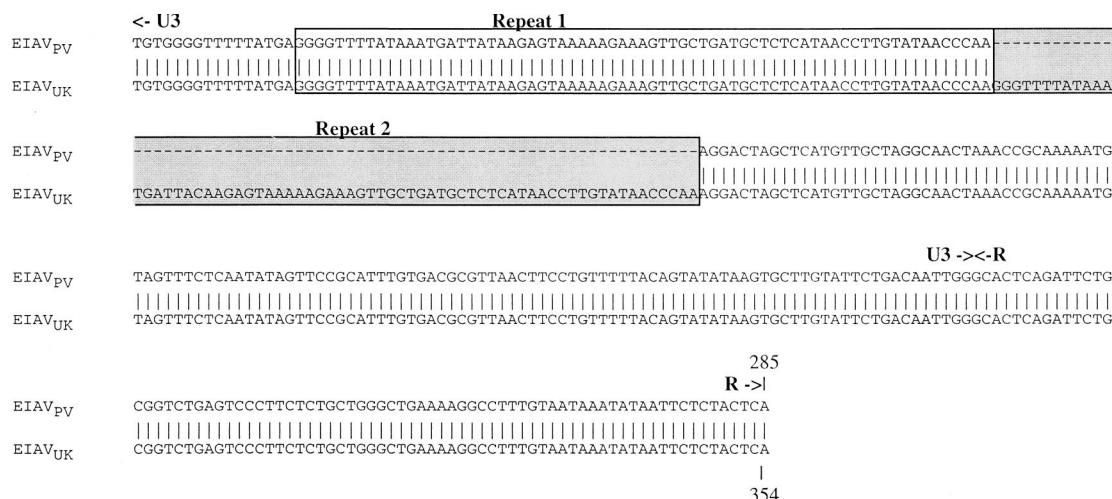


FIG. 5. Comparison of nucleotide sequences between EIAV<sub>UK</sub> and EIAV<sub>PV</sub> consensus sequences showing the location of the insertion/duplication in the 3' LTR. The first repeat of the 68-nucleotide region in U3 is indicated by a white box; the second repeat of the same sequence in EIAV<sub>UK</sub> is marked by a shaded box.

nies, two animals experienced only a single bout of disease with a modest 38% decline in platelet count, whereas the remaining two had multiple febrile episodes and a maximum 86% drop in platelets from preinfection levels (20). Likewise, in results reported here, ponies 614 and 700 experienced only a single febrile response, whereas horse 764 had three febrile episodes in 56 days. It appears that the spectrum of clinical responses obtained with viruses from molecular clone EIAV<sub>UK</sub> is very similar to that observed for the EIAV<sub>PV</sub> strain.

The initial reason for selecting the 3.3-kb fragment for generation of chimeric molecular clones was based on nucleotide sequence comparisons between pathogenic and nonpathogenic strain of EIAV. These comparisons demonstrated that most of the differences which could account for the phenotypic variation between strains are located in the 3'-terminal half of the viral genome. For example, compared to the virulent EIAV<sub>WY</sub> strain, the avirulent proviral molecular clone, CL-22, has nucleotide differences of 0.26% in the *gag* gene and 0.67% in the *pol* gene but differences of 3.09% in ORF S1, 3.83% in ORF S3, 5.57% in the SU glycoprotein coding sequences, 3.04% in the TM glycoprotein coding sequence, and 5.3% in the 3' LTR (46). Preliminary studies had also indicated that the majority of differences between the avirulent cell culture adapted Wyoming strain (EIAV<sub>PR</sub>) and the EIAV<sub>PV</sub> strain were also located in the 3' half of the viral genome. It seemed logical to assume that the regions of the viral genome having the greatest variability between pathogenic and nonpathogenic strains of EIAV should contain some, if not all, of the pathogenic determinants. This prediction has been confirmed by the results of this study.

The nucleotide sequence from position 1 in the 5' LTR to position 4889 in EIAV<sub>UK</sub> was identical to that of the parental 19-2-6A molecular clone. This finding demonstrated that no mutations which could alter the biological properties of the chimeric molecular clones had been introduced into the 5' half of the viral genome during the molecular cloning steps. A highly significant finding was that the coding sequences downstream of position 4889 in EIAV<sub>UK</sub> differed by just three nucleotides from the EIAV<sub>PV</sub> consensus sequence. These differences resulted in just a single amino acid change in which tryptophan at position 103 in ORF S3 in the EIAV<sub>PV</sub> consensus sequence is replaced by arginine in EIAV<sub>UK</sub>. An extensive analysis of the EIAV<sub>PV</sub> population both in vitro and in vivo has

been undertaken. Originally this analysis was restricted to the variable regions of the SU glycoprotein and the LTR (34). However, it was recently extended to include amplification and nucleotide sequencing of 3' 3,901-bp fragments obtained both from the EIAV<sub>PV</sub> stock and from plasma samples taken during febrile episodes (32). This demonstrated that the predominant members of the population present in the biologically cloned (50), tissue culture-derived EIAV<sub>PV</sub> stock maintain their dominance during initial replication in ponies and during the first febrile episode (32, 34). Therefore, replication in vivo does not result in major shifts in the population and selection of minority species. Consequently, the predominant members of the population in the EIAV<sub>PV</sub> stock, which may be defined as those closest to the consensus sequence (32), are probably the ones responsible for inducing disease. The nucleotide sequence analysis presented here demonstrates that EIAV<sub>UK</sub> contains amino acid sequences typical of those comprising the majority of the EIAV<sub>PV</sub> population. Complete homology was also observed between EIAV<sub>UK</sub> and the EIAV<sub>PV</sub> consensus sequence in all of the 3' LTR *cis*-acting elements that have been previously associated with the control of viral transcription (4, 5, 13). This extensive homology at the nucleotide level probably accounts for the very similar biological properties between viruses derived from the molecular clone EIAV<sub>UK</sub> and the EIAV<sub>PV</sub> strain. One major difference was, however, detected between the EIAV<sub>PV</sub> consensus sequence and the molecular clone EIAV<sub>UK</sub>. This was the presence of a 68-bp insertion/duplication which was located in a normally conserved area of the U3 region of the 3' LTR, not previously associated with *cis*-acting transcriptional control elements (4, 5, 13, 39). The role of this insertion/duplication in pathogenesis will be examined in future studies, although the available evidence indicates that LTRs containing it, both in the initial viral stock and during the first febrile episode, are a very minor species in the EIAV<sub>PV</sub> population (7, 34). Furthermore, comparisons between the titer in vivo and in vitro of the EIAV<sub>PV</sub> stock suggest that induction of disease is not restricted to a minority of the EIAV<sub>PV</sub> quasispecies (21). Therefore, it is questionable if this 68-bp insertion/duplication in the LTR functions as a major pathogenic determinant in the EIAV<sub>PV</sub> population. However, this does not preclude it from having a role within the specific context of molecular clone EIAV<sub>UK</sub>. It should be noted that substitutions of 3' LTR sequences in 19-2

molecular clones with ones identical to those in the EIAV<sub>PV</sub> strain, but without the insertion/duplication, have failed to produce virulent progeny viruses (34).

In addition to the possible involvement of this duplication in the 3' LTR, some predictions concerning the location of other potential pathogenic determinants can be inferred from the comparative nucleotide sequence analysis between EIAV<sub>UK</sub>, 19-2, and EIAV<sub>PV</sub>. For example, the sequences of integrase from nucleotide position 4889 and ORF S1 are identical between EIAV<sub>UK</sub>, the EIAV<sub>PV</sub> consensus sequence, 19-2, and a cloned representative of the EIAV<sub>PR</sub> strain. Therefore, within the context of 19-2- and EIAV<sub>PV3.3</sub>-based clones, it is unlikely that the integrase or Tat protein play a direct role in determining virulence. This is in contrast to a study based on the production of chimeras between SIV and human immunodeficiency virus type 1, where *tat* was implicated in pathogenicity (23).

In ORF S2, there are two amino acid substitutions at positions 4 and 19 between 19-2 and EIAV<sub>UK</sub> (Fig. 3A). At present, it is difficult to assign a role to these amino acid residues in pathogenicity. The fact that the nucleotide sequence of S2 in the avirulent EIAV<sub>PR</sub> strain is identical to that of EIAV<sub>UK</sub> and EIAV<sub>PV</sub> suggests that S2 is not associated with determinants of virulence. However, it should be noted that although the function of S2 is unknown, the majority of sequences examined (including those of EIAV<sub>PR</sub>, EIAV<sub>PV</sub>, and EIAV<sub>UK</sub>) contain a single nucleoporin motif (Fig. 3A) at the amino terminus (reviewed in reference 41). The leucine-to-valine change found in 19-2 could potentially disrupt this motif and perhaps further reduce the ability of viruses derived from this molecular clone to induce disease in vivo. Obviously further studies, including experiments to examine the functional significance of the nucleoporin motif, will be required to determine the role of S2 in pathogenesis.

A more consistent demarcation between virulent and avirulent viruses is seen in ORF S3, where isoleucine at position 14 and histidine at position 91 in EIAV<sub>UK</sub> (and EIAV<sub>PV</sub>) are represented by valine and tyrosine in 19-2 and EIAV<sub>PR</sub> (Fig. 3B; Table 2). However, less uniform substitutions were also observed in ORF S3. In 19-2, glycine is located at position 8, whereas the other strains contain a serine residue. Similarly, EIAV<sub>UK</sub> contained an arginine residue at position 103 instead of tryptophan found in EIAV<sub>PV</sub>, EIAV<sub>PR</sub>, and 19-2 (Fig. 3B; Table 2). Although the lack of complete correlation between these substitutions and pathogenicity suggests that they are not associated with virulence determinants, it is still possible that they are important in the context of an individual molecular clone. Several other amino acid differences between EIAV<sub>UK</sub>, 19-2, and EIAV<sub>PR</sub> are also apparent in the envelope gene, with most being found in the SU glycoprotein (Fig. 3C; Table 2).

In addition to variation in amino acid residues, there are clearly identifiable differences between EIAV<sub>WY</sub>, EIAV<sub>PR</sub>, and EIAV<sub>PV</sub> in the potential *cis*-acting transcriptional control elements located in the LTR. These differences appear to have evolved in response to viral growth conditions, which in turn suggests that they could have a role in cell tropism and/or pathogenesis. The hypervariable region in the LTR of the macrophage-tropic virulent EIAV<sub>WY</sub> strain contains two CAAT-box motifs, three potential *ets* protein binding domains, and a single AP-1 binding site (Fig. 4). Recently, we have undertaken an extensive analysis of the population of LTR sequences present in the EIAV<sub>PR</sub> strain (29) and found that the majority do not contain a 22-bp deletion as reported previously (35, 49). Instead, this analysis demonstrates that apparently during adaptation of the EIAV<sub>WY</sub> strain to fibroblastic cell cultures, one *ets* and one CAAT-box motif were replaced with two PEA-2

sites and an additional AP-1 binding motif (Fig. 4). Similarly, generation of the EIAV<sub>PV</sub> from the EIAV<sub>PR</sub> strain resulted in further changes, such as an alteration in the position of the CAAT box, the loss of PEA-2 and AP-1 binding sites and the creation of a possible new motif occupying the same relative position as the 5' *ets* site in the EIAV<sub>WY</sub> strain (Fig. 4). This new motif has some resemblance to the *ets* binding site except that the central core sequence is TTTC rather than TTCC (Fig. 4). However, as mentioned above, replacement of a 3' LTR with the EIAV<sub>PV</sub> consensus sequence in molecular clone 19-2-6A (designated EIAV<sub>PV100</sub>) did not result in the production of virulent progeny viruses (34). In fact, 19-2-6A was itself created with a 3' LTR resembling that of the EIAV<sub>WY</sub> strain in terms of having two CAAT-box motifs and three *ets* protein binding domains (Fig. 4). Therefore, if as suggested by the differences in *cis*-acting control elements between EIAV<sub>WY</sub>, EIAV<sub>PR</sub>, and EIAV<sub>PV</sub> the LTR does play a part in pathogenesis, it must act in concert with sequences elsewhere in the viral genome.

Originally it was considered that these additional sequences might be located in the coding region of the SU glycoprotein, as this glycoprotein has been implicated in the pathogenesis of some lentivirus diseases (23, 43, 47, 57). Furthermore, in EIAV most of the differences in amino acid content between isolates are located in this area of the viral genome, and in many cases these differences result in changes in the number of N-linked glycosylation sites (32, 34). However, viruses produced from a chimeric molecular clone containing a consensus EIAV<sub>PV</sub> SU glycoprotein gene sequence in addition to a EIAV<sub>PV</sub> 3' LTR (designated EIAV<sub>PV101</sub>) were also nonpathogenic when tested in one animal (34). If confirmed, this result would demonstrate that areas of the viral genome other than the hypervariable region of the LTR and/or the SU glycoprotein are involved in pathogenicity. These other areas could act either in isolation, with each other, or in combination with LTR and/or SU glycoprotein sequences. Perhaps one of the most intriguing conclusions from the comparative nucleotide sequence analysis and from the previously reported studies (34) is that these other areas must be limited to a very small number of nucleotides. In fact, based on differences between EIAV<sub>UK</sub> and 19-2, confirmation of the result with viruses from EIAV<sub>PV101</sub> would restrict these areas to two nucleotides in ORF S2 (at positions 4 and 19), potentially four nucleotides in ORF S3 (at positions 8, 14, 91, and 103), and two nucleotides in the TM glycoprotein (at positions 589 in the extracellular domain and 701 in the cytoplasmic domain). Interestingly, residues in TM in combination with SU sequences and an intact *nef* gene have recently been demonstrated to be essential for the optimal replication of SIV in brain cells derived from the micro-vessel endothelium and for SIV-mediated neurovirulence in macaques (16, 37). Experiments are in progress to test the involvement of differences observed between EIAV<sub>UK</sub> and 19-2-6A in pathogenesis. It is anticipated that production of additional chimeric molecular clones, coupled with the relatively simple genome organization of EIAV, will eventually enable identification of viral sequences associated with pathogenicity in this and possibly other lentiviruses. A potentially more difficult challenge may be to determine how these viral sequences interact with their host, thereby discovering why variation in relatively few nucleotides between strains of EIAV can produce such dramatic differences in clinical outcome.

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## REFERENCES

- Banapour, B., M. Marthas, R. Ramos, B. Lohman, R. Unger, M. Gardner, N. Pedersen, and P. Luciw. 1991. Identification of viral determinants of macrophage tropism for simian immunodeficiency virus SIVmac. *J. Virol.* **65**: 5798–5805.
- Barnes, W. M. 1994. PCR amplification of up to 35-kb DNA with high fidelity and high yield from  $\lambda$  bacteriophage templates. *Proc. Natl. Acad. Sci. USA* **91**:2216–2220.
- Carpenter, S., and B. Chesebro. 1989. Change in host cell tropism associated with *in vitro* replication of equine infectious anemia virus. *J. Virol.* **63**:2492–2496.
- Carvalho, M., and D. Derse. 1993. Physical and functional characterization of transcriptional control elements in the equine infectious anemia virus promoter. *J. Virol.* **67**:2064–2074.
- Carvalho, M., M. Kirkland, and D. Derse. 1993. Protein interactions with DNA elements in variant equine infectious anemia virus enhancers and their impact on transcriptional activity. *J. Virol.* **67**:6586–6595.
- Cheevers, W. P., and T. C. McGuire. 1985. Equine infectious anemia virus: immunopathogenesis and persistence. *Rev. Infect. Dis.* **7**:83–88.
- Cook, R. F., and C. J. Issel. Unpublished observations.
- Cook, R. F., S. J. Cook, and C. J. Issel. 1992. A non-radioactive micro-assay for released reverse transcriptase activity of a lentivirus. *BioTechniques* **13**:380–386.
- Cook, R. F., S. L. Berger, K. E. Rushlow, J. M. McManus, S. J. Cook, S. Harrold, M. L. Raabe, R. C. Montelaro, and C. J. Issel. 1995. Enhanced sensitivity to neutralizing antibodies in a variant of equine infectious anemia virus is linked to amino acid substitutions in the surface unit envelope glycoprotein. *J. Virol.* **69**:1493–1499.
- Cook, R. F., C. J. Issel, and R. C. Montelaro. 1996. Equine infectious anemia, p. 297–323. *In* M. J. Studdert (ed.), *Virus infections of equines*. Elsevier Science B.V., Amsterdam, The Netherlands.
- Costa, L. R., C. J. Issel, R. C. Montelaro, S. J. Cook, R. F. Cook, K. E. Rushlow, C. Grund, and S. Z.-S. Wang. 1994. Responses of ponies to challenge with equine infectious anemia virus following exposure to recombinant gp90 or viral p26 immunogens, p. 85–94. *In* H. Nakajima and W. Plowright (ed.), *Equine infectious diseases*, vol. VII. R&W Publications, Newmarket, England.
- Cunningham, T. P., R. C. Montelaro, and K. E. Rushlow. 1993. Lentivirus envelope sequences and proviral genomes are stabilized in *Escherichia coli* when cloned into low copy-number plasmid vectors. *Gene* **124**:93–98.
- Derse, D., P. L. Dorn, L. Levy, R. M. Stephens, N. R. Rice, and J. W. Casey. 1987. Characterization of equine infectious anemia virus long terminal repeat. *J. Virol.* **61**:743–747.
- Dorn, P., L. DaSilva, L. Martarano, and D. Derse. 1990. Equine infectious anemia virus Tat: insights into the structure, function, and evolution of lentivirus transactivation proteins. *J. Virol.* **64**:1616–1624.
- Dougherty, R. M. 1964. *In* R. J. C. Harris (ed.), *Techniques in experimental virology*, p. 169–220. Academic Press, London, England.
- Flaherty, M. T., D. A. Hauer, J. L. Mankowski, M. C. Zink, and J. E. Clements. 1997. Molecular and biological characterization of a neurovirulent molecular clone of simian immunodeficiency virus. *J. Virol.* **71**:5790–5798.
- Fridell, R. A., K. M. Partin, S. Carpenter, and B. R. Cullen. 1993. Identification of the activation domain of equine infectious anemia virus rev. *J. Virol.* **67**:7317–7323.
- Genetics Computer Group. 1994. Program manual for the Wisconsin Package. Genetics Computer Group, Madison, Wis.
- Gutekunst, D. E., and C. S. Becvar. 1979. Responses in horses infected with equine infectious anemia virus adapted to tissue culture. *Am. J. Vet. Res.* **40**:974–977.
- Hammond, S. A., S. J. Cook, D. L. Lichtenstein, C. J. Issel, and R. C. Montelaro. 1997. Maturation of the cellular and humoral immune responses to persistent infection in horses by equine infectious anemia is a complex and lengthy process. *J. Virol.* **71**:3840–3852.
- Issel, C. J. Unpublished observations.
- Issel, C. J., D. W. Horohov, D. F. Lea, W. V. Adams, Jr., S. D. Hagius, J. M. McManus, A. C. Allison, and R. C. Montelaro. 1992. Efficacy of inactivated whole-virus and subunit vaccines in preventing infection and disease caused by equine infectious anemia virus. *J. Virol.* **66**:3398–3408.
- Karlsson, G. B., M. Halloran, J. Li, L.-W. Park, R. Gomila, K. A. Reimann, M. K. Axthelm, S. A. Iliff, N. L. Letvin, and J. Sodroski. 1997. Characterization of molecularly cloned simian-human immunodeficiency viruses causing rapid CD4<sup>+</sup> lymphocyte depletion in rhesus monkeys. *J. Virol.* **71**:4218–4225.
- Kobayashi, K. 1961. Studies on the cultivation of equine infectious anemia virus *in vitro*. I. Serial cultivation of the virus in the culture of various horse tissues. *Virus* **11**:177–189.
- Kobayashi, K., and Y. Kono. 1967. Propagation and titration of equine infectious anemia virus in horse leukocyte culture. *Natl. Inst. Anim. Health Q.* **7**:8–20.
- Kono, Y., and Y. Yokomizo. 1968. Attempts to cultivate the equine infectious anemia virus in various cell types. *Natl. Inst. Anim. Health Q.* **8**:182–186.
- Kono, Y., T. Yoshino, and Y. Fukunaga. 1970. Growth characteristics of equine infectious anemia virus in horse leukocyte cultures. *Arch. Virol.* **30**:252–256.
- Kraft, R., J. Tardiff, K. S. Krauter, and L. A. Leinwand. 1988. Using mini-prep plasmid DNA for sequencing double stranded template with Sequenase. *BioTechniques* **6**:544–546.
- Langemeier, J. L., and C. J. Issel. Unpublished observations.
- Langemeier, J. L., S. J. Cook, R. F. Cook, K. E. Rushlow, R. C. Montelaro, and C. J. Issel. 1996. Detection of equine infectious anemia viral RNA in plasma samples from recently infected and long-term inapparent carrier animals by PCR. *J. Clin. Microbiol.* **34**:1481–1487.
- Leroux, C., and R. C. Montelaro. Unpublished observations.
- Leroux, C., C. J. Issel, and R. C. Montelaro. 1997. Novel and dynamic evolution of equine infectious anemia virus genomic quasispecies associated with sequential disease cycles in an experimentally infected pony. *J. Virol.* **71**:9627–9639.
- Lichtenstein, D. L., K. E. Rushlow, R. F. Cook, M. L. Raabe, C. J. Swardson, G. J. Kociba, C. J. Issel, and R. C. Montelaro. 1995. Replication *in vitro* and *in vivo* of an equine infectious anemia virus mutant deficient in dUTPase activity. *J. Virol.* **69**:2881–2888.
- Lichtenstein, D. L., C. J. Issel, and R. C. Montelaro. 1996. Genomic quasispecies associated with the initiation of infection and disease in ponies experimentally infected with equine infectious anemia virus. *J. Virol.* **70**: 3346–3354.
- Madden, C. R., and D. S. Shih. 1996. Analysis of the long terminal repeat from a cytopathic strain of equine infectious anemia virus. *Virology* **225**:395–399.
- Malmquist, W. A., D. Barnett, and C. S. Becvar. 1973. Production of equine infectious anemia antigen in a persistently infected cell line. *Arch. Virol.* **42**:361–370.
- Mankowski, J. L., M. T. Flaherty, J. P. Spelman, D. A. Hauer, P. J. Didier, A. M. Amedee, M. Murphey-Corb, L. M. Kirstein, A. Mumoz, J. E. Clements, and M. C. Zink. 1997. Pathogenesis of simian immunodeficiency virus encephalitis: viral determinants of neurovirulence. *J. Virol.* **71**:6055–6060.
- Marthas, M. L., R. A. Ramos, B. L. Lohman, K. K. van Rompay, R. E. Unger, C. J. Miller, B. Banapour, N. C. Pedersen, and P. L. Luciw. 1993. Viral determinants of simian immunodeficiency virus (SIV) virulence in rhesus macaques assessed by using attenuated and pathogenic molecular clones of SIVmac. *J. Virol.* **67**:6047–6055.
- Maury, W. 1994. Monocyte maturation controls expression of equine infectious anemia virus. *J. Virol.* **68**:6270–6279.
- Montelaro, R. C., J. M. Ball, and K. E. Rushlow. 1993. Equine retroviruses, p. 257–360. *In* J. A. Levy (ed.), *The Retroviridae*, vol. 2. Plenum Press, New York, N.Y.
- Nigg, E. A. 1997. Nucleocytoplasmic transport: signals, mechanisms and regulation. *Nature* **2386**:779–787.
- Noiman, S., A. Yaniv, L. Sherman, S. R. Tronick, and A. Gazit. 1990. Pattern of transcription of the genome of equine infectious anemia virus. *J. Virol.* **64**:1839–1843.
- Novembre, F. J., P. R. Johnson, M. G. Lewis, D. C. Anderson, S. Klumpp, H. M. McClure, and V. A. Hirsch. 1993. Multiple viral determinants contribute to pathogenicity of the acutely lethal simian immunodeficiency virus SIVsmmPBj variant. *J. Virol.* **67**:2466–2474.
- Orrego, A., C. J. Issel, R. C. Montelaro, and W. V. Adams, Jr. 1982. Virulence and *in vitro* growth of a cell-adapted strain of equine infectious anemia virus after serial passage in ponies. *Am. J. Vet. Res.* **43**:1556–1560.
- Payne, S. L., J. Rausch, K. Rushlow, R. C. Montelaro, C. Issel, M. Flaherty, S. Perry, D. Sellon, and F. Fuller. 1994. Characterization of infectious molecular clones of equine infectious anemia virus. *J. Gen. Virol.* **75**:425–429.
- Perry, S. T., M. T. Flaherty, M. J. Kelley, D. L. Clabough, S. R. Tronick, L. Coggins, L. Whetter, C. R. Lengel, and F. Fuller. 1992. The surface envelope protein gene region of equine infectious anemia virus is not an important determinant of tropism *in vitro*. *J. Virol.* **66**:4085–4097.
- Power, C., J. C. McArthur, R. T. Johnson, D. E. Griffin, J. D. Glass, S. Perryman, and B. Chesebro. 1994. Demented and nondemented patients with AIDS differ in brain-derived human immunodeficiency virus type 1 envelope sequences. *J. Virol.* **68**:4643–4649.
- Rosin-Arbesfeld, R., M. Rivlin, S. Noiman, P. Mashiah, A. Yaniv, T. Miki, S. R. Tronick, and A. Gazit. 1993. Structural and functional characterization of *rev*-like transcripts of equine infectious anemia virus. *J. Virol.* **67**:5640–5646.
- Rushlow, K., K. Olsen, G. Stiegler, S. L. Payne, R. C. Montelaro, and C. J. Issel. 1986. Lentivirus genomic organization: the complete nucleotide se-

- quence of the *env* gene region of equine infectious anemia virus. *Virology* **155**:309–321.
50. **Rwambo, P. M., C. J. Issel, K. A. Hussain, and R. C. Montelaro.** 1990. *In vitro* isolation of a neutralization escape mutant of equine infectious anemia virus (EIAV). *Arch. Virol.* **111**:275–280.
  51. **Sambrook, J., E. F. Fritsch, and T. Maniatis.** 1989. *Molecular cloning: a laboratory manual*, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
  52. **Schiltz, R. L., D. S. Shih, S. Rasty, R. C. Montelaro, and K. E. Rushlow.** 1992. Equine infectious anemia virus gene expression: characterization of the RNA splicing pattern and the protein products encoded by open reading frames S1 and S2. *J. Virol.* **66**:3455–3465.
  53. **Sellon, D. C., S. T. Perry, L. Coggins, and F. J. Fuller.** 1992. Wild-type equine infectious anemia virus replicates *in vivo* predominantly in tissue macrophages, not in peripheral blood monocytes. *J. Virol.* **66**:5906–5913.
  54. **Sellon, D. C., F. J. Fuller, and T. C. McGuire.** 1994. The immunopathogenesis of equine infectious anemia virus. *Virus Res.* **32**:111–138.
  55. **Stephens, R. M., J. W. Casey, and N. R. Rice.** 1986. Equine infectious anemia virus gag and pol genes: relatedness to Visna and AIDS virus. *Science* **231**:589–594.
  56. **Stephens, R. M., D. Derse, and N. R. Rice.** 1990. Cloning and characterization of cDNA encoding equine infectious anemia virus tat and putative rev proteins. *J. Virol.* **64**:3716–3725.
  57. **Toggas, S. M., E. Masliah, E. M. Rockenstein, G. F. Rall, C. R. Abraham, and L. Mucke.** 1994. Central nervous system damage produced by expression of the HIV-1 coat protein gp120 in transgenic mice. *Nature* **367**:188–193.
  58. **Torsteinsdottir, S., G. Agnarsdottir, S. Matthiasdottir, B. Rafnar, V. Andressdottir, O. S. Andresson, K. Staskus, G. Petursson, P. A. Palsson, and G. Georgsson.** 1997. *In vivo* and *in vitro* infection with two different molecular clones of visna virus. *Virology* **229**:370–380.
  59. **Ushimi, C., J. B. Henson, and J. R. Gorham.** 1972. Study of the one-step growth curve of equine infectious anemia virus by immunofluorescence. *Infect. Immun.* **5**:890–895.
  60. **Wang, S. Z.-S., K. E. Rushlow, C. J. Issel, R. F. Cook, S. J. Cook, M. L. Raabe, Y.-H. Chong, L. Costa, and R. C. Montelaro.** 1994. Enhancement of EIAV replication and disease by immunization with a baculovirus-expressed recombinant envelope surface glycoprotein. *Virology* **199**:247–251.