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Exploration of muscle from GRMD dogs tranplanted with MuStem cells using "omics" approaches

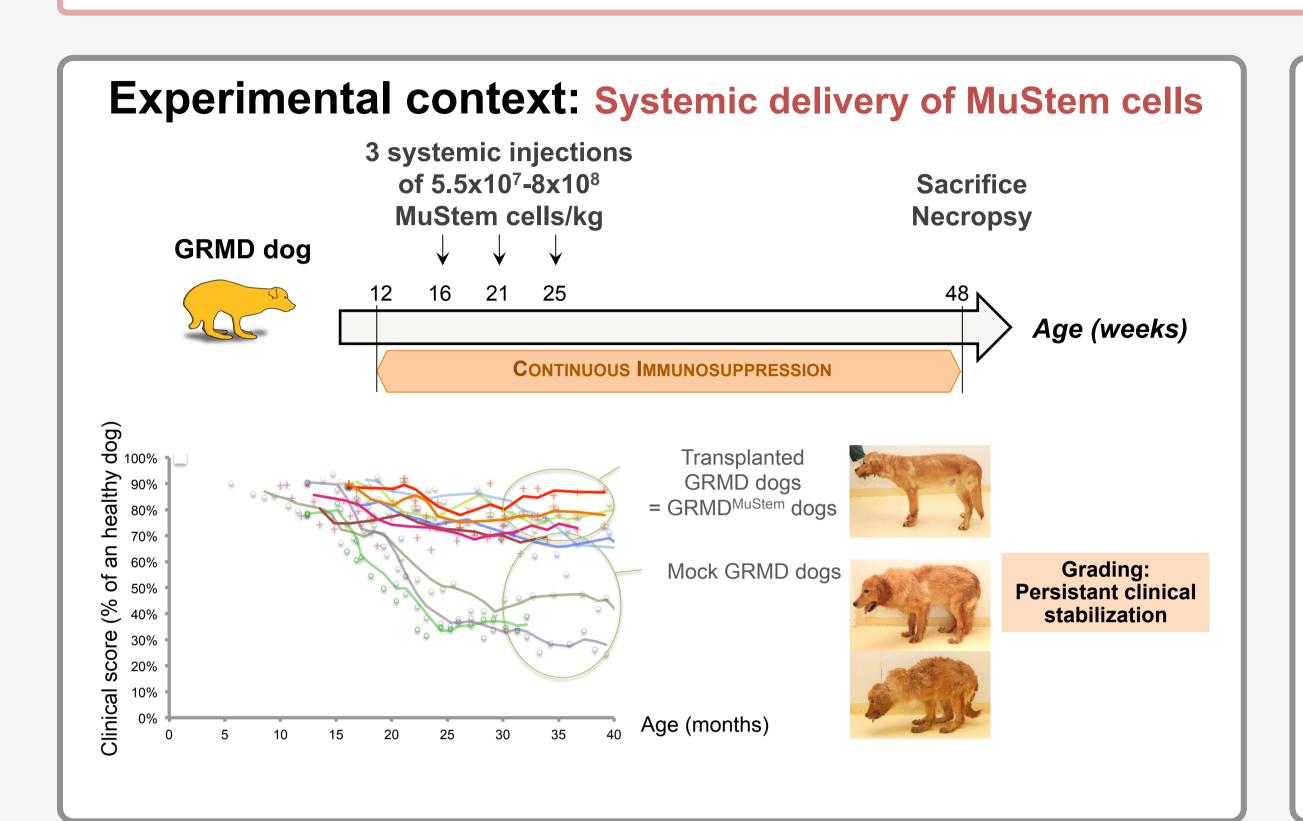
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Introduction - Duchenne Muscular Dystrophy (DMD), the most common form of inherited neuromuscular disorder, is caused by mutations in the dystrophin gene leading to the protein lack. Membrane disorganization and subsequent alterations in signaling pathways and energy metabolism play important roles in muscle fibre necrosis. Systemic delivery of MuStem cells, skeletal muscle-resident stem cells isolated from healthy dog, generates a remodeling of muscle tissue and gives rise to striking clinical benefits in Golden Retriever Muscular Dystrophy (GRMD) dog (Rouger et al., 2011). To pursue investigation of the consequences on the skeletal muscle tissue 6 months after cell transplantation with undedicated approach, we used here a combined analysis of transcriptomics (gene expression microarrays) and quantitative proteomics (ICPL/LC-MS/MS) (Robriquet et al., 2015; Lardenois et al., in preparation). In addition, we establish that the analysis of a limited set of miRNAs in skeletal muscle discriminates MuStem cell therapy effects on GRMD dogs.

Objective - To pave the way to the understanding of the mechanisms underlying the therapeutical effect of the MuStem cells and identify potential biomarkers of the treatment efficiency we employed combined "omics" approaches that allowed us to identify changes in transcript and protein profiles in Healthy, mock GRMD and Treated GRMD muscles (GRMD^{MuStem}).



Material and methods

We performed comparative transcriptomic and proteomic studies between healthy, mock GRMD (received continuous immunosuppressive regimen) and GRMD^{MuStem} dog muscles (MuStem cell transplantation under the same immunosuppression) in order to identify changes in transcript and protein profiles. Biceps femoris muscle samples were collected surgically from the middle portion of the muscle in 9-month-old healthy, mock GRMD dogs and GRMD^{MuStem} dogs (n=2: transcriptomic; n=3: proteomic).

Transcriptomic study

- -The transcriptome experiment was performed by the integrative Genomic core facility (Nantes, France). mRNA expression profiling was obtained using the Agilent-021193 Canine (V2) Gene Expression Microarray.
- Statistical filtration and classification were performed. The detectable probes yielding signals ≥ 7.32 were identified and probes displaying a high expression variation (fold change ≥ 2) were then selected. A LIMMA statistical test was done.

Proteomic study

- •The proteomic experimentation was performed by the Biogenouest Proteomic core facility (Rennes, France). We used isotope-coded protein labeling for quantitative proteomic analysis. LC-MS/MS analyses performed on an ESI-LTQ-Orbitrap mass spectrometer led to the relative quantification of 492 proteins.
- •Statistical filtration and classification were performed. The quantified proteins displaying a high ratio (absolute value of the median of the ratio data in a triplex ≥1.5) in at least two triplex were identified. A LIMMA statistical test was then used to identify the proteins that were significantly differentially expressed (F-value adjusted with the False Discovery Rate, p≤0.05).

miRNA study

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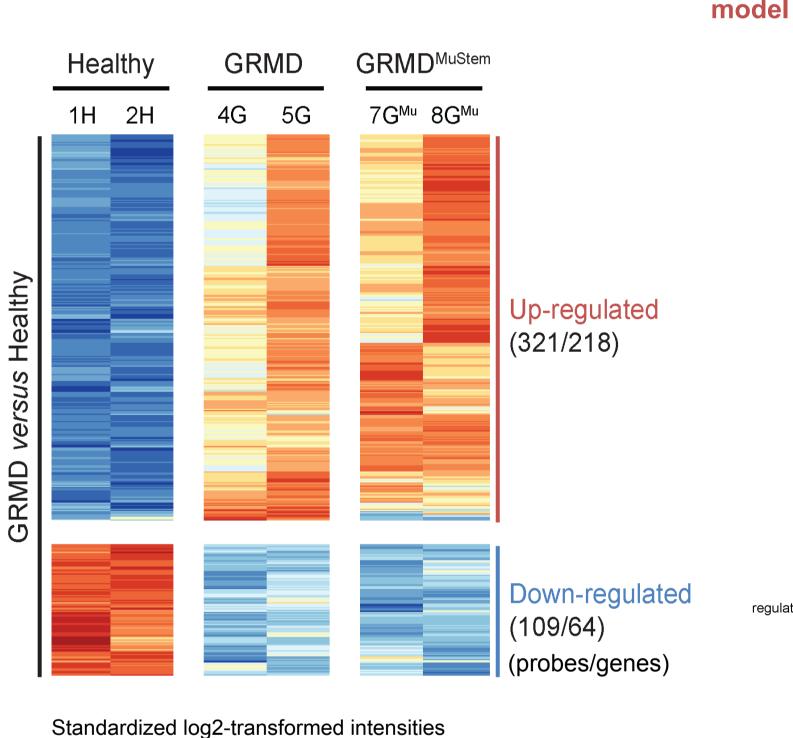
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- The expression levels of 5 miRNAs (miR-1, miR-133, miR-206, miR-222 and miR-486) were determined by RT-qPCR in Biceps femoris of 9month-old healthy, mock GRMD dogs and GRMD^{MuStem} dogs (n=6).

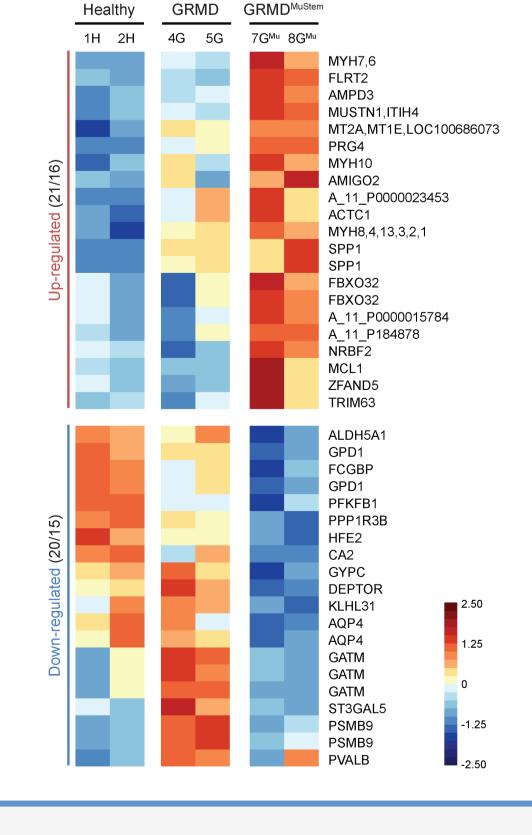
Transcriptomic study

A- Profiling of differentially expressed transcripts



Standardized log2-transformed intensities

C- Impact of MuStem cell delivery



The high-throughput characterization of the GRMD dog model reveals an altered expression profile of 282 genes. Up-regulated genes are significantly associated with genes

implicated in regeneration, extracellular matrix

organization, inflammatory response, cell

adhesion and cell migration.

regulation of transmembrane transporter activity

Molecular function

heat shock protein binding

Cellular component

B- Functional characterization of GRMD dog

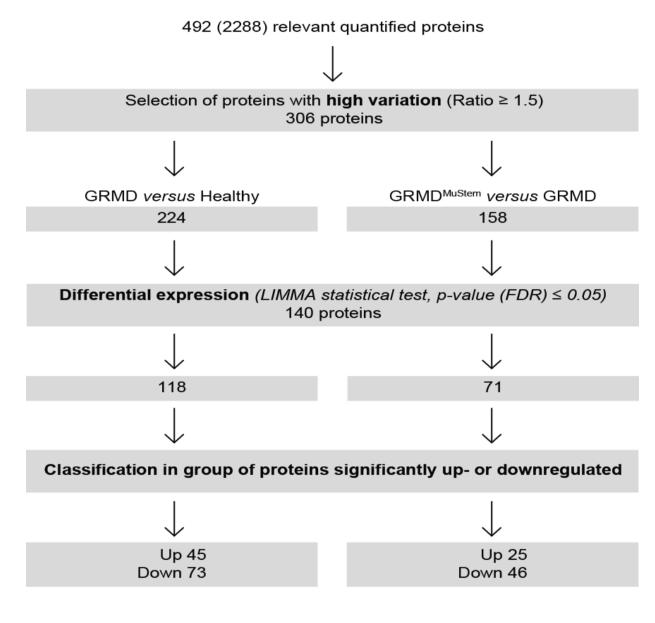
Gene Ontology enrichment analysis

MuStem cell administration is associated with gene expression modification of 31 factors implicated in different biological functions, among them: regeneration, protein degradation and folding, myogenesis, lipid and glucose metabolism, homeostasis.

Robriquet et al., PLoS One (2015)

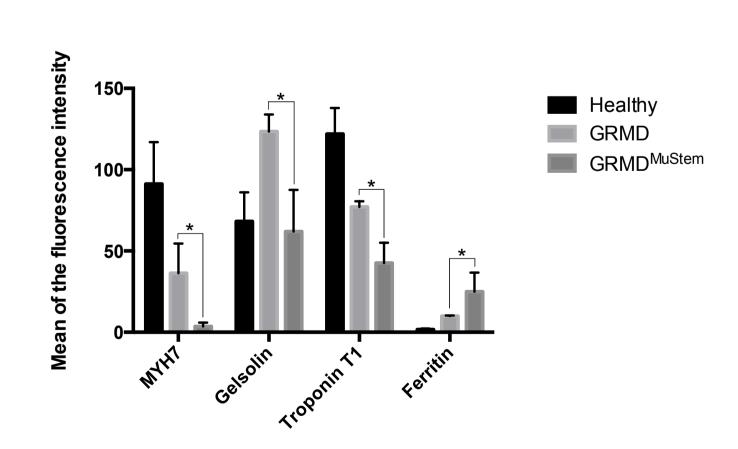
Proteomic study

A- Flow chart of the statistical filtration of the ICPL data



✓ The 492 proteins considered as correctly quantified. were filtered up to groups of **up-** and **downregulated** proteins in the GRMD versus Healthy comparison and in the GRMD^{MuStem} versus GRMD comparison.





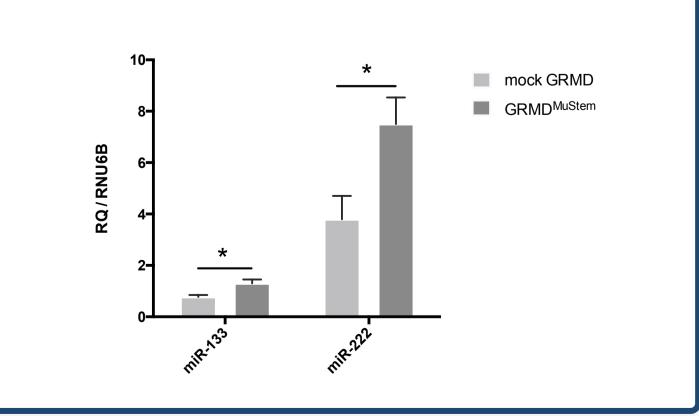
B- ICPL data validation. Ratios ICPL versus Western Blot

Protein name	Triplex	ICPL ratio	Average ICPL ratio	WB ratio	Average WB ratio
MYH7	T ₁	0.55		0.13	
	T ₂	0.18	0.27±0.25	0.29	0.23±0.09
	Т3	0.07		0.27	
Gelsolin	T1	0.42		0.77	
	T ₂	0.81	0.60±0.2	0.8	0.7±0.15
	Т3	0.56		0.52	
Vimentin	T ₁	0.36		0.31	
	T ₂	0.48	0.56±0.24	0.64	0.57±0.23
	T ₃	0.83		0.76	
Haptoglobin	T ₁	3.58		3.35	
	T2	2.36	0.29±0.33	1.20	1.83±1.33
	Т3	1.82		0.93	
Troponin Tı	T ₁	0.18		0.67	
	T ₂	0.03	0.29±0.33	0.73	0.65±0.09
	Т3	0.66		0.55	
Ferritin	T ₁	20.97		4.08	
	T ₂	6.96	11.65±8.07	2.86	3.01±1
	Т3	7.01		2.09	

- ✓ Quantitative MS and WB data comparison. To validate the protein changes detected by proteomic analyses, immunoblotting experiments were performed on a representative range of protein types
- Myobifrillar reorganization. Inhibition of myogenic and sarcomeric proteins after MuStem cell tranplantation (MYH7, MHC, Troponin)
- Protection of oxydative stress. MuStem cell administration enhances skeletal muscle cells protection against oxidative stress (Haptoglobin, Ferritin)

miRNA study

- The expression levels of 5 miRNAs (miR-1, miR-133, miR-206, miR-222, and miR-486) and some targets of relevant miRNA were determined by RT-qPCR and WB in Biceps femoris of all dogs.
- The analysis of a miR-133 and miR-222 in skeletal muscle discriminates MuStem cell therapy effects on GRMD dogs



Conclusions

Overall, the combination of transcriptomics, proteomics and miRNA approaches allowed to pave the way to the understanding of MuStem cell action modalities.

Interestingly, we show that MuStem cell administration is associated with expression pattern modification of many factors belonging to different biological functions such as muscle mass regeneration, myfibrillar reorganization, metabolism, lipid and energy homeostasis. In addition, we demonstrate an effect of MuStem cell transplantation on the muscle expression of miR-133 and miR-222.

This strategy has a great potential to considerably contribute to the identification of therapeutic biomarkers of MuStem cell transplantation and thus represents an interesting tool to monitor therapeutic effects during DMD-dedicated preclinical studies.

Fundings

